



## Hydrogen sulphide and the hyperdynamic circulation in cirrhosis: a hypothesis

M R Ebrahimkhani, A R Mani and K Moore

*Gut* 2005;54;1668-1671; originally published online 20 Sep 2005;  
doi:10.1136/gut.2004.056556

---

Updated information and services can be found at:  
<http://gut.bmjournals.com/cgi/content/full/54/12/1668>

---

*These include:*

### References

This article cites 29 articles, 6 of which can be accessed free at:  
<http://gut.bmjournals.com/cgi/content/full/54/12/1668#BIBL>

### Rapid responses

You can respond to this article at:  
<http://gut.bmjournals.com/cgi/eletter-submit/54/12/1668>

### Email alerting service

Receive free email alerts when new articles cite this article - sign up in the box at the top right corner of the article

---

### Topic collections

Articles on similar topics can be found in the following collections

[Molecular Medicine](#) (999 articles)  
[Liver, including hepatitis](#) (829 articles)

---

### Notes

---

To order reprints of this article go to:  
<http://www.bmjournals.com/cgi/reprintform>

To subscribe to *Gut* go to:  
<http://www.bmjournals.com/subscriptions/>

## LEADING ARTICLE

## Hydrogen sulphide and the hyperdynamic circulation in cirrhosis: a hypothesis

M R Ebrahimkhani, A R Mani, K Moore

Gut 2005;54:1668–1671. doi: 10.1136/gut.2004.056556

Cirrhosis is associated with the development of a hyperdynamic circulation, which is secondary to the presence of systemic vasodilatation. Several mechanisms have been postulated to be involved in the development of systemic vasodilatation, including increased synthesis of nitric oxide, hyperglucagonaemia, increased carbon monoxide synthesis, and activation of  $K_{ATP}$  channels in vascular smooth muscle cells in the systemic and splanchnic arterial circulation. Hydrogen sulphide ( $H_2S$ ) has recently been identified as a novel gaseous transmitter that induces vasodilatation through activation of  $K_{ATP}$  channels in vascular smooth muscle cells. In this brief review, we comment on what is known about  $H_2S$ , vascular and neurological function, and postulate its role in the pathogenesis of the vascular abnormalities in cirrhosis.

cystathionine- $\beta$ -synthase being most prominent in the liver.<sup>5</sup>

$H_2S$  is permeable to plasma membranes as its solubility in lipophilic solvents is fivefold greater than in water.<sup>6</sup> It can be hydrolysed to hydro-sulphide and sulphide ions in the following sequential reactions:



However, even in an aqueous solution, approximately one third of  $H_2S$  remains undissociated at pH 7.4.<sup>6</sup> Cellular concentrations of  $H_2S$  are reported to be in the micromolar range (50–160  $\mu M$  reported in the brain and 45  $\mu M$  in plasma)<sup>4,7</sup> with a short half life due to its rapid reaction with haeme groups or disulphide containing proteins, or its oxidation to thiosulphate ( $S_2O_3$ ) and sulphate.<sup>6</sup> These relatively high concentrations, together with its short half life, suggest that generation or flux of  $H_2S$  is high. The amounts of urinary thiosulphate as well as sulphaemoglobin in erythrocytes are currently believed to be among the best markers of  $H_2S$  formation in vivo,<sup>2,6</sup> although these do have limitations, and recent studies have suggested that fluxes of  $H_2S$  can be measured using polarographic techniques.<sup>8</sup>

Endogenous gaseous transmitters such as nitric oxide (NO) and carbon monoxide (CO) constitute a unique class of mediators which play an important role in cell physiology. The high membrane permeability of these gases enables their rapid transfer across the cell membrane where they bind directly to the haeme group of guanylate cyclase or cytochrome oxidase, resulting in cell signalling in a receptor independent manner. A number of other biologically active gases such as nitrous oxide, ammonia, and hydrogen sulphide ( $H_2S$ ) may also participate in the regulation of cell function. Among them, recent reports have proposed  $H_2S$  as a novel endogenous transmitter with potential roles in both physiology and disease.

