

BIOLOGICAL SCIENCES: Medical Sciences

Pathogenetic mechanisms of AA amyloidosis

Short title: Pathogenesis of AA amyloidosis

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Abstract

Systemic AA amyloidosis is a serious complication of chronic inflammation. Serum amyloid A protein (SAA), an acute phase plasma protein, is deposited extracellularly as insoluble amyloid fibrils which damage tissue structure and function. Clinical AA amyloidosis is typically preceded by many years of active inflammation before presenting, most commonly with renal involvement. Using dose dependent, doxycycline inducible, transgenic expression of SAA in mice, we show that AA amyloid deposition can occur independently of inflammation, and that the time before amyloid deposition is determined by the circulating SAA concentration. High level SAA expression induced amyloidosis in all mice after a short, slightly variable delay. SAA was rapidly incorporated into amyloid, acutely reducing circulating SAA concentrations by up to 90%. Prolonged modest SAA overexpression occasionally produced amyloidosis after long delays and primed most mice for explosive amyloidosis when SAA production subsequently increased. Endogenous priming and bulk amyloid deposition are thus separable events, each sensitive to plasma SAA concentration. Amyloid deposits slowly regressed with restoration of normal SAA production after doxycycline withdrawal. Reinduction of SAA overproduction revealed that, following amyloid regression all mice were primed, especially for rapid glomerular amyloid deposition leading to renal failure, closely resembling the rapid onset of renal failure in clinical AA amyloidosis following acute exacerbation of inflammation. Clinical AA amyloidosis rarely involves the heart but amyloidotic SAA transgenic mice consistently had minor cardiac amyloid deposits, enabling us to extend to the heart the demonstrable efficacy of our novel antibody therapy for elimination of visceral amyloid.

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AA amyloidosis is a rare but important complication of otherwise treatable chronic inflammatory conditions (1) in which the amyloid deposits are derived from serum amyloid A protein (SAA) (2), an apolipoprotein of HDL. SAA is a non-specific acute phase protein, with a dynamic range of 1-> 3,000 mg/L in humans, synthesized by hepatocytes under the control of pro-inflammatory cytokines (3). The deposition of amyloid A (AA) type amyloid in the viscera, connective tissue and blood vessel walls damages tissue structure and function, with the kidney as the major target organ (4). Control of inflammation and hence of SAA production in patients with AA amyloidosis, arrests amyloid accumulation and enables its clinically beneficial regression (4, 5). Although clinical outcomes have improved markedly in recent years through better control of inflammation, AA amyloidosis is still fatal, and there are important gaps in our understanding of the pathogenesis of this disease. For example, the precise nature of the relationship between inflammation and amyloid deposition, and the factors governing the onset, progression and anatomical distribution of amyloid deposition remain unclear.

AA amyloidosis never occurs in the absence of inflammation and the associated acute phase response with increased circulating concentrations of SAA, but only a minority of individuals at risk ever develop AA amyloidosis. Persistently increased plasma SAA concentrations within the range typically seen in chronic inflammatory diseases are thus not generally sufficient for amyloid deposition. Furthermore since all previous animal models of AA amyloidosis depend on induction of chronic inflammation (6-9) to stimulate persistently increased SAA production, it has not hitherto been possible to identify the respective roles of inflammation and of increased abundance of SAA.

Human AA amyloidosis is typically preceded by many years of active inflammatory disease and the clinical presentation is then relatively acute, suggesting a slow or long delayed priming process and/or a rare stochastic event which eventually triggers amyloid deposition in some individuals. In mice greatly accelerated AA deposition can be produced by parenteral administration of an extract of amyloidotic tissue, so called “amyloid-enhancing factor”, the active component of which is amyloid fibrils themselves, followed by a single very aggressive acute phase stimulus. Deposition of AA amyloid in humans may thus also be triggered by such an amyloid fibril seed but it has not hitherto been possible to investigate this mechanism clinically. Another weakness of previous models is that while almost all AA amyloidosis patients present with or develop renal dysfunction, kidney involvement in murine AA amyloidosis is late and rarely causes renal failure. The new tractable and tunable transgenic model reported here, with hepatic SAA production induced independently of inflammation, by oral administration of doxycycline (10) allows the underlying mechanisms to be dissected for the first time.

Results

Regulated Overexpression of SAA in Mice. Without exposure to doxycycline, double transgenic mice carrying both a doxycycline-inducible SAA transgene (Fig. 1A) and a liver-specific reverse tet transactivator transgene (11), expressed SAA at the same baseline levels, ~ 10 mg/L, as wild type C57BL/6 mice. When exposed to drinking water containing doxycycline at 2 mg/mL, SAA production dramatically increased to values otherwise only seen in severe inflammation (Fig. 1B). Doxycycline had no effect on SAA values in non-transgenic and single-transgenic, SAA or rTA^{Lap-1}, mice but there was a clear dose response effect in

double-transgenic mice. Drinking water containing 0.1 mg/mL doxycycline induced SAA production to yield plasma values of 50-250 mg/L (Fig. 1C) while exposure to 2 mg/mL doxycycline induced values of > 1,000 mg/L (Fig. 1B). Peak SAA concentrations of 2,000-5,000 mg/L are much greater than seen in any other experimental or naturally occurring model of AA amyloidosis, although among ~700 patients with clinical AA amyloidosis we have seen values of ~2,500 mg/L in a few individuals. Induction of SAA expression by doxycycline was reversible, with the serum SAA concentration falling to baseline within 72 h of doxycycline withdrawal (Fig. 1D). As with acute phase SAA in wild type mice, the SAA produced in response to doxycycline was associated with HDL, eluting in size exclusion chromatography of whole serum in the trailing edge of the apoAI peak (Fig. 1E), consistent with the known displacement by SAA of apoAI from HDL particles (12).

