

1 **Acoustic identification of Mexican bats based on taxonomic and ecological**
2 **constraints on call design**

3 Veronica Zamora-Gutierrez^{1,2*}, Celia Lopez-Gonzalez³, M. Cristina MacSwiney Gonzalez⁴,
4 Brock Fenton⁵, Gareth Jones⁶, Elisabeth K.V. Kalko^{7,8}, Sebastien J. Puechmaille^{9,10},
5 Vassilios Stathopoulos¹¹ and Kate E. Jones^{2,12*}

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7 ¹Conservation Science Group, Department of Zoology, University of Cambridge, Downing
8 Street, Cambridge, CB2 3EJ, United Kingdom.

9 ²Centre for Biodiversity and Environment Research, Department of Genetics, Evolution and
10 Environment, University College London, Gower Street, London, WC1E 6BT, United
11 Kingdom.

12 ³Centro Interdisciplinario de Investigación para el Desarrollo Integral Regional (CIIDIR)
13 Unidad Durango, Instituto Politécnico Nacional, Calle Sigma 119, Fraccionamiento 20 de
14 Noviembre II, Durango, Durango, 34220, Mexico.

15 ⁴Centro de Investigaciones Tropicales, Universidad Veracruzana, Casco de la ExHacienda
16 Lucas Martin, Privada de Araucarias Col. Periodistas, Xalapa, Veracruz, 91019, Mexico.

17 ⁵Department of Biology, Western University, London, Ontario, N6A 5B7, Canada.

18 ⁶School of Biological Sciences, University of Bristol, 24 Tyndall Avenue, Bristol, BS8 1TQ,
19 United Kingdom.

20 ⁷Institute of Experimental Ecology, University of Ulm, Albert-Einstein-Allee 11, 89069 Ulm,
21 Germany.

22 ⁸Smithsonian Tropical Research Institute, Balboa, Panama.

23 ⁹Zoology Institute, Ernst-Moritz-Arndt University, Greifswald, D-17489, Germany.

24 ¹⁰School of Biology and Environmental Science, University College Dublin, Dublin 4, Ireland.

25 ¹¹Department of Statistics, University of Warwick, Coventry, CV4 7AL, United Kingdom.

26 ¹²Institute of Zoology, Zoological Society of London, Regent's Park, London, NW1 4RY,
27 United Kingdom.

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29 * Corresponding authors: zamora.gtz@gmail (Tel: +44 (0)1223 336600, Fax: +44 (0)1223
30 336676) and kate.e.jones@ucl.ac.uk (Tel: +44 (0)20 31084230)

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39 SUMMARY

- 40 1. Monitoring global biodiversity is critical for understanding responses to anthropogenic
41 change, but biodiversity monitoring is often biased away from tropical, megadiverse
42 areas that are experiencing more rapid environmental change. Acoustic surveys are
43 increasingly used to monitor biodiversity change, especially for bats as they are
44 important indicator species and most use sound to detect, localise and classify
45 objects. However, using bat acoustic surveys for monitoring poses several
46 challenges, particularly in mega-diverse regions. Many species lack reference
47 recordings, some species have high call similarity or differ in call detectability, and
48 quantitative classification tools, such as machine learning algorithms, have rarely
49 been applied to data from these areas.
- 50 2. Here, we collate a reference call library for bat species that occur in a megadiverse
51 country, Mexico. We use 4,685 search-phase calls from 1,378 individual sequences
52 of 59 bat species to create automatic species identification tools generated by
53 machine learning algorithms (Random Forest). We evaluate the improvement in
54 species-level classification rates gained by using hierarchical classifications,
55 reflecting either taxonomic or ecological constraints (guilds) on call design, and
56 examine how classification rate accuracy changes at different hierarchical levels
57 (family, genus, and guild).
- 58 3. Species-level classification of calls had a mean accuracy of 66% and the use of
59 hierarchies improved mean species-level classification accuracy by up to 6%
60 (species within families 72%, species within genera 71.2% and species within guilds
61 69.1%). Classification accuracy to family, genus and guild-level was 91.7%, 77.8%
62 and 82.5%, respectively.
- 63 4. The bioacoustic identification tools we have developed are accurate for rapid
64 biodiversity assessments in a megadiverse region and can also be used effectively to
65 classify species at broader taxonomic or ecological levels. This flexibility increases

66 their usefulness when there are incomplete species reference recordings and also
67 offers the opportunity to characterise and track changes in bat community structure.
68 Our results show that bat bioacoustic surveys in megadiverse countries have more
69 potential than previously thought to monitor biodiversity changes and can be used to
70 direct further developments of bioacoustic monitoring programs in Mexico.

71 *Keywords:* acoustic identification, guild, hierarchical classification, random forest, machine
72 learning, Neotropical, whispering bats.

73

74 INTRODUCTION

75 Effective conservation depends on our ability to define, measure and track ecological
76 communities through time and space (Magurran *et al.* 2010). Although biodiversity
77 monitoring programmes are critical to assess the impact of anthropogenic change, many are
78 biased towards high latitude, temperate countries (Collen *et al.* 2009). Megadiverse
79 countries (e.g., Indonesia, Mexico, Zaire) cover only 34% of the Earth surface, yet they
80 harbour 70% of the world's biodiversity and are undergoing rapid environmental degradation
81 (Mittermeier *et al.* 1997). In spite of the great conservation opportunity these hotspot regions
82 offer, biodiversity monitoring programmes are often lacking, causing considerable knowledge
83 gaps.

84 Bioacoustic surveys, especially for bats, are increasingly used to survey and monitor
85 biodiversity responses to anthropogenic change (Jones *et al.* 2013; Amorim *et al.* 2014).
86 Echolocating bats use sound to detect, localise and classify objects (Schnitzler *et al.* 2003)
87 making them detectable both remotely and non-invasively. Bats are also ideal biodiversity
88 indicators since they have a wide range of ecological traits, different tolerances to
89 environmental variables and play key roles in ecosystems (Jones *et al.* 2009; Russo & Jones
90 2015). However, using bat acoustics as a monitoring tool poses several challenges,
91 especially in megadiverse and tropical regions (Walters *et al.* 2013). First, in spite of the
92 growing efforts to create more bat call reference recording libraries, tropical and
93 megadiverse regions have rarely been included in such initiatives. This is compounded by
94 recording method heterogeneity (e.g., full spectrum, frequency division, heterodyne), which
95 makes compiling comprehensive libraries difficult (Walters *et al.* 2013). Such poor and
96 uneven coverage of intra- and inter-specific variation makes Identification of bat calls for
97 these regions challenging

98 Second, although it is possible to identify many bat species based on their calls,
99 phylogenetic relatedness, ecological similarities, and call plasticity have led to overlapping

100 structures and high call similarity among and within species in some groups (Obrist 1995;
101 Jones & Teeling 2006). For example, species may have similar calls within families and
102 genera (Jung *et al.* 2007, 2014), and ecological guild membership may also reflect foraging
103 and echolocation behaviour (e.g., aerial insectivores, gleaners) (Denzinger & Schnitzler
104 2013). An additional challenge is that bat species differ in detectability of their calls. Aerial
105 insectivores typically produce loud calls of high-intensity and low frequency, whereas
106 'whispering' bats (including many bats in the families Phyllostomidae, Natalidae, and
107 Thyropteridae) often produce low-intensity, high frequency calls (Griffin 1958). However,
108 recent findings suggest that some 'whispering' bat calls are more detectable than previously
109 thought. For example, *Macrophyllum macrophyllum* and *Artibeus jamaicensis* can emit calls
110 as loud as those of many aerial insectivores (Brinkløv *et al.* 2009). *Otonycteris hemprichii*, a
111 passive gleaner, can also operate as an aerial hawk and can adjust its call intensity
112 depending on foraging mode even while flying in the same habitat type (Hackett *et al.* 2014).

