

Genomic Guidance for Future Mental Illness Treatment

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A thesis submitted for the degree of

Doctor of Philosophy

Molecular Psychiatry

University College London

Supervised by:

Professor Andrew McQuillin

Professor Nick Bass

Declaration

I, Kai Yao, confirm that the work presented in my thesis is my own. Where information has been derived from other sources, I confirm that this has been indicated in the thesis.

Abstract

Chapter 1 of this thesis introduces how various genomic approaches can help identify potential biomarkers to enable future personalized treatments for mental illnesses. In Chapter 2, I examined how psychiatric polygenic risk scores (PRS), as an indication of genetic liabilities to different psychiatric disorders, interact with environmental factors, adverse childhood experiences (ACE) for different bipolar disorder (BD) phenotype presentations. Although no significant interaction effects were found, higher ADHD PRS correlated with increased number of ACEs and increased the likelihood of rapid cycling. Elevated BD PRS was associated with the presence of psychotic symptoms. Lithium is the gold standard for treating BD however, only about 30% of patients are excellent responders while others could experience side effects. Chapter 3 employed genome-wide association studies (GWAS) to identify good lithium response biomarkers. SNP rs116927879 (A/G) was found to be genome-wide significantly associated with good lithium response, with splicing QTL analysis suggesting that it affects *ADCY1* splicing across brain regions. Antipsychotics are widely prescribed for schizophrenia, but about 40% of patients develop extrapyramidal side effects (EPSE) such as dyskinesias, Parkinsonism, akathisia, and dystonia. Chapter 4 integrated epigenome-wide association studies (EWAS) on GWAS to identify EPSE biomarkers resulted from long-term antipsychotic use. Notably, cg12044923, which mapped to the *STK32B* gene, showed enrichment for EPSE and has been associated with movement disorders such as tremors. In Chapter 5, I explored how interventions could impact PRS predictions of cardiovascular risks in patients with severe mental illness. The findings highlighted that treatments can attenuate cardiovascular PRS predictions and warrant careful PRS

assessment timing. The results also underscored the complex interplay between BD genetic risks and cardiovascular treatment response. This thesis concludes by reflecting on the employed methods' limitations, the potential challenges with pharmacogenetic testing implementation, and how future research could address these to advance the field.

Impact Statement

While pharmacogenomic testing (PGx) is increasingly recognized as important for its potential to personalize mental illness treatments. Comprehensive genomic profiles associated with different treatments must be systematically documented and analysed to realize their potential. This thesis applies various genomic analysis methods to identify potential genetic biomarkers that can differentiate psychiatric treatment responses.

This thesis starts in Chapter 1 where I discussed how changes in views regarding mental illness have shaped the associated treatments today. Modern mental health care can benefit from PGx to provide more precise, biologically informed, and personalized treatment guidance. Thus, the field must continue refining its understanding of genetic influences on psychiatric disorders and associated treatment responses.

In Chapter 2, I employed multiple psychiatric polygenic risk scores to investigate their interactions with environmental factors like adverse childhood experiences (ACE) on the development of different BD phenotypes. The analyses used robust PRS calculation methods and the latest reference GWAS. We also made clear separation on ACE definitions. The findings provide evidence for the utility of psychiatric PRS in phenotype predictions, which can enable early risk identification.

In Chapter 3, I conducted genome-wide association study (GWAS) meta-analyses to identify lithium response genetic biomarkers. This GWAS is currently the largest to date in terms of size for lithium. We used genomic data to predict the heritability of good lithium response for the first time. The study identified *ADCY1* as a gene that may

influence the brain regions for producing different lithium response. These results pinpoint the importance of examining *ADCY1* for patients with BD in future lithium response research. Future research into its role could help optimize treatment decisions and improve outcomes for individuals with BD.

In Chapter 4, I integrated epigenome-wide association studies (EWAS) on GWAS results to identify biomarkers that can differentiate extrapyramidal side-effects (EPSE) from antipsychotic treatments. We had the largest sample size for EPSE to date in both analyses, revealing that the *STK32B* gene showed significant enrichment for EPSE. Notably, *STK32B* has been previously linked to movement disorders, such as tremors. These findings underscore the importance of examining *STK32B* in patients with schizophrenia in future antipsychotic treatment research. This may help predict and mitigate the risk of EPSE, improving treatment safety and efficacy.

Cardiovascular PRS is increasingly used in risk prediction models and has been shown to add information. However, there were very few papers examining how PRS predictions can change after interventions. In Chapter 5, I explored PRS-based predictions of 12-month cardiovascular treatment responses in patients with severe mental illness. To our knowledge, this analysis represents the first of its kind in psychiatry. The findings highlighted the instability of cardiovascular PRS predictions and warrant careful PRS assessment timing in clinical settings.

This thesis concludes in Chapter 6 where I discussed the limitations of the methods used in relation to the results from each chapter and how future research can improve on these.

Inclusion of Published Works

This paper in chapter 2 has been expanded from my previous MSc project and lead to a publication.

- Yao, K., van der Veen, T., Thygesen, J., Bass, N., & McQuillin, A. (2023). Multiple psychiatric polygenic risk scores predict associations between childhood adversity and bipolar disorder. *Journal of Affective Disorders*, 341, 137–146.
<https://doi.org/10.1016/j.jad.2023.08.116>

This paper in chapter 3 is currently on preprint shown at Research Square and is under review.

- McQuillin, A., Yao, K., Nadeem, A., Van Der Veen, T., Thygesen, J., Jonsson, L., ... & Bass, N. (2024). Implication of the ADCY1 Gene in Lithium Response in Bipolar Disorder by Genome-wide Association Meta-analysis.

This paper in chapter 4 is currently on preprint shown at MedRxiv and is under review.

- Yao, K., Thygesen, J. H., Lock, S. K., Pardinas, A. F., Pritchard, A. L., O'Donovan, M. C., Owen, M. J., Walters, J. T. R., Clair, D. S., Bass, N., & McQuillin, A. (2025). Integrating Genome-wide and Epigenome-wide Associations for Antipsychotic Induced Extrapyramidal Side Effects (p. 2025.02.27.25323006). medRxiv.
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Author Contribution Statement

I led all the writing and main analyses presented in this thesis from chapter 1 to 6 under guidance of my supervisors. Other authors' contributions to each research paper presented in chapters 2 to 5 are listed in each research paper declaration form.

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All authors participated in the design of this study. KY led the writing of the manuscript. TV and AM supported the statistical analyses. All authors contributed to revise drafts.

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KY led main analyses and manuscript writing. AN, TV, JHT, LJ, JS, and ML supported data processing. NB reviewed lithium coding. AM supported study design. All authors contributed to revise drafts.

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KY led the meta-analyses and manuscript writing. JHT, IE, AL, and ZA supported UCL analyses. SKL, AFP, MCO, MJO, and JTRW supported Cardiff analyses. ALP and DSC supported Aberdeen analyses. NB reviewed EPSE coding. AM supported UCL analyses and study design.

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Table of Contents

Abstract	3
Impact Statement.....	5
Inclusion of Published Works.....	7
Author Contribution Statement	8
UCL Research Paper Declaration Form 1.....	9
UCL Research Paper Declaration Form 2.....	11
UCL Research Paper Declaration Form 3.....	13
UCL Research Paper Declaration Form 4.....	15
Acknowledgement.....	17
Abbreviations	24
1. General Introduction.....	26
1.1 A brief history of mental illness treatments	28
1.2 A brief history of mental illness medications	32
1.4 Psychiatric genetics and application in psychiatric treatment	36
1.4.1 Genome-wide Association Analyses	39
1.4.2 Polygenetic Risk Scores.....	41
1.4.3 Epigenome-wide Association Analyses	42
1.5 PhD Overview.....	43
2. Multiple Psychiatric Polygenic Risk Scores Predict Associations between Childhood Adversity and Bipolar Disorder.....	45
2.1 Abstract.....	45
2.2 Introduction.....	46
2.3 Methods.....	51
2.3.1 Participants.....	51
2.3.2 Study measures	52
2.3.3 Genotyping, imputation, and quality control	53
2.3.4 Calculation of the polygenic risk score (PRS).....	54
2.3.5 Statistical analysis	55
2.4 Results.....	57
2.4.1 Sample demographics	57
2.4.2 Correlations between multi-PRSs and the CLEQ scores	60
2.4.3 ACE/BD PRS associations & interaction with phenotypes	62
2.4.4 ADHD/MDD/SCZ PRS associations & interaction with phenotypes	62
2.4.5 PRS quantile analyses	65
2.4.6 PRS quantile interaction analyses results	68
2.5 Discussion	70
3. Implication of the ADCY1 Gene in Lithium Response in Bipolar Disorder by Genome-wide Association Meta-analysis	76
3.1 Abstract	76
3.2 Introduction.....	77
3.3 Methods.....	80

3.3.1	Participants.....	80
3.3.2	Study measures	81
3.3.3	Genotyping, imputation, quality control and meta-analyses	82
3.3.4	Lithium definition heterogeneity testing.....	82
3.3.5	GWAS meta-analysis & follow-on analyses.....	84
3.3.6	PRS calculation and analyses.....	88
3.4	Results.....	89
3.4.1	Sample demographics	89
3.4.2	Lithium definition heterogeneity testing.....	92
3.4.3	GWAS results for lithium responders vs non-responders	94
3.4.4	SNP-heritability estimation & genetic correlations	98
3.4.5	PRS analyses results	98
3.5	Discussion.....	99
4.	Integrating Genome-wide and Epigenome-wide Associations for Antipsychotic Induced Extrapyramidal Side Effects	103
4.1	Abstract	103
4.2	Introduction.....	104
4.3	Methods.....	107
4.3.1	Participants selection and genotyping.....	107
4.3.2	Coding of EPSE data	109
4.3.3	GWAS meta-analyses & follow-up analyses	111
4.3.4	EWAS methylation data.....	112
4.3.5	EWAS analysis and meta-analyses	113
4.3.6	EWAS findings integration and permutation test.....	114
4.3.7	PRS calculation and analyses.....	114
4.4	Results.....	115
4.5	Discussion	127
5.	Polygenic Risk and Cardiovascular Treatment Effects in Severe Mental Illness.....	131
5.1	Abstract	131
5.2	Introduction.....	132
5.3	Methods.....	135
5.3.1	Participants Selection and Genotyping	135
5.3.2	Study Measures	136
5.3.3	Genetic Data Imputation and Quality Control	136
5.3.4	PRS Calculations	137
5.3.5	Statistical analysis	138
5.4	Results.....	140
5.4.1	Sample demographics	140
5.4.2	Cardiovascular PRS Associations Results	144
5.4.3	BD & SCZ PRS Interaction Results	145
5.5	Discussion	151
6.	General discussion.....	155
6.1	PhD summary.....	155
6.2	General limitations.....	157

6.2.1	Limitations in measurements	157
6.2.2	Limitations in study design	159
6.2.3	Limitations in sample size	161
6.3	Genomic method limitations	162
6.3.1	GWAS limitations	162
6.3.2	PRS limitations	165
6.3.3	EWAS and integration limitations	166
6.4	Clinical implications	168
6.5	Future directions	169
6.6	Summary and conclusions	171
References		172
Appendices		229

Table of Tables

Table 2.1 Participants' Demographics and Clinical Characteristics concerning ACE Presence	59
Table 2.2 Results of Multiple Regression Analyses with Adjustments based on ACE (total score) and BD/ADHD PRS	63
Table 2.3 Results of Multiple Regression Analyses with Adjustments based on Total ACE Number and MDD/SCZ PRS	64
Table 2.4 Results of PRS Quantile Group Comparisons for the Odds Plot	66
Table 2.5 Results of Additional Interaction Analyses based on ACE Presence and BD/ADHD PRS Quantile Groups	69
Table 3.1 The Summary of Study Sample Sizes	87
Table 3.2 GWAS Meta-analyses Participants' Demographics and Clinical Characteristics concerning Lithium Response	91
Table 3.3 Binomial Sign Test Results for Response Heterogeneity	93
Table 3.4 Regions of the Genome showing the Strongest Association Signals with the Binary Trait	97
Table 4.1 GWAS Meta-analysis Participants' Demographics and Clinical Characteristics concerning EPSE Presence	117
Table 4.2 GWAS Participants' Demographics concerning EPSE Presence	118
Table 4.3 Regions of the Genome showing the Strongest Association Signals with EPSE Presence	121
Table 4.4 EPSE-associated differentially Methylated Positions	124
Table 4.5 Results of Multiple Regression Analyses	126
Table 5.1 Participant Demographics and Clinical Characteristics by Treatment Groups at Baseline Assessment	142
Table 5.2 Participant Demographics and Clinical Characteristics by Treatment Groups at 12 Month Follow-up	143
Table 5.3 Results of Multiple Regression Analyses with corresponding Cardiovascular PRS	148
Table 5.4 Adjusted Results of Cardiovascular and Psychiatric PRS Interactions	149

Table of Figures

Figure 2.1 Mean Polygenic Risk Scores for ADHD, BD, MDD, and SCZ across ACE Groups.....	61
Figure 2.2 The Odds Ratios of having ACE and Developing the Three Selected Sub-phenotypes with Reference to PRS Quantile Levels	67
Figure 3.1 The Manhattan Plots for Two Sets of GWAS Meta-analyses.....	96
Figure 5.1 By-group Scatter Plots comparing Baseline and 12-Month Follow-up Clinical Values against Associated PRS	147
Figure 5.2 Comparison of Baseline and 12-Month Follow-up Cholesterol Levels Across BD PRS Quartiles	150

Table of Appendices

2.1 OPCRIT Items 54 to 77	229
2.2 CLEQ QUESTIONNAIRE	234
2.3 Supplementary Table 1 Results of Multiple Regression Analyses with Adjustments based on ACE (total score) and BD/ADHD PRS	235
2.4 Supplementary Table 2 Results of Multiple Regression Analyses with Adjustments based on ACE (total score) and MDD/SCZ PRS.....	236
3.1 Supplementary Table 1 UCL Samples' Lithium Response Assessment.....	237
3.2 Supplementary Table 2 UCL samples' demographics and clinical characteristics concerning lithium response	238
3.3 Supplementary Figure 1 The QQ plot for the UCL+Song ^{Sub} +ConLi ⁺ Gen GWAS meta-analysis.....	239
3.4 Supplementary Figure 2 The QQ plot for the UCL+Song ^{Ob} +ConLi ⁺ Gen GWAS meta-analysis.....	240
3.5 Supplementary Figure 3 Regional association plot on chromosome 7 in which the Genome-wide significant SNP is located.....	241
4.1 Supplementary Table 1 Behavioural Key Words used to Identify EPSE Cases ..	242
4.2 Supplementary Table 2 Pharmacological Keywords used to Identify EPSE Cases	243
4.3 Supplementary Table 3 FGA Codes used to Select Participants.....	244
4.4 Supplementary Table 4. SGA Codes used to Select Participants.....	245
4.5 Supplementary Table 5 EPSE Medications Codes used to Select Cases	246
4.6 Supplementary Figure 1 The Manhattan Plot of EPSE GWAS Meta-analysis....	247
4.7 Supplementary Figure 2 The QQ Plot of EPSE GWAS Meta-analysis for FGA and SGA Combined	248
4.8 Supplementary Figure 3 The Manhattan Plot of EPSE GWAS Meta-analysis for any Exposure to FGA	249
4.9 Supplementary Figure 4 The QQ Plot of EPSE GWAS Meta-analysis for any Exposure to FGA	250
4.10 Supplementary Figure 5. The Manhattan Plot of EPSE GWAS Meta-analysis for only Exposure to SGA	251
4.11 Supplementary Figure 6 The QQ Plot of EPSE GWAS Meta-analysis for only Exposure to SGA	252
4.12 Supplementary Table 6 EWAS Participants' Demographics and Clinical Characteristics concerning EPSE Presence	253
4.13 Supplementary Figures 7–15 The Mapping of CpG to the Schizophrenia GWAS Region.....	254

Abbreviations

ACE	Adverse Childhood Experience
ADHD	Attention-Deficit/Hyperactivity Disorder
AIMS	The Abnormal Involuntary Movement Scale
AUDIT-C	Alcohol Use Disorders Identification Test Score
BARS	Barnes Akathisia Rating Scale
BD	Bipolar Disorder
BMI	Body Mass Index
CLEQ	Children Life Event Questionnaire
CM	Childhood Maltreatment
CpG	Cytosine-phosphate-Guanine
CS	Continuous Shrinkage
CVD	Cardiovascular Disease
CYP	Cytochrome P450
DALY	Disability-adjusted Life Year
GTE _x	Genotype-Tissue Expression
GSA	Global Screening Array
GSE	Genomic Structural equation modelling
EA	Emotional Abuse
EN	Emotional Neglect
ESRS	Extrapyramidal Symptom Rating Scale
EWAS	Epigenome-wide Association Study
EPSE	Extrapyramidal Side Effects
FDR	False Discovery Rate
FGA	First Generation Antipsychotic
FUMA	Functional Mapping and Annotation of Genome-Wide Association Studies
GLGC	Global Lipids Genetics Consortium
GWAS	Genome-wide Association Study
HDL	High-density Lipoprotein Cholesterol
IPAQ	International Physical Activity Questionnaire
IVW	Inverse-Variance Weighted
LD	Linkage Disequilibrium
LDL	Low-density Lipoprotein Cholesterol
MDD	Major Depressive Disorder
MMS	Morisky Scale of Adherence
Neff	effective sample sizes
NES	Normalized Effect Size
NHS	National Health Service
NICE	National Institute for Health and Care Excellence
OPCRIT	Operational Criteria Checklist
PA	Physical Abuse
PPD	Premorbid Personality Disorder
PGC	Psychiatric Genomics Consortium

PN	Physical Neglect
PRS	Polygenic Risk Score
SA	Sexual Abuse
SADS-L	Schizophrenia and Affective Disorder Schedule
SD	Standard Deviation
SE	Standard Error
SGA	Second Generation Antipsychotic
SSW	Sample Size Weighted
SCZ	Schizophrenia
SMI	Severe Mental Illness
SNP	Single-nucleotide Polymorphism
UCL	University College London
VIF	Variance Inflation Factor
WURS	Wender Utah Rating Scale
WGS	Whole-genome Sequencing

1. General Introduction

Mental illnesses are recognized as complex disorders that affect the mood, thought, and behaviour. Mental illnesses can be distinct from other medical conditions by their impact on cognition, particularly memory, self-awareness, and consciousness, while also being shaped by the unique life experiences and complexities that each individual carries (Malla et al., 2015).

Mental illnesses can be distinguished by their diverse forms. Varying disorders have different prevalence and impact. For instance, schizophrenia (SCZ) affects nearly 1% of the global population and is characterized by a combination of both positive symptoms such as psychotic symptoms and cognitive dysfunctions, and negative symptoms such as depressive moods and lack of motivations (Charlson et al., 2018). People diagnosed with the disorder can have a 15 year reduction in life expectancy compared with the general population (Hjorthøj et al., 2017). Meanwhile, Major Depressive Disorder (MDD) is characterized by a persistently low mood, diminished interest or pleasure in previously enjoyable activities, and recurrent thoughts of death. MDD affects about 185 million people, approximately 2.4% of the global population according to an estimate in 2019 (Marx et al., 2023). In addition, Bipolar Disorder (BD) is characterized by mood swinging between euthymia, (hypo-)mania and depressive episodes. BD has an estimated lifetime prevalence from 0.6% to 2.4% worldwide (Merikangas et al., 2011).

Mental illnesses are sometimes considered together because of overlapping symptoms. Distinguishing between SCZ and BD phenomenologically remain

1 challenging as they share symptoms, such as hallucinations, delusions, and mood
2 symptoms (Yamada et al., 2020). According to a recent study, more than a third of
3 patients with these disorders could face misdiagnosis (Ayano et al., 2021). Patients with
4 SCZ were likely to be misdiagnosed as having BD, while patients with BD were also
5 more likely to be misdiagnosed as having SCZ. The similar presentations of SCZ and
6 BD might reflect the overlapping biological underpinnings. For instance, genetic
7 findings suggest that these disorders might share a neurodevelopmental gradient of
8 psychopathology, accompanied by shared patterns of cognitive impairments (Doherty
9 & Owen, 2014). Meanwhile, SCZ and BD also exhibit considerable overlaps in terms
10 of morphological brain volume changes and functional connectivity
11 changes (O'Donoghue et al., 2017). Psychosis serves as an intermediate syndrome,
12 bridging SCZ and BD through shared symptomatology and neurological features. Thus,
13 SCZ, psychosis, and BD are often grouped together as severe mental illness (SMI) for
14 their shared functional impairments, treatment needs, and biological underpinnings.

15 Mental illnesses together account for a growing proportion of the global health
16 burden (Patel et al., 2018). Between 1990 and 2019, the proportion of global disability-
17 adjusted life years (DALYs) attributed to mental disorders, which quantify the burden
18 of a disease in terms of both premature death and disability, increased from 3.1% to 4.9%
19 (GBD Mental Disorders Collaborators, 2022). The rise reflects population growth,
20 aging, and improved reporting and recognition of mental illnesses globally. Such
21 findings also highlight the chronic nature of many mental disorders, which lead to
22 significant disability over time rather than early death and long-term treatments for

1 mental health are in need.

2 Views on mental illnesses and associated treatments have evolved significantly
3 throughout history (George et al., 2023). In earlier stages, mental disorders were often
4 attributed to supernatural forces, such as the Devil, with treatments like exorcisms,
5 prayer, and physical punishment (Porter, 2003). In the 1950s, breakthroughs in
6 psychiatric medications revolutionized mental health care and transformed treatments
7 by targeting the biological underpinnings (Duvall & Gallicchio, 2017; Pereira &
8 Hiroaki-Sato, 2018). In recent decades, advancements in neuroscience, genetics, and
9 personalized medicine have further refined treatments, with a focus on tailoring
10 interventions to an individual's psychological and biological profiles (Insel & Cuthbert,
11 2015). Modern mental health care prioritizes an integrative and holistic approach,
12 combining medication, psychotherapy, community-based support, and stigma reduction
13 efforts (George et al., 2023). However, a key challenge lies in determining the most
14 appropriate treatment plans tailored to everyone's unique needs. The following
15 introduction will outline key time points in the evolution of mental illness treatments
16 and explain how genomic methods can guide future mental health care, enabling more
17 personalized treatment planning.

18

19 **1.1 A brief history of mental illness treatments**

20 Throughout history, treatments for mental illnesses have undergone significant
21 transformations (Farreras, 2025). During the Middle Ages, explanations of mental
22 illnesses are dominated by the Church, attributing it to supernatural forces such as

1 possession by the Devil or evil spirits (Porter, 2003). Treatments were religious in
2 nature, involving exorcisms, prayer, and rituals like chanting or using holy water.
3 Severe cases often faced confinement, physical abuse, or execution. Medical theories,
4 such as Hippocrates' views on the biological basis of illness, were largely dismissed.
5 However, toward the end of the Middle Ages, the dominance of mystical explanations
6 began to wane, giving way to a gradual reemergence of scientific and medical
7 approaches to mental illness (Cantor, 2014).

8 The rise of humanism during the Renaissance further changed the situation by
9 emphasizing human welfare and individual uniqueness. In the mid to late 1500s,
10 physicians like Johann Weyer challenged Church dogma, arguing that many accused
11 "witches" were mentally ill rather than demonically possessed. This shift led to the
12 establishment of asylums where the mentally ill could be confined, as families struggled
13 to take care of their afflicted relatives (Laffey, 2003). During the 16th century, the
14 number of asylums increased as governments recognized that the growing population
15 of individuals with mental illnesses could no longer be managed in private homes.
16 However, these institutions, such as London's Bethlem Hospital, quickly became
17 overcrowded and notorious for inhumane treatment, where patients were chained,
18 exhibited for public entertainment, and subjected to harsh conditions (Walsh, 1907).

19 By the end of the 17th century and into the Age of Enlightenment, views on mental
20 illnesses shifted further, with an increasing focus on physical causes rather than moral
21 or spiritual ones. Harsh treatments including restraints and somatic methods persisted
22 but, there was also an emphasis on environmental management in asylums, such as

1 regulating diet and exercise (Laffey, 2003). The late 18th century saw the emergence of
2 the moral treatment movement, spearheaded by figures like Philippe Pinel in France
3 and William Tuke in England. This approach prioritized humane and respectful
4 treatment, emphasizing individual care, social support, and rehabilitation through
5 structured activities. Patients were unchained, given outdoor access, and treated with
6 kindness, leading to notable improvements in their conditions.

7 The success of moral treatment in the early 19th century fuelled the rapid expansion
8 of asylums across Europe and the United States. However, overpopulation, limited
9 resources, and discriminatory practices against immigrants led to overcrowding and a
10 decline in care quality (Scull, 2015). By the mid-19th century, asylums had evolved into
11 large and impersonal institutions, often prioritizing containment over therapeutic
12 intervention. While moral treatments highlighted the importance of humane care, it
13 became clear that additional medical advancements were needed to address more severe
14 cases effectively (Crossley, 2006).

15 The turn of the 20th century marked significant developments in understanding and
16 treating mental illnesses. Emil Kraepelin's classification system gained traction, notably
17 distinguishing mood disorders from what would later be termed schizophrenia. This era
18 also saw the rise of psychoanalysis, pioneered by Sigmund Freud, who emphasized the
19 role of unconscious motives and dialogue-based therapy in mental illness (Freud, 2009).
20 Alongside, behaviourism, introduced by John B. Watson and influenced by Ivan
21 Pavlov's conditioning principles, offered an alternative view, focusing on
22 reconditioning maladaptive behaviours (Chand et al., 2023; Malone, 2014). These

1 theoretical frameworks laid the foundation for modern psychopathology and treatment
2 approaches.

3 Institutions for mental healthcare underwent changes in both terminology and
4 function. Asylums were rebranded as hospitals, and patients replaced the stigmatizing
5 term "inmates." The mental hygiene movement gained momentum, advocating for
6 preventive approaches to mental illnesses and the establishment of public health clinics.
7 However, alongside these advances, troubling practices arose, including eugenics-
8 driven sterilization programs targeting institutionalized patients (P. A. Lombardo &
9 Dorr, 2006). Innovative yet controversial treatments emerged during this period. Insulin
10 coma therapy was introduced in 1927 but later abandoned due to its risks (Wellington,
11 2022), while electroconvulsive therapy (ECT) became a common substitute (Suleman,
12 2020). Lobotomy, a drastic psychosurgical procedure, was widely performed during the
13 mid-20th century, reflecting a lack of effective alternatives (Faria, 2013). Post-World
14 War II, the need for standardized mental health care for veterans led to the publication
15 of the first *Diagnostic and Statistical Manual of Mental Disorders* (DSM) in 1952. This
16 framework evolved over time, with the DSM-III (1980) marking a shift toward
17 diagnostic neutrality regarding the causes of mental illnesses, a position maintained in
18 subsequent editions (Pichot, 1986). Nowadays, diagnoses tools like DSM-V and ICD-
19 10 are still widely used although challenges in diagnoses and classifications persisted
20 (Hirsch et al., 2016; Kawa & Giordano, 2012).

1.2 A brief history of mental illness medications

Starting in the 1950s, medications were introduced for the treatment of mental illnesses, bringing an immediate and significant impact (Braslow & Marder, 2019). The first noticeable psychiatric medication, lithium, could date back to 1949 when John Cade first used it to treat maniac patients (Duvall & Gallicchio, 2017). Lithium then became a foundational treatment for bipolar disorder, though its efficacy varies among patients. Two revolutionary drugs reserpine and chlorpromazine, were introduced into psychiatry in the early 1950s (Baumeister, 2013; Braslow & Marder, 2019). Reserpine depletes monoamines in the brain, while chlorpromazine blocks dopamine receptors, a discovery that transformed the understanding of chemical communication in the brain. Early antidepressants, such as monoamine oxidase inhibitors (MAOIs) and tricyclic antidepressants (TCAs) were also introduced in the 1950s then dominated the field until the 1980s (Chockalingam et al., 2019).

The growing range of available drugs led to 47.2 million psychotropic drug prescriptions being dispensed under the National Health Service (NHS) between 1965 and 1970 (Haggett, 2015). However, concerns quickly arose due to inadequate training, aggressive advertising, and the simultaneous rise in prescribing, drawing significant criticism from the medical press. Meanwhile, clinicians started to notice that psychiatric medications could have significant side-effects (Evarts & Butler, 1959). Concerns about side effects have shaped the use of psychiatric medications today. Atypical antipsychotics, introduced as alternatives to typical antipsychotics, offered fewer extrapyramidal side-effects (EPSE) but introduced metabolic issues like weight gain

1 and glucose dysregulation (Nasrallah, 2008). Drugs like clozapine, though highly
2 effective for treatment-resistant SCZ, remain underutilized due to concerns about
3 agranulocytosis and other safety risks (Kendall, 2011). Over the past 70 years,
4 antipsychotic drugs have evolved through first- (e.g., chlorpromazine and fluphenazine)
5 and second-generation medications (e.g., olanzapine and clozapine), with a potential
6 third generation (e.g., aripiprazole). However, there remains a critical need for the
7 development of more effective treatments with a lower side-effect profile than existing
8 ones (Weston-Green, 2022).

9 Nowadays, the use of antipsychotics and antidepressants is widespread, with up to
10 17% of adults being prescribed antidepressants in high-income countries (Pillinger et
11 al., 2023). The prevalence of antipsychotic prescriptions doubled in England increased
12 from 0.6% in 2000 to 1.2% in 2014 and continues to grow (Shoham et al., 2021). While
13 these drugs offer relief for many individuals, around 74% of patients with chronic
14 schizophrenia could discontinue their assigned treatment before 18 months owing to
15 inefficacy or intolerable side effects (Lieberman et al., 2005). Around 75% of patients
16 report experiencing side effects, which range from mild to severe (Iversen et al., 2018).
17 Non-adherence to psychotropic medication due to concerns of inefficacy and side-
18 effects is widespread, with an estimated overall prevalence of 49% (Semahegn et al.,
19 2020). Non-adherence occurs in approximately 56% of individuals with SCZ, 50% with
20 MDD, and 44% with BD. These statistics nevertheless underscore the variability in
21 treatment response and highlight the pressing need for more robust predictive
22 approaches to guide and improve treatment planning.

Adverse side effects can diminish quality of life, hinder daily functioning, contribute to morbidity and mortality, and foster stigma surrounding mental illness (Tandon et al., 2020). Additionally, concerns about inefficacy and side effects often lead to poor adherence, increasing the likelihood of relapse (Haddad et al., 2014). For individuals with depression and schizophrenia, the impact of side effects plays a significant role in medication decisions, often becoming a primary concern when evaluating treatment options (Hopwood, 2020; Kaar et al., 2019).

The psychiatric medication advancements provided effective treatment options for individuals with mental illnesses. However, treatment response is influenced by multiple factors (McCutcheon et al., 2015). In addition, different psychiatric medications vary widely in their side effect profiles, further complicating treatment decisions (Pillinger et al., 2023). To optimize psychiatric treatment, it is crucial to identify characteristics that distinguish patients who respond best to specific medications, allowing for more personalized and effective therapeutic strategies.

1.3 Predictions of treatment response

To increase the success of treatments, specific clinical characteristics have been identified as potential indicators of treatment response. Factors such as medication plasma levels, adherence, metabolic differences, co-prescriptions, lifestyle habits, sex, and the possibility of alternative diagnoses could all influence treatment responses (McCutcheon et al., 2015). For depression, factors such as depression subtype, symptom history, age of onset, episode duration, and recurrence all play roles in

1 determining treatment effectiveness. For instance, early-onset depression is linked to
2 increased resistance to certain antidepressants, while recurrent and prolonged
3 depressive episodes also correlate with poorer outcomes (Perlman et al., 2019). In
4 addition, patients with severe baseline symptoms, histories of trauma, cognitive
5 impairments, or anxious symptomatology tend to exhibit reduced antidepressant
6 efficacy. In BD, factors predicting lithium response include the sequence of manic and
7 depressive episodes, the absence of rapid cycling or psychotic symptoms, a family
8 history of BD, and a shorter duration of illness before lithium treatment begins (Hui et
9 al., 2019). Conversely, higher body mass index and a greater number of prior episodes
10 or hospitalizations have been associated with poorer lithium response. In schizophrenia,
11 a higher severity of positive symptoms and the use of atypical antipsychotics,
12 particularly clozapine, are associated with the greatest likelihood of treatment response
13 (Seppälä et al., 2021). Risk factors for extrapyramidal side-effects (EPSE) from
14 antipsychotics include a history of prior episode and high medication dose (Hedenmalm
15 et al., 2006) and older age (Jeste, 2004).

16 Given the substantial variability in individual responses to psychiatric medications,
17 careful monitoring of physical health parameters has been recommended to optimize
18 treatment (Hiemke et al., 2018). Taking lithium as an example, clinical guidelines
19 emphasize the importance of tracking body mass index, kidney function, body lithium
20 levels and thyroid activity in patients with bipolar disorder receiving lithium treatment,
21 with ongoing dose adjustments to ensure therapeutic efficacy (Godden, 2024; NICE,
22 2014a). However, this process can be time-consuming, requiring extensive trial and

1 producing error before an optimal plan is tailored to everyone's needs. One promising
2 alternative approach to improve this process is the integration of pharmacogenomic
3 (PGx) testing, which examines genetic variations that affect drug metabolism and
4 response. By guiding medication selection and dosing decisions, PGx testing can help
5 personalize treatment, reduce adverse effects, and enhance therapeutic outcomes from
6 the onset. The following sections will introduce how PGx testing could guide future
7 treatments and showcase several relevant genomic methods which could enrich PGx
8 testing.

9

10 **1.4 Psychiatric genetics and application in psychiatric treatment**

11 For centuries, observations have indicated that mental illnesses tend to run in
12 families, a pattern that has also been confirmed by large-scale population studies
13 (Arribas-Ayllon et al., 2019). Individuals with a first-degree relative affected by BD or
14 schizophrenia face a significantly higher risk of developing these conditions themselves,
15 approximately six to ten times greater (Andreassen et al., 2023; Lichtenstein et al.,
16 2009). Furthermore, familial risk is not limited to a single disorder; relatives of
17 individuals with psychiatric illnesses also have an increased likelihood of developing
18 other mental health conditions, suggesting a shared underlying aetiology (Cardno &
19 Owen, 2014). Additional complexity arises from non-inherited genetic variations that
20 accumulate in brain tissue over time, as well as random biological processes that further
21 shape individual risk profiles (Andreassen et al., 2023). Nevertheless, genetic variations
22 in parents could also influence the environments they create for their offspring, adding

1 further complexity (Kong et al., 2018).

2 With the advent of molecular genetics in the 1980s and 1990s, researchers began
3 using linkage and candidate gene studies to identify genetic variants associated with
4 mental disorders (Owen et al., 2000). However, many early findings failed to replicate
5 due to small sample sizes and the complex polygenic nature of psychiatric genetics
6 (Sullivan et al., 2012). It became increasingly evident that most psychiatric conditions
7 result from the combined effects of multiple genes, each contributing only a small
8 increase or decrease in liability, along with environmental influences, a concept known
9 as polygenic multifactorial causation (Owen & Cardno, 1999). The field advanced
10 significantly in the 2000s with the introduction of genome-wide association studies
11 (GWAS) and large-scale collaborative efforts, particularly from the Psychiatric
12 Genomics Consortium (PGC; Sullivan et al., 2018). The PGC has played a crucial role
13 in systematically evaluating common single-nucleotide polymorphisms (SNPs), rare
14 variants, gene sets, and pathways, shedding light on the genetic and biological
15 foundations of mental illnesses (Watson et al., 2020). More recently, approaches such
16 as polygenic risk scores (PRS), whole-genome sequencing (WGS), and multi-omics
17 integration have provided deeper insights into the genetic architecture of mental
18 illnesses and their interactions with environmental factors (Mullins et al., 2021, 2022;
19 O'Connell et al., 2025). These advancements continue to pave the way for a more
20 comprehensive understanding of mental disorders and become the foundation for
21 potential precision medicine approaches in psychiatry, guiding future psychiatric
22 treatment medication development.

1 Meanwhile, the importance of applying genomic methods to predict current
2 psychiatric medication efficacy and side effect profiles is increasingly recognized,
3 offering new insights into personalized treatment strategies (Goetz & Schork, 2018).
4 For instance, the cytochrome P450 (CYP) enzymes, *CYP2C19* and *CYP2D6*, hold
5 significant clinical relevance for antidepressants (Carvalho Henriques et al., 2020).
6 Research indicates that genetic variants in *CYP2C19* and *CYP2D6* affect antidepressant
7 blood concentrations, adverse drug reactions, and, to an extent, clinical outcomes such
8 as treatment discontinuation and symptom response (Bråten et al., 2020; Jukić et al.,
9 2018). However, it is important to note that not all psychiatric medications' genetic
10 profiles are fully documented yet, and additional investigations are needed for other
11 components of the medication response such as side-effects (Bousman et al., 2021).

12 As psychiatric genetics continues to evolve, it holds the promise of transforming
13 mental health treatments by providing more precise and biologically informed
14 approaches. Genetic research has already shed light on the heritability and familial
15 patterns of psychiatric disorders, and its integration with pharmacogenomics may soon
16 truly enable personalized medication strategies. Moving forward, the field must
17 continue refining its understanding of genetic influences on psychiatric conditions and
18 their treatment responses, paving the way for more effective and individualized
19 therapeutic interventions. The following sections will introduce several genomic
20 methods that could be applied to study psychiatric medication responses.

1.4.1 Genome-wide Association Analyses

GWAS on psychiatric treatment responses stands out as a promising approach for pinpointing potential genes linked to medication response and side effects. GWAS analyses genome-wide SNP data to detect associations between genetic variants and specific phenotypes. While whole-genome and exome sequencing provide alternative approaches, most GWAS rely on genotyped SNP arrays combined with imputed variants to enhance genomic coverage (Tam et al., 2019). Imputation, based on reference haplotypes, enables researchers to infer untyped variants, reducing the need for direct genotyping while increasing cost efficiency and sample sizes (Das et al., 2016). Given that GWAS tests millions of SNPs across the genome, stringent multiple-testing correction is required. The widely accepted genome-wide significance threshold is $p < 5 \times 10^{-8}$, accounting for approximately one to two million independent statistical tests (M. I. McCarthy et al., 2008).

With the increasing number of GWAS, researchers have realized that most psychiatric disorders could be polygenic which each SNP may only carry small to modest phenotypic effects (de Bakker et al., 2008). Meta-analysis of GWAS, which enhance detection of SNP with modest effect size through increased sample size and power from collaborative efforts, is increasingly recognized as important. Tools like METAL has been developed to efficiently meta-analyse GWAS summary statistics while using Inverse-Variance Weighted (IVW) or Sample Size Weighted (SSW) methods considering either the standard error or sample size to generate combined outputs (Willer et al., 2010).

1 Psychiatric genetics benefited hugely with PGC's collaborative efforts worldwide
2 on GWAS meta-analyses. Taking BD genetics as an example, one of the first PGC BD
3 GWAS identified 34 independent SNPs with strong evidence for replication in 4,493
4 independent BD cases and 42,542 independent controls (Sklar et al., 2011). Then, with
5 an increased sample size of 41,917 BD cases and 371,549 controls of European ancestry,
6 64 associated genomic loci were identified (Mullins et al., 2021). More recently, by
7 combining participants of European, East Asian, African American and Latino
8 ancestries covering 158,036 cases with BD and 2.8 million controls, PGC reported 298
9 genome-wide significant loci which mapped to 36 credible genes in the aetiology of
10 BD (O'Connell et al., 2025). Notably, one of the top SNP from the meta-analysis,
11 rs6693503, had an odds ratio of 1.048 with a standard error of 0.007, while most other
12 reported significant SNPs had odds ratios ranging from 0.9 to 1.1 (O'Connell et al.,
13 2025). Such findings again highlight the polygenic nature of psychiatric disorders and
14 emphasize the importance of larger sample size from collaborations for detecting true
15 genetic effects.

16 More and more researchers have started to apply similar GWAS methods to
17 investigate psychiatric medication responses (Li et al., 2020; Zai et al., 2023). The genes
18 identified through GWAS provide insights into the underlying mechanisms of these
19 medications which could be used as targets to monitor in the clinic. With clearer
20 understanding of each psychiatric medication's metabolism and effectiveness profiles,
21 more effective drugs could also be developed. However, most of current psychiatric
22 medication response GWAS had limited sample size and restricted population groups.

1 Collaborative efforts are needed to further advance our understanding and guide future
2 discoveries.

3

4 **1.4.2 Polygenetic Risk Scores**

5 Although GWAS could be used to investigate common SNPs, each SNP's
6 influence is often modest with an odds ratio typically ranging from 0.7 to 1.3 (Mullins
7 et al., 2021; Trubetskoy et al., 2022). Polygenic risk scores (PRS), could summarize the
8 potential risk alleles from the GWAS, weighted by their respective effect sizes to
9 produce a summary score which indicate an individual's genetic liability to the
10 phenotype (Dudbridge, 2013). Before computing PRS, a pruning and thresholding (P +
11 T) approach is typically applied (Purcell et al., 2007). Pruning (or clumping) removes
12 highly correlated SNPs based on linkage disequilibrium (LD), ensuring that only
13 independent SNPs are retained. Thresholding involves applying different p-value
14 cutoffs to exclude SNPs with weak associations with the trait, optimizing PRS
15 predictive power. Advanced methods for PRS computation have been developed to
16 capture complex genetic architectures, such as those using a Bayesian framework (Ge
17 et al., 2019; Lloyd-Jones et al., 2019).

18 Researchers also discovered psychiatric PRS' effectiveness in the prediction of
19 medication responses. For instance, studies by ConLi+Gen, (2018) and Amare et al.
20 (2021) have demonstrated that PRS for SCZ and MDD among patients with BD are
21 negatively associated with their response to lithium. However, PRS could only explain
22 less variance than the total genetic variance identified in GWAS, as they are constrained

by the limitations of current GWAS methodologies. Thus, more advanced GWAS may also further enhance findings from psychiatric medication response PRS.

1.4.3 Epigenome-wide Association Analyses

Epigenome-Wide Association Studies (EWAS) investigate the relationship between epigenetic modifications, primarily DNA methylation and complex disease phenotypes. Unlike GWAS, which focus on inherited genetic variants, EWAS assess modifications that are heritable and/or environmentally modifiable, providing insights into gene regulation mechanisms beyond DNA sequence variation (Campagna et al., 2021; Villicaña & Bell, 2021). The most widely studied epigenetic mechanism is DNA methylation, where methyl groups are added to cytosine-phosphate-guanine (CpG) dinucleotides, influencing gene expression. The typical significance threshold for EWAS is $p < 1 \times 10^{-7}$. This threshold accounts for the large number of CpG sites tested (~450,000 on Illumina arrays) and is similar to the multiple testing correction used in GWAS (Liu et al., 2013).

EWAS can capture dynamic, environmentally influenced epigenetic modifications, such as DNA methylation changes induced by long-term medication use, stress, or lifestyle factors. EWAS have also been applied to investigate psychiatric medication responses, offering a promising approach to understand gene-environment interactions in treatment outcomes (Engelmann et al., 2022). Despite the potential, EWAS of psychiatric medication response are often limited by smaller sample sizes, as large, well-characterized cohorts with both methylation data and detailed medication response

1 phenotypes remain scarce.

2

3 **1.5 PhD Overview**

4 This PhD aimed to explore the genome to identify potential biomarkers that
5 differentiate psychiatric phenotype development and treatment responses. The findings
6 emphasized the importance of personalized treatment approaches, which could
7 significantly improve patient outcomes. To achieve this, the research incorporated
8 large-scale genetic analyses, including GWAS, EWAS, and PRS predictions, while
9 integrating significant discoveries to better understand the complex biological effects.

10 In my first research paper (Chapter 2), I examined how multiple psychiatric PRS
11 interact with environmental factors, adverse childhood experiences (ACE), to influence
12 the development of various BD phenotypes. This included age of onset, the presence of
13 psychotic symptoms, suicide ideation, and rapid cycling. The analyses aimed to shed
14 light on how genetic and environmental factors combine to shape the clinical
15 presentation of BD. In Chapter 3, I used a GWAS meta-analysis approach to identify
16 SNP associated with a positive lithium response among BD patients. I also examined if
17 psychiatric PRS could predict positive lithium response. This investigation sought to
18 pinpoint genetic markers that could predict which individuals are more likely to benefit
19 from lithium treatment. In Chapter 4, I integrated an EWAS meta-analysis with
20 associated GWAS findings to uncover biomarkers linked to extrapyramidal side effects
21 (EPSE) in individuals with long-term exposure to antipsychotic medications. This
22 chapter provided insights into the influence from both genetic and epigenetic factors

1 resulting from environmental treatment factors. In Chapter 5, I explored how genetic
2 risk factors, assessed through PRS, could predict cardiovascular treatment responses in
3 patients with severe mental illness (SMI). This research aimed to understand the role of
4 genetics in shaping the effectiveness of cardiovascular treatments. These results could
5 help patients in planning more personalized and effective health treatment. This thesis
6 concluded in chapter 6 where I discussed the potential implications and limitations from
7 current findings on using genomic analyses to guide PGx testing and support
8 personalized treatment planning.

2. Multiple Psychiatric Polygenic Risk Scores Predict Associations between Childhood Adversity and Bipolar Disorder

This paper has been expanded from my previous MSc project and a version of this paper has been published at Journal of Affective Disorder (Yao et al., 2023).

2.1 Abstract

Background: It remains unclear how adverse childhood experiences (ACE) and increased genetic risk for bipolar disorder (BD) interact to influence BD symptom outcomes. Here we calculated multiple psychiatric polygenic risk scores (PRS) and used the measures of ACE to understand these gene-environment interactions.

Method: 885 BD subjects were included for analyses. BD, ADHD, MDD and SCZ PRSs were calculated using the PRS-CS-auto method. ACEs were evaluated using the Children Life Event Questionnaire (CLEQ). Participants were divided into groups based on the presence of ACE and the total number of ACEs. The associations between total ACE number, PRSs and their interactions were evaluated using multiple linear and logistic regressions. Secondary analyses were performed to evaluate the influence of ACE and PRS on sub-phenotypes of BD.

Results: The number of ACEs increased with the ADHD PRS. BD participants who had experienced ACE showed an earlier age of BD onset and higher odds of having rapid cycling. Increased BD PRS was associated with increased odds of developing psychotic symptoms. Higher ADHD PRS was associated with increased odds of having rapid cycling. No prediction effect was observed from MDD and SCZ PRS. And, we found no significant interaction between ACE numbers and any of the PRSs in

1 predicting any selected BD sub-phenotypes. The study was limited by sample size, ACE
2 definition, and cross-sectional data collection method.

3 *Conclusions:* The findings consolidate the importance of considering multiple
4 psychiatric PRSs in predicting symptom outcomes among BD patients.

6 **2.2 Introduction**

7 BD is recognised as a complex psychiatric disorder because of its variable clinical
8 presentation. The first episode of mania or depression of BD commonly occurs between
9 the ages of 18 and 24 (McMahon et al., 1994). However, early onset before age 18 is
10 not rare and such cases are often associated with more severe or complex BD
11 phenotypes, such as increased comorbidity and delayed treatment response (Joslyn et
12 al., 2016). While some BD patients may experience rapid cycling of episodes within
13 hours or days, in others, episodes are separated by months or years (Carvalho et al.,
14 2014). BD patients may also have common comorbidities such as anxiety (Spoorthy et
15 al., 2019), substance abuse (Jawad et al., 2018), and psychotic features (Maggioni et al.,
16 2017). In addition, BD patients are at a higher risk of performing suicidal behaviour
17 compared to other psychiatric patients (Jamison, 2001).

18 Both genetic and environmental factors are recognised to be important in the
19 development of BD phenotypes. Environmental risk factors such as adverse childhood
20 experiences (ACE) and childhood maltreatment (CM) have been closely associated
21 with BD phenotype development. ACEs describe adverse events, such as the death or
22 divorce of a caregiver, serious illness, or hospitalisation. According to the past meta-

1 analysis by Palmier-Claus et al., (2016), BD patients were 2.63 times more likely to
2 have experienced ACE. ACE has also been found to play a significant role in the risk
3 of relapse in BD patients (Hosang et al., 2010). Meanwhile, CM describes traumatic
4 abusive events. CM is separated into emotional abuse (EA), physical abuse (PA), and
5 sexual abuse (SA), as well as emotional neglect (EN) and physical neglect (PN) (Gilbert
6 et al., 2009). CM is commonly recognised as a more severe form of ACE. CM has also
7 been associated with more severe and complex BD phenotypes (Agnew-Blais & Danese,
8 2016). These include earlier age of onset and a higher risk of comorbidities such as
9 anxiety disorder, substance misuse disorder, suicide attempts, and increased mood
10 episodes.

11 Examining the influence of both genes and the environment is essential for
12 understanding BD symptom outcomes (Quidé et al., 2020). It is commonly believed
13 that existing biological vulnerabilities interact with the later experiences such ACE and
14 lead to more complex mental health conditions. Thus, individuals who already carry a
15 high genetic risk for BD are at even higher risk of developing BD if they have
16 experienced ACE (Duffy et al., 2020). Recently, researchers have begun exploring such
17 gene and environment interaction effects in predicting the development of BD sub-
18 phenotypes (Aas et al., 2020; Anand et al., 2015; Park et al., 2020). These studies were
19 based on the development of genome-wide association studies (GWAS) and polygenic
20 risk score (PRS) calculations (see sections 1.4.1 and 1.4.2 for details). The calculation
21 of polygenic risk scores generally has two primary purposes (1) to predict the likelihood
22 of developing the outcome of interest by synthesizing existing GWAS information into

1 a single measure; (2) to estimate the predictive ability of known effects. Therefore,
2 PRSs can classify the relative risk of a specific outcome for an individual within a
3 population and have been applied in different gene-environment interaction studies.

4 The gene-environment interaction can be additive that the effects of genes and
5 environment both independently contribute to the risk thus, the effects simply sum
6 together (Rami et al., 2025). In contrast, the interaction effects can also be multiplicative
7 that the effects of genes and environment amplify one another beyond a simple sum.
8 Thus, the presence of both risk factors together may result in a higher risk than would
9 be expected if their effects were additive (Lin et al., 2025).

10 However, there have been recent conflicts regarding how BD PRS is associated
11 with the number of ACEs and how BD PRS interacts with ACE in the predictions of
12 various BD sub-phenotypes (Aas et al., 2020; Park et al., 2020). For example, Aas et al.
13 (2020) showed that BD PRS might only interact with childhood maltreatment in
14 predicting the risk of rapid cycling. In comparison, Park et al. (2020) observed another
15 interaction effect in predicting the age at onset (AAO) of BD. These differences
16 between results may be due to different ACE definitions or clinical heterogeneity
17 between sample cohorts. For instance, Aas et al.'s (2020) measures of ACE were mainly
18 CM, while Park et al.'s (2020) measures were mainly ACE, with an additional CM item
19 on physical abuse.

