

# Relationship between Membrane Phosphatidylinositol-4,5-Bisphosphate and Receptor-Mediated Inhibition of Native Neuronal M Channels

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The relationship between receptor-induced membrane phosphatidylinositol-4'5'-bisphosphate (PIP<sub>2</sub>) hydrolysis and M-current inhibition was assessed in single-dissociated rat sympathetic neurons by simultaneous or parallel recording of membrane current and membrane-to-cytosol translocation of the fluorescent PIP<sub>2</sub>/inositol 1,4,5-trisphosphate (IP<sub>3</sub>)-binding peptide green fluorescent protein-tagged pleckstrin homology domain of phospholipase C (GFP-PLCδ-PH). The muscarinic receptor agonist oxotremorine-M produced parallel time- and concentration-dependent M-current inhibition and GFP-PLCδ-PH translocation; bradykinin also produced parallel time-dependent inhibition and translocation. Phosphatidylinositol-4-phosphate-5-kinase (PI5-K) overexpression reduced both M-current inhibition and GFP-PLCδ-PH translocation by both oxotremorine-M and bradykinin. These effects were partly reversed by wortmannin, which inhibits phosphatidylinositol-4-kinase (PI4-K). PI5-K overexpression also reduced the inhibitory action of oxotremorine-M on PIP<sub>2</sub>-gated G-protein-gated inward rectifier (Kir3.1/3.2) channels; bradykinin did not inhibit these channels. Overexpression of neuronal calcium sensor-1 protein (NCS-1), which increases PI4-K activity, did not affect responses to oxotremorine-M but reduced both fluorescence translocation and M-current inhibition by bradykinin. Using an intracellular IP<sub>3</sub> membrane fluorescence-displacement assay, initial mean concentrations of membrane [PIP<sub>2</sub>] were estimated at 261 μM (95% confidence limit; 192–381 μM), rising to 693 μM (417–1153 μM) in neurons overexpressing PI5-K. Changes in membrane [PIP<sub>2</sub>] during application of oxotremorine-M were calculated from fluorescence data. The results, taken in conjunction with previous data for KCNQ2/3 (Kv7.2/Kv7.3) channel gating by PIP<sub>2</sub> (Zhang et al., 2003), accorded with the hypothesis that the inhibitory action of oxotremorine-M on M current resulted from depletion of PIP<sub>2</sub>. The effects of bradykinin require additional components of action, which might involve IP<sub>3</sub>-induced Ca<sup>2+</sup> release and consequent M-channel inhibition (as proposed previously) and stimulation of PIP<sub>2</sub> synthesis by Ca<sup>2+</sup>-dependent activation of NCS-1.

**Key words:** PIP<sub>2</sub>; M current; neuronal excitability; G-protein-coupled receptors; PLC; sympathetic neurons

## Introduction

M channels are low-threshold K<sup>+</sup> channels that regulate neuronal excitability in both the CNS and the PNS (Brown and Adams, 1980; Brown, 1988; Marrion, 1997). They are composed of subunits of the KCNQ (Kv7) gene family (Wang et al., 1998; Jentsch, 2000). The channels are inhibited by stimulating G<sub>q/11</sub>-coupled receptors, for example in sympathetic neurons by M<sub>1</sub> muscarinic acetylcholine receptors (M<sub>1</sub>-mAChRs) (Marrion et al., 1989;

Bernheim et al., 1992) and by bradykinin (BK) B<sub>2</sub> receptors (Jones et al., 1995). The activated G-protein subunit [principally Gα<sub>q</sub> in the rat sympathetic neuron (Haley et al., 1998)] does not appear to gate the M channels directly but instead induces their closure by some indirect mechanism (Selyanko et al., 1992; Marrion, 1993).

Recent evidence suggests that receptor-induced closure results from hydrolysis of the membrane phospholipid, phosphatidylinositol-4'5'-bisphosphate (PIP<sub>2</sub>) (Suh et al., 2004). As with a number of other channels (Hilgemann et al., 2001; Runnels et al., 2002; Loussouarn et al., 2003), PIP<sub>2</sub> is required to maintain KCNQ channels in their open state (Zhang et al., 2003), and resynthesis of PIP<sub>2</sub> is necessary for recovery from M-channel inhibition produced by mAChRs (Suh and Hille, 2002) and by nucleotide receptors (Ford et al., 2003). Thus, mAChR-induced closure might result directly from the local depletion of PIP<sub>2</sub> consequent on its hydrolysis. In contrast, BK-induced M-channel inhibition more likely stems from the action of a downstream product of PIP<sub>2</sub> hydrolysis, inositol 1,4,5-trisphosphate (IP<sub>3</sub>), which releases Ca<sup>2+</sup> from intracellular stores and thereby closes

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channels via binding to calmodulin (Selyanko and Brown, 1996; Cruzblanca et al., 1998; Bofill-Cardona et al., 2000; Delmas et al., 2002; Gamper and Shapiro, 2003). Another product of PIP<sub>2</sub> hydrolysis, diacylglycerol, might contribute to mAChR-induced inhibition of M channels through activation of protein kinase C and subsequent channel phosphorylation (Hoshi et al., 2003). These alternative scenarios are summarized by Delmas et al. (2004).

Because both muscarinic agonists and bradykinin are clearly capable of activating phospholipase C (PLC) in sympathetic neurons (Del Rio et al., 1999), we examined more closely the relationship between PIP<sub>2</sub> hydrolysis and M-channel closure in these neurons. For this, we used a fluorescent IP<sub>3</sub>/PIP<sub>2</sub>-binding peptide [the green fluorescent protein-tagged pleckstrin homology domain of PLC $\delta$  (GFP-PLC $\delta$ -PH)] (Stauffer et al., 1998; Varnai and Balla, 1998; Nahorski et al., 2003). We then asked what happens when the synthesis of PIP<sub>2</sub> is enhanced by overexpressing the enzyme phosphatidylinositol-4-phosphate-5-kinase (PI5-K) (Ishihara et al., 1996) or the neuronal calcium sensor-1 protein (NCS-1; frequenin), which increases the activity of phosphatidylinositol-4-kinase (PI4-K) (Burgoyne et al., 2004). Finally, we compared these effects of PI5-K overexpression with its effects on receptor-induced inhibition of G-protein-gated inward rectifier (GIRK; Kir3.1+Kir3.2) channels, which has been attributed to PIP<sub>2</sub> depletion (Kobrinisky et al., 2000; Lei et al., 2001; Meyer et al., 2001; Bender et al., 2002).

Preliminary reports of our observations have been published previously (Winks and Marsh, 2003; Winks et al., 2003)

## Materials and Methods

**Neuron cultures.** Superior cervical ganglia (SCGs) were isolated from 17-d-old Sprague Dawley rats (either gender) after terminal anesthesia using CO<sub>2</sub>, followed by decapitation in accordance with United Kingdom Home Office regulatory procedures. Ganglia were digested in collagenase solution (800 international units of collagenase class 2; Sigma-Aldrich, Dorset, UK), 12 mg of bovine serum albumin (BSA; Sigma-Aldrich), and 2 ml of HBSS plus HEPES for 15 min at 37°C. Ganglia were then rinsed with HBSS plus 10 mM HEPES and placed in trypsin solution [2 mg of trypsin, bovine type XII S (Sigma-Aldrich), 12 mg of BSA, 2 ml of HBSS plus 10 mM HEPES] for 30 min at 37°C. Digested fragments were then transferred into 2 ml of growth medium [42 ml of L-15 plus GlutaMAX (Sigma-Aldrich), 8.5 ml of 1.26% NaHCO<sub>3</sub> (24 mM; Invitrogen, San Diego, CA), 5 ml of fetal calf serum (Sigma-Aldrich), 1.15 ml of 30% glucose, 0.61 ml of 200 mM L-glutamine (2 mM; Invitrogen), 50 ng/ml NGF 7S (Sigma-Aldrich)], and the mixture was triturated and then centrifuged five times, after which the remaining cell suspension was spun for 5 min at 800 rpm. The supernatant was then discarded, and the cells were resuspended in 5 ml of growth media. For electrophysiological recording/imaging, ~500  $\mu$ l of cell solution was added to each sterile culture/recording chamber consisting of a glass ring (diameter, 16 mm; height, 3 mm) attached to a laminin substrate-coated borosilicate glass coverslip (22  $\times$  22 mm; thickness, 0.13–0.17 mm; BDH Chemicals, Poole, UK) placed inside a 35 mm Petri dish. Chambers were then incubated at 37°C in 95% O<sub>2</sub>/5% CO<sub>2</sub> for at least 4 h to allow neurons to adhere to the coverslip. Neurons were usually transfected/recorded between 1 and 4 d after plating.

**Transfection.** Neurons were transfected with cDNA 1–2 d after culture by intranuclear microinjection of cDNA plasmids. Sharp electrodes (>50 M $\Omega$  resistance) were filled with plasmid dissolved in injection solution (in mM: 154 NaCl, 5 HEPES, 2.5 KCl, and 0.5 MgCl<sub>2</sub>, pH adjusted to 7.4 using NaOH) at a concentration of 100 ng/ $\mu$ l of each cDNA plasmid to be injected. Injections were performed using an Eppendorf Scientific (Westbury, NY) automated microinjector (Transjector 5246) or (for Kir 3.1/3.2 cDNAs) a Narishige (Tokyo, Japan) micromanipulator. Neurons were incubated for 1 d before use unless otherwise stated. The following plasmids were used: pleckstrin-homology domain of

phospholipase C- $\delta$ <sub>1</sub> in enhanced GFP (eGFP)-C1 vector (GFP-PLC- $\delta$  PH) [obtained from T. Meyer (Stanford University, Stanford, CA) via K. Young (University of Leicester, Leicester, UK)]; rat neuronal calcium sensor protein-1 in pcDNA 3.1 vector (obtained from J. Weiss, University of Sheffield, Sheffield, UK); murine phosphatidylinositol 4-phosphate-5-kinase-I $\beta$  in pcDNA 3.1 vector containing GFP sequence (GFPmPIP5K-I $\beta$ ; here, PI5-K; obtained from J. N. Davis, Louisiana State University Health Sciences Center, Shreveport, LA); mutant phosphatidylinositol 4-phosphate-5-kinase-I $\beta$ , lacking kinase activity, in pcDNA 3.1 vector containing GFP sequence [GFPmPIP5K-I $\beta$ (K179M); here, mutant PI5-K; obtained from J. N. Davis]; Kir3.1 and Kir3.2 (GIRK1 and GIRK2) in pCI and pRC/cytomegalovirus (CMV) vectors, respectively (from F. Lesage, Sophia Antipolis, Valbonne, France); enhanced green fluorescent protein in pcDNA 3.1 vector (eGFP) and red fluorescent cloning vector (pDSRed2-N1; Clontech, Cambridge, UK). All plasmids used contained the CMV promoter. Plasmids were propagated in DH5 $\alpha$  (Invitrogen) *Escherichia coli* and purified using Qiagen (Hilden, Germany) maxiprep columns.

**Electrophysiological recording.** The perforated-patch configuration of the patch-clamp technique was used (except where otherwise stated) with amphotericin B as the pore-forming agent (Rae et al., 1991). Neurons were superfused via a gravity-driven system (10–15 ml/min; bath volume, 500  $\mu$ l) with an external solution containing the following (in mM): 136 NaCl, 11.1 glucose, 5 HEPES, 3 KCl, 2.5 CaCl<sub>2</sub>, and 1.5 MgCl<sub>2</sub>, pH adjusted to 7.4 with NaOH. For inward rectifier (Kir 3.1/3.2, GIRK) current recording, the KCl concentration in the bath solution was increased to 6 mM (KCl substituted for NaCl) and tetrodotoxin (0.5  $\mu$ M) added to block the Na<sup>+</sup> current. Pipettes with 2–3 M $\Omega$  resistance were filled with an internal solution containing the following (in mM): M current, 30 KCl, 100 K acetate, 20 HEPES, 10 EGTA, and 3 MgCl<sub>2</sub>, pH 7.2 with KOH; Kir/GIRK current, 20 KCl, 90 K acetate, 3 MgCl<sub>2</sub>, 40 HEPES, and 0.1 BAPTA, pH 7.4 with KOH; and N-type Ca<sup>2+</sup> current, 140 CsCl, 1 MgCl<sub>2</sub>, 0.1 BAPTA, 0.1 CaCl<sub>2</sub>, and 10 HEPES, CsOH to pH 7.4. All recordings were made at room temperature (19–22°C) unless otherwise stated. Currents were recorded using an Axopatch 200A or 200B amplifier (except where otherwise stated), filtered at 1 kHz, and digitized at 1–10 kHz. Pipette series resistances were compensated 60–90%.