113 Third, although acoustic species identification tools for different species are developing
114 rapidly (e.g., European bats Walters *et al.* 2012; birds Stowell & Plumbley 2014), they
115 remain rare for megadiverse regions. The immense amount of data obtained from acoustic
116 monitoring can be daunting and automatic analytical tools are extremely useful in analysing
117 such data (Walters *et al.* 2013). Bat call identification tools have been mainly developed
118 using multivariate statistical techniques such as discriminant function analysis (e.g.,
119 Vaughan *et al.* 1997; Russo & Jones 2002; Avila-Flores & Fenton 2005; MacSwiney *et al.*
120 2008) or machine learning algorithms (e.g., Skowronski & Harris 2006, Walters *et al.* 2012),
121 the latter generally providing higher species-level classification accuracy (Armitage & Ober
122 2010; Britzke *et al.* 2011; Keen *et al.* 2014). Machine learning algorithms have mostly been
123 applied to classify data at one level of categorisation (e.g., species) and have rarely
124 incorporated hierarchical information to aid classification accuracy (e.g., species within
125 families or orders). Hierarchical classification approaches have been shown to improve
126 general species classification accuracy for European bat calls up to 13% (Parsons & Jones

127 2000; Walters *et al.* 2012). Assigning taxa to classes within a hierarchy may reduce model
128 complexity and minimise misclassifications outside their hierarchy (Vens *et al.* 2008).
129 However, if an erroneous hierarchy is applied, then classification errors are added
130 cumulatively across different levels, leading to a reduction in classification accuracy.

131 A hierarchical classification approach may be useful to classify calls to broader classes (e.g.,
132 genera, families or guilds) when reference material is missing for species, or where
133 discrimination at species-level is difficult. For example, where there is high call variability
134 within species, or a high overlap of call parameters between species. Although identification
135 to species is most desirable, monitoring the status of the same recognizable signal over time
136 without specific identification may be sufficient in some situations (Redgwell *et al.* 2009;
137 Armitage & Ober 2010). Finding alternatives to species-level studies is needed in
138 megadiverse areas, which usually face considerable financial and data constraints but are a
139 priority for rapid conservation assessments.

140 Here, we collate a reference call library for bat species that occur in a megadiverse country
141 to create acoustic identification tools using machine learning algorithms. We focus on
142 Mexico because it contains one of the highest number of species in the world and has one of
143 the highest rates of species extinction and habitat loss (Myers *et al.* 2000; Brooks *et al.*
144 2002). We also evaluate the improvement in species-level classification rates gained by
145 using hierarchical classifications reflecting either taxonomic or ecological constraints on call
146 design. Our results show that accurate bioacoustic identification tools can be developed for
147 rapid biodiversity assessments in megadiverse regions where hierarchies generally improve
148 species-level classifications. These tools can also be used effectively to classify calls at
149 broader levels, so increasing the usefulness of the tool when there are incomplete species
150 reference recordings.

151

152 MATERIALS AND METHODS

153 Reference call library

154 We collated reference search-phase echolocation calls for bat species that occur in Mexico
155 through a combination of field work and donated material. Field work was conducted in
156 central and northern Mexico from June 2012 to May 2013 at 35 sites (Fig. 1a). Bats were
157 caught with mist nets and identified to species level using field keys (Reid 1997; Medellín *et*
158 *al.* 2008), before being released. Full spectrum, real time recordings were made from all
159 individuals in the habitat in which they were captured using a Pettersson D1000x detector,
160 sampling rate 500kHz, high pass filter off (Pettersson Elektronik AB, Uppsala, Sweden).
161 Files were saved in WAV format on a flash card. We obtained 907 recordings of 39 species
162 from 6 families (see Table S1 in Supporting Information). Additionally, 1,403 full spectrum
163 recordings of bat calls from 87 species that occur in Mexico were donated by colleagues,
164 giving a total of 2,310 recordings (each recording was assumed to contain one individual call
165 sequence) from 92 species in 8 families (68% of species and 100% of families of bats
166 occurring in Mexico). These recordings were obtained from bats released in different ways
167 using several different real time or time-expanded full spectrum detectors, and in a range of
168 habitats across species' distributions (including localities outside Mexico) (Table S2). The
169 inclusion of call variation in the dataset avoids generating biases for any particular recording
170 situation or method (Walters *et al.* 2013), and provides the acoustic identification tools with
171 more flexibility and generality (see Walters *et al.* 2012).

172 Taxonomy followed Simmons (2005), but because of taxonomic changes since 2005 we
173 assume that Natalidae contains only one species, *Natalus stramineus* (López-Wilchis *et al.*
174 2012). Data from *Molossus sinaloae* and the new species *M. alvarezi* (González-Ruiz *et al.*
175 2011) were analysed together as *M. sinaloae* because most of the material was recorded
176 prior to the description of the new species. As some species are hard to identify in the field,
177 we only used the material which were confidently identified. To examine the taxonomic and
178 geographic coverage of the reference call library within Mexico, distribution maps were
179 downloaded from the IUCN mammal assessments (IUCN 2012) and species richness within

180 each 50 km² grid cell was estimated by overlaying and counting how many of those range
181 maps overlap in each grid cell (Hawths Tools, Beyer, 2004). We then calculated the
182 proportion of species both recorded and used in our classifiers from out of those potentially
183 distributed in each cell.

184 Acoustic Identification Tools

185 We visually inspected all recorded sequences using the sound analysis software BatSound
186 Pro v.3.31b (Pettersson Elektronik AB, Uppsala Sweden) to remove non search-phase calls.
187 We distinguished search-phase calls from approach-phase and terminal-phase calls as
188 these phase shifts are characterized by a decrease in call duration and interval, and
189 increase in repetition rate (Schnitzler & Kalko 2001). Social calls were distinguished from
190 echolocation calls by their duration, frequency and pattern of change over time, with social
191 calls being more sporadic and often of a lower frequency range (Fenton 2003). In addition,
192 bats were recorded in situations that significantly minimized the presence of social calls and
193 approach and end-phase echolocation calls (e.g., recorded in open spaces upon release).
194 We then automatically extracted and parameterised search-phase calls using the in-built
195 algorithms in Sonobat v.3 (Szewczak 2010) (following methods in Walters *et al.* 2012). For
196 species which used harmonics, we used measurements from the call used as the main
197 harmonic. We measured a total of 21,064 search-phase echolocation calls from 1,692
198 sequences and 85 species in 8 families, with each sequence assumed to be from a different
199 individual. Material recorded in Mexico contained 16,344 calls, 1,187 sequences from 65
200 species in 7 families across 91 different localities (Fig. 1a).

201 We used Random Forest (RF) models (randomForest package, Liaw & Wiener 2002) to train
202 the classifiers, rejecting species that had less than five sequences. RF models consist of a
203 collection or ensemble of decision tree classifiers where each classifier is randomly built
204 using a bootstrapped sample of the training dataset (Breiman 2001). Each classifier is
205 estimated based on probabilities using a selection of the predictor variables (in our case call
206 parameters) that best separate the classes of interest (e.g., species, families) at different

207 branching splits or nodes in the tree. RF model classifications are then derived from
208 averages of the tree ensembles. RF models possess several advantages over other
209 machine learning algorithms as they are not affected by heteroscedasticity or distributional
210 errors in the data, are not sensitive to outliers or irrelevant variables, can deal with mixed
211 data and missing variables, and are relatively simple to train using reasonable computational
212 resources (Olden *et al.* 2008). We selected 27 of the relevant call parameter variables
213 (following methods in Walters *et al.* 2012) extracted and parameterised by Sonobat (Table
214 S3), and ran a grid search to find the mtry value (optimal number of variables to be randomly
215 sampled at each node). This value was allowed to range from 2-10, in steps of one. Each
216 forest was grown to 2,000 trees and the final mtry value and number of trees were selected
217 for their highest accuracy. The final set of parameters used was 1,000 trees and an mtry
218 value of three. We used the coefficient of the Gini impurity index (used by the RF models to
219 select the most informative variables at nodes during training), as an indicator of call
220 parameter variable importance (Breiman 2001).

