

1 Changes in body composition following haemodialysis as assessed by
2 bioimpedance spectroscopy

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29 Abstract

30 Background

31 Patients with chronic kidney disease treated by haemodialysis (HD) are
32 at increased risk of sarcopenia. Bioelectrical impedance spectroscopy (BIS) can
33 be used to determine body composition, and is one of several potential screening
34 tools for sarcopenia. The newer generation of portable hand held devices can be
35 readily used in dialysis centres. The results from BIS devices using a two
36 compartmental model of body composition can be affected by hydration status

37 and so ideally measurements should be made when patients are not
38 overhydrated. More recently BIS devices using a three compartmental body
39 model, which separate normally hydrated lean tissues from extracellular water
40 (ECW) excess. We wished to determine whether body composition measured
41 using such a BIS device was affected by hydration status.

42 Methods

43 We performed BISs pre and post-haemodialysis using a three body
44 compartmental model .

45 Results

46 BISs were recorded in 48 patients; 68.8% male; mean age 67.70 ± 14.21
47 years, weight pre dialysis 70.54 ± 18.07 , which fell post to 68.58 ± 17.78 kg,
48 Extracellular water fell (16.92 ± 4.76 vs 15.66 ± 4.43 L, $p < 0.001$), whereas there
49 was no change for intracellular water (14.84 ± 4.27 vs 14.90 ± 4.68 L). Fat free mass
50 index (FFMI) fell (7.87 ± 3.98 vs 6.78 ± 3.97 kg/m², $p < 0.001$), whereas fat mass
51 index (FMI) increased from (7.87 ± 3.98 vs 8.12 ± 3.81 kg/m², $p = 0.002$). A fall in
52 FFMI was associated with an increase in FMI ($r = 0.804$, $p < 0.001$).

53 Conclusion

54 FMI and FFMI measured by BIA are both confounded by hydration
55 status. Although pre-dialysis measurements are more convenient, we suggest
56 BIS should preferably be performed post-dialysis when patients are less over-
57 hydrated and have less electrolyte imbalances.

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76 Introduction

77 Patients with chronic kidney disease (CKD) at increased risk of

78 sarcopenia, and once sarcopenia has developed then patients are at increased

79 risk for mortality. Muscle mass can be determined from anthropomorphic

80 measurements and muscle wasting assessed as part of the subjective global

81 assessment (SGA) [1]. However these methods are relatively insensitive in

82 detecting early changes, observer dependent, and potentially time consuming. As

83 such other screening tests including dual energy X ray absorptiometry (DXA)

84 and bioelectrical impedance assessment (BIA) have been recommended as
85 alternatives [2,3].

86 DXA scanning and most bioimpedance devices divide the body into 2
87 compartments; fat mass and fat free mass [4]. Previous reports have
88 demonstrated a strong correlation between body composition as assessed by
89 multi-frequency BIA and DXA scanning in dialysis patients [5,6]. However BIA
90 assessments of body hydration status and body composition can be affected by
91 localised fluid, such as ascites and intra-peritoneal fluid [7,8], and also by over
92 hydration [9]. Haemodialysis patients gain fluid between dialysis sessions, and
93 then this excess fluid is removed during the haemodialysis session. This
94 relatively rapid change in body fluid following dialysis has been demonstrated to
95 lead to a change in body composition as determined by BIA [10]. As such BIA
96 should preferably be made when the haemodialysis patient is closest to their
97 dry or target weight and not pre-dialysis when they are volume overloaded.

98 In clinical practice it is much more convenient to make BIAs prior to the
99 haemodialysis session, rather than asking patients to remain behind for BIAs.
100 More recently BIS devices using 3 compartmental body composition models have
101 been developed [11]. These devices determine the normally hydrated lean body
102 mass, and as such should not be affected by the changes in hydration that occur
103 with haemodialysis. We therefore decided to review body composition data from
104 patients who had corresponding pre and post haemodialysis BIS to determine
105 whether there were no changes in body composition with dialysis, so that pre-
106 dialysis measurements of body composition alone would suffice.

