

Introduction

 Any consideration of feeding in vertebrates will include detailed discussion of the marginal dentition. Far less attention has been paid to the palatal dentition, although characters of the palatal dentition are used in phylogenetic analysis (early tetrapods, Sigurdsen & Bolt, 2010; Diapsida, Benton, 1985; Evans, 1988; Archosauria, e.g. Sereno, 1991; Lepidosauromorpha, e.g. Evans, 1991; Parareptilia, Tsuji, 2006; Rhynchosauria, Dilkes, 1998; Synapsida, Sidor, 2003; Abdala et al. 2008; Campione & Reisz, 2010; and Choristodera (Evans, 1990; Matsumoto, 2011). There is a general acceptance that an extensive palatal dentition is plesiomorphic for amniotes. However, the evolutionary history of this dentition is poorly understood, and detailed studies of its structure and function in either extant or extinct tetrapods are rare (e.g. Regal, 1966; Kordikova, 2002; Mahler & Kearney, 2006; Diedrich, 42 2010). During feeding, the jaws, tongue, and palate cooperate in food prehension, intra-oral transport, and swallowing, thus changes in the palatal dentition should reflect changes in feeding behaviour and/or changes in the anatomy of the oral soft tissues. Potentially, therefore, a better understanding of the functional morphology of the palatal dentition may provide an additional source of information on the biology of extinct tetrapods. Here we review the main trends in the evolutionary history of the tetrapod palatal dentition and then discuss them in relation to changes in the anatomy of the skull and oral soft tissues.

Material and Methods

 Palatal tooth arrangements were mapped onto phylogenetic trees for the tetrapodomorph *Eusthenopteron,* early tetrapods, and basal Amniota (Ruta et al. 2003; Ruta & Coates, 2007; Snitting, 2008); Synapsida (Sidor, 2001); Parareptilia (Tsuji & Müller, 2009; Tsuji et al. 2012); and Diapsida (DeBraga & Rieppel, 1997; Rieppel & Reisz, 1999; Brusatte et al. 2010; Borsuk −Białynicka & Evans, 2009a; Dilkes & Sues, 2009). The data on palatal tooth arrangement patterns for each taxon were collected from descriptions in the literature or data matrices for phylogenetic analysis. For some synapsids and early diapsids, the palatal tooth arrangement has not been described, and specimens were examined first hand (see Appendix 1-7).

Evolutionary patterns in the palatal dentition of early tetrapods and

amphibians

 Early tetrapods (e.g. *Acanthostega*, Clack, 1994; *Ichthyostega*, Clack, 2012; *Pederpes*, Clack & Finney, 2005; *Crassigyrinus*, Clack, 2012; *Greererpeton*, Smithson, 1982; *Megalocephalus*, Beaumont, 1977) inherited the basic pattern of the palatal dentition (vomer, palatine, and ectopterygoid) from that of ancestral sarcopterygians (e.g. *Eusthenopteron*, Clack, 2012). There was a single lateral palatal tooth row on each side, running parallel to the jaw margin and with teeth of similar size (and/or larger) to those of the marginal dentition. In *Eusthenopteron,* the parasphenoid intervened between the vomers and the pterygoids in the midline, with the latter element expanded posterior to the marginal tooth row. Small teeth were randomly and widely distributed across the parasphenoid and pterygoid, forming a shagreen dentition. Early tetrapods retained shagreen teeth on the pterygoid (e.g. *Ichthyostega*, *Acanthostega*; Fig. 1), with parasphenoid teeth in a more limited area (e.g. *Acanthostega*, Clack, 1994; *Pederpes*, Clack & Finney, 2005*; Greererpeton*, Smithson, 1982; Fig. 1). This primitive arrangement was conserved in many Temnospondyli (e.g. *Phonerpeton*, Dilkes, 1990; *Doleserpeton*, Sigurdsen & Bolt, 2010), Anthracosauria (*Silvanerpeton*, Ruta & Clack, 2006; *Proterogyrinus*, Holmes, 1984; *Pholiderpeton*, Clack, 1987) and Seymouriamorpha (*Seymouria*, Klembara et al., 2005; *Discosauriscus*, Klembara, 1997; *Utegenia*, Laurin, 1996), with a tooth shagreen on all palatal elements but a reduction in the number of large lateral palatal teeth (Fig. 1). However, in temnospondyls enlargement 81 of the interpterygoid vacuity separated the pterygoids with loss of their anterior midline contact (Fig. 1). As a result, the shagreen teeth on the pterygoid became more laterally 83 restricted. In addition, the ventral surface of the interpterygoid vacuity was sometimes covered by a bony plate bearing patches of loosely set denticles (Schoch & Milner, 2000). Many lepospondyls retained the primitive arrangement with a lateral palatal tooth row parallel to the jaw margin, but there is more variation in the presence and/or arrangement of the shagreen teeth on the palate and the parasphenoid (Fig. 1: e.g. *Odonterpeton*; *Tambachia,* Sumida et al. 1998). *Pantylus* (Romer, 1969) had teeth scattered across the palate (various sizes distributed randomly), *Brachydectes* (Wellstead, 1991) possessed 90 longitudinally aligned midline vomerine tooth rows, and some derived taxa (e.g.

Cardiocephalus, Ptyonius, Carroll et al. 1998) had reduced or lost the shagreen teeth

completely (Fig. 1). Further variations are listed in Appendix 1.

 Living lissamphibians (Gymnophiona, Caudata, and Anura) have reduced shagreen teeth, and palatal teeth are usually restricted to the vomer and parasphenoid, although some species also bear teeth on a palatine/pterygopalatine (e.g. the caudates *Siren* and *Necturus*) or maxillopalatine (e.g. the gymnophionan *Dermophis*, Trueb, 1993). Gymnophiona generally have a single lateral vomerine tooth row parallel to the jaw margin (e.g. *Epicrionops,* Nussbaum, 1977) whereas in frogs (Anura) there is more often a transverse tooth row lying parallel, or nearly parallel, to the anterior part of the marginal tooth row (e.g. *Pelobates*, Roček, 1981; the hylid *Triprion*, Trueb, 1993) (see Appendix 1). The pattern in caudates is much more variable and ranges from a transverse anterior vomerine row (e.g. *Ambystoma;* the plethodontid *Desmognathus*, Trueb, 1993), a medial longitudinal row (e.g. the salamandrids *Notophthalmus* and *Taricha*, Trueb, 1993, Duellman & Trueb, 1994), a roughly "T" shaped combination row (e.g. the plethodontids *Pseudotriton* and *Stereochilus*, Regal, 1966, Wake, 1966), an anterior row parallel to the marginal tooth row (e.g. *Necturus*, Trueb, 1993; *Cryptobranchus*, Elwood & Cundall, 1994)(Fig. 2A), or a tooth platform in either the anterior (*Siren*, Trueb, 1993) or posterior part of the mouth in combination with a transverse anterior vomerine row (e.g. the plethodontids *Bolitoglossa* and *Plethodon*, Wake, 1966).

Evolutionary patterns in the palatal dentition of amniotes

111 A dramatic change occurred in the palatal dentition of Diadectomorpha, the sister taxon of the Amniota (e.g. Ruta et al. 2003; Ruta & Coates, 2007). They lost the early tetrapod pattern (a 113 lateral palatal row and median tooth shagreen) and replaced it with an arrangement of longitudinally oriented rows of conical teeth on the anterior palatal elements (e.g. *Diadectes*, Olson, 1947; Berman et al. 1998; *Orobates*, Berman et al. 2004) and/or a transverse posterior row on the pterygoid flange (*Limnoscelis,* Williston, 1911, Berman et al. 2010; *Tseajaia*, Moss, 1972). This palatal morphology would have been inherited by early members of both Synapsida (mammals and stem-mammals) and Reptilia (Parareptilia+Eureptilia) when these 119 two major clades diverged in the Late Carboniferous.

Synapsida

 Recent phylogenetic analyses place either Caseidae or Ophiacodontidae + Varanopidae as the basal synapsid clade (Benson, 2012). In members of the Caseidae (e.g. *Cotylorhynchus*, Reisz & Sues, 2000; *Ennatosaurus*, Maddin et al. 2008) and Varanopidae (*Mesenosaurus*, Reisz & Berman, 2001, detailed information shown in Appendix 2), there were palatal teeth 126 on the vomer, palatine, pterygoid, and, in some cases, the parasphenoid (Caseidae) and ectopterygoid (e.g. *Edaphosaurus*, Modesto, 1995). However, there was a general trend towards simplification and reduction of the longitudinal palatal tooth rows, while retaining the transverse pterygoid flange tooth row, which was usually located posterior to the marginal tooth row (Fig. 3). The vomerine tooth row tended to become narrower as the choanae elongated anteroposteriorly, and it was lost in Sphenacodontidae (e.g. *Dimetrodon*, Case, 1904; *Secodontosaurus*, Reisz et al. 1992; *Tetraceratops*, Laurin & Reisz, 1996). The posterior elongation of the choanae also had the effect of restricting the longitudinal palatine and pterygoid tooth rows to the back of the mouth (Fig. 3). In these non-therapsid synapsids, particularly in the carnivorous *Haptodus* (Laurin, 1993), *Dimetrodon* (Case, 1904)(Fig. 2B), and *Tetraceratops* (Laurin & Reisz, 1996), the pterygoid flange teeth were often larger than those of the longitudinal tooth rows (vomer, palatine, pterygoid). By contrast, the herbivorous *Edaphosaurus* lacked pterygoid flange teeth but developed a large plate of closely packed 139 palatine and pterygoid teeth level with the posterior teeth of the marginal row (Fig. 3). Further reductions occurred within the clade Therapsida (including Biarmosuchia, Dinocephalia, Anomodontia, and Theriodontia). Although some Biarmosuchia and Dinocephalia retained the transverse pterygoid flange tooth row, they lost vomerine teeth (the dinocephalian *Estemmenosuchus* is an exception, King, 1988) (Fig. 3). The longitudinal tooth rows were rearranged into either circular patches (e.g. the biarmosuchian *Lycaenodon*, Sigogneau-Russell,1989 and the dinocephalian *Syodon*, King, 1988), or a predominantly transverse, M-shaped anterior tooth row (e.g. *Biarmosuchus*, Ivakhnenko, 1999, and the dinocephalian *Titanophoneus*, King, 1988). Loss of the palatal dentition occurred independently within Anomodontia (except the basal *Biseridens*, Liu et al. 2009) and Theriodontia (Modesto et al. 1999). In the latter group, a

palatal dentition was retained in Gorgonopsidae and some Therocephalia (Fig. 4). The palatal

dentition of gorgonopsids was similar to that in non-therapsids (e.g. Biarmosuchia), with

- posteriorly located circular tooth patches on the palatine and pterygoids (Fig. 4). The
- presence of a pterygoid flange row varied, even between species (e.g. *Cyonosaurus*, see
- Appendix 2). In Therocephalia, the medial palatal teeth were further restricted to a small area
- well posterior to the marginal tooth row (e.g. *Regisaurus*, Mendrez, 1972; Fourier & Rubidge,
- 2007; *Theriognathus*, Brink, 1956; *Viatkosuchus*, Tatarinov, 1995), or were lost completely
- (e.g. *Bauria*, Kemp, 1982; *Moschorhinus*, Battail & Surkov, 2000). Palatal teeth were absent
- in Cynodontia (the lineage leading to mammals).
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Reptilia (Parareptilia+Eureptilia)

161 In contrast to Synapsida, many basal members of both Parareptilia and Eureptilia retained longitudinal palatal tooth rows, in conjunction with those on the pterygoid flange (Fig. 5–7; Appendix 3–4).

