LINE-1 Hypomethylation in Blood and Tissue Samples as an Epigenetic Marker for Cancer Risk: A Systematic Review and Meta-Analysis



Martina Barchitta¹, Annalisa Quattrocchi¹, Andrea Maugeri¹, Manlio Vinciguerra^{2,3}*, Antonella Agodi¹*

1 Department GF Ingrassia, University of Catania, Catania, Italy, 2 University College London, Institute for Liver and Digestive Health, Royal Free Campus, London, United Kingdom, 3 Gastroenterology Unit, Department of Medical Sciences, IRCCS Casa Sollievo della Sofferenza, San Giovanni Rotondo, Italy

Abstract

Objective: A systematic review and a meta-analysis were carried out in order to summarize the current published studies and to evaluate LINE-1 hypomethylation in blood and other tissues as an epigenetic marker for cancer risk.

Methods: A systematic literature search in the Medline database, using PubMed, was conducted for epidemiological studies, published before March 2014. The random-effects model was used to estimate weighted mean differences (MDs) with 95% Confidence Intervals (CIs). Furthermore, subgroup analyses were conducted by sample type (tissue or blood samples), cancer types, and by assays used to measure global DNA methylation levels. The Cochrane software package Review Manager 5.2 was used.

Results: A total of 19 unique articles on 6107 samples (2554 from cancer patients and 3553 control samples) were included in the meta-analysis. LINE-1 methylation levels were significantly lower in cancer patients than in controls (MD: -6.40, 95% CI: -7.71, -5.09; p<0.001). The significant difference in methylation levels was confirmed in tissue samples (MD -7.55; 95% CI: -9.14, -65.95; p<0.001), but not in blood samples (MD: -0.26, 95% CI: -0.69, 0.17; p=0.23). LINE-1 methylation levels were significantly lower in colorectal and gastric cancer patients than in controls (MD: -8.33; 95% CI: -10.56, -6.10; p< 0.001 and MD: -5.75; 95% CI: -7.75, -3.74; p<0.001) whereas, no significant difference was observed for hepatocellular cancer.

Conclusions: The present meta-analysis adds new evidence to the growing literature on the role of LINE-1 hypomethylation in human cancer and demonstrates that LINE-1 methylation levels were significantly lower in cancer patients than in control samples, especially in certain cancer types. This result was confirmed in tissue samples, both fresh/frozen or FFPE specimens, but not in blood. Further studies are needed to better clarify the role of LINE-1 methylation in specific subgroups, considering both cancer and sample type, and the methods of measurement.

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* Email: agodia@unict.it (AA); m.vinciguerra@ucl.ac.uk (MV)

Introduction

Epigenetic alterations, heritable DNA modifications that do not involve changes in the DNA sequence, are associated with changes in gene expression and are important in maintaining genomic stability [1]. Among epigenetic mechanisms, DNA methylation is the most commonly studied and involved in various biological processes including cancer [2–5]. Global hypomethylation, an overall genome-wide reduction in DNA methylation content, is associated with genomic instability and an increased number of mutational events [6]. Genomic DNA hypomethylation is likely to result from demethylation in repetitive elements, which account for about 55% of the human genome and determine gene regulation and genomic stability [7,8]. Long Interspersed Nucleotide Element 1 (LINE-1) and Alu repetitive elements are major constituents of interspersed DNA repeats. Due to their high occurrence throughout the genome, methylation in repetitive elements have been shown to correlate with global genomic DNA methylation content and demethylation has been associated with genome instability and chromosomal aberrations. Thus, LINE-1 and Alu have been used as global surrogate markers for estimating the genomic DNA methylation level in cancer tissues [6,9–10] and in peripheral blood leukocytes [11]. LINE-1 hypomethylation was observed in several types of cancer [12–14] and was associated with a poor prognosis [15]. In a meta-analysis [11], global DNA hypomethylation in peripheral blood leukocytes was associated with increased cancer risk. Another meta-analysis, investigating genome-wide DNA methylation in peripheral blood DNA and

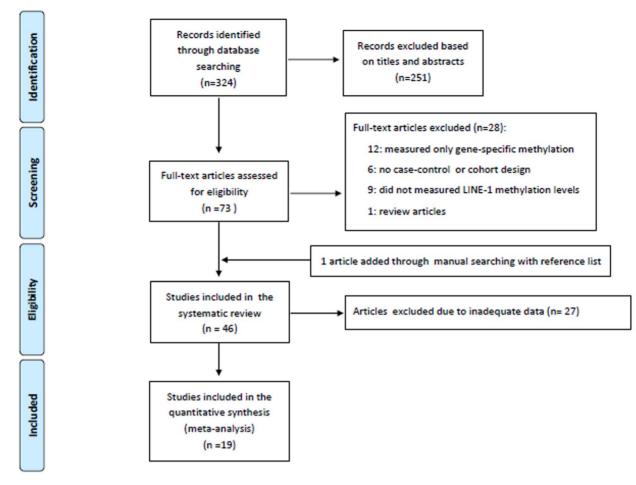


Figure 1. Flow diagram of study selection. doi:10.1371/journal.pone.0109478.g001

cancer risk, reports a significant inverse association between genomic 5-methylcytosine levels and cancer risk, but no overall risk association using surrogates for genomic methylation, including methylation at the LINE-1 and Alu repetitive elements was found [16]. The aim of the present study was to carry out a more comprehensive systematic review and a meta-analysis in order to summarize the current published studies and to evaluate LINE-1 hypomethylation in blood and other tissues as an epigenetic marker for cancer risk.

Methods

Search strategy and selection criteria

A systematic literature search in the Medline database, using PubMed, was carried out for epidemiological studies, published before March 2014, investigating the association between LINE-1 hypomethylation and cancer risk. The searches were limited to studies written in English; abstracts and unpublished studies were not included. Literature search was conducted independently by two Authors. The following selection criteria were used to search articles and abstracts: ("cancer" or "tumor" or "carcinoma") AND ("LINE-1" or "Long Interspersed Element-1" or "global") AND ("hypomethylation" or "methylation"). Moreover, the reference lists from selected articles were checked to search for further relevant studies. No studies were excluded a priori for weakness of design or data quality. Articles were included in the quantitative analysis only if they satisfied the following criteria: (1) case-control or cohort study designs; and (2) studies that reported mean values and standard deviations (SD) of DNA methylation level in cancer patients and in control group. Furthermore exclusion criteria were as follows: (1) the study reporting only results as median of the methylation levels or through graphic display, or 95% confidence intervals (CIs) with adjusted odds ratios (OR) or relative risks for cancer risk in subjects with the lowest level of global DNA methylation (tertile, quartile or decile) compared to group with the highest level, (2) the study reporting only gene-specific DNA methylation analysis, and (3) review articles.

Where there were missing data or additional information were required, study Authors were contacted by email.

The preferred reporting items for systematic reviews and metaanalysis (PRISMA) guidelines for the conduct of meta-analysis were followed [17].

Data collection and extraction

Two of the Authors independently reviewed all the eligible studies and abstracted the following information in a standard format: first Author's last name, year of publication, country where the study was performed, study design, cancer sites and types, sample type, experimental methods to measure global DNA methylation levels, number of cases and controls, mean values and SD of global DNA methylation levels for each group and main results.