**M currents.** Cells were held at –20 mV to preactivate the current. Currents were then deactivated at intervals of 10–15 s by 1 s hyperpolarizing steps to –50 mV. Current amplitudes were measured from extrapolated deactivation current tails using a biexponential curve-fitting procedure.

**Ca<sup>2+</sup> currents.** N-type Ca<sup>2+</sup> currents were recorded by holding the cell at –70 mV and depolarizing to +5 mV for 200 ms. Currents were leak subtracted by blockade with 5 mM CoCl<sub>2</sub>. Ca<sup>2+</sup> charge entry was determined from the integral of the current (after leak subtraction) as described by Wanaverbecq et al. (2003).

**G-protein-gated inward rectifier potassium currents.** GIRK currents were recorded from cells preinjected with Kir3.1/3.2 and eGFP cDNA plasmids and identified by eGFP fluorescence, as described by Fernandez-Fernandez et al. (1999) but with the differences that perforated-patch recording was used and the external K<sup>+</sup> concentration was 6 mM. Currents were generated by applying 200 ms voltage ramps between –140 and –40 mV every 5 s from a holding potential of –40 mV and recorded using an Axoclamp 2B switch-clamp amplifier in discontinuous mode (sample rate, 6–8 kHz).

**Fluorescence microscopy.** Live neurons were imaged using a Nikon (Tokyo, Japan) Diaphot inverted microscope with a 40 $\times$  oil-immersion lens. Excitation light was provided by a monochromator (Polychrome II; TILL Photonics, Martinsried, Germany) controlled via an Apple Macintosh G4 computer and Openlab imaging software (Improvision, Coventry, UK). Images were captured on a 12-bit grayscale digital camera (C4880-00; Hamamatsu, Shizouka, Japan) and pseudo-color palettes overlaid according to the dye imaged. Images were acquired with minimum delay, according to the exposure time of the camera, but usually more than one image per second.

**Digital deconvolution.** Images obtained with the above equipment are subject to a certain amount of blurring, caused by light “haze” emitted from above and below the plane of focus. To remove this blurring with-

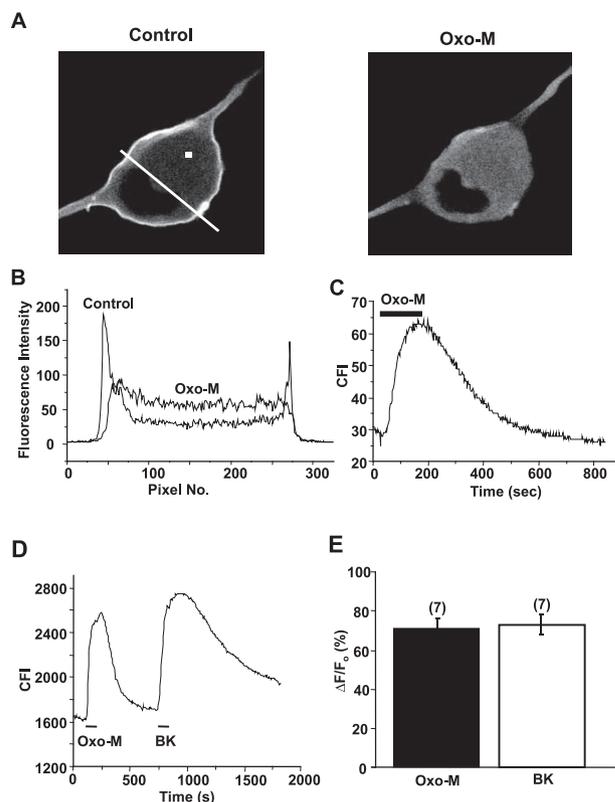
out the use of laser confocal microscopy, digital deconvolution was used. This technique corrects and removes the noise and haze originating from images in other focal planes using mathematical algorithms, the simplest of which is the “nearest neighbor algorithm” (Agard et al., 1989; Castleman, 1996). This algorithm calculates the contribution of light noise from the focal planes above and below the desired slice and subtracts them. An automation was designed in-house, using Openlab software, which automatically focused the microscope to the bottom of the cell, and then programmed it to acquire several optical slices ( $\approx 20$ ) in  $0.5 \mu\text{m}$  steps through the cell. An image from a particular focal plane was corrected using the nearest five-neighbors algorithm (i.e., using five optical slices above and five below the chosen plane). The algorithm calculates the point-spread function of the microscope, which can be estimated once the objective, immersion solution, camera resolution, and binning are known. The images produced by this method were indistinguishable from those obtained from a “conventional” laser-scanning microscope (SP2; Leica, Nussloch, Germany), which essentially (at this resolution) gave the same results.

**Simultaneous electrophysiological and fluorescence recording.** After locating a fluorescent cell, it was then patched at a holding potential of  $-60$  mV using only transmitted light. The light was then switched off while the patched membrane permeabilized, because amphotericin B is light sensitive. When the access resistance was suitably low ( $<15 \text{ M}\Omega$ ), the fluorescent light was switched on and the focus and camera exposure time were adjusted. The membrane potential was then set to the holding potential for M-current activation ( $-20$  to  $-30$  mV). Hyperpolarizing steps to  $-50$  mV to deactivate M currents were initiated by a digital pulse, which was triggered by commencement of the imaging recording, to synchronize the two recordings.

**Ca<sup>2+</sup> imaging.** Cells were incubated with  $2 \mu\text{M}$  fura-2 AM for 30 min at  $37^\circ\text{C}$  and then washed for 20 min before commencing experimentation. Alternate images at 350 and 380 nm excitation wavelengths were recorded at 1 Hz and background subtracted, and the ratio images were calibrated using an algorithm formulated in a separated set of experiments. The equipment was calibrated by cell dialysis in the whole-cell configuration using intracellular solutions from a calcium calibration buffer kit (Molecular Probes, Eugene, OR) containing fura-2 acid ( $100 \mu\text{M}$ ).

**Cell volume measurements.** Cell volume was determined using a Leica SP2 confocal-scanning microscope. A z-stack consisting of a series of  $0.2 \mu\text{m}$  optical slices was taken through cells that had been preloaded with the intracellular fluorescent marker calcein (calcein-AM,  $2 \mu\text{M}$  for 35 min at  $37^\circ\text{C}$ ; excitation/emission wavelengths, 480/530 nm). These two-dimensional slices were then three-dimensionally reconstructed, and the cell surface was rendered using the Imap software package (Bitplane, Zurich, Switzerland). Volumes of the isolated soma were determined by voxel counting of the entire calibrated and reconstructed volume.

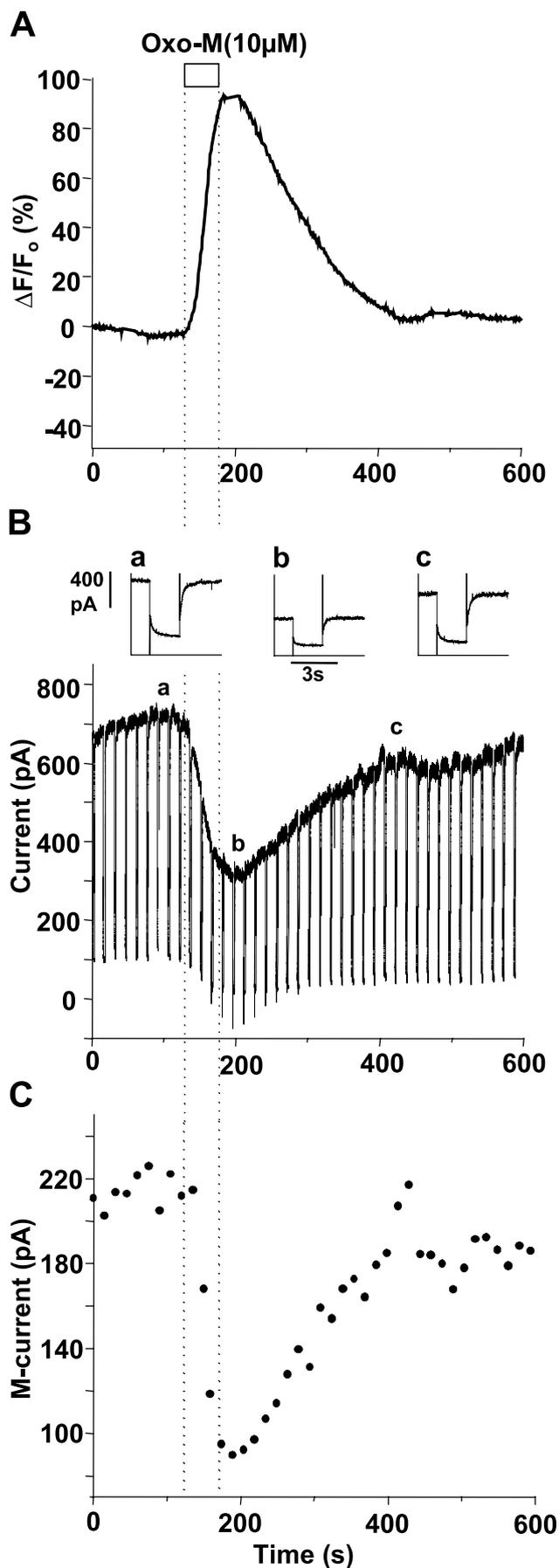
**Free intracellular space.** To determine the free intracellular space available to a small hydrophilic molecule such as inositol-1,4,5-trisphosphate (see Results), we compared the fluorescence intensity of known concentrations of a low-molecular-weight fluorophore [4,4-difluoro-4-bora-3a,4a-diaza-s-indacene fluorophore (BODIPY-FL); molecular weight, 292; Molecular Probes] contained within aqueous vesicles with that of the same concentrations fully dialyzed into patch-clamped sympathetic neurons. Aqueous vesicles in a silicon elastomer (Sylgard 182; Dow Corning, Midland, MI) were fashioned after the methods of Canepari et al. (2001). BODIPY-FL was initially dissolved in intracellular solution and then  $50 \mu\text{l}$  of this solution was rapidly stirred into 1 ml of uncured elastomer. A small aliquot was placed on a microscope slide and covered with a glass coverslip. The microscope slide was then heated on a hotplate at  $60^\circ\text{C}$  for 5 min to set, firmly embedding the vesicles within. The microscope was optimized for recording the fluorescence intensity of the highest concentration of BODIPY-FL ( $100 \mu\text{M}$ ) contained within spherical aqueous vesicles of comparable size to sympathetic neurons ( $19$ – $23 \mu\text{m}$  diameter) by adjusting the excitation intensity ( $480 \text{ nm}$ ) to minimize photobleaching ( $<5\%$  in 10 min) and optimizing the 12-bit image capture by using multiple frame-averaging ( $n = 5$ ) background subtraction and digital deconvolution. Once set, these parameters were used throughout the entire experiment. The dialysis of BODIPY-FL into SCG cells was monitored by taking brief fluorescent images (100 ms) every



**Figure 1.** Agonist-induced translocation of GFP-PLC $\delta$ -PH. **A**, Scanning confocal microscope images (8-bit resolution; i.e., 256 gray scales) of GFP-PLC $\delta$ -PH fluorescence distribution in a sympathetic neuron before and during application of  $10 \mu\text{M}$  Oxo-M. **B**, Plot of fluorescence intensity across a single line scan (**A**, white line) before and during the peak of the Oxo-M response. **C**, Continuous time plot of cytosolic fluorescence intensity recorded in a single region of interest (**A**, white box). **D**, Changes in CFI (12-bit resolution; i.e., 4096 gray scales) in a single neuron produced by successive applications of  $10 \mu\text{M}$  Oxo-M and  $100 \text{ nM}$  BK. **E**, Mean changes in cytosolic fluorescence in seven neurons produced by  $10 \mu\text{M}$  Oxo-M and  $100 \text{ nM}$  BK applied sequentially as in **D**. The change in fluorescence is given as  $\Delta F/F_0 \times 100\%$ , where  $\Delta F = F - F_0$ ,  $F$  is peak fluorescence after agonist application, and  $F_0$  is baseline fluorescence before agonist application.

minute for 15 min, after which a digital deconvolved image was recorded. To minimize errors, we used the same BODIPY-FL-containing intracellular solutions for whole-cell dialysis and aqueous vesicle measurement. Fluorescence intensity was measured using a binary mask generated by image thresholding. This allows an average intensity within boundaries of both the dialyzed cells and aqueous vesicles to be measured. The free intracellular space was determined by subjecting the raw data to linear regression and calculating the ratio of the slopes of the lines generated (see Fig. 8A).