 Parareptilia. Most parareptiles had the same palatal tooth arrangement as diadectidomorphs and basal amniotes, but shagreen teeth were generally absent (the Permian *Macroleter* was an exception, Tsuji, 2006). Several early parareptiles had teeth on the parasphenoid and/or ectopteryoid (e.g. *Millerosaurus,* Carroll, 1988 and *Lanthanosuchus,* Efremov, 1946; *Nyctiphruretus,* Tsuji et al. 2012), but whether as a retention of the primitive condition or a 170 redevelopment is unclear. Most parareptiles retained a tooth row on the pterygoid flange (e.g. *Lanthanosuchus*, Efremov, 1946; *Nycteroleter*, Tverdokhlebova & Ivakhnenko, 1984), although this is absent in Procolophoniodea (including *Procolophon*, Carroll & Lindsay, 1985; Cisneros, 2008; *Barasaurus*, Piveteau, 1955; *Owenetta,* Reisz & Scott, 2002) and *Mesosaurus* (Modesto, 2006). Where present, the orientation of the flange row also varies 175 from clearly transverse (most taxa) to more oblique (\sim 45 \degree to the transverse axis in *Scutosaurus* [Tsuji et al. 2012] and *Pareiasuchus* [Lee et al. 1997]) (Fig. 5). The longitudinal tooth rows are generally straight, but there was some variation within procolophonids. In *Procolophon*, the palatine and pterygoid tooth rows form a "w" shape (Carroll and Lindsay, 1985; Cisneros, 2008); *Owenetta* shows a triangular arrangement composed of vomer,

palatine and pterygoid rows (Fig. 5); and *Bashkyroleter mesensis* had an additional row

running parallel to the marginal dentition (Ivakhnenko, 1997).

 Members of the Permian Bolosauridae (e.g. *Bolosaurus*, *Eudibamus*) generally lacked palatal teeth (Watson, 1954; Berman et al. 2000). This includes *Belebey maximi* and *B. chengi* (Ivakhnenko & Tverdochlebova, 1987; Müller et al. 2008), but pterygoid flange rows were present in *B. vegrandis* (Müller et al. 2008).

 Eureptilia and stem Diapsida. Eureptilia also inherited the primitive amniote pattern of longitudinal and transverse palatal tooth rows, as shown by *Captorhinus* which had teeth on the palatine, pterygoid, and, variably, the parasphenoid (Warren, 1961; Modesto, 1998), but not the ectopterygoid. Warren (1961) recorded sporadic vomerine teeth in *Captorhinus* sp., but other authors recorded them as absent (Fox & Bowman, 1966). Perhaps they were variable like those of the parasphenoid, although *Labidosaurus* had lost both sets (Modesto et al. 2007). Parasphenoid teeth were present in several other stem eureptilian taxa and stem diapsids (e.g. *Paleothyris*, Carroll, 1969; *Petrolacosaurus*, Reisz, 1981; *Orovenator*, Reisz et al. 2011), but ectopterygoid teeth were rare (e.g. *Araeoscelis*, Vaughn, 1955)(Fig. 6). *Claudiosaurus* appears to have been exceptional in replacing the discrete tooth rows with a shagreen of small teeth across all but the ectopterygoid bones (Carroll, 1981)(Fig. 6). The stem diapsid pattern was inherited by members of some descendant clades (e.g. *Youngina*, Gow, 1975) but parasphenoid and ectopterygoid teeth were generally absent. Subsequently, members of the two major crown diapsid clades, Archosauromorpha and 201 Lepidosauromorpha, showed parallel patterns of reduction from the primitive palatal pattern (Fig. 6–7). Basal archosauromophs, like *Protorosaurus* (Late Permian, Seeley, 1887) and *Czatkowiella* (Early Triassic, Borsuk-Białynicka & Evans, 2009a), retained longitudinal tooth rows on the vomer, palatine and pterygoid, but lacked teeth on either the pterygoid flange or parasphenoid (ectopterygoid teeth unknown; Fig. 7). In contrast, Choristodera (if these are archosauromorphs, e.g. Evans, 1988, 1990; Gauthier et al. 1988) generally retained the pterygoid flange row and expanded the longitudinal pterygoid row into a broad tooth battery. Most choristoderes, including the earliest (Middle - Late Jurassic *Cteniogenys*; Evans, 1990),

210 lacked parasphenoid teeth, so their presence in the Early Cretaceous neochoristodere *Ikechosaurus* (Brinkman & Dong, 1993) was probably a reacquisition (Fig. 2C). The broad- snouted Paleocene choristodere *Simoedosaurus* (e.g. Sigogneau-Russell & Russell, 1978) is characterized by shagreen teeth covering the palate, and there may be a relationship between snout width and palatal tooth row width in this group (Matsumoto & Evans, 2015). Members of some early archosauromorph clades (e.g. Rhynchosauridae, Langer & Schultz, 2000; Trilophosauria, Spielmann et al. 2008) independently lost the palatal dentition, possibly in association with the evolution of a specialized marginal dentition, but the primitive arrangement was retained in archosauriform stem taxa (*Tanystropheus* being unusual in having vomerine teeth running parallel to the marginal tooth row [Wild, 1973])(Fig. 7). Most crown-group archosaurs lacked palatal teeth (Dilkes & Sues, 2009), but a 221 longitudinal row persisted on the palatal ramus of the pterygoid in a few taxa, including the early pterosaur *Eudimorphodon* (Wild, 1978), the basal non-avian dinosaur, *Eodromaeus,* and the basal sauropodmorph *Eoraptor* (Martinez et al. 2011; Sereno et al. 2012). Marginal and palatal teeth were both present in the oldest recorded chelonian, the late Triassic aquatic *Odontochelys* (Li et al. 2008), which had longitudinal tooth rows on the 226 vomer, palatine and pterygoid, but not the pterygoid flange. A similar palatal tooth arrangement was present in the terrestrial *Proganochelys* (Gaffney, 1990; Kordikova, 2002)(Fig. 7), but teeth were absent in all known later testudine taxa. Within the aquatic Sauropterygia, Placodontia is exceptional in the possession of plate-like 230 crushing palatal teeth that were larger than those of the marginal dentition (Neenan et al., 2013)(Fig. 6). However, the palatal dentition was lost at an early stage in the Eosauropterygia (e.g. *Nothosaurus*, Albers & Rieppel, 2003; *Simosaurus*, Rieppel, 1994) and Ichthyopterygia (Motani, 1999). A single individual of the basal ichthyosaur *Utatsusaurus hataii* reportedly had teeth on the pterygoid, but some re-examination is needed (Motani, 1999, and personal communication to RM, 2007). In Lepidosauromorpha, the longitudinal rows remained extensive in stem lepidosaurs like the kuehneosaurs and in early rhynchocephalians (e.g. *Gephyrosaurus*, Evans, 1980), but the 238 pterygoid flange row was lost in most taxa (Fig. 6). The palate of early squamates remains unknown but was probably like that of stem-lepidosaurs. Crown rhynchocephalians lost the

 pterygoid teeth but preserved and enlarged the lateral palatine row (e.g. *Palaeopleurosaurus*, Carroll & Wild, 1994; *Priosphenodon*, Apesteguia & Novas, 2003; *Sphenodon*, Jones et al. 242 2012), which was realigned so as to lie parallel to the maxillary tooth row. This arrangement allows the specialized shearing mechanism that characterizes Rhynchocephalia (Jones et al. 244 2012), whereby the teeth of the dentary bite between the maxillary and palatine tooth rows. Squamates only rarely have palatine teeth (e.g. polychrotines, *Lanthanotus*, *Heloderma*) but pterygoid teeth are more common (Mahler & Kearney, 2006; Evans, 2008), usually along the margins of the interpterygoid vacuity (Fig. 6). Without well-preserved early members of major 248 lineages, it is difficult to determine whether palatine teeth were lost multiple times, or have occasionally been regained as has been suggested for the vomerine teeth of the anguid *Ophisaurus apodus* (Evans, 2008)(Fig. 6). In snakes, the small-mouthed scolecophidians, anomochilids, and uropeltids lack any palatal teeth (Cundall & Irish, 2008), but this is likely to be a specialization rather than the primitive condition. 'Primitive' alethinophidian snakes (e.g. cylindrophiids, aniliids, xenopeltids) have a row of teeth on both the palatine and pterygoid, and this arrangement is retained in macrostomatan snakes, where enlarged palatal teeth play an important role in gripping prey as it is drawn into the mouth (Mahler & Kearney, 2006; Cundall & Irish, 2008). Again, the palatine teeth, at least, may have been regained (Cundall & Greene, 2000). The palate is incompletely known in basal fossil snakes like the Cretaceous *Najash* (Zaher et al. 2009) and *Dinilysia* (Zaher & Scanferla, 2012), but the marine simoliophids (e.g. *Haasiophis*, Tchernov et al. 2000) already show the macrostomatan 260 configuration.

Discussion

263 The review presented above highlights the variation in palatal tooth morphology that exists across tetrapods, but also show some clear trends, summarized in Figure 8. The first is a major difference between early tetrapods and Temnospondyli ('amphibians'), on the one 266 hand, and early amniotes on the other. Early amniotes are characterized by a rearrangement of the palatal dentition to produce a series of distinct longitudinal and/or transverse tooth 268 rows. This arrangement was retained in early representatives of both Synapsida and Reptilia, but there followed a similar, but independent, pattern of reduction in both lineages, starting

270 with the teeth on the parasphenoid and ectopterygoid, and then the vomer and/or pterygoid 271 flange. Within synapsids, all remaining palatal teeth were lost in the ancestors of cynodonts, concomitant with the evolution of the secondary palate. However, as most Reptilia have only 273 a primary palate, palatal teeth persisted somewhat longer, especially in parareptiles and early members of both Archosauromorpha and Lepidosauromorpha. Palatal teeth were lost 275 completely in the ancestors of crown-group crocodiles and turtles, and in early non-avian dinosaurs. In contrast, lepidosaurs tended to retain (or regain) at least some palatal teeth, most often on the posterior part of the pterygoid plate. Regain would also help to explain the presence of parasphenoid teeth in some derived members of Choristodera and *Kuehneosaurus*, despite their absence in more primitive members of the same lineages. It seems likely the developmental mechanism for generating palatal teeth was suppressed 281 rather than lost in some lineages, a phenomenon that has been reported for the marginal dentition in, for example, birds and frogs (Harris et al. 2006; Wiens, 2011).

 These trends in the arrangement and subsequent reduction of the palatal dentition raise 284 guestions about the role of palatal teeth generally and of different patterns (e.g. tooth shagreen versus distinct rows) or groups (e.g. transverse pterygoid flange teeth versus longitudinal rows) of palatal teeth. Like the marginal dentition, the palatal dentition would be expected to reflect diet and feeding strategy to some degree, but diet alone is less likely to explain major trends. Palate morphology should also be correlated with structures in the floor of the mouth, notably the tongue, the hyobranchial apparatus, and the pharynx, as well as jaw muscles like the pterygoideus that have palatal attachments, and with other aspects of feeding strategy including skull kinesis and jaw movements.