PHYSIOLOGICAL ACTIONS OF  $H_2S$  AND UNDERLYING MECHANISMS

The first and most important evidence for a physiological role of  $H_2S$  was obtained in 1989 when endogenous sulphide levels in rat brain tissues (1.6  $\mu g/g$ )<sup>9</sup> and in normal human post mortem brainstem (0.7  $\mu g/g$ ) were reported.<sup>10</sup> The study by Awata *et al* in 1995<sup>11</sup> provided the enzymatic mechanisms for this endogenous  $H_2S$  in rat brain, in which the activities of cystathionine- $\beta$ -synthase and cystathionine- $\gamma$ -lyase in six different brain regions were measured, with the activity of cystathionine- $\beta$ -synthase being >30-fold that of cystathionine- $\gamma$ -lyase.

FORMATION AND METABOLISM OF  $H_2S$ 

$H_2S$  is produced endogenously from desulphydration of cysteine (or cystine) by three different enzymes.<sup>1,2</sup> The reaction is catalysed by cystathionine- $\gamma$ -lyase (sometimes termed cystathionase), cystathionine- $\beta$ -synthase, or 3-mercapto-sulphurtransferase (fig 1). The first two enzymes are cytosolic haeme proteins, and the latter is a zinc dependent protein which is present in both the cytoplasm as well as mitochondria.<sup>2</sup> Cystathionase is currently the only identified  $H_2S$  generating enzyme present in the vasculature<sup>3</sup> whereas cystathionine- $\beta$ -synthase is the only  $H_2S$  generating system found in the nervous system.<sup>4</sup> However, all three enzymes are present in the liver and kidney, with

“There has been an explosion of interest in the biochemistry, physiology, and pharmacology of  $H_2S$ , which is rapidly emerging as a new biological mediator”

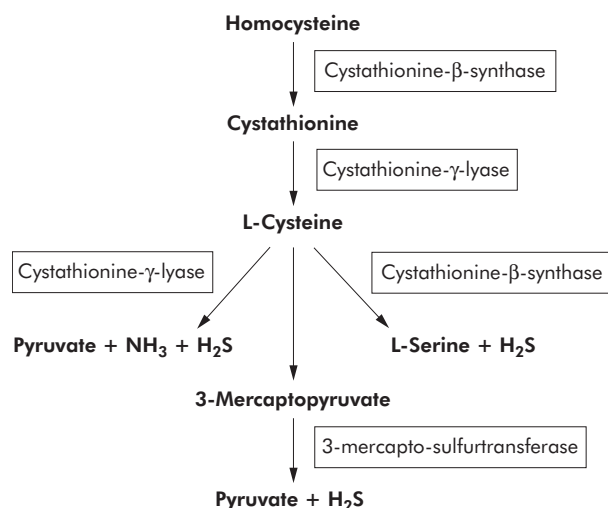
Further evidence for a physiological role of  $H_2S$  was reported by Abe and Kimura in 1996, who suggested that it may act as a neuromodulator as physiological concentrations of  $H_2S$  enhance glutamate mediated transmission via

**Abbreviations:**  $H_2S$ , hydrogen sulphide; NO, nitric oxide; CO, carbon monoxide; NMDA, N-methyl-D-aspartate

See end of article for authors' affiliations

Correspondence to: Professor K Moore, The UCL Institute of Hepatology, Royal Free and University College Medical School, University College London, Rowland Hill St, London NW3 2PF, UK; [kmoore@medsch.ucl.ac.uk](mailto:kmoore@medsch.ucl.ac.uk)

Revised version received 14 July 2005  
Accepted for publication 20 July 2005



**Figure 1** There are three enzymatic pathways involved in the synthesis of hydrogen sulphide ( $H_2S$ ) from cysteine in mammals. Of these only cystathionine- $\gamma$ -lyase is found in the vasculature. All three enzymes are present in the liver and kidney, with most activity residing in cystathionine- $\beta$ -synthase.