**Immunocytochemistry.** Cells for immunocytochemistry were cultured on glass coverslips in four-well plates (Nunc; Nunc, Roskilde, Denmark). Cells were fixed with 0.2% glutaraldehyde/2% paraformaldehyde (in PBS; Sigma-Aldrich) for 20 min at room temperature. Fixed cells were then permeabilized with 0.1% Triton X-100 (Sigma-Aldrich) in PBS for 15 min. After washing the cells several times in PBS, they were incubated for 1 h in a blocking buffer consisting of  $10 \text{ mg}\cdot\text{ml}^{-1}$  BSA (Sigma-Aldrich) in PBS and then for 1 h in the primary antibody, rabbit anti-NCS-1 (1:200). A secondary antibody (swine anti-rabbit IgG conjugated with tetramethylrhodamine isothiocyanate, 1:200) was then used for visualization. After several washes with PBS, the coverslips were removed from the four-well plates and mounted onto slides using a mounting medium, where they were sealed using clear nail varnish. Slides were refrigerated until examination. The level of auto-fluorescence was estimated by performing the above immunocytochemistry procedures in the absence of primary and secondary antibodies. Nonspecific binding was determined in the absence of the primary antibody.



*Data.* Results are expressed as means  $\pm$  SEM (95% confidence limits). Significance levels were determined by one- or two-tailed *t* tests as appropriate, and differences were considered real at  $p < 0.05$ .

*Drugs and chemicals.* Reagents were obtained from BDH Chemicals unless otherwise stated. Oxotremorine-M (Oxo-M), bradykinin (triacetate salt), IP<sub>3</sub> (D-myo-inositol 1,4,5-trisphosphate hexasodium salt), 1-(6-((17 $\beta$ -3-methoxyester-1,3,5(10)-trien-17-yl)amino)hexyl)-1H-pyrrole-2,5-dione (U73122), and amphotericin B were all obtained from Sigma-Aldrich. Calcein-AM, fura-2 AM, and BODIPY FL were obtained from Molecular Probes. The primary anti-NCS-1 antibody used in the immunocytochemistry was kindly provided by J. Weiss; the secondary antibody was obtained from Santa Cruz Biotechnology (Santa Cruz, CA).

## Results

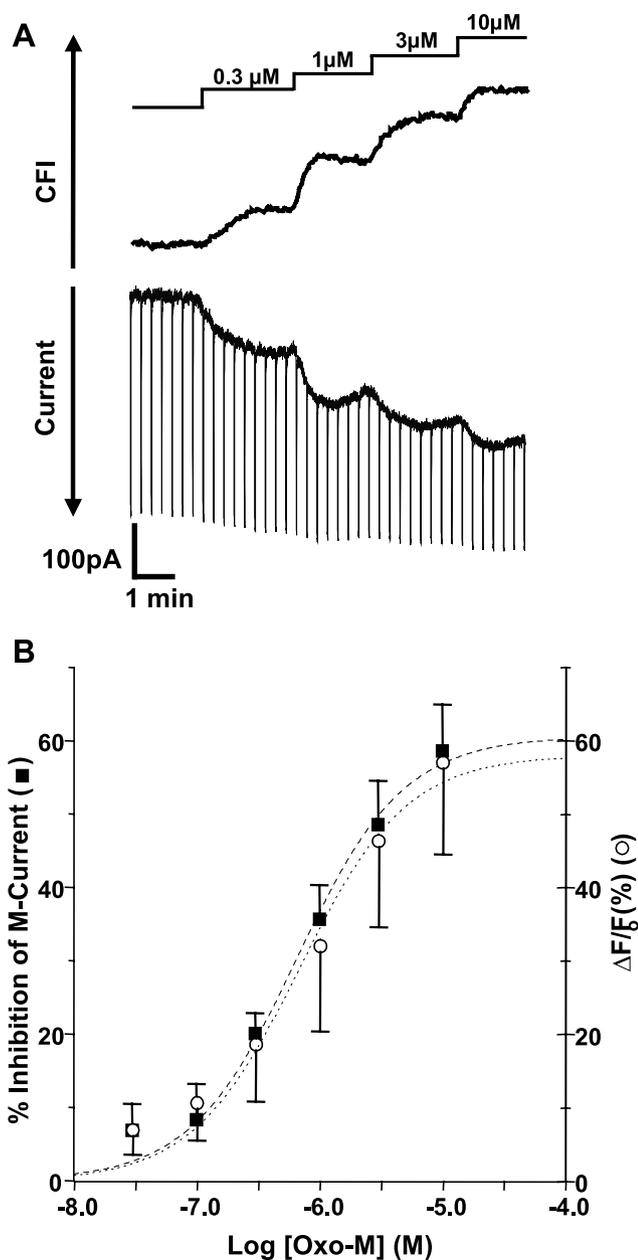
### The response to muscarinic receptor stimulation

In the absence of receptor stimulation, the expressed GFP-PLC $\delta$ -PH construct was highly localized to the plasma membrane (Fig. 1A, control). The addition of the mAChR agonist Oxo-M (10  $\mu$ M) to the bathing fluid produced a rapid and substantial translocation of GFP-PLC $\delta$ -PH into the cytosol. Thus, a confocal line scan showed a clear reduction in membrane fluorescence as well as an increase in cytosolic fluorescence (Fig. 1B). However, because of cell movement, line scans could not be routinely used to quantitate translocation; instead, we did this in the first instance by monitoring the time-dependent increase of fluorescence within a defined region of the cytoplasm, as shown in Fig. 1C.

Figure 2 shows that, when simultaneously recorded, the time courses of the changes in cytosolic fluorescence (Fig. 2A), resting membrane current (Fig. 2B), M-current amplitude (Fig. 2C, deduced from deactivation tail-currents, B), and their recoveries after washout were closely matched. Likewise, the concentration dependence of the cytosolic fluorescence increase mirrored that for membrane current reduction when recorded simultaneously (Fig. 3A), and the mean increase in cytosolic fluorescence ( $\Delta F/F_0$ ) coincided very closely with the mean inhibition of the M current recorded separately (Fig. 3B). Thus, there is a good concordance between PIP<sub>2</sub> hydrolysis, as monitored by GFP-PLC $\delta$ -PH translocation, and M-current inhibition for this agonist.

As reported previously (Suh and Hille, 2002), the aminosteroid phospholipase C inhibitor U73122 (3  $\mu$ M) produced a slow inhibition of the M current and substantially reduced the inhibitory action of 10  $\mu$ M oxotremorine-M on the residual current, from  $66 \pm 2.5\%$  ( $n = 6$ ) to  $23.5 \pm 4.2\%$  ( $n = 12$ ). U73122 did not change the resting cytosolic fluorescence intensity (CFI) in cells expressing GFP-PLC $\delta$ -PH but very strongly reduced the increment in fluorescence after application of 10  $\mu$ M oxotremorine-M, from  $72.2 \pm 4.2\%$  ( $n = 9$ ) to  $5.9 \pm 1.9\%$  ( $n = 7$ ). This reinforces the view that the increased fluorescence induced by oxotremo-

**Figure 2.** Simultaneous recording of changes in cytosolic fluorescence (A), membrane current (B), and M-current amplitude (C) produced by oxotremorine-M in a sympathetic neuron. Membrane current was recorded using an amphotericin perforated-patch electrode from a neuron expressing GFP-PLC $\delta$ -PH. Cytosolic fluorescence was monitored simultaneously from a defined cytoplasmic region of interest as in Figure 1. Oxo-M (10  $\mu$ M) was added to the bathing fluid for 60 s, indicated by the open bar and vertical dashed lines in A–C. A, Percentage change in cytosolic fluorescence from baseline ( $\Delta F/F_0 \times 100\%$ ). B, Membrane current (in picoamperes) recorded at  $-20$  mV. Downward deflections show current transients in response to hyperpolarizing steps to  $-50$  mV. Currents recorded at times a–c are shown on a faster time base above. C, Amplitude of M-current deactivation tails (in picoamperes) measured from the initial amplitudes of the deactivation currents illustrated in B plotted against time. Note that oxotremorine-M produced a large increase in fluorescence coincident with the reduction of outward current and M-current deactivation tails.



**Figure 3.** Concentration dependence of oxotremorine-M-induced increases in cytosolic fluorescence and M-current inhibition. **A**, Simultaneous recording of CFI (arbitrary units) and membrane current (in picoamperes) during step-wise increments in the concentration of oxotremorine-M added to the bath-perfusion fluid. **B**, Mean concentration–response curves for percentage of inhibition of M current (filled squares and dashed line) and percentage increase in cytosolic fluorescence ( $\Delta F/F_0 \times 100\%$ ; open circles and dotted line) produced by increasing concentrations of oxotremorine-M (in log-molar units). Oxotremorine-M was added cumulatively as in **A**. Each point is the mean from measurements in 5–13 neurons; bars show unidirectional SEMs. Curves are drawn according to the equation  $y = y_{\max} \times x/(x + K)$ . Values for  $y_{\max}$  and  $K$  were (mean  $\pm$  SEM) as follows: M-current inhibition,  $61.0 \pm 2.4\%$ ,  $0.68 \pm 0.13 \mu\text{M}$ ; fluorescence,  $58.2 \pm 3.3\%$ ,  $0.68 \pm 0.94 \mu\text{M}$ .

rine-M results from activation of phospholipase C and consequent hydrolysis of PIP<sub>2</sub>.

It might be expected that the PLC $\delta$ -PH peptide could itself reduce M current (by sequestering PIP<sub>2</sub>) and/or reduce the inhibition of M current by agonist (by competing with PLC), as reported previously for thyrotropin-releasing hormone-induced inhibition of GIRK currents (Lei et al., 2001). No significant reduction of M current or oxotremorine-M-induced inhibition

was apparent 1 d after GFP-PLC $\delta$ -PH cDNA injection (as used for the experiments described above). However, M current was substantially and significantly reduced after 3 d of expression when compared with control neurons cultured for 3 d simultaneously, from  $526 \pm 76$  to  $198 \pm 16$  pA ( $n = 7$  for each).

### The response to bradykinin

Bradykinin (10–100 nM) also induced translocation of the GFP-PLC $\delta$ -PH construct, similar to that produced by 10  $\mu\text{M}$  oxotremorine-M (Fig. 1*D,E*). However, the effect of bradykinin differed in two respects from that of oxotremorine-M. First, recovery was much slower. Second, the effect showed strong desensitization, such that it was not possible to obtain two equivalent responses from the same cell. This accords with the slow recovery from, and desensitization of, M-current inhibition by bradykinin (Jones et al., 1995) and unfortunately precluded construction of accurate concentration–response curves.

### PI5-K overexpression reduces the effects of both oxotremorine-M and bradykinin

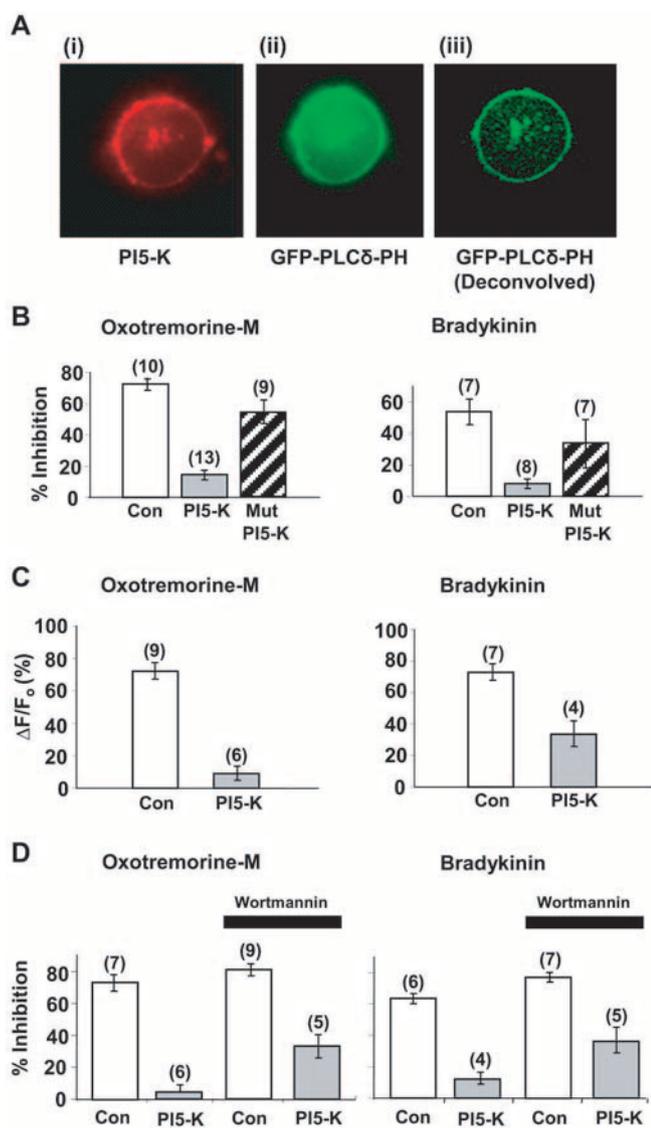
Individual neurons were injected intranuclearly with plasmids encoding GFP-tagged versions of either the wild-type PI5-K or a mutant form PI5-K(K179M) devoid of kinase activity (Davis et al., 1997). In both cases, substantial fluorescence in association with the outer membrane was apparent after 2 d of expression but with additional patches of cytoplasmic fluorescence (perhaps in association with the Golgi/endoplasmic reticulum) (Fig. 4*A*).