Deposition of AA Amyloid Without Inflammation. Neither doxycycline alone, nor massively induced production of mouse SAA, caused increased production of murine SAP (Fig. S1). Mouse SAP is a major murine acute phase reactant which responds sensitively to almost all inflammatory or tissue damaging processes (13). Doxycycline-induced overexpression of SAA is thus totally independent of inflammation. Double-transgenic mice exposed to drinking water containing 2 mg/mL doxycycline developed major systemic amyloid deposits after median (range), 5 (4-16) wk as shown by massive whole body retention of intravenously administered ¹²⁵I-human SAP (¹²⁵I-hSAP), a validated, specific, quantitative, in vivo tracer for systemic amyloid deposits in mouse (14, 15) and man (16-18). Heavy AA amyloid deposits were confirmed histologically in spleen and liver, correlating closely with the major organ localization of ¹²⁵I-hSAP. Significant minor amyloid deposits were also present in kidneys, heart and gut

(Fig. S2). The total amyloid load was much greater than we have ever seen, or has been previously reported, but organ function was preserved (Table S1). A substantial proportion, 13 of 60, of these amyloidotic mice had normal SAP concentrations ≤ 10 mg/L), showing unequivocally that amyloid deposition can occur in the absence of inflammation. The remaining amyloidotic animals had only modestly elevated SAP concentrations, probably as a result of tissue damage caused by the massive amyloid deposits. For example, in one experiment the SAP values were mean (SD), 17.0 (5.9) mg/L in mice after deposition of amyloid compared to 7.0 (1.0) mg/L in the same animals after induction of SAA expression but before amyloid deposition ($n = 18$).

Latency of Amyloid Deposition. Induction of murine AA amyloidosis by sustained chronic inflammation typically takes several weeks (7). However mice in which AA amyloid deposits have regressed following remission of chronic inflammation are primed for rapid reaccumulation when they receive a new inflammatory stimulus (19) and previously untreated mice are readily primed by intravenous injection of amyloidotic tissue extracts or isolated amyloid fibrils, so-called amyloid enhancing factor (AEF) (20, 21). In double-transgenic mice, a single AEF injection 4 d after starting on drinking water containing high dose doxycycline at 2 mg/mL, produced heavy amyloid deposits just 2 d later. By 21 d there was massive amyloidosis in all animals tested, with markedly increased liver and spleen sizes (Fig. 2). In contrast, the median time before amyloid deposition was detected in mice receiving doxycycline treatment alone was 5 wk, demonstrating the requirement, even in the face of abundant SAA, for a delayed and/or rare seeding or triggering event.

When only 0.1 mg/mL doxycycline was present in the drinking water, inducing more modestly increased mouse SAA expression and a mean (SD) concentration of 120 (52) mg/L

($n = 10$), no amyloid was detected by ^{125}I -hSAP retention in over a year. One mouse then became amyloidotic between 58 and 66 wk, resembling the long latency and low incidence of clinical AA amyloidosis. In principle, these findings could reflect delayed seeding of amyloid deposition, insufficient SAA expression to permit net accumulation of amyloid, or both. When mice received AEF and low dose doxycycline at 0.1 mg/mL in their drinking water, only 1 of 10 animals developed amyloidosis between 9-13 wk, 2 became amyloidotic after 34-50 wk, and 7 had no detectable amyloid at 58 wk. The drinking water doxycycline concentration was then increased to 2 mg/mL and all 7 mice became amyloidotic within 4 d, demonstrating both that the primed state persisted, and that these animals had not previously become amyloidotic because SAA abundance was limiting. In contrast, when 8 mice which had received water containing 0.1 mg/mL of doxycycline for 21 mo were switched onto 2 mg/mL, 4 of them promptly developed amyloidosis within 4 d, and a further 2 did so at 7 d while 2 others remained amyloid free at 7 d. Amyloid was deposited significantly later in a control cohort of previously unexposed, double-transgenic, age matched animals started on water containing 2 mg/mL doxycycline at exactly the same time as the dose was increased in the long term low dose group (Fig. S3; $P = 0.022$). These results clearly differentiate between endogenous priming for amyloid deposition and bulk amyloid accumulation and show that priming can occur when levels of SAA are elevated modestly and insufficiently for net amyloid deposition. Sustained modest elevation of circulating SAA concentration does not potently initiate or promote amyloid deposition but can cause amyloidosis occasionally and also primes for rapid deposition if SAA production sharply rises.

Sequestration of Circulating SAA During Active Amyloid Deposition. While amyloid was being deposited, between 2-4 d following AEF injection in mice on 2 mg/mL doxycycline, we observed a precipitous ~ 90% fall in circulating SAA concentrations (Fig. 3A). This was not an effect of exogenous AEF as SAA values fell similarly in mice which, after long term treatment with 0.1 mg/mL doxycycline, were then given 2 mg/mL doxycycline and developed amyloid within 4 d (Fig. 3B). SAA values did not fall in mice which did not develop amyloid despite receiving identical treatment (Fig. 3C). SAA mRNA abundance in the livers of doxycycline treated mice did not differ between those receiving AEF and controls (Fig. S4A) despite the sharp fall in circulating SAA concentration during amyloid deposition in the AEF treated group (Fig. S4B). The evident rapid sequestration of SAA into nascent amyloid deposits was quantitatively consistent with the increase in amyloidotic organ weights. Assuming a peak steady state SAA concentration of 2 mg/mL, blood volume 1.5 mL, and SAA $t_{1/2}$ 90 min (22), maximal SAA production is 24 mg daily. Assuming constant SAA secretion rate and otherwise normal turnover, the 90% fall in circulating SAA concentration during amyloid deposition corresponds to loss of 21.6 mg daily from the plasma. The total of ~0.45 g over 3 wk, based on conservative assumptions, is consistent with the observed ~2 g increase in liver weight of mice after AEF and 3 wk on doxycycline, considering that amyloid deposits contain abundant proteoglycans and SAP as well as amyloid fibrils (1).

Amyloid Regression, Recurrence and Onset of Renal Failure. Consistent with clinical experience, amyloid load diminished very slowly after withdrawal of doxycycline, despite prompt extinction of SAA transgene expression. Retention of ^{125}I -hSAP took at least 4 mo to return to baseline, remained abnormal after > 1 yr in some animals, and specks of typical

Congophilic amyloid (23), undetected by the whole body method, were sometimes present histologically. After 6 and 17 mo off doxycycline, re-exposure of previously amyloidotic mice to drinking water containing 2 mg/mL of doxycycline triggered extremely rapid new amyloid deposition with interestingly different pathological and clinical features. There were massive typical splenic marginal zone and inter-follicular deposits (Fig. 4A and B) but hepatic amyloid was predominantly in and around portal tracts and central veins rather than scattered in the parenchyma (Fig. 4C and D). Most strikingly, major renal glomerular amyloidosis developed (Fig. 4E-H). The glomerular amyloid deposits were far more abundant than we have previously observed in mice and remarkably, all glomeruli were affected. Renal function monitored by serum cystatin C assay (24) was normal before amyloid regression (Fig. 4I) or immediately prior to doxycycline re-exposure but was markedly impaired within 3d of re-induction of SAA expression and the ensuing renewed amyloid deposition (Fig. 4J and K).