 Based on studies of living taxa (as referenced below), Figure 9 presents a summary of some major changes that are thought to have occurred in the soft tissues and/or feeding mechanics of major tetrapod groups. Some of these changes may be correlated with changes in the palatal dentition. However, developing functional hypotheses to explain palatal tooth distribution in extinct taxa is complicated by the fact that, with the exception of snakes (which are highly specialized), most living amniotes have either significantly reduced the palatal dentition (lizards, rhynchocephalians) or lost it completely (chelonians, archosaurs, mammals). Moreover, examination of the palatal surface in a bony skull provides an

 incomplete understanding of its original structure, much of which relies on the presence of overlying soft tissues. Thus, for example, an apparently smooth bone surface may have been covered in life by keratinized oral epithelium that was itself ridged or papillate (Fig. 10). One of the major challenges faced by early land animals was food acquisition (e.g. Lauder & Gillis, 1997). Although aquatic animals often rely on suction feeding to ingest prey and transport it through the mouth toward the pharynx (e.g. Lauder & Shaffer, 1993; Deban & Wake, 2000; Iwasaki, 2002), terrestrial animals must move food physically into the mouth, pass it towards the back of the oral cavity (intra-oral transport, e.g. Smith, 1993; Schwenk, 2000a), and finally push into the pharynx prior to swallowing. The palatal dentition, lying between the teeth of the upper jaws, is positioned to assist the tongue and jaws primarily in intra-oral transport. Very small or thin prey may be moved by the tongue alone (due to surface adhesion) but the development of a palatal gripping surface would have made it easier to manipulate (and perhaps subjugate) larger, potentially resistant, food items. The longitudinal palatal rows of adult terrestrial salamanders have also been correlated with the possession of a mobile tongue (Regal, 1966; Wake & Deban, 2000), the two working together to hold and transport food. However, the absence of intrinsic muscles in most amphibian tongues (Schwenk, 2000a) may limit their mobility and power within the oral cavity. A muscular tongue with both extrinsic and intrinsic muscles is found in many amniotes and probably evolved in stem members of that group, followed by keratinization of the epithelial surface (Iwasaki, 2002). This type of tongue is well adapted to work against the roof of the mouth during intra-oral transport and also to help to roll the food into a bolus at the back of the oral cavity (Schwenk, 2000a). It may therefore be significant that the inferred evolution of this type of tongue (stem-amniotes) was coincident with the change in the pattern of palatal teeth into an ordered arrangement of distinct longitudinal rows. In the absence of a muscular pharynx, a muscular tongue is also used to push the food bolus into the entrance of the pharynx, a process known as pharyngeal packing (Schwenk, 2000a). Teeth on the posterior part of the palate (parasphenoid and pterygoid flanges) may originally have been important in holding the food bolus in place at the entrance to the pharynx, but perhaps became less so as food positioning and swallowing became more efficient (e.g. by expansion of posterior lobes

on the tongue, or by kinetic movements of the jaws and palate, Schwenk 2000a).

 Reacquisition of parasphenoid teeth (as in the Late Triassic kuehneosaurs and the neochoristodere *Ikechosaurus*) may therefore indicate a change in skull biomechanics or feeding strategy whereby an extra gripping surface at the entrance to the pharynx was beneficial. In kuehneosaurs, at least, this may have been correlated with a potential for the quadrates (and attached pterygoids) to splay out laterally to increase pharyngeal width (SE unpublished). Moreover, a subsequent increase in size of the pterygoideus muscle in later lineages, parts of which attach to the pterygoid flange, may have resulted in loss of the pterygoid flange tooth row (e.g. King et al. 1989; Maier et al. 1996).

 The dichotomy in the fate of the palatal dentition between archosauromorphs and lepidosauromorphs may, in part, reflect changes in the archosaurian tongue. Both crocodiles and birds, and thus potentially their common archosaurian ancestor, have lost much of the intrinsic tongue musculature (Schwenk, 2000a). Instead of using the tongue for prehension and transport, they mainly use jaw prehension, inertial feeding, and gravity (Schwenk, 2000a). Loss of the palatal dentition would be consistent with this, as would the development of a secondary palate in derived crocodiles. However, some extant archosaurs (birds, crocodiles) and chelonians (e.g. the sea turtles *Dermochelys coriacea, Chelonia mydas*) have keratinized epithelium forming corny papillae and/or rugae on the palate and/or on the tongue (e.g. Shimada et al. 1990; Kobayashi et al. 1998; Iwasaki, 2002) (Fig. 10). These may have a 348 role analogous to that of the original palatal dentition, especially in turtles where a muscular tongue is retained. In some birds, palatal papillae run transversally across the back of the oral cavity, an arrangement similar to that of a pterygoid flange tooth row. Harrison (1964) suggested that this arrangement, which can also occur across the back of the tongue, facilitates positioning of prey prior to swallowing, a role that we also infer for the pterygoid flange and parasphenoid teeth of more primitive amniote taxa. Most lepidosaurs have a mobile muscular tongue with a papillose surface (Schwenk, 2000b). Although many non-iguanian lizards used jaw prehension to bring food into the

mouth, aided by varying levels of kinesis, most lizards still use the tongue for intraoral

transport and pharyngeal packing, with the latter aided in most taxa by enlarged posterior

- lobes on the tongue (chameleons, varanids and some teiids lack these). The retention of
- clusters or lines of teeth on the posterior part of the pterygoid plate, close to the opening of

 the pharynx (Mahler & Kearney, 2006) may help in positioning/restraining the food bolus during packing. Pharyngeal packing is followed by pharyngeal compression, in which external neck muscles (constrictor colli) contract to squeeze the bolus into the muscular esophagus for swallowing (Schwenk, 2000a). However, the bolus needs to be pushed posterior to the main body of the hyoid before compression begins, to ensure it does not move back up into the mouth instead. In derived anguimorphs and snakes, together or independently depending on the phylogenetic hypothesis, the anterior part of the tongue is bifid and slender, with a purely chemosensory role. In *Varanus*, this change in tongue function is compensated for by the adoption of inertial feeding whereby food items are effectively thrown to the back of the mouth (Schwenk, 2000b). Snakes employ a different strategy, using kinetic jaws and, especially in macrostomatans, enlarged palatine and pterygoid teeth, to draw prey to the back of the mouth for swallowing. As noted above, these may be a secondary development, given that both tongue action and inertial feeding are precluded in snakes.

 The fossil record of synapsids is generally good, permitting many stages in the evolution of the mammalian feeding apparatus, such as heterodonty, reduction of the accessory jaw bones, and formation of a bony secondary palate, to be followed. Coincident changes in oral soft anatomy must also have occurred (Fig. 9), although these are more difficult to pinpoint in time. They include formation of a soft tissue secondary palate prior to the bony one (choanal folds), extension of the bony secondary palate by a muscular soft palate to improve the separation of food and air streams, and muscularization of the pharynx so that the food bolus 380 can be formed within the oropharynx rather than in the mouth, and then swallowed rapidly (e.g. Maier et al. 1996; Schwenk, 2000a). This would have reduced the need for parasphenoid or pterygoid flange teeth. The mammalian tongue remained large and muscular, and reduction of the hyoid apparatus gave it greater mobility for intraoral transport, aided by the development of muscular cheeks. Although palatal teeth were lost, many terrestrial mammals (like birds and turtles) have developed transverse palatal rugae to help to 386 grip food. These rugae are generally reduced in aquatic mammals that feed under water (e.g. suction feeders) where a gripping palatal surface is less useful (Werth, 2000), although Beaked Whales are an exception to this, in developing papillose rugosities to hold their slippery prey (Heyning & Mead, 1996).

Conclusions

 Palatal teeth clearly had an important role in holding and manipulating food within the mouth (although they may occasionally have contributed to food reduction), and it is reasonable to conclude that an extensive palatal dentition was correlated with a well-developed mobile tongue (although the obverse is not necessarily true). The more anterior palatal teeth (vomer, palatine, anterior pterygoid) were probably used mainly during intraoral transport, whereas posterior palatal teeth, notably those on the pterygoid flange and parasphenoid, may have had a greater role in positioning and stabilizing the food bolus at the entrance to the pharynx. Subsequent loss/reduction of the palatal dentition in derived members of most major tetrapod 400 lineages was probably linked to anatomical and functional changes that rendered a palatal gripping surface less important or effective. These include 402 1. reduction of the tongue (e.g. archosaurs, varanid lizards). 2. functional replacement of the palatal dentition with palatal or lingual rugosities (e.g. some turtles, mammals), or with keratinized papillae (e.g. birds). 3. skull or jaw adaptations that improved food holding (e.g. cranial kinesis) 4. changes in feeding strategy (e.g. the adoption of inertial feeding, *Varanus*, crocodiles) 5. invasion of the ventral palatal surface by pterygoid musculature 6. development of an extensive hard and soft palate (e.g. mammals). No single factor can be invoked to explain the loss (or reacquisition) of palatal teeth in any 410 one taxon, and many aspects remain poorly understood (e.g. the relationship between 411 skeletal and soft tissue anatomy in the palate; the developmental biology of the palatal dentition). Nonetheless, palatal tooth patterns have the potential to provide additional information on diet and feeding strategy in extinct taxa and would benefit from further more detailed study.

Acknowledgements

We would like to thank Prof. Ryosuke Motani (University of Toronto), and Drs Takuya Konishi

- (Royal Tyrrell Museum of Palaeontology), Juan C. Cisneros (Universidade Federal do Piauí),
- Pavel Skutschas (St. Petersburg State University), Shin-ichi Fujiwara (Nagoya University),

- **Berman DS, Reisz RR, Bolt JR, et al.** (1995) The cranial anatomy and relationships of the
- synapsid *Varanosaurus* (Eupelycosauria: Ophiacodontidae) from the Early Permian of
- Texas and Oklahoma. *Ann Carnegie Mus* **64**, 99–133.
- **Berman DS, Reisz RR, Scott D, et al.** (2000) Early Permian bipedal reptile. *Science* **290**, 969–972.
- **Berman DS, Reisz RR, Scott D** (2010). Redescription of the skull of *Limmoscelis paludis*
- Williston (Diadectomorpha: Limnoscelidae) from the Pennsylvanian of Cañon del cobre,
- northern New Mexico. *New Mexico Mus Nat Hist Sci Bull* **49**, 185–210.
- **Berman DS, Sumida SS, Martens T** (1998) *Diadectes* (Diadectomorpha, Diadectidae) from
- the Early Permian of central Germany, with description of a new species. *Ann Carnegie Mus* **67**, 53–93.
- **Berman DS, Henrici AC, Kissel, RA, et al.** (2004) A new diadectid (Diadectomorpha),
- *Orobates pabsti*, from the Early Permian of Central Germany. *Bull Carnegie Mus Nat Hist* **35**, 1–36.
- **Bonaparte JF, Sues H-D** (2006) A new species of *Clevosaurus* (Lepidosauria:
- Rhynchocephalia) from the Upper Triassic of Rio Grande do Sul, Brazil. *Palaeontol* **49**, 917–923.
- **Boonstra LD** (1965) The skull of *Struthiocephalus kitchingi*. *Ann S Afr Mus* **48**, 251–265.
- **Borsuk−Białynicka M, Evans SE** (2009a) A long−necked archosauromorph from the Early

Triassic of Poland. *Palaeontol Pol* **65**, 203–234.