107

108 Methods

109 We audited the results of body composition estimations taken pre and
110 post haemodialysis using bioimpedance spectroscopy (BodyStat multiscan 5000
111 ,BodyStat, Douglas, Isle of Man). Four electrodes were placed according to the
112 manufacturer's instructions on the dorsal surface of the contra-lateral hand and
113 foot and ankle to the arterio-venous fistula or arterio-venous graft. Patients
114 were weighed and height recorded prior to dialysis (Marsden Weighing Group,
115 Rotherham, UK). Pre dialysis measurements were made with patients lying supine
116 on a bed. Patients dialysed using Fresenius 4008H (Fresenius Bad Homburg,
117 Germany) or Dialogue R+ (BBraun, Melsungen, Germany) with high flux
118 polysulfone dialyzers (Elisio, Nipro Corporation, Osaka, Japan) [12] and
119 anticoagulated with bolus low molecular weight heparin (Tinzaparin, Leo
120 Laboratories, Hurley, Berkshire, UK) [13]. Dialysis machines were regularly
121 serviced and dialysate conductivity checked against flame photometry [14]. We
122 audited 48 consecutive patients attending for haemodialysis with corresponding
123 pre and post dialysis measurements. Patients were not allowed to eat during
124 dialysis and were restricted to one small drink (180 ml). After the
125 haemodialysis session had been completed and the fistula needle sites had
126 stopped bleeding, then patients were reweighed using the same scales, returned
127 to lying supine on their bed, rested and then post dialysis measurements were
128 repeated. There was no change in the position of the electrodes between
129 measurements.

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131 **Ethics**

132 Standard of care for haemodialysis patients at the Royal Free Hospital
133 has been to measure bioimpedance pre and post dialysis sessions. We audited
134 the results of pre and post haemodialysis BIS measurements to determine
135 whether bioimpedance derived body composition should be measured pre or post
136 dialysis to be the standard of care for routine clinical practice. Ethical approval
137 fulfilled UK NHS clinical service development and audit (UK NHS guidelines for
138 clinical audit and service development ([http://www.hra.nhs.uk/documents](http://www.hra.nhs.uk/documents/2013/09/defining-research.pdf)
139 [/2013/09/defining-research.pdf](http://www.hra.nhs.uk/documents/2013/09/defining-research.pdf))) with all patient data being anonymised.

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141 **Statistical methods**

142 Statistical analysis was by parametric or non-parametric pair testing by
143 Student's t test or Mann Whitney U test, and Anova. Univariate correlation was
144 by Pearson or Spearman correlation (Prism 6.0, Graph Pad, San Diego, USA), and
145 Bland Altman analysis (version 3.0, Analyse It, Leeds, UK), with significance at
146 the $p < 0.05$ level. Data is reported as mean \pm standard deviation, median and
147 interquartile range or percentage.

148

149 **Results**

150 BIS was measured pre and post-dialysis in 48 patients; 68.8% male; mean
151 age 67.70 ± 14.21 years (mean \pm standard deviation), attending for in-centre
152 haemodialysis treatments. Median urine output was <50 (<50 -58) ml/day, Stoke-

153 Davies co-morbidity grade 1 (0-1), and Canadian Society of Geriatrics frailty
154 score 4 (4-5.8). Fourteen patients had diabetic nephropathy, 13 undetermined
155 cause, 5 interstitial renal diseases, 4 hypertensive nephropathy, 4 primary
156 glomerular disease, 3 myeloma kidney, 2 systemic lupus erythematosus, one each
157 of haemolytic uraemic syndrome, pre-eclampsia, and amyloid. All patients were
158 prescribed folic acid and B vitamin supplements, and 50% of patients were
159 additionally prescribed anti-hypertensive medications and 22% diuretics.

160 Mean weight pre dialysis was 70.54 ± 18.07 kg, height 165.80 ± 11.34 cm,
161 and body mass index (BMI) was 25.52 ± 6.11 kg/m².

162 The median dialysis session time was 4.0 (3.5-4) hours, dialysate
163 temperature 35 ($35-35.5$)^oC, sodium 137 (136-138) mmol/l, potassium 2.0 (1.0-
164 2.0) mmol/l, calcium 1.35 (1.25-1.35) mmol/l, magnesium 0.5 mmol/l, bicarbonate
165 32 mmol/l, acetate 3.0 mmol/l.