Therapeutic Reduction of Cardiac Amyloid Deposits. We have recently shown that administration of anti-human SAP antibodies to AA amyloidotic mice in which the amyloid deposits had been loaded with human SAP, leads to macrophage mediated elimination of hepatic and splenic amyloid (25). In contrast to the standard mouse AA amyloidosis model, cardiac amyloid is consistently present in the new SAA transgenic model. SAA transgenic mice with abundant amyloid after exposure to doxycycline were therefore given 3 daily parenteral injections of human SAP to load up the amyloid deposits and 5 d after the last injection, when all human SAP had left the circulation, they were treated with monoclonal mouse IgG2a anti-human SAP antibodies. After a single dose of anti-SAP, spleen and liver amyloid deposits were, as usual, substantially reduced (both $P < 0.001$ compared to amyloidotic controls not given the

antibody), but there was no significant difference in extent of cardiac amyloid. However after a second treatment with SAP and anti-SAP one month later, the cardiac amyloid deposits were substantially diminished ($P = 0.014$; Fig. 5).

Discussion

Systemic AA amyloid deposition is always accompanied by increased SAA production, whether triggered by tissue damage, infection or abnormal pro-inflammatory cytokine activity (26-30). The transgenic mice reported here allowed us to induce expression of the amyloidogenic mouse SAA isoform, SAA2, by oral administration of doxycycline, completely independently of any inflammatory stimulus. Neither doxycycline exposure nor induction of SAA production itself triggered an acute phase response of other proteins. Thus, contrary to claims in the literature, circulating mouse SAA, like authentic human SAA (31), is clearly not intrinsically pro-inflammatory. Furthermore, although AA amyloidosis has hitherto been inextricably linked to systemic inflammation, the present results demonstrate that isolated overproduction of SAA, without inflammation, is sufficient.

In common with models in which AA amyloidosis is evoked by potently inducing inflammation, transgenic mice induced to express SAA at high levels deposited amyloid after a variable latent period, and the process was accelerated and made absolutely consistent by injection of AEF, containing preformed amyloid fibrils which seed amyloid fibrillogenesis.

In contrast with experimental AA amyloidosis in mice, human AA amyloidosis affects only a small minority of patients with inflammatory diseases, usually presents long after the onset of inflammation (median (range) 17 (0-68) yr) (4, 5), most commonly with proteinuria and/or renal failure (4, 5), and plasma SAA concentrations, median (range), 28 (1-1610) mg/L (4, 5), are typically more modest than in the mouse model. The reasons why some patients

develop amyloidosis and most others do not, are obscure. The ability to dissociate priming and deposition experimentally raises the possibility that either (or both) of these could be limiting in patients. Using low dose doxycycline to induce modestly increased SAA production we replicated the long latency of the human disease and the occasional late onset development of amyloidosis, resembling the low frequency of its incidence in patients. Amyloid deposition occurred only occasionally in mice in which prolonged modest overexpression of SAA was induced after priming with AEF. Thus, SAA levels can be elevated for an extended period and yet be insufficient to drive net accumulation of amyloid, whether or not amyloid deposition has been primed. While extended modest overproduction of SAA did not inexorably lead to amyloid deposition, some animals which remained free of detectable amyloid were primed for amyloid deposition when SAA expression was increased. If these findings accurately reflect amyloid pathogenesis in patients, a significant proportion of individuals with prolonged chronic inflammatory disease may already be primed, and amyloid deposition may then be precipitated by an episode of severe inflammation. In patients with a long history of chronic inflammation, it would be wise to treat episodes of severe inflammation aggressively and without delay. Prompt evaluation of such patients may lead to early detection of AA amyloidosis, perhaps before the onset of significant renal failure.

Mice have two distinct acute phase SAA proteins, SAA1 and SAA2, encoded by different genes and produced in roughly equal amounts (2) but only SAA2 is amyloidogenic. As wild type mice with chronic inflammation become amyloidotic, the SAA1:SAA2 concentration ratio increases sharply and the total circulating SAA value falls modestly, consistent with selective sequestration of SAA2 into amyloid (32, 33). Our present finding of a dramatic fall of SAA2 values in all the transgenic mice as amyloid was being deposited was similarly consistent with

such sequestration. In man, there are also two different acute phase SAA proteins (34) encoded by different genes but, unlike in the mouse, both human acute phase SAA proteins are deposited as amyloid fibrils (35). Our transgenic model, overexpressing only amyloidogenic murine SAA2, thus more closely resembles the human situation than studies in wild type mice and the present finding of rapid depletion of circulating SAA may have important implications for clinical management. Plasma SAA values in AA amyloidosis patients correlate closely with progression of amyloid deposition, morbidity and mortality (4, 5) and the aim of treatment is to maximally reduce the SAA concentration. However the transgenic mouse results now suggest that SAA values may be misleadingly low during active amyloid deposition, perhaps explaining clinical observations of amyloid accumulation in the face of modest plasma SAA concentrations (4). If such depression of plasma SAA does occur in patients, is likely to be most significant in those who have the heaviest amyloid loads because the large amounts of amyloid in those individuals will provide an abundant sink for circulating SAA.

The most common important clinical consequence of AA amyloidosis is renal dysfunction, with > 10% of patients having end-stage renal failure at first diagnosis (4). Renal failure developed very rapidly upon re-induction of SAA expression in transgenic mice in which amyloid had previously been induced and spontaneously regressed. This closely parallels the rapid onset or relapse of renal failure precipitated by intercurrent flares of inflammation in some AA amyloidosis patients in remission (4, 5), and suggests that glomerular amyloid deposition may be responsible, as in the transgenic mice. The factors which determine the sites of amyloid deposition in general are unknown but the dramatically altered pattern of amyloid reaccumulation following regression could reflect a different distribution of amyloid template. Amyloid regresses much more rapidly in the spleen and liver than in the kidney, making the

latter more susceptible to amyloid deposition when SAA production increases. Thus, the presence of large amounts of template in spleen and/or liver may, through sequestration of SAA, limit the rate of deposition in the kidney. A related phenomenon may explain the discrepancy between clinical AA amyloidosis which is almost invariably accompanied by renal dysfunction, and experimental murine AA amyloidosis, in which kidney function is rarely affected. Experimental amyloidosis is usually induced rapidly in a single phase whereas clinical amyloidosis typically occurs after many years of inflammatory disease, providing opportunity for cycles of amyloid deposition and regression.