221 We trained four different RF model classifiers: Classifier 1 - Species-level without a
222 hierarchy; Classifier 2 - Species-level within a family hierarchy (see call examples in Fig.
223 S1a-f); Classifier 3 - Species-level within a genus hierarchy (see call examples in Fig. S1g);
224 and Classifier 4 - Species-level within a guild hierarchy, following definitions of guilds from
225 Denzinger & Schnitzler (2013) (see call examples in Fig. S2a-e): Guild 1 represented Open
226 space aerial foragers; Guild 2 - Edge space aerial foragers; Guild 3 - Edge space trawling
227 foragers; Guild 4 - Narrow space flutter detecting foragers; and Guild 5 - Narrow space
228 passive gleaning foragers and Guild 6 - Narrow space passive/active gleaning foragers.
229 Guild 7 - Narrow space active gleaning foragers was not included in the study because of
230 the lack of reference material.

231 We used five-fold cross-validation to assess the accuracy of all four RF classifiers and
232 assigned the individual calls into the five folds by sequence rather than individual calls
233 (Stathopoulos *et al.* 2014). This procedure ensured that calls from the same individual (i.e.,

234 sequence) were not used in the same training and testing run of the cross-validation to avoid
235 over-fitting. We set a maximum of 100 calls per species for Classifier 1 and a minimum of 20
236 calls per species for Classifiers 2, 3 and 4, as a compromise between maximising the
237 number of calls and balancing the datasets, since RF classifiers tend to be biased towards
238 the majority class (species, genus, family or guild with the highest number of training calls)
239 (Chen *et al.* 2004). Only the highest quality calls were selected from each sequence
240 (determined by the signal to noise ratio given by Sonobat), until the selected number of calls
241 was reached. However, for some species with smaller sample sizes, we continued selecting
242 calls from sequences in descending order of quality until we had used all available data or
243 reached the number of calls allowed (Table S4). The number of calls selected per sequence
244 was a compromise between maximising the number of calls and avoiding over-fitting the RF
245 models. Sample sizes after this selection process were 4,685 calls and 1,378 sequences
246 from 8 families, 32 genera, and 59 species that occur in Mexico. See Figure S3 for an outline
247 of the analytical procedure.

248 As we used recordings from locations from both inside and outside of Mexico, we checked
249 that the variation in call parameters recorded in locations outside of Mexico did not impact
250 species classification accuracy. To investigate this, we compared model accuracy using the
251 four classifiers of two datasets consisting of 47 species recorded from locations inside
252 Mexico and the same species recorded from all locations. We found very little difference in
253 classification accuracy between the two datasets. Classifier 1 had the biggest difference in
254 classification accuracy, albeit with only 1.5% reduction in correct classification rates (67.1%
255 and 65.6% for inside Mexico and for all locations, respectively). We therefore used
256 recordings from outside Mexico to complement species with less than five Mexican
257 sequences. All analyses were performed in R version 3.0.2 (R Development Core Team
258 2013).

259

260 RESULTS

261 Database coverage

262 Our collated library of echolocation call recordings covered 69% of the species, 79% of the
263 genera, and 100% of the families occurring in Mexico. Data of high enough quality to build
264 the automatic identification tools covered 43% of the species, 51% of the genera, and 100%
265 of the families (Table S5). There was generally a good representation of species for the
266 identification tools within genera and families (>50%), except for Phyllostomidae, where only
267 19% of the species were represented. Species coverage was more comprehensive within
268 the central and northern parts of Mexico for both the library and identification tools (Fig. 1b-
269 c).

270

271 Acoustic Identification Tools

272 Overall 16 out of the 27 parameters used to train the models contributed most to all classifiers
273 (based on a score >30 for the Gini Coefficient from the RF models) (Table S6, Fig 2a-d).
274 Although different parameters were important for each hierarchy, the most important overall
275 were Fc Characteristic call frequency (kHz), Fctr Frequency at the center of the call duration
276 (kHz), FLed Frequency of the ledge (kHz), StartF Frequency at the start of a call (kHz),
277 HFreq Highest call frequency (kHz), and FMPwr Frequency of the maximum call amplitude
278 (kHz) (Fig. 2a-d, see Table S3 for further variable definitions).

279

280 Overall mean species-level classification accuracies for Mexican bat species varied across
281 the four classifiers between 66.0% (Classifier 1: Species-level without a hierarchy) and
282 72.0% (Classifier 2: Species-level within a family hierarchy), with Classifiers 3 (Species-level
283 with a genus hierarchy) and 4 (Species-level with a guild hierarchy) having accuracies of
284 71.2% and 69.1%, respectively (Table 1). Across all classifiers, on average the highest
285 classification accuracies were found for species within families Natalidae (100%),
286 Mormoopidae (94.6%), Thyropteridae (81.5%), and Emballonuridae (77.7%), with the lowest
287 found within Noctilionidae (70.4%), Molossidae (67%), Vespertilionidae (51.5%), and
288 Phyllostomidae (51.4%) (Fig. 3). Phyllostomid species were mostly misclassified with other

289 phyllostomids or with vespertilionids, whereas vespertilionids were commonly misclassified
290 with other vespertilionids or with molossids (Table 1). For the ecological classifier, species
291 within Guild 4 (narrow space flutter detecting foragers) (100%), Guild 3 (edge space trawling
292 foragers) (74.6%), and Guild 1 (open space aerial foragers) (63.8%) had on average the
293 highest classification rates. The lowest average classification rates for species were found
294 within in the gleaners (Guild 5 58.5% and Guild 6 57.7%) Guild 2 (edge space aerial
295 foragers) (54.5%) (Fig. 4).

296

297 Classification accuracy at different hierarchical levels was highest at family-level with a mean
298 of 91.7% across all families (Table 1, Fig. 3), where Natalidae and Mormoopidae had the
299 highest classification accuracies (100% and 97.3%, respectively). Noctilionidae had the
300 lowest classification accuracy (72.8%) and was frequently misclassified as Molossidae (17%
301 of the calls). Genus-level mean classification accuracy was 77.8% across all genera (Table
302 1), *Natalus* (Natalidae) and *Rhynchonycteris* (Emballonuridae) had the highest classification
303 accuracies (100%), and 18 genera had accuracies >80% (Fig. 3). The genus *Myotis* yielded
304 a classification accuracy of 73.8%, with two species over >80% (*Myotis thysanodes* and
305 *Myotis keaysi*) and only 4 with <50%. Genera with the lowest classification accuracies
306 (<50%) were in the Phyllostomidae and Vespertilionidae (Fig. 3). Phyllostomids were mostly
307 misclassified as other phyllostomids, while vespertilionids were misclassified as other
308 vespertilionids and molossids. Mean guild-level classification accuracy was 82.5% across all
309 guilds (Table 1, Fig. 4). Guild 4 (narrow space flutter detecting foragers) had the highest
310 classification accuracy (100%), followed by Guild 6 (88.3%) although 6% of these calls were
311 misclassified with Guild 5. Guild 5 had the worst classification accuracy (68%), and 18% of
312 calls were misclassified as Guild 6 (Fig. 4).

313

314 DISCUSSION

315 We have collated the most extensive bat acoustic library for a megadiverse region (all
316 families and over half of the species occurring in Mexico) and developed the most
317 comprehensive bat acoustic automated species-level classifiers to date. The mean species-
318 level classification accuracy rate of 66-72% (depending on which hierarchy is chosen) is
319 reasonable given the high level of call similarity of the bat species in this area (Walters *et al.*
320 2013). The species-level classifiers also contain a large variation in accuracy rates, where
321 some species are classified to >80% accuracy (species of Emballonuridae, Mormoopidae,
322 Natalidae and Thyropteridae), with the poorest results overall from species of
323 Vespertilionidae and Phyllostomidae. This suggests that acoustic monitoring may be more
324 feasible focusing on a few species whose calls can be reliably classified.

