

Table 1: Functional components of neuropathic and chronic pain pathways, key anatomical substrates and their importance

Process and underlying mechanism	Major neurotransmitter/s (target /tissue)	Time of release/ activation	Consequences	Importance/Remarks
<i>Pain signalling/peripheral sensitisation at primary afferent neurons</i>				
Peripheral nociceptor sensitisation (hyperexcitability)	Substance P (receptors on peripheral terminals and NK1 receptors, plasma membrane of cell bodies, dendrites of non-stimulated neurons [19, 22, 98-100])	Early in the development of neuropathic pain	Sensitisation of peripheral terminals, increased firing rate.	This mechanism explains hyperalgesia as consequence of hypersensitisation
Activation of purinoceptors on microglia	Purinergic pathways[19,22]		Induction of neuropathic pain state	
Release of excitatory amino acids (EAA)			Release of TNF- α	

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<p>Cytokine release following tissue injury is released by macrophages and nerve cell</p> <p>Inflammation (active macrophage infiltrate)</p> <p>Activation of phospholipase A₂ (PLA₂) enzyme on cell membranes</p>	<p>Cytokines (Receptors on blood monocytes)</p> <p>TNF-α</p> <p>Release of arachidonic acid from the cell membrane phospholipid</p>	<p>Early, within 24 h of the onset of inflammatory response</p>	<p>Mediates the inflammatory state</p> <p>Activation and release of platelet derived growth factor (PGDF)</p> <p>Increase in prostaglandin concentrations, which in turn increase the production of glutamate</p>	<p>Ectopic hyper-excitability due to increase in nerve cell interaction, resulting in a vicious cycle of inflammation</p> <p>TNF-α is the primary inflammatory mediator involved in certain nerve injuries (e.g. lumbar disc herniation)</p>
PLA ₂ activation triggers two	Prostaglandins (Peripheral nociceptors, PGE ₂)		Sensitisation of peripheral nociceptors,	While IL-6 is the primary chemical mediator in pain, IL-10 is a natural

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<p>competing pathways, i.e., cyclo-oxygenase (COX) and lipo-oxygenase (LOX)</p> <p>Release of interleukins</p>	<p>receptors in smooth muscle)</p> <p>Thromboxane (TXA₂ receptors on platelets)</p> <p>Leukotrienes (receptors on smooth muscle)</p> <p>IL-1β, IL-6, IL-8, IL-10 (peripheral nociceptors)[101]</p>	<p>Within the first few hours of tissue injury</p>	<p>localised pain, hypersensitivity in uninjured tissue</p> <p>Leukotriene-induced platelet activation and constriction of smooth muscle</p> <p>Increased vascular permeability and leukocyte attraction</p> <p>Stimulation of the production of pro-inflammatory mediators such as PGE₂, COX-2, and matrix metallo-proteases (MMP)</p>	<p>anti-inflammatory cytokine. The net inflammatory response is the result from these opposing effects</p>

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<i>Pain processing</i>				
Central sensitisation (spinal cord)	Glutamate (Presynaptic opioid, glutamate receptors) Substance P (Calcium channels-(α_2 - δ)) Protein kinase C (NMDA receptors) and purinoceptors [22, 23, 102]	Unknown	Dynamic mechanical allodynia Punctate mechanical allodynia	Spread of spinal hyper-excitability Expansion of neuronal fields [22,23,102]
Phenotypical switch Nociceptor peptides normally expressed by A δ and C fibres are expressed by large myelinated A β fibres	Calcitonin gene-related peptide, substance P (dorsal horn receptors)	Unknown	Input from mechanoreceptor A fibres is perceived as pain (dynamic and punctuate allodynia)	Increased synaptic transmission, which is considered the most important steps in the development of chronic pain [25]

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Descending dysinhibition	GABA (GABA receptors) Endogenous opioids (μ receptors)	Late manifestation, months to years after neurological insult[103]	Loss of inhibitory synaptic currents	Selective apoptotic loss of GABAergic neurons in superficial dorsal horn of the spinal cord
Functional degeneration of interspinal inhibitory interneurons Decreased supraspinal descending modulation Descending facilitation	Serotonin/norepinephrine, dopamine (α -2, 5-HT receptors at the dorsal horn inhibitory interneurons) Glutamate (glutamate receptors, purinergic receptors [22, 25, 104])	Protracted several weeks after peripheral nerve injury [1,26,104]	Enhanced signal transmission in the DRG	Inhibition or prevention of apoptotic loss leading to functional degeneration could provide disease modifying effect in neuropathic pain Structures in the mesencephalic reticular formation—possibly the nucleus cuneiformis and the periaqueductal gray area are involved in central sensitisation in neuropathic pain [25] Interestingly, advanced functional MRI (fMRI) techniques show that the same brainstem structures are active in humans with allodynia