As well as providing insights into aspects of the pathogenesis of AA amyloidosis, amyloidotic transgenic mice have proved to be a valuable model for applied studies, and they offer some significant advantages over other models. Although cardiac involvement is very rare in clinical AA amyloidosis (4), the transgenic mice reproducibly developed cardiac amyloid deposits. The heart is usually involved in systemic monoclonal immunoglobulin light chain (AL) disease, which is by far the most common type of human amyloidosis. Cardiac amyloid of transthyretin type is also common in the elderly (36, 37), although until recently it was rarely diagnosed in life. The development of cardiac amyloidosis in the transgenic model allowed us to show that our recently developed antibody therapy (25) effectively reduces cardiac amyloid load, as it does amyloid in the spleen and liver. The present mouse model is also proving useful for development of novel non-invasive in vivo amyloid imaging methods (38).

Methods

Transgenic Mice. The SAA transgene comprises the tetracycline-responsive promoter from pTRE-myc (Clontech) linked within their respective 5' untranslated regions to a 3.7 kb genomic DNA fragment containing the amyloidogenic mouse SAA isoform SAA2 (2), also known as SAA1.1 (9), with 1.25 kb of 3' flanking sequence (ENSEMBL id. ENSMUSG00000057465). The transgene fragment was isolated from the plasmid vector and microinjected into pronuclei of 1-cell eggs obtained from C57BL/6 X CBA F₁ donors. Transgenic mice were identified by PCR with primers GTGTACGGTGGGAGGCCTAT and CCAGGAGGTTCCACTGAGAG from the promoter and the SAA gene, respectively. Ten lines of transgenic mice were established and crossed with rTA^{Lap}-1 reverse tet-transactivator (rtTA) mice (11), which express the reverse tet transactivator rTA2^S-S2 (39) in hepatocytes, obtained from the European Mouse Mutant Archive (stock number EM:00404). Presence of the rtTA transgene was assayed by PCR using primers CTGGGAGTTGAGCAGCCTAC and AGAGCACAGCGGAATGACTT. All of the reported work was done with a single line of mice selected because, in conjunction with rTA^{Lap}-1, they had normal baseline serum SAA values when untreated with doxycycline and highly inducible SAA expression when doxycycline was administered. Double transgenic mice were backcrossed onto the C57BL/6J background. All experiments reported here used hemizygous transgenic mice.

Induction of SAA Expression and of Amyloid Deposition. Doxycycline hydrochloride (Sigma D9891) was administered in drinking water containing 5% sucrose. Amyloid loads were assessed essentially as described (14) by whole-body retention of ¹²⁵I-hSAP 24 and 48 h after injection of tracer; 0.006% w/v KI was included in the drinking water to prevent retention of free ¹²⁵I iodide. AEF (20, 21) was administered by tail vein injection. Abundant amyloid deposits were reliably and consistently produced by a single AEF injection and administration of doxycycline; e.g. 2mg/mL doxycycline was administered for 3 d before and 7 d after AEF injection. The inconvenience of using (double) transgenic mice is mitigated because we have bred mice homozygous for both transgenes. This refined transgenic model of AA amyloidosis minimizes the severity of experimental animal procedures because it eliminates the need for injections to stimulate severe or chronic inflammation. All animal studies were ethically reviewed and carried out in accordance with European Directive 86/609/EEC.

Biochemical Assays. Commercial ELISA assays were used for SAA (Tridelata or Immune Systems Ltd.), apolipoprotein AI (USCN Life Science Inc.) and cystatin C (Biovendor), and were performed according to the manufacturers' instructions. Mouse SAP was assayed by electroimmunoassay with monospecific sheep anti-mouse SAP antiserum (13). All other assays were performed at MRC Harwell using an Olympus AU400 automated clinical chemistry analyzer. Size exclusion chromatography was performed using a Superdex 200 column on the ÄKTA Explorer apparatus (GE Healthcare). The column was equilibrated and eluted at 0.5 mL/min with PBS buffer containing 5 mM EDTA, pH 7.4.

qRT-PCR. Assay of SAA mRNA levels was performed by qRT-PCR using transgene-specific primers ATAGAAGACACCGGGACCGAT and TAGCTTCATCCTGCTGGTGTCT, specific for the tet-responsive promoter and the SAA gene, respectively. The results were normalized against reference genes SDHA (primers TCATCACAGAAGGGTGTTCGT and GCAACAGGTGCGTATCTCTC) and HPRT (primers TCCTCCTCAGACCGCTTTT and ACCTGGTTCATCATCGCTAATC).

Histology. Formalin fixed wax-embedded tissues were sectioned at 7 μ m, stained with alkaline alcoholic Congo red (23) and viewed under strong cross polarized light.

Antibody Therapy. Amyloidosis was induced in female transgenic mice by treatment with AEF and doxycycline, and verified by assaying whole body retention of 125 I-hSAP. For therapy, the amyloid deposits were loaded with unlabelled human SAP by injection of 10 mg per day for 3 d. After a delay of 5 d to allow elimination of circulating hSAP, the mice were injected with 5 mg of monoclonal mouse anti-hSAP antibody (clone SAP-5 supplied by GlaxoSmithKline) (25). Amyloid load was scored as described (15); the values reported are the average of the scores assigned by three expert individuals who were blinded to the treatments given.

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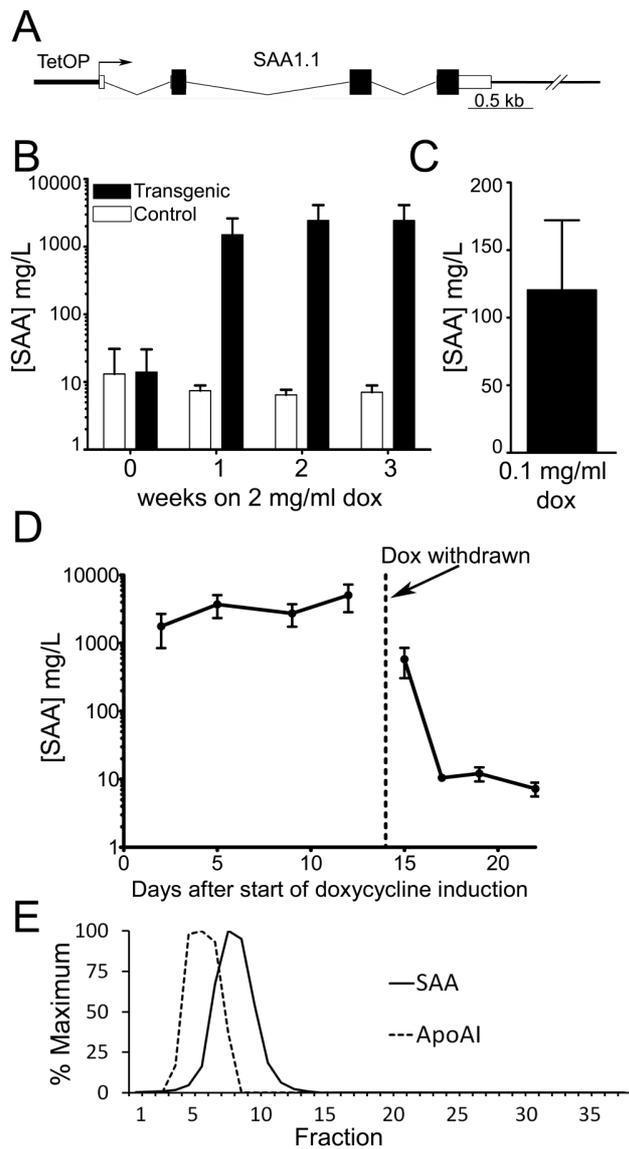


