Acid-cleavable thiomaleamic acid linker for homogeneous antibody-drug conjugation

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General Experimental

All reagents were purchased from Sigma-Aldrich or AlfaAesar and were used as received without further purification. Where described below petrol refers to petrol (b.p. 40-60 °C). All reactions were monitored by thin-layer chromatography (TLC) on pre-coated SIL G/UV254 silica gel plates (254 µm) purchased from VWR. Flash column chromatography was carried out with Kiesegel 60M 0.04/0.063 mm (200-400 mesh) silica gel. ¹H and ¹³C NMR spectra were recorded at ambient temperature on a Bruker Avance 600 instrument operating at a frequency of 600 MHz for ¹H and 150 MHz for ¹³C in a deuterated solvent as described below. The chemical shifts (δ) for ¹H and ¹³C are quoted relative to residual signals of the solvent on the ppm scale. ¹H NMR peaks are reported as singlet (s), doublet (d), triplet (t), quartet (q), quintet (quintet), broad (br) or multiplet (m). Coupling constants (J values) are reported in Hertz (Hz) and are H-H coupling constants unless otherwise stated. Signal multiplicities in ¹³C NMR were determined using the distortionless enhancement by phase transfer (DEPT) spectral editing technique. Infrared spectra were obtained on a Perkin Elmer Spectrum 100 FTIR Spectrometer operating in ATR mode with frequencies given in reciprocal centimetres (cm⁻¹). Melting points were measured with a Gallenkamp apparatus and are uncorrected. Mass spectra were obtained on a VG70-SE mass spectrometer. Optical rotations were measured using a Perkin Elmer 343 polarimeter.

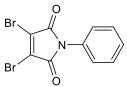
LC-MS was performed on protein samples using a Thermo Scientific uPLC connected to MSQ Plus Single Quad Detector (SQD). Column: Hypersil Gold C4, 1.9 μ m 2.1 x 50 mm. Wavelength: 254 nm. Mobile Phase: 99:1 Water:MeCN (0.1% formic acid) to 1:9 Water: MeCN (0.1% formic acid) gradient over 4 min. Flow Rate: 0.3 mL/min. MS Mode: ES⁺. Scan Range: m/z = 500-2000. Scan time: 1.5 s. Data obtained in continuum mode. The electrospray source of the MS was operated with a capillary voltage of 3.5 kV and a cone voltage of 50 V. Nitrogen was used as the nebulizer and desolvation gas at a total flow of 600 L/h. Ion series were generated by integration of the total ion chromatogram (TIC) over the 3.5-5.0 min range. Total mass spectra for protein samples were reconstructed from the ion series using the pre-installed ProMass software.

3,4-Dibromo-furan-2,5-dione¹



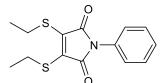
A mixture of maleic anhydride (1.50 g, 15.3 mmol), AlCl₃ (28 mg, 0.21 mmol) and Br₂ (1.57 mL, 4.89 g, 30.6 mmol) was heated at 160 °C in a sealed ampoule for 20 h. After this time, the reaction mixture was allowed to cool to room temperature, and diluted with EtOAc (25 mL). The reaction mixture was then filtered and the solid washed thoroughly with EtOAc (3×25 mL). The filtrate was then concentrated *in vacuo* to afford 3,4-dibromo-furan-2,5-dione as a yellow solid (3.25 g, 12.8 mmol, 83%): m.p. 107-110 °C (*lit. m.p. 113-114* °C)¹; ¹³C NMR (MeOD, 150 MHz) δ 164.4 (C), 125.9 (C); IR (solid) 1769, 1706, 1590 cm⁻¹.

3,4-Dibromo-1-phenyl-pyrrole-2,5-dione 1¹

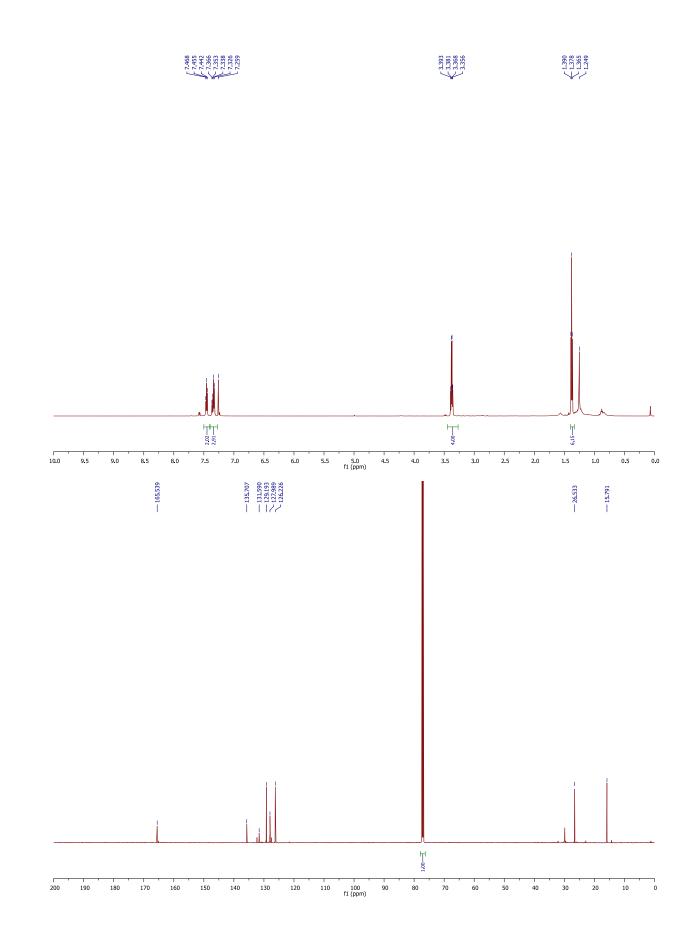


To a solution of 3,4-dibromo-furan-2,5-dione (0.50 g, 1.97 mmol) in AcOH (25 mL) was added aniline (0.18 mL, 1.97 mmol), and the reaction mixture stirred at room temperature for 3 h and then at 130 °C for 90 min. After this time, the solvents were removed *in vacuo*, with the residual traces of AcOH removed by azeotropic distillation using toluene (3×25 mL). The crude residue was purified by flash column chromatography (5% EtOAc/petrol) to afford 3,4-dibromo-1-phenyl-pyrrole-2,5-dione **1** as a pale yellow solid (0.37 g, 1.12 mmol, 57%): m.p. 158-160 °C (*lit. m.p. 162-163* °C)¹; ¹H NMR (CDCl₃, 600 MHz) δ 7.48-7.47 (2H, m, ArH), 7.44-7.40 (1H, m, ArH), 7.33 (2H, d, J = 7.2 Hz, ArH); ¹³C NMR (CDCl₃, 150 MHz) δ 163.0 (C), 132.8 (C), 130.8 (C), 129.5 (CH), 128.8 (CH), 126.2 (CH); IR (solid) 3058, 1727, 1715, 1646, 1610, 1598, 1501 cm⁻¹; LRMS (EI) 333 (50, [M⁸¹Br⁸¹Br]⁺), 331 (100, [M⁸¹Br⁷⁹Br]⁺), 329 (50, [M⁷⁹Br⁷⁹Br]⁺); HRMS (EI) calcd for C₁₀H₅O₂NBr₂ [M⁷⁹Br⁷⁹Br]⁺ 328.8682, observed 328.8685.