20 There has been ongoing debate regarding how ACEs should be defined. The
21 original ACE framework proposed by Felitti et al. (1998) encompassed three domains:
22 abuse, neglect, and household dysfunction (e.g. domestic violence, and parental

1 separation/divorce). However, following scholars have broadened the ACEs definitions
2 to include other adversities, including discrimination, peer victimization, unsafe
3 neighbourhoods, and socioeconomic problems further increasing conceptual
4 inconsistency (Karatekin & Hill, 2019). It is important to distinguish between
5 independent events (e.g. parental death, financial hardship, or divorce) and
6 victimization experiences (e.g. neglect and abuse), as they may differ in developmental
7 impact and intervention implications (Fitzgerald & Gallus, 2025; McLaughlin, 2017).
8 The lack of a clear and consistent definition in past studies obscures the distinct effects
9 of different adverse experiences.

10 Another major limitation of both these past studies is that they did not consider an
11 individual's genetic liability to other psychiatric disorders (Grigoriu-Serbanescu et al.,
12 2020). Given that BD has a high level of overlapping clinical heterogeneity and shared
13 genetic risk with other psychiatric disorders, it seems necessary to include multiple
14 psychiatric PRSs and their interactions with ACE to predict phenotype development
15 (Baldwin et al., 2022; Coombes et al., 2020).

16 Differentiating between attention-deficit/hyperactivity disorder (ADHD) and BD
17 remains as a challenge for clinicians, as these two disorders have extensive symptom
18 overlap, reciprocal comorbidity, and overlapping age of onset periods (Brus et al., 2014;
19 Marangoni et al., 2015). ADHD and BD were proven to be genetically correlated
20 (r_g 0.121) and share common risk variants (O'Connell et al., 2021). Children with
21 ADHD were more likely to have higher ACE exposure than children without ADHD
22 (Brown et al., 2017). And the higher exposure to ACE may also increase their risks to

1 develop ADHD (Crouch et al., 2021). The presence of ADHD symptoms can be an
2 indication of worse depression presentation (Powell et al., 2021) and is linked to rapid
3 cycling between BD episodes (Aedo et al., 2018).

4 In addition, genetic predisposition to MDD can be predictive of specific patterns
5 of depression symptoms (Martínez-Levy et al., 2021). An additive interaction effect
6 between genetic predisposition to depression and trauma exposure has been observed
7 on depressive symptoms among MDD patients (Thorp et al., 2023). Meanwhile, SCZ
8 PRS can also be predictive of schizophrenia patients' psychotic symptoms, cognition,
9 illness severity, and diagnostic changes (K. G. Jonas et al., 2019). However, no previous
10 study specifically examined how these different psychiatric genetic liabilities interact
11 with environment risk factors such as ACE in predicting the development of BD sub-
12 phenotypes.

13 To address these research gaps, we wanted to replicate past gene x environment
14 (GxE) findings on ACE and BD PRS. We also wanted to explore these multiple PRSs'
15 prediction effects of BD sub-phenotype development. Thus, our research had three main
16 aims:

- 17 (1) to clarify the association between BD PRS and ACE;
- 18 (2) to investigate how the number of ACEs interacts with multi-PRSs in the
19 predictions of distinct BD sub-phenotypes;
- 20 (3) to understand whether ACE could predict BD sub-phenotype development
21 and change the interaction effect when CM items are absent.

Our objective was to use multiple linear and logistic regressions to explore the association between the PRSs and the ACE numbers, including their interaction effect in predicting the associated sub-phenotypes. We hypothesized that the multi-PRSs would interact with ACEs in predicting the sub-phenotype development. People with higher PRSs (BD/ADHD/MDD/SCZ) and more ACEs were predicted to have developed BD at a younger age and developed more severe or complex sub-phenotypes, such as psychosis symptoms, suicide ideation, and rapid cycling.

2.3 Methods

2.3.1 Participants

All BD cases received an ICD10 diagnosis of BD from a UK National Health Service (NHS) psychiatrist (World Health Organization, 1992). Ancestrally matched healthy controls (n = 1818) were recruited from the NHS blood transfusion service and from study sites where case participants were also being recruited. The controls were screened for an absence of a lifetime history of the following disorders: schizophrenia and any other psychosis, major affective or schizoaffective disorders, eating disorders, alcohol/drug addiction, and obsessive-compulsive disorders. All participants were of English, Scottish, Welsh, or Irish descent and had at least three out of four grandparents of the same descent. The study was approved by the NHS Metropolitan Multi-centre Research Ethics Committee (MREC/03/11/090). All participants read an approved information sheet and signed a physical informed consent form.

2.3.2 Study measures

Semi-structured interviews were performed with BD participants using the lifetime version of the Schizophrenia and Affective Disorder Schedule (SADS-L; Spitzer et al., 1978), the 90-item Operational Criteria Checklist (OPCRIT; McGuffin et al., 1991) and the Children Life Event Questionnaire (CLEQ; Monaghan et al., 1979). In a subset of participants, the 25-item version of the Wender Utah Rating Scale (WURS) was used to measure childhood ADHD symptoms (Grigoriu-Serbanescu et al., 2020; M. F. Ward et al., 1993). A subset of the participants received lithium treatment prior to the assessment and their response to lithium was scored using data from clinicians and the research participant. Where these scores differed the clinician's rating was used. The data was coded in a binary format to differentiate between responders and non-responders. Participants' premorbid personality disorder (PPD) diagnosis was assessed with OPCRIT item 11. BD age of onset, presence of psychosis symptoms, presence of suicide ideation, and presence of rapid cycling were selected for analyses in the current study. These variables were chosen because of their relevance to BD according to past reviews (Escamilla & Zavala, 2008; Palmier-Claus et al., 2016). We also sought to replicate previous findings from Park et al. and Aas et al. (Aas et al., 2020; Park et al., 2020).

In this research, BD age of onset was defined as the earliest age at which symptoms began to cause subjective distress or impair functioning or at which medical advice was sought for psychiatric reasons. Age at onset was stored as a continuous variable (OPCRIT item 4). The presence of psychotic symptoms was defined as whether the

1 participants disclosed any symptoms of impaired reality testing (any presence of
2 OPCRIT items 54-77; see Appendix 2.1). The presence of suicide ideation was defined
3 as whether the participants with BD reported thoughts of death (not necessarily their
4 own), thinking of suicide, wishing to be dead, or attempting to kill themselves (OPCRIT
5 item 43). The presence of rapid cycling was defined as whether the patient ever had
6 four or more mood disturbances in one year. These latter three variables were stored in
7 binary format.

8 ACEs were measured separately using the CLEQ, an adaption of the children's life
9 events inventory, which records the ACE experienced before the age of 16 (Monaghan
10 et al., 1979). The CLEQ included 13 binary "YES" or "No" questions. The first 12
11 questions had content such as the death of a parent; the death of a brother or sister;
12 serious illness; hospitalisation of a parent; teenage pregnancy; suspension from school
13 (see CLEQ in Appendix 2.2). The last question required the participants to specify other
14 significant adverse life events experienced as a child if they answered "YES." Due to
15 the variability of events recorded with question 13, we only focused on the first 12
16 CLEQ questions. If the participant answered "Yes" to any of the first 12 questions, this
17 indicated the experience of ACE. The number of "YES" answers to all 12 questions
18 indicated how many ACEs each participant had been through.

19

20 **2.3.3 Genotyping, imputation, and quality control**

21 Genome-wide single nucleotide polymorphism data was available for the BD and
22 healthy control subjects. The data was generated in two waves at the Broad Institute,

1 Boston, MA, US, using the Illumina PsychArray and Illumina Global Screening Array
2 (GSA). The quality control and imputation methods used for the PsychArray had been
3 described elsewhere (Grigoriu-Serbanescu et al., 2020). The genotype data from the
4 GSA underwent equivalent quality control and imputation procedures as for the
5 PsychArray.

6

7 **2.3.4 Calculation of the polygenic risk score (PRS)**

8 Patients' multi-PRSs (BD/ADHD/MDD/SCZ) were computed using imputed data
9 from the PsychArray and GSA with the PRS-CS-auto method which provides a single
10 score for each sample without any thresholds (Ge et al., 2019). The PRS-CS-auto
11 method was chosen over other methods since it outperformed other existing methods
12 according to the simulation studies by Ge et al. (2019). Pain et al. (2021) also found
13 PRS-CS-auto to be the best of the pseudo-validation PRS methods. PRS-CS is distinct
14 as it utilises a high-dimensional Bayesian regression framework and places a
15 continuous shrinkage (CS) before SNP effect size calculations. Such procedures result
16 in substantial computational advantages and enable multivariate modelling of local LD
17 patterns, which makes PRS-CS robust to varying genetic architectures.

18 The application of the PRS-CS method required an LD reference panel and
19 reference GWAS summary statistic, which help infer the posterior effect sizes of SNPs.
20 We chose the European sample from The 1000 Genomes Project Consortium (2010) as
21 our LD reference panel. The BD reference GWAS came from Mullins et al. (2021)
22 which included the PsychArray samples used in the current study. We therefore used

1 summary statistics generated without the overlapping samples to avoid confounding.
2 Our ADHD reference GWAS came from the Psychiatric Genomics Consortium, a meta-
3 analysis including 38,691 ADHD cases (Demontis et al., 2022). We used data from
4 Howard et al. (2019) for MDD GWAS which excluded UK Biobank and 23andMe data
5 covering 170,756 cases and 329,443 controls. The SCZ GWAS came from Trubetskoy
6 et al. (2022) which covered 68,676 cases and 96,079 controls. We adapted all GWAS
7 samples to be based on subjects of European ancestry and data from the samples that
8 we had contributed to the PGC schizophrenia analyses excluded our own lab's data.
9 The new GWAS generation followed the same procedures as described by each GWAS
10 paper. The reference GWAS sample sizes were calculated using the effective sample
11 size method (Neff) as $4/(1/N_{\text{cases}} + 1/N_{\text{controls}})$, where N_{cases} is the number of
12 cases and N_{controls} is the number of controls.

13 We used the mean and standard deviation of the PRS from the healthy controls to
14 standardise the PRS data of the BD cases. This was performed separately for the
15 PsychArray and the GSA data. By standardising the PRSs to a normal distribution with
16 mean = 0 and SD = 1, the PRSs from the PsychArray and GSA could be combined
17 directly and applied easily into regression models as continuous variables. Also, the
18 standardisation of the PRS allowed the conversion of an individual's PRS to quantiles
19 for risk comparison across individuals.

20

21 **2.3.5 Statistical analysis**

22 CLEQ and PRS data was available for 885 BD subjects, 640 from the PsychArray

1 and 245 from the GSA. The participants were first divided into two groups based on the
2 presence of ACE and then compared for the difference in characteristics. Wilcoxon rank
3 sum tests, chi-squared tests and independent t-tests were applied according to the
4 variable type and distribution. To illustrate the association between ACEs and multi-
5 PRSs, we first conducted pairwise comparisons using simple t-tests taking ACE as a
6 categorical variable. Then we conducted linear regressions using ACE total number and
7 multi-PRSs taking appropriate adjustments.

8 Next, we performed multiple linear and logistic regression analyses to assess how
9 ACE, multi-PRSs and their interactions predict the development of selected BD sub-
10 phenotypes. The assumptions for linear and logistic regressions were pre-checked and
11 found to be satisfactory for each regression. Then the CLEQ total score, one PRS, or
12 their interaction term, was each added into the model by sequence. Finally, we divided
13 the samples into four quantile groups based on their BD/ADHD PRS values respectively.
14 We conducted additional logistic regressions taking the lowest PRS group as reference
15 to test if people with higher BD/ADHD PRSs would have higher odds of experiencing
16 ACE or developing the selected sub-phenotypes. The presence of ACE was added as a
17 moderator. We carried out additional interaction analyses to examine if there were
18 significant differences between each PRS quantile group and the presence of ACE, and
19 the interactions between ADHD and BD PRS were also examined.

20 The participants' BD age of onset and sex were included as covariates in all
21 analyses except for age of onset where only sex was included. The genotyping chip type
22 and the first three principal components from GWAS population stratification were

1 included in addition for all regression analyses involving PRS to account for chip and
2 ancestry confounding. To further account for genotyping difference, we conducted
3 additional sensitivity analyses using PRSs generated from only overlapping risk
4 variants from both chips (Appendix 2.3 and 2.4). All these described analyses were
5 done using RStudio with R version 4.1.3 (R Core Team, 2021).

6 To account for multiple testing, we applied False Discovery Rate (FDR) correction
7 method (Benjamini & Hochberg, 1995) to the results obtained using the `p.adjust`
8 function in R for each set of analyses. The FDR method was chosen over the others as
9 it gives a good illustration of results and has been applied in previous studies involving
10 PRS (Grigoriu-Serbanescu et al., 2020).

11

12 **2.4 Results**

13 **2.4.1 Sample demographics**

14 Overall, the participants had a median age of 49 when they received the assessment.
15 62% of the participants experienced at least one ACE before the age of 16 (see Table
16 2.1). The samples contained a high proportion of females (60%) and type 1 BD patients
17 (65%). The two groups did not differ in sex or BD type ratios. The participants also did
18 not differ in childhood ADHD (defined using the WURS) and responses to lithium.
19 However, the participants with any presence of ACE were more likely to have PPD
20 ($p=0.029$). The participants had a median BD onset age of 19. The participants' age at
21 interview and BD onset age was earlier in subjects who had experienced at least one
22 ACE (See Table 2.1). The participants also differed in the presence of suicide ideation

- 1 ($p=0.017$) and rapid cycling ($p=0.016$). We found strong evidence to suggest that the
- 2 presence of ACE was associated with an increase in ADHD PRS ($p=0.005$).

Table 2.1 Participants' Demographics and Clinical Characteristics concerning ACE Presence

Variables	N	Overall N = 885	Group without ACE N = 338 (38%)	Group with ACE N = 547 (62%)	p-values
Age at interview	728	49 (39, 59)	50 (42, 60)	48 (38, 57)	0.003^a
Sex (Females)	885	532 (60%)	199 (59%)	333 (61%)	0.600 ^b
BD type	885				0.400 ^b
BD type 1		572 (65%)	224 (66%)	348 (64%)	
BD type 2		141 (16%)	56 (17%)	85 (16%)	
Schizoaffective BD		172 (19%)	58 (17%)	114 (20%)	
Childhood ADHD	191	37.07 (22.58)	39.00 (21.03)	35.83 (23.54)	0.333 ^c
Lithium responders	418	152 (36%)	65 (38%)	87 (35%)	0.612 ^b
PPD diagnosis	824	74 (9%)	19 (2%)	55 (7%)	0.029^b
Age of BD onset	746	19 (16, 29)	22 (17, 29)	18 (15, 28)	0.009^a
Psychotic symptoms	844	598 (71%)	222 (69%)	376 (72%)	0.400 ^b
Suicide ideation	814	613 (75%)	217 (71%)	396 (78%)	0.017^b
Rapid cycling	499	215 (43%)	61 (36%)	154 (47%)	0.016^b
BD PRS	885	0.71 (1.05)	0.72 (1.01)	0.70 (1.08)	0.819 ^c
ADHD PRS	885	0.02 (1.03)	-0.10 (1.05)	0.10 (1.01)	0.005^c
MDD PRS	885	0.26 (1.00)	0.19 (0.99)	0.30 (1.00)	0.113 ^c
SCZ PRS	885	0.52 (0.97)	0.59 (0.96)	0.48 (0.97)	0.101 ^c

Notes. ACE, adverse childhood experience; childhood ADHD (scores from the Wender Utah rating scale); PPD, premorbid personality disorder; BD, bipolar disorder; ADHD, attention deficit hyperactivity disorder; MDD, major depressive disorder; SCZ, schizophrenia disorder; IQR, interquartile range; SD, standard deviation; PRS, polygenic risk score.

^a Wilcoxon rank sum test; median (IQR)

^b Pearson's Chi-squared test of independence; n (%)

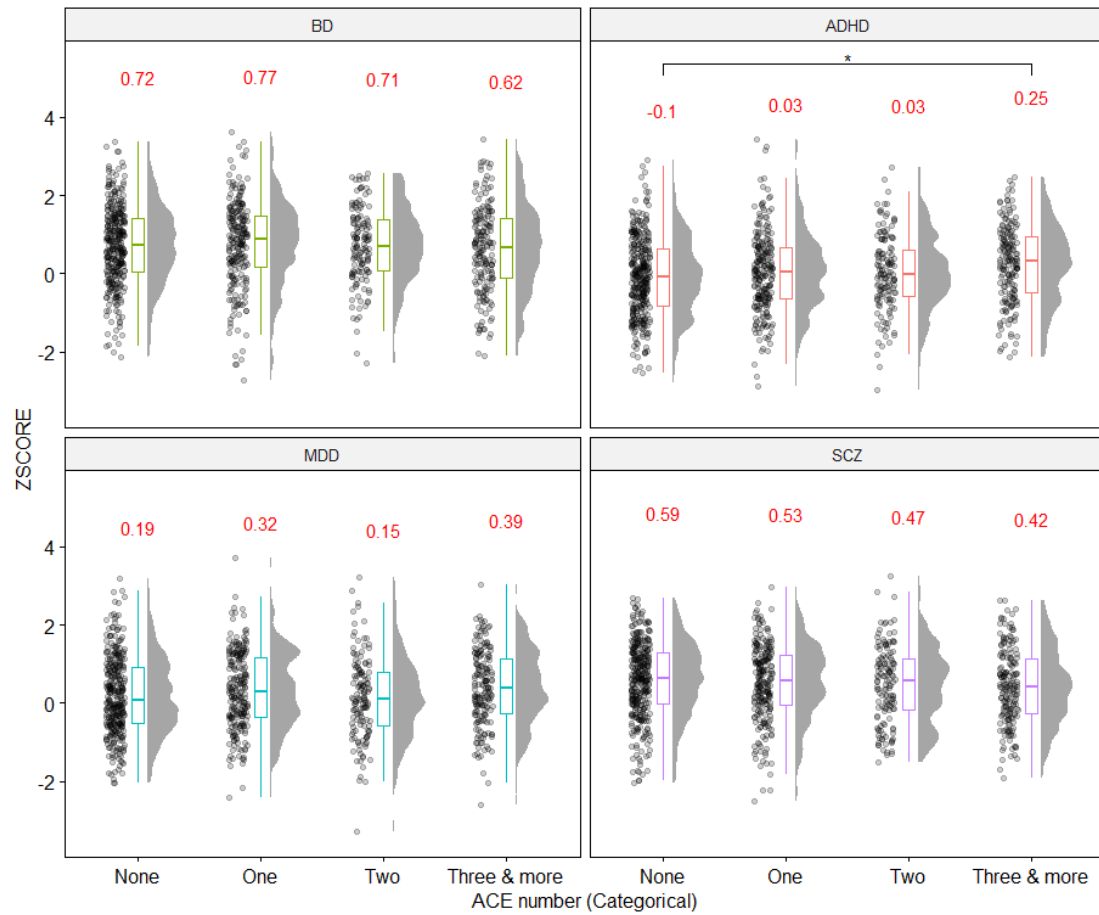
^c Independent t-test; mean (SD)

In bold p values are below 0.05 threshold.

2.4.2 Correlations between multi-PRSs and the CLEQ scores

We found no evidence supporting any associations between BD PRS and ACE scores using pairwise comparisons (see Figure 2.1) or linear regressions with (coefficient=-0.037; 95% CI: -0.146 to 0.073, $p=0.511$) or without any adjustments (coefficient=-0.051; 95% CI: -0.149 to 0.046, $p=0.300$). However, we found strong evidence suggesting that the participants with none and three or more ACE differed in ADHD PRS (see Figure 2.1). The unadjusted linear regression results also indicated that higher ADHD PRS increased the susceptibility to ACE reporting (coefficient=0.198, 95% CI: 0.099 to 0.296, $p<0.001$). The association was even stronger after adding in adjustments (coefficient=0.231, 95% CI: 0.118 to 0.345, $p<0.001$). Weak evidence was found to suggest the association between MDD PRS and ACE scores (coefficient=0.103, 95% CI: <0.001 to 0.205, $p=0.049$) the evidence remained after adding in adjustments (coefficient=0.127; 95% CI: 0.012 to 0.243, $p=0.031$). No evidence could be found for SCZ PRS before (coefficient=-0.099, 95% CI: -0.204 to 0.006, $p=0.066$) or after adjustment (coefficient=-0.043, 95% CI: -0.161 to 0.074, $p=0.470$). And no other pair-wise comparison results survived correction for multiple testing (see Figure 2.1).

Figure 2.1 Mean Polygenic Risk Scores for ADHD, BD, MDD, and SCZ across ACE Groups



Notes. ACE, adverse childhood experience; BD, bipolar disorder; ADHD, attention deficit hyperactivity disorder; MDD, major depressive disorder; SCZ, schizophrenia disorder.

The above numbers corresponded to PRS means for each ACE group.

* in the current plot, p values survived FDR multiple testing correction for simple t-tests for pair-wise comparison.

2.4.3 ACE/BD PRS associations & interaction with phenotypes

We found substantial evidence to suggest that each unit increase in BD PRS would increase the odds of having psychotic symptoms by $\exp(0.258) = 1.294$ (95% CI 1.093 to 1.538; $FDR_p=0.035$; See Table 2.2). We also found strong evidence that each unit increase in ACE number would increase the odds of having rapid cycling by $\exp(0.228) = 1.256$ (95% CI 1.084 to 1.463; $FDR_p=0.035$). We found some tendency that ACE might be associated with an earlier age of onset. We carried out additional subgroup analyses to clarify the effect. We found strong evidence to suggest that only males who experienced ACE would have earlier age of onset (coefficient -1.568, 95% CI: -2.568 to -0.567, $p=0.002$).

Overall, we did not observe any interaction effect between ACE number and BD PRS in predicting any of the sub-phenotypes examined. (See Table 2.2).

2.4.4 ADHD/MDD/SCZ PRS associations & interaction with phenotypes

We found strong evidence to suggest that each unit increase in ADHD PRS led to an increase in the odds of developing rapid cycling BD by $\exp(0.369) = 1.45$ (95% CI 1.20–1.75; $FDR_p=0.002$). No other symptoms were associated with the ADHD PRS and no significant interaction effect between ACE number and ADHD PRS in predicting these sub-phenotypes was observed (see Table 2.2).

We could not find any evidence to suggest MDD and SCZ PRS would predict any of these selected sub-phenotypes (see Table 2.3). And we observed no interaction effect.

Table 2.2 Results of Multiple Regression Analyses with Adjustments based on ACE (total score) and BD/ADHD PRS

Variables	Estimated Coefficient	Standard Error	Confidence Intervals (95%)	Reference Values	<i>p</i>	<i>FDR_p</i>
Age at onset^a						
ACE	-0.835	0.330	-1.482, -0.188	-2.543	0.012	0.104
BD PRS	-0.315	0.466	-1.230, 0.600	-0.677	0.499	0.877
BD Interaction	-0.316	0.328	-0.960, 0.328	-0.963	0.336	0.765
ADHD PRS	-0.624	0.491	-1.588, 0.339	-1.273	0.204	0.717
ADHD Interaction	-0.421	0.341	-1.090, 0.248	-1.236	0.217	0.717
Presence of psychotic symptoms^b						
ACE	0.069	0.063	-0.052, 0.195	1.091	0.275	0.761
BD PRS	0.258	0.087	0.089, 0.430	2.974	0.003	0.035
BD Interaction	-0.074	0.063	-0.197, 0.051	-1.177	0.239	0.718
ADHD PRS	-0.043	0.090	-0.221, 0.134	-0.478	0.633	0.878
ADHD Interaction	0.005	0.065	-0.122, 0.133	0.074	0.941	0.991
Presence of suicide ideation^b						
ACE	0.178	0.074	0.037, 0.328	2.398	0.016	0.119
BD PRS	-0.048	0.093	-0.231, 0.134	-0.514	0.607	0.878
BD Interaction	-0.088	0.072	-0.229, 0.053	-1.229	0.219	0.717
ADHD PRS	0.055	0.098	-0.137, 0.248	0.562	0.574	0.878
ADHD Interaction	-0.007	0.077	-0.156, 0.146	-0.095	0.924	0.991
Presence of rapid cycling^b						
ACE	0.228	0.076	0.081, 0.381	2.992	0.003	0.035
BD PRS	-0.106	0.103	-0.310, 0.095	-1.032	0.302	0.765
BD Interaction	-0.041	0.085	-0.209, 0.125	-0.479	0.632	0.878
ADHD PRS	0.495	0.121	0.262, 0.738	4.080	<0.001	0.002
ADHD Interaction	0.144	0.090	-0.30, 0.324	1.600	0.109	0.563

Notes. ACE, adverse childhood experience; PRS, polygenic risk score; BD, bipolar disorder; ADHD, attention deficit hyperactivity disorder.

^a Multiple linear regression analyses, reference value t.

^b Multiple logistic regression analyses, reference value z.

FDR_p = false discovery rate corrected p values for multiple testing.

In bold p values were significant before correction or survived FDR correction for multiple testing.

All results were adjusted for participants' BD age of onset and sex (except for age of onset where only sex was included).

PRS & interaction results were adjusted for chip type and the first three principal components from GWAS population stratification in addition to sex and age of onset.

Table 2.3 Results of Multiple Regression Analyses with Adjustments based on Total ACE Number and MDD/SCZ PRS

Variables	Estimated Coefficient	Standard Error	Confidence Intervals (95%)	Reference Values	<i>p</i>	<i>FDR_p</i>
Age at onset^a						
MDD PRS	-0.472	0.494	-1.442, 0.498	-0.956	0.340	0.765
MDD Interaction	-0.070	0.331	-0.719, 0.579	-0.212	0.832	0.991
SCZ PRS	0.021	0.501	-0.961, 1.004	0.043	0.966	0.991
SCZ Interaction	-0.299	0.358	-1.001, 0.403	-0.837	0.403	0.853
Presence of psychotic symptoms^b						
MDD PRS	-0.052	0.091	-0.232, 0.127	-0.574	0.566	0.878
MDD Interaction	-0.001	0.064	-0.128, 0.123	-0.017	0.987	0.991
SCZ PRS	0.118	0.092	-0.063, 0.299	1.280	0.201	0.717
SCZ Interaction	-0.134	0.069	-0.251, 0.021	-1.643	0.100	0.563
Presence of suicide ideation^b						
MDD PRS	-0.041	0.098	-0.233, 0.151	-0.418	0.676	0.888
MDD Interaction	-0.002	0.074	-0.149, 0.143	-0.028	0.978	0.991
SCZ PRS	-0.001	0.100	-0.199, 0.195	-0.011	0.991	0.991
SCZ Interaction	0.021	0.086	-0.148, 0.190	0.241	0.810	0.991
Presence of rapid cycling^b						
MDD PRS	0.086	0.111	-0.131, 0.304	0.776	0.438	0.876
MDD Interaction	0.032	0.081	-0.128, 0.193	0.397	0.691	0.888
SCZ PRS	0.052	0.109	-0.161, 0.267	0.477	0.634	0.878
SCZ Interaction	0.052	0.080	-0.105, 0.212	0.641	0.521	0.878

Notes. ACE, adverse childhood experience; MDD, major depressive disorder; SCZ, schizophrenia disorder

^a Multiple linear regression analysis, reference value t.

^b Multiple logistic regression analysis, reference value z.

FDR_p = false discovery rate corrected p values for multiple testing.

All results were adjusted for participants' BD age of onset and sex (except for age of onset where only sex was adjusted). PRS & interaction results were adjusted for chip type and the first three principal components from GWAS population stratification in addition to sex and age of onset.

2.4.5 PRS quantile analyses

Given that no effect could be observed from MDD/SCZ PRS, participants were only divided into quantile groups based on their BD/ADHD PRSs for ACE influence (see Table 2.4). Overall, we found strong evidence to suggest that BD patients in the highest ADHD PRS quantile group had 2.009 higher odds of having experienced ACE than those in the lowest ADHD PRS quantile group (95% CI 1.230 to 3.313, $p=0.006$, $FDR_p=0.046$; Figure 2.2A).

BD patients in the highest BD PRS quantile group had 1.917 higher odds of developing psychotic symptoms than those in the lowest BD PRS quantile group (95% CI 1.166 to 3.182, $p=0.011$, $FDR_p=0.234$; Figure 2.2B). Meanwhile, BD patients in the highest ADHD PRS quantile group had 2.642 higher odds of developing rapid cycling than those in the lowest ADHD PRS quantile group (95% CI 1.393 to 5.097, $p=0.003$, $FDR_p=0.026$; Figure 2D).

Table 2.4 Results of PRS Quantile Group Comparisons for the Odds Plot

Variables	Odds	Confidence Intervals (95%)	Reference Values	<i>p</i>	<i>FDR_p</i>
Presence of ACE					
BD Q1 vs Q2	0.889	0.558, 1.416	-0.495	0.621	1.000
BD Q1 vs Q3	1.106	0.696, 1.758	0.425	0.671	1.000
BD Q1 vs Q4	1.140	0.714, 1.821	0.548	0.584	1.000
ADHD Q1 vs Q2	0.983	0.627, 1.540	-0.073	0.942	1.000
ADHD Q1 vs Q3	1.365	0.854, 2.186	1.299	0.194	0.775
ADHD Q1 vs Q4	2.009	1.230, 3.313	2.764	0.006	0.046
Presence Psychotic Symptoms					
BD Q1 vs Q2	1.567	0.964, 2.562	1.803	0.071	0.209
BD Q1 vs Q3	1.546	0.953, 2.519	1.759	0.079	0.209
BD Q1 vs Q4	1.917	1.166, 3.182	2.547	0.011	0.087
ADHD Q1 vs Q2	0.922	0.560, 1.513	-0.321	0.748	1.000
ADHD Q1 vs Q3	0.696	0.421, 1.143	-1.428	0.153	0.307
ADHD Q1 vs Q4	0.925	0.547, 1.568	-0.289	0.772	1.000
Presence of suicide ideation					
BD Q1 vs Q2	0.949	0.548, 1.642	-0.187	0.852	1.000
BD Q1 vs Q3	1.008	0.583, 1.743	0.029	0.977	1.000
BD Q1 vs Q4	0.800	0.467, 1.366	-0.815	0.415	1.000
ADHD Q1 vs Q2	1.141	0.683, 1.906	0.504	0.615	1.000
ADHD Q1 vs Q3	1.054	0.618, 1.800	0.194	0.846	1.000
ADHD Q1 vs Q4	1.356	0.772, 2.409	1.052	0.293	1.000
Presence of rapid cycling					
BD Q1 vs Q2	0.785	0.427, 1.434	-0.787	0.431	0.575
BD Q1 vs Q3	0.576	0.313, 1.051	-1.789	0.074	0.196
BD Q1 vs Q4	0.727	0.397, 1.323	-1.042	0.297	0.476
ADHD Q1 vs Q2	1.702	0.901, 3.252	1.629	0.103	0.207
ADHD Q1 vs Q3	2.059	1.105, 3.893	2.254	0.024	0.097
ADHD Q1 vs Q4	2.642	1.393, 5.097	2.943	0.003	0.026

Notes. ACE, adverse childhood experience; PRS, polygenic risk score; BD, bipolar disorder; ADHD, attention deficit hyperactivity disorder.

Results came from multiple logistic regression analysis, reference value z.

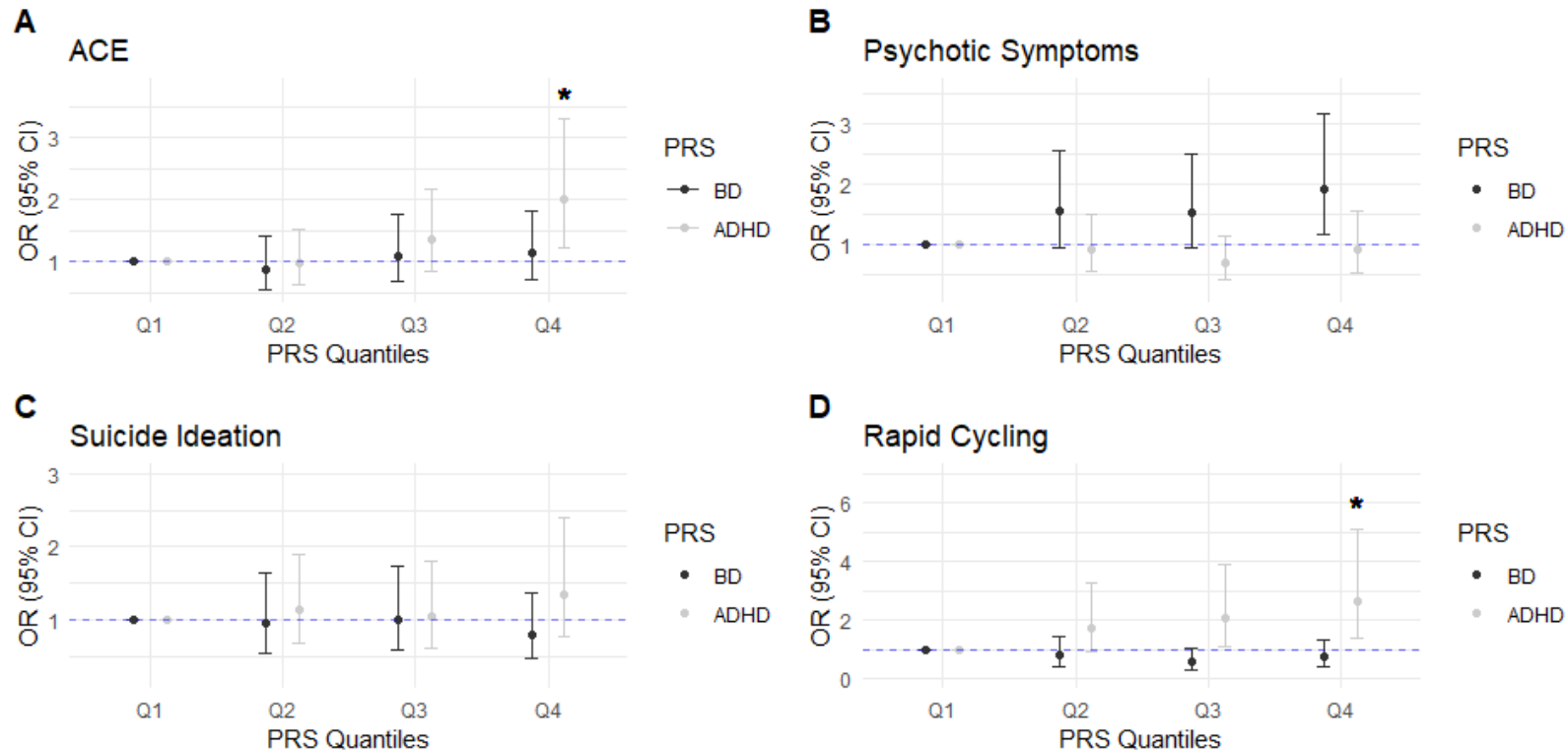
FDR_p = false discovery rate corrected p values for multiple testing.

All results were adjusted for participants' BD age of onset and sex, chip type and the first three principal components from GWAS population stratification.

The presence of ACE was included as a moderator in the PRS models for the three sub-phenotypes.

In bold p values were significant before correction or survived FDR correction for multiple testing.

Figure 2.2 The Odds Ratios of having ACE and Developing the Three Selected Sub-phenotypes with Reference to PRS Quantile Levels



Notes. OR odds ratio, CI confidence intervals, PRS polygenic risk scores

Q1, 0 - 25%; Q2, 25% - 50%; Q3, 50% - 75%; Q4 75% - 100%

The participants' sex, age of onset, the genotyping chip type and the first three principal components from GWAS population stratification were included as covariates in all analyses.

The presence of ACE was included as a moderator in the PRS models for the three sub-phenotypes.

* in the current plot, p values survived FDR multiple testing correctio

2.4.6 PRS quantile interaction analyses results

To further dissect how the presence of ACE can influence the genetic impact and if the two PRSs' associations depend on each other, we carried out additional interaction analyses (see Table 2.5). We only found weak evidence to suggest a positive interaction effect between BD and ADHD PRSs on the presence of ACE (estimated coefficient: 0.158, 95% CI: 0.018, 0.301, $p=0.028$, $FDR_p=0.278$). The positive result suggested that participant who had both high BD and ADHD PRSs are more likely to have experienced ACE than those who had both low BD and ADHD PRSs. We found no other tendency for an interaction between the two PRSs. However, such results should be interpreted with cautions because the participants' ADHD and BD PRS are highly correlated (estimated coefficient: 0.113, 95% CI: 0.047 to 0.178, $p<0.001$). The Variance Inflation Factor (VIF) values for the two PRS interaction variables in the models ranged from around 12 to 13 indicating potential collinearity. Further examinations in future studies are required. The other interaction analyses taking PRS quantile groups and ACE presence showed consistent results to the main analyses as no significant interaction effects could be detected.

Table 2.5 Results of Additional Interaction Analyses based on ACE Presence and BD/ADHD PRS Quantile Groups

Variables	Estimated Coefficient	Standard Error	Confidence Intervals (95%)	Reference Values	<i>p</i>	<i>FDR_p</i>
Presence of ACE						
BD ADHD interaction	0.158	0.072	0.018, 0.301	2.200	0.028	0.278
Presence Psychotic Symptoms						
BD ACE interaction	-0.185	0.169	-0.519, 0.144	-1.099	0.272	0.679
ADHD ACE interaction	0.059	0.171	-0.277, 0.395	0.346	0.729	0.886
BD ADHD interaction	0.011	0.076	-0.138, 0.160	0.144	0.886	0.886
Presence of suicide ideation						
BD ACE interaction	-0.296	0.180	-0.651, 0.056	-1.643	0.100	0.335
ADHD ACE interaction	-0.110	0.186	-0.478, 0.253	-0.589	0.556	0.794
BD ADHD interaction	0.020	0.082	-0.141, 0.181	0.243	0.808	0.886
Presence of rapid cycling						
BD ACE interaction	0.178	0.211	-0.234, 0.595	0.843	0.399	0.736
ADHD ACE interaction	0.384	0.220	-0.047, 0.818	1.745	0.081	0.335
BD ADHD interaction	0.072	0.094	-0.111, 0.257	0.769	0.442	0.736

Notes. ACE, adverse childhood experience; PRS, polygenic risk score; BD, bipolar disorder; ADHD, attention deficit hyperactivity disorder.

Results came from multiple logistic regression analysis, reference value z.

FDR_p = false discovery rate corrected p values for multiple testing.

All results were adjusted for participants' BD age of onset and sex, chip type and the first three principal components from GWAS population stratification.

The presence of ACE was included as a moderator in the two PRS interaction models for the three sub-phenotypes.

In bold p values were significant before correction.

2.5 Discussion

This study used multiple PRSs to dissect their association and interaction with ACE for predicting BD sub-phenotypes. Our study demonstrated that BD participants with higher ADHD PRS tended to report more ACEs, whereas BD PRS was not associated with ACEs reporting. We also found some weak evidence to suggest that BD patients with higher MDD may experience more ACE. In addition, our results suggested that the BD patients with higher BD PRS would have increased odds of developing psychotic symptoms. Although both increased ACE number and ADHD PRS were associated with increased odds of developing rapid cycling among BD patients, no additive interaction effect could be observed.

Past studies had conflicts regarding whether the increase in ACE numbers is associated with BD PRS positively (Park et al., 2020) or negatively (Aas et al., 2020). Our results still could not provide a specific answer since we could not identify any significant results between BD PRS and the reported number of ACE across multiple analyses. However, our results suggested that increased ADHD PRS was associated with more ACE reporting. These results are consistent with National Survey of Children's Health studies which observed that children with ADHD symptoms may more likely experience ACE (Brown et al., 2017; Crouch et al., 2021). We also found weak evidence for the positive association between MDD PRS and ACE. Thus, it seems more likely that BD patients' genetic liability to other psychiatric disorders together accounted for the presence of ACE. However, it should not be assumed that the exposure to ACE is entirely determined by genetic risk, given that most recorded ACE

1 were passive. In this case, a possible explanation could be that these participants'
2 parents not only transmitted genetic risk variants linked to psychiatric disorders to their
3 child but also provided adverse environments (Baldwin et al., 2022). In addition,
4 genetically influenced behaviours in children (e.g., impulsivity, emotional
5 dysregulation) may also evoke harsher parenting or peer rejection (Reiss et al., 2022).

6 Most previous research has shown that an increasing number of ACEs is associated
7 with a lower age of BD onset (Aas et al., 2020; Anand et al., 2015; Park et al., 2020).

8 Our study also identified the same pattern in results for ACE presence. Only the study
9 by Park et al. (2020) found an additive interaction effect between BD PRS and ACE for
10 predicting earlier BD age of onset. Our results and the other research did not identify
11 such an interaction pattern (Aas et al., 2020). It should be noted that the study by Park
12 et al. (2020) was purely based on BD type 1 patients. Our samples contained 65% BD
13 type 1 patients, and the study by Aas et al. (2020) contained 74% BD type 1 patients.

14 Thus, differences between study samples in the genetic liability to other psychiatric
15 disorders may have influenced these results (Guzman-Parra et al., 2021). In addition,
16 although another study by Anand et al. (2015) included 1995 BD type 1 patients, they
17 still could not identify significant interaction between any single SNP and the presence
18 of childhood traumatic events on the prediction of BD age at onset. However, they
19 found that only SNPs in or near genes coding for calcium channel activity-related
20 proteins (Gene Ontology: 0005262) were more likely to show an interaction effect.

21 Thus, if the gene x environment interaction exists in predicting BD age of onset, it might
22 be small and require large sample sizes to detect. Given that the study by Park et al.

1 (2020) contained 1615 BD cases, much larger than our sample size of 885 and Aas et
2 al.'s (2020) sample size of 402. The sample sizes might explain the difference in the
3 results. Another possible explanation for the difference on age of onset could be sex
4 differences. Our additional sub-group analyses found that only males who experienced
5 ACE would have earlier age of onset.

6 Our results on psychosis symptoms were consistent with Aas et al. (2020), which
7 similarly recorded the psychosis variable in terms of the episode. Our results showed
8 that people with a higher BD PRS may be more likely to show psychotic symptoms.
9 One recent study also found that BD PRS might relate to the manic symptoms in
10 participants with a history of psychotic episodes (Ahangari et al., 2022). However, we
11 could not identify any significant association between ACE and psychosis symptoms
12 or evidence for an interaction effect. Upthegrove et al. (2015) argued that the different
13 ACE types might influence the development of psychosis symptoms differently.
14 Childhood abuse had the strongest associations. Although Aas et al.'s (2020) study
15 focused on CM, covering all abuse items, the associations and interaction effects were
16 still insignificant. Thus, it seemed likely that these psychotic symptoms in BD subjects
17 might be more genetically predetermined.

18 We found that the ADHD PRS in BD subjects may be highly associated with the
19 presence rapid cycling. However, we should be careful when interpreting the findings
20 on rapid cycling. In contrast to Aas et al. (2020), we could not find an interaction
21 between BD PRS and ACE or between ADHD PRS and ACE on rapid cycling. Our
22 results showed that the increase in ACE number and ADHD PRS might independently

1 increase the odds of rapid cycling. And they also positively associated with each other.
2 Such results may indicate the presence of mediation effects which requires further
3 examination.

4 In addition, we could not find any prediction effects from MDD or SCZ PRS. The
5 previous studies which found these two PRSs' prediction effects were based on samples
6 which were of the same category MDD and SCZ patients (K. G. Jonas et al., 2019;
7 Thorp et al., 2023). Although these two disorders were also genetically correlated with
8 BD, the current results suggest that they are underpowered to predict the development
9 of any BD sub-phenotype symptoms.

10 Overall, we could not identify any significant interaction effect in any models. We
11 anticipated that cases with more ACE and higher PRSs would develop more severe
12 phenotypes. However, our results suggested that different levels of PRS might not
13 significantly influence the relationship between ACE and other BD phenotypes.
14 Meanwhile, the association between the PRS and sub-phenotypes did not significantly
15 differ according to the number of ACE participants experienced. Such conflicts might
16 be because past studies that found the interaction effects included more severe forms of
17 ACE, such as CM, covering abuse and trauma (Aas et al., 2020; Park et al., 2020).
18 However, the ACE measurement in our study only included ACE without any traumatic
19 events such as abuse and neglect. Thus, the interaction effects may have been attenuated
20 in our study. However, our results also showed evidence that even ACE without any
21 CM items could be significant predictors for predicting different sub-phenotypes.

1 Overall, such results highlight the importance of considering different ACE definitions
2 for future interaction studies.

3 Our study first used the more advanced PRS-CS-auto method and the latest
4 reference GWAS to calculate multi-PRSs. Thus, a straightforward explanation for the
5 difference between our results and those from past studies might be that the multi-PRSs
6 used here were more powerful predictors of a subject's genetic liability to psychiatric
7 disorders. We also used robust correction method for selecting results from the
8 statistical analyses. Concerning the link between multi-PRSs and ACEs, we replicated
9 past studies' analyses and ran additional tests. By focusing on ACE items without any
10 CM items, our results consolidated ACE's potential in predicting different phenotype
11 developments. Such results also highlighted that future studies should consider the ACE
12 definition when conducting gene-environment interaction studies. In addition, our
13 findings confirmed the importance of considering multiple psychiatric PRSs in BD
14 subjects and their interactions with ACE for improving phenotype predictions. Future
15 larger-scale studies with more precise separation of ACE types, more psychiatric PRSs
16 and more of their interactions are encouraged.

17 This study's findings could be limited by its case-control recruitment and cross-
18 sectional data collection methods. Retrospective data such as analysed in this study is
19 always subject to potential recall bias. The participants had a median age of 49 at the
20 interview assessment, but the CLEQ required them to recall life events before age 16.
21 All data was collected directly from the participants. If the BD patients developed
22 delusions or hallucinations regarding their childhood experiences, it would be

1 impossible for us to tell. Also, our study did not separate ACE severity or phenotype
2 severity. We focused on only the ACE without any CM items. Thus, we could only
3 make inferences but not compare ACE and CM items' different effectiveness. In
4 addition, our findings were limited by our sample size and sample characteristics
5 (mixed BD types). Future studies can use mixed models that deal better with missing
6 data, characteristic differences, unequal sample sizes, and non-independence of samples.
7 Given that all samples are Europeans, our findings might also have limited
8 transferability. Finally, neither the PRS nor ACEs could explain a large amount of
9 individual variation. Even though we found some links between PRS and phenotypes
10 that might be important in the clinic, these results were still a long way from letting us
11 predict how an individual's phenotype will develop.

12 BD is a complex and heterogeneous disorder. Both genetic and environmental risk
13 factors influence its phenotypic development. In this context, future larger-scale studies
14 with more precise separation of ACE types, more psychiatric PRSs, and more of their
15 interactions can better illustrate the phenotype development predictions among BD
16 patients. Together, these efforts will support better BD prognosis, risk prediction,
17 treatment allocation, and harm prevention.

3. Implication of the ADCY1 Gene in Lithium Response in Bipolar Disorder by Genome-wide Association Meta-analysis

A version of this paper has been submitted as preprint at Research Square.

3.1 Abstract

Background: Efforts to examine the influence of genetics in the efficacy of lithium using GWAS have identified several loci. Here, we report new cases and meta-analyses to combine previous efforts.

Method: We report data from 1259 participants with BD recruited at University College London who had been treated with lithium. The data comes from three waves of genotyping on different arrays. The GWAS data from each array was analysed separately and then meta-analysed with two published lithium response GWAS datasets. Post-GWAS analyses were conducted to examine lithium response heritability and its genetic correlations with other traits. We also attempted to replicate past polygenic risk scores (PRS) results.

Results: SNP rs116927879 (A/G) was associated with lithium response at a genome-wide significance level ($p=4.509\times 10^{-08}$) with a consistent effect across all cohorts. rs116927879 is located on chromosome 7 and maps to the protein-coding gene *ADCY1* and two pseudo-genes, *GTF2IP13* and *SEPT7P2*. The sQTL results suggested that rs116927879 genotypes may influence the splicing of *ADCY1* across different brain regions. We estimated the SNP heritability (h^2) for good lithium response as 20.3% and 15.6% for subjective/objective response definitions, respectively. We did not observe any genetic correlation or PRS association between the lithium response and

1 schizophrenia or major depression disorder. However, we found weak evidence to
2 suggest that males were more likely to be good responders.

3 *Conclusions:* Our GWAS identifies a genome wide significant finding, and provides
4 updated heritability estimates for lithium efficacy.

5

6 **3.2 Introduction**

7 Lithium is a first-line treatment option for BD and has been recommended by
8 multiple clinical guidelines (UK (NICE, 2014a); Australia & New Zealand (Malhi et
9 al., 2015); Canada (Yatham et al., 2018)). Lithium has been found to be the most
10 effective mood stabilizer (Joas et al., 2017), not only in controlling mania and
11 hypomania symptoms (Cipriani et al., 2011; Yildiz et al., 2011) and reducing depression
12 severity (J. C. Nelson et al., 2014), but also in preventing suicides and suicide attempts
13 among BD patients (Baldessarini & Tondo, 2022; Song, Sjölander, et al., 2017).
14 However, it has been estimated that only around 30% of patients are excellent
15 responders while around 40% of patients do not receive any benefits (Rybakowski et
16 al., 2001). Meanwhile, lithium can cause adverse effects on the kidneys, thyroid, and
17 parathyroid glands (Gitlin, 2016). Exploring the genetic variability underlying
18 heterogenous lithium response may help clinicians improve future treatment plans.

19 To date, GWAS of lithium response have identified several different loci. The
20 earliest lithium response GWAS was reported in 2009 (Perlis et al., 2009). The study
21 included samples from the US Systematic Treatment Enhancement Program for Bipolar
22 Disorder (STEP-BD) cohort and a subset of the University College London (UCL)

1 samples. Although no SNPs reached genome-wide significance, the findings suggested
2 that good lithium response was associated with the *GRIA2* gene which encodes
3 glutamate ionotropic receptor AMPA type subunit 2 (Perlis et al., 2009). Subsequently,
4 a GWAS performed in a Han Chinese sample found that another protein-coding gene
5 *GADLI* (glutamate decarboxylase-like protein 1) was highly associated with lithium
6 response (C.-H. Chen et al., 2014). The top two SNPs (rs17026688 and rs17026651) in
7 their combined series, had p values of 1.66×10^{-49} and 7.07×10^{-50} , respectively.
8 However, these findings could not be replicated in subjects of European ancestry
9 (Cruceanu et al., 2015), or in other Asian samples (Birnbaum, 2014; Kotambail et al.,
10 2015).

