

## Engineering Angiogenesis by Hypoxia-Induced Signaling: Adopting a Physiological Approach

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**INTRODUCTION:** Successful engineering of tissues with clinically relevant size and complexity critically depends on their *in vitro* pre-vascularization which can promote cell survival, differentiation and rapid vascularization post-implantation. However, mimicking *in vitro* the physiological complexity of a vascular network currently presents major obstacles<sup>1</sup>. In this study we tested the hypothesis that a hypoxia-induced signaling (HIS) - cell population can generate the complete angiogenic cascade necessary for inducing endothelial cell sprouting and tubule formation within a 3D construct.

**METHODS:** HUVECs and human-dermal-fibroblasts (HDFs) were co-cultured in 3D spiral or flat collagen constructs<sup>2</sup> for 1 or 2 weeks, with no direct contact between the two cell types. HDFs were seeded at high density ( $23 \times 10^6$  cells/ml) or low density ( $1 \times 10^6$  cells/ml) in spiral and flat constructs, respectively. HUVEC-only constructs served as controls. Constructs were cultured in the presence or absence of anti-VEGF neutralizing antibody. O<sub>2</sub> tension within constructs was monitored using an optical fibre-based system<sup>3</sup>. ELISA was used to quantify HIF-1 $\alpha$  and VEGF in 5 and 10 day cultures.

**RESULTS:** Cell O<sub>2</sub> consumption in high-HDF-density co-cultures resulted in hypoxic O<sub>2</sub> levels (<3%) in the HDF region's core, within 24hrs. In high-HDF-density co-cultures HUVECs formed CD31 and von-Willebrand factor positive capillary-like structures (CLS) with lumens and invaded the HDF region at 1 week. There was a significant increase in the number of sprouts from 1 to 2 weeks which correlated with a reduction in the number of endothelial-cell clusters. No CLS formation was observed in HUVEC-only cultures or in low-HDF-density co-cultures (no hypoxic stimulus). HIF-1 $\alpha$  was present in high-HDF-density co-cultures at 5 and 10 days, while VEGF levels increased by 7 fold from 5 to 10 days. No HIF-1 $\alpha$  or VEGF were detected in HUVEC-only cultures.

Anti-VEGF neutralizing antibody reduced sprout length by 50% in high-HDF-density co-cultures.

### DISCUSSION & CONCLUSIONS:

While it is widely accepted that long-term exposure of cells to hypoxia can be detrimental to cell viability, the results of this study indicate that hypoxia can be employed as a physiological signal for inducing an angiogenic response within a 3D tissue construct. Here we show that the angiogenic response was accompanied by up-regulation of two critical, hypoxia-inducible angiogenic factors, HIF-1 $\alpha$  and VEGF. However, the use of hypoxia as the primary angiogenic signal would be expected to trigger the complete angiogenic cascade required for a physiological angiogenic response. The ability to spatially localize the hypoxic signal within a 3D tissue construct could be an invaluable tool for engineering angiogenesis *in vitro* or for pre-conditioning constructs prior to implantation.

We propose that a HIS - cell population could rapidly and physiologically induce an angiogenic response within a 3D tissue construct.

**REFERENCES:**<sup>1</sup>N.C.Rivron, J.Liu, J. Rouwkema, J. de Boer, C.A. van Blitterswijk (2008) Engineering vascularised tissues in vitro. *European cells and Materials* **15**, 27-40. <sup>2</sup>R.A.Brown, M.Wiseman, C.B.Chuo, U.Cheema, S.N.Nazhat (2005) Ultrarapid engineering of biomimetic materials and tissues: Fabrication of nano- and microstructures by plastic compression. *Advanced Functional Materials* **15**, 1762-1770. <sup>3</sup>U.Cheema, R.A.Brown, B.Alp, A.J.MacRobert (2008) Spatially defined oxygen gradients and vascular endothelial growth factor expression in an engineered 3D cell model. *Cell Mol.Life Sci.* **65**, 177-186.

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