166 Weight fell post dialysis, as did extracellular water (ECW), but there was
167 no overall significant change in intracellular water (ICW) (Table 1). Assessments
168 of body composition showed an increase in measurements for body fat and a fall
169 in fat free tissues (table 1). As changes in ECW and ICW lead to corresponding
170 changes in BIS derived body composition, we then looked for differences at the
171 individual patient level, by Bland Altman analysis comparing the differences
172 between corresponding pre and post-dialysis values

173 There was an overall negative mean bias for ICW (Figure 1), suggesting a
174 small increase in ICW following dialysis and a positive bias for ECW (Figure 2)
175 showing a loss of ECW. As such there was a mean negative bias for fat mass

176 index (FMI) of -0.23 (95% limits of agreement -1.16 to 0.71 kg) corresponding to
177 an increase in FMI post-dialysis, and a mean positive bias for fat free mass
178 index (FFMI) of 0.83 (-0.4 to 1.79), with a loss of FFMI post-dialysis. The
179 corresponding coefficient of variation for FMI and FFMI were 0.49 and 0.24
180 respectively.

181 There were positive correlations between the change in pre and post
182 dialysis ICW and the change in fat free mass ($r= 0.29, p=0.042$) and muscle mass
183 ($r=0.31, p=0.031$), and a negative correlation with the change in fat mass ($r=-$
184 $0.30, p=0.037$). There were positive correlations between the change in pre and
185 post dialysis ECW and change in fat free mass ($r=0.95, p<0.001$), muscle mass
186 (Figure 3) lean dry mass ($r=0.37, p=0.01$) and a negative correlation with fat
187 weight ($r= 0.78, p<0.001$). There was a strong correlation between the change in
188 muscle mass and fat mass ($r=0.82, p<0.001$) (Figure 4).

189 To ensure that patients had re-equilibrated post-dialysis, and that
190 measurements did not change following dialysis, two additional BIS
191 measurements were made post-dialysis . There were no significant differences
192 between 1st, 2nd or 3rd post-dialysis measurements (table 2).

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194 Discussion

195 BIA devices have advanced from single frequency to multiple frequency
196 and bio-impedance spectroscopy (BIS) devices [15] with measurements
197 comparable to isotopic methods [16]. Standard BIA devices use a 2
198 compartmental model of body composition, dividing the body into fat and fat

199 free mass. However this approach is affected by hydration status, and as muscle
200 contains more water than fat, when over hydrated then muscle mass is over
201 estimated [9,10,17]. As such we made measurements with a BIS device using a 3
202 compartmental model approach which aims to distinguish normally hydrated
203 muscle mass from pathologic fluid overload when assessing body composition
204 [11]. This is based on comparing measured ECW and that estimated using the
205 slope of ECW volume and body weight at normovolaemia, where the slope is
206 0.239 L/kg for men and 0.214 L/kg for women [11]. As such these bioimpedance
207 devices potentially have the advantage of being able to assess body composition
208 using a single pre-dialysis measurement, as there should be no change in fat free
209 index or fat index with dialysis, whereas standard bioimpedance devices using a
210 2 compartment model require post-dialysis measurements to prevent errors due
211 to hydration status [10].

212 We observed that whereas both weight and ECW fell post dialysis there
213 was no significant change in ICW. Associated with weight loss there were
214 changes in body composition following haemodialysis, with a fall in fat free mass,
215 muscle and dry lean tissue, and an increase in fat mass. The changes in muscle
216 mass we observed are in keeping with an earlier version of the BodyStat [18],
217 but differ from studies using other manufacturers 3 compartmental BIS
218 devices [18]. When we looked at the individual patient basis, then the greater
219 the fall in ECW, there was a corresponding greater fall in fat free mass and
220 muscle mass, but with a negative correlation with fat mass. We observed that a
221 fall in ICW following haemodialysis was associated with a reduction in muscle

222 mass, whereas a gain in ICW was associated with an increase in muscle mass,
223 which is in keeping with studies using 3 compartmental BIS devices from
224 different manufacturers [18].