11 In 2016, Song and colleagues (Song, Bergen, et al., 2017; Song et al., 2016) reported
12 a GWAS meta-analysis of 3874 BD lithium users from Sweden and the UK. The
13 researchers analysed the lithium response separately according to objective and
14 subjective definitions. While 323 participants were objectively good responders using
15 the clinical definition, 1639 were self-reported good lithium responders. The
16 researchers conducted four sets of GWAS: lithium responders compared with non-
17 responders and lithium responders compared with healthy controls using both objective
18 and subjective measures. Overall, one SNP (rs146727601) reached genome-wide
19 significance from the GWAS of objective measured responders versus controls (Song,
20 Bergen, et al., 2017). rs146727601 is a two-base deletion on chromosome 11q22.4 in
21 the gene PTS (6-pyruvoyltetrahydropterin synthase) which is a catalyst involved in the
22 regulation of serotonin biosynthesis and nitric oxide synthase activity.

1 In 2016, Hou et al. (2016) also reported a GWAS for lithium response among BD-I
2 and II participants. The data included 2343 European BD participants treated with
3 lithium. The data was collected from 22 sites as part of the International Consortium on
4 Lithium Genetics (ConLi⁺Gen). Lithium response was rated on the Retrospective
5 Assessment of the Lithium Response Phenotype Scale (Alda scale) which is a well-
6 established measurement scale for lithium response (Scott et al., 2020). In the
7 consortium study, the researchers reported that four linked SNPs on chromosome 21
8 were associated with good lithium response (rs79663003; rs78015114; rs74795342;
9 and rs75222709) at a genome-wide level of significance. The chromosomal locus of the
10 four SNPs contains genes for two long, non-coding RNAs (lncRNA), AL157359.3 and
11 AL157359.4. Subsequent analyses of the ConLi⁺Gen data provided insights into the
12 potential genetic architecture of good lithium response. ConLi+Gen (2018) and Amare
13 et al. (2021) reported that higher schizophrenia (SCZ) and major depression disorder
14 (MDD) polygenic risk scores (PRS) were both associated with reduced lithium response
15 in BD participants. These findings were further consolidated by Schubert et al. (2021),
16 who demonstrated that a combination of the SCZ and the MDD PRSs provided
17 improved prediction of lithium treatment response, whereas BD PRS had no impact on
18 the prediction.

19 Overall, prior findings from GWAS have highlighted different loci for the
20 mechanistic action of lithium. This may in part be due to different measures of lithium
21 response and small sample sizes. In addition, past PRS analyses were only calculated
22 using samples from ConLi⁺Gen. The aim of the present study is to report a much-

1 enlarged University College London (UCL) sample of participants with BD taking
2 lithium and conduct GWAS meta-analyses with other compatible European samples.
3 We meta-analysed our GWAS results with the summary statistics from Song et al. (Song,
4 Bergen, et al., 2017; Song et al., 2016) and Hou et al. (2016), the two largest sets of
5 GWAS on the European population to date. We estimated SNP heritability for good
6 lithium response and we conducted exploratory analyses based on the GWAS meta-
7 analysis outputs as follow-up. Finally, SCZ and MDD PRSs were calculated in the
8 samples to replicate past findings.

9

10 **3.3 Methods**

11 **3.3.1 Participants**

12 All UCL participants had received an ICD10 diagnosis of BD (World Health
13 Organization, 1992) from a UK National Health Service (NHS) psychiatrist. A total of
14 1259 BD participants had a recorded lithium response. Ancestrally matched healthy
15 control subjects (n=1782) were recruited from the NHS blood transfusion service and
16 from study sites where case participants were also being recruited. 1323 controls were
17 screened for an absence of a lifetime history of the following disorders: schizophrenia
18 and any other psychosis, major affective or schizoaffective disorders, eating disorders,
19 alcohol/drug addiction, and obsessive-compulsive disorders. 459 unscreened controls
20 were recruited from the same sites. All participants were of English, Scottish, Welsh, or
21 Irish descent and had at least three out of four grandparents of the same descent. The
22 study was approved by the NHS Metropolitan Multi-centre Research Ethics Committee

(MREC/03/11/090). All participants read an approved information sheet and signed a physical informed consent form.

3.3.2 Study measures

The SADS-L (Spitzer et al., 1978) and the 90-item OPCRIT (McGuffin et al., 1991) were administered to all participants with BD by a psychiatrist or trained researcher. The participants' diagnosis with PPD was assessed with OPCRIT item 11. The assessment of lithium treatment response was determined by the interviewer based on the rate and severity of BD episodes before and after the treatment using both face-to-face and case note-derived information. Researchers were trained in the application of all the assessment measures. The rating contained a mixture of the interviewer's judgements and the participants' self-report. The lithium response was first rated on a 5-point Likert scale ranging from very good to very poor response (see Appendix 3.1). Then only subjects reporting a "very good" or "good response" were coded as "good responders" while subjects reporting "moderate response", "poor response", or "very poor response" were coded as non-responders. We chose the dichotomous conversion to differentiate excellent responders and to be consistent with the definitions used in previous GWAS of lithium response. For instance, ConLi⁺Gen participants were classified as "responders" if had Part A score of 7 or higher (total score 10), with moderate responders excluded. These data were reviewed by a research psychiatrist (N.B.).

3.3.3 Genotyping, imputation, quality control and meta-analyses

Genome-wide single nucleotide polymorphism data was generated in three waves at the Broad Institute, Boston, MA, USA, using the Affymetrix 500K Array (Affymetrix Inc., Santa Clara, CA, US), the Illumina PsychArray (Illumina Inc., San Diego, CA, US), and the Illumina Global Screening Array (GSA; Illumina Inc.). The three waves of data underwent equivalent quality control and imputation methods which, have been described in detail elsewhere (Grigoriu-Serbanescu et al., 2020).

We first performed three separate GWAS with imputed SNP genotypes, from each wave of genotyping using PLINK v2.00a2LM (Chang et al., 2015), with the dichotomous lithium response (responders vs non-responders) as the case-control phenotype. The first three principal components of population structure were included as covariates to control for population stratification. Sensitivity analysis including sex as a covariate was conducted, we only observed slight attenuation towards the null. Thus, to be consistent with previous studies and to avoid potential heterogeneity between studies that were to be meta-analysed, age and sex were not included as covariates in the primary analyses.

3.3.4 Lithium definition heterogeneity testing

Song et al. (Song, Bergen, et al., 2017; Song et al., 2016) reported GWAS results from their objective and subjective binary lithium response definitions. We chose to use data from Song et al.'s subjective definition GWAS because this most closely matched the definition used in the UCL sample and because this measure had the largest overall

1 sample size.

2 Data from a dichotomous coded lithium response ConLi⁺Gen European GWAS
3 (GCST012486) published by Hou et al. (2016) was downloaded from the NHGRI-EBI
4 GWAS Catalog (Sollis et al., 2023) on 24/10/2023. Hou et al.'s lithium definition came
5 from the Alda scale for the long-term treatment response to lithium (Hou et al., 2016).
6 Thus, potential heterogeneity within the response definition existed.

7 We conducted binomial sign tests to compare the direction of SNP associations at
8 three different p-value thresholds (P_T ; 1×10^{-3} , 1×10^{-4} ; and 1×10^{-5}) with GWAS summary
9 statistics from the Hou et al. study and from the UCL data using the Song et al.'s
10 subjective definition GWAS as the reference study. We expected that there would be
11 strong convergence of SNP associations between the datasets.

12 LD Score Regression (LDSC; Bulik-Sullivan et al., 2015) was used to further
13 explore the genetic architecture of lithium response and to investigate the effect of
14 differences in lithium response definition. The LDSC method requires large sample
15 sizes, and the algorithm did not converge when all three data sets were analysed
16 separately. We therefore performed LDSC analyses comparing data from Hou et al.,
17 with the data from a meta-analysis of data from the UCL study and the data from Song
18 et al.'s subjective GWAS (UCL+Song^{sub}). The meta-analysis was performed using
19 METAL (Willer et al., 2010). We also examined genetic correlation between the
20 UCL+Song^{sub} and the Hou et al data with BD GWAS summary statistics from the PGC
21 BD working group (Mullins et al., 2021). Next, we used Genomic Structural equation
22 modelling (GSEM; Grotzinger et al., 2019) to fit a one-factor model to investigate

1 factor loadings for the UCL+Song^{sub} data and the Hou et al data on the PGC BD
2 summary statistics.

3 **3.3.5 GWAS meta-analysis & follow-on analyses**

4 After we found no evidence for definition heterogeneity, the GWAS results were
5 together meta-analysed separately with subjective and objective lithium response
6 GWAS results from Song et al. (Song, Bergen, et al., 2017; Song et al., 2016) and with
7 the ConLi⁺Gen GWAS results from Hou et al. (Hou et al., 2016). We conducted fixed-
8 effect meta-analyses with METAL (Willer et al., 2010) using each GWAS's Neff
9 number as weights (see Table 3.1). The two sets of GWAS meta-analyses were labelled
10 as UCL+Song^{Sub}+ConLi⁺Gen and UCL+Song^{Ob}+ConLi⁺Gen with corresponding data
11 input order. The UCL+Song^{Sub}+ConLi⁺Gen meta-analysis was used as the primary
12 analysis, given that Song et al.'s subjective definition mostly aligns with the UCL
13 definition, and that we found good evidence of overlap genetic association in these three
14 datasets. Correction for multiple testing was not attempted as Song et al.'s two sets of
15 GWAS results had highly overlapping samples. The genome-wide significance
16 threshold was set at $P < 5 \times 10^{-08}$.

17 The meta-analysis results were uploaded to Functional Mapping and Annotation of
18 Genome-Wide Association Studies (FUMA) for data interpretation(Watanabe et al.,
19 2017). Single tissue Expression QTL (eQTL) and single tissue splicing QTL (sQTL)
20 data for the top SNP was extracted from the GTEx project data V8 (dbGaP Accession
21 phs000424.v8.p2; GTEx Consortium, 2020) on 14/12/2023 focusing on brain regions.
22 We conducted functional fine-mapping using Polyfun+SuSiE (Weissbrod et al., 2020)

1 with the set number of causal variants as five per locus to identify potential causal SNPs.
2 We took the pre-computed summary LD information from the UK Biobank as the
3 reference and set the range to cover the start and end positions of the target locus. We
4 estimated the heritability of good lithium response in BD cases using LDSC taking the
5 GWAS meta-analysis results (Bulik-Sullivan et al., 2015). We examined the genetic
6 correlation between lithium responsive BD and SCZ (Trubetskoy et al., 2022) and
7 MDD (Howard et al., 2019). The top tissue from the MAGMA tissue expression
8 analysis was testis. We therefore examined the genetic correlation using summary
9 statistics of testosterone levels from Ruth et al. (2020).

10 Given that the lithium response phenotype was conditional on BD status, there was
11 potential collider bias. We applied the Slope-Hunter method (Mahmoud et al., 2022) to
12 examine if the results would be strongly influenced by the use of the PGC BD GWAS
13 summary statistics (Mullins et al., 2021). The summary statistics that we used excluded
14 data that the UCL group had contributed to the PGC and followed the same procedures
15 described in the original paper. However, some additional sample overlap remained for
16 the data from Song et al. (Song, Bergen, et al., 2017; Song et al., 2016) and ConLi⁺Gen
17 (Hou et al., 2016). This overlap meant that some of the statistical assumptions of the
18 Slope-Hunter method were broken. Thus, the Slope-Hunter method was only applied
19 to see if there would be major change in the results patterns, rather than producing
20 adjusted results.

21 The meta-analysis results were uploaded to FUMA for data interpretation
22 (Watanabe et al., 2017). Single tissue Expression QTL (eQTL) and single tissue splicing

1 QTL (sQTL) data for the top SNP was extracted from the GTEx project data V8 (dbGaP
2 Accession phs000424.v8.p2) on 12/14/2023 focusing on brain regions. We conducted
3 functional fine-mapping using Polyfun+SuSiE (Weissbrod et al., 2020) with the set
4 number of causal variants as five per locus to identify potential causal SNPs. We took
5 the pre-computed summary LD information from the UK Biobank as the reference and
6 set the range to cover the start and end positions of the target locus. We also estimated
7 the heritability of good lithium response BD cases using LDSC taking the GWAS meta-
8 analysis results (Bulik-Sullivan et al., 2015). And we examined the genetic correlation
9 between lithium responsive BD and the other two relevant disorders SCZ (Howard et
10 al., 2019), MDD (Trubetskoy et al., 2022) and the top trait from the MAGMA tissue
11 expression analysis which is testosterone using summary statistics from (Ruth et al.,
12 2020).

Table 3.1 The Summary of Study Sample Sizes

Sources	UCL Affy	UCL Psych	UCL GSA	Song et al.'s (Subjective)	Song et al.'s (Objective)	Hou et al.'s (ConLi+Gen)	UCL+Song ^{Sub} +ConLi+Gen	UCL+Song ^{Ob} +ConLi+Gen
Responders	164	247	29	1639	387	659	2738	1486
Non-responders	202	503	114	1059	792	1684	3562	3295
Total (R vs N)	366	750	143	2698	1179	2343	6300	4781
Neff (R vs N)	362	663	92	2573	1040	1895	5585	4052

Affy, Affymetrix 500K Array; Psych, Illumina PsychArray; GSA, Illumina Global Screening Array; UCL+Song^{Sub}+ConLi+Gen, the meta-analysis taking Song et al.'s subjective measures with the corresponding input order; UCL+Song^{Ob}+ConLi+Gen, the meta-analysis taking Song et al.'s objective measures with the corresponding input order.

Neff, the effective sample size used for METAL; R vs N, responders versus non-responders. The Neff numbers were calculated using the recommended methods as $4/(1/N \text{ cases} + 1/N \text{ controls})$, where N cases is the number of cases and N controls is the number of controls. The Neff numbers for UCL+Song^{Sub}+ConLi+Gen and UCL+Song^{Ob}+ConLi+Gen were calculated as the sum of corresponding samples' Neff numbers for each meta-analysis given that these were put as weights.

3.3.6 PRS calculation and analyses

PRSs for MDD and SCZ were computed using the PRS-CS-auto method (Ge et al., 2019). We chose the European samples from The 1000 Genomes Project Consortium (2010) as our LD reference panel. Once weights were produced, individual PRSs were calculated using PLINK v2.00a2LM (Chang et al., 2015). We then used the mean and standard deviation of the PRSs from the healthy controls to standardize the PRSs for the BD subjects.

We used data from Howard et al. (2019) for MDD GWAS which excluded the UK Biobank and 23andMe data. The SCZ GWAS was from Trubetskoy et al. (2022). We adapted all GWAS samples to be based on subjects of European ancestry. The summary statistics that were used excluded the data that the UCL group had contributed to the PGC SCZ analyses, but followed the same procedures as described by each GWAS paper. The effective GWAS sample sizes were calculated using the Neff equation (Willer et al., 2010). Thus, the reference GWAS sample sizes were 449,856 for MDD and 160,197 for SCZ. We performed multiple logistic regression analyses to assess how these PRSs predict lithium response in our own samples. The participants' age of BD onset, sex, genotyping chip, and first three principal components from population stratification were included as covariates in all regressions. The statistical assumptions were pre-checked and found to be satisfactory for each regression.

3.4 Results

3.4.1 Sample demographics

A total of 6300 and 4781 participants were included in the two sets of GWAS meta-analyses UCL+Song^{Sub}+ConLi⁺Gen and UCL+Song^{Ob}+ConLi⁺Gen respectively (see Table 3.1). Sample demographics were first combined to examine differences between lithium responders and non-responders (see Table 3.2). Overall, the participants had mean ages of 48.82 (SD 13.34) and 48.47 (SD 12.67) years. In the two meta-analyses, 43% and 31% of the participants responded well to lithium. The overall samples for the meta-analyses contained a high proportion of females (62% vs 59%) and type 1 BD patients (66% vs 68%). Responders were slightly older at the assessments (50.34 vs 47.64 & 50.60 vs 47.42; $p < 0.001$ in both datasets). Males were overrepresented as good responders to lithium (40% vs 37%, $p = 0.007$; 44% vs 40% $p = 0.005$) in both meta-analyses. In the UCL+Song^{Sub}+ConLi⁺Gen meta-analysis, there was an increased proportion of BD I participants coded as good lithium responders (68% vs 64%; $p = 0.016$). In the UCL samples, we observed the same pattern of sex differences in lithium response ($p = 0.039$) but not the bipolar subtype difference ($p = 0.189$, See Table S2). The UCL non-responders more likely had diagnosis of PPD (3% vs 1%, $p = 0.027$). To further validate the evidence for sex difference, we examined the heterogeneity across the samples. We did not observe any evidence for heterogeneity in the sex difference for the UCL+Song^{Sub}+ConLi⁺Gen meta-analysis ($Q = 0.31$, d.f.=2, $p = 0.859$, $I^2 = 0\%$) or the UCL+Song^{Ob}+ConLi⁺Gen meta-analysis ($Q = 2.04$, d.f.=2, $p = 0.361$, $I^2 = 1.9\%$). Participants who were lithium responders had risk ratios of 1.14

- 1 (UCL+Song^{Sub}+ConLi⁺Gen, 95%CI: 1.07 to 1.22) and 1.15 (UCL+Song^{Ob}+ConLi⁺Gen,
- 2 95%CI: 1.06 to 1.25) for being male according to the common effect model.

Table 3.2 GWAS Meta-analyses Participants' Demographics and Clinical Characteristics concerning Lithium Response

UCL+Song^{Sub}+ConLi⁺Gen	N	Overall N = 6300	Lithium Responders N = 2738 (43%)	Lithium Non-responders N = 3562 (57%)	p-values
Age at interview	6194	48.82 (13.34)	50.34 (14.05)	47.64 (12.77)	<0.001¹
Sex	6300				0.007²
Male		2408 (38%)	1082 (40%)	1326 (37%)	
Female		3892 (62%)	1656 (60%)	2236 (63%)	
BD types	3957				0.016 ²
Type 1		2607 (66%)	1406 (68%)	1201 (64%)	
Others		1350 (34%)	673 (32%)	677 (36%)	
UCL+Song^{Ob}+ConLi⁺Gen	N	Overall N = 4781	Lithium Responders N = 1486 (31%)	Lithium Non-responders N = 3295 (69%)	p-values
Age at interview	3733	48.47 (12.67)	50.60 (13.09)	47.42 (12.46)	<0.001¹
Sex	3839				0.005²
Male		1586 (41%)	556 (44%)	1020 (40%)	
Female		2253 (59%)	697 (56%)	1556 (60%)	
BD types	1496				0.059 ²
Type 1		1020 (68%)	429 (71%)	591 (66%)	
Others		476 (32%)	175 (29%)	301 (34%)	

UCL+Song^{Sub}+ConLi⁺Gen, the meta-analysis taking Song et al.'s subjective measures with the corresponding input order.

UCL+Song^{Ob}+ConLi⁺Gen, the meta-analysis taking Song et al.'s objective measures with the corresponding input order.

BD, bipolar disorder; SD, standard deviation.

¹ Two Sample t-test; mean (pooled SD)

² Pearson's Chi-squared test of independence; n (%)

Bold in current table for p<0.05.

Demographics for the ConLi⁺Gen EUR samples were estimated from their overall samples' demographics which included 220 East Asians. And Hou et al. did not report ConLi⁺Gen samples' BD types with the split of lithium response. The demographic information of the Swedish samples for Song et al.'s objective measures is not included because Song et al. reported a correction for the Swedish sample but did not report the updated demographics for the sample.

3.4.2 Lithium definition heterogeneity testing

Binomial sign-tests provided strong evidence for non-random convergence of SNP associations between the lithium response GWAS datasets. These findings became stronger at more stringent p-value thresholds (P_T : 1×10^{-3} , 1×10^{-4} , and 1×10^{-5} ; see Table 3.3). For the comparison of the Song^{Sub} data with the Hou et al., ConLi⁺Gen GWAS, 58% ($p=5.58 \times 10^{-53}$), 66% ($p=6.36 \times 10^{-22}$), and 83% of SNPs ($p=3.30 \times 10^{-30}$) had the same direction of effect at p-value thresholds of 1×10^{-3} , 1×10^{-4} , and 1×10^{-5} respectively. For the UCL meta-analysis, the percentages of SNPs with the same direction of effect as the Song^{Sub} data were similar with 52% ($p=2.17 \times 10^{-4}$), 70% ($p=1.51 \times 10^{-27}$), and 86% ($p=3.04 \times 10^{-33}$) at p-value thresholds of 1×10^{-3} , 1×10^{-4} , and 1×10^{-5} respectively.

LDSC analysis demonstrated a genetic correlation of 0.904 (SE 0.500) between UCL+Song^{Sub} and ConLi⁺Gen. There was low genetic correlation with BD (UCL+Song^{Sub} $r^2 = 0.143$, SE 0.093; ConLi⁺Gen $r^2 = -0.045$, SE 0.139). Additionally, according to the GSEM one-factor model, the UCL+Song^{Sub} and ConLi⁺Gen GWAS had high factor loadings of 0.997 and 0.907, respectively, indicating they load strongly onto the same factor, whereas BD had a low factor loading of 0.139 to that factor. These results indicated low genetic heterogeneity between lithium response GWAS datasets using clinical assessments or the Alda scale and indicate that these results were not highly related to genetic risk for BD.

Table 3.3 Binomial Sign Test Results for Response Heterogeneity

	N total	N SNP same direction	N SNP opposite direction	P
Song^{Sub} 10⁻³	7604			
ConLi ⁺ Gen	5635	3278 (58%)	2348 (42%)	5.583x10⁻⁵³
UCL	5674	2970 (52%)	2704 (48%)	2.169x10⁻⁴
Song^{Sub} 10⁻⁴	983			
ConLi ⁺ Gen	832	553 (66%)	279 (34%)	6.362x10⁻²²
UCL	734	512 (70%)	222 (30%)	1.509x10⁻²⁷
Song^{Sub} 10⁻⁵	318			
ConLi ⁺ Gen	276	229 (83%)	47 (17%)	3.303x10⁻³⁰
UCL	244	211 (86%)	33 (14%)	3.036x10⁻³³

Song^{Sub}, Song et al.'s subjective definition of lithium response GWAS

3.4.3 GWAS results for lithium responders vs non-responders

For the UCL+Song^{Sub}+ConLi⁺Gen meta-analysis, we observed little evidence for genomic inflation ($\lambda=1.01$, See Appendix 3.3). We found one SNP rs116927879 (A/G) reaching the genome-wide significance ($p=4.509 \times 10^{-08}$) on chromosome 7 with agreement for the direction of effect in all constituent samples (see Figure 3.1 & Table 3.4). The clumping from FUMA indicated that 196 GWAS-tagged candidate SNPs fell within the genomic locus from position chr7:45697297-45869101 (hg19, see Appendix 3.5). We conducted follow-up fine mapping with Polyfun+SuSiE to identify potential causal SNPs within the region. We observed no SNP with posterior causal probability (PIP) higher than 0.10. The three SNPs with the highest PIP were rs77411260, rs1128602, and rs116927879. The SNPs rs77411260 (chr7:45823152-45823652) and rs1128602 (chr7:45762446-45762946) were close to the top SNP rs116927879 (chr7:45864477-45864977) and did not map to any additional genes.

Genotype-Tissue Expression (GTEx) data (Release V8; dbGaP Accession phs000424.v8.p2) were analysed using the SNP rs116927879 (G/A) and the genes implicated by FUMA analyses focusing on brain areas (see details in Tables S4 & S5). Single-tissue eQTL suggested that rs116927879 genotypes influence expression of *GTF2IP13* in the cortex (Normalized effect size (NES)=-0.66, $p=1.4 \times 10^{-20}$), caudate (NES=-0.66, $p=1.8 \times 10^{-17}$), hippocampus (NES=-0.63, $p=7.5 \times 10^{-16}$), and other brain areas. The single-tissue sQTL analyses suggested that rs116927879 genotypes influence splicing of *SEPT7P2* in the cerebellum (NES=-1.3, $p=3.4 \times 10^{-25}$), nucleus accumbens (NES=-1.4, $p=1.1 \times 10^{-20}$), cortex (NES=-1.1, $p=2.3 \times 10^{-20}$), and other

1 brain areas. sQTL analyses also suggested that rs116927879 genotypes influenced
2 splicing of *ADCY1* in the cortex (NES=-0.66, $p=1.4 \times 10^{-10}$), and the frontal cortex
3 (BA9) (NES=-0.66, $p=2.9 \times 10^{-9}$) at junction 10 (chr7:45662214:45677869, hg38) of
4 *ADCY1*.

5 For the UCL+Song^{Ob}+ConLi⁺Gen meta-analysis, we found little evidence for
6 inflation ($\lambda=1.03$, see Appendix 3.4) but we observed no SNP reaching the genome-
7 wide significance. We conducted the same eQTL and sQTL analyses for the top SNPs
8 from this meta-analysis however, no significant outputs could be observed. The top four
9 lead SNPs from the UCL+Song^{Sub}+ConLi⁺Gen meta-analysis, rs116927879, rs4761584,
10 rs12296932 and rs28728196 were tested for the directions of effect in the
11 UCL+Song^{Ob}+ConLi⁺Gen meta-analysis. The directions for all remained unchanged,
12 suggesting consistent effect direction.

13 With the application of the Slope-Hunter method, the adjustment factors obtained
14 for the two sets of meta-analyses were negative implying that variants of BD and good
15 lithium response were of concordant net directions of effect. We observed no major
16 change in the result patterns from the adjusted outputs but only slight attenuation
17 towards the null. These results suggested that the identified top SNPs from the meta-
18 analyses were distinct from BD risks.

Figure 3.1 The Manhattan Plots for Two Sets of GWAS Meta-analyses

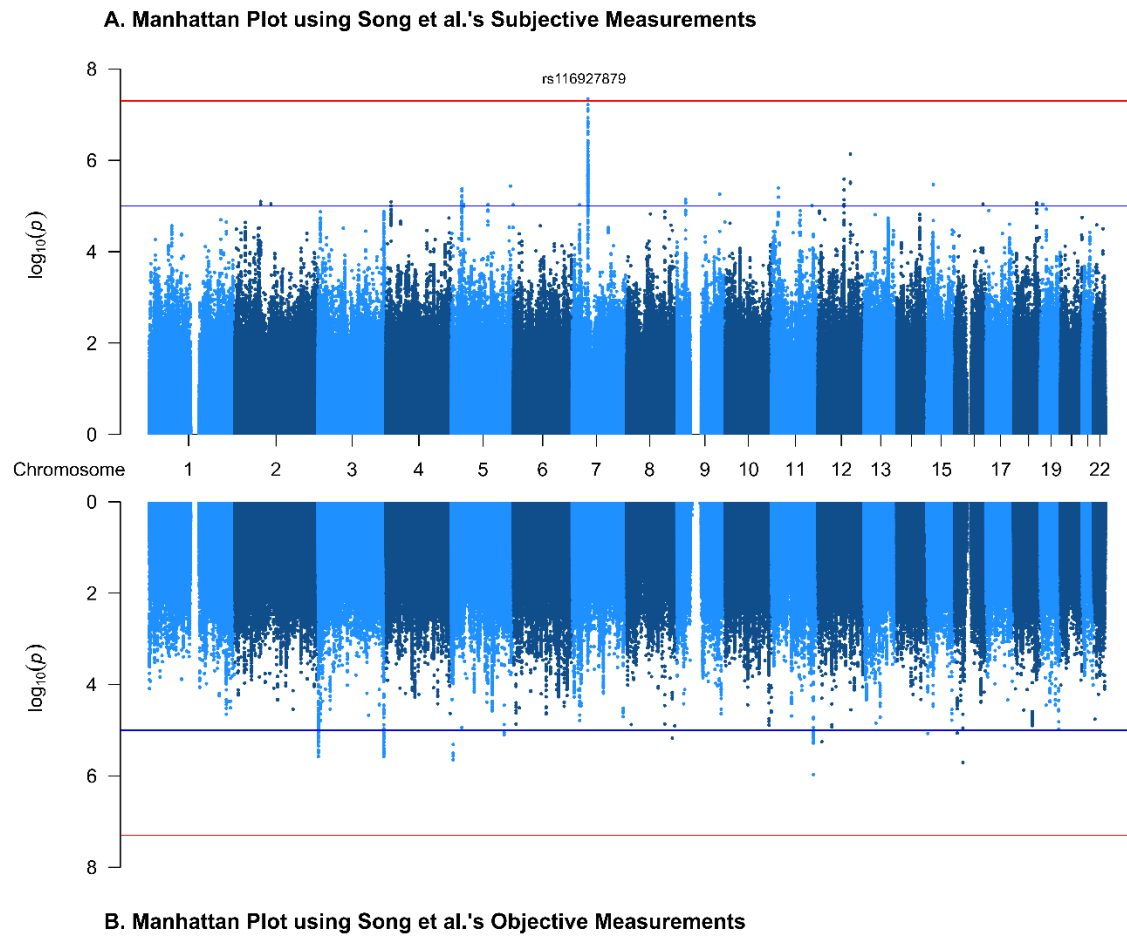


Table 3.4 Regions of the Genome showing the Strongest Association Signals with the Binary Trait

Index SNP	Chr	Position	A1/A2	EAF	Z Score	P-values	Match	Weight (Neff)	Mapped Genes
<i>Responders vs non-responders (using Song et al.'s subjective measurements, UCL+Song^{Sub}+ConLi⁺ Gen, 2738 cases 3562 controls, max Neff 5585)</i>									
rs116927879	7	45697297-45869101	A/G	0.152	-5.470	4.509×10^{-08}	-----	5585	<i>ADCY1, SEPT7P2, GTF2IP13, C1CP20</i>
rs4761584	12	94518829-94527590	T/C	0.250	4.953	7.302×10^{-07}	+++++	5585	<i>RP11-1105G2.3, CCDC41, TMCC3</i>
rs12296932	12	76062748-76111969	T/G	0.352	-4.703	2.569×10^{-06}	-----	5585	<i>KRRI, PHLDA1, NAPILI</i>
rs28728196	15	39533087-39562972	A/G	0.827	-4.645	3.401×10^{-06}	-----	5585	<i>C15orf53, C15orf54, THBS1, FSIP1</i>
<i>Responders vs non-responders (using Song et al.'s objective measurements, UCL+Song^{Ob}+ConLi⁺ Gen, 1486 cases 3295 controls, max Neff 4052)</i>									
rs78026375	11	121673147-121768986	T/G	0.082	4.879	1.066×10^{-06}	+++++	4052	<i>SC5D, CRTAM, ARHGEF12, BLID</i>
rs9933339	16	23756216-23865532	A/C	0.816	-4.758	1.957×10^{-06}	???--	2935	<i>PALB2, ERN2, PLK1, CHP2</i>
rs2913631	5	5234709-5234709	A/G	0.260	4.729	2.254×10^{-06}	?++++	3690	<i>ADAMTS16</i>
rs709122	3	191425855-191549043	T/C	0.375	4.728	2.273×10^{-06}	?++++	3690	<i>PYDC2, FGF12</i>

Index SNP, the single-nucleotide polymorphism with the strongest association in the genomic region and each is independent at $r^2 < 0.1$; Chr, chromosome; Position, the start and end position (UCSC hg19) of the SNP locus where near-by SNPs were clumped to with nominal associations ($p < 0.05$) and LD ($r^2 < 0.1$) within 250-kb windows taking the 1000 genomes project phase 3 EUR as LD reference; A1/A2, effect and alternate allele; EAF, the effect allele frequency based on 1000 genomes EUR; Z-score, the meta-analysis output reference score for the SNP; P-values, the corresponding p-values to the candidate SNP; Match, the agreement across the five datasets, + means individuals who carry the A1 allele have better lithium response, - means negative, ? means missing, the orders are: three sets of GWAS from our own samples (1) Affymetrix Array, (2) Illumina PsychArray, (3) GSA, Illumina Global Screening Array, (4) Song et al.'s samples using subjective/objective measures, and (5) Hou et al.'s EUR samples (ConLi⁺Gen) ; Weight, the overall Neff of the sample for the SNP; Mapped genes, top 4 genes mapped by positional mapping criterion with maximum distance 10 kb. In bold SNP passed genome-wide significant threshold.

3.4.4 SNP-heritability estimation & genetic correlations

SNP heritability (h^2) for good lithium response was 20.3% (SE 8.4%) for the UCL+Song^{Sub}+ConLi⁺Gen meta-analysis and 15.6% (SE 9.1%) for the UCL+Song^{Ob}+ConLi⁺Gen meta-analysis. We observed no genetic correlation between good lithium response and SCZ for the two sets of GWAS meta-analyses (subjective: $Z=-0.625$, $p=0.532$; objective: $Z=-1.239$, $p=0.215$). Nor did we observe any genetic correlation between good lithium response and MDD for the two sets of GWAS meta-analyses (subjective: $Z=-1.753$, $p=0.080$; objective: $Z=-1.336$, $p=0.182$). However, we found weak evidence to suggest the genetic correlation between good lithium response and total testosterone levels for both sets (subjective: 0.231 (SE 0.112); $Z=2.064$, $p=0.039$; objective: 0.347 (SE 0.161), $Z=2.158$, $p=0.031$). Importantly, there was no genetic correlation between BD and total testosterone levels ($Z=0.505$, $p=0.614$) suggesting the association was more relevant to lithium response.

3.4.5 PRS analyses results

We attempted to replicate previous studies, which reported that lithium response could be predicted using PRS for SCZ and MDD. We found no evidence to suggest that SCZ PRS was predictive of lithium response in the UCL sample (OR 1.040, 95%CI: 0.916 to 1.182, $p=0.542$). We also found no association between MDD PRS and lithium response (OR 0.985, 95%CI: 0.868 to 1.119, $p=0.818$).

3.5 Discussion

Here we present the largest GWAS meta-analyses of lithium response on European samples to date. By including data from the UCL study, together with Song et al.'s subjective and objective measures and ConLi⁺Gen's European samples, our GWAS meta-analyses included 6,300 and 4,781 participants respectively. We identified one SNP, rs116927879, that reached the genome-wide significance level. The genomic interval implicated by this finding included the *ADCY1* (Adenylate Cyclase 1) gene and two pseudo-genes, *GTF2IP13* and *SEPT7P2*. While the function of *GTF2IP13* and *SEPT7P2* remain poorly understood, *ADCY1* plays roles in mediating responses to increased cellular Ca²⁺/calmodulin levels which is relevant to regulatory processes in the central nervous system, as well as memory and learning.

Song et al. (Song, Bergen, et al., 2017; Song et al., 2016) also reported association between rs116927879 and subjective lithium response, but this finding did not achieve genome-wide significance in their original study ($p=4.99\times10^{-7}$). With additional samples from the current UCL study and from ConLi⁺Gen, rs116927879 became genome-wide significant with agreement for the direction of effect from all sources. Potential association with *ADCY1* and SCZ has been reported previously (Goes et al., 2015; Sundararajan et al., 2018) and it was also the only gene identified in a cross-trait GWAS meta-analysis of SCZ and lithium response (ConLi+Gen, 2018).

ADCY1 plays essential roles in the regulatory processes in the central nervous system that are critical for neurodevelopment and neuroplasticity (J. Chen et al., 2022). It has been suggested that the circadian modulation of contrast sensitivity is associated

1 with Dopamine D4 Receptors (D4Rs) primarily through the *ADCY1* signalling pathway
2 (Hwang et al., 2013). Sleep disturbances and circadian rhythm dysfunction have been
3 commonly noted among patients with BD (Takaesu, 2018). In addition, the sQTL data
4 suggested that rs116927879 genotypes may influence expression and splicing of
5 *ADCY1* across different brain regions.

6 Three main isoforms of *ADCY1* have been reported, these include the canonical
7 longer isoform (ENST00000297323.12) and two shorter isoforms
8 (ENST00000432715.5 and ENST00000621543.1). The sQTL data from GTEx
9 suggests that SNP alleles associated with poor lithium led to increased splicing at
10 junction 10 (chr7:45662214:45677869, hg38) that in turn is likely to result in the
11 production of the shorter isoforms of *ADCY1*. The peptide encoded by the long isoform
12 of *ADCY1* contains two adenylate cyclase domains, twelve transmembrane domains
13 and two calmodulin interaction domains. The shorter isoforms contain the first
14 adenylate cyclase domain and is therefore likely to have reduced activity. Further
15 research is needed to fully elucidate the specific functions of these isoform types and
16 their associations with response to lithium.

17 We estimated the SNP heritability (h^2) for good lithium response to be 20.3%
18 (subjective meta-analysis) and 15.6% (objective meta-analysis). Inverse correlations
19 between lithium response and PRS for SCZ and MDD have been reported for the
20 ConLi+Gen data (Amare et al., 2021; ConLi+Gen, 2018; Schubert et al., 2021). In our
21 own subject level data, we were not able to replicate these findings. Additionally, we
22 did not observe genetic correlation between lithium response and SCZ/MDD.

1 We found weak evidence to suggest a genetic correlation between lithium response
2 and genetically predicted testosterone levels which was consistent with our finding of
3 a sex difference in lithium response. Testosterone, together with other sex hormones
4 have been suggested to influence the development of mood disorders (G. Lombardo et
5 al., 2021). Lithium administration in rats has been reported to reduce male fertility
6 parameters such as testosterone and gonadosomatic index (Abdelwahab et al., 2022).
7 Data on the effect of sex on lithium response have been contradictory. Two large-scale
8 meta-analytical studies did not observe sex differences in terms of lithium response
9 (Hui et al., 2019; Rybakowski, 2014). However, a nationwide study conducted in
10 Denmark (N=3762) found that females were more likely to be lithium non-responders
11 (Kessing et al., 2011). Although we observed sex differences in the overall samples, the
12 ratio differences were still quite small.

13 The primary limitation to the study is that we used a cross-sectional study design
14 to collect lithium response data for our own samples which may be hampered by recall
15 bias. Another concern could be whether to treat lithium response as binary or
16 continuous due to potential differences in results. In addition, the lithium response
17 coding in our and Song et al.'s samples were essentially based on clinical impression
18 whilst the ConLi⁺Gen group used criteria-based Alda scale. Even though we found no
19 evidence for definition heterogeneity and a strong genetic correlation between GWAS
20 summary statistics generated using these different approaches, differences in recording
21 lithium response may have impacted the results. The findings from the current study
22 suggest that sex may slightly influence lithium response and future analyses should

1 consider formally adjusting for it. Finally, we only included European samples in the
2 analyses, and therefore the results presented may have limited transferability to people
3 from different ancestries.

4 Overall, our GWAS meta-analyses provided new insights for lithium response
5 among BD patients. We identified one SNP rs116927879 (A/G) reaching genome-wide
6 significance on chromosome 7 in a region that included the genes *ADCY1*, *SEPT7P2*,
7 and *GTF2IP13*, which could further explain lithium's bio-mechanisms. Furthermore,
8 we first identified sex differences between lithium responders and non-responders on a
9 genetic level. Future large-scale studies exploring lithium response among BD subjects
10 that properly account for potential sex differences are encouraged.

4. Integrating Genome-wide and Epigenome-wide Associations for Antipsychotic Induced Extrapyrarnidal Side Effects

A version of this paper has been submitted as a preprint at medRxiv.

4.1 Abstract

Background: Antipsychotic medications are the first-line treatment for schizophrenia. However, around 40% of people with schizophrenia who are treated with antipsychotics could develop extrapyramidal side-effects (EPSE) including: 1) Dyskinesias, 2) Parkinsonism, 3) Akathisia, and 4) Dystonia. This study aimed to identify genetic risk factors for EPSE presence following antipsychotic treatment.

Method: We conducted Genome-wide association (GWAS) and Epigenome-wide association (EWAS) meta-analyses of EPSE, with subset analyses separating first and second generation antipsychotic (FGA/SGA) exposure. We integrated significant EWAS findings from a between-case design to a comparable GWAS for association enrichment. We investigated whether polygenic risk scores (PRS) for schizophrenia, Parkinson's disease, and Lewy-body dementia could predict EPSE.

Results: The primary GWAS top SNP rs2709733 (A/G) ($p=5.755 \times 10^{-07}$) mapped to a long intergenic non-protein coding RNA, *LINC01162* with consistent effects across all cohorts. Subset analyses with distinct FGA exposure indicated suggestive genes such as *NAV2*, *NRG3*, *LSAMP* and SGA exposure indicated *SHISA9* and *CNBD1* which are relevant for schizophrenia, autism, and epilepsy. In our primary EWAS, the most significant differentially methylated position (DMP) was cg05599348 (3.181×10^{-07}), located at chrX:103174718 (hg19) mapping to *TMSB15B*. Comparing EPSE cases to

1 healthy controls, we identified nine DMPs associated with EPSE. The DMP
2 cg12044923 (chr2:241453995, hg19), located within the *STK32B* gene, showed
3 significant enrichment for EPSE association (permutation $p=0.010$). *STK32B* is relevant
4 to both psychiatric and movement disorders, suggesting potential shared mechanisms.
5 *Conclusion:* Our study sheds new light on the potential biological mechanisms
6 underlying EPSE development in schizophrenia, highlighting the importance of
7 exploring both methylation shifts and common SNP associations. Further research with
8 larger samples sizes and a focus on the role of *STK32B* are encouraged.

9

10 **4.2 Introduction**

11 Antipsychotic medications are the first-line treatment for schizophrenia (Sabe et al.,
12 2022). Although many people benefit, around 70% may experience treatment failure
13 such as psychiatric rehospitalization, suicide attempt, discontinuation or switch to other
14 medication (Tiihonen et al., 2017). Extrapyramidal side-effects (EPSE) are common
15 with antipsychotic treatment (Carbon et al., 2018; Huhn et al., 2019), with
16 approximately 40% of patients treated with first-generation antipsychotics (FGA)
17 experiencing EPSEs (Wubeshet et al., 2019). EPSEs still occur with second-generation
18 antipsychotics (SGA), although at lower rates in comparison with FGA (Divac et al.,
19 2014). FGAs are primarily dopamine D2 receptor antagonists, which reduce
20 dopaminergic activity to alleviate positive symptoms of psychosis. This, however, often
21 leads to motor side effects such as EPSE (Kapur & Remington, 2001). Meanwhile, SGA
22 targets both dopamine D2 and other receptors such as serotonin 5-HT_{2A} receptors.

1 Serotonin modulation offsets some dopamine blockade effects thus reducing EPSE
2 (Leucht et al., 2009; Zhang et al., 2013). EPSEs describe the movement abnormalities
3 induced by antipsychotics including:

4 1) *Dyskinesia*, hyperkinetic choreiform involuntary movements of the face,
5 extremities, and the trunk (Lim et al., 2021a). When dyskinesia persists for more than
6 one month it is termed tardive dyskinesia which can sometimes become chronic.

7 2) *Parkinsonism*, symptoms of rigidity, tremor and impaired or slow movement
8 (bradykinesia) (Keener & Bordelon, 2016).

9 3) *Akathisia*, characterised by subjective inner restlessness and objective increase
10 in motor activity such as pacing (Factor et al., 2008).

11 4) *Dystonia*, characterised by sustained and abnormal contractions, that can result
12 in abnormal movements and postures (van Harten & Kahn, 1999).

13 These movement abnormalities can lead to severe impairment and reduction in the
14 quality of life of individuals with schizophrenia (D'Souza & Hooten, 2023), by
15 interfering with daily living activities and social functioning (Fujimaki et al., 2012;
16 Schouten et al., 2012). In a meta-analysis, the prevalence of spontaneous dyskinesias
17 and parkinsonism was found to be higher in antipsychotic-naïve patients with
18 schizophrenia and in first-degree relatives of patients with schizophrenia as compared
19 to healthy controls, indicating a heritable, non-drug induced component to these
20 abnormalities (Koning et al., 2010).

21 Parkinsonism seen in EPSE can be clinically indistinguishable from the movement
22 abnormalities seen in the neurological disorders like Parkinson's disease (PD) and

1 Lewy Body Dementia (LBD). Previous studies have identified shared significant loci
2 between schizophrenia and PD (Nalls et al., 2019; Smeland et al., 2021). For example,
3 schizophrenia and PD are both associated with the 22q11.2 deletion syndrome (R. K.
4 Jonas et al., 2014). A duplication of the *SNCA* gene, for which pathogenic variants are
5 associated with autosomal dominant Parkinson's and encodes α -synuclein, a major
6 constituent of LBD, was reported in an individual diagnosed with schizophrenia nine
7 years prior to the development of mild Parkinsonism (Takamura et al., 2016). A recent
8 neuroimaging study on individuals with first episode psychosis found that higher iron
9 loading in the basal ganglia correlated with greater motor abnormalities including EPSE
10 (Cuesta et al., 2021). Similar associations were found with motor abnormalities in PD
11 (Kim & Wessling-Resnick, 2014; R. J. Ward et al., 2014). In view of this, it is plausible
12 that there are shared genetic features between these disorders which also contribute to
13 the shared phenotypical features including movement abnormalities like EPSE in
14 schizophrenia.

15 Genome-wide Association Studies (GWAS) are a promising approach to identify
16 potential genes associated with development of EPSE given the often-complex
17 biological pathways implicated in psychiatric traits (Duncan et al., 2019). However, to
18 our knowledge, only one past study investigated antipsychotic induced EPSE using
19 GWAS comparing EPSE cases versus EPSE controls among European schizophrenia
20 samples (Åberg et al., 2010). The genotype data in that study had somewhat limited
21 genomic coverage compared to contemporary studies and furthermore there was no
22 imputation of genotypes not captured on the genotyping array. Other studies have

1 examined EPSE presence by comparing EPSE cases with healthy controls (Levchenko
2 et al., 2021) and by analysing mixed ancestry cohorts (Lim et al., 2021b). Epigenome-
3 wide Association Study (EWAS) allows for the examination of environmentally
4 induced methylome variation which could directly result from chronic antipsychotic
5 exposure (Murphy & Mill, 2014; Wagner et al., 2014). To date, there has been no EWAS
6 on EPSE to examine the influence from antipsychotics.

7 Our understanding of the molecular mechanisms underlying EPSE may be
8 improved using an integrated functional genomics strategy. The overall aim of this
9 study was to conduct an integrated GWAS and EWAS meta-analysis of EPSE data from
10 existing schizophrenia studies. We also investigated whether Polygenic Risk Scores
11 (PRS) for schizophrenia, PD and LBD could be used to predict the risk of the
12 development of EPSEs. The findings could provide a better understanding of the
13 genetic underpinnings of EPSE and pave the way for the identification of informative
14 genetic biomarkers that could allow for specific tailoring of treatments in the future.

15

16 **4.3 Methods**

17 **4.3.1 Participants selection and genotyping**

18 *UCL Participants* All UCL participants received an ICD10 diagnosis of schizophrenia
19 from a UK National Health Service (NHS) psychiatrist. Details have been reported
20 elsewhere (Trubetskoy et al., 2022; World Health Organization, 1992). The participants'
21 PPD diagnosis was assessed with OPCRIT item 11. Ancestrally matched healthy
22 controls were recruited from the National Health Service (NHS) blood transfusion
23 service and from study sites where case participants were also being recruited. The

1 healthy controls were screened for an absence of a lifetime history of the following
2 disorders: schizophrenia and any other psychosis, major affective or schizoaffective
3 disorders, eating disorders, alcohol/drug addiction, and obsessive-compulsive disorders.
4 All participants read an approved information sheet and signed a physical informed
5 consent form. The study was approved by the NHS Metropolitan Multi-centre Research
6 Ethics Committee (MREC/03/11/090). Genome-wide single nucleotide polymorphism
7 data were generated in three waves at the Broad Institute, Boston, MA, US, using the,
8 Affymetrix Array, Illumina PsychArray, and Illumina Global Screening Array (GSA).
9 The three waves of data underwent equivalent quality control and imputation methods
10 which had been described in details elsewhere (Grigoriu-Serbanescu et al., 2020).

11 *Aberdeen Participants* The Aberdeen case-control sample has been described
12 elsewhere.(Stone et al., 2008) Briefly, the cohort contains participants with
13 schizophrenia and healthy controls who have self-identified as born in the British Isles
14 (95% in Scotland). All participants with schizophrenia met the Diagnostic and
15 Statistical Manual for Mental Disorders fourth edition (DSM-IV; American Psychiatric
16 Association, 1994) and ICD-10 criteria for schizophrenia (World Health Organization,
17 1992). Controls were volunteers recruited through general practices in Scotland.
18 Volunteers who replied to a written invitation were interviewed using a short
19 questionnaire to exclude major mental illness in the individual themselves and their
20 first-degree relatives. The study was approved by both local and multiregional academic
21 ethical committees and all cases and controls gave informed consent. The samples were
22 genotyped at the Broad Institute, as described for the UCL participants.

Cardiff Participants Participants were recruited from community mental health teams in Wales and England on the basis of a clinical diagnosis of schizophrenia or schizoaffective disorder (depressed sub-type) as described previously (Carroll et al., 2011). Diagnosis was confirmed following a SCAN interview (Wing et al., 1990) and review of case notes followed by consensus diagnosis according to DSM-IV criteria (American Psychiatric Association, 1994). The UK Multicentre Research Ethics Committee (MREC) approved the study and all participants provided informed consent. The samples were genotyped at the Broad Institute, as described for the UCL participants.

UK Biobank (UKB) Participants UKB is a biomedical database and research resource of approximately 500,000 individuals from across the UK aged 40 to 69 years at recruitment between 2006 and 2010 (Sudlow et al., 2015). Potential participants in UKB were selected using diagnosis of schizophrenia from ICD10, including codes from F20.0 to F20.9 and excluding participants with any primary Parkinson disorder with G20.

4.3.2 Coding of EPSE data

Participant EPSE status was derived from the data described above following: (1) prescription of antipsychotic medications (FGA or SGA); (2) recorded clinical features of EPSE side-effects; and/or recorded medications prescribed to alleviate EPSE side-effects. Medication dose information was unavailable for most participants thus, not analysed. We used key terms to classify participants with schizophrenia as cases

1 (having life-time EPSE) or controls (not having life-time EPSE) from all available data
2 sources including interviews, medication, and health records. These key words covered
3 two main areas: behavioural and pharmacological:

4 *1) Behavioural features of the four types of EPSE, (dystonia, akathisia,*
5 *parkinsonism, and dyskinesia).* To compile a list of keywords for each of these EPSE
6 types, we consulted several rating scales that are frequently employed to measure
7 EPSE including: The Abnormal Involuntary Movement Scale (AIMS; Munetz &
8 Benjamin, 1988) the Extrapyrmidal Symptom Rating Scale (ESRS; Chouinard &
9 Margolese, 2005), The Simpson Angus Scale (Hawley et al., 2003) and the Barnes
10 Akathisia Rating Scale (BARS; Barnes, 1989). In addition, we searched reliable sources
11 of clinical information for each of these abnormalities including the National Institute
12 for Health and Care Excellence (NICE) guidelines (NICE, 2014b) and the BMJ Best
13 Practice (BMJ Best Practice, 2021).

14 *2) Pharmacological treatments for EPSE.* To generate key words for
15 pharmacological treatments for EPSE, we searched The NICE guidelines(NICE, 2014b)
16 and The Maudsley Prescribing Guidelines in Psychiatry(Taylor et al., 2021) for the most
17 recent recommendations on managing EPSE to identify a list of medications.