M-current amplitudes (measured from deactivation tail-currents) (see Materials and Methods) in GFP-PI5-K-expressing cells and mutant PI5-K(K179M)-expressing cells were not significantly different from those in control cells. Mean amplitudes (in picoamperes)  $\pm$  SEM were as follows: controls,  $215 \pm 16$  ( $n = 15$ ); PI5-K,  $236 \pm 27$  ( $n = 19$ ); PI5-K(K179M),  $223 \pm 35$  ( $n = 11$ ). Mean deactivation time constants also were not significantly different between the three groups, but time constants varied considerably in PI5-K-expressing neurons and, in some cells, were noticeably slow. This might suggest a PIP<sub>2</sub>-dependent effect on gating, as previously noted for KCNQ1/KCNE1 channels by Loussouarn et al. (2003). Because of the variability, however, we could not explore this in more detail.

M-current inhibition by both oxotremorine-M and bradykinin was strikingly reduced in wild-type PI5-K overexpressing neurons by  $\sim 80$  and  $85\%$ , respectively (Fig. 4*B*). This effect was not observed in cells overexpressing the kinase-deficient mutant PI5-K(K179M). It should be noted that the concentrations of oxotremorine-M and bradykinin used were maximally effective in normal neurons (Fig. 3) (Jones et al., 1995).

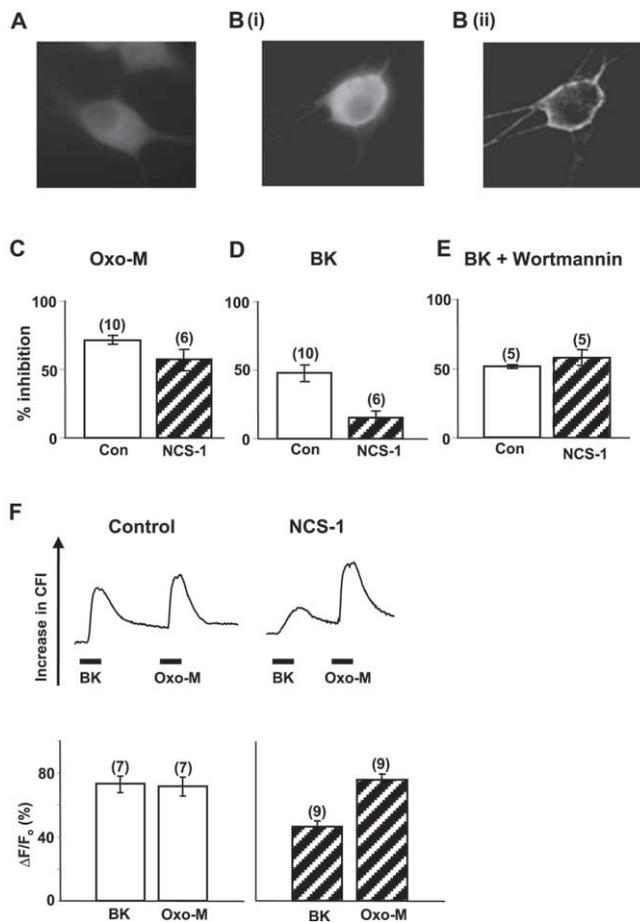
To assess the effect of PI5-K overexpression on GFP-PLC $\delta$ -PH translocation, we replaced the wild-type GFP-tagged PI5-K construct with a construct encoding the red fluorescent protein, pDSRed, and then recorded responses from cells that expressed both red and green fluorescence (Fig. 4*A*). The elevation of cytosolic GFP-PLC $\delta$ -PH fluorescence produced by both oxotremorine-M and bradykinin was strongly reduced by  $\sim 88$  and  $55\%$ , respectively (Fig. 4*C*). We interpret this to result from the apparent increased level of PIP<sub>2</sub> caused by increased PI5-K activity and consequent competitive inhibition of cytosolic IP<sub>3</sub> binding to GFP-PLC $\delta$ -PH (see below).

To assess whether IP<sub>3</sub> production was modified by PI5-K overexpression, we recorded the elevation of intracellular Ca<sup>2+</sup> produced by bradykinin using fura-2 AM. [Oxotremorine-M does not produce a detectable Ca<sup>2+</sup> signal in these cells, but bradykinin produces a small rise (Cruzblanca et al., 1998; Delmas



**Figure 4.** Overexpression of PI5-K reduces M-current inhibition and GFP-PLCδ-PH translocation induced by both oxotremorine-M and bradykinin and is partly reversed by inhibiting phosphatidylinositol-4-kinase with wortmannin. **A**, Sympathetic neuron expressing both a red-tagged (pSRED) PI5-K (*i*) and GFP-PLCδ-PH (*ii*, *iii*). **iii** shows a deconvolved image of that in *ii* (see Materials and Methods). **B**, Mean inhibition of the M current (%) in control (Con), PI5-K, and mutant (Mut) PI5-K overexpressing neurons by 10 μM oxotremorine-M and 100 nM bradykinin. The mutant PI5-K was devoid of kinase activity. M-current amplitudes were measured from deactivation tail currents. **C**, Mean rise in cytosolic fluorescence intensity ( $\Delta F/F_0$ ; %) in GFP-PLCδ-PH-expressing cells produced by 10 μM oxotremorine-M or 100 nM bradykinin in control and PI5-K overexpressing cells. **D**, Mean percentage M-current inhibition in control (noninjected) and PI5-K overexpressing neurons in the absence or presence of 15 μM wortmannin (added 10–15 min before agonist). Error bars indicate SEM of the number of cells indicated.

et al., 2002.) To avoid desensitization, we used brief pressure applications of 100 nM bradykinin. Ca<sup>2+</sup> signals, although highly variable, were not clearly altered in PI5-K overexpressing cells (controls, 221 ± 120 nM, *n* = 5; PI5-K, 198 ± 108 nM, *n* = 6). This suggests that the total amount of IP<sub>3</sub> production remained approximately the same, despite the apparent elevation of the initial membrane PIP<sub>2</sub> concentration (see below), although we cannot exclude the possibility that the release mechanism might have been partly desensitized at the (presumed) higher resting IP<sub>3</sub> concentration.



**Figure 5.** The effects of NCS-1 overexpression in sympathetic neurons. **A**, **B**, Immunocytochemistry of NCS-1 antibody staining in a noninjected neuron (**A**) and a neuron preinjected intranuclearly (**B**) 1 d previously with NCS-1 cDNA. NCS-1 is endogenously expressed, but staining is enhanced in the NCS-1 cDNA-injected neuron. **Bii** is a deconvolved image of that in **Bi**, showing that most of the staining is associated with the plasma membrane. **C**, **D**, Overexpression of NCS-1 reduces M-current inhibition by 100 nM BK (**D**) but not that produced by 10 μM Oxotremorine-M (**C**). **E**, Addition of 20 μM wortmannin (15 min before) reversed the effect of NCS-1 on BK-induced inhibition. Bars show percentage inhibition of M-current measured from deactivation tails. **F**, NCS-1 reduces bradykinin-induced translocation of GFP-PLCδ-PH. Upper records show representative traces of increases in CFI (arbitrary units) in a noninjected neuron (control) and an NCS-1-expressing neuron produced sequentially by BK (100 nM) and Oxotremorine-M (10 μM). Lower bar charts show mean values of percentage  $\Delta F/F_0$  produced by 100 nM BK and 10 μM Oxotremorine-M in control and NCS-1 overexpressing neurons. Error bars indicate SEMs of the number of cells indicated.

If the effects of PI5-K overexpression on M-current inhibition result from increased levels of PIP<sub>2</sub>, these should be reduced or reversed by inhibiting the supply of the substrate phosphatidylinositol-4-phosphate (PIP). To do this, we used 15–20 μM wortmannin, which, at such concentrations, is an effective membrane-permeable inhibitor of PI4-kinase (Nakanishi et al., 1995). As reported previously (Suh and Hille, 2002; Ford et al., 2003), wortmannin itself produced a slow “rundown” of the resting M current (by 28.7 ± 4.7%; *n* = 7; after 10 min in 20 μM wortmannin) and greatly slowed recovery from oxotremorine-M-induced inhibition. It also slowed the recovery of the cytosolic GFP-PLCδ-PH signal; this effect was variable, but, in some neurons, several minutes were required for full recovery. (Attempts to test whether it affected recovery from bradykinin-induced inhibition were frustrated by the very variable duration of bradykinin action in the absence of wortmannin.) However, wortmannin (15 μM) did not significantly affect the fractional inhibition of

the residual M current by either oxotremorine-M or bradykinin (Fig. 4D). In contrast, it enhanced the (small) inhibition observed in cells overexpressing PI5-K and, hence, partly reversed the effect of the PI5-K (Fig. 4D, filled bars). This provides evidence that the effects of PI5-K overexpression are indeed a result of the increased synthesis of PIP<sub>2</sub>.

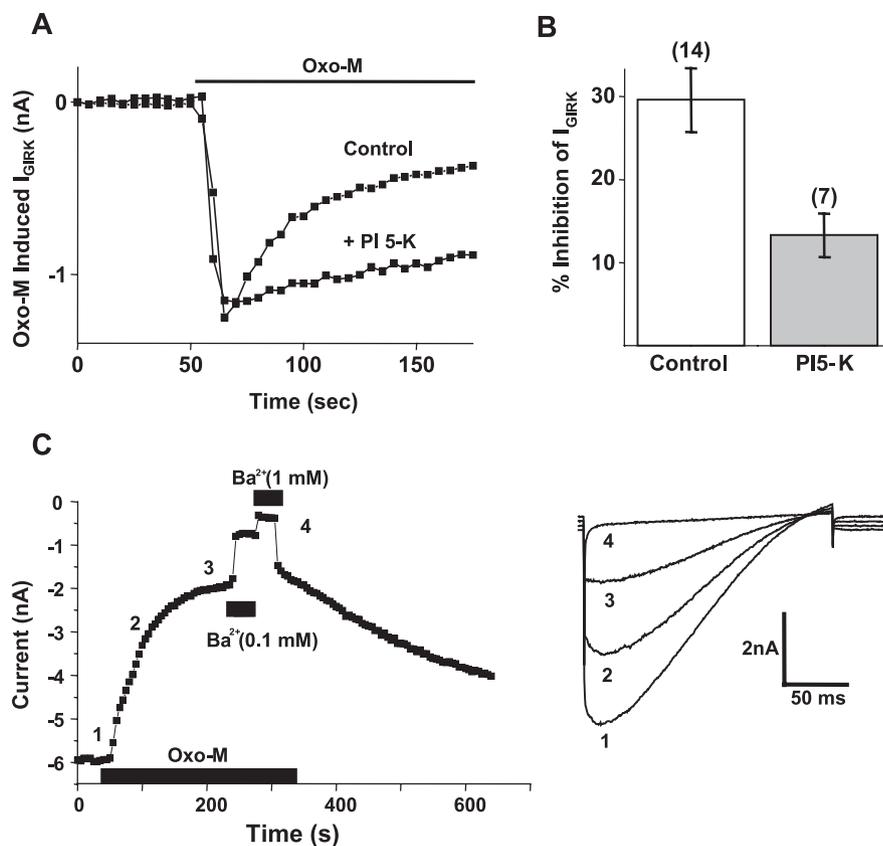
#### NCS-1 overexpression selectively reduces the effects of bradykinin

NCS-1 is a myristoylated protein that enhances the activity of PI4-kinase (see the Introduction). In PC12 cells, overexpression of NCS-1 has been reported to increase cellular levels of PIP and PIP<sub>2</sub> to enhance IP<sub>3</sub> production and Ca<sup>2+</sup> release by UTP and to increase Ca<sup>2+</sup> release by bradykinin (Koizumi et al., 2002; Rajebhosale et al., 2003). In the present experiments, previous intranuclear injection of an NCS-1 cDNA plasmid increased levels of NCS-1 protein in sympathetic neurons as determined by immunofluorescent antibody staining 2 d later (Fig. 5A,B). Deconvolution imaging indicated that much of the increased staining was in, or near, the plasma membrane (Fig. 5Bii).