325 The bat call library and classifiers incorporate both extensive geographic (from 9 countries
326 within the species range of Mexican bats) and intra-specific variation in call types (e.g., the
327 classifiers were trained on the different search-phase echolocation call types found within
328 molossid species, Jung *et al.* 2014). However, the species-level classifiers have a very low
329 coverage of Phyllostomidae and results should be interpreted with caution. It has been
330 traditionally assumed that whispering bats, which include all phyllostomids, echolocate at
331 intensities that were too low for the inclusion of these species in acoustic studies. However,
332 recent field studies of their echolocation behaviour challenged these assumptions about their
333 echolocation characteristics (Brinkløv *et al.* 2009; Hackett *et al.* 2014). Future work should
334 focus on collecting more reference material for the family, to better assess its potential for
335 acoustic monitoring programmes.

336 Our classifiers will be the most accurate in regions where there is a higher coverage of the
337 species present, such as the less species-rich arid and semi-arid regions of Mexico. These
338 ecosystems (e.g., xerophytic scrubland and grasslands) cover at least 40% of the territory
339 (Rzedowski 2006), and together with other North American drylands, support some of the
340 biggest concentrations of mammalian abundance, because bats can form colonies of several
341 millions of individuals (O'Shea & Bogan 2003). These bat populations can provide important

342 ecosystem services such as pollination and control of insect populations (Cleveland *et al.*
343 2006; Munguía-Rosas *et al.* 2009). These important arid and semi-arid environments are
344 increasingly threatened by environmental changes (Villers-Ruiz & Trejo-Vázquez 2003;
345 Rodríguez-Estrella 2007) and future efforts should focus on these arid areas where there are
346 considerable information gaps.

347

348 Our species-level classifier mean accuracy was similar to that of previous studies of bats for
349 species shared with this study (Mexico - MacSwiney *et al.* 2008; Stathopoulos *et al.* 2014;
350 West Indies - Pio *et al.* 2010; United States - Skowronski & Harris 2006; Britzke *et al.* 2011).
351 However, our classification accuracies were slightly lower for some species compared with
352 previous work. This is a consequence of the higher number of classes (species) included in
353 our classifiers compared to all previous studies. Higher numbers of species increases the
354 similarity in the call parameters of several species. For example, we included 26
355 vespertilionids and 8 *Myotis* species, compared to 6 vespertilionids and one *Myotis* in
356 MacSwiney *et al.* (2008) and 9 vespertilionids and two *Myotis* in Stathopoulos *et al.* (2014).
357 Our study nearly triples the number of species used compared to any other quantitative bat
358 call classification study in the Americas or any other megadiverse tropical region in the
359 world. We also included a wide range of ecological, technological and methodological
360 variation in the training dataset, which on one hand increases the classification challenge,
361 but on the other makes the classifiers more robust to real-world recording situations. In spite
362 of the great difference in the number of species used here, we also obtained higher
363 classification accuracies to species level and better mean accuracies than previous studies
364 (e.g., Pio *et al.* 2010; Stathopoulos *et al.* 2014).

365

366 Our use of taxonomic and ecological guild hierarchies improved mean species-level
367 classification rates. By using hierarchical classification approaches the number of final
368 classes is considerably reduced and misclassifications are limited to classes within the
369 respective hierarchy (Vens *et al.* 2008). Mean species-level classification accuracies were

370 most improved using a family hierarchy, closely followed by genera (72% and 71%,
371 respectively), although not all species improved their accuracies (contrary to other studies -
372 Parsons & Jones 2000; Walters *et al.* 2012). The genus-level hierarchy produced the highest
373 number of species-level classifications with >80% accuracy but for many genera not all
374 species were included in the analysis and genus-level taxonomic names can be subject to
375 rapid changes (Simmons 2005). This may suggest that using a genus-level hierarchy may
376 be more problematic than a family hierarchy, especially with incomplete reference material.
377 For example, accuracy may decrease as more species are included, whereas variation
378 within a family may be already adequately represented. In contrast, classification to genus
379 level may be more helpful to reduce the number of options of possible misclassifications
380 inside the hierarchy and further methods for call identification could then be applied (e.g.,
381 visual inspection).

382 Although we found species-level classification rates within an ecological guild-level hierarchy
383 were worse than species-level classification rates within either taxonomic hierarchy,
384 classification of calls to guild-level performed well and could provide a useful alternative to
385 taxonomic-level classifiers. Gleaners, in particular the speciose family Phyllostomidae, are
386 the most abundant and diverse in bat communities in the Americas, yet poorly represented
387 in acoustic libraries. Our results at family and guild-level suggest that there is a good
388 potential for accurate acoustic identification of gleaners. As more sensitive microphones with
389 better signal/noise ratios become available, the detectability of these species will improve, as
390 will the potential for monitoring them acoustically. Guilds 5 and 6, representing gleaning
391 foragers, were frequently confused with each other, so these should be grouped into one
392 class, since the main difference among them is how they use other non-echolocation cues to
393 forage (Denzinger & Schnitzler 2013).

394 Acoustic analysis techniques are evolving rapidly and there is a growing tendency to replace
395 classifications based on parameters extraction with those of whole signal analysis. However,
396 applications of these approaches have mainly focused on bird and marine mammal

397 acoustics (e.g., Ren *et al.* 2009; Damoulas *et al.* 2010) and most bat acoustic classification
398 tasks still represent classifications with a few parameters and further classify them using
399 manual or non-parametric techniques. Such whole signal analyses in bat acoustics are
400 growing (Obrist *et al.* 2004; Skowronski & Harris 2006; Stathopoulos *et al.* 2014) but should
401 be further explored. However, exploration of new approaches requires adequate reference
402 material collected in a systematic way, controlling for variation introduced by the use of
403 different methods, and we strongly encourage further efforts to collect comprehensive
404 reference bat call libraries.

405

406 Applications

407 Standardized identification tools such as these, offer the opportunity for objective and
408 repeatable identifications of monitoring 'units' to identify changes in populations, distributions
409 or community structures through time and space. Furthermore, hierarchical approaches offer
410 the flexibility to adapt the identification tools to the purpose of the study or monitoring
411 programme and the geographic and taxonomic coverage of the reference material available.
412 Although the accuracy reached for some groups might not be sufficient for studies targeting
413 their particular species (e.g., *Myotis* spp.), the hierarchical classifiers can act as filters for
414 large amounts of data. The use of hierarchies considerably reduces the list of species to
415 which an unknown call could belong, thus making detailed inspections and further
416 validations more feasible.

417 Hierarchical classifications, in particular at family-level, could help reduce the costs of
418 monitoring tropical bat communities, which is crucial due to the limited funding these regions
419 often devote to conservation efforts. Despite the relatively poor classification accuracies to
420 species-level within the guild-level hierarchy, classification to guild-level could be used to
421 rapidly characterize ensemble/environment associations or to track changes in community
422 structure. The hierarchical approach may be improved through the use of regional classifiers
423 which allow the reduction of the number of classes and the improvement of classification

424 accuracy. However, such an approach should be used with caution as least known species
425 or those with expanding ranges could be ignored.

426

427 CONCLUSIONS

428 Our study shows that there is more potential for bat acoustic monitoring in megadiverse
429 countries than previously considered. Hierarchies considerably reduced the complexity of
430 call identification at different levels and provided sufficient confidence in the classification of
431 unknown calls into higher taxonomic levels and ecological guilds. While the classifiers did
432 not provide high classification accuracies for several species, they did offer the opportunity
433 to have objective and repeatable identification of monitoring 'units' to implement in national
434 acoustic monitoring programmes.

435

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444 permit was granted by SEMARNAT, Mexico (No. 03374).