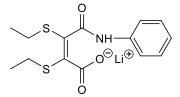
3,4-Bis-ethylsulfanyl-1-phenyl-pyrrole-2,5-dione 2



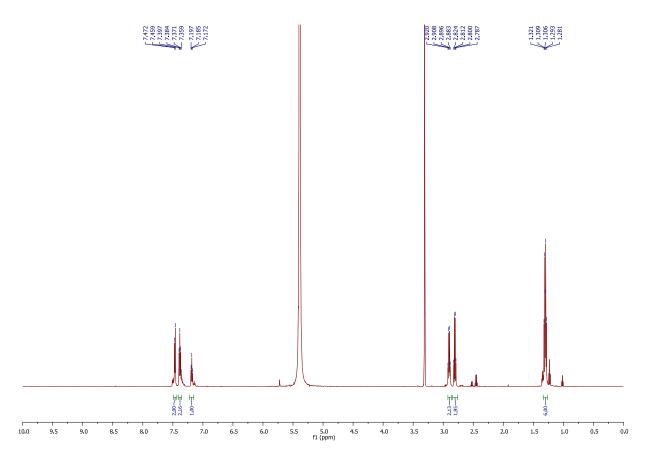
To a solution of 3,4-dibromo-1-phenyl-pyrrole-2,5-dione **1** (237 mg, 0.72 mmol) in CH₂Cl₂ (30 mL) was added ethanethiol (0.11 mL, 1.50 mmol) and NEt₃ (0.21 mL, 1.50 mmol), and the reaction mixture stirred at room temperature for 15 min. After this time, the solvents were removed *in vacuo* and the crude residue purified by flash column chromatography (5% EtOAc/petrol) to afford 3,4-bis-ethylsulfanyl-1-phenyl-pyrrole-2,5-dione **2** as a yellow oil (195 mg, 0.66 mmol, 93%): ¹H NMR (CDCl₃, 600 MHz) δ 7.48-7.44 (2H, m, Ar*H*), 7.37-7.33 (3H, m, Ar*H*), 3.37 (4H, q, *J* = 7.4 Hz, CH₃CH₂S), 1.38 (6H, t, *J* = 7.4 Hz, CH₃CH₂S); ¹³C NMR (CDCl₃, 150 MHz) δ 165.5 (C), 135.7 (C), 131.6 (C), 129.2 (CH), 128.0 (CH), 126.2 (CH), 26.5 (CH₂), 15.8 (CH₃); IR (neat) 2965, 2926, 2851, 1706, 1598, 1501 cm⁻¹; LRMS (EI) 293 (100, [M]⁺); HRMS (EI) calcd for C₁₄H₁₅O₂NS₂ [M]⁺ 293.0539, observed 293.0540.

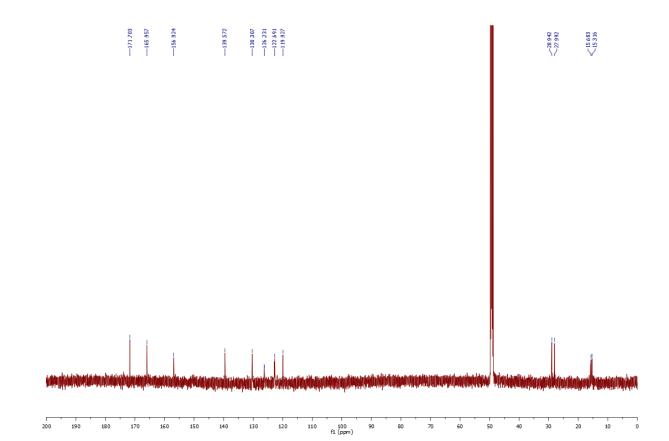


Lithium 2,3-bis-ethylsulfanyl-3-phenylcarbamoyl-acrylate 3



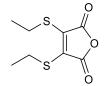
To a solution of 3,4-bis-ethylsulfanyl-1-phenyl-pyrrole-2,5-dione **2** (19.0 mg, 0.065 mmol) in a 1:1 mixture of CD₃OD:D₂O (1 mL:1 mL) was added LiOH.H₂O (146 mg, 3.48 mmol) and the reaction mixture stirred at room temperature for 24 h to afford lithium 2,3-bis-ethylsulfanyl-3-phenylcarbamoyl-acrylate **3**: ¹H NMR (CD₃OD/D₂O, 600 MHz) δ 7.46 (2H, d, *J* = 7.8 Hz, Ar*H*), 7.38 (2H, t. *J* = 7.8 Hz, Ar*H*), 7.18 (1H, t, *J* = 7.8 Hz, Ar*H*), 2.90 (2H, q, *J* = 7.5 Hz, CH₃CH₂S), 2.80 (2H, q, *J* = 7.5 Hz, CH₃CH₂S), 1.31 (3H, t, *J* = 7.5 Hz, CH₃CH₂S), 1.29 (3H, t, *J* = 7.5 Hz, CH₃CH₂S); ¹³C NMR (CD₃OD/D₂O, 150 MHz) δ 171.7 (C), 166.0 (C), 156.9 (C), 139.6 (C), 130.3 (CH), 126.3 (C), 122.7 (CH), 119.9 (CH), 28.9 (CH₂), 28.0 (CH₂), 15.7 (CH₃), 15.3 (CH₃).



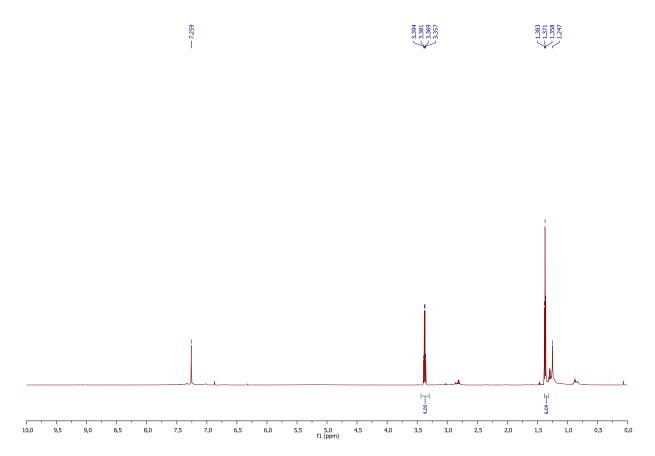


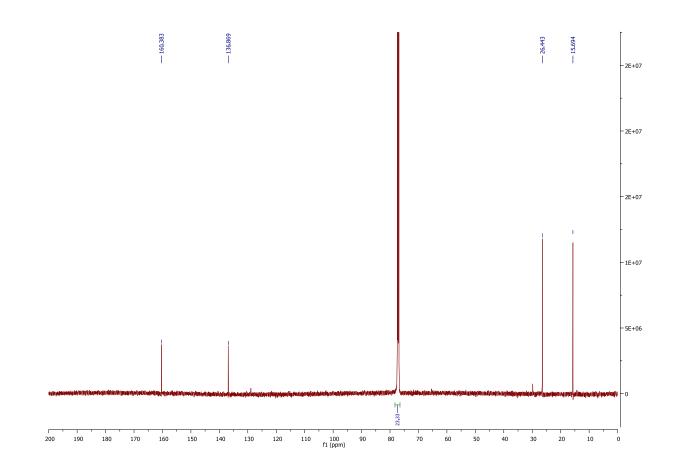
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3,4-Bis-ethylsulfanyl-furan-2,5-dione 4

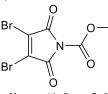


To a solution of lithium 2,3-bis-ethylsulfanyl-3-phenylcarbamoyl-acrylate **3** (0.065 mmol) in a 1:1 mixture of CD₃OD:D₂O (1 mL:1 mL) was added 2M HCl and the reaction mixture acidified to pH 4. After this, 3,4-bis-ethylsulfanyl-furan-2,5-dione **4** was extracted from the reaction mixture into EtOAc (3 × 20 mL), washed with sat. NaCl, dried (MgSO₄), filtered and the solvents removed *in vacuo* to afford 3,4-bis-ethylsulfanyl-furan-2,5-dione **4** as an oil (14.0 mg, 0.065 mmol, >99%% over two steps): ¹H NMR (CDCl₃, 600 MHz) δ 3.37 (4H, q, *J* = 7.4 Hz, CH₃CH₂S), 1.36 (6H, t, *J* = 7.4 Hz, CH₃CH₂S); ¹³C NMR (CDCl₃, 150 MHz) δ 160.4 (C), 136.9 (C), 26.4 (CH₂), 15.7 (CH₃); IR (neat) 2969, 2930, 1761, 1518 cm⁻¹; LRMS (EI) 218 (100, [M]⁺); HRMS (EI) calcd for C₈H₁₀O₃S₂ [M]⁺ 218.0066, observed 218.0071.