M-current amplitudes in NCS-1 overexpressing neurons were not significantly different from those in uninjected neurons or in sham (GFP-plasmid)-injected neurons. Mean amplitudes were as follows: controls, 129 ± 27 pA (*n* = 11); NCS-1, 105 ± 12 pA (*n* = 9); GFP, 122 ± 18 pA (*n* = 9). There was also no significant change in the inhibition of M current produced by 10 μM oxotremorine-M (Fig. 5C). However, the inhibition produced by 100 nM bradykinin was reduced from 43 ± 9.0% (*n* = 5) to 14 ± 4.4% (*n* = 6) (Fig. 5D). This effect was reversed by the PI4-K inhibitor wortmannin (20 μM) (Fig. 5E).

NCS-1 overexpression also did not affect the resting membrane:cytosol distribution of GFP-PLCδ-PH. However, as with M-current inhibition, NCS-1 selectively reduced the increase in cytosolic fluorescence produced by bradykinin (Fig. 5F).

A component, at least, of the inhibition of M current by bradykinin appears to be mediated by Ca<sup>2+</sup> release (see the Introduction). Because NCS-1 is a Ca<sup>2+</sup>-binding protein (Schaad et al., 1996), the reduction of bradykinin-induced inhibition in NCS-1 overexpressing neurons in Figure 5D might have resulted from increased buffering of intracellular Ca<sup>2+</sup>. This was difficult to assess directly because of the small amplitude and variability of the bradykinin-induced Ca<sup>2+</sup> signals (see above). Instead, we tested for changes in buffering capacity by comparing the integral of the Ca<sup>2+</sup> current evoked by 200 ms depolarizing voltage steps to 0 from -70 mV with the peak rise in intracellular free [Ca<sup>2+</sup>] recorded using fura-2 as described by Wanaverbecq et al. (2003). From this, the Ca<sup>2+</sup>-binding ratio may be calculated as  $\kappa = \Delta[\text{Ca}^{2+}]_{\text{bound}}/\Delta[\text{Ca}^{2+}]_i = (\Delta[\text{Ca}^{2+}]_{\text{total}} - \Delta[\text{Ca}^{2+}]_i)/\Delta[\text{Ca}^{2+}]_i$ , where  $\Delta[\text{Ca}^{2+}]_i$  is the increase in free cytosolic Ca<sup>2+</sup> measured with

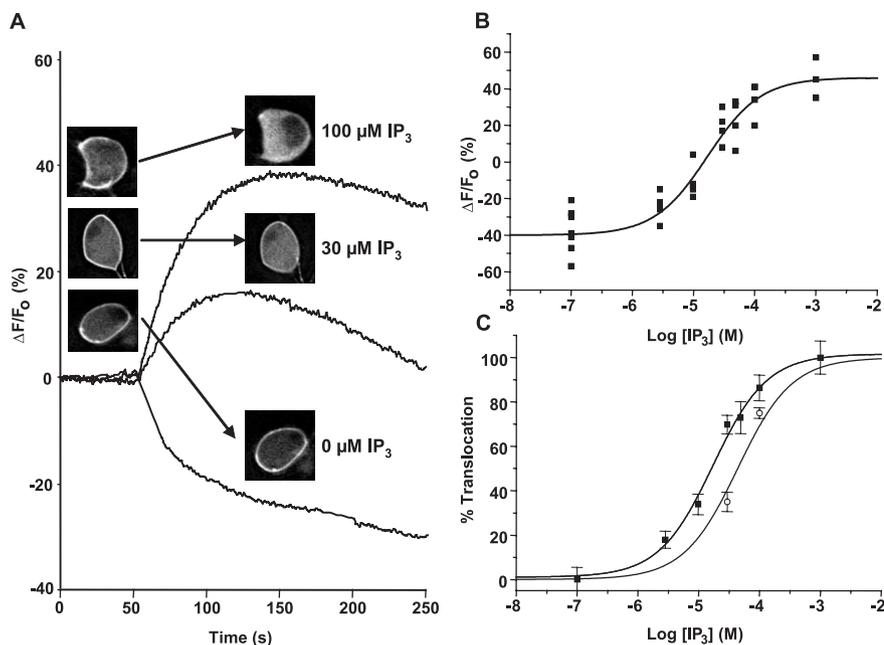


**Figure 6.** Activation and inhibition of GIRK by Oxo-M and reduction of inhibition after overexpression of PI5-K. **A**, Representative time plot of currents activated by 10 μM Oxo-M in individual control and PI5-K-overexpressing neurons. Currents were generated by 200 ms voltage ramps at 5 s intervals from -140 to -40 mV (**C**) and measured at -130 mV. Zero current baseline is that remaining after addition of 1 mM Ba<sup>2+</sup>. Note that Oxo-M produced initial current activation followed by inhibition and that inhibition was slowed in the PI5-K-overexpressing neuron. **B**, Mean ± SEM percentage inhibition of GIRK by 10 μM Oxo-M measured 30 s from the current peak in control neurons and neurons overexpressing PI5-K. Error bars indicate SEMs of the number of cells indicated. **C**, Muscarinic inhibition of GIRK preactivated by coexpression of G-protein βγ subunits. Currents were preactivated by injecting cDNA plasmids encoding Gβ<sub>3</sub> and Gγ<sub>2</sub> subunits 24 h previously. Cells were pretreated with Pertussis toxin (500 ng/ml overnight) to prevent any residual activation of GIRK by oxotremorine-M. The time plot shows current amplitudes recorded at -130 mV from voltage ramps. Records on the right show ramped currents recorded at the corresponding numbered times in the time plot. Oxo-M (10 μM) inhibited the current without previous activation (compare with **A**). Ba<sup>2+</sup> (0.1 and 1 mM) further inhibited the current.

fura-2, and  $\Delta[\text{Ca}^{2+}]_{\text{total}}$  is the total Ca<sup>2+</sup> entry calculated from the clamp current. The baseline for the latter was set using 1 mM Co<sup>2+</sup> to suppress the Ca<sup>2+</sup> current. NCS-1-expressing cells were identified using pDSRed instead of GFP to prevent interference with the fura-2 signals. NCS-1 had no significant effect on the relationship between Ca<sup>2+</sup> entry and fura-2 fluorescence or on the mean calculated binding ratio  $\kappa$  deduced therefrom (control, 650 ± 140, *n* = 6; NCS-1, 830 ± 180, *n* = 7). Also, NCS-1 had no effect on the resting levels of [Ca<sup>2+</sup>]<sub>i</sub> as measured by fura-2 fluorescence (controls, 231 ± 12 nM, *n* = 18; NCS-1, 210 ± 12 nM, *n* = 10; GFP, 224 ± 25 nM, *n* = 8). Thus, the effects of NCS-1 are unlikely to be the result of any appreciable increase in Ca<sup>2+</sup> buffering.

#### Effect of PI5-K overexpression on inhibition of GIRK currents

GIRK currents were generated in these neurons by intranuclear coinjection of plasmids encoding GIRK1 and GIRK2 subunits (Kv3.1 and Kv3.2) along with a marker plasmid encoding GFP, as described previously (Ruiz-Velasco & Ikeda, 1998; Fernandez-Fernandez et al., 1999; Filippov et al., 2004). Currents were generated by voltage ramps (from -140 to -40 mV) and identified



**Figure 7.** Displacement of membrane-associated GFP-PLC $\delta$ -PH by intracellular dialysis with IP<sub>3</sub>. **A**, Records showing the effect of breaking through with patch pipettes containing 0, 30, and 100  $\mu\text{M}$  IP<sub>3</sub> on cytosolic fluorescence in three sympathetic neurons expressing the GFP-PLC $\delta$ -PH construct. Initial fluorescence was measured over a cytoplasmic region of interest (Fig. 1) with the pipette in cell-attached configuration. Records show percentage change in this fluorescence after disrupting the patch ( $\Delta F/F_0 \times 100\%$ , where  $\Delta F = F - F_0$ ,  $F$  is fluorescence after breakthrough, and  $F_0$  is baseline fluorescence before breakthrough). Inset, Images showing the distribution of fluorescence before and after pipette breakthrough. Note that, before breakthrough, fluorescence is localized primarily to the outer membrane but that, at the highest concentration of IP<sub>3</sub>, the fluorescence distributes uniformly with no clear membrane localization. **B**, Concentration dependence of IP<sub>3</sub>-induced change in cytosolic fluorescence  $\Delta F/F_0 \times 100\%$  (**A**), where  $F$  is peak fluorescence after breakthrough. Each point represents a measurement in a separate neuron. **C**, Concentration–response curve for IP<sub>3</sub>-induced GFP-PLC $\delta$ -PH membrane-to-cytosol translocation. The mean fluorescence change in 0 [IP<sub>3</sub>] (–40%) was subtracted from the individual values for  $\Delta F/F_0$  for each finite IP<sub>3</sub> concentration in **B** and scaled to the mean value at 1 mM [IP<sub>3</sub>] (100%). Concentrations were corrected for phosphatase-induced IP<sub>3</sub> breakdown as described in supplemental material 1 (available at [www.jneurosci.org](http://www.jneurosci.org)). Filled symbols indicate means and SEM of the transposed data points in **B**; open symbols indicate means and SEM of equivalent data at 30 and 100  $\mu\text{M}$  [IP<sub>3</sub>] in cells preinjected with a cDNA plasmid encoding PI5-K. Curves are least-squares fits to the equation  $y = y_{\text{max}} \cdot [\text{IP}_3]/(\text{IC}_{50} + [\text{IP}_3])$ . Mean values for  $\text{IC}_{50}$  were as follows: controls,  $16.1 \pm 1.24 \mu\text{M}$ ; +PI5-K,  $42.7 \pm 9.4 \mu\text{M}$ .

from their current–voltage relationship and inhibition by 100  $\mu\text{M}$  to 1 mM Ba<sup>2+</sup> (Filippov et al., 2004) (Fig. 6C).

Oxotremorine-M produced a biphasic effect: generation of an inward current, followed by a decline toward baseline (Fig. 6A, B). We attribute this to an initial activation of the GIRK current via M<sub>2</sub>-mAChRs (Fernandez-Fernandez et al., 1999) with subsequent inhibition via M<sub>1</sub>-mAChRs (Hill and Peralta, 2001). That the latter effect was attributable to inhibition of the M<sub>2</sub>-activated current (rather than desensitization) is confirmed by the fact that oxotremorine-M also inhibited GIRK currents preactivated by stimulating  $\alpha 2$ -adrenoceptors with 10  $\mu\text{M}$  norepinephrine (data not shown) or by expressing G-protein  $\beta_1\gamma_2$ -subunits (Fig. 6C) (Filippov et al., 2004). In the latter case, inhibition was measured in the presence of Pertussis toxin (to prevent M<sub>2</sub>-mAChR activation). Oxotremorine-M produced a concentration-dependent inhibition of the preactivated current of up to 60% of that inhibited by 1 mM Ba<sup>2+</sup>, with an  $\text{IC}_{50}$  value of  $1.16 \pm 0.14 \mu\text{M}$  (pooled data from four cells), quite comparable with that for M-current inhibition (compare with Fig. 3).

In PI5-K overexpressing neurons, the initial (M<sub>2</sub>) activation of the GIRK current noted in Figure 6 was unchanged, but the subsequent (M<sub>1</sub>) inhibitory effect was strongly reduced compared with that in control cells (Fig. 6A). When measured 30 s after the peak of the M<sub>2</sub>-activated current, inhibition was reduced

by ~57%, from  $29.6 \pm 3.3\%$  ( $n = 12$ ) to  $12.8 \pm 2.5\%$  ( $n = 4$ ) (Fig. 6B). We also tested the effect of bradykinin (100 nM), but this agonist failed to inhibit the GIRK current in any of six cells tested.

#### Quantification of membrane PIP<sub>2</sub> levels from GFP-PLC $\delta$ -PH measurements

The PLC $\delta$ -PH peptide binds to both PIP<sub>2</sub> and IP<sub>3</sub> (Garcia et al., 1995; Lemmon et al., 1995). Hence, translocation from membrane to cytosol during PIP<sub>2</sub> hydrolysis reflects (to differing extents) both the reduction of membrane [PIP<sub>2</sub>] and the rise in cytosolic [IP<sub>3</sub>] and indeed has been used as an index of both changes (Stauffer et al., 1998; Varnai and Balla, 1998; Hirose et al., 1999; Van der Wal et al., 2001; Nahorski et al., 2003). Notwithstanding, it is possible (in principle) to estimate individual changes in [PIP<sub>2</sub>] and [IP<sub>3</sub>] from translocation measurements (Xu et al., 2003). Because the inhibition of M current by oxotremorine-M is suggested to result from changes in [PIP<sub>2</sub>] rather than [IP<sub>3</sub>] (see the Introduction), we attempted to use our measurements of GFP-PLC $\delta$ -PH fluorescence to calculate both the resting concentrations of membrane [PIP<sub>2</sub>] and the changes in these concentrations after application of oxotremorine-M. For this, we used a different approach to the “model cell” approach used by Xu et al. (2003), based on (so far as possible) data obtained directly from our experiments on ganglion cells.