Author (Year)	Country	Study design	Cancer type	Sample type	Assay	Number of cases/controls	Mean (SD) Cases	Mean (SD) Controls	Results
Antelo (2012) ^a *	Argentina	Retrospective	Colorectal (Early Onset)	Tissue	Pyrosequencing	185/32	56.6 (8.6)	75.5 (1.5)	Early-onset CRC had significantly lower LINE-1 methylation levels than any other group
Antelo (2012) ^b *	Argentina	Retrospective	Colorectal (Lynch Syndrome)	Tissue	Pyrosequencing	20/32	66.3 (4.5)	75.5 (1.5)	
Antelo (2012) ^c *	Argentina	Retrospective	Colorectal (Older Onset sporadic MSI-high)	Tissue	Pyrosequencing	46/32	67.1 (5.5)	75.5 (1.5)	
Antelo (2012) ^d *	Argentina	Retrospective	Colorectal (Older Onset sporadic MSS/MSI-low)	Tissue	Pyrosequencing	89/32	65.1 (6.3)	75.5 (1.5)	
Choi (2007)*	USA	Retrospective	Neuroendocrine	Tissue	Pyrosequencing	35/35	68.5 (10.0)	80.0 (7.1)	LINE-1 methylation levels were significantly lower in cancer tissues than in normal adjacent tissues
Daskalos (2009)*	Ч	Retrospective	Lung	Tissue	Pyrosequencing	48/48	54.36 (10.52)	69.56 (1.1)	LINE-1 methylation levels were significantly lower in cancer tissues than in normal adjacent tissues
Estecio (2007)*	USA	Retrospective	Colorectal and various cancer Tissue cell lines	Tissue	Pyrosequencing	60/60	54.9 (1.1)	64.3 (0.5)	LINE-1 methylation levels were significantly lower in cancer tissues than in normal adjacent tissues
Hur (2014)*	Spain	Retrospective	Colorectal	Tissue	Pyrosequencing	71/77	66.2 (5.3)	75.8 (3.10)	Compared with normal adjacent mucosa, both primary cancer and metastasis tissue were significantly hypomethylated at LINE-1 elements
lwagami (2013)*	Japan	Prospective	Esophageal	Tissue	Pyrosequencing	50/50	63.3 (12.7)	78.8 (6.2)	LINE-1 methylation levels were significantly lower in cancer tissues than in normal adjacent tissues
Lee (2008)*	Sweden	Retrospective	Thyroid	Tissue	Pyrosequencing	21/21	71.3 (2.6)	71.8 (3.4)	LINE-1 methylation changes are not observed between cancer and normal tissues
Lee (2011)*	South Korea	Retrospective	Gastric	Tissue	COBRA LINE-1	53/53	40.23 (0.92)	45.94 (1.78)	LINE-1 methylation levels were significantly lower in cancer tissues than in normal adjacent tissues
Matsunoki (2012)*	Japan	Retrospective	Colorectal	Tissue	MulticolorMethyLight Assay 48/48	ssay 48/48	63.61 (13.91)	62.54 (14)	LINE-1 methylation levels were significantly lower in cancer tissues than in normal adjacent tissues

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Author (Year)	Country	Study design	Cancer type	Sample type	Assay	Number of cases/controls	Mean (SD) Cases	Mean (SD) Controls	Results
Pavicic (2012) ^a *	Finland	Retrospective	Colorectal (Sporadic MSS)	Tissue	MS-MLPA	55/55	85 (6)	93 (2)	LINE-1 methylation levels were significantly lower in cancer tissues than in normal adjacent tissues
Pavicic (2012) ^b *	Finland	Retrospective	Colorectal (Sporadic MSI)	Tissue	MS-MLPA	52/52	87 (5)	91 (4)	LINE-1 methylation levels were significantly lower in cancer tissues than in normal adjacent tissues
Pavicic (2012) ^c *	Finland	Retrospective	Colorectal (Lynch Syndrome)	Tissue	MS-MLPA	43/43	84 (6)	90 (5)	LINE-1 methylation levels were significantly lower in cancer tissues than in normal adjacent tissues
Pavicic (2012) ^d *	Finland	Retrospective	Colorectal (FCCX)	Tissue	MS-MLPA	18/18	80 (8)	84 (6)	LINE-1 methylation levels were significantly lower in cancer tissues than in normal adjacent tissues
Pavicic (2012) ^e *	Finland	Retrospective	Gastric (Sporadic MSS)	Tissue	MS-MLPA	34/34	79 (12)	90 (5)	LINE-1 methylation levels were significantly lower in cancer tissues than in normal adjacent tissues
Pavicic (2012) ^f *	Finland	Retrospective	Gastric (Sporadic MSI)	Tissue	MS-MLPA	11/11	88 (4)	90 (4)	LINE-1 methylation levels were significantly lower in cancer tissues than in normal adjacent tissues
Pavicic (2012) ^{9*}	Finland	Retrospective	Gastric (Lynch Syndrome)	Tissue	MS-MLPA	13/13	86 (5)	90 (5)	LINE-1 methylation levels were significantly lower in cancer tissues than in normal adjacent tissues
Pavicic (2012) ^h *	Finland	Retrospective	Endometrial (Lynch Syndrome) Tissue	:) Tissue	MS-MLPA	50/50	88 (7)	(2) 06	LINE-1 methylation levels were significantly lower in cancer tissues than in normal adjacent tissues
Shigaki (2013)*	Japan	Prospective	Gastric	Tissue	Pyrosequencing	74/74	72.3 (10.1)	79.2 (5.6)	LINE-1 methylation levels were significantly lower in cancer tissues than in normal adjacent tissues
Shuangshoti (2007)*	Thailand	Retrospective	Cervix uterine	Tissue	Cobra Line-1	7/15	35.63 (7.32)	40.6 (8.86)	LINE-1 methylation levels were significantly lower in cancer patients than in healthy subjects
Subbalekha (2008)*	Thailand	Retrospective	Head and Neck	Oral rinses	Cobra Line-1	38/37	37.53 (2.61)	41.78 (2.84)	LINE-1 methylation levels were significantly lower in cancer patients than in

Author (Year)	Country	Study design	Cancer type	Sample type	Assay	Number of cases/controls	Mean (SD) Cases	Mean (SD) Controls	Results
Cash (2012)*	China	Retrospective	Bladder	Blood	Pyrosequencing	510/528	81.86 (1.82)	81.96 (1.89)	LINE-1 methylation were comparable in cases and controls
Liao (2011)*	Central and eastern Europe	Retrospective	Renal cell	Blood	Pyrosequencing	328/654	82.13 (1.86)	81.74 (1.98)	LINE-1 methylation levels were significantly higher in cancer patients than in healthy subjects
Mirabello (2010)*	USA	Retrospective	Testicular	Blood	Pyrosequencing	152/255	79.1 (0.177)	79.3 (0.128)	There was no significant difference between LINE-1 methylation levels in cases and controls
Ramzy (2011)*	Egypt	Retrospective	Hepatocellular	Blood	COBRA LINE-1	50/10	41.86 (10.06)	54.00 (7.82)	LINE-1 methylation levels were significantly lower in cancer patients than in healthy subjects
Tangkijvanich (2007)*	Thailand	Retrospective	Hepatocellular	Blood	COBRA LINE-1	85/30	46.83 (7.74)	53.45 (4.29)	LINE-1 methylation levels were significantly lower in cancer patients than in healthy subjects
Wu (2012)*	Taiwan	Retrospective	Hepatocellular	Blood	Pyrosequencing	302/1250	76.2 (2.2)	76.2 (2.1)	There was no significant difference between LINE-1 methylation levels in cases and controls
Baba (2010)	USA	Prospective	Colorectal	Tissue	Pyrosequencing	869/NA			Tumor LINE-1 methylation data indicate enormous epigenomic diversity of individual colorectal cancers
Bae (2012)	South Korea	Prospective	Gastric	Tissue	Pyrosequencing	447(two sets of 249 and 198)/NA	0		LINE-1 hypomethylation is an early event in carcinogenesis and it may be a prognostic indicator independent of cancer stage
Bollati (2009)	Italy	Retrospective	Multiple Myeloma	Bone marrow aspirates	Bone marrow Pyrosequencing aspirates	76/11			Cases showed a decrease of LINE-1 methylation levels compared to controls
Chalitchagorn (2004)	Thailand	Retrospective	Gastric	Blood	COBRA LINE-1	71/71			Cases showed a decrease of LINE-1 methylation levels compared to controls
Choi (2009)	USA	Retrospective	Breast	Blood	Pyrosequencing	19/18			There was no significant difference between LINE-1 methylation levels in cases and controls