N-methyl-D-aspartate (NMDA) receptors which promote neuronal long term potentiation.<sup>4</sup> Since these initial publications there has been an explosion of interest in the biochemistry, physiology, and pharmacology of  $H_2S$ , which is rapidly emerging as a new biological mediator.

At present, the functional role of endogenous  $H_2S$  in the cardiovascular system is still the subject of much ongoing research and so far there have been no human studies on the physiological role of endogenously produced  $H_2S$  in the cardiovascular system. However, expression of cystathionine- $\gamma$ -lyase mRNA and endogenous production of  $H_2S$  have been demonstrated in the aorta,<sup>3,7</sup> mesenteric artery,<sup>12</sup> portal vein,<sup>3</sup> as well as cardiac tissue<sup>13</sup> in rats. Hosoki *et al* have demonstrated that  $H_2S$  could be produced in the portal vein, mesenteric artery, pulmonary artery, and thoracic aorta.<sup>3</sup>  $H_2S$  is only generated from vascular smooth muscle cells as no  $H_2S$  generating enzyme systems are expressed in the endothelial layer.<sup>7</sup> This is in contrast with NO and CO which can be produced from both endothelial and vascular smooth muscle cells. Moreover, unlike NO or CO,  $H_2S$  relaxed vascular tissues independent of activation of the cGMP pathway.<sup>8</sup>

" $H_2S$  induces vasorelaxation through activation of ATP sensitive  $K^+$  channels in vascular smooth muscles in vitro and in vivo"

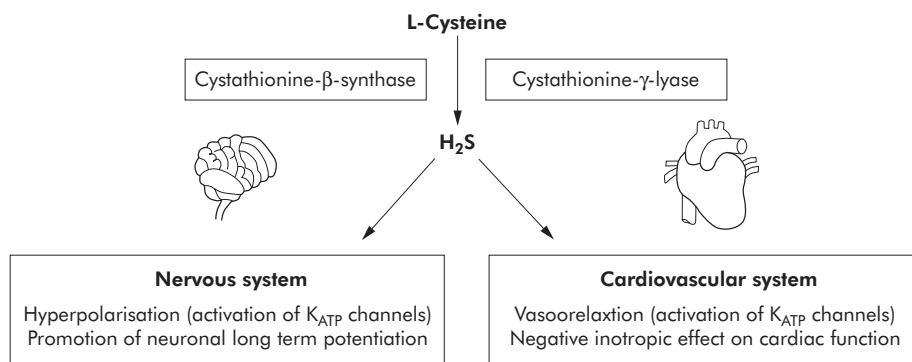
Whereas vasorelaxation induced by NO is virtually abolished by ODQ, a specific inhibitor of soluble guanylyl cyclase,  $H_2S$  induced vasorelaxation is not inhibited by ODQ.<sup>7</sup> The vasorelaxant activity of  $H_2S$  is mimicked by ATP sensitive  $K^+$  channel ( $K_{ATP}$ ) openers, and antagonised by glibenclamide (a  $K_{ATP}$  channel blocker).<sup>12</sup> In a series of studies, Wang and colleagues have shown that  $H_2S$  induces vasorelaxation through activation of ATP sensitive  $K^+$  channels ( $K_{ATP}$ ) in vascular smooth muscles in vitro and in vivo.<sup>7, 12, 13</sup>  $H_2S$  may also exert effects on adjacent cell types. Thus  $H_2S$  released from vascular smooth muscle cells may stimulate endothelial cells of small peripheral resistant arteries to release endothelium derived hyperpolarising factor which further hyperpolarises vascular smooth muscle cells and potentiates vascular relaxation.<sup>12, 13</sup> In vitro studies have also shown that  $H_2S$  exerts a negative inotropic effect on cardiac function, primarily through activation of  $K_{ATP}$  channels.<sup>14</sup> A summary of the major physiological effects of  $H_2S$  is presented at fig 2.