225 So in keeping with other 2 and 3 compartmental BIA and BIS models this
226 device over estimated muscle mass and under estimated fat mass pre dialysis
227 compared to post dialysis values. As such despite a model designed to separate
228 ECW excess from normo-hydrated fat free tissues this model was affected by
229 over hydration. This may not be surprising as these models use predictive
230 models based on ECW relative to ICW, and the relative increase in ECW and
231 ICW between dialysis sessions varies between patients due to differences in
232 sodium and water gains. In addition, previous reports have suggested that BIA
233 measurements can also be affected by body temperature, electrolyte
234 imbalances, and haematocrit [19].

235 Previous studies of patient pre dialysis, when patients were over
236 hydrated, showed that the BIS over estimated total body skeletal muscle mass
237 compared to magnetic resonance in patients with lower muscle mass and
238 conversely under estimated total body skeletal muscle mass in those patients
239 with greater muscle mass [20]. These differences may be due to errors in
240 estimating normo-hydrated fat free tissue [11], as muscle in dialysis patients
241 may not simply just contain more water pre dialysis, but also urea and other
242 azotaemic toxins, and more recently studies have additionally reported higher
243 sodium content, and that sodium can be removed from tissue stores during a
244 haemodialysis session [21].

245 Following a haemodialysis session there is redistribution of urea and
246 other electrolytes, and fluid, which tends to be greater with faster blood flows
247 [22]. To overcome this rebound phenomenon, dialysis adequacy studies based on
248 changes in serum urea concentrations advocate taking a delayed post dialysis
249 serum urea sample to overcome this effect [23]. It is therefore important that
250 BIS is performed after re-equilibration [24,25]. We waited for fistula needle
251 sites to have stopped bleeding, and then re-weighed patients, waited for
252 patients to return to their beds, resume the supine posture, allowed additional
253 time for the postural change and then performed BISs [26]. We then repeated
254 BISs to ensure that there were no discernible changes to establish that
255 patients were in a stable state post dialysis.

256 Patients with chronic kidney disease are at increased risk of sarcopenia,
257 and portable hand held BIS devices could be used to screen patients for
258 sarcopenia [2]. Although there are different classifications for defining
259 sarcopenia, many simply use a cut-off point of more than 2 standard deviations
260 from the normal healthy adult lean body mass index. As such, there could
261 potentially be a difference in the number of patients being classified with
262 sarcopenia depending upon whether BIS is measured pre or post haemodialysis,
263 with fewer patients being recorded as having sarcopenia when using pre-dialysis
264 compared to post dialysis estimations.

265 Bioimpedance measurements are dependent upon the length of the circuit
266 and the cross sectional area. As such, moving the electrode placements changes
267 the results, so one advantage of the Multiscan 5000 was that the larger

268 electrodes remained in position during dialysis, and no electrodes had to be
269 replaced. However this device measures whole body fluid volumes and body
270 composition using paired ipsi-lateral hand and foot electrodes, with the
271 assumption that patients are symmetrical. However dialysis patients may be
272 asymmetrical due to amputations, previous deep venous thrombosis, or central
273 venous occlusion, stroke, polio etc.

274 Most haemodialysis patients are over hydrated and following a
275 haemodialysis session there are changes in ICW and ECW [27,28]. Despite using
276 a BIS device based on a 3 compartmental body model, there were still changes in
277 body composition associated with over hydration. So although pre dialysis BIA
278 measurements are more convenient to both patients and dialysis centre staff,
279 for greater reliability BIS measurements should preferably be made when a are
280 closest to their normo-hydrated state to reduce errors in estimation of body
281 composition when patients are over hydrated.

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284 References

- 285 1. Clinical practice guidelines for nutrition in chronic renal failure. K/DOQI,
286 National Kidney Foundation. Am J Kidney Dis. 2000;35(6 Suppl 2):S1-140
- 287 2. Kondrup J, Allison SP, Elia M, Vellas B, Plauth M; Educational and Clinical
288 Practice Committee, European Society of Parenteral and Enteral Nutrition
289 (ESPEN). ESPEN guidelines for nutrition screening 2002. Clin Nutr.
290 2003;22(4):415-21
- 291 3. Kyle UG, Bosaeus I, De Lorenzo AD, Deurenberg P, Elia M, Gómez JM, et
292 al ; Composition of the ESPEN Working Group. Bioelectrical impedance
293 analysis-part II: utilization in clinical practice. . Clin Nutr. 2004; 23(6):
294 1430-53
- 295 4. Kyle UG, Bosaeus I, De Lorenzo AD, Deurenberg P, Elia M, Gómez JM,
296 Heitmann BL, Kent-Smith L, Melchior JC, Pirlich M, Scharfetter H, Schols
297 AM, Pichard C; Composition of the ESPEN Working. Bioelectrical