18 The UCL and Aberdeen participants' EPSE status was derived using the same list
19 of key words described in Appendices 4.1 and 4.2. The Cardiff participants' EPSE
20 status coding had a few minor adaptations. The keywords "dribbling" was added as it
21 better captured other saliva-related key-words; 'shakes' was removed as it was
22 described in the context of anxiety; "still" was removed as it referred to still doing

1 something not being physically still; “tap” was removed as it was in the context of
2 ‘tapered’; “march” was removed as it referenced the month of March; “irritable” was
3 removed as it was in the context of IBS/irritable bowel syndrome; “parkin” was
4 removed as it referred to Parkinson’s disease not parkinsonism; ‘tropin’ was excluded
5 as it captured atropine as opposed to benztropine. The UKB participants were retained
6 if they received any first or second generation of antipsychotics (See medication codes
7 in Appendices 4.3 and 4.4), then stratified by whether participants received any
8 medication to treat EPSE (See EPSE medication codes in Appendix 4.5); diagnosis of
9 other drug-induced secondary Parkinsonism in G21.1; Drug-induced dystonia in G24.0
10 or Drug-induced tremor in G25.1 were selected as cases.

11

12 **4.3.3 GWAS meta-analyses & follow-up analyses**

13 For the main analysis, we took a within case design comparing participants with
14 exposure to FGA or SGA with EPSE vs not having EPSE. We also conducted subset
15 analyses separating participants who had any exposure to FGA (including those also
16 exposed to a SGA) and participants only exposed to SGA. The Cardiff samples were
17 only included in the SGA subset given most participants only had SGA exposure. We
18 stratified the antipsychotic exposure into separate groups to compare their differential
19 effects, given FGAs' higher EPSE prevalence and distinct mechanistic profiles.

20 We applied logistic regressions taking the participants’ EPSE status to evaluate the
21 association between imputed SNP dosages. For UCL participants, we performed
22 separate GWAS for data from each wave using PLINK v2.00a2LM (Purcell et al., 2007).

1 We conducted the same sets of analyses for Aberdeen, Cardiff, and UKB samples
2 separately. The participants' age, sex and the first ten principal components of
3 population structure were included as covariates to control for population stratification.

4 We conducted fixed-effect meta-analysis taking each GWAS's effective sample
5 sizes (Neff) as weights using METAL (See calculation of Neff in Supplementary Table
6 6) (Willer et al., 2010). The genome-wide significance threshold was set at $P < 5 \times 10^{-8}$.
7 The output results were uploaded to FUMA for interpretation (Watanabe et al., 2017).
8 We also conducted a binomial sign test to evaluate the SNP associations between the
9 FGA and SGA subsets at 10^{-3} level. If there were no SNPs associations between the
10 FGA and SGA subsets, the expectation is that 50% of the Z scores from the meta-
11 analyses would be in the same direction.

12

13 **4.3.4 EWAS methylation data**

14 Methylation data was only available for a proportion of the UCL and Aberdeen
15 samples. The EZ-96 DNA Methylation kit (Zymo Research, CA, USA) was used to
16 treat 500ng of DNA from each sample with sodium bisulfite in duplicate. DNA
17 methylation was quantified using the Illumina Infinium HumanMethylation450
18 BeadChip (Illumina Inc.) run on an Illumina iScan System (Illumina) using the
19 manufacturers' standard protocol. Detailed data collection and imputation process has
20 been described elsewhere (Hannon et al., 2016). As smoking status information was not
21 present for all samples, we estimated a proxy based on the DNA methylation profile at
22 sites known to be associated with smoking status following a previously described

1 approach (Elliott et al., 2014). Cell composition data were not available for these DNA
2 samples, therefore these were estimated Houseman algorithm (Houseman et al., 2012;
3 Koestler et al., 2013) for seven variables recommended in the documentation. We also
4 estimated the participants' methylation age using the Epigenetic Clock software
5 (Horvath, 2013).

6

7 **4.3.5 EWAS analysis and meta-analyses**

8 We employed the same design as in the GWAS to analyse the association of EPSE
9 status on DNA methylation profiles. This included comparisons of EPSE presence
10 among participants with any FGA/SGA exposure (111 EPSE cases, 203 EPSE controls),
11 any FGA exposure (87 EPSE cases, 87 EPSE controls), and only SGA exposure (17
12 EPSE cases, 106 EPSE controls). DNA methylation values for each probe were
13 regressed with covariates for methylation age, gender, seven cell composition scores,
14 and smoking score. Then the results from UCL and Aberdeen were combined with
15 fixed-effect meta- analyses.

16 These within-case analyses may be limited by sample size constraints, potentially
17 reducing statistical power to detect subtle methylation changes. We also performed
18 EPSE case with any antipsychotic exposure vs healthy control analyses to boost
19 statistical power (111 EPSE cases, 748 healthy controls). To eliminate the influence of
20 schizophrenia from the case control study design we included participants
21 schizophrenia PRS as an additional covariate. Thus, the EPSE case control design may
22 help reveal EPSE-associated methylation changes attributable to long-term

antipsychotic exposure after accounting for potential schizophrenia risks. The EWAS meta-analysis significance threshold was set at 1×10^{-07} .

4.3.6 EWAS findings integration and permutation test

We performed separate GWAS on the same participants used in the between-case design EWAS following the same procedure described. The results were combined using METAL and clumped to represent LD independent loci in lead using the 1000 genome European samples as a reference (1000 Genomes Project Consortium et al., 2015). Any significant CpG sites from the EWAS were mapped to within 250 kb of each in the associated GWAS results to identify an enrichment in the region. To quantify significance, 5000 random permutations were generated. Empirical P values for each region were calculated by counting how many of the permutations had more significant P values than the mapped P value from GWAS and dividing by the total number of permutations performed. The CpG sites' locations were also mapped to clumped schizophrenia GWAS results within 250 kb for comparisons (Trubetskoy et al., 2022). Regional plots were produced using GWASLab (He et al., 2023).

4.3.7 PRS calculation and analyses

We calculated the participants' PRSs for schizophrenia, Lewy body dementia and Parkinson's disease using the PRS-CS method with the latest available reference GWAS (Ge et al., 2019). We chose the European samples from the 1000 Genomes Project Consortium as our LD reference panel given all samples included were of

1 European Ancestry (1000 Genomes Project Consortium et al., 2015). Once weights
2 were produced, individual PRSs were calculated using PLINK v2.00a2LM.(Chang et
3 al., 2015) We then used the mean and standard deviation of the healthy controls' PRSs
4 from each sample to standardize their cases' PRSs. The SCZ GWAS came from
5 Trubetskoy et al. (PGC wave 3), which were derived exclusively from European
6 samples (Trubetskoy et al., 2022). The GWAS statistics for Parkinson's disease came
7 from European samples of Nalls et al. excluding 23andMe data (Nalls et al., 2019). The
8 GWAS statistics for Lewybody dementia came from Chia et al., only including
9 European samples (Chia et al., 2021). We adapted the schizophrenia GWAS to exclude
10 each sample's participants used in the current study to avoid sample overlap. The new
11 GWAS generation followed the same procedures as previously described (Trubetskoy
12 et al., 2022).

13 We performed multiple logistic regression analyses to assess how these various
14 PRSs predict the presence of EPSE in each sample. Then the results were meta-analysed
15 using a fixed effect model. The assumptions for logistic regressions were pre-checked
16 and found to be satisfactory for each regression. The significant threshold was kept as
17 0.0167 (i.e. 0.05/3), for multiple testing correction.

18

19 **4.4 Results**

20 **4.4.1 GWAS sample demographics**

21 Overall, the GWAS meta-analysis included 2471 participants with schizophrenia,
22 of whom 1178 (48%) had EPSE. The participants had a mean age of 46.57 (SD 12.22)

1 years old and were mostly males (70%; Table 1) as is typical of genomic studies of
2 schizophrenia. All participants had antipsychotic exposure and most of the participants
3 had taken at least one type of SGA (78%). The participants with and without EPSE did
4 not differ in terms of age at assessment (46.53 vs 46.62, $p=0.855$) nor sex (males 71%
5 vs 69%, $p=0.213$). EPSE was more prevalent in those who had taken FGA or were on
6 both (FGA 51%, SGA 41%, both 61%; $p<0.001$). The participants' characteristics
7 differed between sample sets (see Table 4.2). The participants who developed EPSE
8 were at an older age at assessment than those who did not in the UCL (46.33 vs 42.72,
9 $p<0.001$) and UKB samples (56.06 vs 53.94, $p=0.020$; Table 4.2). In the Cardiff sample,
10 participants who developed EPSE had an earlier age of schizophrenia onset (24.30 vs
11 27.50, $p=0.006$). The pattern of EPSE being more prevalent in those who have taken
12 the first generation of antipsychotics were consistent across most cohorts (UCL 60% vs
13 40% $p<0.001$; Aberdeen 64% vs 39% $p<0.001$; UKB 69% vs 30% $p<0.001$) except for
14 the Cardiff samples where most participants only had exposure to SGA (first 18% vs
15 second 82%). From the UCL sample, there was no evidence to suggest any association
16 between PPD and EPSE ($p=0.999$) nor association between EPSE and prescriptions of
17 any antidepressants ($p=0.276$).

Table 4.1 GWAS Meta-analysis Participants' Demographics and Clinical Characteristics concerning EPSE Presence

	N	Overall	EPSE Presence	EPSE Absence	p-values
Meta-Analyses	<i>2471</i>	<i>n = 1178 (48%) n = 1293 (52%)</i>			
Age at assessment	2471	46.57 (12.22)	46.53 (11.82)	46.62 (12.59)	0.855 ^a
Sex	2471				
Male		1727 (70%)	838 (71%)	889 (69%)	0.213 ^b
Female		744 (30%)	340 (29%)	404 (31%)	
Antipsychotics	2425				
First Generation		488 (20%)	249 (22%)	239 (19%)	<0.001^b
Second Generation		1436 (59%)	595 (52%)	841 (66%)	
Both Generations		501 (21%)	305 (26%)	196 (15%)	

Notes. EPSE, extrapyramidal side effects; SD, standard deviation

^a Two Simple t-test; mean (SD)

^b Pearson's Chi-squared test of independence; n (%)

In bold p passed significance threshold

Table 4.2 GWAS Participants' Demographics concerning EPSE Presence

	N	Overall	EPSE Presence	EPSE Absence	Neff	p-values
UCL	1017		<i>n</i> = 587 (58%)	<i>n</i> = 430 (42%)	983	
Age at assessment	1017	44.80 (12.24)	46.33 (12.10)	42.72 (12.14)		<0.001^a
Age of onset	761	23.34 (8.18)	23.28 (8.07)	23.44 (8.34)		0.795 ^a
Sex	1017					0.145 ^b
Male		735 (72%)	435 (74%)	300 (70%)		
Female		282 (28%)	152 (26%)	130 (30%)		
Antipsychotics	973					<0.001^b
First generation		102 (10%)	77 (14%)	25 (6%)		
Second generation		452 (47%)	208 (37%)	244 (59%)		
Both generations		419 (43%)	274 (49%)	145 (35%)		
Antidepressants	1020	45 (4%)	22 (2%)	23 (2%)		0.276 ^b
PPD diagnosis	742	105 (14%)	60 (8%)	45 (6%)		0.999 ^b
Aberdeen	414		<i>n</i> = 90 (22%)	<i>n</i> = 324 (78%)	282	
Age at assessment	414	44.40 (13.20)	45.70 (13.12)	44.03 (13.21)		0.292 ^a
Age of onset	401	24.08 (8.05)	23.27 (7.76)	24.29 (8.13)		0.293 ^a
Sex	414					0.426 ^b
Male		311 (75%)	71 (79%)	240 (74%)		
Female		103 (25%)	19 (21%)	84 (26%)		
Antipsychotics	414					<0.001^b
First generation		135 (33%)	42 (47%)	93 (29%)		
Second generation		231 (56%)	32 (35%)	199 (61%)		
Both generations		48 (11%)	16 (18%)	32 (10%)		
UKB	507		<i>n</i> = 90 (18%)	<i>n</i> = 417 (82%)	296	
Age at assessment	507	54.32 (8.08)	56.06 (7.61)	53.94 (8.14)		0.020^a
Sex	507					0.322 ^b
Male		335 (66%)	64 (71%)	271 (65%)		
Female		172 (34%)	26 (29%)	146 (35%)		
Antipsychotics	507					<0.001^b
First generation		158 (31%)	52 (58%)	106 (25%)		
Second generation		320 (63%)	28 (31%)	292 (70%)		
Both generations		29 (6%)	10 (11%)	19 (5%)		
Cardiff	533		<i>n</i> = 411 (77%)	<i>n</i> = 122 (12%)	376	
Age at assessment	533	44.30 (11.70)	44.90 (10.90)	42.20 (14.10)		0.057 ^a
Age of onset	508	25.00 (8.80)	24.30 (7.60)	27.50 (11.60)		0.006^a
Sex	533					0.900 ^b
Male		346 (65%)	268 (65%)	78 (64%)		
Female		187 (35%)	143 (35%)	44 (36%)		
Antipsychotics	531					0.100 ^b
First generation		93 (17%)	78 (19%)	15 (12%)		
Second generation		433 (82%)	327 (80%)	106 (88%)		
Both generations		5 (1%)	5 (1%)	0 (0%)		

Notes. EPSE, extrapyramidal side effects; PPD, premorbid personality disorder; SD, standard deviation.

In bold p passed significance threshold

^a Two Simple t-test; mean (SD); ^b Pearson's Chi-squared test of independence; n (%)

Neff, effective sample sizes, calculated as 4/(1/n_cases + n_controls)

4.4.2 GWAS results

For the main meta-analysis including all participants, we did not observe any SNP passing the genome-wide significance threshold at 5×10^{-8} (Table 4.3 and Appendix 4.6). We found no evidence for population inflation across the samples given the lambda value of 1, suggesting the test statistics are not inflated by population stratification or cryptic relatedness (see Appendix 4.7). We observed no evidence for excessive heterogeneity across the samples. The top index SNP rs2709733 (A/G; $Z=4.999$; $p=5.755 \times 10^{-07}$) mapped to a long intergenic non-protein coding RNA, *LINC01162* and its effect was consistent across all cohorts (Table 4.3). The other affiliated protein-coding genes from the suggestive SNP rs11077391 ($p=3.765 \times 10^{-06}$) included *USP36* and *CYTH1*.

For the subset analyses separating FGA and SGA exposures, we did not observe any SNP passing the genome-wide significance threshold (Table 4.3; Appendices 4.8 & 4.10). There was also no evidence for population inflation for these two subsets with lambda values of 1.030 and 0.992 (Appendices 4.9 & 4.11). The top suggestive SNPs from both subsets showed no excessive heterogeneity. The top affiliated genes from the FGA subset such as *NAV2*, *NRG3*, and *LSAMP* have been associated with autism and schizophrenia disorders (Kao et al., 2010; Must et al., 2008; Pretzsch & Ecker, 2023). In addition, one of the top affiliated protein coding genes, *SHISA9* from the SGA subset has shown associations with epilepsy and autism disorder (Pfisterer et al., 2020; Woolf et al., 2023).

1 Among overlapping SNPs at the 10^{-03} level, 1,463 (53%) SNPs in the SGA subset
2 showed concordant effect directions with the FGA subset's ($p=0.004$), whereas 1,345
3 (48%) SNPs in the FGA subset aligned with the SGA subset's SNPs ($p=0.995$). The
4 stronger directional agreement from SGA to FGA may reflect the broader
5 pharmacological coverage of the FGA subset (which included mixed exposures). In
6 contrast, the weaker concordance from FGA to SGA subset may suggest that top-
7 associated SNP effects within the SGA subset may be more distinct given this subset
8 included only exposure to SGA.

Table 4.3 Regions of the Genome showing the Strongest Association Signals with EPSE Presence

Index SNP	Chr	Position	A1/A2	EAF	Z Score	P-values	P-values Het	Match	Weight (Neff)	Mapped Genes
EPSEs with exposure to any antipsychotic										
rs2709733	7	20878995-20955370	A/G	0.452	4.999	5.755×10 ⁻⁰⁷	0.268	++++++	1937	LINC01162
rs12662039	6	99546454-99598593	C/G	0.057	4.948	7.492×10 ⁻⁰⁷	0.461	+++++?	1561	
rs11077391	17	76661207-76789754	A/G	0.311	4.624	3.765×10 ⁻⁰⁶	0.952	+++++?	1561	USP36, CYTH1
rs62530097	9	7590958-7667847	T/G	0.090	4.494	6.978×10 ⁻⁰⁶	0.560	++++++	1937	
EPSEs with exposure to any type of FGA*										
rs2028609	11	19918741-19933123	T/C	0.494	-5.144	2.693×10 ⁻⁰⁷	0.955	--?--	660	NAV2
rs1416851	10	84329093-84380431	T/C	0.233	-4.927	8.365×10 ⁻⁰⁷	0.290	-----	780	NRG3
rs17723244	3	117509984-117822025	A/G	0.721	-4.751	2.022×10 ⁻⁰⁶	0.595	-----	780	LSAMP, LINC03051
rs2840001	3	168714097-168862366	A/G	0.275	4.725	2.306×10 ⁻⁰⁶	0.563	+++++	780	
EPSEs with exposure to SGA only										
rs72800384	10	54799931-54839608	T/C	0.244	4.567	4.937×10 ⁻⁰⁶	0.179	-+++++	1063	CNBD1
rs117545352	8	87786629-87894786	A/G	0.933	-4.522	6.117×10 ⁻⁰⁶	0.272	----+-	1063	
rs4781355	16	13021889-13101555	A/G	0.637	4.509	6.524×10 ⁻⁰⁶	0.776	++++++	1063	

Notes. Index SNP, the single-nucleotide polymorphism with the strongest association in the genomic region and each is independent at $r^2 < 0.1$; Chr, chromosome; Position, the start and end position (UCSC hg19) of the SNP locus where near-by SNPs were clumped to with nominal associations ($p < 0.05$) and LD ($r^2 < 0.1$) within 250-kb windows taking the 1000 genomes project phase 3 EUR as LD reference; A1/A2, effect and alternate allele; EAF, the effect allele frequency based on 1000 genomes EUR; Z-score, the meta-analysis output reference score for the SNP; P-values, the corresponding p-values to the candidate SNP; P-values Het, corresponding p-values to the degree of variability in effect sizes from METAL analysis; Match, the agreement across the six datasets, + means individuals who carry the A1 allele have positive EPSE association, - means negative, ? means missing, the orders are: three sets of GWAS from UCL samples (1) Affymetrix Array, (2) Illumina PsychArray, (3) GSA, Illumina Global Screening Array, (4) Aberdeen samples, (5) UKB samples, and (6) Cardiff samples; Weight, the overall Neff of the sample for the SNP; Mapped genes, the top genes mapped by positional mapping criterion with maximum distance 10kb to the locus position. No SNP passed genome-wide significant threshold. * Includes participants who had also been exposed to SGAs.

4.4.3 EWAS sample demographics

The UCL participants who developed EPSE were younger than the healthy controls in terms of age at assessment (36.90 vs 44.48, $p<0.001$; see Appendix 4.12) and methylation age (39.54 vs 44.08, $p=0.008$). The UCL EPSE cases also had higher ratios of males (81% vs 44%, $p<0.001$). The Aberdeen participants' methylation age (54.29 vs 53.04, $p=0.473$), and males' ratio (70% vs 74%, $p=0.737$) were balanced between the EPSE and the control group.

4.4.4 EWAS meta-analysis results and permutation testing

In our primary within-case analysis including all EPSE samples, the most significant methylated position (DMP) associated with EPSE presence was cg05599348 (3.181×10^{-07}), mapping to *TMSB15B* on chromosome X (hg19 position 103174718). This DMP was also one of the top identified DMPs in the FGA exposure subset (Table 4.4). However, this DMP was only present in the UCL samples while all other top identified DMPs were present in both samples. Thus, its overall signal may have been influenced by the relatively small sample size of the UCL samples. From the FGA/SGA exposure EWAS subsets, we also did not identify any DMP passing the threshold at 1×10^{-07} (Table 4.4).

Comparing EPSE cases with healthy controls, we identified 9 DMPs associated with EPSE presence after controlling for schizophrenia PRS in addition (Table 4.4). Five of these identified DMPs have been implicated by past schizophrenia EWAS meta-analysis, cg12524168, $p=7.61\times 10^{-20}$; cg05419385, $p=3.08\times 10^{-18}$; cg22583147,

1 $p=5.66\times 10^{-22}$; cg12044923, $p=1.32\times 10^{-19}$; and cg20730966, $p=4.90\times 10^{-24}$ (Hannon et
2 al., 2016). The other four DMPs cg14531564, cg20647656, cg12004641, cg22845912,
3 and their affiliated genes *SDF4*, *ANKMY1*, *TNSI*, *SLA* were not identified in past
4 schizophrenia or smoking EWAS (Elliott et al., 2014; Zeilinger et al., 2013).

5 We next examined whether the locations of these 9 DMPs could map to the
6 corresponding GWAS of the same samples or to previously published schizophrenia
7 GWAS. The GWAS summary statistics were first clumped so that multiple non-
8 independent associations were collapsed into single associated loci. None of the
9 identified DMPs were found to be associated with any genome-wide significant loci
10 from past schizophrenia GWAS according to our regional mapping (Appendix 4.13)
11 (Trubetskoy et al., 2022). The SNP rs7622757 within a 250kb window with
12 cg22583147 was closest to genome-wide significance at $p=4.44\times 10^{-07}$ (Appendix 13).

13 Our mapping of the DMPs to the GWAS of associated samples found that
14 cg12044923 was significantly associated (permutation $p=0.010$) with index SNP
15 rs13108591 which had a GWAS p value of 7.482×10^{-05} . cg20647656 was associated
16 (permutation $p=0.030$) with index SNP rs75037293 which had a GWAS p value of
17 2.73×10^{-04} . According to the past schizophrenia GWAS (Trubetskoy et al., 2022), the
18 SNPs rs13108591 (T/C) had p value of 0.761 and rs75037293 (G/C) had p value of
19 0.117 indicating minor relevance to schizophrenia (Trubetskoy et al., 2022). The SNP
20 rs13108591 is located on chr4:5162317 (hg19), mapping to the intron of *STK32B*. SNP
21 rs75037293 is located on chr2:241453995 (hg19) mapping to the intron of *ANKMY1*.

22

Table 4.4 EPSE-associated differentially Methylated Positions

Probe ID	CHR	Position	Methylation difference (%)	SE (%)	P Value	Gene Annotation
EPSEs with exposure to any antipsychotic						
cg05599348	X	103174718	-5.439	0.980	3.181×10^{-07}	<i>TMSB15B</i>
cg07679219	12	77417738	6.742	1.462	3.998×10^{-06}	<i>E2F7</i>
cg06484572	6	41605494	1.686	0.369	4.769×10^{-06}	<i>MDFI</i>
cg25194055	17	8125180	0.527	0.116	5.690×10^{-06}	
cg00145438	13	113105097	-0.991	0.220	6.620×10^{-06}	
cg26912312	20	61274254	-0.452	0.101	6.971×10^{-06}	<i>SLCO4A1</i>
EPSEs with exposure to any type of FGA*						
cg00500167	6	100841663	0.464	0.098	2.093×10^{-06}	<i>SIMI</i>
cg19185544	8	22595422	-1.748	0.371	2.418×10^{-06}	<i>PEBP4</i>
cg05599348	X	103174718	-5.340	1.156	3.854×10^{-06}	<i>TMSB15B</i>
cg05450477	6	20426845	1.654	0.369	7.293×10^{-06}	<i>E2F3</i>
cg26875877	2	133346858	-0.040	0.886	7.742×10^{-06}	<i>GPR39</i>
cg25030888	1	67156909	-3.013	0.692	1.327×10^{-05}	<i>SGIP1</i>
EPSEs with exposure to SGA only						
cg11411904	1	153935719	1.051	0.215	3.341×10^{-06}	<i>SLC39A1</i>
cg02388709	3	4910253	0.959	0.196	3.563×10^{-06}	
cg21130374	21	42734266	-1.636	0.344	5.928×10^{-06}	<i>MX2</i>
cg15977096	8	34857831	-4.410	0.965	1.268×10^{-05}	
cg09583379	16	19133877	-6.745	1.479	1.316×10^{-05}	
cg23814365	13	36429936	1.979	0.436	1.419×10^{-05}	<i>DCLK1</i>
EPSEs with exposure to any antipsychotic compared to healthy controls						
cg14531564	1	1154853	2.949	0.423	3.073×10^{-12}	<i>SDF4</i>
cg20647656	2	241439612	-1.413	0.236	2.098×10^{-09}	<i>ANKMY1</i>
cg12524168	5	76028910	1.931	0.326	3.207×10^{-09}	<i>F2R</i>
cg12004641	2	218750749	1.784	0.320	2.377×10^{-08}	<i>TNSI</i>
cg05419385	12	27352945	1.565	0.281	2.549×10^{-08}	
cg22583147	3	44331824	2.055	0.374	3.828×10^{-08}	<i>TOPAZ1</i>
cg22845912	8	134059874	1.861	0.341	4.858×10^{-08}	<i>SLA</i>
cg20730966	3	33095886	1.736	0.323	7.442×10^{-08}	<i>GLB1</i>
cg12044923	4	5207312	1.695	0.316	8.414×10^{-08}	<i>STK32B</i>

Notes. Listed are all differentially methylated positions (DMPs) associated with different sets of EPSE samples. Sample sizes were: (1) 314 (UCL 64 EPSE cases, 33 EPSE controls; Aberdeen 47 EPSE cases, 170 EPSE controls); (2) 174 (UCL EPSE 57 cases, 23 EPSE controls; Aberdeen EPSE 30 cases, 64 controls); (3) 123 (UCL only 17 samples in total thus excluded; Aberdeen 17 EPSE cases, 106 EPSE controls); and (4) 859 (UCL 64 EPSE cases, 315 healthy controls; Aberdeen 47 cases, 433 healthy controls). Results came from fixed model meta-analysis adjusted for participants' methylation age, sex, and cell compositions. The comparison between EPSE cases and healthy controls adjusted for schizophrenia polygenic risk scores in addition to alleviate schizophrenia genetic risks. Positions are in hg19. Genes in bold had p meeting threshold at 1×10^{-07} . * Includes participants who had also been exposed to SGAs.

1 **4.4.5 PRS results**

2 We found no evidence to suggest any of the selected PRS could predict the
3 development of EPSE (Table 4.5). According to the fixed model meta-analysis, the
4 participants' genetic predisposition to Schizophrenia ($p=0.566$), Parkinson's disease
5 ($p=0.492$), and Lewy-body dementia ($p=0.765$) were not associated with the presence
6 of EPSE.

Table 4.5 Results of Multiple Regression Analyses

Variables	Estimated Coefficient	Standard Error	Confidence Intervals (95%)	Reference Values	<i>p</i>
Schizophrenia PRS					
UCL	0.013	0.017	-0.020, 0.047	0.780	0.436
Aberdeen	0.006	0.020	-0.033, 0.046	0.321	0.749
UKB	0.032	0.017	-0.036, 0.032	1.941	0.053
Fixed-effect model	0.019	0.010	-0.001, 0.039	1.828	0.566
Parkinson's disease PRS					
UCL	0.015	0.017	-0.018, 0.049	0.892	0.373
Aberdeen	0.025	0.022	-0.017, 0.068	1.162	0.246
UKB	-0.010	0.016	-0.042, 0.022	-0.637	0.525
Fixed-effect model	0.007	0.010	-0.013, 0.027	0.687	0.492
Lewy-body dementia PRS					
UCL	-0.022	0.016	-0.053, 0.009	-1.390	0.165
Aberdeen	0.017	0.023	-0.029, 0.062	0.718	0.473
UKB	0.026	0.017	-0.008, 0.060	1.481	0.139
Fixed-effect model	0.003	0.011	-0.017, 0.024	0.299	0.765

Notes. PRS, polygenic risk scores

All results were adjusted for participants' age, sex, genotyping chip-type, and the first three principal components from GWAS population stratification. UKB results were adjusted for 3 additional principal components. Reference values were t values for individual models and z values for the fixed effect model.

4.5 Discussion

In the present study, we report the largest GWAS meta-analysis of EPSE and the first EWAS meta-analysis of EPSE in European populations to date with exploration on the effects of FGA and SGA exposure. The prevalence of any type of EPSE was found to be 48% among participants who have taken either FGA or SGA. EPSEs were found to be more prevalent among those who had taken FGA or both. No SNP passed the genome-wide threshold of significance. The top index SNP rs2709733 from the GWAS of all antipsychotic exposure mapped to a long intergenic non-protein coding RNA, *LINC01162* with consistent effect across all cohorts. SNPs associated with EPSEs from exposure to FGAs at the suggestive level mapped to *NAV2*, *NRG3*, and *LSAMP*. SNPs associated with EPSEs from exposure to SGA mapped to *SHISA9* and *CNBD1*. The primary EWAS meta-analysis indicated suggestive gene *TMSB15B*, located on chromosome X. In addition, we identified multiple DMPs associated with EPSE passing the significance threshold comparing EPSE cases to healthy controls. The *STK32B* gene which was implicated by methylation probe cg12044923 has been associated with psychiatric and movement disorders. We found no evidence that PRSs for schizophrenia, Parkinson's, and Lewy-body dementia predict EPSE development.

The GWAS meta-analysis results may represent a false negative due to the limited sample size and power. Other factors may also be relevant. For example, our sign tests revealed weak SNP effect alignment between the FGA and the SGA GWAS results, suggesting that there may be differences in the genetic architecture of these traits. Thus, combining participants who have taken either FGA or SGA may have impacted our

1 ability to identify drug-specific genetic risks. Conversely, stratifying the sample by
2 antipsychotic exposure (FGA vs SGA) substantially reduced sample sizes,
3 compromising statistical power. However, several of the implicated genes (*NAV2*,
4 *NRG3*, *LSAMP* and *SHISA9*) from the subset analyses were previously reported to be
5 associated with psychiatric disorders and were and with neuronal function. Thus, these
6 findings warrant cautious interpretation due to the suggestive nature of the associations.
7 The results produced here are the result of a concerted effort to increase sample size as
8 a starting point for future studies.

9 The limited GWAS and PRS findings lead us to speculate that EPSE may be more
10 strongly driven by epigenetic modifications over time. Studies have suggested that
11 methylation changes in dopaminergic or serotonergic pathway genes may impact motor
12 control pathways more dynamically than SNP-based variations (Loke et al., 2015). This
13 dynamic epigenetic regulation aligns with how EPSE can vary significantly among
14 patients and change with continued antipsychotic use, whereas GWAS-derived SNPs
15 only offer a static view of genetic risk. Therefore, integrating EWAS may provide
16 insights into the gene-environment interactions involved in EPSE development.

17 Our primary EWAS analysis of EPSE status may have been again limited by
18 modest sample size, potentially reducing statistical power to detect robust epigenetic
19 associations. However, permutation testing in our expanded EWAS comparing EPSE
20 cases with healthy controls identified two genes, *ANKMY1* and *STK32B*, showing
21 significant relevance to the presence of EPSE. *ANKMY1* encodes the protein Ankyrin
22 Repeat and MYND Domain Containing 1, which has a role for protein-protein

1 interactions and cellular signalling. This could indirectly influence pathways relevant
2 to neurodevelopment or dopamine signalling. However, we have found little additional
3 corroborating evidence directly linking *ANKMY1* to schizophrenia or EPSE. The other
4 implicated gene *STK32B* encodes for a member of the human N-myristoylated proteins,
5 which are involved in various cellular signalling and transduction pathways, although
6 its exact biological function remains insufficiently defined (Takamitsu et al., 2015). A
7 520-kb homozygous deletion encompassing *STK32B* has been described in Ellis-Van-
8 Creveld syndrome, which is a rare genetic disorder that primarily affects the skeletal
9 system and other tissues (Temtamy et al., 2008).

10 Notably, changes in the methylation of the *STK32B* promoter region have been
11 previously linked to both schizophrenia and anxiety disorders. This protein may play a
12 role in executive functions such as working memory and selective attention (Ciuculete
13 et al., 2018; Hannon et al., 2016). Moreover, *STK32B* was implicated in a GWAS of
14 essential tremor (Müller et al., 2016), and patients with essential tremor showed
15 increased expression of *STK32B* in the cerebellar cortex, highlighting a potential
16 relevance to movement abnormalities.

17 The current study has several limitations. First, the study's EPSE classification was
18 based on cross-sectional data from existing studies. Individuals classified as not having
19 EPSE at the time of assessment may develop EPSE later in life with increasing exposure
20 to antipsychotics, introducing potential pseudo-negatives. In addition, we could not
21 differentiate between acute and chronic EPSE. Medication dose information was
22 unavailable for most participants thus, not analysed. We used a mixed definition of

1 EPSE and mixed antipsychotic medications and EPSE medications, each of which may
2 have distinct profiles of EPSE risk. Other dynamic factors such as aging, comorbid
3 conditions and drug-drug interactions may influence the recognition of EPSE as well.
4 Differences in sample ascertainment may have contributed to variability in EPSE
5 detection sensitivity and the predominance of male participants could be another source
6 of bias. These variability and potential miss-classification could impact the consistency
7 of our findings and warrant careful consideration in future studies to clarify the effects
8 of specific antipsychotic medications on EPSE with increased sample size to do so.
9 Finally, although we have implemented strategies to control for collider bias related to
10 schizophrenia, our results may still be influenced by genetic risk to schizophrenia.

11 Overall, our study provides new insights into the biological mechanisms underlying
12 EPSE development in patients with schizophrenia. Notably, our approach integrated
13 findings from EWAS with GWAS results, allowing us to explore EPSE-associated
14 methylation shifts using accessible SNP data. The findings of this study indicate that
15 further investigation of the epigenetics of EPSE and the role of *STK32B* in EPSE is
16 likely to enhance our understanding and inform future research and treatment directions.

5. Polygenic Risk and Cardiovascular Treatment Effects in Severe Mental Illness

5.1 Abstract

Background: Patients with severe mental illness (SMI) face increased cardiovascular risks, leading to reduced life expectancy. Polygenic risk scores (PRS) prediction is a promising method to assess cardiovascular risks in SMI. However, the PRS utility and the impact from interventions on these predictions were underexplored.

Methods: Using samples from the PRIMROSE, which involved longitudinal cardiovascular interventions within primary-care, we calculated 7 cardiovascular and 2 psychiatric PRS (bipolar/schizophrenia) to predict 7 corresponding cardiovascular measures (total cholesterol/HDL/LDL/triglyceride/systolic blood pressure/diastolic blood pressure/BMI) assessed at baseline and 12-month follow-up. We applied multiple linear regression models and explored the interactions between cardiovascular and psychiatric PRS on treatment outcomes.

Results: At baseline, most PRS were predictive of corresponding measures, indicating strong genetic associations. However, these associations could attenuate if the treatment effectively altered the measures, particularly for total cholesterol and systolic blood pressure. LDL measures became negatively associated with bipolar PRS after treatment, though no significant interaction effects were found. Participants in the highest bipolar PRS quartile showed 0.58 mmol/L reduction in LDL measurement on average after treatments suggesting a potential protective effect from higher bipolar genetic risks. These results were robust to potential power reduction, participants' age, sex,

1 medication use, smoking habits, alcohol consumption, and physical activity levels.
2 *Conclusions:* Our findings underscored the dynamic interplay between genetic risks
3 and treatment effects and warrant careful PRS assessment timing. While the clinical
4 application of PRS has not yet been fully realized, further research with precise BD
5 subtype measurements is needed to clarify how BD genetic risks interact with
6 interventions.

7

8 **5.2 Introduction**

9 Patients diagnosed with bipolar disorder (BD), schizophrenia (SCZ), or psychosis
10 are often jointly referred to as patients with severe mental illness (SMI). Patients with
11 SMI have a higher mortality rate leading to a 15–20 years reduction in life expectancy
12 in comparison to the general population (Nordentoft et al., 2013). Cardiovascular
13 disease (CVD) contributes 17.4% and 22.0% of life years lost in males and females
14 with SMI, respectively (Jayatilleke et al., 2017; Nielsen et al., 2021). CVD was
15 recorded as the cause of death for approximately 24% of patients with SCZ and 38% of
16 patients with BD (Nielsen et al., 2013; Westman et al., 2013). The life expectancy gap
17 between the general population and patients with SMI appears to be widening due to
18 the lack of effective interventions (Hayes et al., 2017).

19 Multiple factors underlie the increased risks of cardiovascular events among
20 patients with SMI (Nielsen et al., 2021). Patients with SMI are more likely to have an
21 unhealthy lifestyle, increased sedentary time, lack of exercise, poor diet, and increased
22 rates of smoking and heavy alcohol use (Hartz et al., 2014; Vancampfort et al., 2015,

1 2016, 2017). Treatments with antipsychotic drugs can lead to weight gain and higher
2 cardiovascular mortality (Rotella et al., 2020). In addition, antidepressants and mood
3 stabilizers can cause adverse metabolic effects (Correll et al., 2015; Pérez-Piñar et al.,
4 2016).

5 People with SMI may also have a higher genetic predisposition to CVD compared
6 with the general population (O’Sullivan et al., 2022). A meta-analysis of GWAS and
7 candidate gene studies identified 24 potential pleiotropic genes that are likely to be
8 shared between mood disorders and cardiometabolic disease risk (Amare et al., 2017).
9 So et al. (2019) calculated PRS for SCZ and BD in patients with SMI, summarizing
10 their genetic predispositions to these two disorders. These psychiatric PRS were
11 compared with 28 additional cardiometabolic traits. The results showed that SCZ was
12 genetically associated with several cardiometabolic abnormalities including glucose
13 metabolism abnormalities and adverse adipokine profiles independent of medication
14 use. In contrast, BD showed polygenic associations with an overall more favourable
15 cardiometabolic profile, suggesting potential protective metabolic traits linked to BD
16 genetic risks thus, cardiometabolic abnormalities in BD are more likely to be secondary.
17 In addition, Strawbridge et al. (2021) applied multidimensional scaling to shared
18 genetic variants between both psychiatric and cardiometabolic disorders. They found
19 that schizophrenia patients had distinct metabolic profiles.

20 Various behavioural interventions have been developed to control cardiovascular
21 risks for patients with SMI (Goldfarb et al., 2022). Most of these developed
22 interventions were found to be effective in controlling cardiovascular risks among

1 people with SMI (Aschbrenner et al., 2022; Jakobsen et al., 2017). The added value
2 from incorporating cardiovascular PRS in risk predictions is increasingly recognized
3 (Samani et al., 2024; Sun et al., 2021). Individuals with higher cardiovascular genetic
4 risks were found to benefit more from statin and alirocumab treatment suggesting
5 different response profiles (Damask et al., 2020; Natarajan et al., 2017). However,
6 genomic and clinical risk factor predictions for CVD may vary over the life course
7 (Urbut et al., 2025). In addition, no previous studies have specifically examined how
8 interventions or treatments can influence PRS-based predictions of cardiovascular risks
9 in patients with SMI over time. Additionally, the impact of genetic risk for psychiatric
10 disorders on cardiovascular treatment effectiveness remains unclear.

11 The PRIMROSE group developed a pragmatic intervention aimed at reducing
12 cardiovascular disease risk factors among people with SMI in primary care in England
13 (Burton et al., 2015). The PRIMROSE intervention had a similar effect on total
14 cholesterol concentration reduction at 12 months as treatment-as-usual groups with
15 decreased costs through decreased psychiatric relapses and readmissions (Osborn et al.,
16 2018). A subset of participants provided consent for genetic data analysis to determine
17 whether their genetic profiles can predict variations in treatment response. We used the
18 data to explore longitudinal changes in PRS-based risk predictions.

19 The overall aim of this study was to use 7 cardiovascular (total
20 cholesterol/HDL/LDL/triglyceride/systolic blood pressure/diastolic blood
21 pressure/BMI) and 2 psychiatric (bipolar/schizophrenia) PRS to understand how
22 genetic risks influence cardiovascular treatment effectiveness. Specifically, we wanted

1 to explore if these PRS predictions would change after treatments and how psychiatric
2 genetic risks modify these predictions. We hypothesized that the cardiovascular PRS
3 would interact with psychiatric PRSs (BD/SCZ) in predicting treatment effectiveness.
4 People with higher genetic risks for both cardiovascular risks and psychiatric disorders
5 were expected to have reduced treatment response given this group of patients' potential
6 adverse metabolic profiles.

7

8 **5.3 Methods**

9 **5.3.1 Participants Selection and Genotyping**

10 Participants were recruited from GP practices across both rural and urban areas of
11 England (see details for participant specification in Osborn et al., 2018). Participants
12 were defined as having increased cardiovascular risk if they were aged 30–75 years,
13 listed on the Quality and Outcomes Framework register for severe mental illness
14 (schizophrenia, bipolar disorder, or other non-organic psychosis), with elevated
15 cholesterol (total ≥ 5.0 mmol/L or total-to-HDL ratio ≥ 4.0) and at least one additional
16 risk factor (hypertension, diabetes, elevated HbA1c, obesity, or smoking). Ethics
17 approval was obtained from the City Road and Hampstead Research Ethics Committee
18 (Reference No: 12/LO/1934, approval granted 10 January 2013). Local NHS approvals
19 were obtained before the start of each recruitment wave. At baseline, the recruited
20 participants were randomized into a PRIMROSE intervention group or treatment as
21 usual group for improving cardiovascular risks. Each treatment lasted for 12 months
22 and there were no major difference found in terms of the participants' treatment
23 response between the two groups (see details in Osborn et al., 2018). A subset of the

participants (N=194/326) consented to provide a saliva sample for genotyping and the conduct of genetic research. These participants were included in the present study.

5.3.2 Study Measures

The participants' total cholesterol concentrations, high-density lipoprotein cholesterol (HDL) and low-density lipoprotein cholesterol (LDL), lipid concentration (triglyceride), two blood pressures (systolic and diastolic), and BMI were included for analyses. These measures were collected at baseline using usual GP practice equipment and procedures and repeatedly taken at 12-month follow-up following the same procedure. Other health and lifestyle measures included medication use in the past 12-month (antipsychotic/antidepressant/mood stabilizer/antihypertensive/other medications); smoking habits (number of cigarettes per day); alcohol drinking habit from Alcohol Use Disorders Identification Test Score (AUDIT-C); and physical activity measures from International Physical Activity Questionnaire (IPAQ). These data were collected either directly from participant interviews, clinical measures or from the GP practice medical records (see details for all measures in Osborn et al., 2018).

5.3.3 Genetic Data Imputation and Quality Control

194 participants from the PRIMROSE gave consent for genetic analyses and provided saliva sample for genotyping. The participants' genetic data were genotyped by UCL Genomics on the Global Screening Array then received stringent quality control (QC; Anderson et al., 2010). The process focused on excluding samples with

1 discordant sex, missing genotype data above 10%, excessive heterozygosity (more than
2 3 standard deviations above the mean) and evidence of relatedness (PIHAT of > 0.2).
3 SNPs which deviated substantially from the Hardy-Weinberg equilibrium ($P < 10^{-6}$) or
4 had a minor allele frequency $< 0.5\%$ were also excluded. All of these described quality
5 control steps were performed using PLINK 1.9 (Chang et al., 2015).

6 Imputation was performed using the Sanger Imputation server. Prior to the upload,
7 the data genotypes were prepared as instructed and checks were performed using the
8 HRC-1000G-check-bim tool Version 4.2.3 (Rayner, W., 2015). SNPs with an
9 MAF > 0.01 were uploaded for imputation, applying the Haplotype Reference
10 Consortium reference panel (release 1.1; McCarthy et al., 2016). Pre-phasing was done
11 with EAGLE v.2.3.3 and imputation was done with PBWT (Durbin, 2014). Post-
12 imputation QC repeated the prior QC procedure and further excluded all SNPs with
13 $info < 0.9$. A total of 188 participants' genetic data survived quality control and had
14 complete datasets.

15

16 **5.3.4 PRS Calculations**

17 Patients' cardiovascular and psychiatric PRS were computed from the imputed
18 genetic data with the PRS-CS-auto method which employs the Bayesian theorem to
19 provide a single score for each sample (Ge et al., 2019). The PRS-CS-auto method was
20 chosen over other methods since it outperformed other existing methods according to
21 the simulation studies (Ge et al., 2019; Pain et al., 2021).

22 The application of the PRS-CS method required a LD reference panel and reference

1 GWAS summary statistic, which are used to infer the posterior effect sizes of SNPs. We
2 chose the European sample from The 1000 Genomes Project Consortium (2010) as our
3 LD reference panel. We used GWAS summary statistics on European samples from the
4 Global Lipids Genetics Consortium (GLGC) for cholesterol and triglyceride PRS
5 calculations (Graham et al., 2021). These included total cholesterol, HDL cholesterol,
6 LDL cholesterol, and triglycerides. The summary statistics for blood pressure
7 (diastolic/systolic) came from Keaton et al. (2024). BMI European GWAS summary
8 statistics came from Yengo et al. (2018). The GWAS summary statistics for BD came
9 from European samples of O'Connell et al. (2025). The summary statistics for
10 schizophrenia came from Trubetskoy et al. (2022). The calculated PRS were
11 standardized with healthy controls before analyses.

12

13 **5.3.5 Statistical analysis**

14 The participants' demographic and summary statistics were first presented in
15 respect to their treatment group for all variables. We conducted t and chi-square tests
16 based on the variable type for both baseline and 12-month follow-up measures to test
17 for group differences. Given we found little evidence for group differences in these
18 measures, we then combined the two groups for the following analyses where the group
19 allocation was included as an additional covariate. Before conducting the analyses, we
20 systematically removed outliers using a 3 SD approach on the raw clinical values of the
21 7 selected measures at baseline and 12-month follow-up to minimize the impact of
22 extreme values.

1 We conducted multiple linear regressions taking the 7 cardiovascular measures at
2 baseline as outcome measures and their associated PRS as predictor for associations
3 prior to any treatment. We conducted the same set of analyses on cardiovascular
4 measures at 12-month follow-up to explore how treatments can influence PRS
5 predictions. To address potential power reduction from participant dropout at follow-
6 up, we calculated and compared the power of each baseline and 12-month follow-up
7 regression model using Cohen's f^2 method at 0.05 significance level as a sensitivity
8 check (Cohen, 2013). We then explored the interactions between cardiovascular and
9 psychiatric PRS on the 12-month follow-up measures to test how psychiatric genetic
10 risks can interfere with cardiovascular treatment effectiveness. In addition, we split the
11 BD PRS into 4 quartiles for further analyses and conducted subgroup analyses for the
12 two treatment groups as sensitivity analyses.

13 Covariates in all analyses included the participants' age, sex, treatment allocation,
14 diagnosis, smoking habits, alcohol consumption, and physical activity measures. Each
15 model was also adjusted for the corresponding medication presence recorded at baseline
16 or 12-month assessment including antipsychotic, antidepressant, antihypertensive,
17 mood stabilizers and other medications in binary format. Some of the 12-month
18 analyses also included the corresponding baseline measure as an additional covariate to
19 assess the independence of the genetic prediction. Hypotheses and assumptions for each
20 regression were pre-checked and found to be satisfactory. We conducted sensitivity
21 analyses including only European participants given the PRS were calculated based on
22 GWAS of European ancestry. No significant difference in results were observed given

1 95% of the participants had the same ethnicity.

2 For multiple testing correction, we used the FDR correction methods (Benjamini &
3 Hochberg, 1995). The FDR method was chosen over the others as it gives a good
4 illustration of results and has been applied in previous studies involving PRS
5 (Grigoriu-Serbanescu et al., 2020; Yao et al., 2023). The p value threshold was set at
6 0.05 for adjusted p values. All reported analyses were conducted using Rstudio with R
7 4.3.2 (R Core Team, 2021).

8

9 **5.4 Results**

10 **5.4.1 Sample demographics**

11 Overall, 188 participants' genetic data passed quality control at baseline and had
12 complete datasets. 92 (49%) of them received the PRIMROSE intervention while the
13 rest received treatment as usual which was taken as the control group (See Table 5.1).
14 The whole sample was reasonably balanced in sex (47% males) while the control group
15 had slightly higher ratios of males (53% vs 40%), however, there was no evidence to
16 suggest the imbalance ($p=0.104$). The overall sample contained mostly European (95%)
17 participants and about half of them had diagnoses of bipolar disorder (52%) and were
18 non/ex-smokers (53%). The overall mean AUDIT-C score of 3.52 (SD 3.47) suggested
19 that the participants were generally at a low risk for hazardous drinking or alcohol use
20 disorder. The overall IPAQ total score at 2133 suggested that the participants on average
21 meet recommended activity levels for health. However, the high SD (2249) indicated
22 that the physical activity time varied largely between individuals. The two groups of
23 participants did not differ in ethnicity, diagnoses, smoking/drinking habits, physical

1 activity time or medication uses (see details in Table 5.1). The PRIMROSE intervention
2 group's systolic blood pressure was slightly lower than the control group at the baseline
3 (125.32 vs 130.81, $p=0.027$). The two groups of participants did not differ in other
4 clinical measures including total cholesterol, HDL, LDL, triglycerides, diastolic blood
5 pressure, and BMI.

6 At the 12-month follow-up, only 158 participants had complete dataset for analyses
7 and 77 (49%) of them had the PRIMROSE intervention. We did not observe any
8 significant difference in any of these selected measures between these two groups (See
9 results in Table 5.2). Overall, the interventions were effective in reducing total
10 cholesterol measures ($t=2.277$, $p=0.024$). Such results were consistent with the original
11 PRIMROSE study suggesting that the PRIMROSE intervention had the same level of
12 effect as the treatment as usual. Comparing these measures at the two time points, the
13 participants' overall AUDIT-C score dropped from 3.52 to 3.04 and their IPAQ total
14 score increased from 2133 to 3711. These indicated that the participants had positive
15 changes in some of the life habits in general. The participants' overall systolic blood
16 pressure dropped from 128.12 (SD 17.17) mmHg to 124.36 (SD 13.95) mmHg and their
17 diastolic blood pressure also dropped from 81.68 (SD 10.34) mmHg to 79.68 (9.90)
18 mmHg (see details in Table 5.1 & 5.2). These reductions were statistically significant
19 for systolic blood pressure ($t=2.243$, $p=0.026$) and close for diastolic blood pressure
20 ($t=1.828$, $p=0.068$). These changes further indicated the benefits from both
21 interventions. However, changes in all the other measures were small overall
22 considering both interventions.