- **Borsuk−Białynicka M, Evans SE** (2009b) Cranial and mandibular osteology of the Early
- Triassic archosauriform *Osmolskina czatkowicensis* from Poland. *Palaeontol Pol* **65**, 235– 281.
- **Brink AS** (1956) On *Aneugomphius ictidoceps* Broom and Robinson. *Palaeont Afr* **4**, 97–115.
- **Brinkman DB, Dong ZM** (1993) New material of *Ikechosaurus sunailinae* (Reptilia:
- Choristodera) from the Early Cretaceous Langhongdong Formation, Ordos Basin, Inner
- Mongolia, and the interrelationships of the genus. *Can J Earth Sci* **30**, 2153–2162.
- **Broom R** (1904) On two new therocephalian reptiles (*Glanosuchus macrops* and
- *Pristerognathus baini*). *Phil Trans S Afr Phil Soc* **15**, 85–88.
- **Broom R** (1937) A further contribution to our knowledge of the fossil reptiles of the Karroo.
- *Proc Zool Soc B* **1937**, 299–318.
- **Brusatte SL, Nesbitt SJ, Irmis RB, et al**. (2010) The origin and early radiation of dinosaurs. *Earth-Sci Rev* **101**, 68–100.
- **Campione NE, Reisz RR** (2010) *Varanops brevirostris* (Eupelycosauria: Varanopidae) from
- the Lower Permian of Texas, with discussion of varanopid morphology and
- interrelationships *J Vertebr Paleontol* **30**, 724–746.
- **Carroll RL** (1967) Labyrinthodonts from the Joggins Formation. *J Paleontol* **41**, 111–142.
- **Carroll RC** (1969) [A Middle Pennsylvanian captorhinomorph, and the interrelationships of](http://www.jstor.org/stable/1302357)
- [primitive reptiles.](http://www.jstor.org/stable/1302357) *J Paleont* **43**, 151**–**170.
- **Carroll RC** (1981) Plesiosaur ancestors from the Upper Permian of Madagascar. *Phil Trans R Soc B* **293**, 315–383.
- **Carroll RL** (1988) *Vertebrate Paleontology and Evolution*. 698 pp. New York: W.H. Freeman and Company.
- **Carroll RL, Lindsay W** (1985) Cranial anatomy of the primitive reptile *Procolophon*. *Can. J.*
- *Earth Sci* **22**, 1571–1587.
- **Carroll RL, Wild R** (1994) Marine members of the Sphenodontia. In: *In the shadow of the*
- *dinosaurs: early Mesozoic tetrapods*. (eds Fraser NC, Sues H-D), pp 70–83, Cambridge:
- Cambridge University Press.
- **Carroll RL, Bossy KA, Milner AC**, **et al.** (1998) *Handbuch der Paläoherpetologie*, Teil 1,
- *Lepospondyli* (ed. Wellnhofer P). 216 pp. Munich: Verlag Dr Freidrich Pfeil.
- **Case EC** (1904) The osteology of the skull of the pelycosaurian genus, *Dimetrodon*. *J Geol* **12**, 304–311.
- **Cisneros JC** (2008) Taxonomic status of the reptile genus *Procolophon* from the Gondwanan
- Triassic. *Palaeont Afr* **43**, 7–17.
- **Clack JA** (1987) *Pholiderpeton scutigerum* Huxley, an amphibian from the Yorkshire Coal
- Measures. *Phil Trans R Soc B* **318**, 1–107.
- **Clack JA** (1994) *Acanthostega gunnari*, a Devonian tetrapod from Greenland; the snout,
- palate and ventral parts of the braincase, with a discussion of their significance. *Medd*
- *Groen Geosci* **31**, 1–24.
- **Clack JA** (2012) *Gaining Ground: The Origin and Evolution of Tetrapods*, pp. 523.
- Bloomington: Indiana University Press.
- **Clack JA, Finney SM** (2005) *Pederpes finneyae*, an articulated tetrapod from the
- Tournaisian of western Scotland. *J Syst Palaeontol* **2**, 311–346.
- **Cundall D, Greene HW** (2000) Feeding in snakes. In: *Feeding: Form, Function and Evolution*
- *in Tetrapod Vertebrates.* (ed. Schwenk K), pp. 293–333. San Diego and London:
- Academic Press.
- **Cundall D, Irish F** (2008) The snake skull. In: *Biology of the Reptilia, Volume 20, Morphology*
- *H*. (eds Gans C, Gaunt AS, Adler K), pp. 349–692. Ithaca, NY: Society for the Study of Amphibians and Reptiles.
- **DeBraga M, Reisz RR** (1996) *Acleistorhinus*, the oldest known parareptile from the Permian of Oklahoma. *J Vertebr Paleontol* **16**, 384–395.
- **DeBraga M, Rieppel O** (1997) Reptile phylogeny and the interrelationships of turtles. *Zool J Linn Soc* **120**, 281–354.
- **Deban, SM, Wake DW** (2000) Aquatic feeding in salamanders. In: *Feeding: Form, Function*
- *and Evolution in Tetrapod Vertebrates.* (ed. Schwenk K), pp. 65–94. San Diego and
- London: Academic Press.
- **Diedrich CG** (2010) Palaeoecology of *Placodus gigas* (Reptilia) and other placodontids —
- Middle Triassic macroalgae feeders in the Germanic Basin of central Europe— and
- evidence for convergent evolution with Sirenia. *Palaeogeogr, Palaeoclim, Palaeoecol* **285**, 287–306.
- **Dilkes DW** (1990) A new trematopid amphibian (Temnospondyli: Dissorophoidea) from the Lower Permian of Texas. *J Vertebr Paleontol* **10**, 222–243.
- **Dilkes DW** (1998) The Early Triassic rhynchosaur *Mesosuchus browni* and the
- interrelationships of basal archosauromorph reptiles. *Phil Trans R Soc B* **353**, 501–541.
- **Dilkes DW, Sues H-D** (2009) Redescription and phylogenetic relationships of *Doswellia*
- *kaltenbachi* (Diapsida: Archosauriformes) from the Upper Triassic of Virginia. *J Vertebr Paleontol* **29**, 58–79.
- **Downs JP, Daeschler EB, Jenkins FA, Shubin NH** (2008) The cranial endoskeleton of *Tiktaalik roseae*. *Nature* **455**, 925–929.
- **Duellman WE, Trueb L** (1994) Biology of Amphibians. Baltimore: The Johns Hopkins
- University Press. pp. 670.
- **Eberth DA** (1985) The skull of *Sphenacodon ferocior*, and comparisons with other
- sphenacodontines (Reptilia: Pelycosauria). *New Mexico Bur Mines Mineral Resour*
- *Circular* **90**, 1–40.
- **Efremov IA** (1946) O Podklasse Batrachosauria Gruppe Form, Promezhutochnykh Mezhdu
- Zemnovodnymi i Presmykayoushchimisya [On the Subclass Batrachosauria a group of
- forms intermediate between amphibians and reptiles]. *Izvestiya Akademii Nauk SSSR*
- *Otdelenie Biologicheskikh Nauk [Bulletin of the Academy of Science of the USSR Section*
- *of Biological Sciences]* **6**, 615–638.
- **Elwood JRL, Cundall** D (1994) Morphology and behaviour of the feeding apparatus in
- *Cryptobranchus alleganiensis* (Amphibia: Caudata). *J Morph* **220**, 47–70.
- **Erdman S, Cundall D** (1984) The feeding apparatus of the salamander *Amphiuma*
- *tridactylum*: Morphology and behavior. *J Morph* **181**, 175–204.
- **Evans SE** (1980) The skull of a new eosuchian reptile from the Lower Jurassic of South
- Wales. *Zool J Linn Soc* **70**, 203–264.
- **Evans SE** (1988) The early history and relationships of the Diapsida. In: *The Phylogeny and*
- *Classification of the Tetrapoda*, *Vol. 1*, *Amphibians, Reptiles, Birds*. (ed. Benton MJ), pp.
- 221–260. Oxford: Clarendon Press.
- **Evans SE** (1990) The skull of *Cteniogenys*, a choristodere (Reptilia: Archosauromorpha) from
- the Middle Jurassic of Oxfordshire. *Zool J Linn Soc* **99**, 205–237.
- **Evans SE** (1991) A new lizard-like reptile (Diapsida: Lepidosauromorpha) from the Middle
- Jurassic of England. *Zool J Linn Soc* **103**, 391–412.
- **Evans SE** (2008) The skull of lizards and tuatara*. In*: *Biology of the Reptilia, Volume 20*,
- *Morphology H.* (eds Gans C, Gaunt AS, Adler K), pp.1–344. Ithaca, US: Society for the
- Study of Amphibians and Reptiles,.
- **Ewer RF** (1965) The anatomy of the thecodont reptile *Euparkeria capensis* Broom. *Phil Trans R Soc B* **248**, 379–435.
- **Fox RC, Bowman M** (1966) Osteology and relationships of *Captorhinus aguti* (Cope)
- (Reptilia: Captorhinomorpha). *Univ Kans Paleontol Inst* **11**, 1–79.
- **Fourier H, Rubidge BS** (2007) The postcranial skeletal anatomy of the therocephalian
- *Regisaurus* (Therapsida: Regisauridae) and its utilization for biostratigraphic correlation.

Palaeont. afr **42**, 1–16.

- **Gaffney ES** (1990) The comparative osteology of the Triassic turtle *Proganochelys*. *Bull Am Mus Nat Hist* **194**, 1–263.
- **Gauthier JA, Kluge A, Rowe T** (1988) Amniote phylogeny and the importance of fossils.
- Cladistics, **4**,105–209.
- **Germain D** (2010) The Moroccan diplocaulid: the last lepospondyl, the single one on
- Gondwana. *Hist Biol* **22**, 4–39.
- **Gow CE** (1975) The morphology and relationships of *Youngina capensis* Broom and
- *Prolacerta broomi* Parrington. *Palaeont Afr* **18**, 89–131.
- **Gow CE** (1997) A reassessment of *Eunotosaurus africanus* Seeley (Amniota: Parareptilia).
- *Palaeontol Afr* **34**, 33–42.
- **Harris MP, Hasso SM, Ferguson MWJ, Fallon JF** (2006) The development of archosaurian
- first-generation teeth in a chicken mutant. *Curr Biol* **16**, 371–377.
- **Harrison JG** (1964) Tongue. In: *A New Dictionary of Birds*. (ed. Thomason AL), pp.398–399. London: Nelson.
- **Heyning JE, Mead JG** (1996) Suction feeding in beaked whales: morphological and
- observational evidence. *Contr Sci, LACM* **464**, 1-12.
- **Holmes RB** (1984) The Carboniferous amphibian *Proterogyrinus scheelei* Romer, and the
- early evolution of tetrapods. *Philos Trans R Soc Lond B* **306**, 431–527.
- **Holmes RB, Carroll RL, Reisz RR** (1998) The first articulated skeleton of *Dendrerpeton*
- *acadianum* (Temnospondyli, Dendrerpetontidae) from the Lower Pennsylvanian locality of
- Joggins, Nova Scotia, and a review of its relationships. *J Vertebr Paleontol* **18**, 64–79.
- **Holmes RB** (2000) Paleozoic temnospondyls. In: Amphibian Biology, Volume 4,
- Palaeontology (eds Heatwole H, Carroll RL), pp. 1081–1120. Chipping Norton, NSW,
- Australia: Surrey Beatty & Sons.
- **Huene F von** (1956) *Pälaontologie und Phylogenie der Niederen Tetrapoden*, 716 pp. Gustav
- Fischer Verlag, Jena.

Ivakhnenko MF (1997) New Late Permian nycteroleterids from Eastern Europe. *Paleontol J*

31, 552–528

Ivakhnenko MF (1999) *Biarmosuchus* from the Ocher faunal assemblage of Eastern

Europe. *Paleontol J* **33**, 289–296.