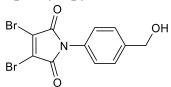


3,4-Dibromo-2,5-dioxo-2,5-dihydro-pyrrole-1-carboxylic acid methyl ester 5²



To a solution of 3,4-dibromo-furan-2,5-dione (1.0 g, 3.9 mmol) and *N*-methylmorpholine (0.43 mL, 3.9 mmol) in THF (35 mL) was added methyl chloroformate (0.30 mL, 3.9 mmol) and the reaction mixture stirred at room temperature for 20 min. After this time, CH₂Cl₂ (40 mL) was added, the organic phase washed with H₂O (3×40 mL), dried (MgSO₄) and the solvent removed *in vacuo* to afford 3,4-dibromo-2,5-dioxo-2,5-dihydro-pyrrole-1-carboxylic acid methyl ester **5** as a pink powder (1.18 g, 3.8 mmol, 97%): m.p. 114-116 °C (*lit. m.p. 115-118* °C)²; ¹H NMR (CDCl₃, 600 MHz) δ 4.00 (3H, s, CH₃); ¹³C NMR (CDCl₃, 150 MHz) δ 159.3 (C), 147.0 (C), 131.5 (C), 54.9 (CH₃); IR (solid) 3236, 2962, 1809, 1769, 1730, 1602 cm⁻¹; LRMS (CI) 314 (50, [M⁸¹Br⁸¹Br +H]⁺), 312 (100, [M⁸¹Br⁷⁹Br]⁺), 310 (50, [M⁷⁹Br⁷⁹Br]⁺); HRMS (EI) calcd for C₆H₃O₄NBr₂ [M⁷⁹Br⁷⁹Br]⁺ 310.8423, observed: 310.8427.

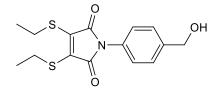
3,4-Dibromo-1-(4-hydroxymethyl-phenyl)-pyrrole-2,5-dione 6²



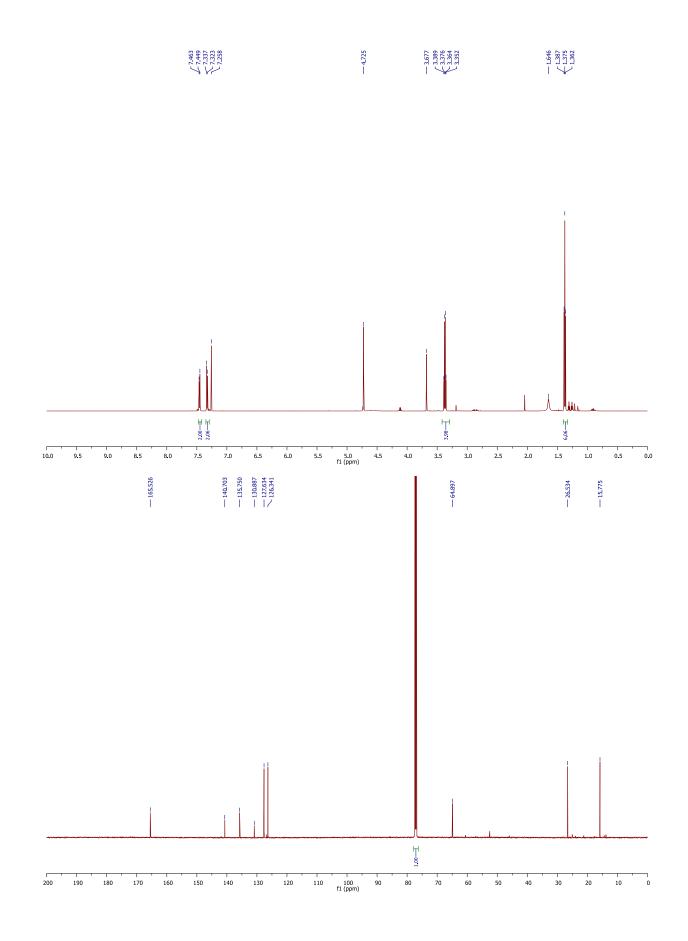
To a solution of 3,4-dibromo-2,5-dioxo-2,5-dihydro-pyrrole-1-carboxylic acid methyl ester 5 (300 mg, 0.965 mmol) in CH_2Cl_2 (42 mL) was added 4-aminobenzyl alcohol (119 mg, 0.965

mmol), and the reaction mixture stirred at room temperature for 2 h. After this time, the solvents were removed *in vacuo* and the crude residue purified by flash column chromatography (50% EtOAc/petrol) to afford 3,4-dibromo-1-(4-hydroxymethyl-phenyl)-pyrrole-2,5-dione **6** as a yellow solid (345 mg, 0.955 mmol, 99%): m.p. 215-217 °C (*lit. m.p. 217-220* °C)²; ¹H NMR (CDCl₃, 600 MHz) δ 7.49 (2H, d, *J* = 8.4 Hz, Ar*H*), 7.33 (2H, d, *J* = 8.4 Hz, Ar*H*), 4.75 (2H, s, CH₂OH); ¹³C NMR (CDCl₃, 150 MHz) δ 163.0 (C), 141.6 (C), 130.2 (C), 130.0 (C), 127.8 (CH), 126.3 (CH), 64.7 (CH₂); IR (solid) 3364, 1732, 1714, 1610, 1519 cm⁻¹; LRMS (EI) 361 (50, [M⁸¹Br⁸¹Br]⁺), 359 (100, [M⁸¹Br⁷⁹Br]⁺), 357 (50, [M⁷⁹Br⁷⁹Br]⁺); HRMS (EI) calcd for C₁₁H₇O₃NBr₂ [M⁷⁹Br⁷⁹Br]⁺ 358.8787, observed: 358.8798.

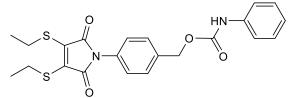
3,4-Bis-ethylsulfanyl-1-(4-hydroxymethyl-phenyl)-pyrrole-2,5-dione



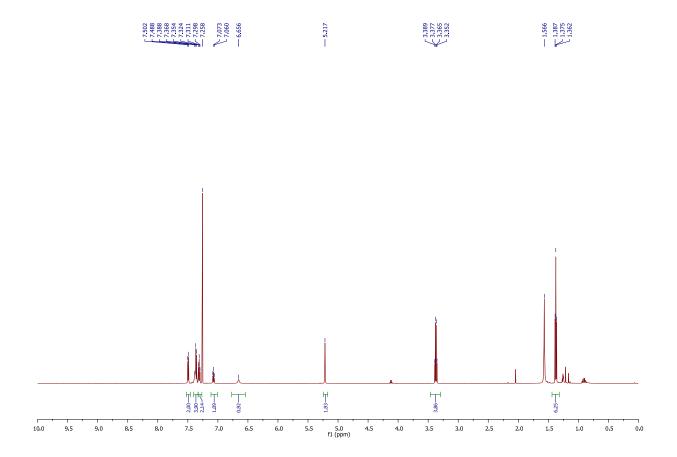
To a solution of 3,4-dibromo-1-(4-hydroxymethyl-phenyl)-pyrrole-2,5-dione **6** (315 mg, 0.872 mmol) in CH₂Cl₂ (40 mL) was added ethanethiol (0.18 mL, 2.33 mmol) and NEt₃ (0.33 mL, 2.33 mmol) and the reaction mixture stirred at room temperature for 1 h. After this time, the reaction mixture was concentrated *in vacuo* and the crude residue purified by flash column chromatography (50% EtOAc/petrol) to afford 3,4-bis-ethylsulfanyl-1-(4-hydroxymethyl-phenyl)-pyrrole-2,5-dione as a yellow solid (242 mg, 0.748 mmol, 86%): m.p. 50-52 °C; ¹H NMR (CDCl₃, 600 MHz) δ 7.45 (2H, d, *J* = 8.4 Hz, Ar*H*), 7.33 (2H, d, *J* = 8.4 Hz, Ar*H*), 4.72 (2H, s, C*H*₂OH), 3.37 (4H, q, *J* = 7.4 Hz, CH₃CH₂S), 1.37 (6H, t, *J* = 7.4 Hz, CH₃CH₂S); ¹³C NMR (CDCl₃, 150 MHz) δ 165.5 (C), 140.7 (C), 135.7 (C), 130.9 (C), 127.6 (CH), 126.3 (CH), 64.9 (CH₂), 26.5 (CH₂), 15.8 (CH₃); IR (solid) 3381, 2967, 2928, 2869, 1702, 1514 cm⁻¹; LRMS (EI) 323 (100, [M]⁺); HRMS (EI) calcd for C₁₅H₁₇O₃NS₂ [M]⁺ 323.0644, observed: 323.0634.