Table 5.1 Participant Demographics and Clinical Characteristics by Treatment

Groups at Baseline Assessment

	N	Overall	PRIMROSE	Control	p-values
Overall Samples	188		<i>n</i> = 92 (49%)	<i>n</i> = 96 (51%)	
Age at assessment	188	50.21 (10.22)	50.18 (10.31)	50.23 (10.19)	0.976 ^a
Sex	188				0.104 ^b
Male		88 (47%)	37 (40%)	51 (53%)	
Female		100 (53%)	55 (60%)	45 (47%)	
Ethnicity	188				0.693 ^b
European (white)		178 (95%)	86 (93%)	92 (96%)	
Other		10 (5%)	6 (7%)	4 (4%)	
Diagnosis	188				0.263 ^b
Bipolar		97 (52%)	46 (50%)	51 (53%)	
Schizophrenia		44 (23%)	26 (28%)	18 (19%)	
Other diagnoses		47 (25%)	20 (22%)	27 (28%)	
Smoking	188				0.361 ^b
Heavy (≥ 20 a day)		42 (22%)	24 (26%)	18 (19%)	
Moderate (10-19 a day)		28 (15%)	10 (11%)	18 (19%)	
Light (≤ 9 a day)		19 (10%)	10 (11%)	9 (9%)	
Non/Ex-smoker		99 (53%)	48 (52%)	51 (53%)	
AUDIT-C Score	188	3.52 (3.47)	3.52 (3.41)	3.51 (3.55)	0.982 ^a
IPAQ Total Score	188	2133 (2249)	2059 (2273)	2203 (2236)	0.663 ^a
Medications	188				
Antipsychotic		117 (62%)	54 (59%)	63 (66%)	0.407 ^b
Antidepressant		90 (48%)	45 (49%)	45 (47%)	0.894 ^b
Mood stabiliser		55 (29%)	25 (27%)	30 (31%)	0.650 ^b
Antihypertensive		40 (21%)	17 (18%)	23 (24%)	0.460 ^b
Other Medications		162 (86%)	77 (84%)	85 (89%)	0.453 ^b
Clinical & Blood Tests					
Total cholesterol	187	5.71 (0.85)	5.73 (0.85)	5.68 (0.86)	0.685 ^a
HDL	185	1.28 (0.40)	1.30 (0.38)	1.25 (0.41)	0.405 ^a
LDL	120	3.43 (0.81)	3.49 (0.84)	3.37 (0.77)	0.417 ^a
Triglycerides	127	2.21 (1.22)	2.27 (1.21)	2.14 (1.23)	0.537 ^a
Systolic blood pressure	188	128.12 (17.16)	125.32 (14.74)	130.81 (18.88)	0.027^a
Diastolic blood pressure	186	81.68 (10.34)	81.20 (10.26)	81.68 (10.34)	0.531 ^a
Body mass index (BMI)	185	31.60 (5.35)	31.86 (5.86)	31.34 (4.82)	0.510 ^a

Notes. AUDIT-C, Alcohol Use Disorders Identification Test Score; IPAQ, International Physical Activity Questionnaire; HDL, high-density lipoprotein; LDL, low-density lipoprotein. In bold p met significance threshold at 0.05

Other diagnoses included schizoaffective disorder, psychosis, and persistent delusional disorder.

^a Two Simple t-test; mean (SD); ^b Pearson's Chi-squared test of independence; n (%)

Table 5.2 Participant Demographics and Clinical Characteristics by Treatment Groups at 12 Month Follow-up

	N	Overall	PRIMROSE	Control	p-values
Overall Samples	158		<i>n</i> = 77 (49%)	<i>n</i> = 81 (51%)	
Age at assessment	158	51.30 (9.97)	51.21 (10.05)	51.40 (9.95)	0.907 ^a
Sex	158				0.321 ^b
Male		71 (45%)	31 (40%)	40 (49%)	
Female		87 (55%)	46 (60%)	41 (51%)	
Ethnicity	158				0.999 ^b
White		150 (95%)	73 (95%)	77 (95%)	
Other		8 (5%)	4 (5%)	4 (5%)	
Diagnosis	158				0.435 ^b
Bipolar		85 (54%)	41 (53%)	44 (54%)	
Schizophrenia		35 (22%)	20 (26%)	15 (19%)	
Other diagnoses		38 (24%)	16 (21%)	22 (27%)	
Smoking	158				0.688 ^b
Heavy (≥ 20 a day)		28 (18%)	16 (21%)	12 (15%)	
Moderate (10-19 a day)		25 (16%)	11 (14%)	14 (17%)	
Light (≤ 9 a day)		15 (9%)	6 (8%)	9 (11%)	
Non/Ex-smoker		90 (57%)	44 (57%)	46 (55%)	
AUDIT-C Score	158	3.08 (3.34)	2.82 (3.28)	3.33 (3.39)	0.333 ^a
IPAQ Total Score	158	3694 (5218)	3620 (4515)	3764 (5836)	0.863 ^a
Medications	158				
Antipsychotic		94 (59%)	47 (61%)	47 (58%)	0.823 ^b
Antidepressant		81 (51%)	40 (52%)	41 (51%)	0.994 ^b
Mood stabiliser		40 (25%)	17 (22%)	23 (28%)	0.466 ^b
Antihypertensive		48 (30%)	21 (27%)	27 (33%)	0.513 ^b
Other		144 (91%)	67 (87%)	77 (95%)	0.134 ^b
Clinical & Blood Tests					
Total cholesterol	158	5.46 (1.11)	5.47 (1.11)	5.45 (1.13)	0.888 ^a
HDL	156	1.28 (0.42)	1.28 (0.42)	1.29 (0.42)	0.919 ^a
LDL	94	3.27 (0.98)	3.25 (0.91)	3.29 (1.05)	0.840 ^a
Triglycerides	102	2.20 (1.26)	2.11 (1.38)	2.28 (1.14)	0.502 ^a
Systolic blood pressure	156	124.36 (13.95)	122.78 (13.52)	125.86 (14.28)	0.168 ^a
Diastolic blood pressure	157	79.68 (9.90)	80.01 (9.42)	79.36 (10.39)	0.679 ^a
Body mass index (BMI)	156	31.77 (5.77)	31.94 (6.26)	31.60 (5.30)	0.709 ^a

Notes. AUDIT-C, Alcohol Use Disorders Identification Test Score; IPAQ, International Physical Activity Questionnaire; HDL, high-density lipoprotein; LDL, low-density lipoprotein.

Other diagnoses included schizoaffective disorder, psychosis, and persistent delusional disorder.

^a Two Simple t-test; mean (SD); ^b Pearson's Chi-squared test of independence; n (%)

5.4.2 Cardiovascular PRS Associations Results

The PRIMROSE and control groups were combined for the following analyses with the group allocation as an additional covariate given that we did not find evidence for significant differences in measures. According to our baseline models, most cardiovascular PRSs were predictive of the actual cardiovascular measures among participants with SMI prior to any treatments except for LDL (Table 5.3). Most of the associations remained robust after interventions with only small attenuations in association coefficients (Table 5.3). After adjustment of the corresponding baseline measures, only HDL PRS remained predictive of the actual HDL measures regardless of the treatment. This might suggest that the interventions had little impact on HDL and/or HDL is more genetically modified among patients with SMI.

The interventions were most effective in reducing total cholesterol ($t=2.277$, $p=0.024$) and systolic blood pressure ($t=2.243$, $p=0.026$) measures. The longitudinal comparison revealed a clear attenuation of genetic associations after effective treatment, with the predictive effects of total cholesterol and systolic blood pressure PRS weakening towards the null at the 12-month follow-up. This trend was visually evident in Figure 5.1 (Panels A & E), where the fitted regression lines became notably flatter compared to the baseline. In contrast, measures with minimal changes maintained stable associations, as demonstrated by overlapping or near-parallel fitted lines in Figure 5.1 (Panels B, C, D, F, & G).

The association coefficients for total cholesterol PRS dropped from 0.196 at baseline to 0.155 at 12-month follow-up and the association coefficients for systolic

1 blood pressure PRS dropped from 4.269 at baseline to 2.093 at 12-month follow-up
2 (Table 5.3). Formal comparison of the coefficients between the two time points using
3 Z-score tests revealed no statistically significant differences for total cholesterol
4 ($\Delta=0.041$, $SE=0.113$, $Z=0.362$, $p=0.717$) or systolic blood pressure ($\Delta=2.176$,
5 $SE=1.788$, $Z=1.217$, $p=0.224$). However, even these modest attenuations were
6 sufficient to substantially weaken PRS predictive accuracy following effective
7 treatment interventions (Table 5.3).

8 Our sensitivity analyses comparing the baseline and 12-month regression models
9 revealed no significant difference in model power. Therefore, the observed attenuations
10 were more likely due to changes in participants' actual measures. In addition, our
11 sensitivity analyses separating the PRIMROSE intervention and control group did not
12 produce any major change in the result patterns.

13

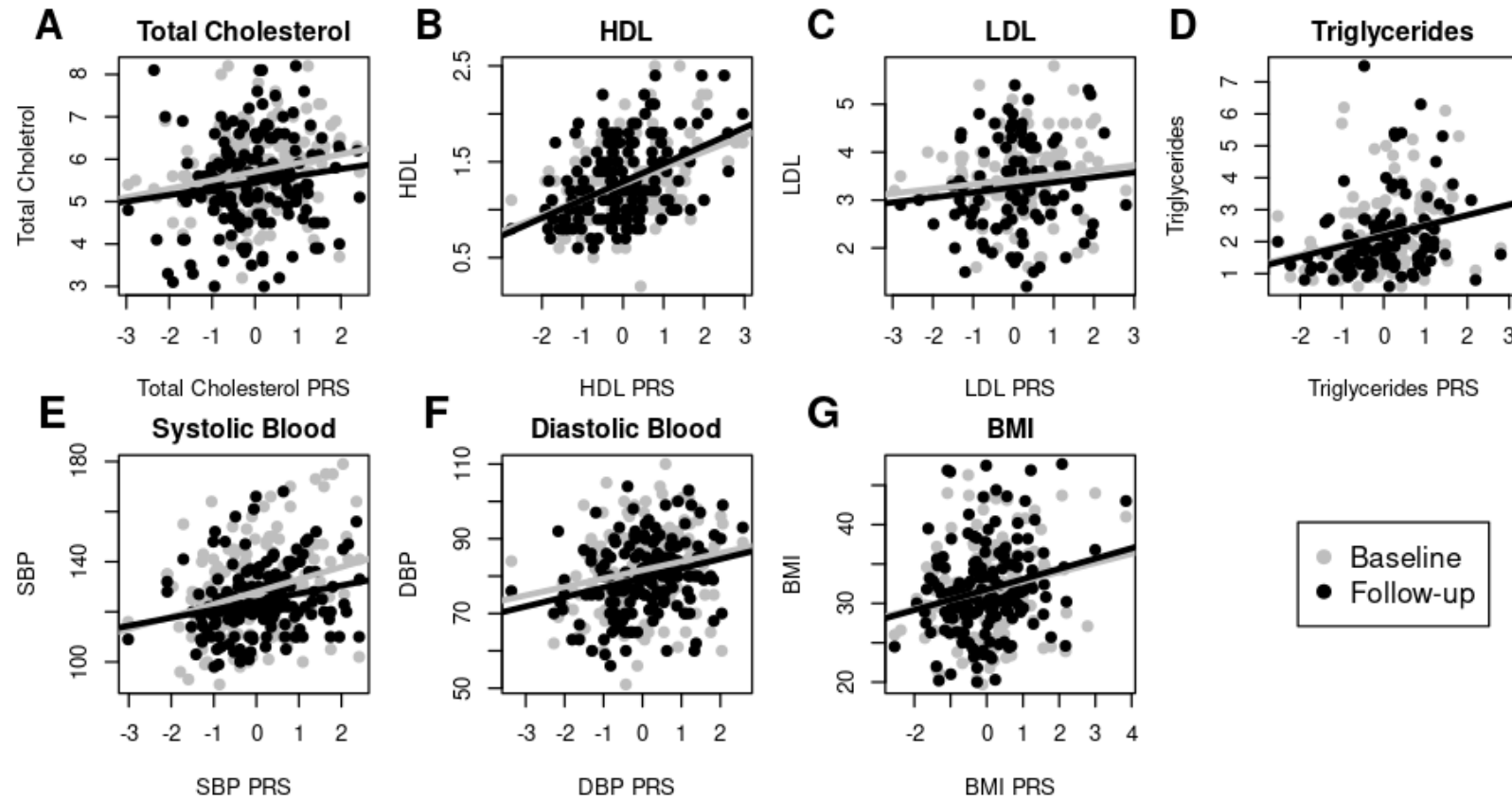
14 **5.4.3 BD & SCZ PRS Interaction Results**

15 We conducted multiple linear regressions taking interactions between the two
16 psychiatric PRS (bipolar/schizophrenia) and each cardiovascular PRS on the
17 corresponding 12-month follow-up measures to explore the impact from psychiatric
18 genetic risk. We first examined potential collinearity between the two psychiatric PRS
19 and each cardiovascular PRS. We only found weak associations between schizophrenia
20 PRS and BMI PRS (coefficient=-0.171; $t=-2.247$, $p=0.026$). However, the model's
21 variables only had the highest Variance Inflation Factor (VIF) number of 2.414
22 suggesting no exceptional collinearity.

1 From the models, we did not find any evidence to suggest that there was any
2 significant interaction effect between BD or SCZ PRS and the corresponding
3 cardiovascular PRS on these clinical measures (See Table 5.4). At 12-month follow-up,
4 total cholesterol PRS was no longer predictive of the actual total cholesterol measure
5 ($p_{adj}=0.166$; Table 5.3). However, for each unit increase in BD PRS, there was a
6 decrease in total cholesterol of 0.276 mmol/L ($p_{adj}=0.043$). In addition, LDL PRS was
7 not predictive of the actual LDL levels at both time points. However, for each unit
8 increase in BD PRS, LDL decreased by 0.331 mmol/L ($p_{adj}=0.043$) at the 12-month
9 follow-up, suggesting a potential protective effect from higher BD genetic risks.

10 To further explore BD PRS's associations with different cholesterol levels, we split
11 BD PRS into 4 quartiles and compared its influence between baseline and 12-month
12 follow-up where the 12-month follow-up models included the corresponding baseline
13 measure as an additional covariate. At baseline, we found no association between BD
14 PRS and any of these cholesterol levels (Figure 5.2 A). However, at 12-month follow-
15 up, the associations became obvious even if controlled for the baseline measure's
16 impact (Figure 5.2 B). If the participants were in the top BD PRS quartile group, they
17 had 0.58 mmol/L fewer LDL measures on average ($t=-0.860$; 95% CI: -1.516, -0.204;
18 $p_{adj}=0.039$). The overall model including all covariates could explain 28% of total
19 variance in LDL measures at 12-month follow-up while BD PRS accounted for 7% of
20 these variations. These findings further indicated BD genetic risk's potential impact on
21 LDL treatment responses. And the relationship between LDL and BD PRS was likely
22 to account for the association of total cholesterol levels and BD PRS.

Figure 5.1 By-group Scatter Plots comparing Baseline and 12-Month Follow-up Clinical Values against Associated PRS



Notes. SBP, systolic blood pressure; DBP, diastolic blood pressure; BMI, body mass index.

Fitted lines came from corresponding linear regressions.

Table 5.3 Results of Multiple Regression Analyses with corresponding Cardiovascular PRS

Variables	Coefficient	SE	t-statistics	CI (95%)	<i>p</i>	<i>p.adj</i>
Total Cholesterol						
Baseline	0.196	0.063	3.116	0.072, 0.319	0.002	0.008
12 Month	0.155	0.094	1.643	-0.032, 0.342	0.103	0.166
12 Month BA	0.001	0.091	0.011	-0.179, 0.181	0.991	0.991
HDL						
Baseline	0.166	0.024	6.822	0.118, 0.214	<0.001	<0.001
12 Month	0.198	0.030	6.604	0.139, 0.257	<0.001	<0.001
12 Month BA	0.056	0.025	2.257	0.007, 0.105	0.026	0.049
LDL						
Baseline	0.084	0.077	1.083	-0.07, 0.237	0.281	0.367
12 Month	0.119	0.117	1.022	-0.114, 0.352	0.311	0.367
12 Month BA	0.037	0.118	0.314	-0.199, 0.273	0.754	0.792
Triglycerides						
Baseline	0.440	0.139	3.171	0.165, 0.715	0.002	0.008
12 Month	0.434	0.163	2.657	0.109, 0.759	0.010	0.025
12 Month BA	0.253	0.175	1.443	-0.098, 0.604	0.154	0.216
Systolic blood pressure						
Baseline	4.269	1.339	3.187	1.624, 6.914	0.002	0.008
12 Month	2.093	1.185	1.765	-0.252, 4.438	0.080	0.140
12 Month BA	1.099	1.089	1.009	-1.055, 3.253	0.315	0.367
Diastolic blood pressure						
Baseline	1.924	0.811	2.373	0.322, 3.525	0.019	0.044
12 Month	1.890	0.829	2.280	0.25, 3.53	0.024	0.049
12 Month BA	1.126	0.745	1.513	-0.347, 2.599	0.133	0.199
Body Mass Index						
Baseline	1.451	0.416	3.486	0.629, 2.273	0.001	0.004
12 Month	1.376	0.500	2.752	0.387, 2.365	0.007	0.020
12 Month BA	0.092	0.243	0.379	-0.389, 0.573	0.705	0.780

Notes. SE, standard errors; CI, confidence interval.

Presented results are outputs from linear regressions with corresponding cardiovascular polygenic risk score regressed on each measure. Participants' sex, age, treatment allocation, diagnosis, smoking scores, alcohol scores, IPAQ scores and the first ten principal components from population stratification were added as covariates to all models. Each model was also adjusted for the corresponding medication recorded at baseline or 12-month assessment including antipsychotic, antidepressant, antihypertensive, mood stabilizers and other medications. 12 Week BA (baseline adjusted) models were further adjusted for the corresponding baseline measure.

P.adj are p values corrected using FDR method. P values in bold are smaller than the threshold at 0.05.

Table 5.4 Adjusted Results of Cardiovascular and Psychiatric PRS Interactions

Variables	Coefficient	SE	t-statistics	CI (95%)	p	p.adj
Total Cholesterol						
BD PRS	-0.276	0.091	-3.016	-0.457, -0.095	0.003	0.043
BD PRS Interaction	0.138	0.097	1.422	-0.054, 0.331	0.157	0.670
SCZ PRS	-0.210	0.099	-2.117	-0.407, -0.014	0.036	0.254
SCZ PRS Interaction	0.068	0.098	0.693	-0.126, 0.262	0.490	0.940
HDL						
BD PRS	0.004	0.030	0.139	-0.055, 0.063	0.889	0.940
BD PRS Interaction	0.012	0.035	-0.352	-0.082, 0.057	0.726	0.940
SCZ PRS	-0.033	0.032	-1.028	-0.096, 0.03	0.306	0.804
SCZ PRS Interaction	-0.003	0.036	-0.075	-0.074, 0.069	0.940	0.940
LDL						
BD PRS	-0.331	0.104	-3.195	-0.539, -0.124	0.002	0.043
BD PRS Interaction	0.106	0.105	1.011	-0.103, 0.315	0.316	0.804
SCZ PRS	-0.212	0.122	-1.738	-0.456, 0.032	0.087	0.487
SCZ PRS Interaction	0.038	0.114	0.335	-0.189, 0.266	0.739	0.940
Triglycerides						
BD PRS	-0.190	0.136	-1.395	-0.461, 0.081	0.167	0.670
BD PRS Interaction	-0.093	0.156	-0.597	-0.405, 0.218	0.553	0.940
SCZ PRS	-0.161	0.149	-1.082	-0.457, 0.135	0.283	0.804
SCZ PRS Interaction	0.032	0.161	0.199	-0.29, 0.354	0.842	0.940
Systolic blood pressure						
BD PRS	1.189	1.086	1.094	-0.961, 3.338	0.276	0.804
BD PRS Interaction	-2.306	1.065	-2.165	-4.414, -0.198	0.032	0.254
SCZ PRS	-0.536	1.263	-0.424	-3.035, 1.963	0.672	0.940
SCZ PRS Interaction	0.164	0.979	0.168	-1.772, 2.101	0.867	0.940
Diastolic blood pressure						
BD PRS	-0.327	0.830	-0.394	-1.969, 1.314	0.694	0.940
BD PRS Interaction	-0.496	0.818	-0.607	-2.114, 1.122	0.545	0.940
SCZ PRS	-0.603	0.925	-0.651	-2.433, 1.228	0.516	0.940
SCZ PRS Interaction	-0.194	0.811	-0.239	-1.799, 1.411	0.812	0.940
Body Mass Index						
BD PRS	0.319	0.466	0.684	-0.603, 1.241	0.495	0.940
BD PRS Interaction	-0.049	0.505	-0.097	-1.048, 0.95	0.923	0.940
SCZ PRS	-0.063	0.520	-0.121	-1.091, 0.965	0.904	0.940
SCZ PRS Interaction	-0.057	0.460	-0.124	-0.968, 0.853	0.901	0.940

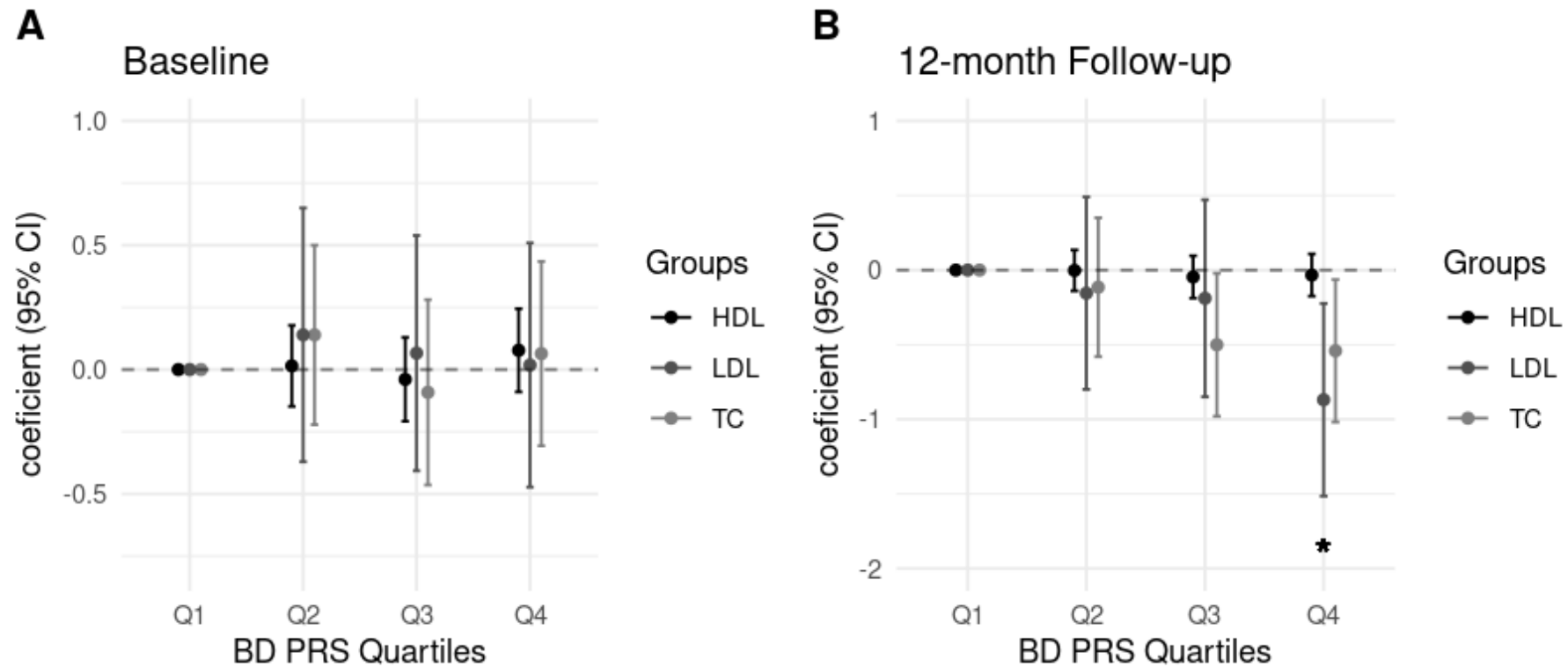
Notes. SE, standard errors; CI, confidence interval; SCZ, schizophrenia; BD, bipolar disorder

Presented results are outputs from linear regressions with corresponding cardiovascular polygenic risk scores (PRS) and their interactions with BD or SCZ PRS regressed on each measure.

Adjustments are the same as regressions in Table 3.

P.adj are p values corrected using FDR method. P values in bold are smaller than threshold at 0.05.

Figure 5.2 Comparison of Baseline and 12-Month Follow-up Cholesterol Levels Across BD PRS Quartiles



5.5 Discussion

In summary, we used multiple cardiovascular and psychiatric PRS to investigate how different genetic risks can impact cardiovascular behavioural treatment responses in patients with SMI within primary care. Prior to treatment, most of these selected PRSs were predictive of the actual cardiovascular measures. However, if the interventions were effective in reducing the measures, significant attenuations in the associations would be expected. For example, the total cholesterol and systolic blood pressure PRS became non-predictive at the 12-month follow-up. The interventions had little effect on HDL measures, suggesting that the HDL PRS may be independent of the baseline measures. The LDL measures were not associated with LDL PRS at both time points but became negatively associated with BD PRS after treatments even though we found no significant interaction effects. These findings collectively suggest that PRS predictions could be influenced by environmental interventions, such as treatment, particularly if the treatment was effective.

The findings of the study may provide valuable insights into the potential heterogeneity within PRS predictions when applied to variable health measures. Our sensitivity analyses comparing baseline and 12-month regression models revealed no significant difference in model power. This suggested that the attenuations in associations were more likely driven by changes in participants' actual measures rather than limitations due to sample drop at follow-up. Most of our selected cardiovascular PRS showed significant association with the corresponding measures prior to the treatments. The observed attenuations in these associations indicated that intervention

1 could play a substantial role in influencing outcomes over time. While individuals'
2 genetic risks always stay stable, the changes in their prediction coefficients could reflect
3 the treatment effects as coefficients represent the strength of associations between
4 predictors and outcomes. For PRS coefficients which mostly remained stable or even
5 increased after treatment such as HDL, this may suggest that the overall treatment likely
6 had little impact. These findings underscored the importance of carefully considering
7 the timing of PRS assessments for accurate risk predictions. Such findings also
8 highlighted the need for more nuanced models that integrate both genetic and
9 environmental factors to better predict individual treatment responses for patients with
10 SMI.

11 For the participants' psychiatric genetic impact on the treatment outcomes, we
12 initially hypothesized that the participants with higher both genetic risks might exhibit
13 reduced treatment effectiveness. However, the results at the 12-month follow-up were
14 in the opposite direction for BD PRS. While it is well-documented that patients with
15 BD can have altered lipid profiles, previous findings have been inconsistent (Hiller et
16 al., 2023). Notably, studies that did find associations often reported a directional trend:
17 depressive episodes tended to correlate with increased lipid levels, whereas manic
18 episodes were more frequently associated with decreased lipid levels (Fusar-Poli et al.,
19 2021; Hiller et al., 2023). Using total cholesterol, triglycerides, LDL, and HDL as
20 biomarkers is inherently complex due to numerous potential confounders, including
21 dietary intake, comorbid somatic conditions, and medication use (Katcher et al., 2009).
22 In this study, no prior association was observed between BD PRS and LDL levels at

1 baseline; however, a negative association with BD PRS emerged after treatment. Again,
2 such findings highlighted the instability of PRS predictions.

3 Notably, BD does not seem to share any genetic correlation with LDL according to
4 the most recent and largest BD GWAS to date (O'Connell et al., 2025). However, a
5 recent comprehensive MR study covering 179 lipid species and 5 psychiatric
6 disorders provided evidence of a potential causal relationship specifically between
7 genetic susceptibility in the plasma lipidome and BD (Yu et al., 2025). The authors
8 identified a protective effect in BD patients, characterized by higher levels of two sterol
9 esters and eight phosphatidylcholines (PCs). These lipids may contribute to regulating
10 neuroinflammation and maintaining neuronal cell membrane stability, potentially
11 explaining their protective role (van der Veen et al., 2017). However, the role of their
12 PCs also appeared dualistic: while some PCs exhibit protective effects, others acted as
13 risk factors to promote the development of BD. Our findings also highlighted the
14 complex relationship between BD and LDL levels. The observed increase in association
15 at follow-up could be partially explained by reductions in other confounding variables,
16 as the participants indeed showed positive changes in some lifestyle and health
17 measures. However, environmental risk factors continue to play a role, and future
18 studies with increased sample size and specific split on BD subtypes may provide
19 further clarity.

20 The study's strengths were evident as we firstly used longitudinal behavioural
21 treatment data to indicate how cardiovascular PRS predictions within patients with SMI
22 could change over time. We employed robust methodologies and utilized the latest

1 GWAS for PRS calculations. The results presented here remained consistent and robust
2 after accounting for the participants' age, sex, group allocation, diagnoses, medication
3 use, smoking habits, alcohol consumption, and physical activity levels. Additionally,
4 we conducted comprehensive sensitivity analyses to validate the results, further
5 reinforcing the credibility and generalizability of our findings. The findings we present
6 here may provide a solid foundation for future studies.

7 The study also had limitations which should be acknowledged. For instance, some
8 of our health measures such as medication use were purely extracted from clinical
9 records which may have been incomplete or inconsistent. The participants in the study
10 had mixed diagnoses and lacked specific symptom measurements thus could not allow
11 us for more in-depth investigations. It is also important to recognize that participants'
12 cardiovascular risk profiles may evolve over time due to a range of external life factors
13 that are independent of the intervention itself. In addition, our samples were
14 predominantly of European ancestry which may have limited generalizability and
15 transferability.

16 In summary, our findings highlighted the instability of PRS for cardiovascular risk
17 predictions within patients with SMI. Researchers should carefully consider the
18 measurement time points as these could produce diverse results. Future studies with
19 increased sample size and more precise symptom measurements on BD subtypes may
20 clarify the complex associations between BD and cardiovascular traits.

6. General discussion

6.1 PhD summary

Overall, this thesis applied different genomic methods to identify potential genomic biomarkers that could guide future mental illness treatments.

The first project (Chapter 2) investigated the interactions between environmental risk factors, ACE and psychiatric genetic risk factors predicted by multiple PRS for different BD phenotype developments. Although we could not find any significant interaction effects, ACE numbers and the PRS were associated with different phenotypes. Participants with ACEs had an earlier age of BD onset and higher odds of having rapid cycling. Higher ADHD PRS correlated with increased ACEs and could increase the likelihood of rapid cycling. Meanwhile, BD PRS was linked to psychotic symptoms and higher ADHD PRS. Thus, the results provided evidence for potential monitoring indexes which may improve phenotype management and guide treatment decisions.

In Chapter 3, I have taken a GWAS meta-analysis approach to identify SNP associated with a positive lithium response among BD patients. The SNP, rs116927879 reached the genome-wide significance level and had a consistent direction of effect from all studies. rs116927879 is located on chromosome 7 and maps to two pseudo-genes *GTF2IP13*, *SEPT7P2*, and the protein coding gene *ADCY1*. By integrating the results with expression data, we found that *ADCY1* plays a role in the regulatory processes in the central nervous system, memory, and learning. The alternative splicing of *ADCY1* could impact different brain regions. We also managed to estimate the SNP

1 heritability (h^2) for subjective lithium response as 20.3% for the first time using
2 genomic data. Thus, the results provided evidence for a potential monitoring gene,
3 *ADCY1* which may guide personalized medication allocation.

4 In Chapter 4, I integrated an EWAS meta-analysis with associated GWAS results to
5 uncover biomarkers linked to EPSE in individuals with long-term exposure to
6 antipsychotic medications. This chapter provided insights into the influence of both
7 genetic and epigenetic factors resulting from environmental treatment factors. Among
8 the 9 identified DMPs associated with EPSE, four of them cg14531564, cg20647656,
9 cg12004641, cg22845912, and their affiliated genes (*SDF4*, *ANKMY1*, *TNSI*, *SLA*)
10 were associated with the risk of developing EPSE and not with schizophrenia or
11 smoking risk. One DMP (cg12044923) which maps to the *STK32B* gene, showed
12 significant enrichment for association with the risk of EPSE and plays a role in tremor
13 development. The results highlighted the importance of exploring both methylation
14 shifts and common SNP associations for medication-induced side-effects.

15 In Chapter 5, I explored how cardiovascular and psychiatric PRS predictions can
16 reflect treatment response using samples from the PRIMROSE trial which included
17 participants with SMI receiving longitudinal cardiovascular treatments. At baseline,
18 most cardiovascular PRS were predictive of corresponding measures. At 12-month
19 follow-up, participants' total cholesterol and systolic blood pressure measures
20 improved; however, the PRS associations weakened. LDL measures became negatively
21 associated with BD PRS after treatment, though no significant interaction effects were
22 found. Participants in the highest BD PRS quartile had 0.58 mmol/L lower LDL on

1 average after treatments suggesting a potential protective effect from higher BD genetic
2 risks. The results showcased the insatiability of PRS predictions, underscored the
3 dynamic interplay between genetic risks and treatment effects, and warrant careful PRS
4 assessment timing for predictions in the future.

5 This thesis concludes in this chapter where I discuss potential limitations inherent
6 in the datasets and used methods, which may impact the accuracy and generalizability
7 of the results.

8

9 **6.2 General limitations**

10 **6.2.1 Limitations in measurements**

11 In the first three projects, the main measures were collected retrospectively through
12 self-reports and clinician interviews. In the reporting of ACE, it has been found that the
13 participants' current mood can impact recall (Hosang et al., 2023). Specifically,
14 participants who reported abuse had higher depression and mania scores compared to
15 those without such reports. The report of ACE may also relate to how individuals
16 process and integrate adverse experiences across development (Baldwin et al., 2019).
17 For medication responses, patients may struggle to accurately remember past treatment
18 courses when doing the assessments. Patients with SCZ and BD can experience
19 episodic or fluctuating symptoms (Perlis et al., 2010). The participants' subjective
20 perception of benefit or side effects may be distorted by current mood state or long-
21 term cognitive biases. For example, it has been shown that patients can possess a
22 "resilience"-like period before realizing the improvements from antidepressants
23 (Stassen et al., 2007). Thus, using retrospective recall for measurements is prone to

1 recall biases and may introduce inaccuracies.

2 In addition, the way in which the measures were recorded may also introduce
3 inconsistencies. For instance, ACEs can be recorded in a binary format for simplicity.
4 However, past studies have found a graded, dose-response relationship between the
5 number of ACEs and risk for adverse mental and physical outcomes (Felitti et al., 1998).
6 This suggests ACEs should be measured in a total number which assumes equal
7 weighting of all adversities while neglect severity, chronicity, and timing. In contrast,
8 more recent studies showed that individuals with at least four ACEs were at increased
9 risks for all health outcomes compared to those with none, suggesting potential
10 categorical measurement methods (Hughes et al., 2017). Across past studies,
11 considerable heterogeneity (I^2 of >75%) has been observed in past ACE studies,
12 particularly due to differences in how ACEs were defined and measured. Nowadays,
13 how to best define and measure ACEs is still under debate. Further research is needed
14 to quantify how each ACE should be weighted and how they cluster together (Lacey &
15 Minnis, 2020).

16 Similar heterogeneity exists in the measurement of medication response. Response
17 to medication can be treated as a binary variable for simplicity, yet it can be difficult to
18 determine the threshold at which a person is consider as a good responder. In contrast,
19 response can be measured as a continuous or categorical variable, ranging from
20 excellent responders, to medium responders and non-responders. Take lithium as an
21 example, the Alda scale is widely regarded as the gold standard for retrospective
22 assessment of lithium response in BD (Grof et al., 2002). It is a two-part scale for

1 continuous response measurements. Part A (0–10) quantifies the degree of clinical
2 improvement during lithium treatment, and part B (0–10) adjusts for confounders (e.g.,
3 adherence, duration of treatment, use of concomitant medications, number of episodes
4 prior to lithium). Researchers have found divergent results when using a binary
5 calcification (a total score ≥ 7 as “responders”) versus taking the Alda scale as a
6 measure (Hou et al., 2016). Such measurement differences can undermine the
7 consistency of study findings, as other past studies defined response in binary formats
8 and did not explore their continuous forms. However, no measure is without limitations,
9 especially for complex traits such as medication response. Another study on Alda scale
10 showed that the part B is vulnerable to error measurement, with some items contributing
11 little yet may impact the overall total score (Scott et al., 2020). Thus, further research
12 is needed to clarify how differences in measurement can impact findings.

13

14 **6.2.2 Limitations in study design**

15 The first three studies used cross-sectional collection methods, which can only
16 provide a partial view compared to cohort designs. Cross-sectional retrospective self-
17 report carries both advantages and limitations. Such measures are relatively
18 inexpensive, scalable, and allow data collection from large samples across the life
19 course (Teicher et al., 2016). However, this design assumes that retrospective reports
20 and prospective measures identify the same, or at least similar, groups of individuals.
21 In reality, taking ACE measurement as an example, the agreement between prospective
22 and retrospective measures of CM is generally low ($k \approx 0.19$), suggesting that these

1 methods often capture different individuals. While prospective measures may capture
2 exposure more objectively, retrospective recall may index subjective appraisal and
3 perceived impact, both of which are relevant, but distinct dimensions of mental health
4 outcomes (Baldwin et al., 2019).

5 Using cross-sectional study designs to assess medication response also be more
6 problematic. Patients with mental illnesses often experience mood transitions from
7 depression to manic or mixed states, many of which are not specific to treatment effects
8 (Perlis et al., 2010). In addition, polypharmacy is common in psychiatric care (Wolff et
9 al., 2021). Patients may frequently switch between antipsychotics or undergo dose
10 titrations due to side effects, partial response, or non-adherence, increasing the risk for
11 adverse effects and drug–drug interactions (Möller et al., 2014; Wolff et al., 2021).
12 Dose effects also accumulate with increasing exposure. Longer cumulative exposure
13 and higher lifetime dose of antipsychotics are associated with increased risk of tardive
14 dyskinesia and parkinsonism (Correll et al., 2017). Furthermore, the pharmacokinetic
15 and pharmacodynamic processes can change with age. Pharmacokinetic changes
16 include reduced renal and hepatic clearance and an increased volume of distribution of
17 lipid soluble drugs, leading to prolonged elimination half-life. Pharmacodynamic
18 changes typically involve increased sensitivity to several drugs classes such as
19 anticoagulants, cardiovascular and psychotropic drugs (Mangoni & Jackson, 2004). For
20 these reasons, many drugs are considered unsuitable for the elderly (Hefner et al., 2021;
21 Motter et al., 2018).

22 Given these complexities, cross-sectional designs may not be optimal. Time-

1 varying models can capture dynamic treatment patterns more accurately than static
2 measures (Leucht et al., 2013). Nonetheless, cross-sectional and retrospective designs
3 continue to dominate large-scale psychiatric genetic research due to feasibility but,
4 findings must still be interpreted with caution.

5

6 **6.2.3 Limitations in sample size**

7 The power of GWAS to detect significant associations depends on multiple
8 parameters, including the minor allele frequency (MAF) of SNPs, the prevalence of the
9 trait, the effect size of genetic variants, and the sample size available (Visscher et al.,
10 2017a). Common variants with higher allele frequencies are generally easier to detect
11 than rare variants of similar effect size. Likewise, traits with higher prevalence provide
12 greater statistical power, as more cases are available for comparison, whereas rare
13 disorders often require very large sample sizes to achieve sufficient power. The increase
14 in sample size has been the foundation for continued and increased discoveries from
15 past GWAS as other factors are largely fixed (Abdellaoui et al., 2023). The conventional
16 GWAS significance threshold at 5×10^{-8} was based on a minimum sample of roughly
17 5,000 cases and 5,000 controls, with SNPs having MAF of at least 5% in European
18 population (Hoggart et al., 2008). Now, take MDD as an example, if MDD has a
19 heritability of 0.3 with 3,000 independent significant SNPs, the sample size required to
20 detect at least 95% of them is estimated to be a minimum of 7.56×10^7 sample (Wu et
21 al., 2022). Comparing these numbers to our sample size of only a few thousands, it is
22 evident that the reported samples are only the starting point for future more powered

and robust findings on the traits.

The limitations in sample also apply to PRS-based predictions. Typically, PRS studies test the associations between PRS and different traits in the target data and evaluate its potential effect. The association can be quantified using standard association or goodness-of-fit metrics such as the proportion of phenotypic variance explained (R^2). For BD, the latest PRS calculated from PRS-CS was estimated to explain a median of approximately 7.4% variance pooling from all European cohorts (O’Connell et al., 2025). For SCZ, the authors conducted leave-one-sample out PRS analyses using different p thresholds for PRS constructions (Trubetskoy et al., 2022). At a 0.05 threshold, which maximises out-of-sample prediction, the median variance in PRS liability explained was found to be 7.3%. If both testing and prediction are conducted within a single sample, allocating approximately 2000 cases and 2000 controls to the replication set is recommended to achieve reliable predictive accuracy (Dudbridge, 2013). Our PRS analyses did not achieve these sample sizes, limiting predictive performance and the ability to fully capture the PRS’s potential. Moreover, even if sufficient sample sizes were reached, the variance explained would remain relatively low, particularly given that recent psychiatric GWAS with hundreds of thousands of participants report modest R^2 estimates.

6.3 Genomic method limitations

6.3.1 GWAS limitations

GWAS is a powerful method in extend to family-based linkage studies which can provide valuable genetic insights into complex psychiatric disorders. Findings from

1 past psychiatric GWAS have advanced our understanding of the aetiology of many
2 different disorders and these findings continue to guide future research and medication
3 developments (O'Connell et al., 2025). However, due to the polygenic nature of
4 psychiatric disorders, GWAS often identify variants with small effect sizes. These
5 identified loci may only explain a tiny fraction of the heritability and a small minority
6 of the inferred genetic variance (Manolio et al., 2009). This is known as the "missing
7 heritability" problem. For instance, BD is commonly estimated to have a 60% to 80%
8 heritability from twin studies (Johansson et al., 2019; McGuffin et al., 2003). However,
9 the most recent PGC BD multi-ancestry GWAS (excluding self-reported data), which
10 explained the most genetic variance, could still only explain 9% of total genetic
11 variance while other meta-analyses only explain even less variance (O'Connell et al.,
12 2025). Such statistic is already coming from the largest study to date with 158,036
13 bipolar disorder cases and 2.8 million controls. Comparing these numbers with our
14 current lithium response and EPSE GWAS meta-analysis sample sizes, we could
15 already expect potential false positives, lack of power, and limited transferability (M. I.
16 McCarthy et al., 2008). Thus, applying GWAS to study psychiatric medication response
17 is still in its early stages. The data we produced here could only act as a starting point
18 for future studies to increase the sample size for more meaningful and insightful
19 findings.

20 Part of the reason why GWAS findings often had modest effect sizes was because
21 GWAS intends to primarily focus on common genetic variants (with minor allele
22 frequency > 5%). Rare variants, which may have larger effects, are often missing or

1 intentionally removed (Gibson, 2012). In the two included GWAS meta-analyses, we
2 also removed rare variants during the genetic data quality control steps leaving effects
3 from these variants as unknown. In addition, our analyses also did not include the sex
4 chromosomes due to complexities (e.g., dosage compensation, and hemizyosity in
5 males, Lee et al., 2019). Although we observed potential sexual differences in lithium
6 response which may yield findings from these sex chromosomes, we could not
7 investigate these further.

8 The interpretations of GWAS findings could be another challenge. For instance, the
9 identified loci could fall outside protein coding regions. Meanwhile, the identified loci
10 may not provide direct information about the biological mechanisms underlying these
11 associations. Follow-up functional studies are required (Visscher et al., 2017b).
12 Because of LD between SNPs, identifying the most plausible causal SNP can be another
13 challenge. Fine-mapping is required to pinpoint causal variants (Schork et al., 2013).
14 However, as shown in our lithium response GWAS meta-analysis, even with one of the
15 most cutting-edge fine-mapping methods Polyfun+SuSiE, pinpointing the exact causal
16 SNP failed due to limited sample size and power (Weissbrod et al., 2020). Building on
17 the LD problem, different ancestry groups may also have ancestry-specific patterns
18 however, all samples reported in this thesis are of European ancestry with limited
19 generalizability to other populations (Price et al., 2006). This is also the scenario for
20 most reported GWAS to date (Martin et al., 2019; Sirugo et al., 2019).

6.3.2 PRS limitations

In the past decade, the numbers of PRS studies increased substantially due to the increasing number of available GWAS on different traits and diseases (Visscher et al., 2017b). PRS which provides a summary of an individual's genetic predisposition, is a promising tool for risk predictions and have been found to be associated with many heritable traits. Notably, all studies we presented here involved PRS predictions including phenotype development and treatment responses predictions. However, as we have also shown in the last project (Chapter 5), PRS predictions can be unstable.

In addition, the generation of PRS relies on the reference GWAS. As a result, the power of PRS is largely influenced by the reference GWAS quality, sample size and population structure (Choi et al., 2020). Most GWAS only focused on common variants, excluding rare variants, and used imputed data which did not capture the dosage from all SNPs for cost-effectiveness reasons. If genetic effects from GWAS could be estimated without error, and these effects were accurately aggregated by PRS, the variance in phenotype explained by PRS would be expected to equal to the SNP heritability (h^2) of the trait. However, this is often not the case in reality. With these limitations in data, PRS often only account for only a small fraction of the heritability of complex traits or diseases (Wray et al., 2013).

Although many PRS calculation methods now use Bayesian approaches to infer genetic architecture, they still do not account for non-linear interactions between genes. This limitation makes it challenging for PRS to explain more complex traits effectively (Boyle et al., 2017; Ge et al., 2019). PRS also do not account for gene-environment

1 interactions, which can significantly influence disease risk. For example, lifestyle
2 factors like diet or smoking may modify the effect of genetic risk. Additionally, PRS
3 are highly dependent on the ancestry of the training dataset. Scores derived from one
4 population (e.g., European ancestry) often perform poorly in other populations (e.g.,
5 African or Asian ancestry), exacerbating health disparities (Martin et al., 2019). Most
6 GWAS have been conducted in populations of European ancestry, the findings have
7 limited generalizability to other ethnic groups, which further widen health disparities
8 (Martin et al., 2019; Sirugo et al., 2019).

9

10 **6.3.3 EWAS and integration limitations**

11 Given the limitation that GWAS typically do not account for gene-environment
12 interactions, EWAS is a powerful approach to identify traits' or diseases' epigenetic
13 modifications which change over time in response to environmental exposures (Rakyan
14 et al., 2011). In our case, we were interested in side effects from antipsychotic
15 medication use which can accumulate epigenic changes over time. However, EWAS
16 has similar limitation profiles like GWAS as the associations identified may not directly
17 determine mechanisms and establish causality (Relton & Davey Smith, 2012). EWAS
18 results do not provide direct insights into the functional consequences of epigenetic
19 changes. Differences in ancestry or population structure can also confound EWAS
20 results, as epigenetic patterns vary between ethnic groups. The epigenome can also vary
21 across study populations. If any of these differences are linked to the disease of interest,
22 confounding may be introduced (Michels, 2010). Adequately large sample sizes are

1 crucial for ensuring both the validity and reliability of study results. The EWAS we
2 reported here had participants of European ancestry only and may have had limited samples
3 size and power (Michels et al., 2013). Meanwhile, the EWAS was cross-sectional which
4 may not capture dynamic changes over time (Jones et al., 2015).

5 In addition to these, EWAS also suffer from its unique limitations. GWAS can use
6 DNA from most tissue types to identify germline genetic variation, typically extracted
7 from blood or blood-cell-derived cell lines. However, epigenetic changes can be tissue-
8 specific, meaning that findings in one tissue (e.g., blood) may not reflect changes in
9 other tissues (e.g., brain). As most EWAS are conducted with living individuals, DNA
10 is generally limited to easily accessible sources such as blood or saliva. Given our
11 samples' data only came from blood, this may limit the generalizability of our EWAS
12 results to understand mechanisms happening in the brain (Smith et al., 2015).

13 EWAS involve sensitive genetic and epigenetic data, which can also raise concerns
14 about privacy, data security, and potential misuse of information (Heyn & Esteller,
15 2012). Combinations of hypermethylated DNA biomarkers have indicated high
16 sensitivity in detecting cancer cells and predicting tumour progression (Heyn & Esteller,
17 2012). The key to success lies in combinatorial approaches. By integrating GWAS and
18 EWAS, researchers can uncover the functional mechanisms underlying genetic
19 associations and improve the translational potential of the findings. However, for
20 biomarkers to be clinically useful, their detection sensitivity and specificity in
21 biological fluids must be improved in the future.

22

6.4 Clinical implications

Personalized medicine represents one of the most promising frontiers in contemporary medicine, aiming to tailor therapies to individual patients by integrating clinical, genetic, and other molecular predictions. A patient's DNA only needs to be collected once, but it can be reanalysed over time to identify newly discovered risk genes and to calculate updated polygenic risk scores (PRS), making it a valuable and enduring resource in precision care.

GWAS findings have multiple clinical implications to strengthen future treatments. For instance, the genetic variants identified through medication response studies can help guide personalized prescriptions (Giacomini et al., 2017). Treatments could be prioritized for patients with high expression of genes associated with favourable response and reconsidered for those at high risk of adverse effects. The findings may also contribute to the development of risk prediction algorithms, enabling earlier identification of individuals at heightened risk for poor treatment response or adverse effects to facilitate more personalized intervention strategies in a timely manner. In addition, insights into the underlying medication biological pathways for good response can support the development of targeted medications to maximize treatment while minimize potential side-effects. Drugs with supporting genetic evidence are more likely to succeed in the development pipeline (M. R. Nelson et al., 2015).

PRS predictions also have multiple important clinical implications, especially for risk predictions. They may be most useful in cohorts with a higher prior probability of disease, where they can assist diagnosis or to inform treatment choices (C. M. Lewis &

1 Vassos, 2020). Our findings from the last study further suggest that the timing of PRS
2 assessment is critical as predilections may be more informative at specific stages of
3 disease progression. Determining the optimal time to collect or apply this genetic
4 information remains an important area for future research. Nevertheless, incorporating
5 PRS into clinical workflows could enhance early intervention strategies, support
6 personalized treatment planning, and ultimately improve patient outcomes.

7

8 **6.5 Future directions**

9 There are many ways future studies can improve on the current psychiatric
10 treatment response GWAS. One option is to promote global collaboration to increase
11 sample sizes and cover multi-ancestry groups. Psychiatric genetics benefited hugely
12 through global collaborations as also discussed in Chapter 1 (O'Connell et al., 2025).
13 Another option could be conducting a multi-trait analysis of GWAS (MTAG) which
14 combines GWAS summary statistics of related traits using multi-trait analysis tools. By
15 analysing schizophrenia and lithium response GWAS summary statistics together as a
16 cross-trait meta-GWAS, ConLi+Gen (2018) identified 15 genetic loci that may have
17 overlapping effects on lithium treatment response and susceptibility to SCZ. In addition,
18 future research could also update genotyping arrays to include rare and structural
19 variants and employ whole-genome/exome sequencing data for more comprehensive
20 analyses. Large samples can be linked with electronic health records to capture more
21 comprehensive treatment histories than using cross-sectional collection methods alone.