### INTERACTION OF $H_2S$ WITH NO AND CO

NO can regulate endogenous production of  $H_2S$  in vascular tissues by increasing cystathionine- $\gamma$ -lyase gene expression; this is evident by the fact that incubating cultured vascular smooth muscle cells with an NO donor significantly increased the transcriptional level of cystathionine- $\gamma$ -lyase.<sup>7</sup> On the other hand,  $H_2S$ , even at a very low concentration, can enhance relaxation of smooth muscle induced by NO by approximately 10-fold.<sup>3</sup> This effect is independent of free thiol groups as both cysteine and glutathione do not have such an effect.<sup>3</sup>

"There is a dynamic interplay between not only the  $H_2S$  and NO pathways but also between the  $H_2S$  and CO systems"

$H_2S$  has also recently been shown to upregulate CO synthesis through induction of haeme oxygenase.<sup>15</sup> Altered synthesis of  $H_2S$  may also affect the pulmonary circulation.<sup>15</sup> Thus Qingyou *et al* have shown that administration of sodium hydrosulphide (a donor of  $H_2S$ ) causes a decrease in pulmonary artery pressure in rats with hypoxic pulmonary hypertension, and administration of an inhibitor of cystathionine- $\gamma$ -lyase led to an increase in pulmonary artery pressure and a decrease in CO synthesis.<sup>15</sup> This suggests that there is a



**Figure 2** Major physiological actions of endogenously produced hydrogen sulphide ( $H_2S$ ). Activation of ATP sensitive  $K^+$  channels ( $K_{ATP}$ ) is a common mechanism of  $H_2S$  physiological effects, which induce vasorelaxation and neuronal hyperpolarisation in the cardiovascular system and nervous system, respectively.  $H_2S$  can also promote glutamate mediated transmission via N-methyl-D-aspartate receptors which enhance neuronal long term potentiation.

dynamic interplay between not only the H<sub>2</sub>S and NO pathways but also between the H<sub>2</sub>S and CO systems.

### POTASSIUM CHANNELS AND CONTROL OF VASCULAR FUNCTION IN CIRRHOSIS

Hypotension, low systemic vascular resistance, and reduced responsiveness to vasoconstrictors are all features of the hyperdynamic circulation in cirrhosis. These changes have been attributed to increased synthesis of NO, CO, anandamide, and calcitonin gene related polypeptide<sup>16–19</sup>; however, the precise mechanisms underlying the cardiovascular changes in cirrhotic subjects are not completely understood. In 1994, Moreau *et al* showed that there was activation of K<sub>ATP</sub> channels in vascular smooth muscle cells in rats with cirrhosis, and that this was partly responsible for the development of systemic vasodilatation in this animal model.<sup>20, 21</sup> In arterial smooth muscle cells, plasmalemmal K<sub>ATP</sub> channels play an important role in arterial vasodilatation by modulating membrane potential.<sup>22</sup> In cirrhosis, activation of K<sub>ATP</sub> leads to membrane hyperpolarisation which results in closure of the L-type Ca<sup>2+</sup> channel and subsequent decrease in Ca<sup>2+</sup> entry and vasorelaxation.<sup>20, 21</sup> One potential mechanism of K<sub>ATP</sub> channel activation involves prostaglandins such as prostacyclin as K<sub>ATP</sub> activation can be partially inhibited by cyclooxygenase inhibitors.<sup>20</sup> However, the observation that H<sub>2</sub>S can cause K<sub>ATP</sub> activation in a variety of experimental systems lends support to the idea that H<sub>2</sub>S may be involved in K<sub>ATP</sub> channel activation in cirrhosis.