Fig. 1. Inducible expression of SAA. (A) Schematic of the doxycycline-responsive SAA transgene. (B) Sustained high-level expression of SAA specifically in transgenic mice on drinking water containing 2 mg/mL doxycycline (mean \pm SD; $n = 5$ transgenic, $n = 6$ controls). (C) Modest SAA expression in transgenic mice on drinking water containing 0.1 mg/mL doxycycline (mean \pm SD; $n = 10$ (5 ♂; 5 ♀)). (D) SAA concentrations rapidly return to normal baseline values following doxycycline removal (mean \pm SD; $n = 8$ (4 ♂; 4 ♀)). (E) Elution profile of SAA and apoAI in size exclusion chromatography of whole serum from mouse with high level transgenic SAA expression.

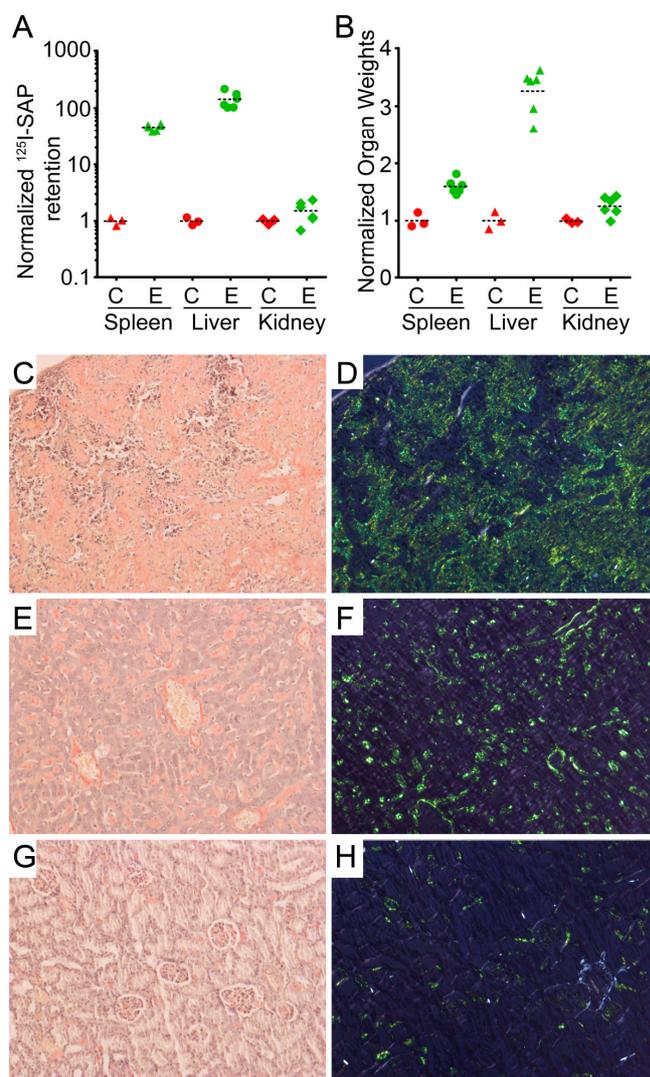


Fig. 2. Amyloidosis in SAA overexpressing transgenic mice. (A) Retention of ¹²⁵I-human SAP in organs of control (C) and experimental transgenic (E) mice treated with doxycycline for 3 wk after AEF injection. The ¹²⁵I retained in spleens, livers and kidneys was counted 24 h after injection of 10⁶ cpm of ¹²⁵I-labeled human SAP. The counts retained are shown normalized to the means of the control values (spleen 8.1 × 10² cpm; liver 4.5 × 10³ cpm; kidney 2.8 × 10³ cpm). (B) Weights of amyloidotic and control mouse organs normalized to the means of the control values (spleen 0.072 g; liver 0.846 g; kidney 0.265 g). (C-H) Representative histology of amyloidotic spleen (C and D), liver (E and F) and kidneys (G and H) of transgenic mice stained with Congo red. C, E, G: bright field; D, F, H: crossed polarized light.

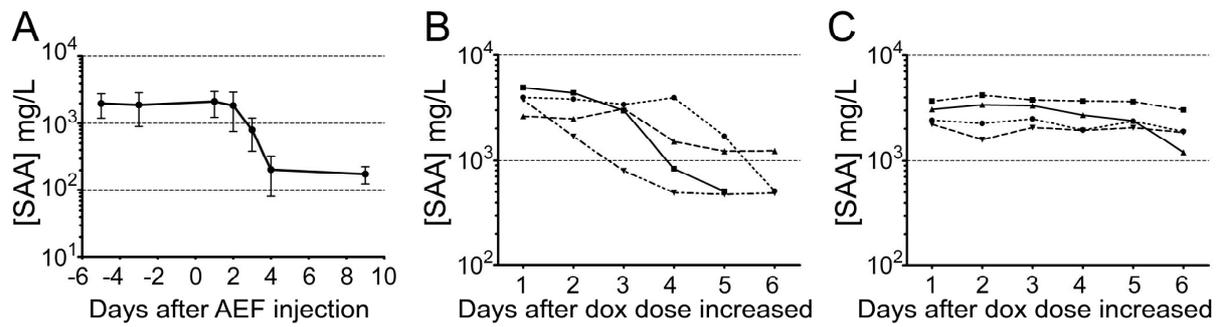


Fig. 3. Circulating SAA concentration falls during amyloid deposition. (A) SAA concentrations fell sharply following AEF injection as amyloid was deposited (note the logarithmic scale for SAA values; mean \pm SD; $n = 8$; $P < 0.0001$, repeated measures ANOVA). (B and C) Circulating SAA concentrations following increase of drinking water doxycycline content to 2 mg/mL in mice treated long-term with water containing 0.1 mg/mL doxycycline. (B) Mice amyloidotic by day 4: SAA concentrations fell ($n = 4$; $P = 0.0062$, repeated measures ANOVA). (C) Mice not detectably amyloidotic on day 4: SAA concentrations were stable ($n = 4$; $P = 0.3378$, repeated measures ANOVA).