22 PRS predictions are not yet widely implemented in clinical settings due to several

1 challenges, including risk prediction instability, limited actionable intervention plans,
2 and ethical concerns such as information privacy, discrimination, and stigmatization (A.
3 C. F. Lewis & Green, 2021; Torkamani et al., 2018). Looking ahead, as GWAS sample
4 sizes continue to grow and genotyping methods continue to advance, PRS are expected
5 to play an increasingly central role in biomedical research and personalized medicine.
6 However, their successful translation into clinical practice will depend on
7 advancements in methodological development, accurate analysis, appropriate
8 interpretation, and a thorough understanding of their strengths and limitations (Massi
9 et al., 2023).

10 Even if all risk DNA variants were identified with perfect accuracy, imperfect
11 predictions from genetics are expected. This is because genetic factors are not the only
12 risk factors for disorders (Wray et al., 2021). Predictions can only be improved if
13 combined with other clinical risk factors, such as sex, age, and medical histories
14 (Hippisley-Cox et al., 2017). While no single factor is a strong predictor on its own,
15 their combination can meaningfully guide clinical decision-making. Future progress
16 will require developing new approaches to conceptualising and quantifying
17 environmental risks, such as ACE, to enable more comprehensive models of risk
18 prediction. In addition, future studies should investigate the extent to which
19 environmental or cultural differences between populations influence specific traits
20 (Abdellaoui et al., 2023).

1 **6.6 Summary and conclusions**

2 An individual's response to treatment is diverse and influenced by multiple factors.
3 By incorporating various genomic research methods combined with environmental risk
4 factors, projects included in this thesis tried to identify potential biomarkers that
5 differentiate mental illness treatment responses. These findings highlight the need for
6 further investigations to validate and expand upon the results obtained. Future research
7 should involve larger sample sizes, meta-analyses with diverse ancestral groups, and
8 the integration of advanced methods and technologies. These efforts will help the field
9 move towards a more comprehensive understanding of the genetic underpinnings of
10 mental illness treatment response and pave the way for the development of more
11 effective prevention and more personalized treatment strategies.

References

- 1000 Genomes Project Consortium, Auton, A., Brooks, L. D., Durbin, R. M., Garrison, E. P., Kang, H. M., Korbel, J. O., Marchini, J. L., McCarthy, S., McVean, G. A., & Abecasis, G. R. (2015). A global reference for human genetic variation. *Nature*, 526(7571), 68–74.
<https://doi.org/10.1038/nature15393>
- Aas, M., Bellivier, F., Bettella, F., Henry, C., Gard, S., Kahn, J.-P., Lagerberg, T. V., Aminoff, S. R., Melle, I., Leboyer, M., Jamain, S., Andreassen, O. A., & Etain, B. (2020). Childhood maltreatment and polygenic risk in bipolar disorders. *Bipolar Disorders*, 22(2), 174–181.
<https://doi.org/10.1111/bdi.12851>
- Abdellaoui, A., Yengo, L., Verweij, K. J. H., & Visscher, P. M. (2023). 15 years of GWAS discovery: Realizing the promise. *The American Journal of Human Genetics*, 110(2), 179–194.
<https://doi.org/10.1016/j.ajhg.2022.12.011>
- Abdelwahab, A. A., Eid, A. H., Balah, A. I., Abd Elraouf, O. M., & Ali, A. (2022). L- Carnitine mitigates infertility induced by lithium and carbamazepine in protein malnourished rats. *Azhar International Journal of Pharmaceutical and Medical Sciences*, 2(2), 116–130.
<https://doi.org/10.21608/aijpms.2022.116887.1107>
- Åberg, K., Adkins, D. E., Bukszár, J., Webb, B. T., Caroff, S. N., Miller, D. D., Sebat, J., Stroup, S., Fanous, A. H., Vladimirov, V. I., McClay, J. L., Lieberman, J. A., Sullivan, P. F., & van den Oord, E. J. C. G. (2010). Genomewide Association Study of Movement-Related Adverse Antipsychotic Effects. *Biological Psychiatry*, 67(3), 279–282.
<https://doi.org/10.1016/j.biopsych.2009.08.036>
- Aedo, A., Murru, A., Sanchez, R., Grande, I., Vieta, E., & Undurraga, J. (2018). Clinical characterization of rapid cycling bipolar disorder: Association with attention deficit

hyperactivity disorder. *Journal of Affective Disorders*, 240, 187–192.

<https://doi.org/10.1016/j.jad.2018.07.051>

Agnew-Blais, J., & Danese, A. (2016). Childhood maltreatment and unfavourable clinical outcomes in bipolar disorder: A systematic review and meta-analysis. *The Lancet Psychiatry*, 3(4), 342–349. [https://doi.org/10.1016/S2215-0366\(15\)00544-1](https://doi.org/10.1016/S2215-0366(15)00544-1)

Ahangari, M., Bustamante, D., Kirkpatrick, R., Nguyen, T.-H., Verrelli, B. C., Fanous, A., Kendler, K. S., Webb, B. T., Bacanu, S.-A., & Riley, B. P. (2022). Relationship between polygenic risk scores and symptom dimensions of schizophrenia and schizotypy in multiplex families with schizophrenia. *The British Journal of Psychiatry*, 1–8. <https://doi.org/10.1192/bjp.2022.179>

Amare, A. T., Schubert, K. O., Hou, L., Clark, S. R., Papiol, S., Cearns, M., Heilbronner, U., Degenhardt, F., Tekola-Ayele, F., Hsu, Y.-H., Shekhtman, T., Adli, M., Akula, N., Akiyama, K., Arda, R., Arias, B., Aubry, J.-M., Backlund, L., Bhattacharjee, A. K., ... Baune, B. T. (2021). Association of polygenic score for major depression with response to lithium in patients with bipolar disorder. *Molecular Psychiatry*, 26(6), Article 6. <https://doi.org/10.1038/s41380-020-0689-5>

Amare, A. T., Schubert, K. O., Klingler-Hoffmann, M., Cohen-Woods, S., & Baune, B. T. (2017). The genetic overlap between mood disorders and cardiometabolic diseases: A systematic review of genome wide and candidate gene studies. *Translational Psychiatry*, 7(1), Article 1. <https://doi.org/10.1038/tp.2016.261>

American Psychiatric Association. (1994). *American Psychiatric Association. Diagnostic and Statistical Manual of Mental Disorders (DSM-IV)*.

- Anand, A., Koller, D. L., Lawson, W. B., Gershon, E. S., & Nurnberger, J. I. (2015). Genetic and childhood trauma interaction effect on age of onset in bipolar disorder: An exploratory analysis. *Journal of Affective Disorders*, 179, 1–5. <https://doi.org/10.1016/j.jad.2015.02.029>
- Anderson, C. A., Pettersson, F. H., Clarke, G. M., Cardon, L. R., Morris, A. P., & Zondervan, K. T. (2010). Data quality control in genetic case-control association studies. *Nature Protocols*, 5(9), Article 9. <https://doi.org/10.1038/nprot.2010.116>
- Andreassen, O. A., Hindley, G. F. L., Frei, O., & Smeland, O. B. (2023). New insights from the last decade of research in psychiatric genetics: Discoveries, challenges and clinical implications. *World Psychiatry*, 22(1), 4–24. <https://doi.org/10.1002/wps.21034>
- Arribas-Ayllon, M., Bartlett, A., & Lewis, J. (2019). *Psychiatric Genetics: From Hereditary Madness to Big Biology*. Routledge. <https://doi.org/10.4324/9781315657967>
- Aschbrenner, K. A., Naslund, J. A., Gorin, A. A., Mueser, K. T., Browne, J., Wolfe, R. S., Xie, H., & Bartels, S. J. (2022). Group Lifestyle Intervention With Mobile Health for Young Adults With Serious Mental Illness: A Randomized Controlled Trial. *Psychiatric Services*, 73(2), 141–148. <https://doi.org/10.1176/appi.ps.202100047>
- Ayano, G., Demelash, S., yohannes, Z., Haile, K., Tulu, M., Assefa, D., Tesfaye, A., Haile, K., Solomon, M., Chaka, A., & Tsegay, L. (2021). Misdiagnosis, detection rate, and associated factors of severe psychiatric disorders in specialized psychiatry centers in Ethiopia. *Annals of General Psychiatry*, 20, 10. <https://doi.org/10.1186/s12991-021-00333-7>
- Baldessarini, R. J., & Tondo, L. (2022). Testing for Antisuicidal Effects of Lithium Treatment. *JAMA Psychiatry*, 79(1), 9–10. <https://doi.org/10.1001/jamapsychiatry.2021.2992>

- Baldwin, J. R., Reuben, A., Newbury, J. B., & Danese, A. (2019). Agreement Between Prospective and Retrospective Measures of Childhood Maltreatment: A Systematic Review and Meta-analysis. *JAMA Psychiatry*, 76(6), 584–593. <https://doi.org/10.1001/jamapsychiatry.2019.0097>
- Baldwin, J. R., Sallis, H. M., Schoeler, T., Taylor, M. J., Kwong, A. S. F., Tielbeek, J. J., Barkhuizen, W., Warrier, V., Howe, L. D., Danese, A., McCrory, E., Rijdsdijk, F., Larsson, H., Lundström, S., Karlsson, R., Lichtenstein, P., Munafò, M., & Pingault, J.-B. (2022). A genetically informed Registered Report on adverse childhood experiences and mental health. *Nature Human Behaviour*, 1–22. <https://doi.org/10.1038/s41562-022-01482-9>
- Barnes, T. R. E. (1989). A Rating Scale for Drug-Induced Akathisia. *The British Journal of Psychiatry*, 154(5), 672–676. <https://doi.org/10.1192/bjp.154.5.672>
- Baumeister, A. A. (2013). The Chlorpromazine Enigma. *Journal of the History of the Neurosciences*. <https://www.tandfonline.com/doi/abs/10.1080/0964704X.2012.664087>
- Benjamini, Y., & Hochberg, Y. (1995). Controlling the False Discovery Rate: A Practical and Powerful Approach to Multiple Testing. *Journal of the Royal Statistical Society: Series B (Methodological)*, 57(1), 289–300. <https://doi.org/10.1111/j.2517-6161.1995.tb02031.x>
- Birnbaum, R. (2014). Variant GADL1 and Response to Lithium in Bipolar I Disorder. *The New England Journal of Medicine*, 370(19), 1855–1860. <https://doi.org/10.1056/NEJMc1401817>
- BMJ Best Practice. (2021). Retrieved 13 September 2021, from <https://bestpractice.bmj.com/>.
- Bousman, C. A., Bengesser, S. A., Aitchison, K. J., Amare, A. T., Aschauer, H., Baune, B. T., Asl, B. B., Bishop, J. R., Burmeister, M., Chaumette, B., Chen, L.-S., Cordner, Z. A., Deckert,

- J., Degenhardt, F., DeLisi, L. E., Folkersen, L., Kennedy, J. L., Klein, T. E., McClay, J. L., ... Müller, D. J. (2021). Review and Consensus on Pharmacogenomic Testing in Psychiatry. *Pharmacopsychiatry*, 54(1), 5–17. <https://doi.org/10.1055/a-1288-1061>
- Boyle, E. A., Li, Y. I., & Pritchard, J. K. (2017). An expanded view of complex traits: From polygenic to omnigenic. *Cell*, 169(7), 1177–1186. <https://doi.org/10.1016/j.cell.2017.05.038>
- Braslow, J. T., & Marder, S. R. (2019). History of Psychopharmacology. *Annual Review of Clinical Psychology*, 15(1), 25–50. <https://doi.org/10.1146/annurev-clinpsy-050718-095514>
- Bråten, L. S., Haslemo, T., Jukic, M. M., Ingelman-Sundberg, M., Molden, E., & Kringen, M. K. (2020). Impact of CYP2C19 genotype on sertraline exposure in 1200 Scandinavian patients. *Neuropsychopharmacology*, 45(3), Article 3. <https://doi.org/10.1038/s41386-019-0554-x>
- Brown, N. M., Brown, S. N., Briggs, R. D., Germán, M., Belamarich, P. F., & Oyeku, S. O. (2017). Associations Between Adverse Childhood Experiences and ADHD Diagnosis and Severity. *Academic Pediatrics*, 17(4), 349–355. <https://doi.org/10.1016/j.acap.2016.08.013>
- Brus, M. J., Solanto, M. V., & Goldberg, J. F. (2014). Adult ADHD vs. Bipolar Disorder in the DSM-5 Era: A Challenging Differentiation for Clinicians. *Journal of Psychiatric Practice®*, 20(6), 428. <https://doi.org/10.1097/01.pra.0000456591.20622.9e>
- Bulik-Sullivan, B. K., Loh, P.-R., Finucane, H. K., Ripke, S., Yang, J., Patterson, N., Daly, M. J., Price, A. L., & Neale, B. M. (2015). LD Score regression distinguishes confounding from polygenicity in genome-wide association studies. *Nature Genetics*, 47(3), Article 3. <https://doi.org/10.1038/ng.3211>
- Burton, A., Osborn, D., Atkins, L., Michie, S., Gray, B., Stevenson, F., Gilbert, H., & Walters, K.

- (2015). Lowering Cardiovascular Disease Risk for People with Severe Mental Illnesses in Primary Care: A Focus Group Study. *PLOS ONE*, 10(8), e0136603. <https://doi.org/10.1371/journal.pone.0136603>
- Campagna, M. P., Xavier, A., Lechner-Scott, J., Maltby, V., Scott, R. J., Butzkueven, H., Jokubaitis, V. G., & Lea, R. A. (2021). Epigenome-wide association studies: Current knowledge, strategies and recommendations. *Clinical Epigenetics*, 13(1), 214. <https://doi.org/10.1186/s13148-021-01200-8>
- Cantor. (2014). *In the Wake of the Plague: The Black Death and the World It Made—Norman F. Cantor—Google Books*. [https://books.google.co.uk/books?hl=en&lr=&id=L3qvBAAQBAJ&oi=fnd&pg=PA1&dq=Cantor,+N.+F.+\(2001\).+In+the+Wake+of+the+Plague:+The+Black+Death+and+the+World+It+Made.+Free+Press&ots=e4dQraDAU8&sig=214r4ZeBE6BbgCcwMukJD91x360#v=onepage&q&f=false](https://books.google.co.uk/books?hl=en&lr=&id=L3qvBAAQBAJ&oi=fnd&pg=PA1&dq=Cantor,+N.+F.+(2001).+In+the+Wake+of+the+Plague:+The+Black+Death+and+the+World+It+Made.+Free+Press&ots=e4dQraDAU8&sig=214r4ZeBE6BbgCcwMukJD91x360#v=onepage&q&f=false)
- Carbon, M., Kane, J. M., Leucht, S., & Correll, C. U. (2018). Tardive dyskinesia risk with first- and second-generation antipsychotics in comparative randomized controlled trials: A meta-analysis. *World Psychiatry*, 17(3), 330–340. <https://doi.org/10.1002/wps.20579>
- Cardno, A. G., & Owen, M. J. (2014). Genetic Relationships Between Schizophrenia, Bipolar Disorder, and Schizoaffective Disorder. *Schizophrenia Bulletin*, 40(3), 504–515. <https://doi.org/10.1093/schbul/sbu016>
- Carroll, L. s., Williams, H. j., Walters, J., Kirov, G., O'Donovan, M. c., & Owen, M. j. (2011). Mutation screening of the 3q29 microdeletion syndrome candidate genes DLG1 and PAK2 in schizophrenia. *American Journal of Medical Genetics Part B: Neuropsychiatric*

- Genetics*, 156(7), 844–849. <https://doi.org/10.1002/ajmg.b.31231>
- Carvalho, A. F., Dimellis, D., Gonda, X., Vieta, E., McIntyre, R. S., & Fountoulakis, K. N. (2014). Rapid Cycling in Bipolar Disorder: A Systematic Review. *The Journal of Clinical Psychiatry*, 75(6), 16864. <https://doi.org/10.4088/JCP.13r08905>
- Carvalho Henriques, B., Yang, E. H., Lapetina, D., Carr, M. S., Yavorsky, V., Hague, J., & Aitchison, K. J. (2020). How Can Drug Metabolism and Transporter Genetics Inform Psychotropic Prescribing? *Frontiers in Genetics*, 11. <https://www.frontiersin.org/articles/10.3389/fgene.2020.491895>
- Chand, S. P., Kuckel, D. P., & Huecker, M. R. (2023). Cognitive Behavior Therapy. In *StatPearls*. StatPearls Publishing. <http://www.ncbi.nlm.nih.gov/books/NBK470241/>
- Chang, C. C., Chow, C. C., Tellier, L. C., Vattikuti, S., Purcell, S. M., & Lee, J. J. (2015). Second-generation PLINK: Rising to the challenge of larger and richer datasets. *GigaScience*, 4(1), s13742-015-0047–0048. <https://doi.org/10.1186/s13742-015-0047-8>
- Charlson, F. J., Ferrari, A. J., Santomauro, D. F., Diminic, S., Stockings, E., Scott, J. G., McGrath, J. J., & Whiteford, H. A. (2018). Global Epidemiology and Burden of Schizophrenia: Findings From the Global Burden of Disease Study 2016. *Schizophrenia Bulletin*, 44(6), 1195–1203. <https://doi.org/10.1093/schbul/sby058>
- Chen, C.-H., Lee, C.-S., Lee, M.-T. M., Ouyang, W.-C., Chen, C.-C., Chong, M.-Y., Wu, J.-Y., Tan, H. K.-L., Lee, Y.-C., Chuo, L.-J., Chiu, N.-Y., Tsang, H.-Y., Chang, T.-J., Lung, F.-W., Chiu, C.-H., Chang, C.-H., Chen, Y.-S., Hou, Y.-M., Chen, C.-C., ... Cheng, A. T.-A. (2014). Variant GADL1 and Response to Lithium Therapy in Bipolar I Disorder. *New England Journal of Medicine*, 370(2), 119–128. <https://doi.org/10.1056/NEJMoa1212444>

- Chen, J., Ding, Q., An, L., & Wang, H. (2022). Ca²⁺-stimulated adenylyl cyclases as therapeutic targets for psychiatric and neurodevelopmental disorders. *Frontiers in Pharmacology*, 13. <https://www.frontiersin.org/articles/10.3389/fphar.2022.949384>
- Chia, R., Sabir, M. S., Bandres-Ciga, S., Saez-Atienzar, S., Reynolds, R. H., Gustavsson, E., Walton, R. L., Ahmed, S., Viollet, C., Ding, J., Makarious, M. B., Diez-Fairen, M., Portley, M. K., Shah, Z., Abramzon, Y., Hernandez, D. G., Blauwendraat, C., Stone, D. J., Eicher, J., ... Scholz, S. W. (2021). Genome sequencing analysis identifies new loci associated with Lewy body dementia and provides insights into its genetic architecture. *Nature Genetics*, 53(3), Article 3. <https://doi.org/10.1038/s41588-021-00785-3>
- Chockalingam, R., Gott, B. M., & Conway, C. R. (2019). Tricyclic Antidepressants and Monoamine Oxidase Inhibitors: Are They Too Old for a New Look? *Handbook of Experimental Pharmacology*, 250, 37–48. https://doi.org/10.1007/164_2018_133
- Choi, S. W., Mak, T. S.-H., & O'Reilly, P. F. (2020). Tutorial: A guide to performing polygenic risk score analyses. *Nature Protocols*, 15(9), 2759–2772. <https://doi.org/10.1038/s41596-020-0353-1>
- Chouinard, G., & Margolese, H. C. (2005). Manual for the Extrapyrarnidal Symptom Rating Scale (ESRS). *Schizophrenia Research*, 76(2–3), 247–265. <https://doi.org/10.1016/j.schres.2005.02.013>
- Cipriani, A., Barbui, C., Salanti, G., Rendell, J., Brown, R., Stockton, S., Purgato, M., Spineli, L. M., Goodwin, G. M., & Geddes, J. R. (2011). Comparative efficacy and acceptability of antimanic drugs in acute mania: A multiple-treatments meta-analysis. *Lancet (London, England)*, 378(9799), 1306–1315. [https://doi.org/10.1016/S0140-6736\(11\)60873-8](https://doi.org/10.1016/S0140-6736(11)60873-8)

- Ciuculete, D. M., Boström, A. E., Tuunainen, A.-K., Sohrabi, F., Kular, L., Jagodic, M., Voisin, S., Mwinyi, J., & Schiöth, H. B. (2018). Changes in methylation within the STK32B promoter are associated with an increased risk for generalized anxiety disorder in adolescents. *Journal of Psychiatric Research*, 102, 44–51. <https://doi.org/10.1016/j.jpsychires.2018.03.008>
- Cohen, J. (2013). *Statistical Power Analysis for the Behavioral Sciences* (2nd edn). Routledge. <https://doi.org/10.4324/9780203771587>
- Coombes, B. J., Markota, M., Mann, J. J., Colby, C., Stahl, E., Talati, A., Pathak, J., Weissman, M. M., McElroy, S. L., Frye, M. A., & Biernacka, J. M. (2020). Dissecting clinical heterogeneity of bipolar disorder using multiple polygenic risk scores. *Translational Psychiatry*, 10(1), Article 1. <https://doi.org/10.1038/s41398-020-00996-y>
- Correll, C. U., Detraux, J., De Lepeleire, J., & De Hert, M. (2015). Effects of antipsychotics, antidepressants and mood stabilizers on risk for physical diseases in people with schizophrenia, depression and bipolar disorder. *World Psychiatry*, 14(2), 119–136. <https://doi.org/10.1002/wps.20204>
- Correll, C. U., Kane, J. M., & Citrome, L. L. (2017). Epidemiology, Prevention, and Assessment of Tardive Dyskinesia and Advances in Treatment. *The Journal of Clinical Psychiatry*, 78(8), 1136–1147. <https://doi.org/10.4088/JCP.tv17016ah4c>
- Crossley, N. (2006). *Contesting Psychiatry: Social Movements in Mental Health*. Psychology Press.
- Crouch, E., Radcliff, E., Bennett, K. J., Brown, M. J., & Hung, P. (2021). Examining the Relationship Between Adverse Childhood Experiences and ADHD Diagnosis and Severity. *Academic Pediatrics*, 21(8), 1388–1394. <https://doi.org/10.1016/j.acap.2021.03.009>

- Cruceanu, C., Alda, M., Dion, P. A., Turecki, G., & Rouleau, G. A. (2015). No Evidence for GADL1 Variation as a Bipolar Disorder Susceptibility Factor in a Caucasian Lithium-Responsive Cohort. *American Journal of Psychiatry*, 172(1), 94–95.
<https://doi.org/10.1176/appi.ajp.2014.14070855>
- Cuesta, M. J., Lecumberri, P., Moreno-Izco, L., López-Ilundain, J. M., Ribeiro, M., Cabada, T., Lorente-Omeñaca, R., Erausquin, G. de, García-Martí, G., Sanjuan, J., Sánchez-Torres, A. M., Gómez, M., & Peralta, V. (2021). Motor abnormalities and basal ganglia in first-episode psychosis (FEP). *Psychological Medicine*, 51(10), 1625–1636.
<https://doi.org/10.1017/S0033291720000343>
- Damask, A., Steg, P. G., Schwartz, G. G., Szarek, M., Hagström, E., Badimon, L., Chapman, M. J., Boileau, C., Tsimikas, S., Ginsberg, H. N., Banerjee, P., Manvelian, G., Pordy, R., Hess, S., Overton, J. D., Lotta, L. A., Yancopoulos, G. D., Abecasis, G. R., Baras, A., ... On behalf of the Regeneron Genetics Center and the ODYSSEY OUTCOMES Investigators. (2020). Patients With High Genome-Wide Polygenic Risk Scores for Coronary Artery Disease May Receive Greater Clinical Benefit From Alirocumab Treatment in the ODYSSEY OUTCOMES Trial. *Circulation*, 141(8), 624–636.
<https://doi.org/10.1161/CIRCULATIONAHA.119.044434>
- Das, S., Forer, L., Schönherr, S., Sidore, C., Locke, A. E., Kwong, A., Vrieze, S. I., Chew, E. Y., Levy, S., McGue, M., Schlessinger, D., Stambolian, D., Loh, P.-R., Iacono, W. G., Swaroop, A., Scott, L. J., Cucca, F., Kronenberg, F., Boehnke, M., ... Fuchsberger, C. (2016). Next-generation genotype imputation service and methods. *Nature Genetics*, 48(10), 1284–1287.
<https://doi.org/10.1038/ng.3656>

- de Bakker, P. I. W., Ferreira, M. A. R., Jia, X., Neale, B. M., Raychaudhuri, S., & Voight, B. F. (2008). Practical aspects of imputation-driven meta-analysis of genome-wide association studies. *Human Molecular Genetics*, 17(R2), R122–R128. <https://doi.org/10.1093/hmg/ddn288>
- Demontis, D., Walters, G. B., Athanasiadis, G., Walters, R., Therrien, K., Farajzadeh, L., Voloudakis, G., Bendl, J., Zeng, B., Zhang, W., Grove, J., Als, T. D., Duan, J., Satterstrom, F. K., Bybjerg-Grauholm, J., Bækved-Hansen, M., Gudmundsson, O. O., Magnusson, S. H., Baldursson, G., ... Børglum, A. D. (2022). *Genome-wide analyses of ADHD identify 27 risk loci, refine the genetic architecture and implicate several cognitive domains* (p. 2022.02.14.22270780). medRxiv. <https://doi.org/10.1101/2022.02.14.22270780>
- Divac, N., Prostran, M., Jakovcevski, I., & Cerovac, N. (2014). Second-generation antipsychotics and extrapyramidal adverse effects. *BioMed Research International*, 2014, 656370. <https://doi.org/10.1155/2014/656370>
- Doherty, J. L., & Owen, M. J. (2014). Genomic insights into the overlap between psychiatric disorders: Implications for research and clinical practice. *Genome Medicine*, 6(4), 29. <https://doi.org/10.1186/gm546>
- D’Souza, R. S., & Hooten, W. M. (2023). Extrapyramidal Symptoms. In *StatPearls*. StatPearls Publishing. <http://www.ncbi.nlm.nih.gov/books/NBK534115/>
- Dudbridge, F. (2013). Power and Predictive Accuracy of Polygenic Risk Scores. *PLoS Genetics*, 9(3), e1003348. <https://doi.org/10.1371/journal.pgen.1003348>
- Duffy, A., Carlson, G., Dubicka, B., & Hillegers, M. H. J. (2020). Pre-pubertal bipolar disorder: Origins and current status of the controversy. *International Journal of Bipolar Disorders*,

8(1), 18. <https://doi.org/10.1186/s40345-020-00185-2>

Duncan, L. E., Ostacher, M., & Ballon, J. (2019). How genome-wide association studies (GWAS) made traditional candidate gene studies obsolete. *Neuropsychopharmacology*, 44(9), 1518–1523. <https://doi.org/10.1038/s41386-019-0389-5>

Durbin, R. (2014). Efficient haplotype matching and storage using the positional Burrows–Wheeler transform (PBWT). *Bioinformatics*, 30(9), 1266–1272. <https://doi.org/10.1093/bioinformatics/btu014>

Duvall, A., & Gallicchio, V. (2017). Lithium Treatment in Clinical Medicine: History, Current Status and Future Use. *Journal of Cell Science & Therapy*, 08. <https://doi.org/10.4172/2157-7013.1000270>

Elliott, H. R., Tillin, T., McArdle, W. L., Ho, K., Duggirala, A., Frayling, T. M., Davey Smith, G., Hughes, A. D., Chaturvedi, N., & Relton, C. L. (2014). Differences in smoking associated DNA methylation patterns in South Asians and Europeans. *Clinical Epigenetics*, 6(1), 4. <https://doi.org/10.1186/1868-7083-6-4>

Engelmann, J., Zillich, L., Frank, J., Wagner, S., Cetin, M., Herzog, D. P., Müller, M. B., Tadic, A., Foo, J. C., Sirignano, L., Braus, D. F., Dahmen, N., Sordon, S., Riemenschneider, M., Spaniol, C., Gasparoni, G., Rietschel, M., Witt, S. H., Lieb, K., & Streit, F. (2022). Epigenetic signatures in antidepressant treatment response: A methylome-wide association study in the EMC trial. *Translational Psychiatry*, 12(1), 1–9. <https://doi.org/10.1038/s41398-022-02032-7>

Escamilla, M. A., & Zavala, J. M. (2008). Genetics of bipolar disorder. *Dialogues in Clinical Neuroscience*, 10(2), 141–152. <https://doi.org/10.31887/DCNS.2008.10.2/maescamilla>

- Evarts, E. V., & Butler, R. N. (1959). A Review of the Effects of Chlorpromazine and Reserpine in Patients with Mental Disorders. In J. O. Cole & R. W. Gerard (Eds), *Psychopharmacology: Problems in evaluation*. (pp. 64–82). National Academy of Sciences.
<https://doi.org/10.1037/11259-004>
- Factor, S., Lang, A., & Weiner, W. (2008). *Drug Induced Movement Disorders*. John Wiley & Sons.
- Faria, M. A. (2013). Violence, mental illness, and the brain – A brief history of psychosurgery: Part 1 – From trephination to lobotomy. *Surgical Neurology International*, 4, 49.
<https://doi.org/10.4103/2152-7806.110146>
- Farreras, I. G. (2025). *History of Mental Illness*. Noba. <https://nobaproject.com/modules/history-of-mental-illness>
- Felitti, V. J., Anda, R. F., Nordenberg, D., Williamson, D. F., Spitz, A. M., Edwards, V., Koss, M. P., & Marks, J. S. (1998). Relationship of Childhood Abuse and Household Dysfunction to Many of the Leading Causes of Death in Adults: The Adverse Childhood Experiences (ACE) Study. *American Journal of Preventive Medicine*, 14(4), 245–258.
[https://doi.org/10.1016/S0749-3797\(98\)00017-8](https://doi.org/10.1016/S0749-3797(98)00017-8)
- Fitzgerald, M., & Gallus, K. L. (2025). Conceptualizing and measuring childhood adversity: A comprehensive critique of the adverse childhood experiences measure and offering a new conceptualization of childhood adversity. *American Journal of Orthopsychiatry*, 95(3), 274–287. <https://doi.org/10.1037/ort0000785>
- Freud, S. (2009). Introductory Lectures on Psycho-Analysis. In *Myths and Mythologies*. Routledge.
- Fujimaki, K., Morinobu, S., Yamashita, H., Takahashi, T., & Yamawaki, S. (2012). Predictors of quality of life in inpatients with schizophrenia. *Psychiatry Research*, 197(3), 199–205.

<https://doi.org/10.1016/j.psychres.2011.10.023>

Fusar-Poli, L., Amerio, A., Cimpoesu, P., Natale, A., Salvi, V., Zappa, G., Serafini, G., Amore, M.,

Aguglia, E., & Aguglia, A. (2021). Lipid and Glycemic Profiles in Patients with Bipolar Disorder: Cholesterol Levels Are Reduced in Mania. *Medicina*, 57(1), Article 1.

<https://doi.org/10.3390/medicina57010028>

GBD Mental Disorders Collaborators. (2022). Global, regional, and national burden of 12 mental disorders in 204 countries and territories, 1990–2019: A systematic analysis for the Global Burden of Disease Study 2019. *The Lancet Psychiatry*, 9(2), 137–150.

[https://doi.org/10.1016/S2215-0366\(21\)00395-3](https://doi.org/10.1016/S2215-0366(21)00395-3)

Ge, T., Chen, C.-Y., Ni, Y., Feng, Y.-C. A., & Smoller, J. W. (2019). Polygenic prediction via Bayesian regression and continuous shrinkage priors. *Nature Communications*, 10(1), Article 1. <https://doi.org/10.1038/s41467-019-09718-5>

George, P., Jones, N., Goldman, H., & Rosenblatt, A. (2023). Cycles of reform in the history of psychosis treatment in the United States. *SSM. Mental Health*, 3, 100205.

<https://doi.org/10.1016/j.ssmmh.2023.100205>

Giacomini, K. M., Yee, S. W., Mushiroda, T., Weinshilboum, R. M., Ratain, M. J., & Kubo, M. (2017). Genome-wide association studies of drug response and toxicity: An opportunity for genome medicine. *Nature Reviews Drug Discovery*, 16(1), 70–70.

<https://doi.org/10.1038/nrd.2016.234>

Gibson, G. (2012). Rare and common variants: Twenty arguments. *Nature Reviews Genetics*, 13(2), 135–145. <https://doi.org/10.1038/nrg3118>

Gilbert, R., Widom, C. S., Browne, K., Fergusson, D., Webb, E., & Janson, S. (2009). Burden and

- consequences of child maltreatment in high-income countries. *The Lancet*, 373(9657), 68–81. [https://doi.org/10.1016/S0140-6736\(08\)61706-7](https://doi.org/10.1016/S0140-6736(08)61706-7)
- Gitlin, M. (2016). Lithium side effects and toxicity: Prevalence and management strategies. *International Journal of Bipolar Disorders*, 4(1), 27. <https://doi.org/10.1186/s40345-016-0068-y>
- Godden, H. (2024, February 16). *Lithium monitoring and toxicity management*. The Pharmaceutical Journal. <https://pharmaceutical-journal.com/article/ld/lithium-monitoring-and-toxicity-management>
- Goes, F. S., McGrath, J., Avramopoulos, D., Wolyniec, P., Pirooznia, M., Ruczinski, I., Nestadt, G., Kenny, E. E., Vacic, V., Peters, I., Lencz, T., Darvasi, A., Mulle, J. G., Warren, S. T., & Pulver, A. E. (2015). Genome-wide association study of schizophrenia in Ashkenazi Jews. *American Journal of Medical Genetics Part B: Neuropsychiatric Genetics*, 168(8), 649–659. <https://doi.org/10.1002/ajmg.b.32349>
- Goetz, L. H., & Schork, N. J. (2018). Personalized Medicine: Motivation, Challenges and Progress. *Fertility and Sterility*, 109(6), 952–963. <https://doi.org/10.1016/j.fertnstert.2018.05.006>
- Goldfarb, M., De, H. M., Detraux, J., Di, P. K., Munir, H., Music, S., Piña I., & Ringen, P. A. (2022). Severe Mental Illness and Cardiovascular Disease. *Journal of the American College of Cardiology*, 80(9), 918–933. <https://doi.org/10.1016/j.jacc.2022.06.017>
- Graham, S. E., Clarke, S. L., Wu, K.-H. H., Kanoni, S., Zajac, G. J. M., Ramdas, S., Surakka, I., Ntalla, I., Vedantam, S., Winkler, T. W., Locke, A. E., Marouli, E., Hwang, M. Y., Han, S., Narita, A., Choudhury, A., Bentley, A. R., Ekoru, K., Verma, A., ... Willer, C. J. (2021). The power of genetic diversity in genome-wide association studies of lipids. *Nature*,

600(7890), Article 7890. <https://doi.org/10.1038/s41586-021-04064-3>

- Grigoriou-Serbanescu, M., Giaroli, G., Thygesen, J. H., Shenyan, O., Bigdeli, T. B., Bass, N. J., Diaconu, C. C., Neagu, A. I., Forstner, A. J., Degenhardt, F., Herms, S., Nöthen, M. M., & McQuillin, A. (2020). Predictive power of the ADHD GWAS 2019 polygenic risk scores in independent samples of bipolar patients with childhood ADHD. *Journal of Affective Disorders*, 265, 651–659. <https://doi.org/10.1016/j.jad.2019.11.109>
- Grof, P., Duffy, A., Cavazzoni, P., Grof, E., Garnham, J., MacDougall, M., O'Donovan, C., & Alda, M. (2002). Is Response to Prophylactic Lithium a Familial Trait? *The Journal of Clinical Psychiatry*, 63(10), 942–947. <https://doi.org/10.4088/JCP.v63n1013>
- Grotzinger, A. D., Rhemtulla, M., de Vlaming, R., Ritchie, S. J., Mallard, T. T., Hill, W. D., Ip, H. F., Marioni, R. E., McIntosh, A. M., Deary, I. J., Koellinger, P. D., Harden, K. P., Nivard, M. G., & Tucker-Drob, E. M. (2019). Genomic structural equation modelling provides insights into the multivariate genetic architecture of complex traits. *Nature Human Behaviour*, 3(5), 513–525. <https://doi.org/10.1038/s41562-019-0566-x>
- GTEx Consortium. (2020). The GTEx Consortium atlas of genetic regulatory effects across human tissues. *Science (New York, N.Y.)*, 369(6509), 1318–1330. <https://doi.org/10.1126/science.aaz1776>
- Guzman-Parra, J., Streit, F., Forstner, A. J., Strohmaier, J., González, M. J., Gil Flores, S., Cabaleiro Fabeiro, F. J., del Río Noriega, F., Perez Perez, F., Haro González, J., Orozco Diaz, G., de Diego-Otero, Y., Moreno-Kustner, B., Auburger, G., Degenhardt, F., Heilmann-Heimbach, S., Herms, S., Hoffmann, P., Frank, J., ... Rietschel, M. (2021). Clinical and genetic differences between bipolar disorder type 1 and 2 in multiplex families. *Translational*

Psychiatry, 11(1), Article 1. <https://doi.org/10.1038/s41398-020-01146-0>

Haddad, P. M., Brain, C., & Scott, J. (2014). Nonadherence with antipsychotic medication in schizophrenia: Challenges and management strategies. *Patient Related Outcome Measures*, 5, 43–62. <https://doi.org/10.2147/PROM.S42735>

Haggett, A. (2015). Pharmacological Solutions. In *A History of Male Psychological Disorders in Britain, 1945–1980*. Palgrave Macmillan. <https://www.ncbi.nlm.nih.gov/books/NBK333539/>

Hannon, E., Dempster, E., Viana, J., Burrage, J., Smith, A. R., Macdonald, R., St Clair, D., Mustard, C., Breen, G., Therman, S., Kaprio, J., Touloupoulou, T., Pol, H. E. H., Bohlken, M. M., Kahn, R. S., Nenadic, I., Hultman, C. M., Murray, R. M., Collier, D. A., ... Mill, J. (2016). An integrated genetic-epigenetic analysis of schizophrenia: Evidence for co-localization of genetic associations and differential DNA methylation. *Genome Biology*, 17(1), 176. <https://doi.org/10.1186/s13059-016-1041-x>

Hartz, S. M., Pato, C. N., Medeiros, H., Cavazos-Rehg, P., Sobell, J. L., Knowles, J. A., Bierut, L. J., Pato, M. T., & for the Genomic Psychiatry Cohort Consortium. (2014). Comorbidity of Severe Psychotic Disorders With Measures of Substance Use. *JAMA Psychiatry*, 71(3), 248–254. <https://doi.org/10.1001/jamapsychiatry.2013.3726>

Hawley, C., Fineberg, N., Roberts, A., Baldwin, D., Sahadevan, A., & Sharman, V. (2003). The use of the Simpson Angus Scale for the assessment of movement disorder: A training guide. *International Journal of Psychiatry in Clinical Practice*, 7(4), 349–2257. <https://doi.org/10.1080/13651500310002986>

Hayes, J. F., Marston, L., Walters, K., King, M. B., & Osborn, D. P. J. (2017). Mortality gap for

- people with bipolar disorder and schizophrenia: UK-based cohort study 2000–2014. *The British Journal of Psychiatry*, 211(3), 175–181. <https://doi.org/10.1192/bjp.bp.117.202606>
- He, Y., Koido, M., Shimmori, Y., & Kamatani, Y. (2023). *GWASLab: A Python package for processing and visualizing GWAS summary statistics*. <https://doi.org/10.51094/jxiv.370>
- Hedenmalm, K., Güzey, C., Dahl, M.-L., Yue, Q.-Y., & Spigset, O. (2006). Risk Factors for Extrapyramidal Symptoms During Treatment With Selective Serotonin Reuptake Inhibitors, Including Cytochrome P-450 Enzyme, and Serotonin and Dopamine Transporter and Receptor Polymorphisms. *Journal of Clinical Psychopharmacology*, 26(2), 192. <https://doi.org/10.1097/01.jcp.0000203200.96205.34>
- Hefner, G., Hahn, M., Toto, S., Hiemke, C., Roll, S. C., Wolff, J., & Klimke, A. (2021). Potentially inappropriate medication in older psychiatric patients. *European Journal of Clinical Pharmacology*, 77(3), 331–339. <https://doi.org/10.1007/s00228-020-03012-w>
- Heyn, H., & Esteller, M. (2012). DNA methylation profiling in the clinic: Applications and challenges. *Nature Reviews Genetics*, 13(10), 679–692. <https://doi.org/10.1038/nrg3270>
- Hiemke, C., Bergemann, N., Clement, H. W., Conca, A., Deckert, J., Domschke, K., Eckermann, G., Egberts, K., Gerlach, M., Greiner, C., Gründer, G., Haen, E., Havemann-Reinecke, U., Hefner, G., Helmer, R., Janssen, G., Jaquenoud, E., Laux, G., Messer, T., ... Baumann, P. (2018). Consensus Guidelines for Therapeutic Drug Monitoring in Neuropsychopharmacology: Update 2017. *Pharmacopsychiatry*, 51(1/2), 9–62. <https://doi.org/10.1055/s-0043-116492>
- Hiller, J. K., Jangmo, A., Tesli, M. S., Jaholkowski, P. P., Hoseth, E. Z., Steen, N. E., & Haram, M. (2023). Lipid Biomarker Research in Bipolar Disorder: A Scoping Review of Trends,

- Challenges, and Future Directions. *Biological Psychiatry Global Open Science*, 3(4), 594–604. <https://doi.org/10.1016/j.bpsgos.2023.07.004>
- Hippisley-Cox, J., Coupland, C., & Brindle, P. (2017). Development and validation of QRISK3 risk prediction algorithms to estimate future risk of cardiovascular disease: Prospective cohort study. *BMJ*, 357, j2099. <https://doi.org/10.1136/bmj.j2099>
- Hirsch, J. A., Nicola, G., McGinty, G., Liu, R. W., Barr, R. M., Chittle, M. D., & Manchikanti, L. (2016). ICD-10: History and Context. *AJNR: American Journal of Neuroradiology*, 37(4), 596–599. <https://doi.org/10.3174/ajnr.A4696>
- Hjorthøj, C., Stürup, A. E., McGrath, J. J., & Nordentoft, M. (2017). Years of potential life lost and life expectancy in schizophrenia: A systematic review and meta-analysis. *The Lancet Psychiatry*, 4(4), 295–301. [https://doi.org/10.1016/S2215-0366\(17\)30078-0](https://doi.org/10.1016/S2215-0366(17)30078-0)
- Hoggart, C. J., Clark, T. G., De Iorio, M., Whittaker, J. C., & Balding, D. J. (2008). Genome-wide significance for dense SNP and resequencing data. *Genetic Epidemiology*, 32(2), 179–185. <https://doi.org/10.1002/gepi.20292>
- Hopwood, M. (2020). The Shared Decision-Making Process in the Pharmacological Management of Depression. *The Patient - Patient-Centered Outcomes Research*, 13(1), 23–30. <https://doi.org/10.1007/s40271-019-00383-w>
- Horvath, S. (2013). DNA methylation age of human tissues and cell types. *Genome Biology*, 14(10), 3156. <https://doi.org/10.1186/gb-2013-14-10-r115>
- Hosang, G. M., Korszun, A., Jones, L., Jones, I., Gray, J. M., Gunasinghe, C. M., McGuffin, P., & Farmer, A. E. (2010). Adverse life event reporting and worst illness episodes in unipolar and bipolar affective disorders: Measuring environmental risk for genetic research.

Psychological Medicine, 40(11), 1829–1837.

<https://doi.org/10.1017/S003329170999225X>

Hosang, G. M., Manoli, A., Shakoor, S., Fisher, H. L., & Parker, C. (2023). Reliability and convergent validity of retrospective reports of childhood maltreatment by individuals with bipolar disorder. *Psychiatry Research*, 321, 115105. <https://doi.org/10.1016/j.psychres.2023.115105>

Hou, L., Heilbronner, U., Degenhardt, F., Adli, M., Akiyama, K., Akula, N., Arda, R., Arias, B., Backlund, L., Banzato, C. E. M., Benabarre, A., Bengesser, S., Bhattacharjee, A. K., Biernacka, J. M., Birner, A., Brichant-Petitjean, C., Bui, E. T., Cervantes, P., Chen, G.-B., ... Schulze, T. G. (2016). Genetic variants associated with response to lithium treatment in bipolar disorder: A genome-wide association study. *The Lancet*, 387(10023), 1085–1093. [https://doi.org/10.1016/S0140-6736\(16\)00143-4](https://doi.org/10.1016/S0140-6736(16)00143-4)

Houseman, E. A., Accomando, W. P., Koestler, D. C., Christensen, B. C., Marsit, C. J., Nelson, H. H., Wiencke, J. K., & Kelsey, K. T. (2012). DNA methylation arrays as surrogate measures of cell mixture distribution. *BMC Bioinformatics*, 13(1), 86. <https://doi.org/10.1186/1471-2105-13-86>

Howard, D. M., Adams, M. J., Clarke, T.-K., Hafferty, J. D., Gibson, J., Shirali, M., Coleman, J. R. I., Hagenaars, S. P., Ward, J., Wigmore, E. M., Alloza, C., Shen, X., Barbu, M. C., Xu, E. Y., Whalley, H. C., Marioni, R. E., Porteous, D. J., Davies, G., Deary, I. J., ... McIntosh, A. M. (2019). Genome-wide meta-analysis of depression identifies 102 independent variants and highlights the importance of the prefrontal brain regions. *Nature Neuroscience*, 22(3), Article 3. <https://doi.org/10.1038/s41593-018-0326-7>

- Hughes, K., Bellis, M. A., Hardcastle, K. A., Sethi, D., Butchart, A., Mikton, C., Jones, L., & Dunne, M. P. (2017). The effect of multiple adverse childhood experiences on health: A systematic review and meta-analysis. *The Lancet Public Health*, 2(8), e356–e366. [https://doi.org/10.1016/S2468-2667\(17\)30118-4](https://doi.org/10.1016/S2468-2667(17)30118-4)
- Huhn, M., Nikolakopoulou, A., Schneider-Thoma, J., Krause, M., Samara, M., Peter, N., Arndt, T., Bäckers, L., Rothe, P., Cipriani, A., Davis, J., Salanti, G., & Leucht, S. (2019). Comparative efficacy and tolerability of 32 oral antipsychotics for the acute treatment of adults with multi-episode schizophrenia: A systematic review and network meta-analysis. *The Lancet*, 394(10202), 939–951. [https://doi.org/10.1016/S0140-6736\(19\)31135-3](https://doi.org/10.1016/S0140-6736(19)31135-3)
- Hui, T. P., Kandola, A., Shen, L., Lewis, G., Osborn, D. P. J., Geddes, J. R., & Hayes, J. F. (2019). A systematic review and meta-analysis of clinical predictors of lithium response in bipolar disorder. *Acta Psychiatrica Scandinavica*, 140(2), 94–115. <https://doi.org/10.1111/acps.13062>
- Hwang, C. K., Chaurasia, S. S., Jackson, C. R., Chan, G. C.-K., Storm, D. R., & Iuvone, P. M. (2013). Circadian Rhythm of Contrast Sensitivity Is Regulated by a Dopamine–Neuronal PAS-Domain Protein 2–Adenylyl Cyclase 1 Signaling Pathway in Retinal Ganglion Cells. *The Journal of Neuroscience*, 33(38), 14989–14997. <https://doi.org/10.1523/JNEUROSCI.2039-13.2013>
- Insel, T. R., & Cuthbert, B. N. (2015). Brain disorders? Precisely. *Science*, 348(6234), 499–500. <https://doi.org/10.1126/science.aab2358>
- International Consortium on Lithium Genetics (ConLi+Gen). (2018). Association of Polygenic Score for Schizophrenia and HLA Antigen and Inflammation Genes With Response to

- Lithium in Bipolar Affective Disorder: A Genome-Wide Association Study. *JAMA Psychiatry*, 75(1), 65–74. <https://doi.org/10.1001/jamapsychiatry.2017.3433>
- Iversen, T. S. J., Steen, N. E., Dieset, I., Hope, S., Mørch, R., Gardsjord, E. S., Jørgensen, K. N., Melle, I., Andreassen, O. A., Molden, E., & Jönsson, E. G. (2018). Side effect burden of antipsychotic drugs in real life – Impact of gender and polypharmacy. *Progress in Neuro-Psychopharmacology and Biological Psychiatry*, 82, 263–271. <https://doi.org/10.1016/j.pnpbp.2017.11.004>
- Jakobsen, A. S., Speyer, H., Nørgaard, H. C. B., Karlsen, M., Birk, M., Hjorthøj, C., Mors, O., Krogh, J., Gluud, C., Pisinger, C., & Nordentoft, M. (2017). Effect of lifestyle coaching versus care coordination versus treatment as usual in people with severe mental illness and overweight: Two-years follow-up of the randomized CHANGE trial. *PLOS ONE*, 12(10), e0185881. <https://doi.org/10.1371/journal.pone.0185881>
- Jamison, K. R. (2001). Suicide and Bipolar Disorder. In *The Science of Mental Health*. Routledge.
- Jawad, I., Watson, S., Haddad, P. M., Talbot, P. S., & McAllister-Williams, R. H. (2018). Medication nonadherence in bipolar disorder: A narrative review. *Therapeutic Advances in Psychopharmacology*, 8(12), 349–363. <https://doi.org/10.1177/2045125318804364>
- Jayatilleke, N., Hayes, R. D., Dutta, R., Shetty, H., Hotopf, M., Chang, C.-K., & Stewart, R. (2017). Contributions of specific causes of death to lost life expectancy in severe mental illness. *European Psychiatry*, 43, 109–115. <https://doi.org/10.1016/j.eurpsy.2017.02.487>
- Jeste, D. V. (2004). Tardive dyskinesia rates with atypical antipsychotics in older adults. *The Journal of Clinical Psychiatry*, 65 Suppl 9, 21–24.
- Joas, E., Karanti, A., Song, J., Goodwin, G. M., Lichtenstein, P., & Landén, M. (2017).