Table 1. Cont.									
Author (Year)	Country	Study design	Cancer type	Sample type	Assay	Number of cases/controls	Mean (SD) Cases	Mean (SD) Controls	Results
Dammann (2010)	Germany	Retrospective	Ovarian	Tissue	QUBRA	22/NA			High prevalence of LINE1 hypomethylation throughout all tumor stages
Di (2011)	China	Retrospective	Hepatocellular	Blood	Pyrosequencing	315/356			Hypomethylation lead to a significant 2.6-fold increased risk for HCC
Fabris (2011)	Italy	Retrospective	Chronic lymphocytic leukemia Blood	a Blood	Pyrosequencing	7/77			LINE-1 methylation levels were significantly lower in cancer patients than in healthy subjects
Gao (2012)	China	Retrospective	Gastric	Blood	Pyrosequencing	192/384			There was no significant difference between LINE-1 methylation levels in cases and controls
Gao (2013)	China	Prospective	Hepatocellular	Tissue	Sequencing and Real-time qPCR	243/48			Hypomethylation of LINE-1 was associated with tumour progression, larger tumour size, higher recurrence rates, worse tumour stage and poor tumour differentiation
Geli (2008)	Sweden	Retrospective	Pheochromocytoma and Paraganglioma	Tissue	Pyrosequencing	55/NA			Cases showed a decrease LINE-1 methylation levels compared with controls
Hsiung (2007)	USA	Retrospective	Head and Neck	Blood	COBRA LINE-1	278/526			The median methylation level in controls was slightly but significantly higher than the median level in cases. Hypomethylation lead to a significant 1.6-fold increased risk for disease
Hou (2010)	Poland	Retrospective	Gastric	Blood	Pyrosequencing	302/421			Cancer risk was highest among those with lowest level of methylation in LINE-1 relative to those with the highest levels, although the trends were not statistically significant
Igarashi (2010)	Japan	Retrospective	GIST	Tissue	Pyrosequencing	106/NA			LINE-1 hypomethylation correlates significantly with the aggressiveness of tumors and it could be a useful marker for risk assessment

Author (Year)	Country	Study design	Cancer type	Sample type	Assay	Number of cases/controls	Mean (SD) M Cases C	Mean (SD) Controls	Results
Kreimer (2013)	Germany	Retrospective	Bladder	Tissue	Pyrosequencing	23/12			LINE-1 methylation was significantly decreased in cancers compared to normal tissues with striking differences in their percent median values
Ogino (2008)	USA	Prospective	Colorectal	Tissue	Pyrosequencing	643/NA			LINE-1 hypomethylation was linearly associated with a statistically significant increase in cancer - specific mortality
Phokaew (2008)	Thailand	Retrospective	Head and Neck	Tissue	COBRA LINE-1	11/12			LINE-1 methylation level at each locus is different, it can be influenced differentially depending on where the particular sequences are located in the genome
Pobsook (2011)	Thailand	Retrospective	Head and Neck	Various	Cobra Line-1	90/114			LINE-1 partial methylation represents hypomethylation in normal cells but hypermethylation in cancer cells
Saito (2010)	Japan	Retrospective	Lung	Tissue	Real-time PCR	379/333			LINE-1 methylation levels were significantly lower in cancer patients than in healthy subjects
Sigalotti (2011)	Italy	Retrospective	Melanoma	Tissue	Pyrosequencing	42/4			LINE-1 methylation is identified as a molecular marker of prognosis
Sunami (2011)	USA	Retrospective	Colorectal	Tissue	AQAMA-PCR	117/117			LINE-1 hypomethylation was significantly greater in adenoma tissue compared to its contiguous normal epithelium and cancer mesenchymal tissue
Trankenschuh (2010)	Germany	Retrospective	FLC	Tissue	Pyrosequencing	25/15			No evidence of global hypomethylation was found
Van Hoesel (2012)	USA	Prospective	Breast	Tissue	AQAMA-PCR	129/109			LINE-1 hypomethylation is a prognostic biomarker of poor outcome

	1. Cont.	
	Table	

Author (Year)	Country	Study design	Cancer type	Sample type	Assay	Number of cases/controls	Mean (SD) Cases	Mean (SD) Controls	Results
Wilhelm 2010	USA	Retrospective	Bladder	Blood	Pyrosequencing	285/465			Being in the lowest LINE1 methylation decile was associated with a significant 1.8-fold increased risk of cancer
Wolff (2010)	USA	Retrospective	Bladder	Tissue	Pyrosequencing	113/63			Cases showed a decrease LINE-1 methylation levels compared with controls
Yegnasubramanian (2008)	USA	Retrospective	Prostate	Tissue	COMPARE	76/24			Cases showed a decrease LINE-1 methylation levels compared with controls
Zhu (2011)	USA	Retrospective	Various	Blood	Pyrosequencing	205/487			Individuals with lowest LINE-1 methylation levels had a significant 4.4-fold increased incidence of lung cancer. No significant associations were observed for other tumors
(*) Studies included in the meta-analysis (N = 19). AQAMA-PCR: Absolute Quantitative Assessment (digestion, CRC: colorectal Cancer, FCCX: Familial MLPA: Methylation-Specific Multiplex Ligation-de Advi:10.1377/iournal none 0100428.0001	meta-analysis (N antitative Assess Cancer, FCCX: Fa : Multiplex Ligati	l = 19). ment Of Methylated All milial Colorectal Cancer on-dependent Probe A	(*) Studies included in the meta-analysis (N = 19). AQAMA-PCR: Absolute Quantitative Assessment Of Methylated Alleles PCR, COBRA LINE-1: Combined Bisulfite Restriction Analysis LINE-1, COMPARE: Combination Of Methylated DNA Precipitation And Restriction Enzyme digestion, CRC: Colorectal Cancer, FCCX: Familial Colorectal Cancer type X, FLC: Fibrolamellar Carcinoma, GIST: Gastrointestinal Stromal Tumors, LINE-1: Long Interspersed Nucleotide Element 1, MSI: MicroSatellite Instable, ALDA: Methylated DNA Precipitation, MSI: MicroSatellite Instable, ALDA: Methylated DNA Precipitation And Restriction Enzyme at Methylation-Specific Multiplex Ligation-dependent Probe Amplification, MSS: MicroSatellite Stable, QUBRA: Quantitative Bisulfite Restriction Analysis.	mbined Bisulfite Re Carcinoma, GIST: Ga :llite Stable, QUBRA	sstriction Analysis LINE-1, C istrointestinal Stromal Turr : Quantitative Bisulfite Res	COMPARE: Combination (ors, LINE-1: Long Intersp striction Analysis.	Df Methylated Dl bersed Nucleotid	NA Precipitation e Element 1, MS	(*) Studies included in the meta-analysis (N = 19). AQAMA-PCR: Absolute Quantitative Assessment Of Methylated Alleles PCR, COBRA LINE-1: Combined Bisulfite Restriction Analysis LINE-1, COMPARE: Combination Of Methylated DNA Precipitation And Restriction Enzyme digestion, CRC: Colorectal Cancer, FCCX: Familial Colorectal Cancer type X, FLC: Fibrolamellar Carcinoma, GIST: Gastrointestinal Stromal Tumors, LINE-1: Long Interspersed Nucleotide Element 1, MSI: MicroSatellite Instable, MS- MLDA: Restriction Analysis.