### H<sub>2</sub>S AND THE HYPERDYNAMIC CIRCULATION

In this paper, we suggest that H<sub>2</sub>S may contribute to the pathogenesis of vascular dysfunction in cirrhosis (fig 3). This hypothesis is based on the following evidence.

- (1) Plasma H<sub>2</sub>S concentrations increase in rats with endotoxaemia.<sup>23</sup> Endotoxaemia is a common feature of cirrhosis<sup>24</sup> and high concentrations of circulating endotoxins are observed in cirrhotic patients with no clinical evidence of infection, and this may be due to impaired clearance of gut bacteria in cirrhotic liver.<sup>24, 25</sup> Studies are emerging which increasingly link the development of

extrahepatic complications of cirrhosis (for example, hyperdynamic circulation, cirrhotic cardiomyopathy, and hepatic encephalopathy) to the advent of endotoxaemia or sepsis in cirrhosis.<sup>26–28</sup> As endotoxin can induce the synthesis of H<sub>2</sub>S, this may have two consequences. Firstly, there may be increased H<sub>2</sub>S synthesis leading to increased K<sub>ATP</sub> activation in vascular smooth muscle cells and a resulting systemic vasodilatation. Secondly, increased H<sub>2</sub>S formation may lead to altered cardiac function as it has been shown that H<sub>2</sub>S exerts a negative inotropic effect on cardiac function, primarily through activation of K<sub>ATP</sub> channels.<sup>14</sup>

- (2) Increased synthesis of NO is well recognised in cirrhosis and portal hypertension,<sup>16</sup> and may lead to increased expression of cystathionine-γ-lyase, the main H<sub>2</sub>S producing enzyme in vascular smooth muscle cells.<sup>7</sup> Thus increased NO synthesis may enhance the formation of H<sub>2</sub>S in cirrhosis, thus leading indirectly to activation of K<sub>ATP</sub> channels.
- (3) Increased activity of serum cystathionine-γ-lyase has been demonstrated in rats with liver injury due to carbon tetrachloride.<sup>29</sup> Whether this is applicable to other forms of liver injury is unknown but increased cystathionine-γ-lyase activity would be expected to increase H<sub>2</sub>S formation in this model.<sup>23</sup>

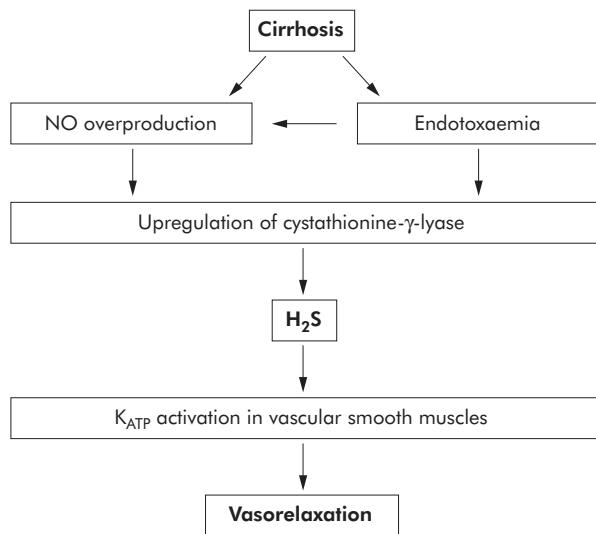
In conclusion, we propose a mechanism by which endotoxaemia, either alone or in combination with increased NO synthesis, leads to upregulation of cystathionine-γ-lyase activity and H<sub>2</sub>S synthesis. Increased synthesis of H<sub>2</sub>S leads to activation of K<sub>ATP</sub> channels and systemic vasodilatation (fig 3). Studies in the future will determine the validity of this hypothesis in humans.

### Authors' affiliations

**M R Ebrahimkhani**, Department of Pharmacology, School of Medicine, Tehran University of Medical Sciences, Tehran, Iran

**A R Mani, K Moore**, The UCL Institute of Hepatology, Department of Medicine, Royal Free and University College Medical School, University College London, London, UK

Conflict of interest: None declared.