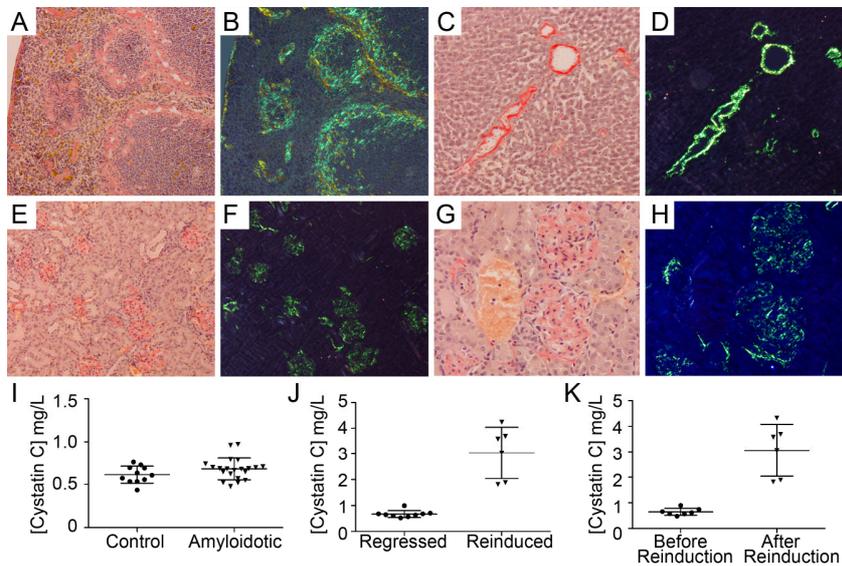


Fig. 4. Remission and relapse lead to glomerular amyloidosis and renal failure. Congo red staining of reaccumulated amyloid in spleen (*A* and *B*), liver (*C* and *D*) and renal cortex (*E* to *H*) showing widespread, heavy glomerular amyloid deposits; bright field illumination (*A*, *C*, *E*, *G*), crossed polarized light (*B*, *D*, *F*, *H*). (*I*) Normal serum cystatin C in mice amyloidotic for the first time (mean \pm SD; $n = 11$ controls; $n = 21$ amyloidotic; $P = 0.135$; t-test). (*J* and *K*) Increased serum cystatin C concentration caused by renal failure in mice with massive glomerular amyloid deposition after reinduction. (*J*) Comparison of reinduced ($n = 6$) and control ($n = 5$) groups (mean \pm SD; $P = 0.0025$; Mann-Whitney test). (*K*) Before and after reinduction values in individual mice ($n = 6$; mean \pm SD; $P = 0.0156$; Wilcoxon matched-pairs signed rank test).

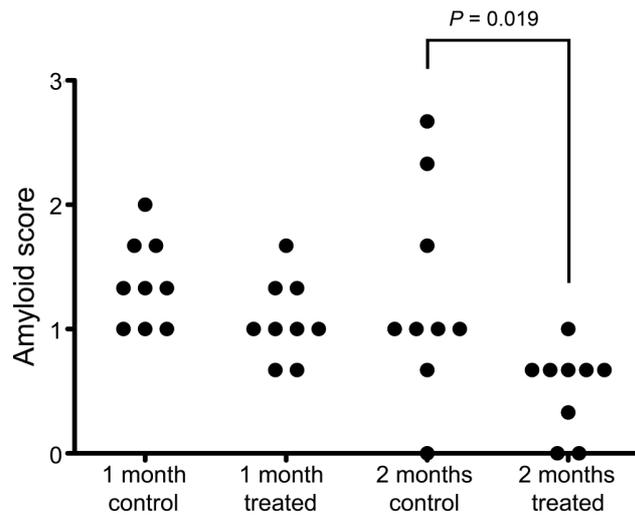
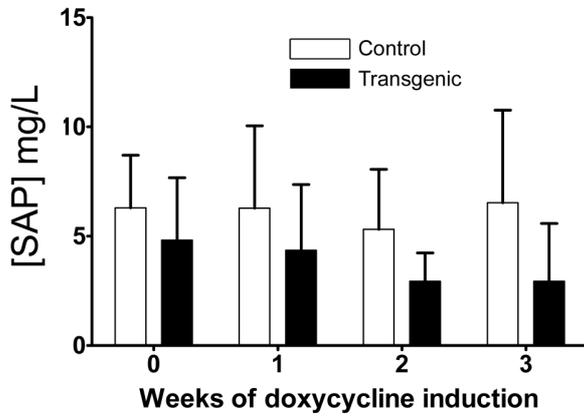
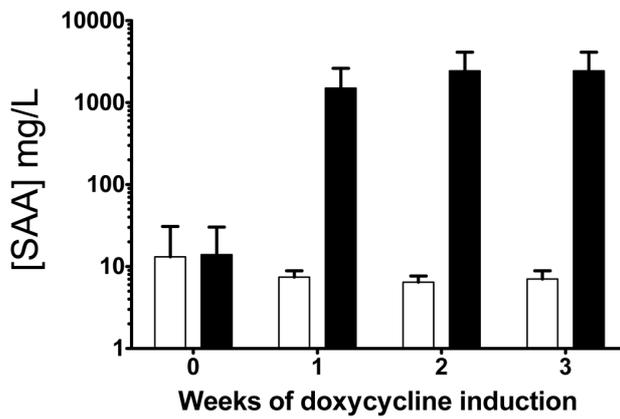
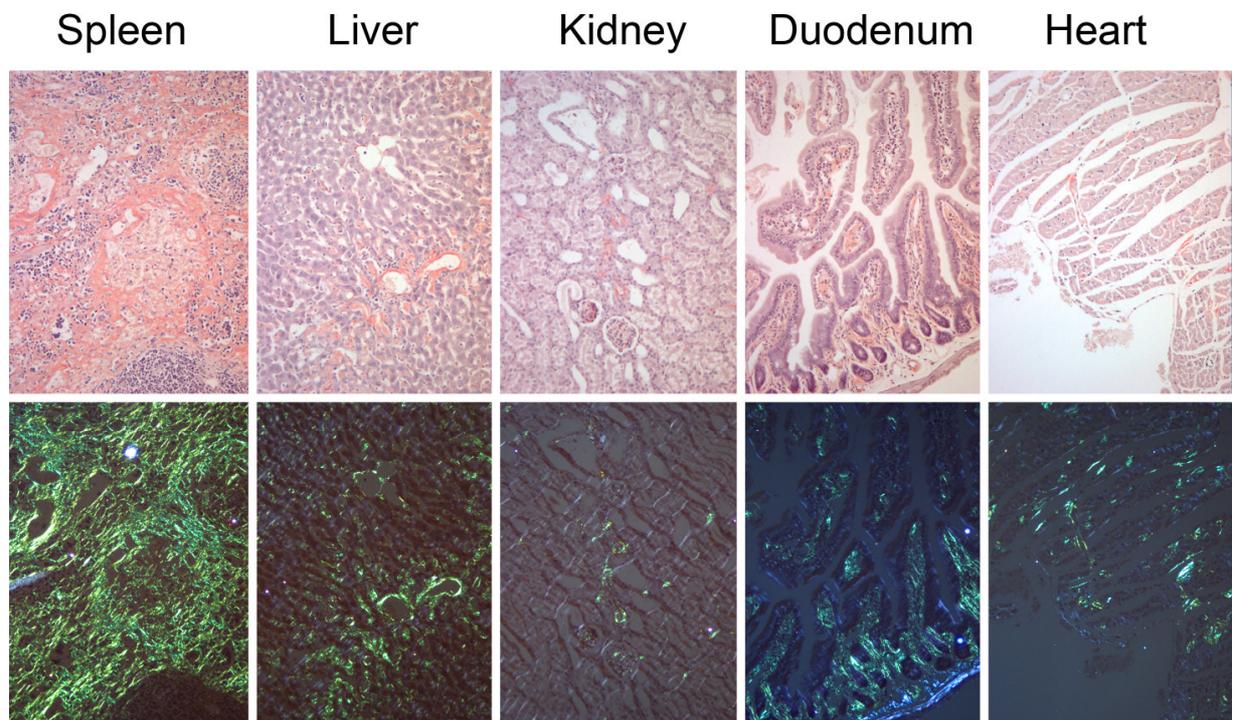