445

446 DATA ACCESSIBILITY

447 Complete call measurements for each of the 4,685 search-phase calls from 1,378 individual
448 sequences of 59 bat species used to create the classification tools and R scripts have been
449 uploaded to DataDyrad (doi:10.5061/dryad.760r8).

450

451 LITERATURE CITED

- 452 Amorim, F., Carvalho, S.B., Honrado, J. & Rebelo, H. (2014). Designing optimized multi-
453 species monitoring networks to detect range shifts driven by climate change: a case
454 study with bats in the north of Portugal. *PLoS ONE*, **9**, e87291.
- 455 Armitage, D.W. & Ober, H.K. (2010). A comparison of supervised learning techniques in the
456 classification of bat echolocation calls. *Ecological Informatics*, **5**, 465–473.
- 457 Avila-Flores, R. & Fenton, M.B. (2005). Use of spatial features by foraging insectivorous bats
458 in a large urban landscape. *Journal of Mammalogy*, **86**, 1193–1204.
- 459 Beyer, H.L. (2004). Hawth's analysis tools for ArcGIS. URL
460 <http://www.spatalecolgy.com/htools> [accessed 1 July 2015]
- 461 Breiman, L. (2001). Random forests. *Journal of Machine Learning Research*, **45**, 5–32.
- 462 Brinkløv, S., Kalko, E.K. V. & Surlykke, A. (2009). Intense echolocation calls from two
463 'whispering' bats, *Artibeus jamaicensis* and *Macrophyllum macrophyllum*
464 (Phyllostomidae). *The Journal of Experimental Biology*, **212**, 11–20.
- 465 Britzke, E.R., Duchamp, J.E., Murray, K.L., Swihart, R.K. & Robbins, L.W. (2011). Acoustic
466 identification of bats in the eastern United States: a comparison of parametric and
467 nonparametric methods. *Journal of Wildlife Management*, **75**, 660–667.
- 468 Brooks, T.M., Mittermeier, R. A., Mittermeier, C.G., da Fonseca, G. A. B., Rylands, A.B.,
469 Konstant, W.R., Flick, P., Pilgrim, J., Oldfield, S., Magin, G. & Hilton-Taylor, C. (2002).
470 Habitat loss and extinction in the hotspots of biodiversity. *Conservation Biology*, **16**,
471 909–923.
- 472 Chen C., Liaw, A. & Breiman, L. (2004). Using random forest to learn imbalanced data, Dept.
473 Statistics, Univ. California, Berkeley, CA, Tech. Rep.666.
- 474 Cleveland, C.J., Betke, M., Federico, P., Frank, J.D., Hallam, T.G., Horn, J., Jr, J.D.L.,
475 Mccracken, G.F., Medellín, R.A., Moreno-Valdez, A., Sansone, C.G., Westbrook, J.K. &
476 Kunz, T.H. (2006). Economic value of the pest control service provided by Brazilian
477 free-tailed bats in south-central Texas. *Frontiers in Ecology and the Environment*, 238–
478 243.
- 479 Collen, B., Loh, J., Whitmee, S., McRae, L., Amin, R. & Baillie, J.E.M. (2009). Monitoring
480 change in vertebrate abundance: the living planet index. *Conservation Biology*, **23**,
481 317–27.
- 482 Damoulas, T., Henry, S., Farnsworth, A., Lanzone, M. & Gomes, C. (2010). Bayesian
483 classification of flight calls with a novel dynamic time warping kernel. *Proceedings of*
484 *the 2010 Ninth International Conference on Machine Learning and Applications*, pp.
485 424–429. IEEE Computer Society, Washington, DC.
- 486 Denzinger, A. & Schnitzler, H.-U. (2013). Bat guilds, a concept to classify the highly diverse
487 foraging and echolocation behaviors of microchiropteran bats. *Frontiers in Physiology*,
488 **4**, 1–15.

- 489 Fenton, M.B. (2003). Eavesdropping on the echolocation and social calls of bats. *Mammal*
490 *Review*, **33**, 193–204.
- 491 González-Ruiz, N., Ramírez-Pulido, J. & Arroyo-Cabrales, J. (2011). A new species of
492 mastiff bat (Chiroptera: Molossidae: *Molossus*) from Mexico. *Mammalian Biology*, **76**,
493 461–469.
- 494 Griffin, D.R. (1958). *Listening in the dark*. Yale University Press, New Haven, Connecticut.
- 495 Hackett, T.D., Korine, C. & Holderied, M.W. (2014). A whispering bat that screams: bimodal
496 switch of foraging guild from gleaning to aerial hawking in the desert long-eared bat.
497 *The Journal of Experimental Biology*, **217**, 3028–3032.
- 498 IUCN. (2012). IUCN Red List of Threatened Species version 2012.14. URL
499 <http://www.iucnredlist.org> [accessed 1 July 2015]
- 500 Jones, G., Jacobs, D.S., Kunz, T.H., Willig, M.R. & Racey, P.A. (2009). Carpe noctem: the
501 importance of bats as bioindicators. *Endangered Species Research*, **8**, 93–115.
- 502 Jones, K.E., Russ, J., Bashta, A.-T., Bilhari, Z., Catto, C., Csösz, I., Gorbachev, A., Györfi,
503 P., Hughes, A., Ivashkiv, I., Koryagina, N., Kurali, A., Langton, S., Collen, A., Margiean,
504 G., Pandourski, I., Parsons, S., Prokofev, I., Szodoray-Paradi, A., Szodoray-Paradi, F.,
505 Tilova, E., Walters, C., Weatherill, A. & Zavarzin, O. (2013). Indicator Bats Program: a
506 system for the global acoustic monitoring of bats. *Biodiversity monitoring and*
507 *conservation: Bridging the gaps between global commitment and local action* (eds B.
508 Collen, N. Pettorell, J.E.M. Baillie & S. Durant). Wiley-Blackwell, London.
- 509 Jones, G. & Teeling, E.C. (2006). The evolution of echolocation in bats. *Trends in Ecology*
510 *and Evolution*, **21**, 149–156.
- 511 Jung, K., Kalko, E.K.V. & von Helversen, O. (2007). Echolocation calls in Central American
512 emballonurid bats: signal design and call frequency alternation. *Journal of Zoology*,
513 **272**, 125–137.
- 514 Jung, K., Molinari, J. & Kalko, E.K.V. (2014). Driving factors for the evolution of species-
515 specific echolocation call design in new world free-tailed bats (Molossidae). *PLoS ONE*,
516 **9**, e85279.
- 517 Keen, S., Ross, J.C., Griffiths, E.T., Lanzone, M. & Farnsworth, A. (2014). A comparison of
518 similarity-based approaches in the classification of flight calls of four species of North
519 American wood-warblers (Parulidae). *Ecological Informatics*, **21**, 25–33.
- 520 Liaw, A. & Wiener, M. (2002). Classification and regression by randomForest. *R News*, **2**,
521 18–22.
- 522 López-Wilchis, R., Guevara-Chumacero, L.M., Pérez, N.Á., Juste, J., Ibáñez, C. & Barriga-
523 Sosa, I.D.L.A. (2012). Taxonomic status assessment of the Mexican populations of
524 funnel-eared bats, genus *Natalus* (Chiroptera: Natalidae). *Acta Chiropterologica*, **14**,
525 305–316.
- 526 MacSwiney, M.C., Clarke, F.M. & Racey, P.A. (2008). What you see is not what you get: the
527 role of ultrasonic detectors in increasing inventory completeness in Neotropical bat
528 assemblages. *Journal of Applied Ecology*, **45**, 1364–1371.