#### Resting membrane [PIP<sub>2</sub>] and the effect of PI5-K overexpression

We estimated resting membrane [PIP<sub>2</sub>] ([PIP<sub>2</sub>]<sub>Rest</sub>) using an intracellular IP<sub>3</sub> competition assay. Starting from the situation in which most of the GFP-PLC $\delta$ -PH fluorescence is localized to the plasma membrane (where it is presumably bound to phosphatidylinositides, including PIP<sub>2</sub>), we patched neurons in open-tip configuration with pipettes containing different concentrations of IP<sub>3</sub> and recorded the rise in cytosolic fluorescence (Fig. 7A). To “freeze” PIP<sub>2</sub> concentrations at their prepatch level, we used an EGTA-buffered Ca<sup>2+</sup>-free intracellular solution, because Ca<sup>2+</sup> is required for PLC function (Rhee, 2001).

In the absence of intrapipette IP<sub>3</sub>, cytosolic fluorescence declined on breaking through with the pipette. This may be attributed to washout not only of resting cytosolic IP<sub>3</sub> but also of any other diffusible molecules with polar groups that might bind GFP-PLC $\delta$ -PH, plus any free (unbound) GFP-PLC $\delta$ -PH. [There may also be some increase in the available “free space” (see below) as a result of disruption of cytoplasmic organelles by whole-cell patching.] With increasing concentrations of IP<sub>3</sub> added to the pipette solution, there was a progressively increasing fluorescence above this zero-IP<sub>3</sub> baseline (Fig. 7B). After baseline subtraction and scaling (Fig. 7C), the concentration–response curve yielded a mean  $\text{EC}_{50}$  value of  $16 \pm 1.2 \mu\text{M}$ . We then repeated these experiments in cells overexpress-

ing PI5-K. For these tests, we used two concentrations of IP<sub>3</sub>, 30 and 100 μM. These concentrations clearly produced less translocation than in the control cells, giving an estimated EC<sub>50</sub> value of ~43 μM from the transposed concentration–response curve (Fig. 7C, filled circles).

With a small correction for intracellular IP<sub>3</sub> breakdown (estimated at <5% at 3 μM intrapipette [IP<sub>3</sub>], <1% at >10 μM) (see supplemental material 1, available at www.jneurosci.org), the data in Figure 7C yielded the following values for the IP<sub>3</sub> EC<sub>50</sub> value (means, with lower and upper 95% confidence limits) (in μM): controls, 16.1 (11.9 – 23.5); +PI5-K, 42.7 (25.7 – 70.9). These results are broadly similar to those obtained by Van der Wal et al. (2001) in neuroblastoma cells using flash photolysis of caged IP<sub>3</sub>, in which IP<sub>3</sub> concentrations between 10 and 100 μM were required to produce substantial translocation. The requirements for such high concentrations may seem surprising, because the dissociation equilibrium constant for IP<sub>3</sub> binding to PLCδ-PH has been reported to be ~0.1–0.2 μM, as opposed to ~2 μM for binding to PIP<sub>2</sub> (Lemmon et al., 1995; Hirose et al., 1999). The reason is that, clearly, the resting concentration of PIP<sub>2</sub> (and other potential GFP-PLCδ-PH-binding phosphoinositides) in the membrane must be even higher.

The other principal PLCδ-PH-binding phosphoinositide is phosphatidylinositol-4-phosphate, which has a binding constant of ~7 μM (Garcia et al., 1995) and is present at ~80% of the concentration of PIP<sub>2</sub> in neuroblastoma cell membranes (Willars et al., 1998) and ~60% in human erythrocyte membranes (Ferrell and Huestis, 1984). We ignored phosphatidylinositol (PI) because, even if present in excess over PIP<sub>2</sub> (Willars et al., 1998; Xu et al., 2003) (but see Ferrell and Huestis, 1984), phosphatidylcholine/phosphatidylserine (PC/PS) liposomes doped with 1% PI show very little extra binding of PLCδ-PH over PC/PS liposomes alone [Garcia et al. (1995), their Fig. 2]. We can also ignore other double-phosphorylated and triple-phosphorylated PIs, because, even though these bind PLCδ-PH with comparable affinity to PI(4,5)P (Garcia et al., 1995), their sum concentration is <1% of that of PI(4,5)P<sub>2</sub> (Vanhaesebroeck et al., 2001).

The proportion of GFP-PLCδ-PH bound to IP<sub>3</sub> ( $P_{IP_3}$ ) is then given by the following standard competitive binding equation:

$$P_{IP_3} = C_{IP_3} / (1 + C_{PIP_2} + C_{PIP} + C_{IP_3}), \quad (1)$$

where  $C_{IP_3} = [IP_3]/K_{IP_3}$ ,  $C_{PIP_2} = [PIP_2]/K_{PIP_2}$ , and  $C_{PIP} = [PIP]/K_{PIP}$ , and  $K_{IP_3}$ ,  $K_{PIP_2}$ , and  $K_{PIP}$  are the dissociation equilibrium constants for GFP-PLCδ-PH to IP<sub>3</sub>, PIP<sub>2</sub>, and PIP, respectively. At the EC<sub>50</sub> value for the IP<sub>3</sub>-induced rise in cytosolic fluorescence (when  $P_{IP_3} = 0.5$ ), this simplifies to the following:

$$C_{PIP_2} + C_{PIP} = C_{IP_3} - 1. \quad (2)$$

This was solved for [PIP<sub>2</sub>] by setting  $K_{IP_3} = 0.1 \mu\text{M}$ ,  $K_{PIP_2} = 2 \mu\text{M}$ ,  $K_{PIP} = 7 \mu\text{M}$ , and  $[PIP] = 0.8[PIP_2]$  (Table 1). The EC<sub>50</sub> values for IP<sub>3</sub>-induced GFP-PLCδ-PH displacement in Figure 7 then yielded the following estimates for [PIP<sub>2</sub>] (means with upper and lower 95% confidence limits): controls, 261 μM (192 – 381); +PI5-K, 693 μM (417–1153).

These estimates of the initial (resting) levels of PIP<sub>2</sub> assume that the instillation of IP<sub>3</sub> does not significantly change the level of PIP<sub>2</sub> in the membrane during the time required for the fluorescence measurements. This seems a reasonable assumption, because the slope of the [IP<sub>3</sub>]/translocation curve in Figure 7C was near unity, whereas elevation of membrane PIP<sub>2</sub> would be expected to produce a progressive reduction in translocation and hence flattening of the curve. The absolute

**Table 1. Numerical values used in Equations 1–5 and Figures 8B and 9**

Parameter	Value	Source
$K_{IP_3}$	0.1 μM	Hirose et al. (1999)
$K_{PIP_2}$	2 μM	Hirose et al. (1999)
$K_{PIP}$	7 μM	Garcia et al. (1995)
CMVR	83	Figure 8A; cell radius, 10.1 μM
$[PIP_2]_{Rest}$	261 μM	Figure 7C, Equation 2
$[PIP_2]_{Rest} (+PI5-K)$	693 μM	Figure 7C, Equation 2
$[PIP]_{Rest}$	208 μM	Figure 7C; $[PIP] = 0.8 * [PIP_2]$ (Willars et al., 1998)
$[PIP]_{Rest} (+PI5-K)$	554 μM	Proportionate to $[PIP_2]_{Rest} (+PI5-K)$
$[IP_3]_{Rest}$	0.16 μM	Fink et al. (1999)
$[IP_3]_{Rest} (+PI5-K)$	0.42 μM	Proportionate to $[PIP_2]_{Rest} (+PI5-K)$
Fractional GFP-PLCδ-PH translocation ( $\Delta F/F_0$ )	0.86	Figure 7B ( $\Delta F/F_0 = 0.86$ at maximal $[IP_3]$ )
M-channel availability		
$K_M$	87.2 μM $[PIP_2]$	Zhang et al. (2003)
$n_H$	1.35	Zhang et al. (2003)

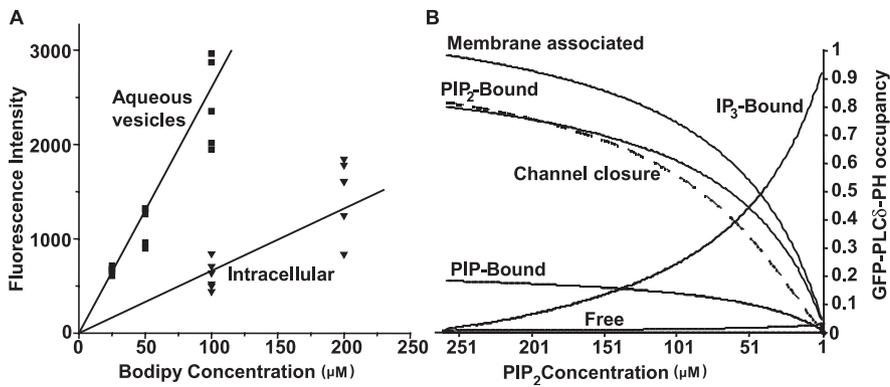
values will also depend critically on the ratio of the binding constants, but the proportionate change on overexpressing PI5-K would be unchanged.

We also used the intracellular IP<sub>3</sub> displacement assay to assess the effect of NCS-1 on membrane [PIP<sub>2</sub>]. For this, we used a single (30 μM) concentration of intracellular IP<sub>3</sub>. Because NCS-1 is a Ca<sup>2+</sup>-binding protein, we measured the IP<sub>3</sub>-induced rise in cytosolic fluorescence using intracellular solutions containing 0 and 200 nM buffered [Ca<sup>2+</sup>]. Mean values (±SEM) for  $\Delta F/F_0$  (%) from the 0 [IP<sub>3</sub>] baseline were as follows: 0 Ca<sup>2+</sup> – NCS-1, 59.8 ± 3.8 ( $n = 5$ ); 0 [Ca<sup>2+</sup>] plus NCS-1, 48.8 ± 1.8 ( $n = 6$ ); 200 nM [Ca<sup>2+</sup>] – NCS-1, 57.3 ± 2.0 ( $n = 6$ ); 200 nM [Ca<sup>2+</sup>] plus NCS-1, 50.3 ± 2.3. None of these values were significantly different from the control values at 30 μM in Fig. 7. Thus, although there might have been some reduction of IP<sub>3</sub> displacement fluorescence in NCS-1 overexpressing cells (which was not dependent on the absence or presence of 200 nM [Ca<sup>2+</sup>]), the effect of NCS-1 was much less than that of PI5-K overexpression (in which the corresponding  $\Delta F/F_0$  at 30 μM [IP<sub>3</sub>] was reduced to 30.1 ± 3.8%). At best, we estimate a maximum increase in resting membrane [PIP<sub>2</sub>] of ~100–150 μM, in line with the data of Koizumi et al. (2002) and Rajebhosale et al. (2003) suggesting rises of 50 and 25%, respectively.

We emphasize that these calculated PIP<sub>2</sub> concentrations are the effective concentrations in the plasma membrane as “seen” by the GFP-PLCδ-PH tag. Assuming they refer only to PIP<sub>2</sub> in the inner (5-nm-thick) leaflet of the membrane, they equate to a density of 783 molecules of PIP<sub>2</sub>/μm<sup>2</sup> membrane area if arranged as a monolayer, rising to 2079/μm<sup>2</sup> after overexpressing PI5-K. These values are substantially less than previous estimates for neuroblastoma cells (4000/μm<sup>2</sup>) (Xu et al., 2003) based on measurements of total membrane phospholipid. In these measurements, PIP<sub>2</sub> comprised 0.43% of total membrane phospholipid, whereas our measurements imply correspondingly lower values of ~0.08%, rising to 0.22% with PI5-K. However, note the following: (1) the calculations of Xu et al. (2003) do not appear to allow for the area occupied by membrane protein, and (2) GFP-PLCδ-PH only bound to the plasma membrane (Fig. 1A) not the intracellular membranes included in biochemical assays.