- **Ivakhnenko MF, Tverdochlebova GI** (1987) A revision of the Permian bolosauromorphs of
- Eastern Europe. *Paleont Zh* **1987**, 98–106.
- **Iwasaki S** (2002) Evolution of the structure and function of the vertebrate tongue. *J Anat* **201**, 1–13.
- **Jones MEH** (2006) *Skull Evolution and Functional Morphology in* Sphenodon *and Other*
- *Rhynchocephalia (Diapsida:Lepidosauria)*, Ph.D. thesis, University College London, UK. 565 pp.
- **Jones MEH, O'Higgins P, Fagan M, et al.** (2012) Shearing mechanics and the influence of
- a flexible symphysis during oral food processing in *Sphenodon* (Lepidosauria:
- Rhynchocephalia). *Anat Rec* **295**, 1075–1091.
- **Kemp TS** (1982) *Mammal-Like Reptiles and the Origin of Mammals*. pp.363. London and
- New York: Academic Press.
- **Klembara J** (1997) The cranial anatomy of *Discosauricsus* [Kuhn, a seymouriamorph tetrapod](http://rstb.royalsocietypublishing.org/content/352/1351/257.short)
- [from the Lower Permian of the Boskovice Furrow \(Czech Republic\)](http://rstb.royalsocietypublishing.org/content/352/1351/257.short). *Phil Trans R Soc B*
- **352**, 257–302.
- **Klembara J, Berman DS, Henrici AC, et al.** (2005) New structures and reconstructions of
- the skull of the seymouriamorph *Seymouria sanjuanensis* Vaughn. *Ann Carnegie Mus* **74**,
- 217–224.
- **King GM** (1988) Anomodontia. In: *Encyclopedia of Paleoherpetology*, Part 17C (ed.
- Wellnhofer P), pp.1–174. Stuttgart: Gustav Fischer Verlag.
- **King GM, Oelofsen BW, Rubidge BS** (1989) The evolution of the *Dicynodon* feeding system.
- *Zool J Linn Soc* **96**, 185–211.
- **Kobayashi K, Kumakura M, Yoshimura K, et al.** (1998) Fine structure of the tongue and
- lingual papillae of the penguin. *Arch Histol Cytol* **61**, 37–46.
- **Kordikova EG** (2002) Comparative morphology of the palate dentition in *Proganochelys*
- *quenstedti* Baur 1887 from the Upper Triassic of Germany and chelonian ancestry. *Neues*
- *Jahrb Geol Paläontol-Abh* **225**, 195–249.
- **Kuhn O** (1969) Cotylosauria Part 6. Encyclopedia of Paleoherpetology. 137pp Gutsav
- Fischer Verlag, Stuttgart and New York.
- **Kuhn O** (1976) Thecodontia Part 13, Encyclopedia of Paleoherpetology. 137pp. Gutsav
- Fischer Verlag, Stuttgart and New York.
- **Langer MC, Schultz CL** (2000) A new species of the Late Triassic rhynchosaur
- *Hyperodapedon* from the Santa Maria Formation of south Brazil. *Palaeontology* **43**, 633– 652.
- **Langston WJr, Reisz RR** (1981) *Aerosaurus wellesi*, new species, a varanopseid mammal-
- like reptile (Synapsida: Pelycosauria) from the Lower Permian of New Mexico. *J Vertebr*
- *Paleontol* **1**, 73–96.
- **Lauder GV, Gillis GB** (1997) Origin of amniote feeding. In: *Amniote Origins: Completing the*
- *Transition to Land*. (eds Sumida SS, Martin KLM), pp. 169–206. London and New York:
- Academic Press.
- **Lauder GV, Shaffer HB** (1993) Design of feeding systems in aquatic vertebrates: major
- patterns and their evolutionary interpretations. In: *The Skull, volume 3, Functional and*
- *Evolutionary Mechanisms*. (eds Hanken J, Hall BK), pp. 113–149. Chicago: The University
- of Chicago Press.
- **Laurin M** (1993) [Anatomy and relationships of](http://mnhn.academia.edu/MichelLaurin/Papers/1242210/Anatomy_and_relationships_of_Haptodus_garnettensis_a_Pennsylvanian_synapsid_from_Kansas) *Haptodus garnettensis*, a Pennsylvanian
- [synapsid from Kansas.](http://mnhn.academia.edu/MichelLaurin/Papers/1242210/Anatomy_and_relationships_of_Haptodus_garnettensis_a_Pennsylvanian_synapsid_from_Kansas) *J Vertebr Paleontol* **13**, 200–229.
- **Laurin M** (1996) A reappraisal of *Utegenia*[, a Permo-Carboniferous seymouriamorph](http://www.tandfonline.com/doi/abs/10.1080/02724634.1996.10011327)
- [\(Tetrapoda: Batrachosauria\) from Kazakhstan.](http://www.tandfonline.com/doi/abs/10.1080/02724634.1996.10011327) *J Vertebr Paleontol* **16**, 374**–**383.
- **Laurin M, Reisz RR** (1996) The osteology and relationships of *Tetraceratops insignis*, the
- oldest known therapsid. *J Vertebr Paleontol* **16**, 95–102.
- **Lee MSY, Gow CE, Kitching JW** (1997) [Anatomy and relationships of the parieasaur](http://www.ncbi.nlm.nih.gov/pmc/articles/PMC1692386/)*,*
- *Pareiasuchus nasicornis* [from the Upper Permian of Zambia.](http://www.ncbi.nlm.nih.gov/pmc/articles/PMC1692386/) *Palaeontology* **40**, 307–335.
- **Li C, Wu XC, Rieppel O, et al.** (2008) An ancestral turtle from the Late Triassic of
- southwestern China. *Nature* **456**, 497–501.
- **Liu J, Rubidge B, Li J** (2009) A new specimen of *Biseridens qilianicus* indicates its
- phylogenetic position as the most basal anomodont. *Proc R Soc B* **277**, 285–292.
- **Maddin HC, Sidor CA, Reisz RR** (2008) Cranial anatomy of *Ennatosaurus tecton*
- (Synapsida: Caseidae) from the Middle Permian of Russia and the evolutionary
- relationships of Caseidae. *J Vertebr Paleontol* **28**,160–180.
- **Mahler DL, Kearney M** (2006) The palatal dentition in squamate reptiles: morphology,
- development, attachment, and replacement. *Fieldiana Zool* **108**, 1–61.
- **Martinez RN, Apaldetti C, Colombi CE, Praderio A, Fernandez E, Malnis PS, Correa GA,**
- **Abelin D, Alcober O** (2013) A new sphenodontian (Lepidosauria: Rhynchocephalia) from
- the Late Triassic of Argentina and the early origin of the herbivore opisthodontians. *Proc R*
- *Soc B Biol Sci* **280**, 2013–2057.
- **Maier W, Heever J van den, Durand F** (1996) New therapsid specimens and the origin of
- the secondary hard and soft palate of mammals. *J Zool Syst Evol Res* **34**, 9–19.
- **Martínez RN, Sereno PC, Alcober OA, et al.** (2011) A basal dinosaur from the dawn of the
- dinosaur era in southwestern Pangaea. *Science* **331**, 206–210.
- **Matsumoto R** (2011) The Palaeobiology of Choristodera (Reptilia: Diapsida). Ph.D.
- dissertation, University College London, UK, 437 pp.
- **Matsumoto R, Evans SE** (2015) Morphology and function of the palatal dentition in
- Choristodera. *J Anat*, doi:10.1111/joa.12414
- **Mendrez CH** (1972) On the skull of *Regisaurus jacobi*, a new genus and species of
- Bauriamorpha Watson and Romer 1956 (= Scaloposauria Boonstra 1953), from the
- *Lystrosaurus*−zone of South Africa. In: *Studies in Vertebrate Evolution*. (eds Joysey KA,
- Kemp TS), pp. 191–212. Edinburgh: Oliver and Boyd.
- **Mendrez CH** (1974a) Etude du crâne d'un jeune specimen de *Moschorhinus kitchingi* Broom,
- 1920 (? *Tigrisuchus simus* Owen, 1876), Therocephalia, Pristerosauria, Moschorhinidae,
- d'Afrique austral. *Ann S Afr Mus* **64**, 71–115.
- **Mendrez ChH** (1974b) A new specimen of *Promoschorhynchus platyrhinus* Brink 1954
- (Moschorhinidae) from the Daptocephalus-Zone (Upper Permian) of South Africa.
- *Palaeontol Afr* **17**, 69–85.
- **Modesto SP** (1995) The skull of the herbivorous synapsid *Edaphosaurus boanerges* from the
- Lower Permian of Texas. *Palaeontology* **38**, 213–239.
- **Modesto SP** (1998) New information on the skull of the Early Permian reptile *Captorhinus aguti*. *Paleobios* **18**, 21–35.
- **Modesto SP** (1999) Observations on the structure of the Early Permian reptile *Stereosternum*
- *tumidum*. *Palaeontol Afr* **35,** 7–19.
- **Modesto SP** (2006) The cranial skeleton of the Early Permian aquatic reptile *Mesosaurus*
- *tenuidens*: implications for relationships and palaeobiology. *Zool J Linn Soc* **146**, 345–368.
- **Modesto SP, Rubidge B, Welman J** (1999) The most basal anomodont therapsid and the
- primacy of Gondwana in the evolution of the anomodonts. *Proc R Soc* **266**, 331–337.
- **Modesto SP, Scott DM, Reisz RR** (2009) A new parareptile with temporal fenestration from
- the Middle Permian of South Africa. *Can J Earth Sci* **46**, 9–20.
- **Modesto SP, Scott DM, Berman DS, et al.** (2007) The skull and the palaeoecological
- significance of *Labidosaurus hamatus*, a captorhinid reptile from the Lower Permian of
- Texas. *Zool J Linn Soc* **149**, 237–262.
- **Moodie RL** (1909) Carboniferous air-breathing vertebrates of the United States National Museum. *Proc U S Natl Mus* **37**, 11–28.
- **Moodie RL** (1912) The skull structure of *Diplocaulus magnicornis* Cope and the amphibian
- order Diplocaulia. *J Morph* **23**, 31–44.
- **Moss JL** (1972) The morphology and phylogenetic relationships of the Lower Permian
- tetrapod *Tseajaia campi* Vaughn (Amphibia: Seymouriamorpha). *Univ California Pub Geol Sci* **98**, 1–72.
- **Motani R** (1999) Phylogeny of the Ichthyopterygia. *J Vertebr Paleontol* **19**, 472–495.
- **Müller J** (2007) First record of a thalattosaur from the Upper Triassic of Austria. *J Vertebr*
- *Paleontol* **27**, 236–240.
- **M r , Li J-L, Reisz RR** (2008) A new bolosaurid parareptile, *Belebey chengi* sp. nov.,
- from the Middle Permian of China and its paleogeographic significance. *Naturwiss* **95**,
- 1169–1174.
- **Müller J, Renesto S, Evans SE** (2005) The marine diapsid reptile *Endennasaurus* (Reptilia:
- Thalattosauriformes) from the Late Triassic of Italy. *Palaeontology* **48**, 15–30.
- **Neenan JM, Klein N, Scheyer TM** (2013) European origin of placodont marine reptiles and the evolution of crushing dentition in Placodontia. *Nature Comm* 4, no. **1621**. doi:10,1038/ncomms2633. **Nussbaum RA** (1977) Rhinatrematidae: a new family of caecilians (Amphibia: Gymnophiona). *Occ Pap Mus Zool, Univ Michigan 682*, 1–30. **Olson EC** (1947) The family Diadectidae and its bearing on the classification of reptiles. *Fieldiana Geol* **11**, 1–53. **Olson EC** (1962) Late Permian terrestrial vertebrates, USA and USSR. *Trans Amer Phil Soc* **52**, 1–224. **Olson EC** (1970) New and little known genera and species of vertebrates from the Lower Permian of Oklahoma. *Fieldiana Geol* **18**, 359–434. **Piveteau J** (1955) Existence d'un reptile du groupe des Procolophonidés à Madagascar - conséquences stratigraphiques et paléontologiques. *C R Acad Sci Paris* **241**,1325–1327. **Pyron RA, Burbrink FT, Wiens JJ** (2013) A phylogeny and revised classification of Squamata, including 4161 species of lizards and snakes. BMC *Evol Biol* **13**, **93**, 1-53. **Rage J-C, Roč k Z** (1989) Redescription of *Triadobatrachus massinoti* (Piveteau, 1936) an anuran amphibian from the Early Triassic". *Palaeontogr Abt A Palaeozool-Stratigr* **206**, 1– 16. **Regal PJ** (1966) Feeding specializations and the classification of terrestrial salamanders. *Evolution* **20**, 392–407. **Reisz RR** (1972) Pelycosaurian reptiles from the Middle Pennsylvanian of North America. *Bull Mus Comp Zool* **144**, 27–62. **Reisz RR** (1981) A diapsid reptile from the Pennsylvanian of Kansas. *Paleont Contrib Univ Kansas, Sp Pub* **7**, 1–74. **Reisz RR, Berman DS** (2001) The skull of *Mesenosaurus romeri*, a small varanopseid (Synapsida: Eupelycosauria) from the Upper Permian of the Mezen River Basin, northern Russia. *Ann Carnegie Mus* **70**, 113–132. **Reisz RR, Dilkes DW, Berman DS** (1998) Anatomy and relationships of *Elliotsmithia longiceps* Broom, a small synapsid (Eupelycosauria: Varanopseidae) from the Late Permian of South Africa. *J Vertebr Paleontol* **18**, 602–611.
	-
- **Reisz RR, Godfrey SJ, Scott D** (2009) *Eothyris* and *Oedaleops*: do these Early Permian
- synapsids from Texas and New Mexico form a clade? *J Vertebr Paleontol* **29**, 39-47.
- **Reisz RR, Scott DM** (2002) *Owenetta kitchingorum*, n. sp., a small parareptile from the