	Can	cer gro	up	Con	trol gro	up		Mean Difference	Mean Difference
Study or Subgroup	Mean	SD	Total	Mean	SD	Total	Weight	IV, Random, 95% CI	IV, Random, 95% CI
1.1.1 Tissue									
Antelo (a) 2012	56.6	8.6	185	75.5	1.5	32	3.9%	-18.90 [-20.24, -17.56]	+
Antelo (b) 2012	66.3	8.6	20	75.5	1.5	32	3.0%	-9.20 [-13.00, -5.40]	
Antelo (c) 2012	67.1	5.5	46	75.5	1.5	32	3.8%	-8.40 [-10.07, -6.73]	-
Antelo (d) 2012	65.1	6.3	89	75.5	1.5	32	3.9%	-10.40 [-11.81, -8.99]	+
Choi 2007	68.5	10	35	80	7.1	35	2.9%	-11.50 [-15.56, -7.44]	
Daskalos 2009	54.36		48	69.56	1.1	48		-15.20 [-18.19, -12.21]	
Estecio 2007	54.9	1.1	60	64.3	0.5	60	4.1%	-9.40 [-9.71, -9.09]	
Hur 2014	66.2	5.3	77	75.8	3.1	77	3.9%	-9.60 [-10.97, -8.23]	-
lwagami 2013	63.3	12.7	50	78.8	6.2	50	3.0%	-15.50 [-19.42, -11.58]	<u> </u>
Lee 2008	71.3	2.6	21	71.8	3.4	21	3.8%	-0.50 [-2.33, 1.33]	-
Lee 2011	40.23	0.92	53	45.94	1.78	53	4.0%	-5.71 [-6.25, -5.17]	•
Matsunoki 2012	63.61	13.91	48	62.54	14	48	2.3%	1.07 [-4.51, 6.65]	
Pavicic (a) 2012	85	6	55	93	2	55	3.8%	-8.00 [-9.67, -6.33]	+
Pavicic (b) 2012	87	5	52	91	4	52	3.8%	-4.00 [-5.74, -2.26]	+
Pavicic (c) 2012	84	6	43	90	5	43	3.6%	-6.00 [-8.33, -3.67]	-
Pavicic (d) 2012	80	8	18	84	6	18	2.7%	-4.00 [-8.62, 0.62]	
Pavicic (e) 2012	79	12	34	90	5	34	2.8%	-11.00 [-15.37, -6.63]	
Pavicic (f) 2012	88	4	11	90	4	11	3.2%	-2.00 [-5.34, 1.34]	
Pavicic (q) 2012	86	5	13	90	5	13	3.0%	-4.00 [-7.84, -0.16]	
Pavicic (h) 2012	88	7	50	90	7	50	3.5%	-2.00 [-4.74, 0.74]	
Shigaki 2013	72.3	10.1	74	79.2	5.6	74	3.5%	-6.90 [-9.53, -4.27]	
Shuangshoti 2007	35.63	7.32	7	40.6	8.86	15	1.9%	-4.97 [-12.01, 2.07]	
Subbalekha 2008	37.53	2.61	38	41.78	2.84	37	3.9%	-4.25 [-5.49, -3.01]	+
Subtotal (95% CI)			1127			922	77.8%	-7.55 [-9.14, -5.95]	•
Heterogeneity: Tau ² =	12.95; (Chi ^z = 61	18.34. 0	df = 22 (P < 0.00	0001); I	² = 96%		
Test for overall effect:									
1.1.2 Blood									
Cash 2012	81.86	1.82	510	81.96	1.89	528	4.1%	-0.10 [-0.33, 0.13]	4
Liao 2011	82.13	1.86		81.74	1.98	654	4.1%	0.39 [0.14, 0.64]	
Mirabello 2010	79.1		152	79.3	0.128	255	4.1%	-0.20 [-0.23, -0.17]	4
Ramzy 2011	41.86	10.06	50	54	7.82	10	2.3%	-12.14 [-17.73, -6.55]	
Tangkijvanich 2007	46.83	7.74		53.45	4.29	30	3.6%	-6.62 [-8.87, -4.37]	
Wu 2012	76.2	2.2	302	76.2	2.1	1250	4.1%	0.00 [-0.27, 0.27]	+
Subtotal (95% CI)			1427			2727	22.2%	-0.26 [-0.69, 0.17]	
Heterogeneity: Tau² = Test for overall effect:				÷5 (P ≺	0.00001	1); I² = 9			
Total (95% CI)			2554			3649	100.0%	-6.40 [-7.71, -5.09]	◆
Heterogeneity: Tau ² = Test for overall effect: Test for subgroup diff	Z=9.56	(P < 0.0	00001)						-20 -10 0 10 20

Figure 2. Forest plot of the mean difference of LINE-1 methylation levels between cancer and control groups in tissue and blood

samples. Subgroup analysis based on sample type. doi:10.1371/journal.pone.0109478.g002

Statistical Analysis

All data were analyzed using the REVIEW MANAGER 5.2 software provided by the Cochrane Collaboration (http://ims. cochrane.org/revman).

The random-effects model was used to estimate weighted mean differences (MDs) with 95% CI [18] and thus, no adjustment for environmental effects was taken into account. Furthermore, subgroup analyses were conducted by sample type (tissue or blood samples), by sample source (fresh tissue or formalin-fixed, paraffin-embedded, FFPE tissue), by cancer types (colorectal, stomach, hepatocellular), and by assays used to measure global DNA methylation levels. Forest plots were generated to illustrate the study-specific effect sizes along with a 95% CI. Heterogeneity across studies, was measured using the Q-test based on the $\chi 2$

statistic, considering significant statistical heterogeneity as p<0.1. As Cochran's test only indicates the presence of heterogeneity and not its magnitude, we also reported the I² statistic, which estimates the percentage of outcome variability that can be attributed to heterogeneity across studies. An I² value of 0% denotes no observed heterogeneity, whereas, 25% is "low", 50% is "moderate" and 75% is "high" heterogeneity [19]. We also estimated the between-study variance using tau-squared (τ^2) statistic [20].

To determine the presence of publication bias, the symmetry of the funnel plots in which mean differences were plotted against their corresponding standard errors were assessed.

	Can	cer gro	up	Cont	rol gro	up		Mean Difference	Mean Difference
Study or Subgroup	Mean	SD	Total	Mean	SD	Total	Weight	IV, Random, 95% CI	IV, Random, 95% CI
1.4.1 fresh/frozen									
Choi 2007	68.5	10	35	80	7.1	35	3.8%	-11.50 [-15.56, -7.44]	
Daskalos 2009	54.36	10.52	48	69.56	1.1	48	4.3%	-15.20 [-18.19, -12.21]	
Estecio 2007	54.9	1.1	60	64.3	0.5	60	5.1%	-9.40 [-9.71, -9.09]	•
lwagami 2013	63.3	12.7	50	78.8	6.2	50	3.9%	-15.50 [-19.42, -11.58]	<u> </u>
Lee 2008	71.3	2.6	21	71.8	3.4	21	4.8%	-0.50 [-2.33, 1.33]	-
Lee 2011	40.23	0.92	53	45.94	1.78	53	5.1%	-5.71 [-6.25, -5.17]	-
Shigaki 2013	72.3	10.1	74	79.2	5.6	74	4.5%	-6.90 [-9.53, -4.27]	
Subbalekha 2008	37.53	2.61	38	41.78	2.84	37	5.0%	-4.25 [-5.49, -3.01]	
Subtotal (95% CI)			379			378	36.5%	-8.19 [-10.54, -5.84]	•
Heterogeneity: Tau² =	: 9.99; C	hi² = 28	5.99, di	f=7 (P ·	< 0.00(001); I ^z a	= 98%		
Test for overall effect:	Z = 6.83	8 (P ≤ 0.	00001)						
1.4.2 FFPE									
Antelo (a) 2012	56.6	8.6	185	75.5	1.5	32	4.9%	-18.90 [-20.24, -17.56]	+
Antelo (b) 2012	66.3	8.6	20	75.5	1.5	32	4.0%	-9.20 [-13.00, -5.40]	
Antelo (c) 2012	67.1	5.5	46	75.5	1.5	32	4.8%	-8.40 [-10.07, -6.73]	-
Antelo (d) 2012	65.1	6.3	89	75.5	1.5	32	4.9%	-10.40 [-11.81, -8.99]	+
Hur 2014	66.2	5.3	77	75.8	3.1	77	4.9%	-9.60 [-10.97, -8.23]	+
Matsunoki 2012	63.61	13.91	48	62.54	14	48	3.1%	1.07 [-4.51, 6.65]	_
Pavicic (a) 2012	85	6	55	93	2	55	4.8%	-8.00 [-9.67, -6.33]	-
Pavicic (b) 2012	87	5	52	91	4	52	4.8%	-4.00 [-5.74, -2.26]	-
Pavicic (c) 2012	84	6	43	90	5	43	4.6%	-6.00 [-8.33, -3.67]	-
Pavicic (d) 2012	80	8	18	84	6	18	3.6%	-4.00 [-8.62, 0.62]	<u> </u>
Pavicic (e) 2012	79	12	34	90	5	34	3.7%	-11.00 [-15.37, -6.63]	
Pavicic (f) 2012	88	4	11	90	4	11	4.2%	-2.00 [-5.34, 1.34]	
Pavicic (g) 2012	86	5	13	90	5	13	3.9%	-4.00 [-7.84, -0.16]	
Pavicic (h) 2012	88	7	50	90	7	50	4.4%	-2.00 [-4.74, 0.74]	
Shuangshoti 2007	35.63	7.32	7	40.6	8.86	15	2.6%	-4.97 [-12.01, 2.07]	<u> </u>
Subtotal (95% CI)			748			544	63.5%	-6.96 [-9.73, -4.20]	◆
Heterogeneity: Tau² =	26.91; 0	Chi² = 3	18.35, i	df = 14 (P < 0.0)0001);	l² = 96%		
Test for overall effect:	Z = 4.94	I(P < 0.	00001)						
Total (95% CI)			1127			922	100.0%	-7.55 [-9.14, -5.95]	•
Heterogeneity: Tau ² =	: 12.95: 0	Chi ² = 6	18.34	df = 22 (P < 0.0				
Test for overall effect:				and a second sec					-20 -10 0 10 20
Toot for oubgroup diff		•			- 0 51	12 - 0	N		