Fig. 5. Reduction of cardiac amyloid deposits following antibody therapy. Amyloidotic mice received an injection of human SAP to load the amyloid deposits, followed by injection of monoclonal anti-human SAP antibody. Control amyloidotic mice received no treatment. Half the treated and control mice were killed 30 d later and the remaining treated animals received a second identical treatment. All remaining mice were then killed after a further 30 d. Amyloid scores of individual mice are plotted. Kruskal-Wallis test $P = 0.002$; multiple comparison test: one anti-SAP dose vs control, $P = 0.38$; two anti-SAP doses vs control, $P = 0.019$.

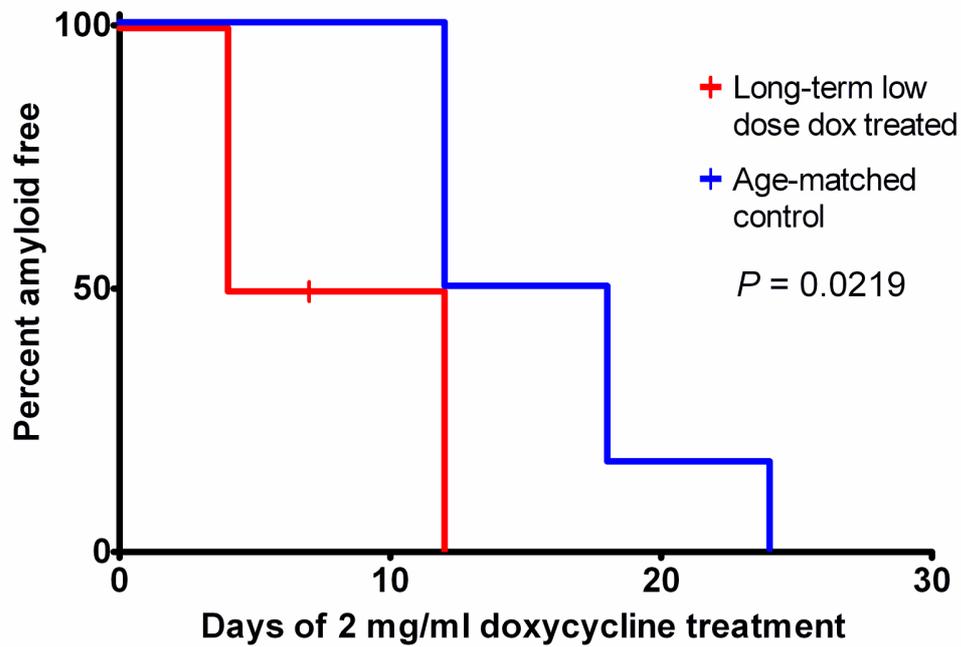
A**B**

Supplemental Figure 1. Lack of systemic inflammation on induction of SAA expression.

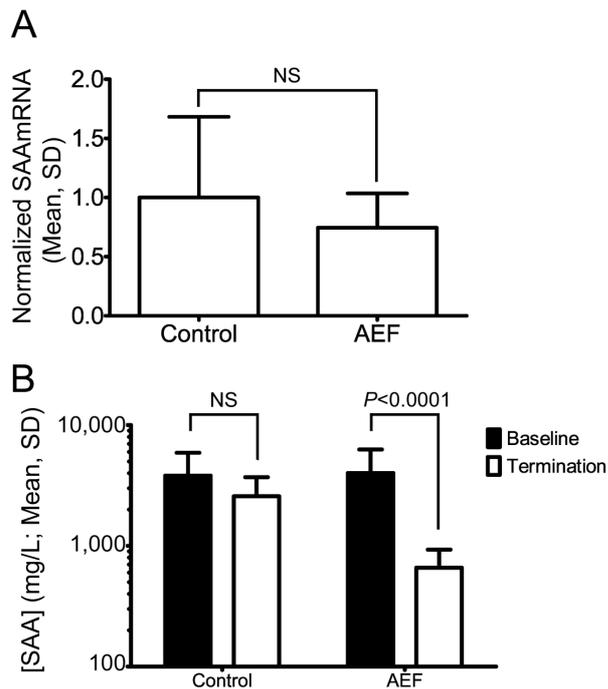
(A) Serum concentrations of SAP (mean \pm SD) remained unchanged when double transgenic ($n = 5$) or control ($n = 7$) mice were treated with 2 mg/mL doxycycline. **(B)** Serum SAA concentrations (mean \pm SD) in the same animals showing normal baseline SAA concentrations before exposure to doxycycline and sustained high-level SAA production when receiving doxycycline.