- 298 impedance analysis--part I: review of principles and methods. *Clin Nutr.*
299 2004;23(5):1226-43.
- 300 **5.** Fürstenberg A, Davenport A. Comparison of multifrequency bioelectrical
301 impedance analysis and dual-energy X-ray absorptiometry assessments in
302 outpatient haemodialysis patients. *Am J Kidney Dis.* 2010;57(1):123-129
- 303 **6.** Fürstenberg A, Davenport A. Assessment of body composition in
304 peritoneal dialysis patients using bioelectrical impedance and dual-energy
305 x-ray absorptiometry. *Am J Nephrol.* 2011;33(2):150-6
- 306 **7.** Davenport A. Does peritoneal dialysate affect body composition
307 assessments using multi-frequency bioimpedance in peritoneal dialysis
308 patients? *Eur J Clin Nutr.* 2013;67(2):223-5
- 309 **8.** Davenport A, Argawal B, Wright G, Mantzoukis K, Dimitrova R, Davar J,
310 Vasianopoulou P, Burroughs AK. Can non-invasive measurements aid clinical
311 assessment of volume in patients with cirrhosis? *World J Hepatol.*
312 2013;5(8):433-8
- 313 **9.** Konings CJ, Kooman JP, Schonck M, van Kreel B, Heidendal GA, Cheriex
314 EC, van der Sande FM, Leunissen KM.. Influence of fluid status on
315 techniques used to assess body composition in peritoneal dialysis patients.
316 *Perit Dial Int.* 2003;23(2):184-90
- 317 **10.** Panorchan K, Nongnuch A, El-Kateb S, Goodlad C, Davenport A. Changes in
318 muscle and fat mass in haemodialysis patients detected by multifrequency
319 bioelectrical impedance analysis. *Eur J Clin Nutr.* 2015;69(10):1109-12
- 320 **11.** Chamney PW, Krämer M, Rode C, Kleinekofort W, Wizemann V. A new
321 technique for establishing dry weight in haemodialysis patients via whole
322 body bioimpedance. *Kidney Int.* 2002;61(6):2250-8
- 323 **12.** Vernon K, Peasegood J, Riddell A, Davenport A. Dialyzers designed to
324 increase internal filtration do not result in significantly increased platelet
325 activation and thrombin generation. *Nephron Clin Pract.* 2011;117(4):c403-
326 8
- 327 **13.** Davenport A. Low-molecular-weight heparin as an alternative
328 anticoagulant to unfractionated heparin for routine outpatient
329 haemodialysis treatments. *Nephrology (Carlton).* 2009;14(5):455-61
- 330 **14.** Sandhu E, Crawford C, Davenport A. Weight gains and increased blood
331 pressure in outpatient haemodialysis patients due to change in acid
332 dialysate concentrate supplier. *Int J Artif Organs.* 2012;35(9):642-7
- 333 **15.** Davies SJ, Davenport A. The role of bioimpedance and biomarkers in
334 helping to aid clinical decision-making of volume assessments in dialysis
335 patients. *Kidney Int.* 2014;86(3):489-96
- 336 **16.** Raimann JG, Zhu F, Wang J, Thijssen S, Kuhlmann MK, Kotanko P, et al .
337 Comparison of fluid volume estimates in chronic haemodialysis patients by
338 bioimpedance, direct isotopic, and dilution methods. *Kidney Int.*
339 2014;85(4):898-908
- 340 **17.** Chua HR, Xiang L, Chow PY, Xu H, Shen L, Lee E, et al. Quantifying acute
341 changes in volume and nutritional status during haemodialysis using
342 bioimpedance analysis. *Nephrology (Carlton).* 2012;17(8):695-702