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<i>Pain perception/plasticity in the brain</i>				
Intense and persistent nociceptive input involving limbic circuitry. Long term down-regulation of dopamine receptors and dopamine production, enhanced glutaminergic transmission from prefrontal cortex to nucleus accumbens. [105]	Dopamine, glutamine	Plasticity onset occurs at a late stage; associated with chronicity of pain	<p>Maintain synaptic plasticity.</p> <p>Develop and maintain inflammatory hyperalgesia</p>	<p>Similar changes occur in the brain, particularly in the cortex and can be measured experimentally and by functional magnetic resonance Imaging or PET.</p> <p>Dramatic alterations in cortical spatial maps can be detected after nerve injury that may contribute to phantom pain[26, 105]</p>

Table 2: Overview of commonly used experimental models of pain in human subjects.

Model	Description	Clinical manifestation	Mechanisms	Limitations /Application
Mechanical stimulation models				
Mechanical stimulation (pinprick, pressure)	Cutaneous stimulation using von Frey filaments, cotton swab, pin-prick or pressure algometers	Allodynia, pin-prick hyperalgesia	Stimulation of nociceptors and mechanoreceptors A δ and C fibres are stimulated	<ul style="list-style-type: none"> Truly noxious stimuli cannot be induced by non-specific cutaneous stimulation Cutaneous techniques do not mimic nociception NSAIDs, systemic ketamine, tramadol show analgesic activity [57,106]
Chemical-, heat- or cold-evoked hyperalgesia				
UVB (ultraviolet B or sunburn)	Hyperalgesia induced by exposing skin area to graded individualised doses of UV B radiation, resulting in dose related erythema	Inflammatory response, allodynia and hyperalgesia Secondary hyperalgesia in erythematous area	Central sensitisation A δ and C fibres are stimulated	<ul style="list-style-type: none"> This model is not sensitive to drugs administered systemically, applied locally or to drug combinations acting via complementary mechanisms of action NSAID activity has been identified, but no analgesia produced by opioids[57,58]
Capsaicin-induced pain	Capsaicin is applied topically, intradermally or intramuscularly Capsaicin exposure leads to acute severe burning pain	Primary or secondary hyperalgesia up to 24 h	Activation of TRPV1 receptor Stimuli mimic the symptoms of hyperalgesia observed in neuropathic pain	<ul style="list-style-type: none"> Hyperalgesia is variable as it depends on capsaicin absorption Opioids, NMDA receptor antagonists, and calcium channel α_2-δ ligands attenuate capsaicin-induced hyperalgesia Limited activity observed with tricyclic antidepressants and cannabinoids More C than Aδ fibres activated Inconsistent results observed during evaluation of drugs with anti-neuropathic pain activity Lamotrigine, desipramine showed no effects, while gabapentin suppressed hyperalgesia[57,58,106]

Mustard oil	<p>Model of acute peripheral sensitisation</p> <p>Topical application for a few minutes leads to burning pain followed by an inflammatory reaction in the exposed area</p>	Secondary hyperalgesia and allodynia in surrounding unaffected area	<p>Activation of cation channel TRP amkyrin type I in noiceptive neurons C fibres thought to mediate burning pain</p> <p>A fibres believed to mediate allodynia to light mechanical stimuli</p>	<ul style="list-style-type: none"> • It has not been widely used in analgesia testing • Limitations similar to those reported with the capsaicin model[57,58,106]
Thermode burn	Hyperalgesia secondary to first degree burn by exposing healthy subjects to a heat stimulus using a contact thermode	Primary hyperalgesia at the site of exposure, secondary hyperalgesia in adjacent tissue	<p>Central sensitisation</p> <p>Aδ and C fibres are stimulated along with co-activation of Aβ fibres</p>	<ul style="list-style-type: none"> • NMDA receptor antagonists attenuate mechanical hyperalgesia, but effects are inconsistent with opioids • Intracellular Na channel blockers, opioid receptor antagonists, and purinergic receptor activators [57,58,106]