Lower Triassic of South Africa. *J Vertebr Paleontol* **22**, 244–256.

- **Reisz RR, Sues H** (2000) Herbivory in late Paleozoic and Triassic terrestrial vertebrates. In:
- *Evolution of Herbivory in Terrestrial Vertebrates.* (ed. Sues HD), pp. 9–41. Cambridge:
- Cambridge University Press.
- **Reisz RR, Berman DS, Scott D** (1992) The cranial anatomy of *Secodontosaurus obtusidens*,
- an unusual mammal-like reptile (Synapsida: Sphenacodontidae) from the Lower Permian
- of Texas. *Zool J Linn Soc* **104**,127–184.
- **Reisz RR, Modesto SP, Scott D** (2011) A new Early Permian reptile and its significance in early diapsid evolution. *Proc R Soc B* **278**, 3731–3737.
- **Rieppel O** (1994) Osteology of *Simosaurus gaillardoti* and the relationships of stem-group
- Sauropterygia. *Fieldiana Geol* **28**, 1–61.
- **Rieppel O, Reisz RR** (1999) The origin and early evolution of turtles. *Ann Rev Ecol Syst* **30**,1–22.
- **Rubidge BS** (1991) A new primitive dinocephalian mammal-like reptile from the Permian of southern Africa. *Palaeontology* **34**, 547–599.
- **Rubidge BS** (1994) *Australosyodon*, the first primitive anteosaurid dinocephalian from the
- Upper Permian of Gondwana. *Palaeontology* **37**, 579–594.
- **Rubidge BS, Heever JA** (1997) Morphology and systematic position of the dinocephalian
- *Styracocephalus platyrhinus*. *Lethaia* **30**, 157–168.

Robinson PL (1962) Gliding lizards from the Upper Keuper of Great Britain. *Proc Geol Soc,*

- *London* **1601**, 137–146.
- **Roč k Z** (1981) Cranial anatomy of frogs of the Family Pelobatidae Stannius, 1856, with
- outlines of their phylogeny and systematics. Acta Univ Carolinae Biol **1980**, 1–164.
- **Romer AS** (1946) The primitive reptile *Limnoscelis* restudied. *Am J Sci* **244**, 149–188.
- **Romer AS** (1956) *Osteology of the Reptiles*, pp 772. , Chicago: University of Chicago Press.
- **Romer AS** (1969) The cranial anatomy of the Permian amphibian *Pantylus*. *Breviora* **314**, 1–
- 37.

Romer AS, Witter RV (1942) *Edops*, a primitive rhachitomous amphibian from the Texas

Red Beds. *J Geol* **50**, 925–960.

- **Ruta M, Clack JA** (2006) A review of *Silvanerpeton miripedes*, a stem amniote from the
- Lower Carboniferous of East Kirkton, West Lothian, Scotland. *Trans R Soc Edin Earth Sci* **97**, 31–63.
- **Ruta M, Coates MI** (2007) Dates, nodes and character conflict: addressing the lissamphibian origin problem. *J Syst Palaeont* **5**, 69–122.
- **Ruta M, Jeffery JE, Coates MI** (2003) A supertree of early tetrapods. *Proc. R. Soc London B* **270**, 2507–2516.
- **SäiIä LK** (2010) The phylogenetic position of *Nyctiphruretus acudens*, a parareptile from the Permian of Russia. *J Iber Geol* **36**, 123–143.
- **Schwenk K** (2000a) An introduction to tetrapod feeding. In: *Feeding: Form, Function and*
- *Evolution in Tetrapod Vertebrates.* (ed. Schwenk K), pp. 21–63. San Diego and London: Academic Press.
- **Schwenk K (**2000b) Feeding in Lepidosaurs. In: *Feeding: Form, Function and Evolution in*
- *Tetrapod Vertebrates.* (ed. Schwenk K), pp. 175–291. San Diego and London: Academic Press.
- **Schoch RR, Milner AR** (2000). Stereospondyli. In: *Handbuch der Paläoherpetologie* 3B. (ed.
- Wellnhofer P), pp. 203. Munich: Verlag Dr. Friedrich Pfeil..
- **Seeley HG** (1887) [Researches on the structure, organization, and classification of the Fossil](http://www.jstor.org/stable/91700)
- Reptilia. 1. On *[Protorosaurus speneri](http://www.jstor.org/stable/91700)* (von Meyer). *Phil Trans R Soc B* **178**, 187**–**213.
- **Sereno PC** (1991) Basal archosaurs: phylogenetic relationships and functional implications. *Mem, Soc Vertebr Paleontol* **2**, 1–53.
- **Sereno PC, Martínez RN, Alcober OA** (2012) Osteology of *Eoraptor lunensis* (Dinosauria,
- Sauropodomorpha). Basal sauropodomorphs and the vertebrate fossil record of the
- Ischigualasto Formation (Late Triassic: Carnian-Norian) of Argentina. *J Vertebr Paleontol*
- **32**, 83–179.
- **Shimada K, Sato I, Yokoi A, et al.** (1990) The fine structure and elemental analysis of
- keratinized epithelium of the filiform papillae analysis on the dorsal tongue in the
- American alligator (*Alligator mississippiensis*). *Okajimas Folia Anat Japonica* **66**, 375–391.

Sidor CA (2001) Simplification as a trend in synapsid cranial evolution. *Evolution* **55**, 1419–

1442.

Sidor CA (2003) Evolutionary trends and the origin of the mammalian lower jaw.

Paleobiology **29**, 605–640.

- **Sidor CA, Rubidge BS** (2006) *Herpetoskylax hopsoni*, a new biarmosuchian (Therapsida:
- Biarmosuchia) from the Beaufort Group of South Africa; 76–113 pp in Carrano MT, Gaudin
- T, Blob R, Wible J (eds.), Amniote Paleobiology: Perspectives on the Evolution of
- Mammals, Birds, and Reptiles. University of Chicago Press, Chicago, Illinois.
- **[Sigogneau-Russell](http://en.wikipedia.org/w/index.php?title=Denise_Sigogneau-Russell&action=edit&redlink=1) D** (1989) Theriodontia I Phthinosuchia, Biarmosuchia, Eotitanosuchia,
- Gorgonopsia. In: *Encyclopedia of Paleoherpetology, Part 17 B.* (ed*.* Wellnhofer P), pp. 1–
- 815 127. Stuttgart: Gustav Fischer Verlag.
- **Sigogneau-Russell D, Russell DE** (1978) Étude ostéologique du Reptile *Simoedosaurus*
- (Choristodera). *Ann Paléontol (Vertébrés)* **64**, 1–84.
- **Sigurdsen T, Bolt JR** (2010) The Lower Permian amphibamid *Doleserpeton*
- 819 (Temnospondyli: Dissorophoidea): the interrelationships of amphibamids, and the origin of
- modern amphibians. *J Vertebr Paleontol* **30**, 1360–1377.
- **Sill WD** (1967) *Proterochampsa barrionuevoi* and the early evolution of the Crocodilia. *Bull Mus Comp Zool* **135**, 415–446.
- **Smith KK** (1993) The form of the feeding apparatus in terrestrial vertebrates: studies of
- adaptation and constraint. In: *The Skull volume 3, Functional and Evolutionary*
- *Mechanisms*. (eds Hanken J, Hall BK). pp. 150–196. Chicago: The University of Chicago Press.
- **Smithson TR** (1982) The cranial morphology of *Greererpeton burkemorani* (Amphibia:
- Temnospondyli). *Zool J Linn Soc* **76**, 29–90.
- **Snitting D** (2008) A redescription of the anatomy of the Late Devonian *Spodichthys buetleri*
- Jarvik, 1985 (Sarcopterygii, Tetrapodomorpha) from East Greenland. *J Vertebr Paleontol* **28**, 637–655.
- **Spielmann JA, Lucas SG, Rhinehart LF, et al.** (2008) The Late Triassic *Trilophosaurus*.
- *Bull New Mexico Mus Nat Hist Sci* **43**, 1–177.
- **Sumida SS, Berman SD, Martens T.** (1998) [A new trematopid amphibian](http://www.palass.org/publications/palaeontology-journal/archive/41/4/article_pp605-629) from the Lower
- Permian of central Germany. *Palaeont* **41**, 605–629.
- **Tatarinov LP** (1968) Morphology and systematics of the Northern *Dvina* cynodonts (Reptilia,

Therapsida); Upper Permian. *Postilla* **126**, 1–51.