Test for subgroup differences: Chi² = 0.44, df = 1 (P = 0.51), l² = 0%

Figure 3. Forest plot of the mean difference of LINE-1 methylation levels between cancer and control groups in tissue samples. Subgroups analysis based on sample source.

doi:10.1371/journal.pone.0109478.g003

Results

Data extraction

The detailed steps of the systematic review and meta-analysis process are given as a PRISMA flow chart in Figure 1. A total of 324 articles were retrieved from the database, one article was added through manual searching with reference list and thus 46 papers, published between 2004 and 2014, were included in the systematic review and summarized in Table 1 by cancer site or type.

Data characteristics and quality assessment

A total of 18 studies were from Asian countries (40%), 13 from European countries (28%), 13 from USA (28%) and 1 from Argentina and from Egypt (2%, each).

Thirty-eight retrospective longitudinal studies compared LINE-1 methylation levels between cancer patients and healthy subjects or normal adjacent tissues in cancer patients. Eight prospective longitudinal studies analysed LINE-1 methylation levels in cohorts of cancer patients, in relation to the life expectancy, the outcome of the disease or the malignancy of the tumor, identifying the role of LINE-1 hypomethylation as a biomarker of poor prognosis in cancer patients [15,21–27].

In 41 studies LINE-1 methylation levels were evaluated both in tumor and in healthy controls tissues, and in the remaining 5 studies only in cancer patients.

Overall, the studies detected LINE-1 methylation levels in 15332 samples: 8103 from cancer patients (4679 tissue samples, 3276 blood samples ,72 oral rinses and 76 bone marrow plasma cells) and 7136 control samples (6277 from healthy subjects and 859 from normal adjacent tissues in cancer patients).

Regarding the experimental methods to measure LINE-1 methylation levels, the "gold standard" method, used in 63% of studies, was the pyrosequencing of bisulphite converted DNA. Furthermore, 9 studies used combined bisulphite restriction analysis of LINE-1 (COBRA LINE-1) and 8 studies used other methods, i.e. sequencing, real-time PCR, AQAMA PCR, COM-PARE methylation assay, MulticolorMethyLight Assay and MS-MLPA.

	Can	cer gro	up	Cont	rol gro	oup		Mean Difference	Mean Difference
Study or Subgroup	Mean	SD	Total	Mean	SD	Total	Weight	IV, Random, 95% CI	IV, Random, 95% CI
1.3.1 Colorectal									
Antelo (a) 2012	56.6	8.6	185	75.5	1.5	32	5.5%	-18.90 [-20.24, -17.56]	+
Antelo (b) 2012	66.3	8.6	20	75.5	1.5	32	5.0%	-9.20 [-13.00, -5.40]	
Antelo (c) 2012	67.1	5.5	46	75.5	1.5	32	5.5%	-8.40 [-10.07, -6.73]	-
Antelo (d) 2012	65.1	6.3	89	75.5	1.5	32	5.5%	-10.40 [-11.81, -8.99]	-
Estecio 2007	54.9	1.1	60	64.3	0.5	60	5.6%	-9.40 [-9.71, -9.09]	
Hur 2014	66.2	5.3	77	75.8	3.1	77	5.5%	-9.60 [-10.97, -8.23]	+
Matsunoki 2012	63.61	13.91	48	62.54	14	48	4.5%	1.07 [-4.51, 6.65]	_ -
Pavicic (a) 2012	85	6	55	93	2	55	5.5%	-8.00 [-9.67, -6.33]	-
Pavicic (b) 2012	87	5	52	91	4	52	5.5%	-4.00 [-5.74, -2.26]	
Pavicic (c) 2012	84	6	43	90	5	43	5.4%	-6.00 [-8.33, -3.67]	
Pavicic (d) 2012	80	8	18	84	6	18	4.8%	-4.00 [-8.62, 0.62]	
Subtotal (95% CI)			693			481	58.5%	-8.33 [-10.56, -6.10]	◆
Heterogeneity: Tau² =	: 12.56; (Chi² = 20	60.84,	df = 10 (P < 0.0)00001);	I² = 96%		
Test for overall effect:	Z = 7.32	(P < 0.0	00001)						
1.3.2 Hepatocellular									
Ramzy 2011	11 06	10.06	50	64	7.82	10	4.5%	-12.14 [-17.73, -6.55]	
Tangkijvanich 2007	41.80	7.74		53.45		30	4.0%	-6.62 [-8.87, -4.37]	
Wu 2012	40.03	2.2	302	76.2		1250	5.6%	0.00 [-0.27, 0.27]	
Subtotal (95% CI)	70.2	2.2	437	70.2	2.1	1290	15.5%	-5.76 [-12.03, 0.51]	
Heterogeneity: Tau ² =	27 95.0	Chi² = 5I		= 2 (P s	: 0 000			0110[12100,0101]	
Test for overall effect:				- 2 (i	. 0.000	,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,	- 30 %		
	2 - 1.00	ų – 0.	,						
1.3.3 Gastric									
Lee 2011	40.23	0.92	53	45.94	1.78	53	5.6%	-5.71 [-6.25, -5.17]	•
Pavicic (e) 2012	79	12	34	90	5	34	4.9%	-11.00 [-15.37, -6.63]	
Pavicic (f) 2012	88	4	11	90	4	11	5.2%	-2.00 [-5.34, 1.34]	
Pavicic (g) 2012	86	5	13	90	5	13	5.0%	-4.00 [-7.84, -0.16]	
Shigaki 2013	72.3	10.1	74	79.2	5.6	74	5.3%	-6.90 [-9.53, -4.27]	
Subtotal (95% CI)			185			185	26.0%	-5.75 [-7.75, -3.74]	◆
Heterogeneity: Tau² =	: 3.09; Cl	hi² = 11.	.91, df=	= 4 (P =	0.02);	l² = 669	6		
Test for overall effect:	Z = 5.61	(P < 0.0	00001)						
Total (95% CI)			1315			1956	100.0%	-7.17 [-9.82, -4.52]	•
Heterogeneity: Tau ² =	: 32 40 0	$Chi^2 = 2i$		df = 18	(P < 0				
Test for overall effect:				- contract - contraction	v . 0		// = 007	×	-20 -10 Ó 10 20
Test for subgroup diff		•			- 0.22	18-2	26%		