Supplemental Figure 2. Representative histology of amyloidotic transgenic mice. Congo red staining shows massive amyloid deposits in spleen and liver, with minor but significant deposits in kidney, duodenum and heart. Above, bright field; below, crossed polarized illumination showing the pathognomonic green birefringence of amyloid.



Supplemental Figure 3. Priming induced by long-term low-level overexpression of SAA. Kaplan-Meier curves showing the accelerated development of amyloidosis induced by 2 mg/mL doxycycline treatment in mice that had been maintained for 21 mo on 0.1 mg/mL doxycycline ($n = 8$), compared to age-matched controls maintained on water ($n = 6$).



Supplemental Figure 4. The fall in circulating SAA concentration during amyloid deposition is not caused by reduced gene expression. **(A)** mRNA levels were no different between the AEF-injected and control groups ($P = 0.149$; unpaired t test with Welch's correction, one-tailed). **(B)** Baseline concentration of SAA in serum of mice after induction with 2 mg/mL doxycycline and SAA values 4 d after injection of AEF or vehicle (note the logarithmic scale; $n = 10$ each group). Circulating SAA concentrations fell only in the AEF-injected group ($P < 0.0001$; repeated measures 2-way ANOVA).

Supplementary Table 1. Serum biochemistry results.

	Sodium (mmol/L)		Potassium (mmol/L)		Chloride (mmol/L)		Urea (mmol/L)		Creatinine (μmol/L)	
	Control (n = 10)	Amyloidotic (n = 8)	Control (n = 10)	Amyloidotic (n = 8)	Control (n = 10)	Amyloidotic (n = 8)	Control (n = 10)	Amyloidotic (n = 8)	Control (n = 9)	Amyloidotic (n = 8)
Mean	153.2	155.8	4.33	4.025	111.5	107.3	7.34	6.975	15.4	17
SD	3.19	5.97	0.1829	0.2252	4.062	4.4	0.9709	1.547	2.956	4.66
P value	0.2818		0.0059		0.0494		0.5487		0.6296	
Test	Mann-Whitney		unpaired t test		unpaired t test		unpaired t test		Mann-Whitney	

	Calcium (mmol/L)		Inorganic phosphate (mmol/L)		ALP (units/L)		ALT (units/L)		AST (units/L)	
	Control (n = 10)	Amyloidotic (n = 8)	Control (n = 8)	Amyloidotic (n = 8)	Control (n = 10)	Amyloidotic (n = 8)	Control (n = 10)	Amyloidotic (n = 8)	Control (n = 10)	Amyloidotic (n = 8)
Mean	2.284	2.649	2.229	2.233	104.3	87.25	36.23	85.7	122.2	136.5
SD	0.09009	0.1323	0.4428	0.4229	39.41	17.81	16.58	68.43	87.22	75.14
P value	<0.0001		0.9864		0.3494		0.0858		0.7178	
Test	unpaired t test		unpaired t test		Mann-Whitney		unpaired t test with Welch's correction for different variances		unpaired t test	

	Total Protein (g/L)		Albumin (g/L)		Total Cholesterol (mmol/L)		HDL Cholesterol (mmol/L)		LDL Cholesterol (mmol/L)	
	Control (n = 10)	Amyloidotic (n = 8)	Control (n = 10)	Amyloidotic (n = 8)	Control (n = 10)	Amyloidotic (n = 8)	Control (n = 10)	Amyloidotic (n = 8)	Control (n = 10)	Amyloidotic (n = 8)
Mean	48.84	60.24	26.8	31.74	2.088	4.874	1.283	2.488	0.388	1.283
SD	4.559	5.654	2.772	3.144	0.7838	0.8647	0.5511	0.4644	0.08879	0.2669
P value	0.0002		0.0027		<0.0001		0.0002		<0.0001	
Test	unpaired t test		unpaired t test		unpaired t test		unpaired t test		unpaired t test with Welch's correction for different variances	

	Glucose (mmol/L)		Triglycerides (mmol/L)		Glycerol (μmol/L)		Free Fatty Acids (mmol/L)		Total Billirubin (μmol/L)	
	Control (n = 10)	Amyloidotic (n = 8)	Control (n = 10)	Amyloidotic (n = 8)	Control (n = 8)	Amyloidotic (n = 8)	Control (n = 10)	Amyloidotic (n = 8)	Control (n = 10)	Amyloidotic (n = 8)
Mean	8.582	8.28	1.064	1.965	703	939.5	0.79	1.626	4.28	4.138
SD	1.527	2.069	0.4387	0.6413	249.7	238.2	0.3746	0.4002	1.515	0.5829
P value	0.7259		0.0027		0.0730		0.0003		0.7893	
Test	unpaired t test		unpaired t test		unpaired t test		unpaired t test		unpaired t test with Welch's correction for different variances	

	LDH (units/L)		Iron (μmol/L)		Amylase (units/L)		CK (units/L)		Uric Acid (μmol/L)	
	Control (n = 10)	Amyloidotic (n = 8)	Control (n = 10)	Amyloidotic (n = 8)	Control (n = 10)	Amyloidotic (n = 8)	Control (n = 9)	Amyloidotic (n = 8)	Control (n = 10)	Amyloidotic (n = 8)
Mean	1429	1172	25.69	24.47	745	823.9	562.2	290	172.2	191
SD	480.6	474.3	7.473	6.889	213	247.5	730.3	86.18	43.49	55.8
P value	0.2732		0.6965		0.2743		0.2994		0.4328	
Test	unpaired t test		Mann-Whitney		Mann-Whitney		unpaired t test with Welch's correction for different variances		unpaired t test	

	Ammonia (μmol/L)		Dox-induced SAA (mg/L)		Duration of dox treatment (days)	
	Control (n = 8)	Amyloidotic (n = 8)	Control (n = 9)	Amyloidotic (n = 8)	Control (n = 10)	Amyloidotic (n = 8)
Mean	170.6	155	7.7	1797	51.0	42.38
SD	36.37	23.3	3.3	1717	18.91	13.09
P value	0.3243		<0.0001		0.2900	
Test	unpaired t test		Mann-Whitney, one-tailed		unpaired t test	

Data that did not pass the D'Agostino & Pearson omnibus normality test were analyzed by the Mann-Whitney U test; other data were analyzed by unpaired t test (with Welch's correction when the variances were different). All two-tailed tests, except where stated.