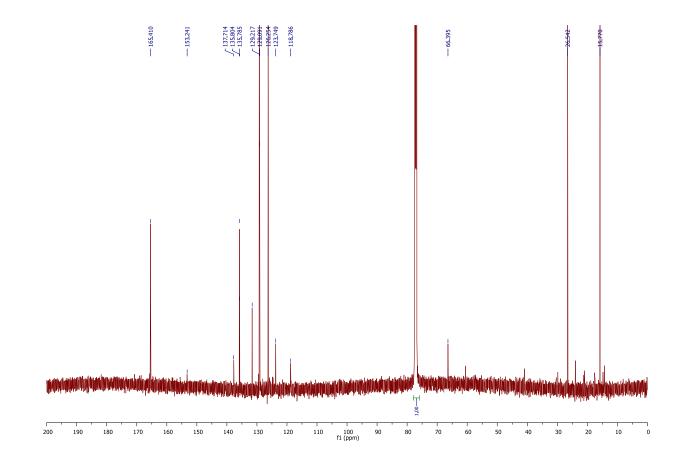


Phenyl-carbamic acid 4-(3,4-bis-ethylsulfanyl-2,5-dioxo-2,5-dihydro-pyrrol-1-yl)-benzyl ester 7

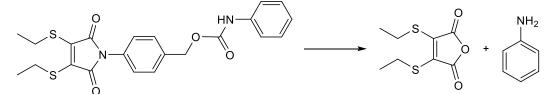


To a solution of 3,4-bis-ethylsulfanyl-1-(4-hydroxymethyl-phenyl)-pyrrole-2,5-dione (100 mg, 0.309 mmol) in CH₂Cl₂ (14 mL) was added phenylisocyanate (0.035 mL, 0.322 mmol) and NEt₃ (0.091 mL, 0.653 mmol), and the reaction mixture stirred at room temperature for 2 days. After this time, the reaction mixture was concentrated *in vacuo* and the crude residue purified by flash column chromatography (20% EtOAc/petrol) to afford phenyl-carbamic acid 4-(3,4-bis-ethylsulfanyl-2,5-dioxo-2,5-dihydro-pyrrol-1-yl)-benzyl ester **7** as a yellow solid (84 mg, 0.190 mmol, 62%): m.p. 97-99 °C; ¹H NMR (CDCl₃, 600 MHz) δ 7.49 (2H, d, *J* = 8.3 Hz, Ar*H*CH₂O), 7.39-7.35 (4H, m, Ar*H*CH₂O and Ar*H*NH), 7.32-7.30 (2H, m, Ar*H*NH), 7.07 (1H, t, *J* = 7.4 Hz, Ar*H*NH), 6.66 (1H, br s, N*H*), 5.22 (2H, s, CH₂OCONH), 3.37 (4H, q, *J* = 7.4 Hz, CH₃CH₂S), 1.37 (6H, t, *J* = 7.4 Hz, CH₃CH₂S); ¹³C NMR (CDCl₃, 150 MHz) δ 165.4 (C), 153.2 (C), 137.7 (C), 135.8 (C), 131.6 (C), 129.2 (CH), 129.1 (CH), 126.3 (CH), 126.2 (CH), 123.7 (CH), 118.8 (C), 66.4 (CH₂), 26.5 (CH₂), 15.8 (CH₃); IR (solid) 3374, 2964, 2928, 1733, 1705, 1598, 1532 cm⁻¹; LRMS (ESI) 465 (100, [M+Na]⁺); HRMS (ESI) calcd for C₂₂H₂₂N₂O₄NaS₂ [M+Na]⁺ 465.0919, observed: 465.0923.



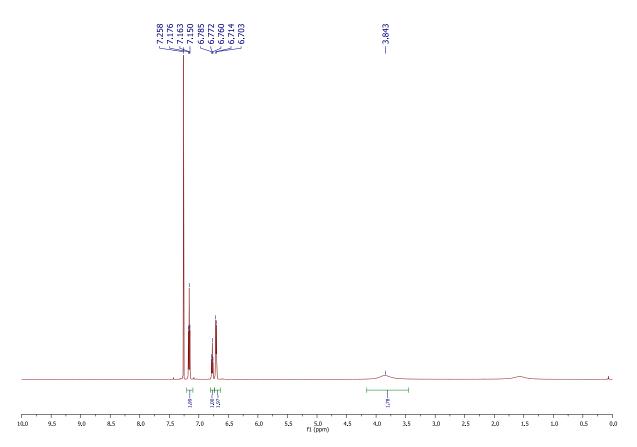


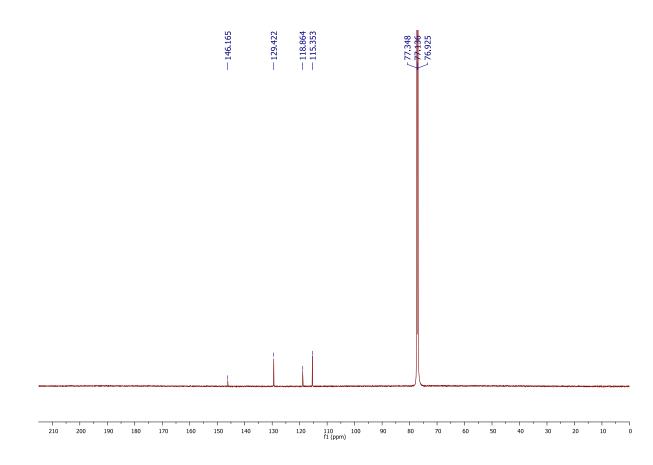
3,4-Bis-ethylsulfanyl-furan-2,5-dione 4 and aniline



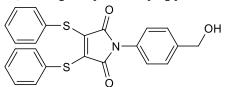
To a solution of phenyl-carbamic acid 4-(3,4-bis-ethylsulfanyl-2,5-dioxo-2,5-dihydro-pyrrol-1yl)-benzyl ester **7** (19.5 mg, 0.044 mmol) in a 1:1 mixture of $CD_3OD:D_2O$ (0.8 mL:0.8 mL) was added LiOH.H₂O (102 mg, 2.44 mmol) and the reaction mixture stirred at room temperature for 2 h. After this time, 2M HCl was added and the reaction mixture acidified to pH 4. Then, 3,4-bis-ethylsulfanyl-furan-2,5-dione **4** was extracted from the reaction mixture into EtOAc (3 × 20 mL), washed with sat. NaCl, dried (MgSO₄), filtered and the solvents removed *in vacuo* to afford 3,4-bis-ethylsulfanyl-furan-2,5-dione **4** as an oil (9.6 mg, 0.044 mmol, >99%). Data matched that described above.

The pH 4, aqueous solution was neutralised by addition of 1M NaOH. Then, the organic materials were extracted into CHCl₃ (3×5 mL), dried (MgSO₄), filtered and the solvent was removed *in vacuo*. The crude residue was purified by flash column chromatography (10% EtOAc/petrol) to afford aniline as an oil (3.9 mg, 0.042 mmol, 95%). ¹H NMR (CDCl₃, 600 MHz) δ 7.16 (2H, t, *J* = 8.0 Hz, Ar*H*), 6.77 (1H, t, *J* = 8.0 Hz, Ar*H*), 6.70 (2H, d, *J* = 8.0 Hz, Ar*H*), 3.84 (2H, br s, NH₂); ¹³C NMR (CDCl₃, 150 MHz) δ 146.2 (C), 129.4 (CH), 118.9 (CH), 115.4 (CH); IR (neat) 3445, 3354, 3210, 3068, 3032, 1619, 1602, 1497 cm⁻¹.