### Changes in fluorescence after PIP<sub>2</sub> hydrolysis

Stimulation of muscarinic acetylcholine receptors accelerates PIP<sub>2</sub> hydrolysis and produces a fall in membrane [PIP<sub>2</sub>], with an equiva-



**Figure 8.** *A*, Determination of the free cytosolic volume. Graphs show the mean fluorescence intensity (ordinates) after the dispersion of different concentrations of a fluorescent low-molecular-weight compound (BODIPY-FL) in aqueous vesicles (squares) and mean cytosolic intensity of sympathetic neurons whole-cell patched with electrodes containing the same fluorescent compound (triangles) (see Materials and Methods). Mean slopes (fluorescence intensity per micromolar concentration [BODIPY])  $\pm$  SEM were as follows: aqueous vesicles,  $26 \pm 0.9$ ; intracellular,  $6.6 \pm 0.66$ . The ratio of the slopes provides a measure of the free cytosolic space available to BODIPY, from which cytosolic concentration of IP<sub>3</sub> generated by PIP<sub>2</sub> hydrolysis could be estimated (see Results). *B*, Changes in distribution of GFP-PLCδ-PH between membrane and cytosol with progressive hydrolysis of PIP<sub>2</sub>, calculated from Equations 3–7, for a sympathetic neuron with a diameter of 20 μm, an initial membrane PIP<sub>2</sub> concentration of 261 μM, an effective CMVR of 83, and binding constants of 0.1 and 2 μM for IP<sub>3</sub> and PIP<sub>2</sub>, respectively (Table 1). The dashed line shows how the fractional M-channel availability would be expected to change during PIP<sub>2</sub> hydrolysis, as calculated from the data in the study by Zhang et al. (2003).

lent fall in [PIP], presumably through conversion to PIP<sub>2</sub> (Willars et al., 1998). Knowing the initial membrane concentrations of PIP<sub>2</sub> and PIP, one can calculate the amount of IP<sub>3</sub> formed as it is hydrolyzed and its concentration in the cytosol from the cytosolic/membrane volume ratio (CMVR) as follows:

$$[IP_3] = \frac{1.8 \cdot \Delta[PIP_2]}{CMVR} + [IP_3]_{Rest}, \quad (3)$$

where  $\Delta[PIP_2] = [PIP_2]_{Rest} - [PIP_2]$  (the multiplicand 1.8 reflects the additional formation of IP<sub>3</sub> by PIP through conversion to PIP<sub>2</sub>). The proportions of GFP-PLCδ-PH remaining in the membrane bound to PIP<sub>2</sub> and PIP as the hydrolysis proceeds is then given by the following:

$$P_{PIP_2} = \frac{[PIP_2]}{K_{PIP_2} \left[ 1 + \frac{[IP_3]}{K_{IP_3}} + \frac{0.8 \cdot [PIP_2]}{K_{PIP}} \right] + [PIP_2]} \quad (4)$$

and

$$P_{PIP} = \frac{0.8 \cdot [PIP_2]}{K_{IP_3} \left[ 1 + \frac{[PIP_2]}{K_{PIP_2}} + \frac{[IP_3]}{K_{IP_3}} \right] + 0.8 \cdot [PIP_2]} \quad (5)$$

Of the GFP-PLCδ-PH that accumulates in the cytosol, most is bound to IP<sub>3</sub>, but some is free. These proportions are given by the following:

$$P_{IP_3} = \frac{[IP_3]}{K_{IP_3} \left[ 1 + \frac{[PIP_2]}{K_{PIP_2}} + \frac{0.8 \cdot [PIP_2]}{K_{PIP}} \right] + [IP_3]} \quad (6)$$

and

$$P_{Free} = 1 - P_{IP_3} - P_{PIP_2} - P_{PIP}. \quad (7)$$

To solve these equations, we need to determine the following: (1) the cell size and cell volume available to IP<sub>3</sub> to calculate CMVR

(Eq. 3) and (2) the relationship between the measured increase in cytosolic fluorescence ( $\Delta F/F_0$ ) and proportion of total GFP-PLCδ-PH in the cytosol (i.e., the fractional translocation of GFP-PLCδ-PH from membrane to cytosol).

### Cell volume and cytoplasmic:membrane volume ratio

We estimated the total cell volume from three-dimensional reconstructions of calcein-loaded neurons (see Materials and Methods). In a sample of five cells, the mean volume was  $4289 \pm 382$  fl (SEM). This would correspond to a spherical cell of 10.1 μm radius and would yield a CMVR of  $\sim 330$ , assuming a membrane thickness of 10 nm. However, not all of the intracellular space will be available to IP<sub>3</sub>. To estimate the volume of distribution of IP<sub>3</sub>, we compared the mean overall cell fluorescence in neurons patched with pipettes containing a low molecular weight hydrophilic fluorescent marker (BODIPY) with that in equivalent-sized aqueous vesicles (Fig. 8A) (see Materials and Meth-

ods). Intracellular fluorescence was clearly less than that expected from the vesicular fluorescence, suggesting that the volume of distribution of BODIPY was  $\sim 25\%$  of the total intracellular volume. Assuming that the volume of distribution of IP<sub>3</sub> is similar to that of BODIPY, the effective CMVR in Equation 1 is then 83. This implies that, in a normal neuron (with membrane [PIP<sub>2</sub>] of  $\sim 261$  μM and [PIP] of  $\sim 209$  μM; see above), cytosolic [IP<sub>3</sub>] would rise to 5.82 μM from an assumed baseline value of 0.16 μM (Fink et al., 1999) when all PIP<sub>2</sub> and PIP (after conversion to PIP<sub>2</sub>) is hydrolyzed. Again, this is not unreasonable, because this does not allow for any hydrolysis of IP<sub>3</sub>. There have been no direct measurements in sympathetic neurons; however, Fink et al. (1999) estimated a value of  $\sim 2$  μM in neuroblastoma cells after strong stimulation with bradykinin. However, these cells are appreciably larger than sympathetic neurons (giving a larger CMVR), and bradykinin is unlikely to produce complete hydrolysis of PIP<sub>2</sub> (see below).

### Relationship between cytosolic fluorescence increase and membrane-to-cytosol GFP-PLCδ-PH translocation

We assume that the increase in cytosolic fluorescence  $\Delta F/F_0$  produced by saturating concentrations of intracellular IP<sub>3</sub> reflects total membrane-to-cytosol translocation of GFP-PLCδ-PH. This value ( $+86\%$  from the zero-IP<sub>3</sub> baseline) (Fig. 7B) is greater than that following maximum concentrations of oxotremorine-M ( $+60\%$ ) (Fig. 3), suggesting that oxotremorine-M does not produce full GFP-PLCδ-PH translocation. We rechecked this by superimposing a breakthrough with a pipette filled with a saturating (100 μM) concentration of IP<sub>3</sub> at the peak of the response to a single application of 10 μM oxotremorine-M. This produced an additional increase in cytosolic fluorescence from which (after baseline correction) we calculated that oxotremorine-M produced  $77.4 \pm 2.5\%$  ( $n = 5$  cells) of the maximum translocation produced by IP<sub>3</sub> (in reasonable agreement with the difference in maximum values of  $\Delta F/F_0$  for IP<sub>3</sub> and oxotremorine-M recorded separately). Accordingly, we took the  $86\% (\Delta F/F_0)_{max}$  for intra-

cellular IP<sub>3</sub> as representing 100% translocation and divided agonist-induced  $\Delta F/F_0$  by 0.86 to obtain fractional translocation.

### Distribution of the fluorescence signal and calculation of [PIP<sub>2</sub>]

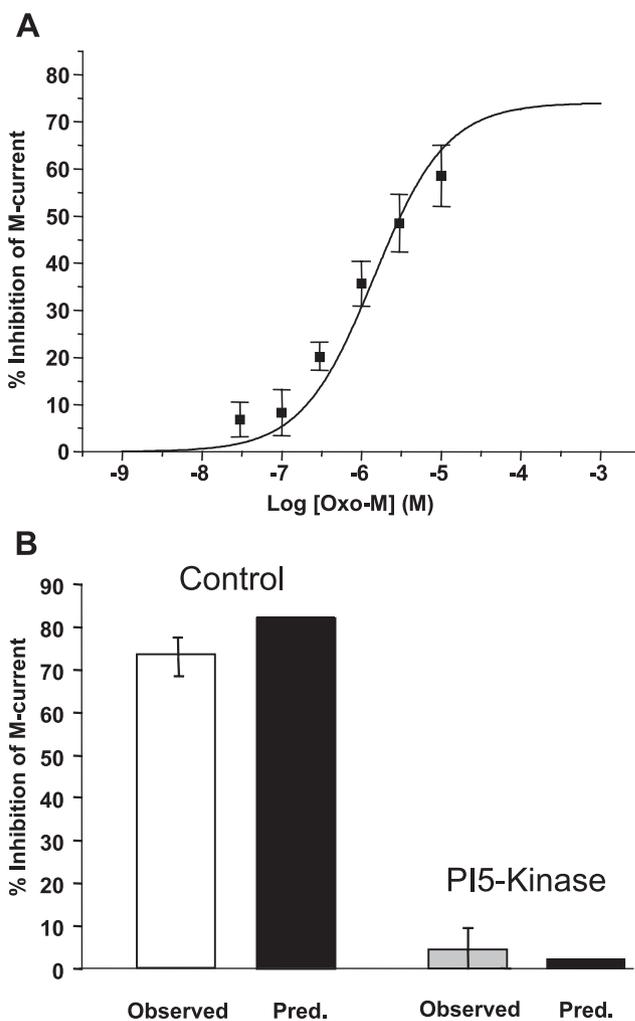
Starting from the estimated mean values for [PIP<sub>2</sub>]<sub>Rest</sub> as deduced from the IP<sub>3</sub> displacement results (261  $\mu$ M in controls and 679  $\mu$ M with PI5-K), we solved Equations 3–7 numerically for diminishing values of [PIP<sub>2</sub>], because PIP<sub>2</sub> is hydrolyzed. Figure 8B shows the results of such calculations for normal cells with 261  $\mu$ M [PIP<sub>2</sub>]<sub>Rest</sub> using numerical values listed in Table 1. From this (and the equivalent readout for PI5-K overexpressing cells), we determined the concentration of [PIP<sub>2</sub>] that corresponded to the fractional translocation calculated from the observed cytosolic fluorescence [ $(\Delta F/F_0)/0.86$ ; see above]. [PIP<sub>2</sub>] can also be solved numerically from an expanded version of Equation 1, if the small fraction of free cytoplasmic GFP-PLC $\delta$ -PH is ignored (see supplemental material 2, available at www.jneurosci.org).

### Relationship between fluorescence change and M-current inhibition

First, we determined how far the close relationship between the increase in cytosolic fluorescence and M-current inhibition produced by increasing concentrations of oxotremorine-M (Fig. 3) might be expected from the “PIP<sub>2</sub>-gating” hypothesis. Taking the values of [PIP<sub>2</sub>] in Figure 8B that corresponded to the fractional GFP-PLC $\delta$ -PH translocation derived from the fluorescence signals in Fig. 3, we calculated the relationship between membrane [PIP<sub>2</sub>] and M channel availability using the experimental data of Zhang et al. (2003) for the activation of KCNQ2/3 channels by DiC<sub>8</sub>-PIP<sub>2</sub>, with an EC<sub>50</sub> value of 87  $\mu$ M and Hill slope of 1.35. This gives a maximum channel availability of 81% at [PIP<sub>2</sub>] of 261  $\mu$ M. Starting here, we calculated the reduction in M-channel availability (and hence percentage inhibition of M current) with increasing concentrations of oxotremorine-M as predicted from the observed fluorescence increase. As shown in Fig. 9A, this provided a reasonable fit to the experimentally observed concentration-inhibition data. The maximum observed fluorescence signal corresponds to an 83% decrease in calculated [PIP<sub>2</sub>], in broad agreement with that (75%) reported by Willars et al. (1998) after strong carbachol stimulation in neuroblastoma cells.