**Figure 3** Postulated role of hydrogen sulphide (H<sub>2</sub>S) in the development of a hyperdynamic circulation in cirrhosis. Endotoxaemia leads to increased nitric oxide (NO) synthesis and upregulation of the enzyme responsible for H<sub>2</sub>S production (cystathionine-γ-lyase). H<sub>2</sub>S causes activation of K<sub>ATP</sub> channels which causes vasodilatation in liver disease. Increased NO synthesis may also cause vasodilatation directly.

### REFERENCES

- 1 Stipanuk MH, Beck PW. Characterization of the enzymic capacity for cysteine desulphhydration in liver and kidney of the rat. *Biochem J* 1982;**206**:267–77.
- 2 Kamoun P. Endogenous production of hydrogen sulfide in mammals. *Amino Acids* 2004;**26**:243–54.
- 3 Hosoki R, Matsuki N, Kimura H. The possible role of hydrogen sulfide as an endogenous smooth muscle relaxant in synergy with nitric oxide. *Biochem Biophys Res Commun* 1997;**237**:527–31.
- 4 Abe K, Kimura H. The possible role of hydrogen sulfide as an endogenous neuromodulator. *J Neurosci* 1996;**16**:1066–71.
- 5 Zhao W, Ndisang JF, Wang R. Modulation of endogenous production of H<sub>2</sub>S in rat tissues. *Can J Physiol Pharmacol* 2003;**81**:848–53.
- 6 Wang R. Two's company, three's a crowd: can H<sub>2</sub>S be the third endogenous gaseous transmitter? *FASEB J* 2002;**16**:1792–8.
- 7 Zhao W, Zhang J, Lu Y, *et al*. The vasorelaxant effect of H<sub>2</sub>S as a novel endogenous gaseous K(ATP) channel opener. *EMBO J* 2001;**20**:6008–16.
- 8 Doeller JE, Isbell TS, Benavides G, *et al*. Polarographic measurement of hydrogen sulfide production and consumption by mammalian tissues. *Anal Biochem* 2005;**341**:40–51.
- 9 Warenycia MW, Goodwin LR, Benishin CG, *et al*. Acute hydrogen sulfide poisoning: demonstration of selective uptake of sulfide by the brainstem by measurement of brain sulfide levels. *Biochem Pharmacol* 1989;**38**:973–81.
- 10 Goodwin LR, Francom D, Dieken FP, *et al*. Determination of sulfide in brain tissue by gas dialysis/ion chromatography: postmortem studies and two case reports. *J Anal Toxicol* 1989;**13**:105–9.
- 11 Awata S, Nakayama K, Suzuki I, *et al*. Changes in cystathionine gamma-lyase in various regions of rat brain during development. *Biochem Mol Biol Int* 1995;**35**:1331–8.
- 12 Cheng Y, Ndisang JF, Tang G, *et al*. Hydrogen sulfide-induced relaxation of resistance mesenteric artery beds of rats. *Am J Physiol Heart Circ Physiol* 2004;**287**:H2316–23.
- 13 Zhao W, Wang R. H<sub>2</sub>S-induced vasorelaxation and underlying cellular and molecular mechanisms. *Am J Physiol Heart Circ Physiol* 2002;**283**:H474–80.