- 343 **18.** El-Kateb S, Davenport A. Changes in Intracellular Water Following
344 Hemodialysis Treatment Lead to Changes in Estimates of Lean Tissue
345 Using Bioimpedance Spectroscopy. *Nutr Clin Pract.* 2015 Dec 18. pii:
346 0884533615621549 PMID: 26684440
- 347 **19.** Dehghan M, Merchant AT. Is bioelectrical impedance accurate for use in
348 large epidemiological studies? *Nutr J.* 2008;7:26. doi: 10.1186/1475-2891-
349 7-26 PMID: 18778488
- 350 **20.** Kaysen GA1, Zhu F, Sarkar S, Heymsfield SB, Wong J, Kaitwatcharachai
351 C, Kuhlmann MK, Levin NW. Estimation of total-body and limb muscle mass
352 in haemodialysis patients by using multifrequency bioimpedance
353 spectroscopy1,2,3. *Am J Clin Nutr.* 2005;82(5):988-95
- 354 **21.** Dahlmann A, Dörfelt K, Eicher F, Linz P, Kopp C, Mössinger I, Horn S,
355 Büschges-Seraphin B, Wabel P, Hammon M, Cavallaro A, Eckardt KU,
356 Kotanko P, Levin NW, Johannes B, Uder M, Luft FC, Müller DN, Titze JM.
357 Magnetic resonance-determined sodium removal from tissue stores in
358 haemodialysis patients. *Kidney Int.* 2015;87(2):434-41
- 359 **22.** Abbas SR, Zhu F, Kaysen GA, Kotanko P, Levin NW. Effect of change in
360 fluid distribution in segments in hemodialysis patients at different
361 ultrafiltration rates on accuracy of whole body bioimpedance
362 measurement. *J Appl Physiol (1985).* 2014;116(11):1382-9
- 363 **23.** Daugirdas JT, Depner TA, Gotch FA, Greene T, Keshaviah P, Levin NW,
364 Schulman G. Comparison of methods to predict equilibrated Kt/V in the
365 HEMO Pilot Study. *Kidney Int.* 1997;52(5):1395-405
- 366 **24.** El-Kateb S, Davenport A. Changes in hydration following haemodialysis
367 estimated with bioimpedance spectroscopy. *Nephrology.* 2016; doi:
368 10.1111/nep.12645 PMID: 26436338
- 369 **25.** Kaysen GA1, Zhu F, Sarkar S, Heymsfield SB, Wong J, Kaitwatcharachai
370 C, Kuhlmann MK, Levin NW. Estimation of total-body and limb muscle mass
371 in haemodialysis patients by using multifrequency bioimpedance
372 spectroscopy1,2,3. *Am J Clin Nutr.* 2005;82(5):988-95
- 373 **26.** Gibson AL, Beam JR, Alencar MK, Zuhl MN, Mermier CM. Time course of
374 supine and standing shifts in total body, intracellular and extracellular
375 water for a sample of healthy adults. *Eur J Clin Nutr.* 2015;69(1):14-9
- 376 **27.** Papakrivopoulou E, Booth J, Pinney J, Davenport A. Comparison of volume
377 status in asymptomatic haemodialysis and peritoneal dialysis outpatients.
378 *Nephron Extra.* 2012 ;2(1):48-54
- 379 **28.** Kumar S, Khosravi M, Massart A, Potluri M, Davenport A. Haemo-
380 diafiltration results in similar changes in intracellular water and
381 extracellular water compared to cooled haemodialysis. *Am J Nephrol.*
382 2013;37(4):320-4
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Figure 1. Bland Altman analysis comparing pre haemodialysis intracellular water (ICW) and post haemodialysis ICW in litres. Mean bias -0.23, 95% limits of agreement -1.16 to 0.71 L.

Figure 2. Bland Altman analysis comparing pre haemodialysis extracellular water (ECW) and post haemodialysis ECW in litres. Mean bias 0.83, 95% limits of agreement -0.39 to 2.83 L.

Figure 3. Change in intracellular water (ECW) pre and post-dialysis, comparing with the change muscle mass. Pearson correlation.

Figure 4. Change in fat free mass and fat mass following a haemodialysis session. Pearson correlation.

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