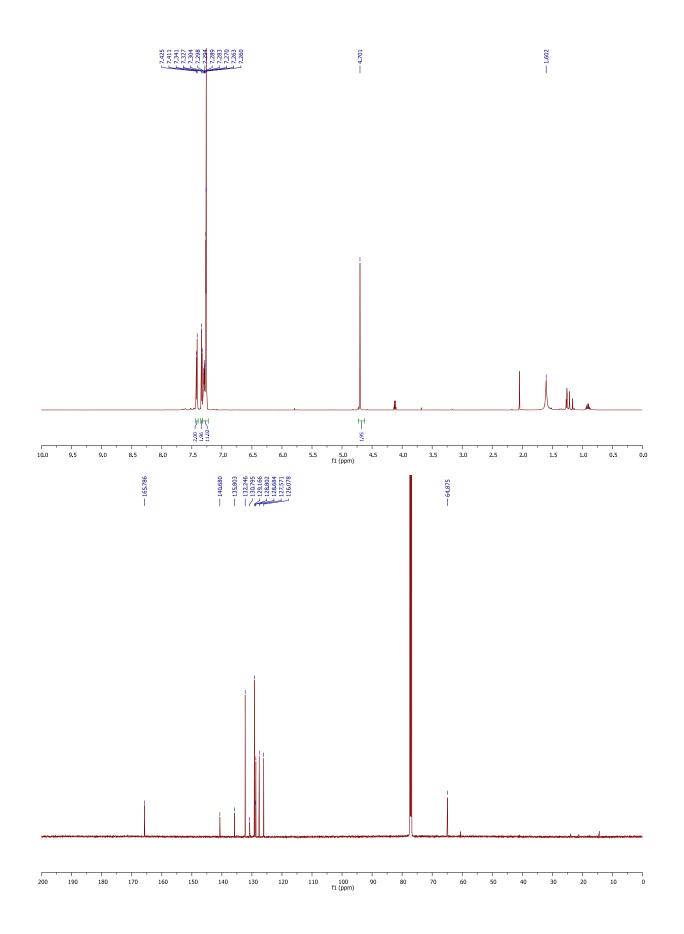




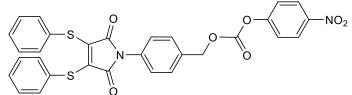
1-(4-Hydroxymethyl-phenyl)-3,4-bis-phenylsulfanyl-pyrrole-2,5-dione 8



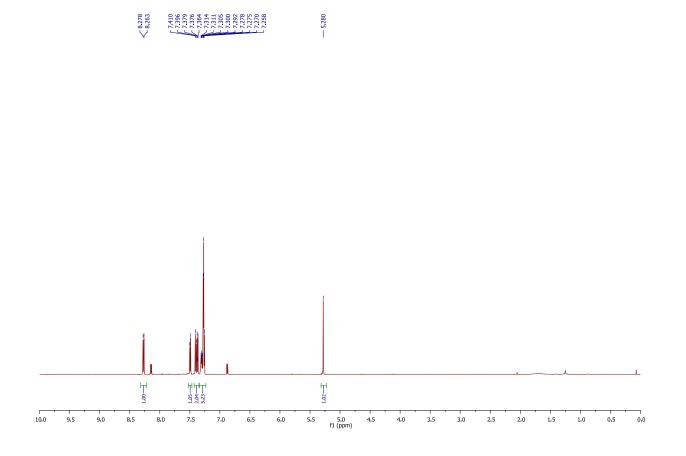
To a solution of 3,4-dibromo-1-(4-hydroxymethyl-phenyl)-pyrrole-2,5-dione **6** (139 mg, 0.385 mmol) in CH₂Cl₂ (18 mL) was added NEt₃ (0.145 mL, 1.04 mmol) and phenylthiol (0.08 mL, 0.809 mmol), and the reaction mixture stirred at room temperature for 30 min. After this time, the reaction mixture was concentrated *in vacuo* and the crude residue purified by flash column chromatography (20% EtOAc/petrol) to afford 1-(4-hydroxymethyl-phenyl)-3,4-bis-phenylsulfanyl-pyrrole-2,5-dione **8** as a yellow solid (159 mg, 0.378 mmol, 98%): m.p. 111-113 °C; ¹H NMR (CDCl₃, 600 MHz) δ 7.46 (2H, d, *J* = 8.4 Hz, Ar*H*CH₂OH), 7.32 (2H, d, *J* = 8.4 Hz, Ar*H*CH₂OH), 7.31-7.26 (10H, m, Ar*H*S), 4.70 (2H, s, CH₂OH); ¹³C NMR (CDCl₃, 150 MHz) δ 165.8 (C), 140.7 (C), 135.8 (C), 132.2 (CH), 130.8 (C), 129.2 (CH), 128.8 (C), 128.7 (CH), 127.6 (CH), 126.1 (CH), 64.9 (CH₂); IR (solid) 3388, 3057, 2928, 2874, 1705, 1515 cm⁻¹; LRMS (EI) 419 (100, [M]⁺); HRMS (EI) calcd for C₂₃H₁₇O₃NS₂ [M]⁺ 419.0650, observed: 419.0653.

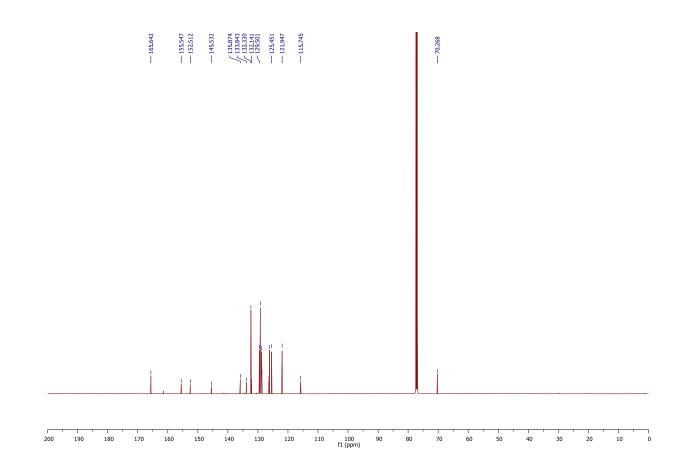


Carbonic acid 4-(2,5-dioxo-3,4-bis-phenylsulfanyl-2,5-dihydro-pyrrol-1-yl)-benzyl ester 4nitro-phenyl ester 9

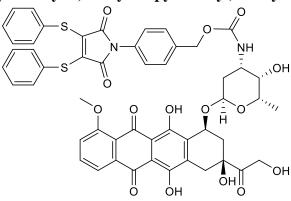


To a solution of 1-(4-hydroxymethyl-phenyl)-3,4-bis-phenylsulfanyl-pyrrole-2,5-dione **8** (30 mg, 0.071 mmol) in CH₂Cl₂ (2 mL) was added 4-nitrophenylchloroformate (17 mg, 0.085 mmol) and pyridine (0.007 mL, 0.085 mmol, and the reaction mixture was stirred at room temperature for 24 h. After this time, CH₂Cl₂ (10 mL) and 10% aqueous citric acid (10 mL) were added, the organic phase washed with H₂O (10 mL) and sat. NaCl (10 mL), dried (MgSO₄), filtered and the solvent removed in *vacuo*. The resultant crude residue was purified by flash column chromatography (20% EtOAc/petrol) to afford carbonic acid 4-(2,5-dioxo-3,4-bis-phenylsulfanyl-2,5-dihydro-pyrrol-1-yl)-benzyl ester 4-nitro-phenyl ester **9** as an oil (30 mg, 0.051 mmol, 72%): ¹H NMR (CDCl₃, 600 MHz) δ 8.27 (2H, d, *J* = 8.3 Hz, Ar*H*NO₂), 7.49 (2H, d, *J* = 8.3 Hz, Ar*H*NO₂), 7.40 (2H, d, *J* = 8.3 Hz, Ar*H*HO₂O₁, 7.37 (2H, d, *J* = 8.3 Hz, Ar*H*NO₂), 7.40 (2H, d, *J* = 8.3 Hz, Ar*H*CH₂O), 7.37 (2H, d, *J* = 8.3 Hz, Ar*H*CH₂O), 7.35-7.26 (10H, m, Ar*H*S), 5.28 (2H, s, CH₂OCO₂); ¹³C NMR (CDCl₃, 150 MHz) δ 165.6 (C), 155.5 (C), 152.5 (C), 147.6 (C), 135.9 (C), 133.8 (C), 132.3 (CH), 132.1 (C), 129.5 (CH), 129.2 (CH), 128.8 (CH), 128.7 (C), 126.2 (CH), 125.5 (CH), 121.9 (CH), 70.3 (CH₂); IR (neat) 3080, 1767, 1714, 1594, 1520 cm⁻¹; LRMS (EI) 584 (100, [M]⁺); HRMS (EI) calcd for C₃₀H₂₀O₇N₂S₂ [M]⁺ 584.0712, observed: 584.0718.