- **Tatarinov LP** (1995) *Viatkosuchus sumini*, a new late Permian therocephalian from the Kirov
- Region: *Paleont J* **19**, 84–97.
- **Tchernov E, Rieppel O, Zaher H, et al.** (2000). A new fossil snake with limbs. *Science* **287**, 2010–2012.
- **Trueb L** (1993) Patterns of cranial diversity among the Lissamphibia. In: *The Skull: Patterns*
- *of Structural and Systematic Diversity.* (eds Hanken J, Hall BK), pp. 255–343. Chicago:
- 844 University of Chicago Press.
- **Tsuji LA** (2006) Cranial anatomy and phylogenetic affinities of the Permian parareptile
- *Macroleter poezicus*. *J Vertebr Paleontol* **26**, 849–865.
- **Tsuji LA, Müller J** (2009) Assembling the history of the Parareptilia: phylogeny,
- diversification, and a new definition of the clade. *Fossil Rec* **12**, 71–81
- **Tsuji LA, Müller J, Reisz RR** (2012) Anatomy of *Emeroleter levis* and the phylogeny of the nycteroleter parareptiles. *J Vertebr Paleontol* **32**, 45–67.
- **Tsuji LA, Müller J, Reisz RR** (2010) *Microleter mckinzieorum* gen. et sp. nov. from the
- Lower Permian of Oklahoma: the basalmost parareptile from Laurasia. *J Syst Palaeontol*
- **18**, 245–255.
- **Tverdokhlebova GI, Ivakhnenko MF** (1984) [Nycteroleterids from the Upper Permian of
- Eastern Europe]. *Paleont Zh* **1984**, 98–111. [Russian]
- **Vallin, G, Laurin M** (2004) Cranial morphology and affinities of *Microbrachis*, and a
- reappraisal of the phylogeny and lifestyle of the first amphibians. *J Vertebr Paleontol* **24**,
- 56–72.
- **Vaughn PP** (1955) The Permian reptile *Araeoscelis* restudied. *Bull Mus Comp Zool 113*,
- 305–467.
- **Wake DB** (1966) Comparative osteology and evolution of the lungless salamanders, Family
- Plethodontidae. *Mem South California Acad Sci* **4**, 1–111.
- **Wake DB, Deban SM** (2000) Terrestrial feeding in salamanders. In: *Feeding: Form, Function*
- *and Evolution in Tetrapod Vertebrates.* (ed. Schwenk K), pp. 95–116. San Diego and

London: Academic Press.

- **Walker AD** (1990) A revision of *Sphenosuchus acutus* Haughton, a crocodylomorph reptile
- from the Elliot Formation (Late Triassic or Early Jurassic) of South Africa. *Phil Trans R Soc B* **330**, 1–120.
- **Warren JW** (1961) The basicranial articulation of the early Permian cotylosaur, *Captorhinus*.
- *J Paleont* **35**, 561–563.
- **Watson DMS** (1913) *Batrachiderpeton lineatum* Hancock & Atthey, a Coal-Measure
- Stegocephalian. *J Zool* **83**, 949–962.
- **Watson DMS** (1954) On *Bolosaurus* and the origin and classification of reptiles. *Bull Mus*

Comp Zool **111**, 299–449.

- **Weems RE** (1980) An unusual newly discovered archosaur from the Upper Triassic of
- Virginia, U.S.A. *Trans Amer Phil Soc* **70**, 1–53.
- **Wellstead C** (1991) [Taxonomic revision of the Lysorophia, Permo-Carboniferous lepospondyl](http://digitallibrary.amnh.org/dspace/handle/2246/904) amphibians. *[Bull Amer Mus Nat Hist](http://digitallibrary.amnh.org/dspace/handle/2246/904)* **209**, 1**–**90.
- **Werneburg R, Lucas SG, Schneider JW, Rinehart LF** (2010) First Pennsylvanian *Eryops*
- (Temnospondyli) and its Permian record from New Mexico. *Bull New Mexico Mus Nat Hist*
- *Sci* **49**, 129–136.
- **Werneburg R, Ronchi A, Schneider JW** (2007) The Early Permian branchiosaurids
- (Amphibia) of Sardinia (Italy): systematic palaeontology, palaeoecology, piostratigraphy
- and palaeobiogeographic problems. *Palaeogeogr Palaeoclimatol Palaeoecol* **252**, 383**–** 404.
- **Werth A** (2000) Feeding in marine mammals. In: *Feeding: Form, Function and Evolution in*
- *Tetrapod Vertebrates.* (ed. Schwenk K), pp. 487–526. San Diego and London: Academic Press.
- **Wiens JJ** (2011) Re-evolution of lost mandibular teeth in frogs after more than 200 million years, and re-evaluating Dollo's Law. *Evolution* **65**, 1283–1296.
- **Wild R** (1973) Die Triasfauna der Tessiner Kalkalpen. XXIII. *Tanystropheus longobardicus*
- (Bassani) (Neue Ergebnisse). *Schweize Pälaont Abh* **95**,1–162.
- **Wild R** (1978) Die Flugsaurier (Reptilia, Pterosauria) aus der Oberen Trias von Cene bei
- Bergamo, Italien. *Boll Soc Paleont Italiana* **17**, 176–256.
- **Williston SW** (1911) A new family of reptiles from the Permian of New Mexico. *Amer J Sci* **31**, 378–398.
- **Williston SW** (1915) A new genus and species of American Theromorpha: *Mycterosaurus*
- *longiceps*. *J Geol* **23**, 554–559.

Wiens JJ, Kuczynski CA, Townsend T, Reeder TW, Mulcahy DG, Sites JWJnr (2010)

- Combining phylogenomics and fossils in higher-level squamate reptile phylogeny:
- molecular data change the placement of fossil taxa. *Syst Biol* **59**, 674-688.
- **Zaher H, Scanferla CA** (2012). The skull of the Upper Cretaceous snake *Dinilysia*
- *patagonica* Smith-Woodward, 1901, and its phylogenetic position revisited. *Zool J Linn Soc* **164**, 194–238.
- **Zaher H, Apesteguia S, Scanferla CA** (2009). The anatomy of the Upper Cretaceous snake
- *Najash rionegrina* Apesteguia & Zaher, 2006, and the evolution of limblessness in snakes. *Zool J Linn Soc* **156**, 801–826.
-
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Figure captions

- **Fig. 1** Phylogenetic tree for early tetrapods and amphibians showing arrangement of palatal
- 913 dentition. Colour coding of the palatal figures is consistent in all figures (tree modified from
- Ruta & Coates, 2007). Palatal figures as follows: 1, *Eusthenopteron*; 2, *Acanthostega*; 3,
- *Pederpes*; 4, *Crassigyrinus*; 5, *Greerepeton*; 6, *Edops*; 7, *Balanerpeton* (original image
- reflected); 8, *Phonerpeton*; 9, *Doleserpeton*; 10, *Dermophis mexicanus*, Gymnophiona; 11,
- *Stereochilus marginatum,* Caudata; 12, *Gastrotheca walker*, Anura; 13, *Silvanerpeton*; 14,
- *Proterogyrinus*; 15, *Seymouria*; 16, *Odonterpeton*; 17, *Rhynchonkos*; 18, *Cardiocephalus*
- (original image reflected); 19, *Pantylus*; 20, *Brachydectes*; 21, *Batrachiderpeton*; 22,
- *Ptyonius*; 23, *Diadectes*. Image sources: 1,2,4, Clack, 2012; 3, Clack & Finney, 2005; 5,
- Smithson, 1982; 6, Romer & Witter, 1942; 7, Holmes 2000; 8, Dilkes, 1990; 9, Sigurdsen &
- Bolt, 2010; 10-12, Duellman & Trueb, 1994; 13, Ruta & Clack, 2006; 14, Holmes, 1984; 15,

 Klembara et al. 2005; 16-22, Carroll et al. 1998; 23, Reisz & Sues, 2000;1, 10-13, 20 original without scale. Abbreviations: ANTH, Anthracosauria; LISS, Lissamphibia; SEY, Seymouriamorpha.

 Fig. 2 Photographs of the palatal tooth arrangement in various lineages: A, *Andrias japonicas* (Lisamphibia; NSM-PO-H-447); B, *Dimetrodon limbatus* (Synapsida; AMNH FR 4001); C, *Ikechosaurus sunailinae* (Choristodera, Diapsida; IVPP V9611-3), grey coloured area marks nasopalatal trough and blue coloured area marks the distribution of the palatal dentition. 931 Institutional abbreviations: American Museum Natural History (AMNH); IVPP Institute of 932 Vertebrate Paleontology and Paleoanthropology, Beijing, China (IVPP); National Museum of 933 Nature and Science, Tokyo (NSM). Anatomical abbreviations: d, dentary; ept, ectopterygoid; 934 hy, hyoid; pal, palatine; psh, parasphenoid; pt, pterygoid; pt fl, pterygoid flange; v, vomer. **Fig. 3** Skulls of synapsids in palatal view (phylogeny based on Sidor, 2001): 1 *Cotylorhynchus*; 2, *Ennatosaurus*; 3, *Mesenosaurus*; 4, *Varanosaurus*; 5, *Edaphosaurus*; 6, *Haptodus*; 7, *Secodontosaurus*; 8, *Tetraceratops*; 9, *Biarmosuchus*; 10, *Lycaenodon*; 11, *Herpetoskylax*; 12, *Titanophoneus*; 13, *Syodon*; 14, *Styracocephalus* (original without scale); 15, *Estemmenosuchu*s; 16, *Struthiocephalus*. Image sources: 1, Reisz & Sues, 2000; 2, Maddin et al. 2008; 3, Reisz & Berman, 2001; 4, Berman et al. 1995; 5, Modesto, 1995; 6, Laurin, 1993; 7, Reisz et al. 1992; 8, Laurin & Reisz, 1996; 9, Ivakhnenko, 1999; 10-11, Sigogneau-Russell, 1989; 12-13, 15, King 1988; 14, Rubidge & van den Heever, 1997; 16, Rubidge, 1991. Abbreviations: BIAR, Biarmosuchia; CASE, Caseasauria; DINO, Dinocephalia; OPHI, Ophiacodontidae; SPHE, Sphenacodontidae; VARA, Varanopidae. **Fig. 4** Skulls of synapsids in palatal view (phylogeny based on Sidor, 2001), continued from Figure 4: 1, *Aelurosaurus*; 2, *Arctognathus*; 3, *Leontocephalus*; 4, *Scylacops*; 5, *Aloposaurus*; 6, *Gorgonops*; 7, *Arctops*; 8, *Prorubidgea*; 9, *Dinogorgon*; 10, *Rubidgea* (original without scale); 11, *Theriognathus*; 12 *Viatkosuchus* (original without scale); 13 *Regisaurus*. Image sources: 1-10, Sigogneau-Russell, 1989; 11,13, Kemp, 1982; 12, Tatarinov, 1995.