- 529 Magurran, A.E., Baillie, S.R., Buckland, S.T., Dick, J.M., Elston, D.A., Scott, E.M., Smith,
530 R.I., Somerfield, P.J. & Watt, A.D. (2010). Long-term datasets in biodiversity research
531 and monitoring: assessing change in ecological communities through time. *Trends in*
532 *Ecology and Evolution*, **25**, 574–82.
- 533 Medellín, R.A., Arita, H.T. & Sánchez, O. (2008). *Identificación de los murciélagos de*
534 *México, clave de campo*, Segunda Edn. Instituto de Ecología, Universidad Nacional
535 Autónoma de México-CONABIO, Mexico D.F.
- 536 Mittermeier, R.A., Mittermeier, C.G. & Robles-Gil, P. (1997). *Megadiversity: earth's*
537 *biologically wealthiest nations*. Cemex, Mexico, D.F.
- 538 Munguía-Rosas, M.A., Sosa, V.J., Ojeda, M.M. & De-Nova, J.A. (2009). Specialization clines
539 in the pollination systems of agaves (Agavaceae) and columnar cacti (Cactaceae): a
540 phylogenetically controlled meta-analysis. *American Journal of Botany*, **96**, 1887–1895.
- 541 Myers, N., Mittermeier, R.A., da Fonseca, G.A.B. & Kent, J. (2000). Biodiversity hotspots for
542 conservation priorities. *Nature*, **403**, 853–858.
- 543 O'Shea, T.J. & Bogan, M.A. (2003). *Monitoring trends in bat populations of the United States*
544 *and territories: Problems and prospects*.
- 545 Obrist, M.K. (1995). Flexible bat echolocation: the influence of individual, habitat and
546 conspecifics on sonar signal design. *Behavioral Ecology and Sociobiology*, **36**, 207–
547 219.
- 548 Obrist, M.K., Boesch, R. & Flückiger, P.F. (2004). Variability in echolocation call design of 26
549 Swiss bat species: consequences, limits and options for automated field identification
550 with a synergetic pattern recognition approach. *Mammalia*, **68**, 307–322.
- 551 Olden, J.D., Lawler, J.J. & Poff, N.L. (2008). Machine learning methods without tears: a
552 primer for ecologists. *The Quarterly Review of Biology*, **83**, 171–193.
- 553 Parsons, S. & Jones, G. (2000). Acoustic identification of twelve species of echolocating bat
554 by discriminant function analysis and artificial neural networks. *The Journal of*
555 *Experimental Biology*, **203**, 2641–2656.
- 556 Parsons, S. & Szewczak, J.M. (2009). Detecting, recording and analyzing the vocalizations
557 of bats. *Ecological and behavioral methods for the study of bats* (eds T.H. Kunz & S.
558 Parsons), pp. 91–111. Hopkins University Press, Baltimore.
- 559 Pio, D.V.V., Clarke, F.M., MacKie, I. & Racey, P.A. (2010). Echolocation calls of the bats of
560 Trinidad, West Indies: is guild membership reflected in echolocation signal design?
561 *Acta Chiropterologica*, **12**, 217–229.
- 562 R Development Core Team. (2013). R: a language and environment for statistical
563 computing. Vienna. URL <http://www.R-project.org> [accessed 1 July 2015]
- 564 Redgwell, R.D., Szewczak, J.M., Jones, G. & Parsons, S. (2009). Classification of
565 echolocation calls from 14 species of bat by support vector machines and ensembles of
566 neural networks. *Algorithms*, **2**, 907–924.

- 567 Reid, F. (1997). *A field guide to the mammals of Central America and southeast Mexico*.
568 Oxford University Press, New York.
- 569 Ren, Y., Johnson, M.T., Clemins, P., Darre, M., Glaeser, S.S., Osiejuk, T.S. & Out-Nyarko,
570 E. (2009). A framework for bioacoustic vocalization analysis using hidden Markov
571 models. *Algorithms*, **2**, 1410–1428.
- 572 Rodríguez-Estrella, R. (2007). Land use changes affect distributional patterns of desert birds
573 in the Baja California peninsula, Mexico. *Diversity and Distributions*, **13**, 877–889.
- 574 Russo, D. & Jones, G. (2002). Identification of twenty-two bat species (Mammalia:
575 Chiroptera) from Italy by analysis of time-expanded recordings of echolocation calls.
576 *Journal of Zoology*, **258**, 91-103.
- 577 Russo, D. & Jones, G. (2015). Bats as bioindicators: an introduction. *Mammalian Biology*,
578 **80**, 157-158.
- 579 Rzedowski, J. (2006). *Vegetación de México*. CONABIO, Mexico, D.F.
- 580 Schnitzler, H.-U. & Kalko, E.K.V. (2001). Echolocation by insect-eating bats. *Bioscience*, **51**,
581 557–569.
- 582 Schnitzler, H.-U., Moss, C.F. & Denzinger, A. (2003). From spatial orientation to food
583 acquisition in echolocating bats. *Trends in Ecology and Evolution*, **18**, 386–394.
- 584 Simmons, N.B. (2005). Order Chiroptera. *Mammal Species of the World* (eds D.E. Wilson &
585 D.M. Reeder), pp. 312–529. John Hopkins University Press, Baltimore.
- 586 Skowronski, M.D. & Harris, J.G. (2006). Acoustic detection and classification of
587 microchiroptera using machine learning: lessons learned from automatic speech
588 recognition. *The Journal of the Acoustical Society of America*, **119**, 1817–1833.
- 589 Stathopoulos, V., Zamora-Gutierrez, V., Jones, K.E. & Girolami, M. (2014). Bat call
590 identification with gaussian process multinomial probit regression and a dynamic time
591 warping kernel. *Proceedings of the 17th International Conference on Artificial
592 Intelligence and Statistics*, **33**, 913–921.
- 593 Stowell, D. & Plumbley, M.D. (2014). Large-scale analysis of frequency modulation in
594 birdsong data bases. *Methods in Ecology and Evolution*, **5**, 901–912.
- 595 Szewczak, J.M. (2010). Sonobat v.3. URL www.sonobat.com [accessed 1 July 2015]
- 596 Vaughan, N., Jones, G. & Harris, S. (1997). Identification of British bat species by
597 multivariate analysis of echolocation call parameters. *Bioacoustics*, **7**, 189–207.
- 598 Vens, C., Struyf, J., Schietgat, L., Dzeroski, S. & Blockeel, H. (2008). Decision trees for
599 hierarchical multi-label classification. *Machine Learning*, **73**, 185–214.
- 600 Villers-Ruiz, L. & Trejo-Vázquez, I. (2003). El cambio climático y la vegetación en México.
601 *México: una visión hacia el siglo XXI. El cambio climático en México* (eds G. Garcia-
602 Carlos), pp. 57–72. INE, UNAM, US Country Studies Program, México, D.F.

603 Walters, C., Collen, A., Lucas, T., Mroz, K., Sayer, C. & Jones, K.E.. (2013). Challenges of
604 using bioacoustics to globally monitor bats. *Bat Evolution, Ecology, and Conservation*
605 (eds R.A. Adams & S.C. Pedersen), pp. 479–500. Springer, New York.

606 Walters, C.L., Freeman, R., Collen, A., Dietz, C., Fenton, M.B., Jones, G., Obrist, M.K.,
607 Puechmaille, S.J., Sattler, T., Siemers, B.M., Parsons, S. & Jones, K.E. (2012). A
608 continental-scale tool for acoustic identification of European bats. *Journal of Applied*
609 *Ecology*, **49**, 1064–1074.