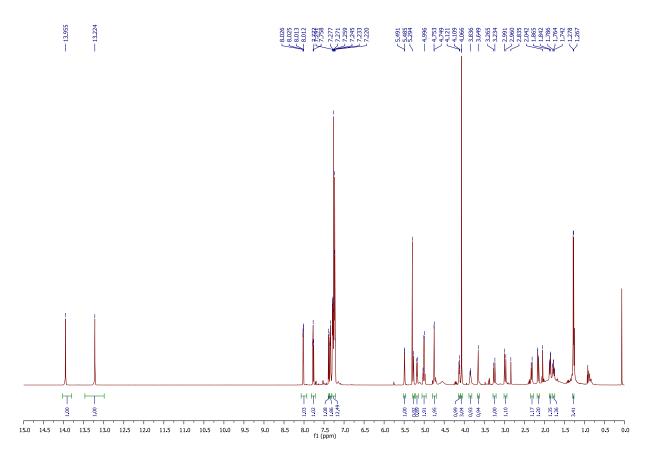


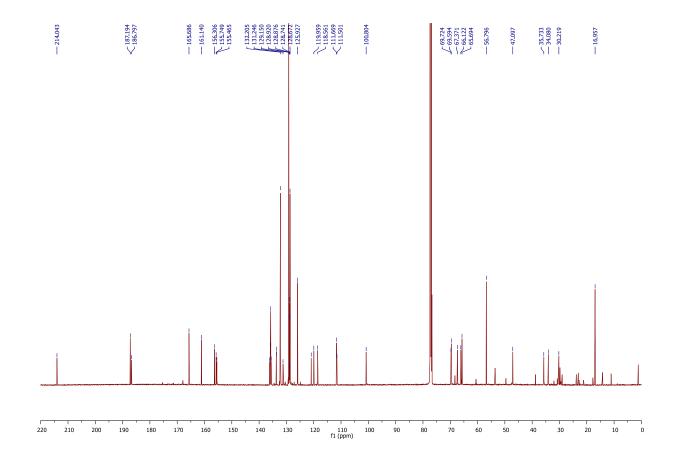


{3-Hydroxy-2-methyl-6-[3,5,12-trihydroxy-3-(2-hydroxy-acetyl)-10-methoxy-6,11-dioxo-1,2,3,4,6,11-hexahydro-naphthacen-1-yloxy]-tetrahydro-pyran-4-yl}-carbamic acid 4-(2,5-dioxo-3,4-bis-phenylsulfanyl-2,5-dihydro-pyrrol-1-yl)-benzyl ester 10



To a solution of carbonic acid 4-(2,5-dioxo-3,4-bis-phenylsulfanyl-2,5-dihydro-pyrrol-1-yl)benzyl ester 4-nitro-phenyl ester **9** (9.8 mg, 0.017 mmol) and DOX·HCl (10 mg, 0.018 mmol) in NMP (0.30 mL) was added NEt₃ (2.5 μ L, 0.018 mmol), and the reaction mixture stirred at room temperature for 3 days. After this time, the reaction mixture was diluted with 10% *i*-propanol/EtOAc, washed with H₂O (10 mL) and sat. NaCl (10 mL), dried (MgSO₄), filtered and the solvent removed in *vacuo*. The crude residue was purified by flash column chromatography (2% MeOH/CH₂Cl₂) to afford {3-Hydrohxy-2-methyl-6-[3,5,12-trihydroxy-3-(2-hydroxy-acetyl)-10-methoxy-6,11-dioxo-1,2,3,4,6,11-hexahydro-naphthacen-1-yloxy]tetrahydro-pyran-4-yl}-carbamic acid 4-(2,5-dioxo-3,4-bis-phenylsulfanyl-2,5-dihydro-pyrrol-1-yl)-benzyl ester **10** as an orange oil (16.4 mg, 0.0166 mmol, 99%): ¹H NMR (CDCl₃, 600 MHz) δ 13.96 (1H, s, DOX), 13.22 (1H, s, DOX), 8.02 (1H, d, *J* = 8.2 Hz, DOX), 7.77 (1H, d, *J* = 8.2 Hz), 7.38 (1H, d, *J* = 8.2 Hz, DOX), 7.34 (2H, d, *J* = 8.2 Hz, Ar*H*CH₂), 7.31-7.22 (12H, m, Ar*H*CH₂ (2H) and Ar*H*S (10H)), 5.49 (1H, br s, DOX), 5.27 (1H, br s, DOX), 5.03-4.98 (2H, m, DOX), 4.75 (2H, s, C*H*₂O), 4.14-4.10 (1H, m, DOX), 4.07 (3H, s, DOX), 3.86-3.82 (1H, m, DOX), 3.65 (1H, s, DOX), 3.25 (1H, d, *J* = 18.7 Hz, DOX), 2.98 (1H, d, *J* = 18.7 Hz), 2.32 (1H, d, *J* = 14.6 Hz, DOX), 2.16 (1H, dd, *J* = 14.6 and 4.0 Hz, DOX), 1.86 (1H, dd, *J* = 13.4 and 4.9 Hz, DOX), 1.79-1.76 (1H, m, DOX), 1.27 (3H, d, *J* = 6.6 Hz, DOX); ¹³C NMR (CDCl₃, 150 MHz) δ 214.0 (C), 187.2 (C), 186.8 (C), 165.7 (C), 161.1 (C), 156.3 (C), 155.7 (C), 155.5 (C), 136.1 (C), 135.9 (C), 135.7 (CH), 135.6 (C), 133.7 (C), 132.2 (CH), 131.2 (C), 129.2 (CH), 129.2 (CH), 128.9 (CH), 128.9 (C), 128.7 (CH), 125.9 (CH), 120.9 (C), 120.0 (CH), 118.6 (CH), 111.7 (C), 111.5 (C), 100.8 (CH), 76.7 (C), 69.7 (CH), 69.6 (CH), 67.4 (CH), 66.1 (CH₂), 65.7 (CH₂), 56.8 (CH₃), 47.1 (CH), 35.7 (CH₂), 34.1 (CH₂), 30.2 (CH₂), 17.0 (CH₃); IR (neat) 3481, 3080, 1716, 1580, 1517 cm⁻¹; LRMS (ESI) 1011 (100, [M+Na]⁺); HRMS (ESI) calcd for C₅₁H₄₄N₂O₁₅NaS₂ [M+Na]⁺ 1011.2081, observed: 1011.2122; [α]_D²⁰ = +91.3 (*c* 1.1, CHCl₃).





Trastuzumab Fab Fragment Preparation

Attempted preparation of a Trastuzumab Fab Fragment using the protocol outlined by K. L. Bennett *et al.*³

In accordance with the report by K. L. Bennett *et al.*,³ trastuzumab (6.41 mg/ml) in digestion buffer was reacted with a 1/10 amount (wt./wt.) of immobilized papain (250 μ g/mL of gel) for 20 h at 37 °C under nitrogen in a buffer containing 20 mM sodium phosphate monobasic, 10 mM disodium EDTA and 80 mM cysteine.HCl (pH 7.0). The cysteine.HCl was incorporated immediately before trastuzumab digestion. After digestion, the mixture was centrifuged at 200 rcf for 5 min and the supernatant was removed for purification. The supernatant was concentrated to a volume of 200 μ L using a diafiltration column (30 KDa MWCO) to purify it from low-molecular-weight proteolytic contaminants, and buffer exchanged into phosphatebuffered saline (PBS, pH 7.0) by passage through diafiltration columns (30 KDa MWCO) four times with excessive PBS (pH 7.0). Finally, the sample was analysed by LCMS and revealed a mixture of Fab products, LCMS observed masses: 47306 and 47673.

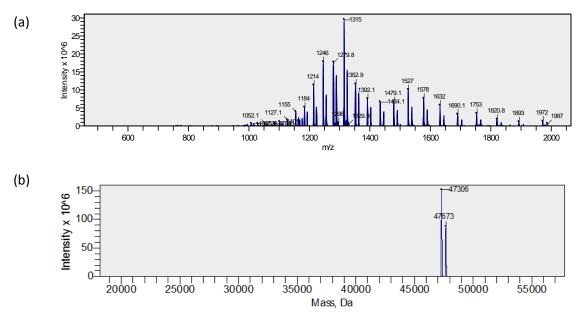


Figure S1. (a) non-deconvoluted and (b) deconvoluted MS data for attempted preparation of a trastuzumab Fab fragment using the protocol outlined by K. L. Bennett *et al.*³

Preparation of trastuzumab Fab fragment (11) using sequential digests with pepsin and papain

Immobilized pepsin (0.15 mL) was washed with digestion buffer (20 mM sodium acetate trihydrate, pH 3.1) four times and trastuzumab (0.5 mL, 6.41 mg/mL in digestion buffer) was added. The mixture was incubated for 5 h at 37 °C whilst shaking (1100 rpm). The resin was separated from the digest using a filter column, and washed with digest buffer (50 mM phosphate, 1 mM EDTA, 150 mM NaCl, pH 6.8) three times. The digest was combined with the washes and the volume adjusted to 0.5 mL. The sample was analysed by LCMS and revealed formation of trastuzumab-F(ab²)₂, LCMS observed mass: 97303.