- 14 **Geng B**, Yang J, Qi Y, *et al.* H<sub>2</sub>S generated by heart in rat and its effects on cardiac function. *Biochem Biophys Res Commun* 2004;**313**:362–8.
- 15 **Qingyou Z**, Junbao D, Weijin Z, *et al.* Impact of hydrogen sulfide on carbon monoxide/heme oxygenase pathway in the pathogenesis of hypoxic pulmonary hypertension. *Biochem Biophys Res Commun* 2004;**317**:30–7.
- 16 **Vallance P**, Moncada S. Hyperdynamic circulation in cirrhosis: a role for nitric oxide? *Lancet* 1991;**337**:776–8.
- 17 **Fernandez M**, Bonkovsky HL. Increased heme oxygenase-1 gene expression in liver cells and splanchnic organs from portal hypertensive rats. *Hepatology* 1999;**29**:1672–9.
- 18 **Batkai S**, Jarai Z, Wagner JA, *et al.* Endocannabinoids acting at vascular CB1 receptors mediate the vasodilated state in advanced liver cirrhosis. *Nat Med* 2001;**7**:827–32.
- 19 **Bendtsen F**, Schifter S, Henriksen JH. Increased circulating calcitonin gene-related peptide (CGRP) in cirrhosis. *J Hepatol* 1991;**12**:118–23.
- 20 **Moreau R**, Komeichi H, Kirstetter P, *et al.* Altered control of vascular tone by adenosine triphosphate-sensitive potassium channels in rats with cirrhosis. *Gastroenterology* 1994;**106**:1016–23.
- 21 **Moreau R**, Lebrec D. Endogenous factors involved in the control of arterial tone in cirrhosis. *J Hepatol* 1995;**22**:370–6.
- 22 **Quayle JM**, Nelson MT, Standen NB. ATP-sensitive and inwardly rectifying potassium channels in smooth muscle. *Physiol Rev* 1997;**77**:1165–232.
- 23 **Hui Y**, Du J, Tang C, *et al.* Changes in arterial hydrogen sulfide (H<sub>2</sub>S) content during septic shock and endotoxin shock in rats. *J Infect* 2003;**47**:155–60.
- 24 **Lin RS**, Lee FY, Lee SD, *et al.* Endotoxemia in patients with chronic liver diseases: relationship to severity of liver diseases, presence of esophageal varices, and hyperdynamic circulation. *J Hepatol* 1995;**22**:165–72.
- 25 **Lumsden AB**, Henderson JM, Kutner MH. Endotoxin levels measured by a chromogenic assay in portal, hepatic and peripheral venous blood in patients with cirrhosis. *Hepatology* 1988;**8**:232–6.
- 26 **Le Moine O**, Soupison T, Sogni P, *et al.* Plasma endotoxin and tumor necrosis factor-alpha in the hyperkinetic state of cirrhosis. *J Hepatol* 1995;**23**:391–5.
- 27 **Liu H**, Ma Z, Lee SS. Contribution of nitric oxide to the pathogenesis of cirrhotic cardiomyopathy in bile duct-ligated rats. *Gastroenterology* 2000;**118**:937–44.
- 28 **Bigatello LM**, Broitman SA, Fattori L, *et al.* Endotoxemia, encephalopathy, and mortality in cirrhotic patients. *Am J Gastroenterol* 1987;**82**:11–15.
- 29 **Taguchi T**, Awata S, Nishioka M, *et al.* Elevation of cystathionine gamma-lyase activity in the serum of rats treated with a single dose of carbon tetrachloride. *Ind Health* 1995;**33**:199–205.

## EDITOR'S QUIZ: GI SNAPSHOT .....

Robin Spiller, *Editor*

### An unusual cause of diarrhoea

#### Clinical presentation

A 70 year old Caucasian female presented with a six month history of intermittent diarrhoea, abdominal pain, and weight loss of 3 stone. Routine blood tests and liver ultrasound were normal.

#### Question

What abnormalities do the computed tomography scan (fig 1) and histology slide (fig 2) demonstrate? What was the cause of her diarrhoea?

See page 1713 for answer

This case is submitted by:

**A Sainsbury, D Clements**  
Airedale General Hospital, Yorkshire, UK

Correspondence to: Dr A Sainsbury, Airedale General Hospital, Skipton Rd, Yorkshire BD20 6TD, UK; anitabansal@yahoo.com

doi: 10.1136/gut.2005.068452



**Figure 1** Computed tomography scan of the abdomen.