### Effect of PI5-K on M-current inhibition

We also tested how far the fluorescence measurements predicted the changes in M-current inhibition by oxotremorine-M after overexpression of PI5-K (compare Figs. 9B and 4B,C). In these experiments, oxotremorine-M increased cytosolic fluorescence ( $\Delta F/F_0$ ) under control conditions by a mean of +72% (Fig. 4C). With conversion to fractional translocation and taking [PIP<sub>2</sub>]<sub>Rest</sub> of 261  $\mu$ M, this equates (from Fig. 8B) to a 93% decrease in [PIP<sub>2</sub>] and thence (from Zhang et al., 2003) to 86% inhibition of available M current (Fig. 9B, filled bars). PI5-K expression increased apparent [PIP<sub>2</sub>]<sub>Rest</sub> to a mean value of 693  $\mu$ M (Fig. 7, Table 1) but reduced  $\Delta F/F_0$  to +9%. This predicts a much smaller decrease in [PIP<sub>2</sub>] of 21.8% and consequently a much-reduced (2.3%) inhibition of M current (Fig. 9B, filled bars, +PI5-K). Thus, although the values for M-current inhibition calculated from the fluorescence signals are slightly greater in controls and slightly less in PI5-K overexpressing neurons than those observed experimentally (Fig. 9B, open bars and filled bars, respectively), the reduction in M-current inhibition calculated from the change in fluorescence signals matches the observed reduction reasonably well. Interestingly, the fluorescence data also suggests that the con-



**Figure 9.** M-current inhibition calculated from the fluorescence measurements. Concentrations of PIP<sub>2</sub> before and during application of oxotremorine-M were calculated from observed  $\Delta F/F_0$  using the numerical data in Figure 8B and converted to fractional M-channel availability (and thence percentage inhibition) from the KCNQ2/3 channel gating data of Zhang et al. (2003) (see Results). Numerical values used for the calculations are listed in Table 1. **A**, M-current inhibition calculated from the fluorescence data in Figure 3 (smooth curve) superimposed on the mean data points for the observed inhibition in Figure 3. The equation for the curve is  $y = y_{\max} \cdot x^{n_H} / (x^{n_H} + K^{n_H})$ , where  $x = \log [\text{Oxo-M}]$  (M),  $K = -5.8 \pm 0.1$  (SEM; mean  $1.6 \mu$ M [Oxo-M]),  $y_{\max} = 74.0 \pm 6.1\%$  inhibition, and  $n_H = 0.96 \pm 0.13$ . **B**, Solid bars, Predicted (Pred.) M-current inhibition produced by 10  $\mu$ M oxotremorine-M in control and PI5-K overexpressing neurons calculated from the fluorescence data in Figure 4C. Open and shaded bars are the observed percentage inhibition in the experiments shown in Figure 4D. Error bars indicate SEMs of observed data ( $n$  values are as in Figs. 3 and 4).

centration of intracellular IP<sub>3</sub> attained after application of oxotremorine-M was, if anything, reduced (from 5.8 to 3.6  $\mu$ M). Although this may seem surprising, it would accord with the absence of any clear increase in the bradykinin-induced Ca<sup>2+</sup> signal after PI5-K overexpression, perhaps implying that PLC-activated PIP<sub>2</sub> hydrolysis induced by high-agonist concentrations is saturated at resting [PIP<sub>2</sub>]. Finally, the apparent increase in resting [PIP<sub>2</sub>] would be expected to increase M-channel availability from 81 to 94%. This would have been undetectable as an increase in resting current in our experiments, because it lies within the scatter of our measurements. Thus, whereas the mean current was  $\sim 10\%$  greater in PI5-K overexpressing neurons, the difference was not significant.

## Discussion

In the present experiments, we were able to record both M-current inhibition and PIP<sub>2</sub> hydrolysis (by fluorescence monitoring of GFP-PLCδ-PH translocation) simultaneously in single sympathetic neurons after stimulation of endogenous G<sub>q/11</sub>-coupled receptors. As a result, we clearly demonstrated that M-current inhibition in these neurons is accompanied by PIP<sub>2</sub> hydrolysis and that (for cholinergic inhibition at least) there is a close correlation between the two events with respect to both time course and concentration dependence. In addition, procedures that modified the extent of PIP<sub>2</sub> hydrolysis, such as inhibition of PLC or overexpression of PI5-K, produced parallel changes in both readouts of receptor activation. This provides strong evidence that M-current inhibition in sympathetic neurons does indeed result from PIP<sub>2</sub> hydrolysis, as previously inferred from other work (see the Introduction).

The following question then arises: does channel closure result directly from the hydrolysis of PIP<sub>2</sub> and hence reduction of PIP<sub>2</sub> M-channel gating or from an effect of a downstream product such as Ca<sup>2+</sup> or DAG?

### Cholinergic inhibition

The present data point strongly toward the upstream “PIP<sub>2</sub>-depletion” hypothesis as the primary mechanism for M-current inhibition by oxotremorine-M for two main reasons. First, overexpression of PI5-K increased apparent resting membrane PIP<sub>2</sub> levels but strongly reduced M-current inhibition. Hence, it is unlikely that inhibition results from the action of a product of PIP<sub>2</sub> hydrolysis. Second, oxotremorine-M produced a quantitatively comparable inhibition of GIRK currents, which are also known to be regulated by PIP<sub>2</sub> (see the Introduction), and this inhibition was equally reduced by PI5-K overexpression.

We used the fluorescence data to calculate the levels of PIP<sub>2</sub> available to the GFP-PLCδ-PH probe and changes therein after oxotremorine-M application and thence, using the experimental data of Zhang et al. (2003) for DiC<sub>8</sub>-PIP<sub>2</sub> activation of KCNQ2/3 channels to assess how far our experimental observations accord with the PIP<sub>2</sub>-gating hypothesis. Although there are a number of uncertainties and limitations to this approach (most of which are considered above), the results are internally consistent (at least, for steady-state conditions) and fairly accurately predict the observed concentration-dependence of oxotremorine-induced M-current inhibition (Fig. 9A). They also predict reasonably well the observed reduction of M-current inhibition on overexpressing PI5-K (Fig. 9B). In the latter case, this appears to result from the fact that the increased PIP<sub>2</sub> levels are not matched by an increased rate of agonist-induced PIP<sub>2</sub> hydrolysis, as determined by the calculated cytosolic IP<sub>3</sub> levels.

We emphasize that these calculations were made solely to determine how far our fluorescence data could be extrapolated to interpret our M-current data. Suh et al. (2004) used a more comprehensive and dynamic “model cell” approach, incorporating PIP<sub>2</sub> channel gating, to analyze the kinetics of mAChR-Gq-mediated inhibition of expressed KCNQ2/3 channels. Apart from the inclusion by Suh et al. (2004) of the various kinetic parameters necessary for time course simulation, one difference is that their model starts from a higher basal density of membrane PIP<sub>2</sub> molecules [5000/μm<sup>2</sup>; closer to that deduced by Xu et al. (2003) from biochemical data]. For PIP<sub>2</sub> gating of M channels, they then assumed a power function of 1.8 with 76% channel availability at resting [PIP<sub>2</sub>]. This provided an excellent fit to their experimental data but implies a much higher apparent dissociation equilibrium constant (~1 mM intramembrane [PIP<sub>2</sub>]) than

that observed experimentally for activation of KCNQ2/3 channels by exogenous DiC<sub>8</sub>-PIP<sub>2</sub> ( $K_M$ , 87 μM;  $n_H$ , 1.35) (Zhang et al., 2003). Our (lower) estimates of membrane [PIP<sub>2</sub>] are more compatible with these experimental data. From this, we reason that the apparent concentration (and density) of PIP<sub>2</sub> as seen by the GFP-PLCδ-PH peptide (and displaceable by intracellular IP<sub>3</sub>), although lower than those deduced from biochemical data, might plausibly reflect the effective concentration as seen by the membrane-subjacent PIP<sub>2</sub>-binding domain of the M channels. Nevertheless, we recognize that the values for [PIP<sub>2</sub>] derived from the IP<sub>3</sub> displacement tests are subject to errors in, for example, assumptions of GFP-PLCδ-PH-binding constants or estimates of CMVR. Thus, if the ratio of binding constants was 10:1 (PIP<sub>2</sub>:IP<sub>3</sub>) rather than 20:1, this would double the calculated [PIP<sub>2</sub>], although this would be offset if we underestimated CMVR by the same factor. Clearly, it would be interesting to assess the IP<sub>3</sub>-displacement method that we used to estimate [PIP<sub>2</sub>] in other cells for which more biochemical data are available, such as the neuroblastoma cells used by Willars et al. (1998) and Xu et al. (2003).

Another difference from the model used by Suh et al. (2004) is that we allowed PIP to fall in parallel with PIP<sub>2</sub> during PIP<sub>2</sub> hydrolysis, similar to the experimental observations of Willars et al. (1998), whereas a much smaller decline of PIP was predicted in the Suh et al. (2004) model. Relaxing this constraint would not have a major effect on our calculations of M-current inhibition, because the amount of GFP-PLCδ-PH bound by PIP is much less than that bound by PIP<sub>2</sub> [and even smaller taking the estimates of relative PIP and PIP<sub>2</sub> concentrations used by Suh et al. (2004)], but it would reduce our estimates of IP<sub>3</sub> production.

### The action of bradykinin

We have less quantitative information regarding the effect of bradykinin, mostly because desensitization limits the scope of experiments. Qualitatively, however, the effects of bradykinin on M currents and on GFP-PLCδ-PH translocation and the changes therein on PI5-K overexpression were similar to those of oxotremorine-M, implying that the primary events were the same. Nevertheless, there were two notable differences between the effects of these two agonists: NCS-1 expression selectively reduced the effects of bradykinin but not those of oxotremorine-M, and bradykinin did not affect GIRK currents.

Both differences might be explained if the release of intracellular Ca<sup>2+</sup> played a key role in the effect of bradykinin (Cruzblanca et al., 1998; Delmas et al., 2002; Gamper and Shapiro, 2003). Thus, NCS-1 is a Ca<sup>2+</sup>-binding protein, and its upregulation of PI4-K activity is enhanced by Ca<sup>2+</sup> (Zhao et al., 2001). Hence, the release of Ca<sup>2+</sup> by bradykinin might accelerate PIP (and subsequently PIP<sub>2</sub>) synthesis in the presence of excess NCS-1 and thereby reduce the depletion of membrane PIP<sub>2</sub> required for effective M-current inhibition (Winks and Marsh, 2003). Although we could not obtain evidence for any substantial changes in [PIP<sub>2</sub>] in NCS-1 overexpressing neurons in the presence of 200 nM intracellular [Ca<sup>2+</sup>], micromolar concentrations are required to enhance PI4-kinase activation (Zhao et al., 2001); such concentrations might well be attained in bradykinin-receptor submembrane “microdomains” (Delmas et al., 2002) after application of bradykinin. An initial increase in PIP<sub>2</sub> synthesis by bradykinin has been noted in neuroblastoma cells (Xu et al., 2003). The concomitant reduction in the fluorescence signal may then be attributed to increased competition with IP<sub>3</sub> for the fluorescent probe by the higher maintained levels of PIP<sub>2</sub>. A similar effect mediated by Ca<sup>2+</sup>-dependent activation of PI4-K by

endogenous NCS-1 may also explain why bradykinin was unable to inhibit the GIRK channels, because (unlike M channels) these are relatively insensitive to intracellular Ca<sup>2+</sup> but bind PIP<sub>2</sub> with a higher apparent affinity than KCNQ channels (EC<sub>50</sub> value for diC<sub>8</sub>-PIP<sub>2</sub>, 17.7 μM) (Rohacs et al., 2003). Gamper et al. (2004) have advanced a similar hypothesis to explain the resistance of PIP<sub>2</sub>-gated Ca<sup>2+</sup> channels to bradykinin and showed that Ca<sup>2+</sup> channels become inhibited by bradykinin when PIP<sub>2</sub> synthesis is reduced by wortmannin or on overexpressing a dominant-negative NCS-1 construct.

How does one explain the reduction in M-current inhibition by bradykinin in PI5-K overexpressing cells on the Ca<sup>2+</sup> hypothesis? A key factor here is that Ca<sup>2+</sup> closes M channels through the intermediation of calmodulin (Gamper and Shapiro, 2003). Because one of the binding sites for calmodulin on the KCNQ2 channel overlaps the putative binding site for PIP<sub>2</sub> [Delmas et al. (2004), their Fig. 4], Ca<sup>2+</sup> might close M channels by reducing their affinity for PIP<sub>2</sub>. For example, an approximately fourfold increase in the dissociation constant (from 87 to 360 μM) without change in [PIP<sub>2</sub>]<sub>Rest</sub> would suffice to produce the ~52% inhibition in Fig. 4B. Because there is no increase (and possibly a decrease) in the Ca<sup>2+</sup> signal in PI5-K overexpressing cells (see Results), at the higher resting PIP<sub>2</sub> concentration, a comparable affinity shift would produce a smaller (~25%) inhibition of M current.

An additional degree of complexity is added by the observation that mAChR-induced activation of protein kinase C enhances the sensitivity of M channels to mAChR-induced inhibition, through phosphorylation of the constituent KCNQ channels by channel-bound A-kinase anchoring protein-attached protein kinase C (Hoshi et al., 2003). This enhancement (amounting to a threefold shift of the concentration–response curve) might also reflect a change in the affinity of PIP<sub>2</sub> for the M channels, perhaps by modifying the binding of calmodulin (which is already partly activated at resting intracellular [Ca<sup>2+</sup>]) (Gamper and Shapiro, 2003), because the phosphorylation sites overlap the second calmodulin binding site (Delmas et al., 2004). If so, modification of PIP<sub>2</sub> binding might provide the unifying common path for the multifarious regulators of M-channel activity. Additional information regarding the nature of the PIP<sub>2</sub>-binding site on M/KCNQ channels and the factors that affect this binding would clearly be helpful.

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