Test for subgroup differences: Chi² = 2.97, df = 2 (P = 0.23), l² = 32.6%

Figure 4. Forest plot of the mean difference of LINE-1 methylation levels between cancer and control groups. Subgroups analysis based on cancer type.

doi:10.1371/journal.pone.0109478.g004

The most frequent tumor type in study was colorectal cancer analyzed in eight studies [15,21,28-33], followed by seven studies that evaluated methylation level in gastric cancer [23,27,32,34-37], five in hepatocellular carcinoma [25,38-41], four in bladder cancer [1,14,42,43] and head and neck carcinoma [10,44-46], two in lung cancer [47,48] and breast cancer [24,49], and single studies assessed methylation levels in renal cell cancer [50], prostate cancer [51], neuroendocrine tumor [52], ovarian cancer [53], thyroid cancer [54], esophageal cancer [26], cervix cancer [55], endometrial cancer [32], skin melanoma [22], testicular cancer [56], leukemia [57], multiple myeloma [58], paraganglioma [59], fibrolamellar carcinoma [60] and gastrointestinal [61]. Four studies evaluated methylation level in several cancer sites [13,28,29,32]. With regard to the assay method, pyrosequencing was used in 29 studies, followed by COBRA in 9 studies, Real-Time PCR and AQAMA-PCR in 2 studies. MulticolorMethy-Light Assay, MS-MLP, COMPARE and QUBRA were adopted in single study each.

Meta-analysis

Of the 46 selected papers, 14 reported means and SD of DNA methylation levels. In addition, means and SDs were independently calculated using data from 2 articles and, among Authors contacted for missing data, 3 responded to the email requests and data were added in the analysis [30,50,56]. Thus, 19 unique articles were included in the quantitative analysis. Furthermore, two papers by Antelo et al. [28] and by Pavicic et al. [32], reported data from different cancer types and thus, they were separated in 4 and 8 sub-studies, respectively (Table 1).

A total of 6107 samples were included in the analysis: 2554 from cancer patients (1127 tissue samples and 1427 blood samples) and 3553 control samples (2811 from healthy subjects and 742 from normal adjacent tissues in cancer patients).

LINE-1 methylation levels were significantly lower in cancer patients than in control samples (MD: -6.40, 95% CI: -7.71, -5.09; p<0.001). However, heterogeneity between studies was significantly high (I²=99%) (**Figure 2**), thus, subgroup analysis based on sample type (tissue or blood samples) was performed.

	Can	cer gro	up	Con	trol gro	up		Mean Difference	Mean Difference
Study or Subgroup	Mean	SD	Total	Mean	SD	Total	Weight	IV, Random, 95% CI	IV, Random, 95% CI
1.2.1 Pyrosequencing	g								
Antelo (a) 2012	56.6	8.6	185	75.5	1.5	32	5.5%	-18.90 [-20.24, -17.56]	+
Antelo (b) 2012	66.3	8.6	20	75.5	1.5	32	4.3%	-9.20 [-13.00, -5.40]	
Antelo (c) 2012	67.1	5.5	46	75.5	1.5	32	5.4%	-8.40 [-10.07, -6.73]	-
Antelo (d) 2012	65.1	6.3	89	75.5	1.5	32	5.5%	-10.40 [-11.81, -8.99]	+
Cash 2012	81.86	1.82	510	81.96	1.89	528	5.7%	-0.10 [-0.33, 0.13]	1
Choi 2007	68.5	10	35	80	7.1	35	4.1%	-11.50 [-15.56, -7.44]	
Daskalos 2009	54.36	10.52	48	69.56	1.1	48	4.7%	-15.20 [-18.19, -12.21]	
Estecio 2007	54.9	1.1	60	64.3	0.5	60	5.7%	-9.40 [-9.71, -9.09]	
Hur 2014	66.2	5.3	77	75.8	3.1	77	5.5%	-9.60 [-10.97, -8.23]	+
lwagami 2013	63.3	12.7	50	78.8	6.2	50	4.2%	-15.50 [-19.42, -11.58]	<u> </u>
Lee 2008	71.3	2.6	21	71.8	3.4	21	5.3%	-0.50 [-2.33, 1.33]	+
Liao 2011	82.13	1.86	328	81.74	1.98	654	5.7%	0.39 [0.14, 0.64]	
Mirabello 2010	79.1	0.177	152	79.3	0.128	255	5.7%	-0.20 [-0.23, -0.17]	1
Shigaki 2013	72.3	10.1	74	79.2	5.6	74	4.9%	-6.90 [-9.53, -4.27]	
Wu 2012	76.2	2.2	302	76.2	2.1	1250	5.7%	0.00 [-0.27, 0.27]	
Subtotal (95% CI)			1997			3180	77.8%	-7.33 [-9.06, -5.59]	•
Heterogeneity: Tau² =					(P < 0.0	00001);	I ² = 1009	6	
Test for overall effect:	Z = 8.27	(P < 0.	00001)						
1.2.2 COBRA									
	40.00			45.04	4 70		5 700	5 74 / O O C . 5 / 71	
Lee 2011	40.23	0.92		45.94	1.78	53	5.7%	-5.71 [-6.25, -5.17]	
Ramzy 2011	41.86		50	54	7.82	10	3.3%	-12.14 [-17.73, -6.55]	
Shuangshoti 2007	35.63	7.32	7	40.6	8.86	15	2.6%	-4.97 [-12.01, 2.07]	
Subbalekha 2008	37.53	2.61	38	41.78	2.84	37	5.5%	-4.25 [-5.49, -3.01]	
Tangkijvanich 2007 Subtotal (95% Cl)	46.83	7.74	85 233	53.45	4.29	30 145	5.1% 22.2%	-6.62 [-8.87, -4.37] -5.75 [-7.13, -4.37]	•
Heterogeneity: Tau ² =	1.17; CI	hi² = 10.	.81, df=	= 4 (P =	0.03); I ^z	= 63%			
Test for overall effect:	Z = 8.16	(P < 0.	00001)						
Total (95% CI)			2230			3325	100.0%	-7.13 [-8.68, -5.58]	•
Heterogeneity: Tau ² =	10.92; (Chi² = 5	375.06,	df = 19	(P < 0.0	00001);	I ² = 1009	6	-20 -10 0 10 20
Test for overall effect:	Z = 9.02	(P < 0.	00001)		835				-20 -10 0 10 20
Test for subgroup diff	erences	: Chi ² =	1.95, d	f=1 (P	= 0.16),	I ² = 48.	7%		

Figure 5. Forest plot of the mean difference of LINE-1 methylation levels between cancer and control groups. Subgroups analysis based on method.

doi:10.1371/journal.pone.0109478.g005

The significant difference in methylation levels was confirmed in tissue samples (MD -7.55; 95% CI: -9.14, -65.95; p<0.001), but not in blood samples (MD: -0.26, 95% CI: -0.69, 0.17; p = 0.23).

A subgroup analysis by sample source was conducted. LINE-1 methylation levels were significantly lower in cancer patients than in control samples in fresh and/or frozen tissue (MD -8.19; 95% CI: -10.54, -5.84; p<0.001) and in FFPE tissue (MD: -6.96; 95% CI: -9.73, -4.20; p<0.001). Heterogeneity between studies, in the two subgroups was significantly high ($I^2 = 98\%$ and 96% respectively) (**Figure 3**).