- Pharmacological treatment and risk of psychiatric hospital admission in bipolar disorder. *The British Journal of Psychiatry: The Journal of Mental Science*, 210(3), 197–202. <https://doi.org/10.1192/bjp.bp.116.187989>
- Johansson, V., Kuja-Halkola, R., Cannon, T. D., Hultman, C. M., & Hedman, A. M. (2019). A population-based heritability estimate of bipolar disorder – In a Swedish twin sample. *Psychiatry Research*, 278, 180–187. <https://doi.org/10.1016/j.psychres.2019.06.010>
- Jonas, K. G., Lencz, T., Li, K., Malhotra, A. K., Perlman, G., Fochtmann, L. J., Bromet, E. J., & Kotov, R. (2019). Schizophrenia polygenic risk score and 20-year course of illness in psychotic disorders. *Translational Psychiatry*, 9(1), Article 1. <https://doi.org/10.1038/s41398-019-0612-5>
- Jonas, R. K., Montoyo, C. A., & Bearden, C. E. (2014). The 22q11.2 Deletion Syndrome as a Window into Complex Neuropsychiatric Disorders Over the Lifespan. *Biological Psychiatry*, 75(5), 351–360. <https://doi.org/10.1016/j.biopsych.2013.07.019>
- Jones, M. J., Goodman, S. J., & Kobor, M. S. (2015). DNA methylation and healthy human aging. *Aging Cell*, 14(6), 924–932. <https://doi.org/10.1111/accel.12349>
- Joslyn, C., Hawes, D. J., Hunt, C., & Mitchell, P. B. (2016). Is age of onset associated with severity, prognosis, and clinical features in bipolar disorder? A meta-analytic review. *Bipolar Disorders*, 18(5), 389–403. <https://doi.org/10.1111/bdi.12419>
- Jukić, M. M., Haslemo, T., Molden, E., & Ingelman-Sundberg, M. (2018). Impact of CYP2C19 Genotype on Escitalopram Exposure and Therapeutic Failure: A Retrospective Study Based on 2,087 Patients. *American Journal of Psychiatry*, 175(5), 463–470. <https://doi.org/10.1176/appi.ajp.2017.17050550>

- Kaar, S. J., Gobjila, C., Butler, E., Henderson, C., & Howes, O. D. (2019). Making decisions about antipsychotics: A qualitative study of patient experience and the development of a decision aid. *BMC Psychiatry*, 19(1), 309. <https://doi.org/10.1186/s12888-019-2304-3>
- Kao, W.-T., Wang, Y., Kleinman, J. E., Lipska, B. K., Hyde, T. M., Weinberger, D. R., & Law, A. J. (2010). Common genetic variation in Neuregulin 3 (NRG3) influences risk for schizophrenia and impacts NRG3 expression in human brain. *Proceedings of the National Academy of Sciences of the United States of America*, 107(35), 15619–15624. <https://doi.org/10.1073/pnas.1005410107>
- Kapur, S., & Remington, G. (2001). Dopamine D2 receptors and their role in atypical antipsychotic action: Still necessary and may even be sufficient. *Biological Psychiatry*, 50(11), 873–883. [https://doi.org/10.1016/S0006-3223\(01\)01251-3](https://doi.org/10.1016/S0006-3223(01)01251-3)
- Karatekin, C., & Hill, M. (2019). Expanding the Original Definition of Adverse Childhood Experiences (ACEs). *Journal of Child & Adolescent Trauma*, 12(3), 289–306. <https://doi.org/10.1007/s40653-018-0237-5>
- Katcher, H. I., Hill, A. M., Lanford, J. L. G., Yoo, J. S., & Kris-Etherton, P. M. (2009). Lifestyle Approaches and Dietary Strategies to Lower LDL-Cholesterol and Triglycerides and Raise HDL-Cholesterol. *Endocrinology and Metabolism Clinics of North America*, 38(1), 45–78. <https://doi.org/10.1016/j.ecl.2008.11.010>
- Kawa, S., & Giordano, J. (2012). A brief historicity of the Diagnostic and Statistical Manual of Mental Disorders: Issues and implications for the future of psychiatric canon and practice. *Philosophy, Ethics, and Humanities in Medicine : PEHM*, 7, 2. <https://doi.org/10.1186/1747-5341-7-2>

- Keaton, J. M., Kamali, Z., Xie, T., Vaez, A., Williams, A., Goleva, S. B., Ani, A., Evangelou, E., Hellwege, J. N., Yengo, L., Young, W. J., Traylor, M., Giri, A., Zheng, Z., Zeng, J., Chasman, D. I., Morris, A. P., Caulfield, M. J., Hwang, S.-J., ... Warren, H. R. (2024). Genome-wide analysis in over 1 million individuals of European ancestry yields improved polygenic risk scores for blood pressure traits. *Nature Genetics*, 56(5), 778–791. <https://doi.org/10.1038/s41588-024-01714-w>
- Keener, A. M., & Bordelon, Y. M. (2016). Parkinsonism. *Seminars in Neurology*, 36(4), 330–334. <https://doi.org/10.1055/s-0036-1585097>
- Kendall, T. (2011). The rise and fall of the atypical antipsychotics. *The British Journal of Psychiatry*, 199(4), 266–268. <https://doi.org/10.1192/bjp.bp.110.083766>
- Kessing, L. V., Hellmund, G., & Andersen, P. K. (2011). Predictors of excellent response to lithium: Results from a nationwide register-based study. *International Clinical Psychopharmacology*, 26(6), 323. <https://doi.org/10.1097/YIC.0b013e32834a5cd0>
- Kim, J., & Wessling-Resnick, M. (2014). Iron and mechanisms of emotional behavior. *The Journal of Nutritional Biochemistry*, 25(11), 1101–1107. <https://doi.org/10.1016/j.jnutbio.2014.07.003>
- Koestler, D. C., Christensen, B. C., Karagas, M. R., Marsit, C. J., Langevin, S. M., Kelsey, K. T., Wiencke, J. K., & Houseman, E. A. (2013). Blood-based profiles of DNA methylation predict the underlying distribution of cell types: A validation analysis. *Epigenetics*, 8(8), 816. <https://doi.org/10.4161/epi.25430>
- Kong, A., Thorleifsson, G., Frigge, M. L., Vilhjalmsdottir, B. J., Young, A. I., Thorgeirsson, T. E., Benonisdottir, S., Oddsson, A., Halldorsson, B. V., Masson, G., Gudbjartsson, D. F.,

- Helgason, A., Bjornsdottir, G., Thorsteinsdottir, U., & Stefansson, K. (2018). The nature of nurture: Effects of parental genotypes. *Science*, 359(6374), 424–428. <https://doi.org/10.1126/science.aan6877>
- Koning, J. P., Tenback, D. E., van Os, J., Aleman, A., Kahn, R. S., & van Harten, P. N. (2010). Dyskinesia and Parkinsonism in Antipsychotic-Naive Patients With Schizophrenia, First-Degree Relatives and Healthy Controls: A Meta-analysis. *Schizophrenia Bulletin*, 36(4), 723–731. <https://doi.org/10.1093/schbul/sbn146>
- Kotambail, A., Mathur, A., Bhat, S. M., Rai, P. S., Sharma, P. S., & Satyamoorthy, K. (2015). GADL1 gene polymorphisms and lithium response in bipolar I disorder: Lack of association from an Indian population. *Psychiatric Genetics*, 25(1), 39. <https://doi.org/10.1097/YPG.0000000000000066>
- Lacey, R. E., & Minnis, H. (2020). Practitioner Review: Twenty years of research with adverse childhood experience scores - Advantages, disadvantages and applications to practice. *Journal of Child Psychology and Psychiatry, and Allied Disciplines*, 61(2), 116–130. <https://doi.org/10.1111/jcpp.13135>
- Laffey, P. (2003). Psychiatric therapy in Georgian Britain. *Psychological Medicine*, 33(7), 1285–1297. <https://doi.org/10.1017/S0033291703008109>
- Lee, P. H., Anttila, V., Won, H., Feng, Y.-C. A., Rosenthal, J., Zhu, Z., Tucker-Drob, E. M., Nivard, M. G., Grotzinger, A. D., Posthuma, D., Wang, M. M.-J., Yu, D., Stahl, E. A., Walters, R. K., Anney, R. J. L., Duncan, L. E., Ge, T., Adolfsson, R., Banaschewski, T., ... Smoller, J. W. (2019). Genomic Relationships, Novel Loci, and Pleiotropic Mechanisms across Eight Psychiatric Disorders. *Cell*, 179(7), 1469-1482.e11.

<https://doi.org/10.1016/j.cell.2019.11.020>

Leucht, S., Cipriani, A., Spineli, L., Mavridis, D., Örey, D., Richter, F., Samara, M., Barbui, C., Engel, R. R., Geddes, J. R., Kissling, W., Stapf, M. P., Lässig, B., Salanti, G., & Davis, J. M. (2013). Comparative efficacy and tolerability of 15 antipsychotic drugs in schizophrenia: A multiple-treatments meta-analysis. *The Lancet*, 382(9896), 951–962.

[https://doi.org/10.1016/S0140-6736\(13\)60733-3](https://doi.org/10.1016/S0140-6736(13)60733-3)

Leucht, S., Corves, C., Arbter, D., Engel, R. R., Li, C., & Davis, J. M. (2009). Second-generation versus first-generation antipsychotic drugs for schizophrenia: A meta-analysis. *The Lancet*, 373(9657), 31–41. [https://doi.org/10.1016/S0140-6736\(08\)61764-X](https://doi.org/10.1016/S0140-6736(08)61764-X)

Levchenko, A., Kanapin, A., Samsonova, A., Fedorenko, O. Y., Kornetova, E. G., Nurgaliev, T., Mazo, G. E., Semke, A. V., Kibitov, A. O., Bokhan, N. A., Gainetdinov, R. R., & Ivanova, S. A. (2021). A genome-wide association study identifies a gene network associated with paranoid schizophrenia and antipsychotics-induced tardive dyskinesia. *Progress in Neuro-Psychopharmacology & Biological Psychiatry*, 105, 110134. <https://doi.org/10.1016/j.pnpbp.2020.110134>

Lewis, A. C. F., & Green, R. C. (2021). Polygenic risk scores in the clinic: New perspectives needed on familiar ethical issues. *Genome Medicine*, 13(1), 14. <https://doi.org/10.1186/s13073-021-00829-7>

Lewis, C. M., & Vassos, E. (2020). Polygenic risk scores: From research tools to clinical instruments. *Genome Medicine*, 12(1), 44. <https://doi.org/10.1186/s13073-020-00742-5>

Li, Z., Votava, J. A., Zajac, G. J. M., Nguyen, J. N., Jaimes, F. B. L., Ly, S. M., Brinkman, J. A., Giorgi, M. D., Kaul, S., Green, C. L., Clair, S. L. S., Belisle, S. L., Rios, J. M., Nelson, D.

- W., Sorci-Thomas, M. G., Lagor, W. R., Lamming, D. W., Yen, C.-L. E., & Parks, B. W. (2020). Integrating Mouse and Human Genetic Data to Move beyond GWAS and Identify Causal Genes in Cholesterol Metabolism. *Cell Metabolism*, 31(4), 741-754.e5. <https://doi.org/10.1016/j.cmet.2020.02.015>
- Lichtenstein, P., Yip, B. H., Björk, C., Pawitan, Y., Cannon, T. D., Sullivan, P. F., & Hultman, C. M. (2009). Common genetic determinants of schizophrenia and bipolar disorder in Swedish families: A population-based study. *The Lancet*, 373(9659), 234–239. [https://doi.org/10.1016/S0140-6736\(09\)60072-6](https://doi.org/10.1016/S0140-6736(09)60072-6)
- Lieberman, J. A., Stroup, T. S., McEvoy, J. P., Swartz, M. S., Rosenheck, R. A., Perkins, D. O., Keefe, R. S. E., Davis, S. M., Davis, C. E., Lebowitz, B. D., Severe, J., & Hsiao, J. K. (2005). Effectiveness of Antipsychotic Drugs in Patients with Chronic Schizophrenia. *New England Journal of Medicine*, 353(12), 1209–1223. <https://doi.org/10.1056/NEJMoa051688>
- Lim, K., Lam, M., Zai, C., Tay, J., Karlsson, N., Deshpande, S. N., Thelma, B. K., Ozaki, N., Inada, T., Sim, K., Chong, S.-A., Lencz, T., Liu, J., & Lee, J. (2021a). Genome wide study of tardive dyskinesia in schizophrenia. *Translational Psychiatry*, 11(1), Article 1. <https://doi.org/10.1038/s41398-021-01471-y>
- Lim, K., Lam, M., Zai, C., Tay, J., Karlsson, N., Deshpande, S. N., Thelma, B. K., Ozaki, N., Inada, T., Sim, K., Chong, S.-A., Lencz, T., Liu, J., & Lee, J. (2021b). Genome wide study of tardive dyskinesia in schizophrenia. *Translational Psychiatry*, 11(1), 351. <https://doi.org/10.1038/s41398-021-01471-y>
- Lin, B. D., Pries, L.-K., Arias-Magnasco, A., Klingenberg, B., Linden, D. E. J., Blokland, G. A. M.,

- van der Meer, D., Luykx, J. J., Rutten, B. P. F., & Guloksuz, S. (2025). Exposome-Wide Gene-By-Environment Interaction Study of Psychotic Experiences in the UK Biobank. *Biological Psychiatry Global Open Science*, 5(3), 100460. <https://doi.org/10.1016/j.bpsgos.2025.100460>
- Liu, Y., Aryee, M. J., Padyukov, L., Fallin, M. D., Hesselberg, E., Runarsson, A., Reinius, L., Acevedo, N., Taub, M., Ronninger, M., Shchetynsky, K., Scheynius, A., Kere, J., Alfredsson, L., Klareskog, L., Ekström, T. J., & Feinberg, A. P. (2013). Epigenome-wide association data implicate DNA methylation as an intermediary of genetic risk in rheumatoid arthritis. *Nature Biotechnology*, 31(2), 142–147. <https://doi.org/10.1038/nbt.2487>
- Lloyd-Jones, L. R., Zeng, J., Sidorenko, J., Yengo, L., Moser, G., Kemper, K. E., Wang, H., Zheng, Z., Magi, R., Esko, T., Metspalu, A., Wray, N. R., Goddard, M. E., Yang, J., & Visscher, P. M. (2019). Improved polygenic prediction by Bayesian multiple regression on summary statistics. *Nature Communications*, 10(1), Article 1. <https://doi.org/10.1038/s41467-019-12653-0>
- Loke, Y. J., Hannan, A. J., & Craig, J. M. (2015). The Role of Epigenetic Change in Autism Spectrum Disorders. *Frontiers in Neurology*, 6, 107. <https://doi.org/10.3389/fneur.2015.00107>
- Lombardo, G., Mondelli, V., Dazzan, P., & Pariante, C. M. (2021). Sex hormones and immune system: A possible interplay in affective disorders? A systematic review. *Journal of Affective Disorders*, 290, 1–14. <https://doi.org/10.1016/j.jad.2021.04.035>
- Lombardo, P. A., & Dorr, G. M. (2006). Eugenics, Medical Education, and the Public Health Service:

- Another Perspective on the Tuskegee Syphilis Experiment. *Bulletin of the History of Medicine*, 80(2), 291–316.
- Maggioni, E., Crespo-Facorro, B., Nenadic, I., Benedetti, F., Gaser, C., Sauer, H., Roiz-Santiañez, R., Poletti, S., Marinelli, V., Bellani, M., Perlini, C., Ruggeri, M., Altamura, A. C., Diwadkar, V. A., Brambilla, P., & Group, E. (2017). Common and distinct structural features of schizophrenia and bipolar disorder: The European Network on Psychosis, Affective disorders and Cognitive Trajectory (ENPACT) study. *PLOS ONE*, 12(11), e0188000. <https://doi.org/10.1371/journal.pone.0188000>
- Mahmoud, O., Dudbridge, F., Davey Smith, G., Munafo, M., & Tilling, K. (2022). A robust method for collider bias correction in conditional genome-wide association studies. *Nature Communications*, 13(1), Article 1. <https://doi.org/10.1038/s41467-022-28119-9>
- Malhi, G. S., Bassett, D., Boyce, P., Bryant, R., Fitzgerald, P. B., Fritz, K., Hopwood, M., Lyndon, B., Mulder, R., Murray, G., Porter, R., & Singh, A. B. (2015). Royal Australian and New Zealand College of Psychiatrists clinical practice guidelines for mood disorders. *Australian & New Zealand Journal of Psychiatry*, 49(12), 1087–1206. <https://doi.org/10.1177/0004867415617657>
- Malla, A., Joobar, R., & Garcia, A. (2015). “Mental illness is like any other medical illness”: A critical examination of the statement and its impact on patient care and society. *Journal of Psychiatry & Neuroscience : JPN*, 40(3), 147–150. <https://doi.org/10.1503/jpn.150099>
- Malone, J. C. (2014). Did John B. Watson Really “Found” Behaviorism? *The Behavior Analyst*, 37(1), 1–12. <https://doi.org/10.1007/s40614-014-0004-3>
- Mangoni, A. A., & Jackson, S. H. D. (2004). Age-related changes in pharmacokinetics and

- pharmacodynamics: Basic principles and practical applications. *British Journal of Clinical Pharmacology*, 57(1), 6–14. <https://doi.org/10.1046/j.1365-2125.2003.02007.x>
- Manolio, T. A., Collins, F. S., Cox, N. J., Goldstein, D. B., Hindorff, L. A., Hunter, D. J., McCarthy, M. I., Ramos, E. M., Cardon, L. R., Chakravarti, A., Cho, J. H., Guttmacher, A. E., Kong, A., Kruglyak, L., Mardis, E., Rotimi, C. N., Slatkin, M., Valle, D., Whittemore, A. S., ... Visscher, P. M. (2009). Finding the missing heritability of complex diseases. *Nature*, 461(7265), 747–753. <https://doi.org/10.1038/nature08494>
- Marangoni, C., De Chiara, L., & Faedda, G. L. (2015). Bipolar Disorder and ADHD: Comorbidity and Diagnostic Distinctions. *Current Psychiatry Reports*, 17(8), 67. <https://doi.org/10.1007/s11920-015-0604-y>
- Martin, A. R., Kanai, M., Kamatani, Y., Okada, Y., Neale, B. M., & Daly, M. J. (2019). Clinical use of current polygenic risk scores may exacerbate health disparities. *Nature Genetics*, 51(4), 584–591. <https://doi.org/10.1038/s41588-019-0379-x>
- Martínez-Levy, G., Renteria, M., Campos, A., Benjet, C., Rabinowits, J., Méndez, E., & Cruz-Fuentes, C. (2021). European Depression Polygenic Risk Score Predicts Suicide Behavior in Mexicans. *Biological Psychiatry*, 89(9, Supplement), S236. <https://doi.org/10.1016/j.biopsych.2021.02.592>
- Marx, W., Penninx, B. W. J. H., Solmi, M., Furukawa, T. A., Firth, J., Carvalho, A. F., & Berk, M. (2023). Major depressive disorder. *Nature Reviews Disease Primers*, 9(1), 1–21. <https://doi.org/10.1038/s41572-023-00454-1>
- McCarthy, M. I., Abecasis, G. R., Cardon, L. R., Goldstein, D. B., Little, J., Ioannidis, J. P. A., & Hirschhorn, J. N. (2008). Genome-wide association studies for complex traits: Consensus,

- uncertainty and challenges. *Nature Reviews Genetics*, 9(5), 356–369.
<https://doi.org/10.1038/nrg2344>
- McCarthy, S., Das, S., Kretschmar, W., Delaneau, O., Wood, A. R., Teumer, A., Kang, H. M., Fuchsberger, C., Danecek, P., Sharp, K., Luo, Y., Sidore, C., Kwong, A., Timpson, N., Koskinen, S., Vrieze, S., Scott, L. J., Zhang, H., Mahajan, A., ... Haplotype Reference Consortium. (2016). A reference panel of 64,976 haplotypes for genotype imputation. *Nature Genetics*, 48(10), 1279–1283. <https://doi.org/10.1038/ng.3643>
- McCutcheon, R., Beck, K., Bloomfield, M. A., Marques, T. R., Rogdaki, M., & Howes, O. D. (2015). Treatment resistant or resistant to treatment? Antipsychotic plasma levels in patients with poorly controlled psychotic symptoms. *Journal of Psychopharmacology*, 29(8), 892–897.
<https://doi.org/10.1177/0269881115576688>
- McGuffin, P., Farmer, A., & Harvey, I. (1991). A Polydiagnostic Application of Operational Criteria in Studies of Psychotic Illness: Development and Reliability of the OPCRIT System. *Archives of General Psychiatry*, 48(8), 764–770.
<https://doi.org/10.1001/archpsyc.1991.01810320088015>
- McGuffin, P., Rijdsdijk, F., Andrew, M., Sham, P., Katz, R., & Cardno, A. (2003). The Heritability of Bipolar Affective Disorder and the Genetic Relationship to Unipolar Depression. *Archives of General Psychiatry*, 60(5), 497–502. <https://doi.org/10.1001/archpsyc.60.5.497>
- McLaughlin, K. A. (2017). Future Directions in Childhood Adversity and Youth Psychopathology. In *Future Work in Clinical Child and Adolescent Psychology*. Routledge.
- McMahon, F. J., Stine, O. C., Chase, G. A., Meyers, D. A., Simpson, S. G., & DePaulo, J. R. (1994). Influence of clinical subtype, sex, and lineality on age at onset of major affective disorder

- in a family sample. *The American Journal of Psychiatry*, 151(2), 210–215.
<https://doi.org/10.1176/ajp.151.2.210>
- Merikangas, K. R., Jin, R., He, J.-P., Kessler, R. C., Lee, S., Sampson, N. A., Viana, M. C., Andrade, L. H., Hu, C., Karam, E. G., Ladea, M., Medina-Mora, M. E., Ono, Y., Posada-Villa, J., Sagar, R., Wells, J. E., & Zarkov, Z. (2011). Prevalence and Correlates of Bipolar Spectrum Disorder in the World Mental Health Survey Initiative. *Archives of General Psychiatry*, 68(3), 241–251. <https://doi.org/10.1001/archgenpsychiatry.2011.12>
- Michels, K. B. (2010). The promises and challenges of epigenetic epidemiology. *Experimental Gerontology*, 45(4), 297–301. <https://doi.org/10.1016/j.exger.2009.12.011>
- Michels, K. B., Binder, A. M., Dedeurwaerder, S., Epstein, C. B., Grealley, J. M., Gut, I., Houseman, E. A., Izzi, B., Kelsey, K. T., Meissner, A., Milosavljevic, A., Siegmund, K. D., Bock, C., & Irizarry, R. A. (2013). Recommendations for the design and analysis of epigenome-wide association studies. *Nature Methods*, 10(10), 949–955. <https://doi.org/10.1038/nmeth.2632>
- Möller, H.-J., Seemüller, F., Schennach-Wolff, R., Stübner, S., Rütther, E., & Grohmann, R. (2014). History, background, concepts and current use of comedication and polypharmacy in psychiatry. *International Journal of Neuropsychopharmacology*, 17(7), 983–996.
<https://doi.org/10.1017/S1461145713000837>
- Monaghan, J. H., Robinson, J. O., & Dodge, J. A. (1979). The children's life events inventory. *Journal of Psychosomatic Research*, 23(1), 63–68. [https://doi.org/10.1016/0022-3999\(79\)90073-4](https://doi.org/10.1016/0022-3999(79)90073-4)
- Motter, F. R., Fritzen, J. S., Hilmer, S. N., Paniz, É. V., & Paniz, V. M. V. (2018). Potentially inappropriate medication in the elderly: A systematic review of validated explicit criteria.

European Journal of Clinical Pharmacology, 74(6), 679–700.

<https://doi.org/10.1007/s00228-018-2446-0>

Müller, S. H., Girard, S. L., Hopfner, F., Merner, N. D., Bourassa, C. V., Lorenz, D., Clark, L. N.,

Tittmann, L., Soto-Ortolaza, A. I., Klebe, S., Hallett, M., Schneider, S. A., Hodgkinson, C.

A., Lieb, W., Wszolek, Z. K., Pendziwiat, M., Lorenzo-Betancor, O., Poewe, W., Ortega-

Cubero, S., ... Rouleau, G. A. (2016). Genome-wide association study in essential tremor

identifies three new loci. *Brain*, 139(12), 3163. <https://doi.org/10.1093/brain/aww242>

Mullins, N., Forstner, A. J., O'Connell, K. S., Coombes, B., Coleman, J. R. I., Qiao, Z., Als, T. D.,

Bigdeli, T. B., Børte, S., Bryois, J., Charney, A. W., Drange, O. K., Gandal, M. J., Hagenaars,

S. P., Ikeda, M., Kamitaki, N., Kim, M., Krebs, K., Panagiotaropoulou, G., ... Andreassen,

O. A. (2021). Genome-wide association study of more than 40,000 bipolar disorder cases

provides new insights into the underlying biology. *Nature Genetics*, 53(6), Article 6.

<https://doi.org/10.1038/s41588-021-00857-4>

Mullins, N., Kang, J., Campos, A. I., Coleman, J. R. I., Edwards, A. C., Galfalvy, H., Levey, D. F.,

Lori, A., Shabalín, A., Starnawska, A., Su, M.-H., Watson, H. J., Adams, M., Awasthi, S.,

Gandal, M., Hafferty, J. D., Hishimoto, A., Kim, M., Okazaki, S., ... Ruderfer, D. M. (2022).

Dissecting the Shared Genetic Architecture of Suicide Attempt, Psychiatric Disorders, and

Known Risk Factors. *Biological Psychiatry*, 91(3), 313–327.

<https://doi.org/10.1016/j.biopsych.2021.05.029>

Munetz, M. R., & Benjamin, S. (1988). How to Examine Patients Using the Abnormal Involuntary

Movement Scale. *Psychiatric Services*, 39(11), 1172–1177.

<https://doi.org/10.1176/ps.39.11.1172>

- Murphy, T. M., & Mill, J. (2014). Epigenetics in health and disease: Heralding the EWAS era. *The Lancet*, 383(9933), 1952–1954. [https://doi.org/10.1016/S0140-6736\(14\)60269-5](https://doi.org/10.1016/S0140-6736(14)60269-5)
- Must, A., Tasa, G., Lang, A., Vasar, E., Kõks, S., Maron, E., & Väli, M. (2008). Association of limbic system-associated membrane protein (LSAMP) to male completed suicide. *BMC Medical Genetics*, 9, 34. <https://doi.org/10.1186/1471-2350-9-34>
- Nalls, M. A., Blauwendraat, C., Vallerga, C. L., Heilbron, K., Bandres-Ciga, S., Chang, D., Tan, M., Kia, D. A., Noyce, A. J., Xue, A., Bras, J., Young, E., Coelln, R. von, Simón-Sánchez, J., Schulte, C., Sharma, M., Krohn, L., Pihlstrøm, L., Siitonen, A., ... Zhang, F. (2019). Identification of novel risk loci, causal insights, and heritable risk for Parkinson's disease: A meta-analysis of genome-wide association studies. *The Lancet Neurology*, 18(12), 1091–1102. [https://doi.org/10.1016/S1474-4422\(19\)30320-5](https://doi.org/10.1016/S1474-4422(19)30320-5)
- Nasrallah, H. A. (2008). Atypical antipsychotic-induced metabolic side effects: Insights from receptor-binding profiles. *Molecular Psychiatry*, 13(1), 27–35. <https://doi.org/10.1038/sj.mp.4002066>
- Natarajan, P., Young, R., Stitzel, N. O., Padmanabhan, S., Baber, U., Mehran, R., Sartori, S., Fuster, V., Reilly, D. F., Butterworth, A., Rader, D. J., Ford, I., Sattar, N., & Kathiresan, S. (2017). Polygenic Risk Score Identifies Subgroup With Higher Burden of Atherosclerosis and Greater Relative Benefit From Statin Therapy in the Primary Prevention Setting. *Circulation*, 135(22), 2091–2101. <https://doi.org/10.1161/CIRCULATIONAHA.116.024436>
- Nelson, J. C., Baumann, P., Delucchi, K., Joffe, R., & Katona, C. (2014). A systematic review and meta-analysis of lithium augmentation of tricyclic and second generation antidepressants

- in major depression. *Journal of Affective Disorders*, 168, 269–275.
<https://doi.org/10.1016/j.jad.2014.05.053>
- Nelson, M. R., Tipney, H., Painter, J. L., Shen, J., Nicoletti, P., Shen, Y., Floratos, A., Sham, P. C., Li, M. J., Wang, J., Cardon, L. R., Whittaker, J. C., & Sanseu, P. (2015). The support of human genetic evidence for approved drug indications. *Nature Genetics*, 47(8), 856–860.
<https://doi.org/10.1038/ng.3314>
- NICE. (2014a). *Bipolar disorder (update): Draft full guideline*. National Institute for Health and Clinical Excellence. [Http://www.nice.org.uk](http://www.nice.org.uk).
- NICE. (2014b). *Psychosis and schizophrenia in adults: Treatment and management (NICE Clinical Guideline CG178)*. Retrieved from <https://www.nice.org.uk/guidance/cg178>.
- Nielsen, R. E., Banner, J., & Jensen, S. E. (2021). Cardiovascular disease in patients with severe mental illness. *Nature Reviews Cardiology*, 18(2), Article 2.
<https://doi.org/10.1038/s41569-020-00463-7>
- Nielsen, R. E., Uggerby, A. S., Jensen, S. O. W., & McGrath, J. J. (2013). Increasing mortality gap for patients diagnosed with schizophrenia over the last three decades—A Danish nationwide study from 1980 to 2010. *Schizophrenia Research*, 146(1), 22–27.
<https://doi.org/10.1016/j.schres.2013.02.025>
- Nordentoft, M., Wahlbeck, K., Hällgren, J., Westman, J., Ösby, U., Alinaghizadeh, H., Gissler, M., & Laursen, T. M. (2013). Excess Mortality, Causes of Death and Life Expectancy in 270,770 Patients with Recent Onset of Mental Disorders in Denmark, Finland and Sweden. *PLOS ONE*, 8(1), e55176. <https://doi.org/10.1371/journal.pone.0055176>
- O’Connell, K. S., Koromina, M., van der Veen, T., Boltz, T., David, F. S., Yang, J. M. K., Lin, K.-

- H., Wang, X., Coleman, J. R. I., Mitchell, B. L., McGrouther, C. C., Rangan, A. V., Lind, P. A., Koch, E., Harder, A., Parker, N., Bendl, J., Adorjan, K., Agerbo, E., ... Andreassen, O. A. (2025). Genomics yields biological and phenotypic insights into bipolar disorder. *Nature*, 1–12. <https://doi.org/10.1038/s41586-024-08468-9>
- O’Connell, K. S., Shadrin, A., Bahrami, S., Smeland, O. B., Bettella, F., Frei, O., Krull, F., Askeland, R. B., Walters, G. B., Davíðsdóttir, K., Haraldsdóttir, G. S., Guðmundsson, Ó. Ó., Stefánsson, H., Fan, C. C., Steen, N. E., Reichborn-Kjennerud, T., Dale, A. M., Stefánsson, K., Djurovic, S., & Andreassen, O. A. (2021). Identification of genetic overlap and novel risk loci for attention-deficit/hyperactivity disorder and bipolar disorder. *Molecular Psychiatry*, 26(8), Article 8. <https://doi.org/10.1038/s41380-019-0613-z>
- O’Donoghue, S., Holleran, L., Cannon, D. M., & McDonald, C. (2017). Anatomical dysconnectivity in bipolar disorder compared with schizophrenia: A selective review of structural network analyses using diffusion MRI. *Journal of Affective Disorders*, 209, 217–228. <https://doi.org/10.1016/j.jad.2016.11.015>
- Osborn, D., Burton, A., Hunter, R., Marston, L., Atkins, L., Barnes, T., Blackburn, R., Craig, T., Gilbert, H., Heinkel, S., Holt, R., King, M., Michie, S., Morris, R., Morris, S., Nazareth, I., Omar, R., Petersen, I., Peveler, R., ... Walters, K. (2018). Clinical and cost-effectiveness of an intervention for reducing cholesterol and cardiovascular risk for people with severe mental illness in English primary care: A cluster randomised controlled trial. *The Lancet Psychiatry*, 5(2), 145–154. [https://doi.org/10.1016/S2215-0366\(18\)30007-5](https://doi.org/10.1016/S2215-0366(18)30007-5)
- O’Sullivan, J. W., Raghavan, S., Marquez-Luna, C., Luzum, J. A., Damrauer, S. M., Ashley, E. A., O’Donnell, C. J., Willer, C. J., Natarajan, P., & null, null. (2022). Polygenic Risk Scores

- for Cardiovascular Disease: A Scientific Statement From the American Heart Association. *Circulation*, 146(8), e93–e118. <https://doi.org/10.1161/CIR.0000000000001077>
- Owen, M. J., & Cardno, A. G. (1999). Psychiatric genetics: Progress, problems, and potential. *The Lancet*, 354, S11–S14. [https://doi.org/10.1016/S0140-6736\(99\)90242-8](https://doi.org/10.1016/S0140-6736(99)90242-8)
- Owen, M. J., Cardno, A. G., & O'Donovan, M. C. (2000). Psychiatric genetics: Back to the future. *Molecular Psychiatry*, 5(1), 22–31. <https://doi.org/10.1038/sj.mp.4000702>
- Pain, O., Glanville, K. P., Hagenaars, S. P., Selzam, S., Fürtjes, A. E., Gaspar, H. A., Coleman, J. R. I., Rimfeld, K., Breen, G., Plomin, R., Folkersen, L., & Lewis, C. M. (2021). Evaluation of polygenic prediction methodology within a reference-standardized framework. *PLOS Genetics*, 17(5), e1009021. <https://doi.org/10.1371/journal.pgen.1009021>
- Palmier-Claus, J. E., Berry, K., Bucci, S., Mansell, W., & Varese, F. (2016). Relationship between childhood adversity and bipolar affective disorder: Systematic review and meta-analysis. *The British Journal of Psychiatry*, 209(6), 454–459. <https://doi.org/10.1192/bjp.bp.115.179655>
- Park, Y.-M., Shekhtman, T., & Kelsoe, J. R. (2020). Interaction between adverse childhood experiences and polygenic risk in patients with bipolar disorder. *Translational Psychiatry*, 10(1), Article 1. <https://doi.org/10.1038/s41398-020-01010-1>
- Patel, V., Saxena, S., Lund, C., Thornicroft, G., Baingana, F., Bolton, P., Chisholm, D., Collins, P. Y., Cooper, J. L., Eaton, J., Herrman, H., Herzallah, M. M., Huang, Y., Jordans, M. J. D., Kleinman, A., Medina-Mora, M. E., Morgan, E., Niaz, U., Omigbodun, O., ... Unützer, J. (2018). The Lancet Commission on global mental health and sustainable development. *Lancet (London, England)*, 392(10157), 1553–1598. <https://doi.org/10.1016/S0140->

- Pereira, V. S., & Hiroaki-Sato, V. A. (2018). A brief history of antidepressant drug development: From tricyclics to beyond ketamine. *Acta Neuropsychiatrica*, 30(6), 307–322.
<https://doi.org/10.1017/neu.2017.39>
- Pérez-Piñar, M., Mathur, R., Foguet, Q., Ayis, S., Robson, J., & Ayerbe, L. (2016). Cardiovascular risk factors among patients with schizophrenia, bipolar, depressive, anxiety, and personality disorders. *European Psychiatry*, 35, 8–15. <https://doi.org/10.1016/j.eurpsy.2016.02.004>
- Perlis, R. H., Ostacher, M. J., Goldberg, J. F., Miklowitz, D. J., Friedman, E., Calabrese, J., Thase, M. E., & Sachs, G. S. (2010). Transition to mania during treatment of bipolar depression. *Neuropsychopharmacology: Official Publication of the American College of Neuropsychopharmacology*, 35(13), 2545–2552. <https://doi.org/10.1038/npp.2010.122>
- Perlis, R. H., Smoller, J. W., Ferreira, M. A. R., McQuillin, A., Bass, N., Lawrence, J., Sachs, G. S., Nimgaonkar, V., Scolnick, E. M., Gurling, H., Sklar, P., & Purcell, S. (2009). A Genomewide Association Study of Response to Lithium for Prevention of Recurrence in Bipolar Disorder. *American Journal of Psychiatry*, 166(6), 718–725.
<https://doi.org/10.1176/appi.ajp.2009.08111633>
- Perlman, K., Benrimoh, D., Israel, S., Rollins, C., Brown, E., Tunteng, J.-F., You, R., You, E., Tanguay-Sela, M., Snook, E., Miresco, M., & Berlim, M. T. (2019). A systematic meta-review of predictors of antidepressant treatment outcome in major depressive disorder. *Journal of Affective Disorders*, 243, 503–515. <https://doi.org/10.1016/j.jad.2018.09.067>
- Pfisterer, U., Petukhov, V., Demharter, S., Meichsner, J., Thompson, J. J., Batiuk, M. Y., Asenjo-Martinez, A., Vasistha, N. A., Thakur, A., Mikkelsen, J., Adorjan, I., Pinborg, L. H., Pers,

- T. H., von Engelhardt, J., Kharchenko, P. V., & Khodosevich, K. (2020). Identification of epilepsy-associated neuronal subtypes and gene expression underlying epileptogenesis. *Nature Communications*, 11(1), 5038. <https://doi.org/10.1038/s41467-020-18752-7>
- Pichot, P. (1986). [DSM-III: The 3d edition of the Diagnostic and Statistical Manual of Mental Disorders from the American Psychiatric Association]. *Revue Neurologique*, 142(5), 489–499.
- Pillinger, T., Howes, O. D., Correll, C. U., Leucht, S., Huhn, M., Schneider-Thoma, J., Gaughran, F., Jauhar, S., McGuire, P. K., Taylor, D. M., Young, A. H., & McCutcheon, R. A. (2023). Antidepressant and antipsychotic side-effects and personalised prescribing: A systematic review and digital tool development. *The Lancet. Psychiatry*, 10(11), 860–876. [https://doi.org/10.1016/S2215-0366\(23\)00262-6](https://doi.org/10.1016/S2215-0366(23)00262-6)
- Porter, R. (2003). *Madness: A Brief History*. OUP Oxford.
- Powell, V., Agha, S. S., Jones, R. B., Eyre, O., Stephens, A., Weavers, B., Lennon, J., Allardyce, J., Potter, R., Smith, D., Thapar, A., & Rice, F. (2021). ADHD in adults with recurrent depression. *Journal of Affective Disorders*, 295, 1153–1160. <https://doi.org/10.1016/j.jad.2021.09.010>
- Pretzsch, C. M., & Ecker, C. (2023). Structural neuroimaging phenotypes and associated molecular and genomic underpinnings in autism: A review. *Frontiers in Neuroscience*, 17. <https://doi.org/10.3389/fnins.2023.1172779>
- Price, A. L., Patterson, N. J., Plenge, R. M., Weinblatt, M. E., Shadick, N. A., & Reich, D. (2006). Principal components analysis corrects for stratification in genome-wide association studies. *Nature Genetics*, 38(8), 904–909. <https://doi.org/10.1038/ng1847>

- Purcell, S., Neale, B., Todd-Brown, K., Thomas, L., Ferreira, M. A. R., Bender, D., Maller, J., Sklar, P., De Bakker, P. I. W., Daly, M. J., & Sham, P. C. (2007). PLINK: A Tool Set for Whole-Genome Association and Population-Based Linkage Analyses. *The American Journal of Human Genetics*, 81(3), 559–575. <https://doi.org/10.1086/519795>
- Quidé, Y., Tozzi, L., Corcoran, M., Cannon, D. M., & Dauvermann, M. R. (2020). The Impact of Childhood Trauma on Developing Bipolar Disorder: Current Understanding and Ensuring Continued Progress. *Neuropsychiatric Disease and Treatment*, 16, 3095–3115. <https://doi.org/10.2147/NDT.S285540>
- R Core Team. (2021). *R: A language and environment for statistical computing*. R Foundation for Statistical Computing, Vienna, Austria. <https://www.R-project.org/>.
- Rakyan, V. K., Down, T. A., Balding, D. J., & Beck, S. (2011). Epigenome-wide association studies for common human diseases. *Nature Reviews Genetics*, 12(8), 529–541. <https://doi.org/10.1038/nrg3000>
- Rami, F. Z., Seo, H., Kang, C., Park, S., Li, L., Le, T.-H., Kim, S.-W., Won, S.-H., Chung, W., & Chung, Y.-C. (2025). Associations of polygenic risk score, environmental factors, and their interactions with the risk of schizophrenia spectrum disorders. *Psychological Medicine*, 55, e111. <https://doi.org/10.1017/S0033291725000753>
- Rayner, W. (2015). *HRC or 1000G Imputation preparation and checking [Online]*. Available: <Http://www.well.ox.ac.uk/~wrayner/tools/> [Accessed].
- Reiss, D., Ganiban, J. M., Leve, L. D., Neiderhiser, J. M., Shaw, D. S., & Natsuaki, M. N. (2022). Parenting in the Context of the Child: Genetic and Social Processes. *Monographs of the Society for Research in Child Development*, 87(1–3), 7–188.

<https://doi.org/10.1111/mono.12460>

- Relton, C. L., & Davey Smith, G. (2012). Two-step epigenetic Mendelian randomization: A strategy for establishing the causal role of epigenetic processes in pathways to disease. *International Journal of Epidemiology*, 41(1), 161–176. <https://doi.org/10.1093/ije/dyr233>
- Rotella, F., Cassioli, E., Calderani, E., Lazzeretti, L., Ragghianti, B., Ricca, V., & Mannucci, E. (2020). Long-term metabolic and cardiovascular effects of antipsychotic drugs. A meta-analysis of randomized controlled trials. *European Neuropsychopharmacology*, 32, 56–65. <https://doi.org/10.1016/j.euroneuro.2019.12.118>
- Ruth, K. S., Day, F. R., Tyrrell, J., Thompson, D. J., Wood, A. R., Mahajan, A., Beaumont, R. N., Wittemans, L., Martin, S., Busch, A. S., Erzurumluoglu, A. M., Hollis, B., O'Mara, T. A., Endometrial Cancer Association Consortium, McCarthy, M. I., Langenberg, C., Easton, D. F., Wareham, N. J., Burgess, S., ... Perry, J. R. B. (2020). Using human genetics to understand the disease impacts of testosterone in men and women. *Nature Medicine*, 26(2), 252–258. <https://doi.org/10.1038/s41591-020-0751-5>
- Rybakowski, J. K. (2014). Factors Associated with Lithium Efficacy in Bipolar Disorder. *Harvard Review of Psychiatry*, 22(6), 353–357. <https://doi.org/10.1097/HRP.0000000000000006>
- Rybakowski, J. K., Chlopocka-Wozniak, M., & Suwalska, A. (2001). The prophylactic effect of long-term lithium administration in bipolar patients entering treatment in the 1970s and 1980s. *Bipolar Disorders*, 3(2), 63–67. <https://doi.org/10.1034/j.1399-5618.2001.030203.x>
- Sabe, M., Pillinger, T., Kaiser, S., Chen, C., Taipale, H., Tanskanen, A., Tiihonen, J., Leucht, S., Correll, C. U., & Solmi, M. (2022). Half a century of research on antipsychotics and schizophrenia: A scientometric study of hotspots, nodes, bursts, and trends. *Neuroscience*

& *Biobehavioral Reviews*, 136, 104608. <https://doi.org/10.1016/j.neubiorev.2022.104608>

Samani, N. J., Beeston, E., Greengrass, C., Riveros-McKay, F., Debiec, R., Lawday, D., Wang, Q., Budgeon, C. A., Braund, P. S., Bramley, R., Kharodia, S., Newton, M., Marshall, A., Krzeminski, A., Zafar, A., Chahal, A., Heer, A., Khunti, K., Joshi, N., ... Nelson, C. P. (2024). Polygenic risk score adds to a clinical risk score in the prediction of cardiovascular disease in a clinical setting. *European Heart Journal*, 45(34), 3152–3160. <https://doi.org/10.1093/eurheartj/ehae342>

Schork, A. J., Thompson, W. K., Pham, P., Torkamani, A., Roddey, J. C., Sullivan, P. F., Kelsoe, J. R., O'Donovan, M. C., Furberg, H., Consortium, T. T. and G., Consortium, T. B. D. P. G., Consortium, T. S. P. G., Schork, N. J., Andreassen, O. A., & Dale, A. M. (2013). All SNPs Are Not Created Equal: Genome-Wide Association Studies Reveal a Consistent Pattern of Enrichment among Functionally Annotated SNPs. *PLOS Genetics*, 9(4), e1003449. <https://doi.org/10.1371/journal.pgen.1003449>

Schouten, H. J., Knol, W., Egberts, T. C. G., Schobben, A. F. A. M., Jansen, P. A. F., & van Marum, R. J. (2012). Quality of Life of Elderly Patients With Antipsychotic-Induced Parkinsonism: A Cross-Sectional Study. *Journal of the American Medical Directors Association*, 13(1), 82.e1-82.e5. <https://doi.org/10.1016/j.jamda.2010.12.003>

Schubert, K. O., Thalamuthu, A., Amare, A. T., Frank, J., Streit, F., Adl, M., Akula, N., Akiyama, K., Arda, R., Arias, B., Aubry, J.-M., Backlund, L., Bhattacharjee, A. K., Bellivier, F., Benabarre, A., Bengesser, S., Biernacka, J. M., Birner, A., Marie-Claire, C., ... Baune, B. T. (2021). Combining schizophrenia and depression polygenic risk scores improves the genetic prediction of lithium response in bipolar disorder patients. *Translational Psychiatry*,

11(1), 606. <https://doi.org/10.1038/s41398-021-01702-2>

- Scott, J., Etain, B., Manchia, M., Brichant-Petitjean, C., Geoffroy, P. A., Schulze, T., Alda, M., Bellivier, F., & Collaborators, C. (2020). An examination of the quality and performance of the Alda scale for classifying lithium response phenotypes. *Bipolar Disorders*, 22(3), 255–265. <https://doi.org/10.1111/bdi.12829>
- Scull, A. (2015). *Madness in Civilization: A Cultural History of Insanity, from the Bible to Freud, from the Madhouse to Modern Medicine on JSTOR*. <https://www-jstor-org.libproxy.ucl.ac.uk/stable/j.ctvc77hvc>
- Semahegn, A., Torpey, K., Manu, A., Assefa, N., Tesfaye, G., & Ankomah, A. (2020). Psychotropic medication non-adherence and its associated factors among patients with major psychiatric disorders: A systematic review and meta-analysis. *Systematic Reviews*, 9, 17. <https://doi.org/10.1186/s13643-020-1274-3>
- Shoham, N., Cooper, C., Lewis, G., Bebbington, P., & McManus, S. (2021). Temporal trends in psychotic symptoms: Repeated cross-sectional surveys of the population in England 2000–14. *Schizophrenia Research*, 228, 97–102. <https://doi.org/10.1016/j.schres.2020.11.057>
- Sirugo, G., Williams, S. M., & Tishkoff, S. A. (2019). The Missing Diversity in Human Genetic Studies. *Cell*, 177(1), 26–31. <https://doi.org/10.1016/j.cell.2019.02.048>
- Sklar, P., Ripke, S., Scott, L. J., Andreassen, O. A., Cichon, S., Craddock, N., Edenberg, H. J., Nurnberger, J. I., Rietschel, M., Blackwood, D., Corvin, A., Flickinger, M., Guan, W., Mattingdal, M., McQuillin, A., Kwan, P., Wienker, T. F., Daly, M., Dudbridge, F., ... Psychiatric GWAS Consortium Bipolar Disorder Working Group. (2011). Large-scale genome-wide association analysis of bipolar disorder identifies a new susceptibility locus

- near ODZ4. *Nature Genetics*, 43(10), Article 10. <https://doi.org/10.1038/ng.943>
- Smeland, O. B., Shadrin, A., Bahrami, S., Broce, I., Tesli, M., Frei, O., Wirgenes, K. V., O'Connell, K. S., Krull, F., Bettella, F., Steen, N. E., Sugrue, L., Wang, Y., Svenningsson, P., Sharma, M., Pihlstrøm, L., Toft, M., O'Donovan, M., Djurovic, S., ... Andreassen, O. A. (2021). Genome-wide Association Analysis of Parkinson's Disease and Schizophrenia Reveals Shared Genetic Architecture and Identifies Novel Risk Loci. *Biological Psychiatry*, 89(3), 227–235. <https://doi.org/10.1016/j.biopsych.2020.01.026>
- Smith, A. K., Kilaru, V., Klengel, T., Mercer, K. B., Bradley, B., Conneely, K. N., Ressler, K. J., & Binder, E. B. (2015). DNA extracted from saliva for methylation studies of psychiatric traits: Evidence tissue specificity and relatedness to brain. *American Journal of Medical Genetics Part B: Neuropsychiatric Genetics*, 168(1), 36–44. <https://doi.org/10.1002/ajmg.b.32278>
- So, H.-C., Chau, K.-L., Ao, F.-K., Mo, C.-H., & Sham, P.-C. (2019). Exploring shared genetic bases and causal relationships of schizophrenia and bipolar disorder with 28 cardiovascular and metabolic traits. *Psychological Medicine*, 49(8), 1286–1298. <https://doi.org/10.1017/S0033291718001812>
- Sollis, E., Mosaku, A., Abid, A., Buniello, A., Cerezo, M., Gil, L., Groza, T., Güneş, O., Hall, P., Hayhurst, J., Ibrahim, A., Ji, Y., John, S., Lewis, E., MacArthur, J. A. L., McMahon, A., Osumi-Sutherland, D., Panoutsopoulou, K., Pendlington, Z., ... Harris, L. W. (2023). The NHGRI-EBI GWAS Catalog: Knowledgebase and deposition resource. *Nucleic Acids Research*, 51(D1), D977–D985. <https://doi.org/10.1093/nar/gkac1010>
- Song, J., Bergen, S. E., Di Florio, A., Karlsson, R., Charney, A., Ruderfer, D. M., Stahl, E. A., Chambert, K. D., Moran, J. L., Gordon-Smith, K., Forty, L., Green, E. K., Jones, I., Jones,

- L., Scolnick, E. M., Sklar, P., Smoller, J. W., Lichtenstein, P., Hultman, C., ... Belliveau, R. A. (2016). Genome-wide association study identifies SESTD1 as a novel risk gene for lithium-responsive bipolar disorder. *Molecular Psychiatry*, 21(9), Article 9. <https://doi.org/10.1038/mp.2015.165>
- Song, J., Bergen, S. E., Di Florio, A., Karlsson, R., Charney, A., Ruderfer, D. M., Stahl, E. A., Chambert, K. D., Moran, J. L., Gordon-Smith, K., Forty, L., Green, E. K., Jones, I., Jones, L., Scolnick, E. M., Sklar, P., Smoller, J. W., Lichtenstein, P., Hultman, C., ... Landén, M. (2017). Erratum: Genome-wide association study identifies SESTD1 as a novel risk gene for lithium-responsive bipolar disorder. *Molecular Psychiatry*, 22(8), 1223. <https://doi.org/10.1038/mp.2016.246>
- Song, J., Sjölander, A., Joas, E., Bergen, S. E., Runeson, B., Larsson, H., Landén, M., & Lichtenstein, P. (2017). Suicidal Behavior During Lithium and Valproate Treatment: A Within-Individual 8-Year Prospective Study of 50,000 Patients With Bipolar Disorder. *American Journal of Psychiatry*, 174(8), 795–802. <https://doi.org/10.1176/appi.ajp.2017.16050542>
- Spitzer, R. L., Endicott, J., & Robins, E. (1978). Research Diagnostic Criteria: Rationale and Reliability. *Archives of General Psychiatry*, 35(6), 773–782. <https://doi.org/10.1001/archpsyc.1978.01770300115013>
- Spoorthy, M. S., Chakrabarti, S., & Grover, S. (2019). Comorbidity of bipolar and anxiety disorders: An overview of trends in research. *World Journal of Psychiatry*, 9(1), 7–29. <https://doi.org/10.5498/wjp.v9.i1.7>
- Stassen, H. H., Angst, J., Hell, D., Scharfetter, C., & Szegedi, A. (2007). Is there a common resilience mechanism underlying antidepressant drug response? Evidence from 2848