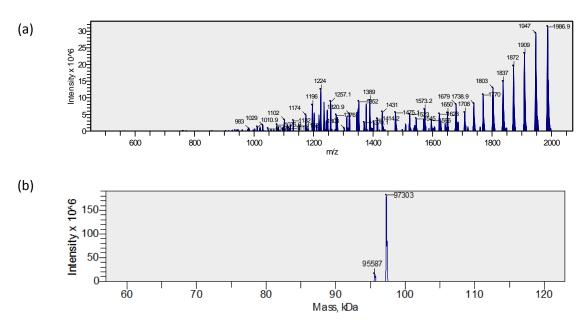
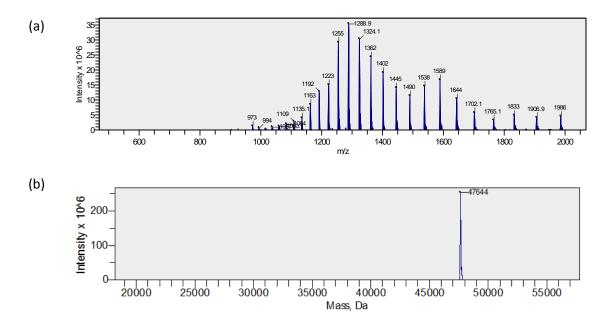


Figure S2. (a) non-deconvoluted and (b) deconvoluted MS data for digestion of trastuzumab with pepsin to afford trastuzumab- $F(ab')_2$.

After this, papain (1 mL, 0.25 mg/mL) was activated with 10 mM DTT (in digest buffer: 50 mM phosphate, 1 mM EDTA, 150 mM NaCl, pH 6.8) under an argon atmosphere whilst shaking (1100 rpm) for 1 h at 25 °C in the dark. The resin was washed with digest buffer (without DTT) four times and the 0.5 mL of Herceptin-F(ab')₂ added. The mixture was incubated for 16 h at 37 °C whilst shaking (1100 rpm) in the dark. Then the resin was separated from the digest using a filter column, and washed with PBS (pH 7.0) three times. The digest was combined with the washes and the buffer was exchanged completely for PBS (pH 7.4) using diafiltration columns (10 KDa MWCO) and the volume adjusted to 0.4 mL. The digest was analysed by SDS-PAGE and LCMS to reveal formation of a single trastuzumab Fab fragment, **11**: observed mass 47644. The concentration of trastuzumab Fab fragment **11** was determined by UV/VIS using a molecular extinction coefficient of $\varepsilon_{280} = 68590 \text{ M}^{-1} \text{ cm}^{-1}$. [trastuzumab Fab fragment **11**] 3.33 mg/mL (0.4 mL), 64%.



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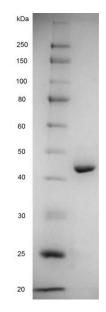


Figure S3. (a) non-deconvoluted, (b) deconvoluted MS data, and (c) SDS-PAGE characterisation of trastuzumab Fab fragment 11 prepared by using sequential digests with pepsin and papain.



Fab ADC Experimental Procedures and Data

Reduced trastuzumab Fab



To a solution of Fab fragment **11** (50 μ L, 1.72 mg/mL, 25 mM sodium borate, 25 mM NaCl, 1 mM EDTA, pH 8.0) was added TCEP (15 μ L, 0.36 mM) to affect reduction of the interchain disulfide. After 1.5 h at 37 °C, the reaction mixture was analysed by LCMS to reveal the heavy and light chains only (*i.e.* reduced trastuzumab Fab fragment).

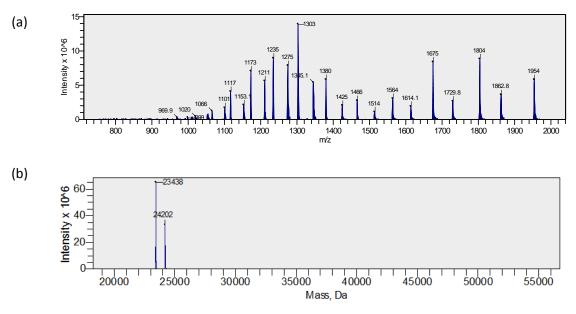
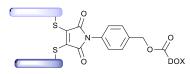


Figure S4. (a) non-deconvoluted and (b) deconvoluted MS data for reduction trastuzumab Fab fragment 11 with TCEP.

Trastuzumab Fab-DOX ADC 12



To the solution of reduced trastuzumab Fab fragment **11** was added MeCN (13 μ L, 20% v/v), DMF (6.5 μ L, 10% v/v) and {3-Hydroxy-2-methyl-6-[3,5,12-trihydroxy-3-(2-hydroxy-acetyl)-10-methoxy-6,11-dioxo-1,2,3,4,6,11-hexahydro-naphthacen-1-yloxy]-tetrahydro-pyran-4-yl}-carbamic acid 4-(2,5-dioxo-3,4-bis-phenylsulfanyl-2,5-dihydro-pyrrol-1-yl)-benzyl ester **10** (1.15 μ L, 7.91 mM in DMF), and the reaction maintained at 37 °C for 1 h. After this time, the reaction mixture was analysed by LCMS to reveal formation of desired trastuzumab Fab-DOX ADC **12** (expected mass: 48414 Da, observed mass: 48413 Da).

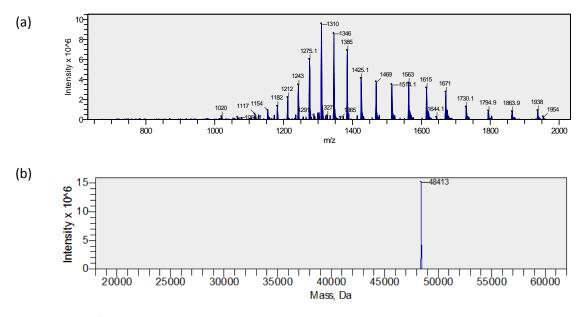
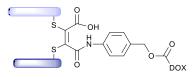


Figure S5. (a) non-deconvoluted and (b) deconvoluted MS data for trastuzumab Fab-DOX ADC 12.

Trastuzumab Fab-DOX ADC 13



The solution of trastuzumab Fab-DOX ADC **12** was prepared as described above, and then buffer exchanged into PBS (pH 7.4, final volume: 50 μ L), and the solution incubated for 20 h at 37 °C. After this time, the reaction mixture was analysed by LCMS to reveal formation of partial maleimide hydrolysis product trastuzumab Fab-DOX ADC **13** (expected mass: 48432 Da, observed mass: 48433 Da). The only degradation product (expected mass: 48036 Da, observed mass: 48031 Da) observed corresponded to that resulting from the known hydrolysis of the sugar component of DOX upon prolonged incubation.⁴

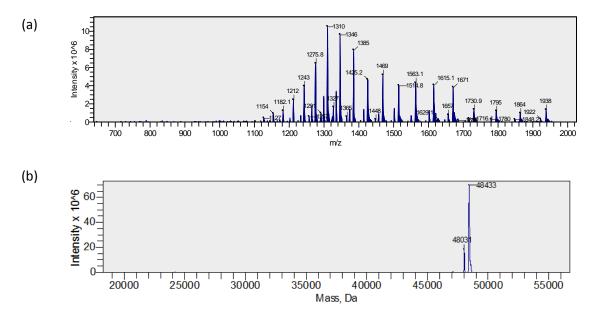


Figure S6. (a) non-deconvoluted and (b) deconvoluted MS data for trastuzumab Fab-DOX ADC 13.

Stability of trastuzumab Fab-DOX ADC (13) at physiological pH and temperature

Trastuzumab Fab-DOX ADC **13** was incubated at physiological temperature and pH (PBS, pH 7.4, final volume: 50 μ L) for 3 days. During this time, no degradation of the linker was observed by LCMS.