Furthermore, a subgroup analysis by specific cancer types, for colorectal, hepatocellular and gastric cancer, was conducted. LINE-1 methylation levels were significantly lower in colorectal and gastric cancer patients than in control samples (MD: -8.33; 95% CI: -10.56, -6.10; p<0.001 and MD: -5.75; 95% CI: -7.75, -3.74; p<0.001). No difference of LINE-1 methylation levels in blood leukocytes was observed for hepatocellular cancer (MD: -5.76; 95% CI: -12.03, +0.51; p=0.23). Heterogeneity between studies, in colorectal and hepatocellular subgroups was significantly high (I²=96%), and moderately high in the gastric subgroups (I²=66%) (**Figure 4**).

A subgroup analysis by assays used to measure the methylation levels, and particularly, between the two commonly used techniques, pyrosequencing and COBRA LINE-1, was performed. The MDs for *pyrosequencing* and *COBRA LINE-1* subgroups were -7.33 (95% CI: -9.06, -5.59; p<0.001) and -5.75 (95% CI: -7.13, -4.37; p=0.03), respectively. Heterogeneity between studies and in the *pyrosequencing* subgroup was significantly high ($I^2 = 100\%$), and moderately high in the *COBRA* subgroup ($I^2 = 63\%$) (**Figure 5**).

A subgroup analysis by sample type, particularly tissue samples, and assay method was conducted. The MDs in the subgroups of studies which detected LINE-1 methylation levels in tissue samples through pyrosequencing and COBRA LINE-1, were -10.42 (95% CI: -12.93, -7.91; p<0.001) and -5.12 (95% CI: -6.33, -3.91; p = 0.10), respectively. Heterogeneity between studies and in the *pyrosequencing* subgroup was significantly high (I² = 97%), moderately high in *COBRA LINE-1* subgroup (I² = 56%) (**Figure 6**). Stratification among studies which detected LINE-1 methylation in blood samples was not performed due to the paucity of studies.

The funnel plots indicate small to moderate asymmetry, suggesting that publication bias cannot be completely excluded as a factor of influence on the present meta-analysis (**Figures 7–12**).

	Cancer group			Control group				Mean Difference	Mean Difference
Study or Subgroup	Mean	SD	Total	Mean	SD	Total	Weight	IV, Random, 95% CI	IV, Random, 95% CI
1.2.1 Pyrosequencing	g								
Antelo (a) 2012	56.6	8.6	185	75.5	1.5	32	7.9%	-18.90 [-20.24, -17.56]	+
Antelo (b) 2012	66.3	8.6	20	75.5	1.5	32	6.3%	-9.20 [-13.00, -5.40]	
Antelo (c) 2012	67.1	5.5	46	75.5	1.5	32	7.7%	-8.40 [-10.07, -6.73]	-
Antelo (d) 2012	65.1	6.3	89	75.5	1.5	32	7.8%	-10.40 [-11.81, -8.99]	+
Choi 2007	68.5	10	35	80	7.1	35	6.1%	-11.50 [-15.56, -7.44]	
Daskalos 2009	54.36	10.52	48	69.56	1.1	48	6.9%	-15.20 [-18.19, -12.21]	
Estecio 2007	54.9	1.1	60	64.3	0.5	60	8.1%	-9.40 [-9.71, -9.09]	-
Hur 2014	66.2	5.3	77	75.8	3.1	77	7.9%	-9.60 [-10.97, -8.23]	-
lwagami 2013	63.3	12.7	50	78.8	6.2	50		-15.50 [-19.42, -11.58]	
Lee 2008	71.3	2.6	21	71.8	3.4	21	7.6%	-0.50 [-2.33, 1.33]	-
Shigaki 2013	72.3	10.1	74	79.2	5.6	74	7.2%	-6.90 [-9.53, -4.27]	
Subtotal (95% CI)			705			493	79.9%		•
Heterogeneity: Tau ² = 16.41; Chi ² = 310.59, df = 10 (P < 0.00001); l ² = 97%									
Test for overall effect:	Z = 8.14	(P < 0.)	00001)						
1.2.2 COBRA									
Lee 2011	40.23	0.92	53	45.94	1.78	53	8.1%	-5.71 [-6.25, -5.17]	•
Shuangshoti 2007	35.63	7.32	7	40.6	8.86	15	4.1%	-4.97 [-12.01, 2.07]	
Subbalekha 2008	37.53	2.61	38	41.78	2.84	37	7.9%	-4.25 [-5.49, -3.01]	-
Subtotal (95% CI)			98			105	20.1%	-5.12 [-6.33, -3.91]	◆
Heterogeneity: Tau ² = 0.58; Chi ² = 4.53, df = 2 (P = 0.10); l ² = 56%									
Test for overall effect:	Z = 8.30	I (P < 0.I	00001)						
Total (95% CI)			803			598	100.0%	-9.32 [-11.34, -7.29]	•
Heterogeneity: Tau ² = 13.14; Chi ² = 528.23, df = 13 (P < 0.00001); I ² = 98%									
Test for overall effect: Z = 9.00 (P < 0.00001) - 20 - 10 0 10 20									
Test for subgroup differences: Chi ² = 13.91, df = 1 (P = 0.0002), I ² = 92.8%									

Figure 6. Forest plot of the mean difference of LINE-1 methylation levels between cancer and control groups in tissue samples. Subgroups analysis based on method.

doi:10.1371/journal.pone.0109478.g006

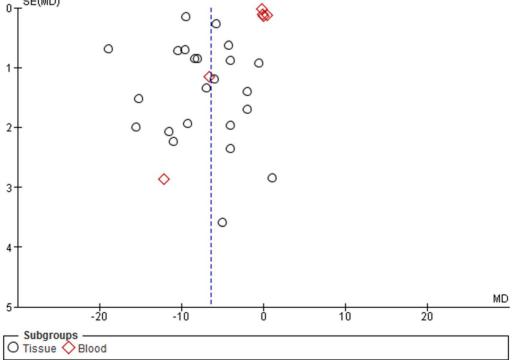


Figure 7. Funnel plot. Subgroup analysis based on sample type. doi:10.1371/journal.pone.0109478.g007

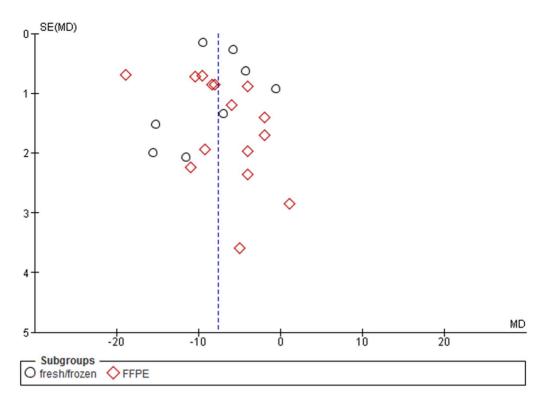


Figure 8. Funnel plot. Subgroup analysis based on tissue specimen types. SE, standard error, MD, mean difference. doi:10.1371/journal.pone.0109478.g008

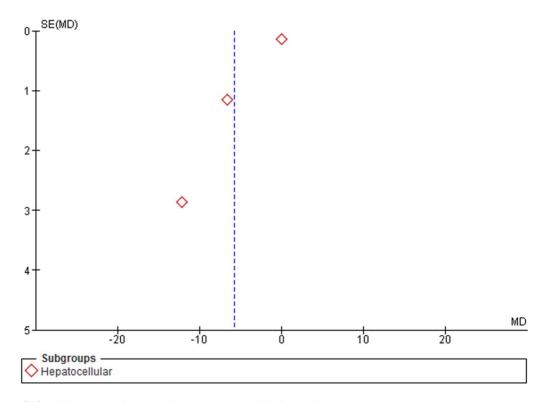


Figure 9. Funnel plot. Subgroup analysis based on cancer type in blood samples. doi:10.1371/journal.pone.0109478.g009