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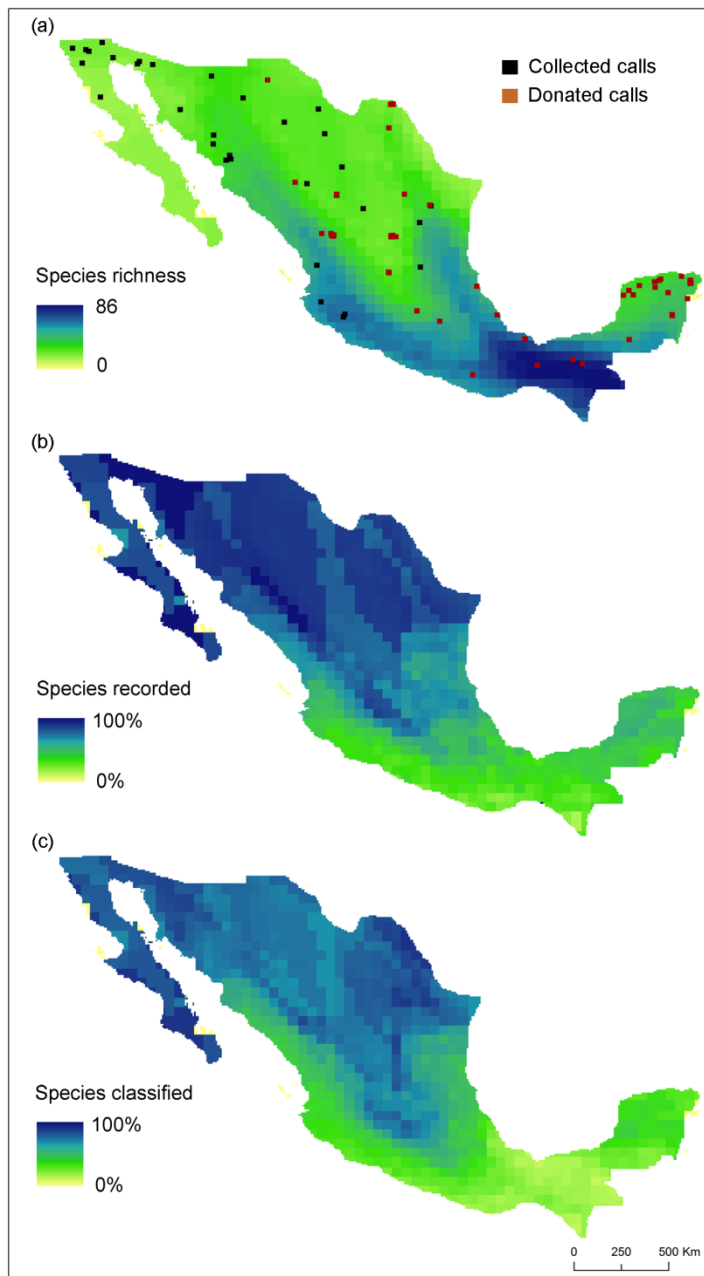
611 **Table 1.** Comparison of classification accuracies of four acoustic classifiers for Mexican bat
 612 species (n=59 species). Where Classifier 1 represents species-level without a hierarchy;
 613 Classifier 2: species-level within families; Classifier 3: species-level within genera; Classifier
 614 4: species-level within guild. Misclassification represents those classes having the most
 615 misclassifications with each other for each classifier and level, where Phyllo Phyllostomidae;
 616 Vesp Vespertilionidae; Molo Molossididae; and Noct Noctilionidae.

617

Classifier	Level	Mean accuracy %	Accuracy range %	% of classes $\geq 80\%$ accuracy	% of classes $\leq 60\%$ accuracy	Misclassifications
1	Species	66	4.2-100	29	41	Species of Phyllo with themselves or Vesp; Vesp with themselves or Molo
2	Species	72	0-100	32	44	Species within families
	Family	91.7	72.8-100	88	0	Noct with Molo
3	Species	71.2	0-100	36	37	Species within genera
	Genus	77.8	0-100	56	16	Phylo with other Phylo and Vesp genera; and Vesp with Phylo and Molo genera
4	Species	69.1	4.5-100	25	44	Species within guilds
	Guild	82.5	68-100	50	0	Guild 5 with Guild 6; Guild 6 with Guild 5

618

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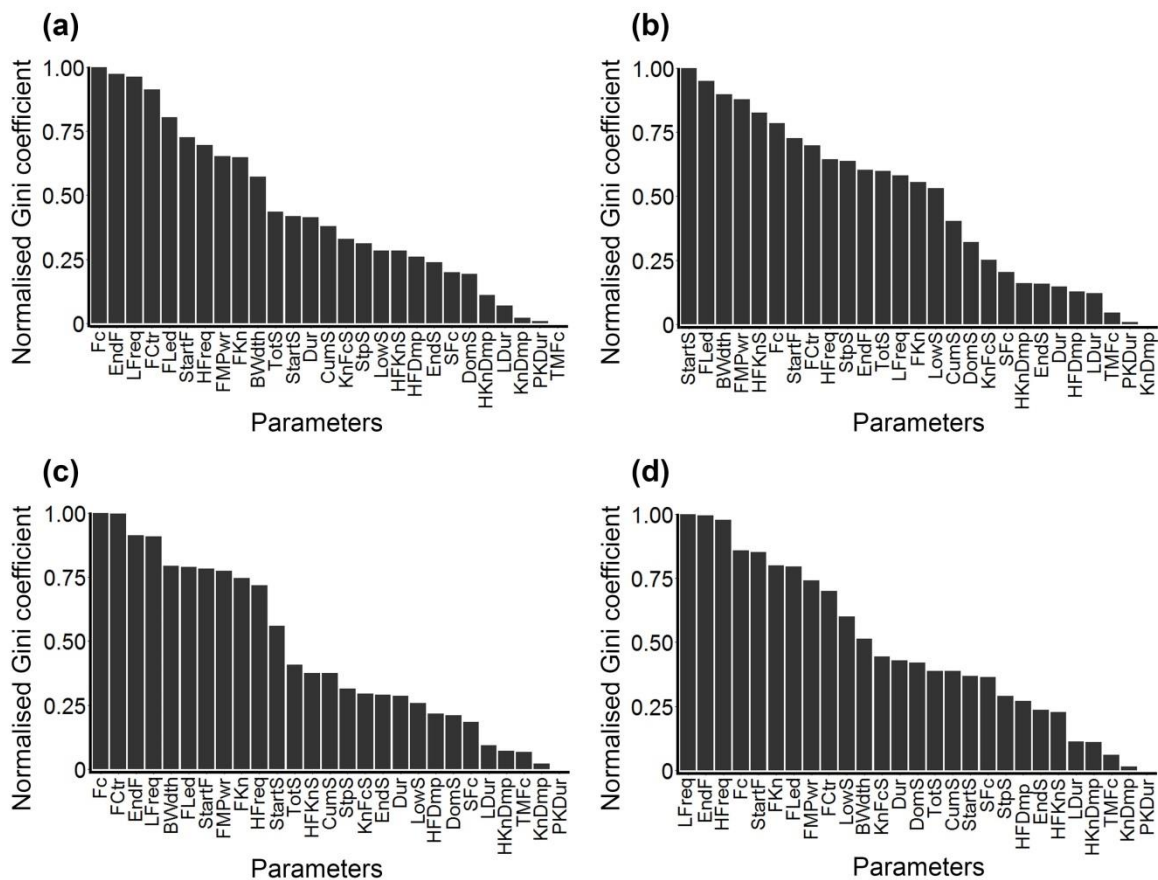


620

621 Figure 1. Spatial coverage of the number of species recorded in Mexico using a grid size of
622 50 km², where (a) shows recording locations in solid squares (n = 91) overlaid with bat
623 species richness, (b) proportion of species recorded compared to potential species richness
624 in each grid, and (c) proportion of species used in the classifiers compared to potential
625 species richness in each grid. A gradient of light green to dark blue indicate higher number
626 of species and higher percentages. Black solid squares represent collection sites which were
627 sampled in this study and red solid squares represent collection sites of donated material.