- **Fig. 5** Skulls of parareptiles in palatal view (phylogeny based on Tsuji et al., 2012). 1,
- *Mesosaurus*; 2, *Millerosaurus*; 3, *Acleistorhinus*; 4, *Nyctiphruretus*; 5, *Procolophon* (original
- without scale); 6, *Owenetta*; 7, *Scutosaurus*; 8, *Pareiasuchus*; 9, *Macroleter*; 10, *Nycteroleter*;

11, *Bashkyroleter mesensis*. Image sources: 1, Modesto, 2006; 2, 4,7, Carroll, 1988; 3,

- DeBraga & Reisz, 1996; 5, Carroll & Lindsay, 1985; 6, Reisz & Scott, 2002; 8, Lee et al.
- 1997; 9, Tsuji, 2006; 10, Tverdokhlebov & Ivakhnenko, 1984; 11, Ivakhnenko, 1997.
- Abbreviations: LANT, Lanthanosuchidae; BOL, Bolosauridae; PROCOL, Procolophonoidea;
- PAREIA, Pareiasauria.
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- **Fig. 6** Skulls of Eureptilia and Diapsida, Sauropterygia, Ichthyopterygia, and
- Lepidosauromorpha in palatal view (phylogeny based on DeBraga & Rieppel, 1997; Pyron et
- al. 2013; Rieppel & Reisz, 1999; Wiens et al. 2010) 1, *Captorhinus*; *2, Paleothyris*; 3,
- *Petrolacosaurus*; 4, *Araeoscelis*; 5, *Claudiosaurus*; 6,*Youngina*; 7, *Placodus*; 8,
- *Kuehneosaurus*; 9, *Marmoretta*; 10, *Gephyrosaurus*; 11, *Clevosaurus*; 12, *Sphenodon*; 13,
- *Lacerta*; 14, *Ctenosaura* (original without scale); 15, *Ophisaurus*; 16 *Heloderma*; 17,
- *Shinisaurus*; 18, *Platecarpus* (original without scale); 19, *Anilius*. Image sources: 1, Reisz &
- Sues, 2000; 2, Benton, 2000; 3, Reisz, 1981; 4, Vaughn, 1955; 5-7, Carroll, 1988; 8,
- Robinson, 1962; 9, Evans, 1991; 10-11, Jones, 2006; 12,18, Romer, 1956; 13-17, Evans,
- 2008; 19, Cundall & Irish, 2008. Abbreviation: Rhyncho, Rhynchocephalia.

- **Fig. 7** Skulls of Archosauromorph in palatal view (phylogeny based on Brusatte et al. 2010;
- Borsuk−Białynicka & Evans, 2009a; Dilkes & Sues, 2009): 1, *Czatkowiella*; 2, *Cteniogenys*; 3,
- *Proganochelys*; 4, *Mesosuchus*; 5, *Tanystropheus*; 6, *Proterosuchus*; 7, *Osmolskina*; 8,
- *Euparkeria*; 9, *Doswellia*; 10, *Proterochampsa*. Image sources: 1, Borsuk−Białynicka &
- Evans, 2009a; 2, Evans, 1990; 3,6, Carroll, 1988; 4, Dilkes, 1998; 5, Wild, 1987; 7,
- Borsuk−Białynicka & Evans, 2009b; 8, Ewer, 1965; 9, Weems, 1980; 10, Sill, 1967.

Fig. 8 Summary of evolutionary patterns in the palatal dentition of tetrapods.

- **Fig. 9** Summary of evolutionary history of soft tissues related to feeding through tetrapod
- 984 evolution (see text for detail and references).

- **Fig. 10** Keratinized oral epithelium in extant taxa; A, *Anas platyrhynchos* (Mallard; KPM-NF
- 2002622, floor of mouth (left) and palate (right); B, *Spheniscus demersus* (African Penguin;
- KPM-NF 2002403), dissection photographs and CT image of a sagittal section; C,
- *Osteolaemus tetraspis* (Dwarf Crocodile; Ueno Zoo, Tokyo Japan, no number), palatal
- surface; D, *Chelonia agassizii* (Galápagos Green Turtle; KPM-NFR 389), palatal surface with
- keratinized keels and serrations. Institutional abbreviation: Kanegawa Prefectural Museum of
- Natural History (KPM-NF).
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- Appendices 1–4
- 995 1. Early tetrapods and amphibians, arrangement of the palatal dentition
- 2. Synapsida, arrangement of the palatal dentition
- 3. Parareptilia, arrangement of the palatal dentition
- 4. Diapsida, arrangement of the palatal dentition
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Supplementary information

- **Sup-Fig. 1.** Skulls of early tetrapods in palatal view. A, *Eusthenopteron* (original without
- scale); B, *Acanthostega*; C, *Pederpes*; D, *Crassigyrius*; E, *Greerepeton*; F, *Edops*; G,
- *Balanerpeton*; H, *Phonerpeton*; I, *Doleserpeton*; J, *Silvanerpeton*; K, *Proterogyrinus*; L,
- *Pholiderpeton*; M, *Seymouria*; N, *Odonterpeton*; O, *Microbrachis*; P, *Hapsidopareion*; Q,
- *Rhynchonkos*; R, *Cardiocephalus* (original image reflected); S, *Pantylus*; T, *Brachydectes*
- (original without scale); U, *Batrachiderpeton*; V, *Ptyonius*; W, *Diadectes*; X, *Dermophis*
- *mexicanus* (Gymnophiona); Y, *Stereochilus marginatum* (Caudata); Z, *Gastrotheca walker*
- (Anura), original without scale. Image sources: A, B,D, Clack, 2012; C, Clack & Finney, 2005;
- E, Smithson, 1982; F, Romer & Witter, 1942; G, Holmes, 2000 (original image reflected); H,
- Dilkes, 1990; I, Sigurdsen & Bolt, 2010; J, Ruta & Clack, 2006; K, Holmes, 1984; L, Clack,
- 1987; M, Klembara et al. 2005; N, P-T, Carroll et al. 1998; O, Vallian & Laurin, 2004; U-V,
- Carroll et al. 1998; W, Reisz & Sues, 2000; X-Z, Duellman & Trueb, 1994 . Colour coding on
- 1013 the palate same as text Figures 1–7.
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- **Sup-Fig. 2** Skulls of synapsids in palatal view, Part 1: A, *Cotylorhynchus* (Caseasauria); B,
- *Ennatosaurus* (Caseasauria); C, *Mesenosaurus* (Varanopidae); D, *Varanosaurus*
- (Ophiacodontidae); E, *Edaphosaurus*; F, *Haptodus*; G, *Secodontosaurus*
- (Sphenacodontidae); H, *Tetraceratops*; I, *Biarmosuchus*; J, *Lycaenodon* (Biarmosuchia); K,
- *Titanophoneus* (Dinocephalia); L, *Syodon* (Dinocephalia); M, *Styracocephalus* (Dinocephalia,
- original without scale bar); N, *Estemmenosuchus* (Dinocephalia); O, *Struthiocephalus*
- (Dinocephalia); P, *Ulemosaurus* (Dinocephalia). Image sources: A, Reisz & Sues, 2000; B,
- Maddin et al. 2008; C, Reisz & Berman, 2001; D, Berman et al. 1995; E, Modesto, 1995; F,
- Laurin, 1993; G, Reisz et al. 1992; H, Laurin & Reisz, 1996; I, Ivakhnenko, 1999; J,
- Sigogneau-Russell,1989; K-L, N-P, King, 1988; M, Rubidge & van den Heever, 1997.
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- **Sup-Fig. 3** Skulls of synapsids in palatal view, Part 2: A, *Otsheria* (Anomodontia, original
- without scale); B, *Aelurosaurus* (Gorgonopsidae); C, *Arctognathus* (Gorgonopsidae); D,
- *Leontocephalus* (Gorgonopsidae); E, *Scylacops* (Gorgonopsidae) ; F, *Arctops*
- (Gorgonopsidae); G, *Prorubidgea* (Gorgonopsidae) ; H, *Dinogorgon* (Gorgonopsidae) ; I,
- *Rubidgea* (Gorgonopsidae); J, *Moschorhinus* (Therocephalia); K, *Theriognathus*
- (Therocephalia); L, *Viatkosuchus* (Therocephalia, original without scale bar); M, *Regisaurus*
- (Therocephalia); N, *Bauria* (Therocephalia, original without scale); O, *Dvinia* (Cynodontia,
- original without scale). Image sources: A, K, M-N, Kemp, 1982; B-I, Sigogneau-Russell, 1989;
- J, Mendrez, 1974a; L, Tatarinov, 1995; O, Tatarinov, 1968.
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- **Sup-Fig. 4** Skulls of Parareptilia in palatal view. A, *Mesosaurus*; B, *Millerosaurus*; C,
- *Lanthanosuchus*; D, *Acleistorhinus* (Lanthanosuchidae); E, *Belebey* (Bolosauridae); F,
- *Nyctiphruretus*; G, *Procolophon* (Procolophonoidea, original without scale); H, *Owenetta*
- (Procolophonoidea); I, *Scutosaurus* (Pareiasauria); J, *Pareiasuchus* (Pareiasauria); K,
- *Macroleter* ('nycteroleter'); L, *Nycteroleter*; M, *Bashkyroleter mesensis* ('nycteroleter', original
- without scale). Image sources: A, Modesto, 2006; B,F,I, Carroll, 1988; C,D, DeBraga &
- Reisz, 1996; E, Ivakhnenko & Tverdochlebova, 1987; G, Carroll & Lindsay, 1985; H, Reisz &
- Scott, 2002; J, Lee et al. 1997; K, Tsuji, 2006; L, Tverdokhlebov & Ivakhnenko, 1984; M,

Ivakhnenko, 1997.

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- **Sup-Fig. 5** Skulls of eureptiles and basal diapsids (A-F), Sauropterygia (G–H),
- Ichthyopterygia (I), and Lepidosauromorpha (J–Z) in palatal view: A, *Captorhinus*; *B,*
- *Paleothyris*; C, *Petrolacosaurus*; D, *Araeoscelis*; E, *Claudiosaurus*; F, *Youngina*; G, *Placodus*;
- H, *Simosaurus*; I, *Ichthyosaurus* (original without scale); J, *Kuehneosaurus*; K, *Marmoretta*; L,
- *Gephyrosaurus* (Rhynchocephalia); M, *Clevosaurus* (Rhynchocephalia) ; N, *Sphenodon*
- (Rhynchocephalia); O, *Hemitheconyx* (Squamata, Gekkota); P, *Tropidophorus* (Squamata,
- Scincoidea); Q, *Lacerta* (Squamata, Lacertoidea); R, *Uromastyx* (Squamata, Iguania); S,
- *Ctenosaura* (Squamata, Iguania: original without scale); T, *Xenosaurus* (Squamata,
- Anguimorpha); U, *Ophisaurus* (Squamata, Anguimorpha); V, *Heloderma* (Squamata,
- Anguimorpha); W, *Shinisaurus* (Squamata, Anguimorpha) ; X, *Varanus* (Squamata,
- Anguimorpha) ; Y, *Platecarpus* (Squamata, Mosasauria: original without scale); Z, *Anilius,*
- Squamata, Serpentes). Image sources: A, Reisz & Sues, 2000; B, Benton, 2000; C, Reisz,
- 1981; D, Vaughn, 1955; E-G, Carroll, 1988; H, Rieppel, 1994; I,N,Y, Romer, 1956; J,
- Robinson, 1962; K, Evans, 1991; L-M, Jones, 2006; O-X, Evans, 2008; Z, Cundall & Irish,
- 2008.

- **Sup-Fig. 6** Skulls of Archosauromorpha in palatal views: A, *Czatkowiella*; B, *Cteniogenys*
- (Choristodera); C, *Proganochelys* (Testudines); D, *Mesosuchus* (Rhynchosauria); E,
- *Trilophosaurus;* F, *Paradapedon* (Rhynchosauria); G, *Tanystropheus*; H, *Proterosuchus*; I,
- *Euparkeria*; J, *Doswellia*; K, *Proterochampsa*; L, *Rutiodon* (Phytosauria); M, *Stagonolepis*
- (Aetosauria); N, *Sphenosuchus* (Crocodylomorpha); O, *Ornithosuchus*. Image sources: A,
- Borsuk−Białynicka & Evans, 2009a; B, Evans, 1990; C, E, F, H, Carroll, 1988; D, Dilkes,
- 1998; G, Wild, 1987; I, Ewer, 1965; J, Weems, 1980; K, Sill, 1967; L-M, O, Kuhn, 1976; N,
- Walker, 1990.
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Figure 3

Figure 6

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Figure 10