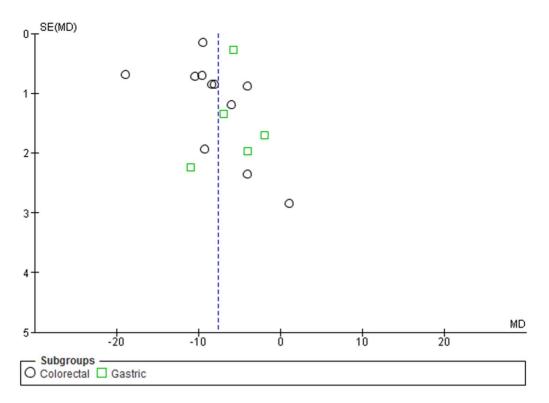


Figure 10. Funnel plot. Subgroup analysis based on cancer type in tissue samples. doi:10.1371/journal.pone.0109478.g010

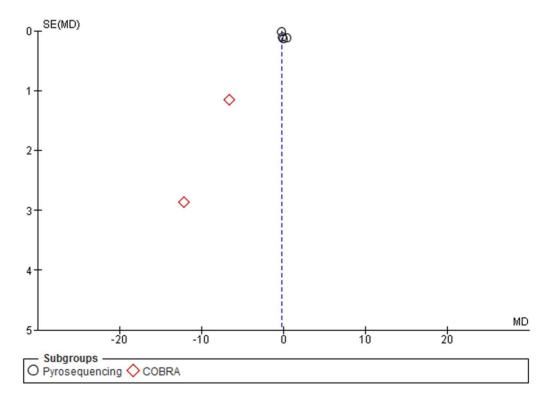


Figure 11. Funnel plot. Subgroup analysis based on detection method in blood samples. doi:10.1371/journal.pone.0109478.g011

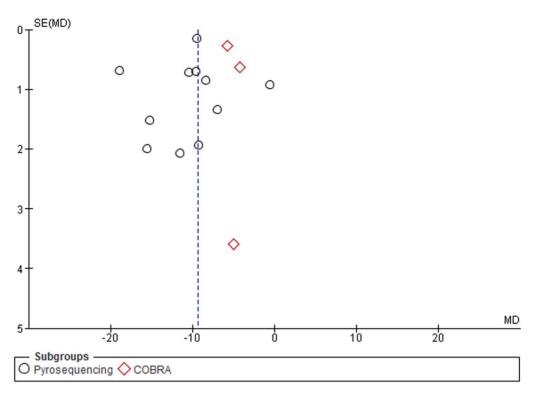


Figure 12. Funnel plot. Subgroup analysis based on detection method in tissue samples. doi:10.1371/journal.pone.0109478.g012

Discussion

The low level of DNA methylation in tumors compared with DNA methylation level in their normal-tissue counterparts was one of the first epigenetic alterations to be found in human cancer [62]. The loss of methylation is mainly due to hypomethylation of repetitive DNA sequences and LINE-1 elements are typically heavily methylated in normal tissues, while LINE-1 hypomethylation has been reported in cancer tissues. Furthermore, Liao et al. [50] reported that LINE-1 methylation levels, measured in leukocyte DNA, were significantly higher in renal cancer patients than in healthy subjects.

Two recent meta-analyses were conducted in order to estimate overall cancer risk according to global DNA hypomethylation in blood leukocytes. The meta-analysis by Woo and Kim [11] reports that global DNA hypomethylation of blood leukocytes was associated with increased cancer risk, although the association varied by the experimental methods used (% 5- methylcytosine method, LINE-1 with pyrosequencing and methyl acceptance assay), the region of DNA targeted and the cancer type. An updated meta-analysis performed by Brennan and Flanagan [16] indicates a significant inverse association between genomic 5methylcytosine levels and cancer risk (OR = 3.65; 1.20-6.09), but no overall risk association for studies using surrogates for genomic methylation, including methylation at the LINE-1 repetitive element (OR = 1.24; 0.76-1.72). Notably, the previous two meta-analyses included studies reporting association analysis between blood methylation levels and cancer risk but did not evaluate studies reporting differences in mean methylation levels in blood and in other tissues. The present meta-analysis of recent reports was conducted including studies reporting methylation levels in blood and in other tissues. This meta-analysis concerned 19 unique articles, but since two articles comprised more than one study conducted on different patient populations, altogether there were 29 non-unique studies included. On a total of 2554 samples from cancer patients and 3553 control samples, this meta-analysis reports that mean methylation levels in cancer patients were significantly 6.4% lower than in control samples.

The association between cancer risk and global DNA methylation has been mostly investigated in blood samples, because harvesting tumor tissue is invasive and cannot be routinely performed. However, several studies have reported that methylation of repetitive elements is tissue specific, most variable in tumor tissue, and is not correlated between tumor and blood [63-65]. Consistently, evidence reveals that genomic hypomethylation in tumor and normal adjacent tissue of bladder and colon cancer was not detectable in blood [43,66], suggesting that hypomethylation is restricted to the disease affected tissue. Interestingly, in the present meta-analysis the significant difference in mean methylation levels was confirmed only in tissue samples, both fresh/ frozen or FFPE specimens, but not in blood samples. Furthermore, the meta-analysis provided sufficient evidence that LINE-1 hypomethylation, significantly increases in colorectal and gastric cancer. On the contrary, no overall association was found for hepatocellular carcinoma. Notably, all studies focusing on colorectal and gastric tumors evaluated LINE-1 methylation in tissue samples, while all the included studies on hepatocellular carcinoma investigated the association only in blood leukocyte samples. Global DNA methylation can be measured by direct and indirect quantification assays. Although the measurement of percentages of 5- methylcytosine to estimate global DNA methylation contents are highly quantitative and reproducible, they require high amount of DNA and are not suitable for large epidemiological studies. Pyrosequencing with bisulfite-treated DNA, the "gold standard" for DNA methylation analysis [67,68], is a high-throughput and accurate method currently available to measure LINE-1 methylation as surrogate marker for global DNA hypomethylation. However, LINE-1 methylation

levels can vary depending on the target CpG sequence detected [69], representing an important factor in the association study with cancer risk. In the present meta-analysis, considering the two most frequently used detection methods (pyrosequencing and COBRA LINE-1) both subgroups report significantly lower LINE-1 methylation levels in cancer patients than in control samples, although heterogeneity between studies was significantly high in the *pyrosequencing* subgroup and moderately high in the *COBRA* subgroup.

The main limitations of this meta-analysis are the small number of studies included (n = 19) and the high heterogeneity across studies. Although a random effects model was performed, in order to take into account the high heterogeneity, the pooled estimates should be interpreted with caution. To overcome this issue, pooled estimates were calculated in more homogeneous subsets of studies (subgroups analysis). In addition, the possible existence of a publication bias was considered. Examination of funnel plots showed small to moderate asymmetry, suggesting that publication bias cannot be completely excluded and may have had at least a moderate impact on the true effect size estimates. In fact, some data, such as conference abstracts, non-English literature, unpublished data and other inconsistent reports according to our selection criteria were excluded. Furthermore, methylation-risk association tend only to be reported if it reveals statistically significant results, and if the authors deem analysis appropriate [16].

Moreover, since most studies (83%) had a case-control design large cohort studies are needed in order to clarify if global hypomethylation is an early cancer-causing aberration or an effect of carcinogenesis [11].

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In conclusion, the present meta-analysis adds new evidence to the growing literature on the role of LINE-1 hypomethylation in human cancer and shows that LINE-1 methylation levels were significantly lower in cancer patients than in controls, especially for certain cancer types. This result was confirmed in tissue samples but not in blood. Further studies are needed to better clarify the role of LINE-1 methylation in specific subgroups, considering both the cancer and sample type, and the methods of measurement.

Supporting Information

Checklist S1 PRISMA Checklist. (DOC)

Checklist S2 Meta-analysis on Genetic Association

Studies Checklist.

(DOC)

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Author Contributions

Conceived and designed the experiments: MV AA. Performed the experiments: MB AQ AM. Analyzed the data: MB AA. Contributed reagents/materials/analysis tools: MB AQ AM. Wrote the paper: MB MV AA.

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