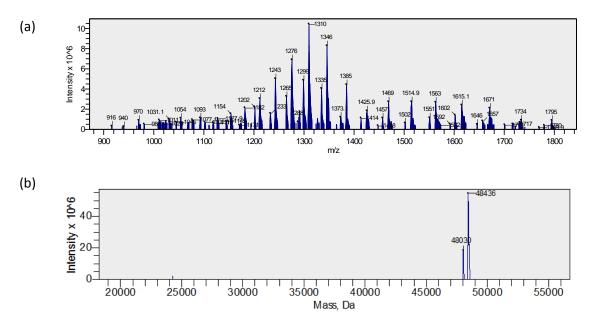
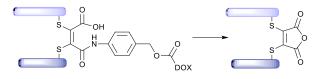
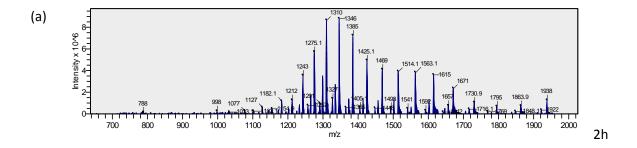


Figure S7. (a) non-deconvoluted and (b) deconvoluted MS data for trastuzumab Fab-DOX ADC 13 at physiological pH and temperature after 3 days.

Release of DOX from ADC 13 at lysosomal pH to yield Conjugate 14



The solution of trastuzumab Fab-DOX ADC **13** was prepared as described above and then buffer exchanged into a low pH buffer (50 mM citric acid, 150 mM sodium chloride, pH 4.5) by ultrafiltration. The solution was incubated at 37 °C. Aliquots of the reaction mixture were analysed by LCMS after 2, 6, 24, 48 and 72 h. Near quantitative release of the cargo was observed (expected mass: 47740 Da, observed mass: 47740 Da).



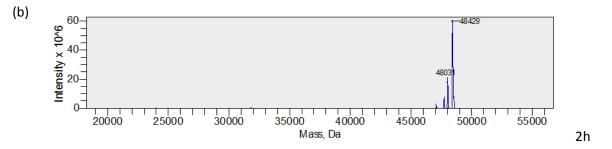


Figure S8. (a) non-deconvoluted and (b) deconvoluted MS data for trastuzumab Fab-DOX ADC **13** at pH 4.5 and 37 °C after 2 h.

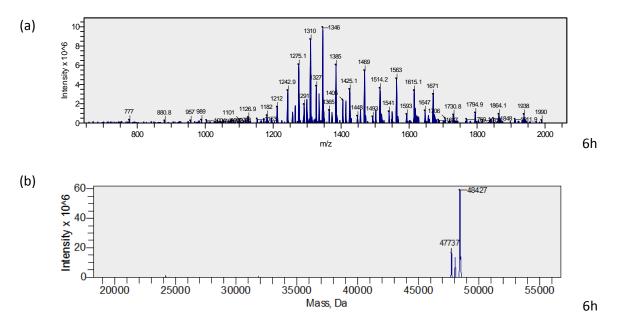


Figure S9. (a) non-deconvoluted and (b) deconvoluted MS data for trastuzumab Fab-DOX ADC **13** at pH 4.5 and 37 °C after 6 h.

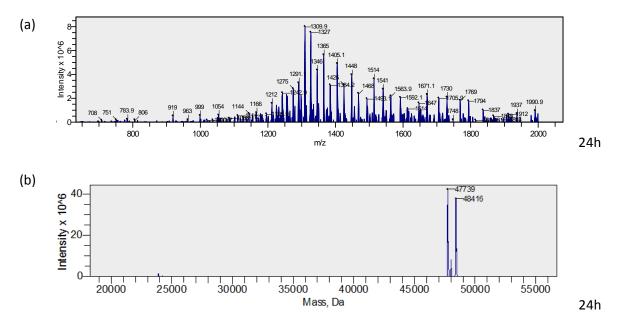


Figure S10. (a) non-deconvoluted and (b) deconvoluted MS data for trastuzumab Fab-DOX ADC 13 at pH 4.5 and 37 °C after 24 h.

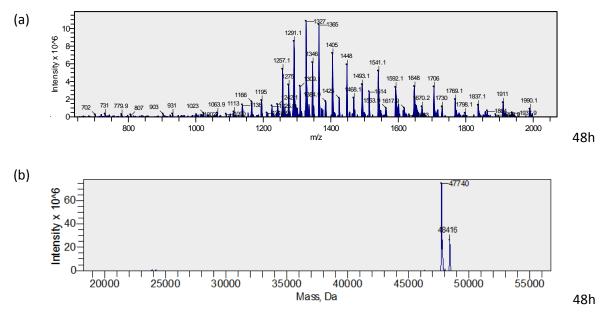


Figure S11. (a) non-deconvoluted and (b) deconvoluted MS data for trastuzumab Fab-DOX ADC 13 at pH 4.5 and 37 °C after 48 h.

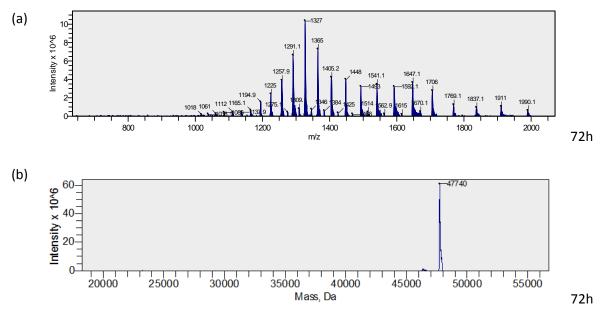


Figure S12. (a) non-deconvoluted and (b) deconvoluted MS data for trastuzumab Fab-DOX ADC 13 at pH 4.5 and 37 °C after 72 h.

Preparation of processed trastuzumab Fab

A solution of Fab fragment **11** (50 μ L, 1.72 mg/mL, 25 mM sodium borate, 25 mM NaCl, 1 mM EDTA, pH 8.0) was incubated for 1.5 h at 37 °C. After this time, to the solution were added MeCN (13 μ L, 20% v/v) and DMF (6.5 μ L, 10% v/v) and the reaction maintained at 37 °C for 1 h. Next, the reaction mixture was buffer exchanged into PBS (pH 7.4, final volume: 50 μ L), and the solution incubated for 20 h at 37 °C. After this time, the reaction mixture was analysed by LCMS to reveal processed Fab (expected mass: 47644 Da, observed mass: 47650 Da).

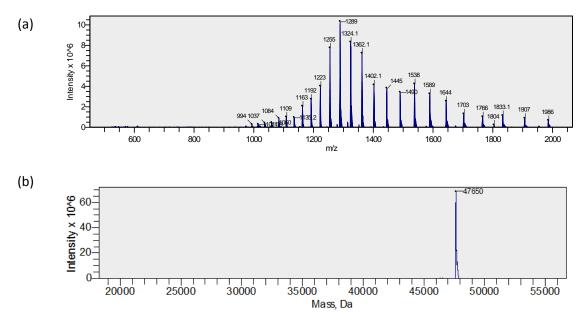


Figure S13. (a) non-deconvoluted and (b) deconvoluted MS data for processed trastuzumab Fab.

Protocol for ELISA

96-Well plates were coated overnight at 4 °C with HER2 (0.25 μ g/mL); PBS was used as negative control. HER2 was removed; the wells washed with PBS and blocked with 200 μ L of 1% BSA solution for 2 h at room temperature. After washing with PBS, the serially diluted test samples (30 nM, 10 nM, 3.33 nM, 1.11 nM, 0.37 nM, 0.12 nM) were added and incubated for 1 h at room temperature. The wells were washed twice with PBS-T (PBS, 0.1% Tween-20) and once with PBS, and anti-human IgG, Fab-specific-HRP antibody (Sigma-Aldrich) was incubated for 1 h at room temperature to detect the bound test sample. The plates were washed again and 100 μ L of 0.5 mg/mL *o*-phenylenediamine hydrochloride (Sigma-Aldrich) in a phosphate-citrate buffer with sodium perborate were added as substrate. Once colour was visible, the reaction was stopped by acidifying with 50 μ L of 4M HCl. Absorbance was immediately measured at 490 nm. ELISA assays were conducted for trastuzumab Fab **11**, processed Fab and Fab ADC **13**.

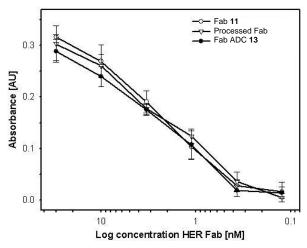


Figure S14. 1 ELISA analysis of Fab 11, processed Fab and Fab ADC 13 binding to the HER2 antigen.

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