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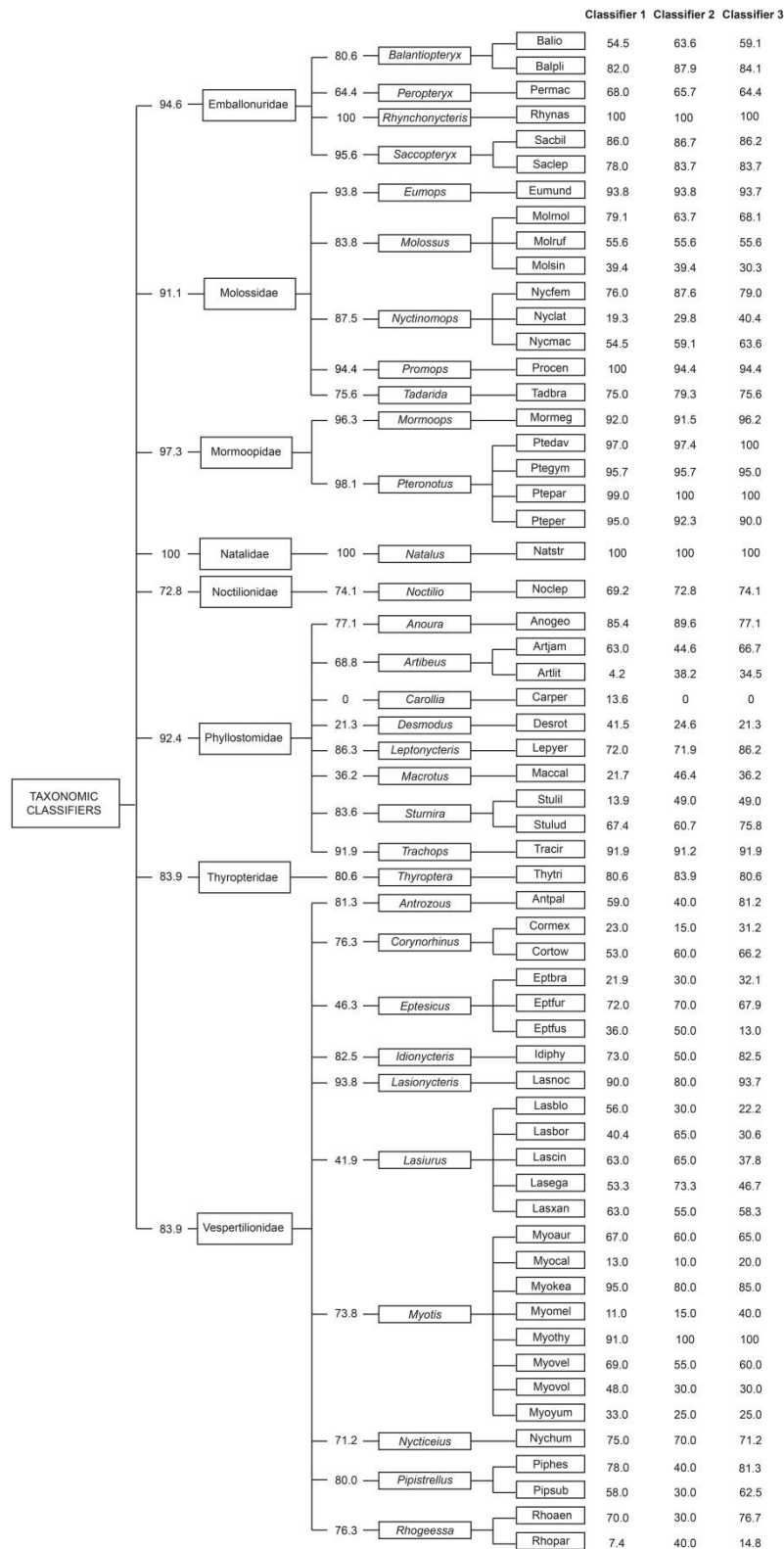
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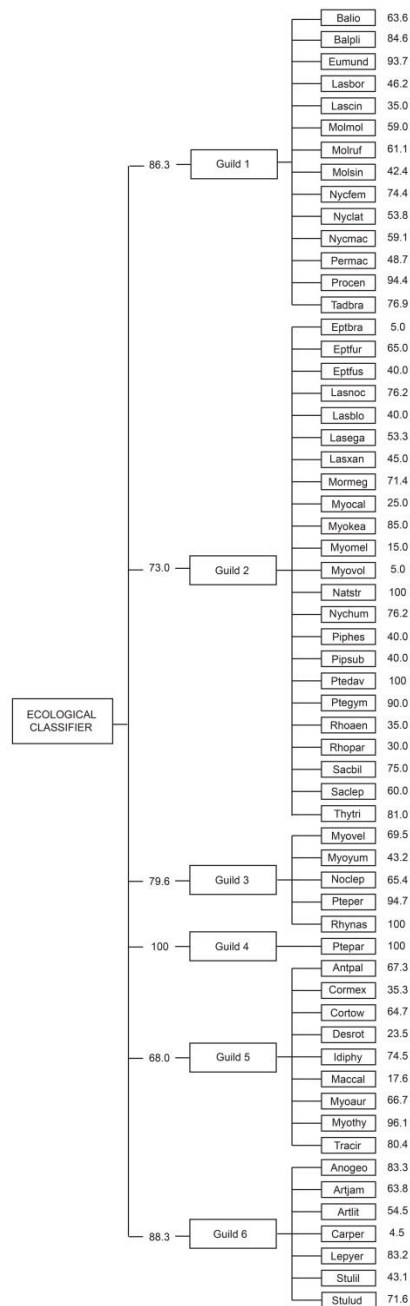
631 **Figure 2.** Echolocation call parameters (n = 27) selected to build each Random Forest
 632 classifier ranked by Gini Coefficient where (a) Classifier 1: Species-level without a hierarchy;
 633 (b) Classifier 2: Species-level within a family hierarchy; (c) Classifier 3: Species-level within a
 634 genus hierarchy; and (d) Classifier 4: Species-level within a guild hierarchy. See Table S3
 635 for parameter definitions.

636



637

638 **Figure 3.** Random Forest percentage classification accuracies obtained for the taxonomic
 639 classifiers (Classifiers 1-3). Species-level accuracies are shown at the end of each branch
 640 for Classifier 1, 2 and 3. Classification accuracies per family and genus are shown in the
 641 middle of each branch (n = 59 species).



642

643 **Figure 4.** Random Forest percentage classification accuracies obtained for the ecological
 644 guild classifier (Classifier 4). Species-level accuracies are shown at the end of each branch.
 645 Classification accuracies per guild are shown in the middle of each branch (n = 59 species).
 646 Guild 1 - Open space aerial foragers; Guild 2 – Edge space aerial foragers; Guild 3 – Edge
 647 space trawling foragers; Guild 4 – Narrow space flutter detecting foragers; Guild 5 - Narrow
 648 space passive gleaning foragers and Guild 6- Narrow space passive/active gleaning
 649 foragers.

650 **Supporting Information**

651 Additional Supporting Information may be found in the online version of this article:

652 **Table S1.** Metadata for the search-phase echolocation calls collected during field work in
653 Mexico from June 2012 to May 2013.

654 **Table S2.** Metadata for the search-phase echolocation calls donated for this study.

655 **Table S3.** Definitions of the 27 call parameters extracted by Sonobat v.3 used for training
656 the Random Forest classifiers.

657 **Table S4.** Number of classes included in each hierarchy of the four classifiers and number of
658 calls used in the training process.

659 **Table S5.** Taxonomic coverage of the bat call library within each family for the number of
660 genera and species recorded / used in the classifiers.

661 **Table S6.** Descriptive statistics (mean and standard deviation) for the 16 most important call
662 parameters ranked by Random Forest Gini Coefficient measured by Sonobat for the 59
663 species.

664 **Figure S1.** Spectrograms showing the inter-specific variability of representative search-
665 phase echolocation calls within taxonomic groups used for the classifiers.

666 **Figure S2.** Spectrograms showing inter-specific variability of representative search-phase
667 echolocation calls within ecological guilds used for the classifiers.

668 **Figure S3.** Schematic representation of the protocol used to build the classifiers.