- patients. *The Journal of Clinical Psychiatry*, 68(8), 1195–1205.
<https://doi.org/10.4088/jcp.v68n0805>
- Stone, J. L., O'Donovan, M. C., Gurling, H., Kirov, G. K., Blackwood, D. H. R., Corvin, A., Craddock, N. J., Gill, M., Hultman, C. M., Lichtenstein, P., McQuillin, A., Pato, C. N., Ruderfer, D. M., Owen, M. J., St Clair, D., Sullivan, P. F., Sklar, P., Purcell (Leader), S. M., Stone, J. L., ... Stanley Center for Psychiatric Research and Broad Institute of MIT and Harvard. (2008). Rare chromosomal deletions and duplications increase risk of schizophrenia. *Nature*, 455(7210), 237–241. <https://doi.org/10.1038/nature07239>
- Strawbridge, R. J., Johnston, K. J. A., Bailey, M. E. S., Baldassarre, D., Cullen, B., Eriksson, P., deFaire, U., Ferguson, A., Gigante, B., Giral, P., Graham, N., Hamsten, A., Humphries, S. E., Kurl, S., Lyall, D. M., Lyall, L. M., Pell, J. P., Pirro, M., Savonen, K., ... Smith, D. J. (2021). The overlap of genetic susceptibility to schizophrenia and cardiometabolic disease can be used to identify metabolically different groups of individuals. *Scientific Reports*, 11(1), Article 1. <https://doi.org/10.1038/s41598-020-79964-x>
- Sudlow, C., Gallacher, J., Allen, N., Beral, V., Burton, P., Danesh, J., Downey, P., Elliott, P., Green, J., Landray, M., Liu, B., Matthews, P., Ong, G., Pell, J., Silman, A., Young, A., Sprosen, T., Peakman, T., & Collins, R. (2015). UK Biobank: An Open Access Resource for Identifying the Causes of a Wide Range of Complex Diseases of Middle and Old Age. *PLOS Medicine*, 12(3), e1001779. <https://doi.org/10.1371/journal.pmed.1001779>
- Suleman, R. (2020). A Brief History of Electroconvulsive Therapy. *American Journal of Psychiatry Residents' Journal*, 16(1), 6–6. <https://doi.org/10.1176/appi.ajp-rj.2020.160103>
- Sullivan, P. F., Agrawal, A., Bulik, C. M., Andreassen, O. A., Børglum, A. D., Breen, G., Cichon, S.,

- Edenberg, H. J., Faraone, S. V., Gelernter, J., Mathews, C. A., Nievergelt, C. M., Smoller, J. W., O'Donovan, M. C., & for the Psychiatric Genomics Consortium. (2018). Psychiatric Genomics: An Update and an Agenda. *American Journal of Psychiatry*, 175(1), 15–27. <https://doi.org/10.1176/appi.ajp.2017.17030283>
- Sullivan, P. F., Daly, M. J., & O'Donovan, M. (2012). Genetic architectures of psychiatric disorders: The emerging picture and its implications. *Nature Reviews. Genetics*, 13(8), 537–551. <https://doi.org/10.1038/nrg3240>
- Sun, L., Pennells, L., Kaptoge, S., Nelson, C. P., Ritchie, S. C., Abraham, G., Arnold, M., Bell, S., Bolton, T., Burgess, S., Dudbridge, F., Guo, Q., Sofianopoulou, E., Stevens, D., Thompson, J. R., Butterworth, A. S., Wood, A., Danesh, J., Samani, N. J., ... Di Angelantonio, E. (2021). Polygenic risk scores in cardiovascular risk prediction: A cohort study and modelling analyses. *PLoS Medicine*, 18(1), e1003498. <https://doi.org/10.1371/journal.pmed.1003498>
- Sundararajan, T., Manzardo, A. M., & Butler, M. G. (2018). Functional analysis of schizophrenia genes using GeneAnalytics program and integrated databases. *Gene*, 641, 25–34. <https://doi.org/10.1016/j.gene.2017.10.035>
- Takaesu, Y. (2018). Circadian rhythm in bipolar disorder: A review of the literature. *Psychiatry and Clinical Neurosciences*, 72(9), 673–682. <https://doi.org/10.1111/pcn.12688>
- Takamitsu, E., Otsuka, M., Haebara, T., Yano, M., Matsuzaki, K., Kobuchi, H., Moriya, K., & Utsumi, T. (2015). Identification of Human N-Myristoylated Proteins from Human Complementary DNA Resources by Cell-Free and Cellular Metabolic Labeling Analyses. *PLOS ONE*, 10(8), e0136360. <https://doi.org/10.1371/journal.pone.0136360>
- Takamura, S., Ikeda, A., Nishioka, K., Furuya, H., Tashiro, M., Matsushima, T., Li, Y., Yoshino, H.,

- Funayama, M., Morinobu, S., & Hattori, N. (2016). Schizophrenia as a prodromal symptom in a patient harboring SNCA duplication. *Parkinsonism & Related Disorders*, 25, 108–109. <https://doi.org/10.1016/j.parkreldis.2016.01.028>
- Tam, V., Patel, N., Turcotte, M., Bossé, Y., Paré, G., & Meyre, D. (2019). Benefits and limitations of genome-wide association studies. *Nature Reviews Genetics*, 20(8), 467–484. <https://doi.org/10.1038/s41576-019-0127-1>
- Tandon, R., Lenderking, W. R., Weiss, C., Shalhoub, H., Barbosa, C. D., Chen, J., Greene, M., Meehan, S. R., Duvold, L. B., Arango, C., Agid, O., & Castle, D. (2020). The impact on functioning of second-generation antipsychotic medication side effects for patients with schizophrenia: A worldwide, cross-sectional, web-based survey. *Annals of General Psychiatry*, 19(1), 42. <https://doi.org/10.1186/s12991-020-00292-5>
- Taylor, D. M., Barnes, T. R. E., & Young, A. H. (2021). *The Maudsley Prescribing Guidelines in Psychiatry*. John Wiley & Sons.
- Teicher, M. H., Samson, J. A., Anderson, C. M., & Ohashi, K. (2016). The effects of childhood maltreatment on brain structure, function and connectivity. *Nature Reviews Neuroscience*, 17(10), 652–666. <https://doi.org/10.1038/nrn.2016.111>
- Temtamy, S. A., Aglan, M. S., Valencia, M., Cocchi, G., Pacheco, M., Ashour, A. M., Amr, K. S., Helmy, S. M. H., El-Gammal, M. A., Wright, M., Lapunzina, P., Goodship, J. A., & Ruiz-Perez, V. L. (2008). Long interspersed nuclear element-1 (LINE1)-mediated deletion of EVC, EVC2, C4orf6, and STK32B in Ellis–van Creveld syndrome with borderline intelligence. *Human Mutation*, 29(7), 931–938. <https://doi.org/10.1002/humu.20778>
- The 1000 Genomes Project Consortium. (2010). A map of human genome variation from population

- scale sequencing. *Nature*, 467(7319), 1061–1073. <https://doi.org/10.1038/nature09534>
- Thorp, J. G., Gerring, Z. F., Colodro-Conde, L., Byrne, E. M., Medland, S. E., Middeldorp, C. M., & Derks, E. M. (2023). The association between trauma exposure, polygenic risk and individual depression symptoms. *Psychiatry Research*, 321, 115101. <https://doi.org/10.1016/j.psychres.2023.115101>
- Tiihonen, J., Mittendorfer-Rutz, E., Majak, M., Mehtälä, J., Hoti, F., Jedenius, E., Enksson, D., Leval, A., Sermon, J., Tanskanen, A., & Taipale, H. (2017). Real-World Effectiveness of Antipsychotic Treatments in a Nationwide Cohort of 29 823 Patients With Schizophrenia. *JAMA Psychiatry*, 74(7), 686–693. <https://doi.org/10.1001/jamapsychiatry.2017.1322>
- Torkamani, A., Wineinger, N. E., & Topol, E. J. (2018). The personal and clinical utility of polygenic risk scores. *Nature Reviews Genetics*, 19(9), 581–590. <https://doi.org/10.1038/s41576-018-0018-x>
- Trubetskoy, V., Pardiñas, A. F., Qi, T., Panagiotaropoulou, G., Awasthi, S., Bigdeli, T. B., Bryois, J., Chen, C.-Y., Dennison, C. A., Hall, L. S., Lam, M., Watanabe, K., Frei, O., Ge, T., Harwood, J. C., Koopmans, F., Magnusson, S., Richards, A. L., Sidorenko, J., ... Schizophrenia Working Group of the Psychiatric Genomics Consortium. (2022). Mapping genomic loci implicates genes and synaptic biology in schizophrenia. *Nature*, 604(7906), 502–508. <https://doi.org/10.1038/s41586-022-04434-5>
- Upthegrove, R., Chard, C., Jones, L., Gordon-Smith, K., Forty, L., Jones, I., & Craddock, N. (2015). Adverse childhood events and psychosis in bipolar affective disorder. *The British Journal of Psychiatry*, 206(3), 191–197. <https://doi.org/10.1192/bjp.bp.114.152611>
- Urbat, S. M., Cho, S. M. J., Paruchuri, K., Truong, B., Haidermota, S., Peloso, G. M., Hornsby, W.

- E., Philippakis, A., Fahed, A. C., & Natarajan, P. (2025). Dynamic Importance of Genomic and Clinical Risk for Coronary Artery Disease Over the Life Course. *Circulation: Genomic and Precision Medicine*, 18(1), e004681. <https://doi.org/10.1161/CIRCGEN.124.004681>
- van der Veen, J. N., Kennelly, J. P., Wan, S., Vance, J. E., Vance, D. E., & Jacobs, R. L. (2017). The critical role of phosphatidylcholine and phosphatidylethanolamine metabolism in health and disease. *Biochimica et Biophysica Acta (BBA) - Biomembranes*, 1859(9, Part B), 1558–1572. <https://doi.org/10.1016/j.bbamem.2017.04.006>
- van Harten, P. N., & Kahn, R. S. (1999). Tardive Dystonia. *Schizophrenia Bulletin*, 25(4), 741–748. <https://doi.org/10.1093/oxfordjournals.schbul.a033415>
- Vancampfort, D., Correll, C. U., Galling, B., Probst, M., De Hert, M., Ward, P. B., Rosenbaum, S., Gaughran, F., Lally, J., & Stubbs, B. (2016). Diabetes mellitus in people with schizophrenia, bipolar disorder and major depressive disorder: A systematic review and large scale meta-analysis. *World Psychiatry*, 15(2), 166–174. <https://doi.org/10.1002/wps.20309>
- Vancampfort, D., Firth, J., Schuch, F. B., Rosenbaum, S., Mugisha, J., Hallgren, M., Probst, M., Ward, P. B., Gaughran, F., De Hert, M., Carvalho, A. F., & Stubbs, B. (2017). Sedentary behavior and physical activity levels in people with schizophrenia, bipolar disorder and major depressive disorder: A global systematic review and meta-analysis. *World Psychiatry*, 16(3), 308–315. <https://doi.org/10.1002/wps.20458>
- Vancampfort, D., Stubbs, B., Mitchell, A. J., De Hert, M., Wampers, M., Ward, P. B., Rosenbaum, S., & Correll, C. U. (2015). Risk of metabolic syndrome and its components in people with schizophrenia and related psychotic disorders, bipolar disorder and major depressive disorder: A systematic review and meta-analysis. *World Psychiatry*, 14(3), 339–347.

<https://doi.org/10.1002/wps.20252>

- Villicaña, S., & Bell, J. T. (2021). Genetic impacts on DNA methylation: Research findings and future perspectives. *Genome Biology*, 22(1), 127. <https://doi.org/10.1186/s13059-021-02347-6>
- Visscher, P. M., Wray, N. R., Zhang, Q., Sklar, P., McCarthy, M. I., Brown, M. A., & Yang, J. (2017a). 10 Years of GWAS Discovery: Biology, Function, and Translation. *The American Journal of Human Genetics*, 101(1), 5–22. <https://doi.org/10.1016/j.ajhg.2017.06.005>
- Visscher, P. M., Wray, N. R., Zhang, Q., Sklar, P., McCarthy, M. I., Brown, M. A., & Yang, J. (2017b). 10 Years of GWAS Discovery: Biology, Function, and Translation. *American Journal of Human Genetics*, 101(1), 5–22. <https://doi.org/10.1016/j.ajhg.2017.06.005>
- Wagner, J. R., Busche, S., Ge, B., Kwan, T., Pastinen, T., & Blanchette, M. (2014). The relationship between DNA methylation, genetic and expression inter-individual variation in untransformed human fibroblasts. *Genome Biology*, 15(2), R37. <https://doi.org/10.1186/gb-2014-15-2-r37>
- Walsh, J. J. (1907). *Bedlam*. In *The Catholic Encyclopedia*. New York: Robert Appleton Company. [Http://www.newadvent.org/cathen/02387b.htm](http://www.newadvent.org/cathen/02387b.htm).
- Ward, M. F., Wender, P. H., & Reimherr, F. W. (1993). The Wender Utah Rating Scale: An aid in the retrospective diagnosis of childhood attention deficit hyperactivity disorder. *The American Journal of Psychiatry*, 150(6), 885–890. <https://doi.org/10.1176/ajp.150.6.885>
- Ward, R. J., Zucca, F. A., Duyn, J. H., Crichton, R. R., & Zecca, L. (2014). The role of iron in brain ageing and neurodegenerative disorders. *The Lancet Neurology*, 13(10), 1045–1060. [https://doi.org/10.1016/S1474-4422\(14\)70117-6](https://doi.org/10.1016/S1474-4422(14)70117-6)

- Watanabe, K., Taskesen, E., van Bochoven, A., & Posthuma, D. (2017). Functional mapping and annotation of genetic associations with FUMA. *Nature Communications*, 8(1), Article 1. <https://doi.org/10.1038/s41467-017-01261-5>
- Watson, H. J., Yilmaz, Z., & Sullivan, P. F. (2020). Chapter 9 - The Psychiatric Genomics Consortium: History, development, and the future. In B. T. Baune (Ed.), *Personalized Psychiatry* (pp. 91–101). Academic Press. <https://doi.org/10.1016/B978-0-12-813176-3.00009-2>
- Weissbrod, O., Hormozdiari, F., Benner, C., Cui, R., Ulirsch, J., Gazal, S., Schoech, A. P., van de Geijn, B., Reshef, Y., Márquez-Luna, C., O'Connor, L., Pirinen, M., Finucane, H. K., & Price, A. L. (2020). Functionally informed fine-mapping and polygenic localization of complex trait heritability. *Nature Genetics*, 52(12), Article 12. <https://doi.org/10.1038/s41588-020-00735-5>
- Wellington, A. (2022). Dr Manfred J. Sakel: Discoverer of insulin shock therapy – psychiatry in history. *The British Journal of Psychiatry*, 221(5), 682–682. <https://doi.org/10.1192/bjp.2022.73>
- Westman, J., Hällgren, J., Wahlbeck, K., Erlinge, D., Alfredsson, L., & Ösby, U. (2013). Cardiovascular mortality in bipolar disorder: A population-based cohort study in Sweden. *BMJ Open*, 3(4), e002373. <https://doi.org/10.1136/bmjopen-2012-002373>
- Weston-Green, K. (2022). Antipsychotic Drug Development: From Historical Evidence to Fresh Perspectives. *Frontiers in Psychiatry*, 13. <https://doi.org/10.3389/fpsy.2022.903156>
- Willer, C. J., Li, Y., & Abecasis, G. R. (2010). METAL: Fast and efficient meta-analysis of genomewide association scans. *Bioinformatics*, 26(17), 2190–2191.

<https://doi.org/10.1093/bioinformatics/btq340>

Wing, J. K., Babor, T., Brugha, T., Burke, J., Cooper, J. E., Giel, R., Jablenski, A., Regier, D., &

Sartorius, N. (1990). SCAN: Schedules for Clinical Assessment in Neuropsychiatry.

Archives of General Psychiatry, 47(6), 589–593.

<https://doi.org/10.1001/archpsyc.1990.01810180089012>

Wolff, J., Hefner, G., Normann, C., Kaier, K., Binder, H., Hiemke, C., Toto, S., Domschke, K.,

Marschollek, M., & Klimke, A. (2021). Polypharmacy and the risk of drug–drug interactions and potentially inappropriate medications in hospital psychiatry.

Pharmacoepidemiology and Drug Safety, 30(9), 1258–1268.

<https://doi.org/10.1002/pds.5310>

Woolf, B., Sallis, H. M., Munafò, M. R., & Gill, D. (2023). Deriving GWAS summary estimates for

paternal smoking in UK biobank: A GWAS by subtraction. *BMC Research Notes*, 16(1),

159. <https://doi.org/10.1186/s13104-023-06438-4>

World Health Organization. (1992). *The ICD-10 classification of mental and behavioural disorders:*

Clinical descriptions and diagnostic guidelines. World Health Organization.

<https://apps.who.int/iris/handle/10665/37958>

Wray, N. R., Lin, T., Austin, J., McGrath, J. J., Hickie, I. B., Murray, G. K., & Visscher, P. M. (2021).

From Basic Science to Clinical Application of Polygenic Risk Scores: A Primer. *JAMA*

Psychiatry, 78(1), 101–109. <https://doi.org/10.1001/jamapsychiatry.2020.3049>

Wray, N. R., Yang, J., Hayes, B. J., Price, A. L., Goddard, M. E., & Visscher, P. M. (2013). Pitfalls

of predicting complex traits from SNPs. *Nature Reviews Genetics*, 14(7), 507–515.

<https://doi.org/10.1038/nrg3457>

- Wu, T., Liu, Z., Mak, T. S. H., & Sham, P. C. (2022). Polygenic power calculator: Statistical power and polygenic prediction accuracy of genome-wide association studies of complex traits. *Frontiers in Genetics, 13*, 989639. <https://doi.org/10.3389/fgene.2022.989639>
- Wubeshet, Y. S., Mohammed, O. S., & Desse, T. A. (2019). Prevalence and management practice of first generation antipsychotics induced side effects among schizophrenic patients at Amanuel Mental Specialized Hospital, central Ethiopia: Cross-sectional study. *BMC Psychiatry, 19*(1), 32. <https://doi.org/10.1186/s12888-018-1999-x>
- Yamada, Y., Matsumoto, M., Iijima, K., & Sumiyoshi, T. (2020). Specificity and Continuity of Schizophrenia and Bipolar Disorder: Relation to ^{SEP}Biomarkers. *Current Pharmaceutical Design, 26*(2), 191–200. <https://doi.org/10.2174/1381612825666191216153508>
- Yao, K., van der Veen, T., Thygesen, J., Bass, N., & McQuillin, A. (2023). Multiple psychiatric polygenic risk scores predict associations between childhood adversity and bipolar disorder. *Journal of Affective Disorders, 341*, 137–146. <https://doi.org/10.1016/j.jad.2023.08.116>
- Yatham, L. N., Kennedy, S. H., Parikh, S. V., Schaffer, A., Bond, D. J., Frey, B. N., Sharma, V., Goldstein, B. I., Rej, S., Beaulieu, S., Alda, M., MacQueen, G., Milev, R. V., Ravindran, A., O'Donovan, C., McIntosh, D., Lam, R. W., Vazquez, G., Kapczinski, F., ... Berk, M. (2018). Canadian Network for Mood and Anxiety Treatments (CANMAT) and International Society for Bipolar Disorders (ISBD) 2018 guidelines for the management of patients with bipolar disorder. *Bipolar Disorders, 20*(2), 97–170. <https://doi.org/10.1111/bdi.12609>
- Yengo, L., Sidorenko, J., Kemper, K. E., Zheng, Z., Wood, A. R., Weedon, M. N., Frayling, T. M., Hirschhorn, J., Yang, J., Visscher, P. M., & the GIANT Consortium. (2018). Meta-analysis

- of genome-wide association studies for height and body mass index in ~700000 individuals of European ancestry. *Human Molecular Genetics*, 27(20), 3641–3649.
<https://doi.org/10.1093/hmg/ddy271>
- Yildiz, A., Vieta, E., Leucht, S., & Baldessarini, R. J. (2011). Efficacy of Antimanic Treatments: Meta-analysis of Randomized, Controlled Trials. *Neuropsychopharmacology*, 36(2), Article 2. <https://doi.org/10.1038/npp.2010.192>
- Yu, L., Long, Q., Zhang, Y., Liu, Y., Guo, Z., Cao, X., Qin, F., Xu, Y., Qian, Q., Gao, B., Chen, J., Liu, J., Zeng, Y., & Teng, Z. (2025). Bidirectional Mendelian randomization analysis of plasma lipidome and psychiatric disorders. *Journal of Affective Disorders*, 379, 871–883.
<https://doi.org/10.1016/j.jad.2024.10.063>
- Zai, C. C., Squassina, A., Tiwari, A. K., Pisanu, C., Pinna, M., Pinna, F., Meloni, A., Paribello, P., Carpiniello, B., Tondo, L., Frye, M. A., Biernacka, J. M., Coombes, B. J., Kennedy, J. L., & Manchia, M. (2023). A genome-wide association study of antidepressant-induced mania. *Progress in Neuro-Psychopharmacology and Biological Psychiatry*, 127, 110800.
<https://doi.org/10.1016/j.pnpbp.2023.110800>
- Zeilinger, S., Kühnel, B., Klopp, N., Baurecht, H., Kleinschmidt, A., Gieger, C., Weidinger, S., Lattka, E., Adamski, J., Peters, A., Strauch, K., Waldenberger, M., & Illig, T. (2013). Tobacco smoking leads to extensive genome-wide changes in DNA methylation. *PloS One*, 8(5), e63812. <https://doi.org/10.1371/journal.pone.0063812>
- Zhang, J.-P., Gallego, J. A., Robinson, D. G., Malhotra, A. K., Kane, J. M., & Correll, C. U. (2013). Efficacy and safety of individual second-generation vs. first-generation antipsychotics in first-episode psychosis: A systematic review and meta-analysis. *International Journal of*

Neuropsychopharmacology,

16(6),

1205–1218.

<https://doi.org/10.1017/S1461145712001277>

Appendices

2.1 OPCRIT Items 54 to 77

54. Persecutory delusions

Includes all delusions with persecutory ideation. (0, 1)

55. Well organised delusions

Illness is characterised by a series of well organised or well systematised delusions.

(0, 1)

56. Increased self esteem

Patient believes that he is an exceptional person with special powers, plans, talents or abilities. Rate positively here if overvalued idea but if delusional in quality also score item 57 (grandiose delusions). Score '1' if duration one week and '2' if lasts two weeks. (0, 1, 2)

57. Grandiose delusions

Patient has grossly exaggerated sense of own importance, has exceptional abilities or believes that he is rich or famous, titled or related to Royalty. Also included are delusions of identification with God, angels, the Messiah etc. (See also item 56). Any duration score '1', if symptom lasts at least 2 weeks score '2'. (0, 1, 2)

58. Delusions of influence

Events, objects or other people in patient's immediate surroundings have a special significance, often of a persecutory nature. Include ideas of reference from the TV or radio, or newspapers, where patient believes that these are providing instructions or prescribing certain behaviour. (0, 1)

59. Bizarre delusions

Strange, absurd or fantastic delusions whose content may have a mystical, magical or 'science fiction' quality. (0, 1)

60. Widespread delusions

Delusions which intrude into most aspects of the patient's life and/or preoccupy the patient for most of his time. (0, 1)

61. Delusions of passivity

Include all 'made' sensations, emotions or actions. Score '1' for all experiences of influence where patient knows that his own feelings, impulses, volitional acts or somatic sensations are controlled or imposed by an external agency. (0, 1).

62. Primary delusional perception

Score '1' where the patient perceives something in the outside world which triggers a special, significant relatively non understandable belief of which he is certain and which is in some way loosely linked to the triggering perception (0, 1).

63. Other primary delusions

Includes delusional mood and delusional ideas. Delusional mood is a strange mood in which the environment appears changed in a threatening way but the significance of the change cannot be understood by the patient who is usually tense, anxious or bewildered. Can lead to a delusional belief. A delusional idea appears abruptly in the patient's mind fully developed and unheralded by any related thoughts. (0, 1)

64. Delusions & hallucinations last for one week

Any type of delusion accompanied by hallucinations of any type lasting one week. (0, 1)

65. Persecutory/jealous delusions & hallucinations

This is self explanatory But note that abnormal beliefs are of delusional intensity and quality and are accompanied by true hallucinations. (0, 1)

66. Thought insertion

Score '1' when patient recognises that thoughts are being put into his head which are not his own and which have probably or definitely been inserted by some external agency. (0, 1)

67. Thought withdrawal

Score '1' when patient experiences thoughts ceasing in his head and may experience 'thought block' which is interpreted as thoughts being removed (or 'stolen') by some external agency. (0, 1)

68. Thought broadcast

Score '1' when patient experiences thoughts diffusing out of his head so that they may be shared by others or even heard by others. (0, 1)

69. Delusions of guilt

Firm belief held by subject that they have committed some sin, crime or have caused harm to others despite absence of any evidence to support this. (0, 1)

70. Delusions of poverty

Firm belief held by subject that they have lost all or much of their money or property impoverished despite absence of any evidence to support this. (0, 1)

71. Nihilistic delusions

Firmly held belief that some part of patient's body has disappeared or is rotting away or is affected by some devastating or malignant disorder despite a lack of any objective supporting evidence. (0, 1)

72. Thought echo

Score '1' if patient experiences thoughts repeated or echoed in his or her head or by a voice outside the head. (0, 1)

73. Third person auditory hallucinations

Two or more voices discussing the patient in the third person. Score '1' if either 'true' or 'pseudo' hallucinations, i.e. differentiation of the source of the voices is unimportant. (0, 1)

74. Running commentary voices

Patient hears voice(s) describing his actions, sensations or emotions as they occur.

Score '1' whether these are possible 'pseudo' hallucinations or definite ('true') hallucinations (0, 1)

75. Abusive/ accusatory/ persecutory voices

Voices talking to the patient in an accusatory, abusive or persecutory manner. (0, 1)

76. Other (non affective) auditory hallucinations

Any other kind of auditory hallucination. Includes pleasant or neutral voices and non verbal hallucinations. (0, 1)

77. Non-affective hallucination in any modality

Hallucinations in which the content has no apparent relationship to elation or depression. Score '1' if present throughout the day for several days or intermittently for One week. (0, 1)

2.2 CLEQ QUESTIONNAIRE

Did you experience as a child (up to age 16 years) any of the following life events?

		<i>Please circle</i>	<i>If yes, how old were you?</i>
1	Death of parent	Yes No	_____
2	Death of a brother/sister	Yes No	_____
3	Death of a close friend	Yes No	_____
4	Divorce of parents	Yes No	_____
5	Marital separation of parents	Yes No	_____
6	Marriage of parent to step parent	Yes No	_____
7	Serious illness needing hospitalisation	Yes No	_____
8	Hospitalisation of a parent	Yes No	_____
9	Acquiring a visible deformity	Yes No	_____
10	Prison sentence of a parent for a year or more	Yes No	_____
11	Teenage pregnancy/fatherhood	Yes No	_____
12	Suspension from school	Yes No	_____
13	Are there any other significant life events you experienced as a child that are not mentioned above	Yes No	

Please provide brief details

Date of Completion: _____

Completed by: Self

☐

Interviewer

☐

2.3 Supplementary Table 1 Results of Multiple Regression Analyses with Adjustments based on ACE (total score) and BD/ADHD PRS

Variables	Estimated Coefficient	Standard Error	Confidence Intervals (95%)	Reference Values	<i>p</i>	<i>FDR_p</i>
Age at onset[†]						
BD PRS	-0.351	0.480	-1.294, 0.592	-0.731	0.465	0.709
BD Interaction	-0.370	0.356	-1.070, 0.330	-1.037	0.300	0.706
ADHD PRS	-0.328	0.506	-1.322, 0.665	-0.649	0.516	0.718
ADHD Interaction	-0.295	0.356	-0.993, 0.404	-0.828	0.408	0.706
Presence of psychotic symptoms[‡]						
BD PRS	0.254	0.090	0.079, 0.430	2.833	0.005	0.074
BD Interaction	-0.089	0.068	-0.223, 0.045	-1.300	0.194	0.706
ADHD PRS	-0.080	0.093	-0.264, 0.102	-0.863	0.388	0.706
ADHD Interaction	0.033	0.068	-0.100, 0.168	0.481	0.631	0.807
Presence of suicide ideation[‡]						
BD PRS	-0.029	0.096	-0.218, 0.157	-0.306	0.760	0.884
BD Interaction	-0.097	0.078	-0.250, 0.055	-1.246	0.213	0.706
ADHD PRS	0.077	0.100	-0.119, 0.275	0.770	0.441	0.706
ADHD Interaction	-0.022	0.079	-0.175, 0.135	-0.276	0.782	0.884
Presence of rapid cycling[‡]						
BD PRS	-0.148	0.106	-0.357, 0.058	-1.406	0.160	0.706
BD Interaction	-0.014	0.088	-0.187, 0.158	-0.157	0.875	0.884
ADHD PRS	0.471	0.124	0.232, 0.721	3.780	<0.001	0.005
ADHD Interaction	0.125	0.094	-0.056, 0.314	1.334	0.182	0.706

Notes. ACE, adverse childhood experience; PRS, polygenic risk score; BD, bipolar disorder; ADHD, attention deficit hyperactivity disorder.

[†] Multiple linear regression analyses, reference value t.

[‡] Multiple logistic regression analyses, reference value z.

FDR_P = false discovery rate corrected p values for multiple testing.

In bold p values survived multiple testing correction in main analyses.

All results were adjusted for participants' BD age of onset and sex (except for age of onset where only sex was adjusted).

PRS & interaction results were adjusted for chip type and the first three principal components from GWAS population stratification in addition to sex and age of onset.

2.4 Supplementary Table 2 Results of Multiple Regression Analyses with Adjustments based on ACE (total score) and MDD/SCZ PRS

Variables	Estimated Coefficient	Standard Error	Confidence Intervals (95%)	Reference Values	<i>p</i>	<i>FDR_p</i>
Age at onset[†]						
MDD PRS	-1.197	0.487	-2.154, -0.241	-2.458	0.014	0.153
MDD Interaction	-0.052	0.355	-0.750, 0.646	-0.145	0.884	0.884
SCZ PRS	-0.442	0.489	-1.403, 0.518	-0.905	0.366	0.706
SCZ Interaction	-0.356	0.350	-1.043, 0.333	-1.015	0.311	0.706
Presence of psychotic symptoms[‡]						
MDD PRS	0.063	0.091	-0.115, 0.241	0.691	0.489	0.712
MDD Interaction	-0.056	0.068	-0.191, 0.077	-0.830	0.406	0.706
SCZ PRS	0.185	0.090	0.009, 0.362	2.060	0.039	0.315
SCZ Interaction	-0.072	0.068	-0.206, 0.061	-1.054	0.292	0.706
Presence of suicide ideation[‡]						
MDD PRS	0.079	0.098	-0.114, 0.272	0.802	0.422	0.706
MDD Interaction	-0.021	0.080	-0.179, 0.137	-0.260	0.795	0.884
SCZ PRS	-0.061	0.100	-0.260, 0.133	-0.610	0.542	0.722
SCZ Interaction	-0.012	0.082	-0.174, 0.148	-0.149	0.881	0.884
Presence of rapid cycling[‡]						
MDD PRS	0.116	0.112	-0.102, 0.337	1.039	0.299	0.706
MDD Interaction	0.114	0.091	-0.064, 0.292	1.231	0.218	0.706
SCZ PRS	-0.023	0.112	-0.242, 0.197	-0.203	0.839	0.884
SCZ Interaction	0.067	0.082	-0.094, 0.230	0.811	0.417	0.706

Notes. ACE, adverse childhood experience; PRS, polygenic risk score; MDD, major depressive disorder; SCZ, schizophrenia disorder.

[†] Multiple linear regression analysis, reference value t.

[‡] Multiple logistic regression analysis, reference value z.

FDR_p = false discovery rate corrected p values for multiple testing.

All results were adjusted for participants' BD age of onset and sex (except for age of onset where only sex was adjusted). PRS & interaction results were adjusted for chip type and the first three principal components from GWAS population stratification in addition to sex and age of onset.

3.1 Supplementary Table 1 UCL Samples' Lithium Response Assessment

Response to Lithium

	<u>Investigators Opinion</u>		<u>Participants Opinion</u>
☐	Very good - no episodes on lithium during long-term treatment	☐	Very good
☐	Good - almost complete elimination of episodes, maintained well on lithium alone, episodes are due to confounding factors e.g.non-compliance	☐	Good
☐	Medium - a significant reduction in episode frequency or severity and/or frequent use of additional psychotropic medication	☐	Medium
☐	Poor - little clinical improvement, often in the presence of therapeutic lithium levels	☐	Poor
☐	Very poor increase in number of episodes	☐	Very poor
☐	Unknown/Unclassifiable	☐	Unknown/Unclassifiable

3.2 Supplementary Table 2 UCL samples' demographics and clinical characteristics concerning lithium response

Variables	N	Overall N = 1259	Lithium Responders N = 440 (35%)	Lithium Non-responders N = 819 (65%)	p-values
Age at interview	1153	50.39 (12.84)	50.34 (13.62)	50.42 (12.40)	0.921 ^a
Age of BD onset	1198	24.66 (10.62)	24.33 (10.68)	24.82 (10.60)	0.453 ^a
Sex	1259				0.039 ^b
Male		499 (40%)	192 (44%)	307 (37%)	
Female		760 (60%)	248 (56%)	512 (63%)	
BD type	1259				0.189 ^b
BD type 1		827 (66%)	297 (68%)	530 (65%)	
BD type 2		169 (13%)	63 (14%)	106 (13%)	
SABP		96 (8%)	34 (8%)	62 (7%)	
No information		167 (13%)	46 (10%)	121 (15%)	
PPD diagnosis	1181	41 (3%)	7 (1%)	34 (3%)	0.027 ^b

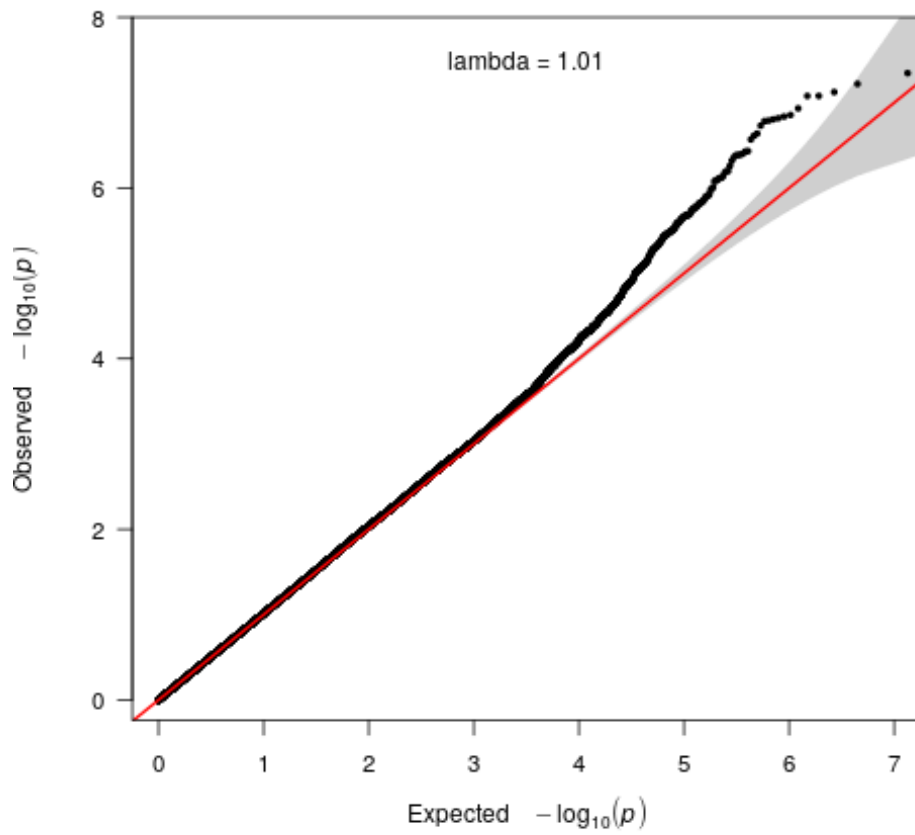
BD, bipolar disorder; SABP, schizoaffective bipolar disorder; PPD, premorbid personality disorder; SD, standard deviation

^a Two Sample t-test; mean (SD)

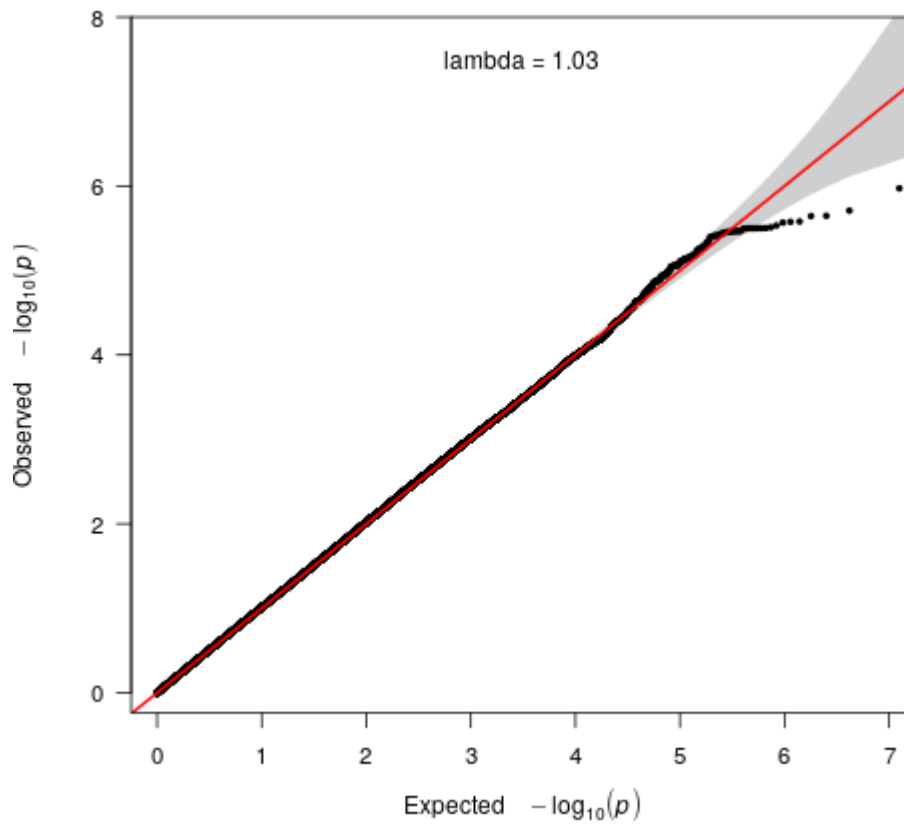
^b Pearson's Chi-squared test of independence; n (%)

In bold, p passed significance at 0.05.

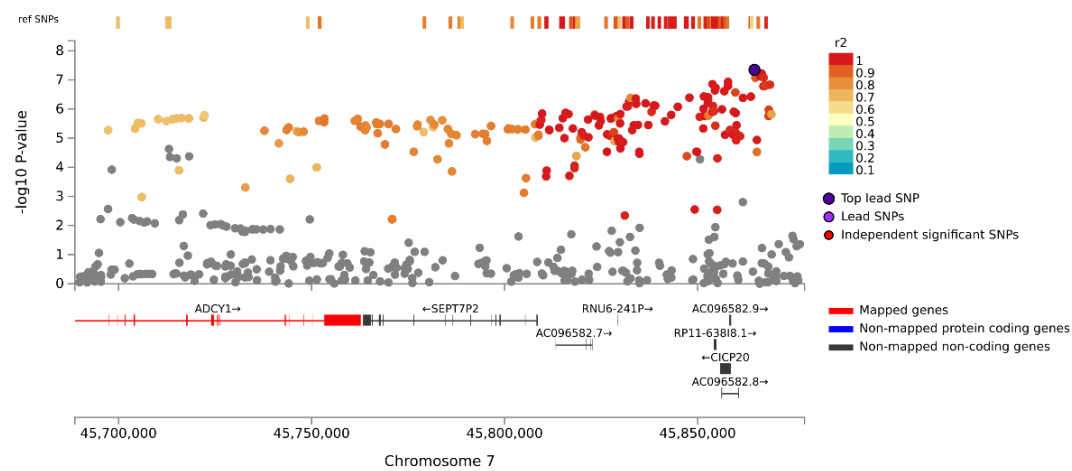
3.3 Supplementary Figure 1 The QQ plot for the UCL+Song^{Sub}+ConLi⁺Gen GWAS meta-analysis



3.4 Supplementary Figure 2 The QQ plot for the UCL+Song^{Ob}+ConLi⁺Gen GWAS meta-analysis



3.5 Supplementary Figure 3 Regional association plot on chromosome 7 in which the Genome-wide significant SNP is located



4.1 Supplementary Table 1 Behavioural Key Words used to Identify EPSE Cases

Symptoms	Keywords
General terms for EPSE	Extrapyramidal; extra pyramidal; epse; EPSE; movement disorder
Parkinsonism (tremor)	Tremor; tremble; shake; quiver
Parkinsonism (rigidity)	Rigid; hypertonic; stiff; inflexible; tight; tense; muscle pain
Parkinsonism (Sialorrhea)	Salivation; sialorrhea; drool
Parkinsonism (bradykinesia and hypokinesia)	Hypokinesia; bradykinesia; freeze; froze; slow; motor block; stride; reduced arm swing; blink; hypophonia; soft voice; voice volume; slurred; micrographia; handwriting
Parkinsonism (mask face)	Mask; hypomimia; expressionless
Parkinsonism (stooped posture)	Stoop; hunch; posture
Parkinsonism (parkinsonian gait)	Shuffle; festinate; parkinsonian
Parkinsonism (parkinsonian gait)	Imbalance; fall
Dystonia (general)	Dystonia; jerk; twist; twitch; spasm; lock
Dystonia (opisthotonos)	Opisthotonos; arch back; bend back; arch spine; bend spine
Dystonia (torticollis)	Torticollis; cervical dystonia; head deviation; head posturing; neck pain
Dystonia (oculogyric crisis)	Oculogyric; fix stare; fix eye; deviate eye
Dystonia (trismus)	Trismus; lock jaw; lockjaw; jawlock; jaw deviation; jaw retraction; clench; grind; mouth pain; restrict mouth; limit mouth
Dystonia (tortipelvic crisis)	Tortipelvic; bend trunk; twist trunk
Dystonia (buccolingual crisis)	Buccolingual; dysphagia; difficult swallow; grimace; protrude; protrusion; risus sardonicus; dysarthria; difficult speak; difficult speech; pseudomacroglossia; swollen tounge; tounge swell
Dystonia (laryngeal dystonia)	Laryngeal; stridor; strangled voice; breathy voice; quiet voice; whispery voice; hoarse; shaky voice; aphonia; interrupt speech; lose voice; voice loss
Tardive Dyskinesia (general)	tardive dyskinesia; dyskinesia; TD
Tardive Dyskinesia (orofacial dyskinesia)	Involuntary; tongue twist; tongue protrusion; chew; biting; bite; suck; clench; lateral jaw movement; sideways jaw movement; smack lip; lip purse; pucker; puff cheek; frown; blink; grimace; blink
Tardive Dyskinesia (limb truncal dyskinesia)	Choreiform; athetoid; choreoathetoid; purposeless; rock; twist; squirm; gyrate; thrust; knee move; tap; heel drop; writhing; rotate; nod; inversion; eversion
Akathisia	Akathisia; akathisia; restless; pace; fidget; irritable; leg cross; leg swing; foot shift; shuffle; tramp; still; march; shift weight; rock

4.2 Supplementary Table 2 Pharmacological Keywords used to Identify EPSE Cases

Medication	Keywords
Trihexyphenidyl	Trihex; benz; artane; agitane; parkin
Benzatropine	Benza; tropin; cogentin
Procyclidine	Procy; lidin; kemad
Orphenadrine	Orphan; drine
Biperiden	Biper; akineton
Hyoscine	Hyos; kwell
Tetrabenazine	Tetrab; ranb; nitom; xenaz

4.3 Supplementary Table 3 FGA Codes used to Select Participants

Medication Name	Medication Type	Medication Code
Benperidol	anquil 250micrograms tablet; benperidol	1140867080; 1140867078
Chlorpromazine	chlorpromazine; cpz - chlorpromazine; largactil 10mg tablet; chloractil 25mg tablet	1140879658; 1140910358; 1140863416; 1140863410
Flupentixol	flupentixol; depixol 3mg tablet; fluanxol 500micrograms tablet; flupenthixol; flupentixol	1140909800; 1140867152; 1140867952; 1140867150; 1140909800
Fluphenazine	decazate 25mg/1ml oily injection; fluphenazine; fluphenazine decanoate; modecate 12.5mg/0.5ml oily injection; moditen 1mg tablet; moditen enanthate 25mg/ml injection	1140867474; 1140882098; 1140867398; 1140867456; 1140867156; 1140856004
Haloperidol	haldol 5mg tablet; haloperidol; serenace 500micrograms capsule	1140867184; 1140867168; 1140867092
Levomepromazine	levomepromazine; nozinan 25mg tablet	1140909802; 1140867122
Loxapine	loxapine; loxapac 10mg capsule	1140867406; 1140867414
Pericyazine	neulactil 2.5mg tablet; pericyazine	1140867136; 1140867134
Perphenazine	fentazin 2mg tablet; perphenazine	1140867210; 1140867208
Pimozide	orap 2mg tablet; pimozide	1140867272; 1140867218
Pipotiazine	piportil depot 50mg/1ml oily injection; pipotiazine	1140867572; 1140909804
Prochlorperazine	prochlorperazine; stemetil 5mg tablet	1140868170; 1140868172
Promazine	promazine	1140879746
Sulpiride	dolmatil 200mg tablet; sulparex 200mg tablet; sulpiride; sulpitil 200mg tablet; sulpor 200mg/5ml oral solution	1140867306; 1140917366; 1140867304; 1140882376; 1141185130
Thioridazine	thioridazine; melleril 10mg tablet	1140879750; 1140867312
Trifluoperazine	stelazine 1mg tablet; tranylcypromine+trifluoperazine 10mg/1mg tablet; trifluoperazine	1140867244; 1140867944; 1140868120
Zuclopenthixol	clopixol 2mg tablet; zuclopenthixol	1140867342; 1140882100

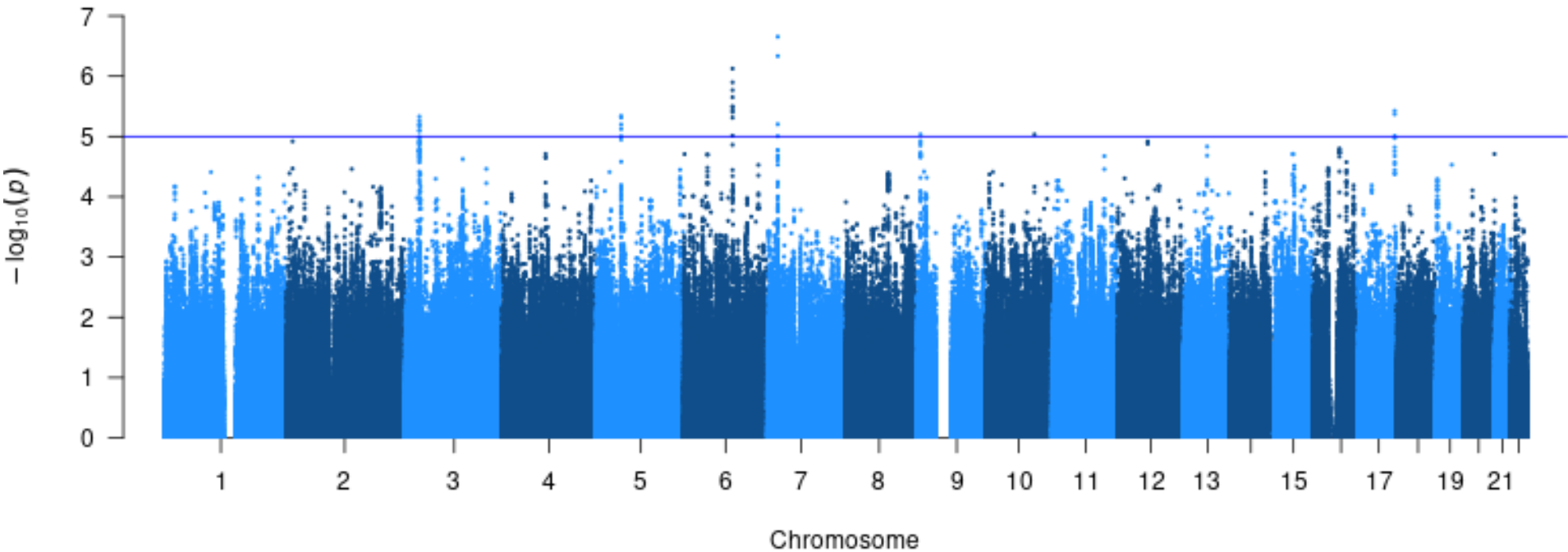
4.4 Supplementary Table 4. SGA Codes used to Select Participants

Medication Name	Medication Type	Medication Code
Amisulpride	amisulpride; solian 100mg/ml s/f oral solution	1141153490; 1141184742
Aripiprazole	abilify 5mg tablet; aripiprazole	1141202024; 1141195974
	clozapine; clozaril 25mg tablet; denzapine	1140867420; 1140882320;
Clozapine	25mg tablet	1141200458
Olanzapine	olanzapine; zyprexa 2.5mg tablet	1140928916; 1141167976
Oxypertine	oxypertine; integrin 10mg capsule	1140879754; 140855978
Quetiapine	quetiapine; seroquel 25mg tablet	1141152848; 1141152860
Remoxipride	remoxipride; roxiam 150mg m/r capsule	1140879704; 1140867432
	dozic 1mg/ml oral liquid; risperdal 0.5mg	1140867180; 1141177762;
Risperidone	tablet; risperidone	1140867444
Sertindole	serdolect 4mg tablet; sertindole	1140927970; 1140927956

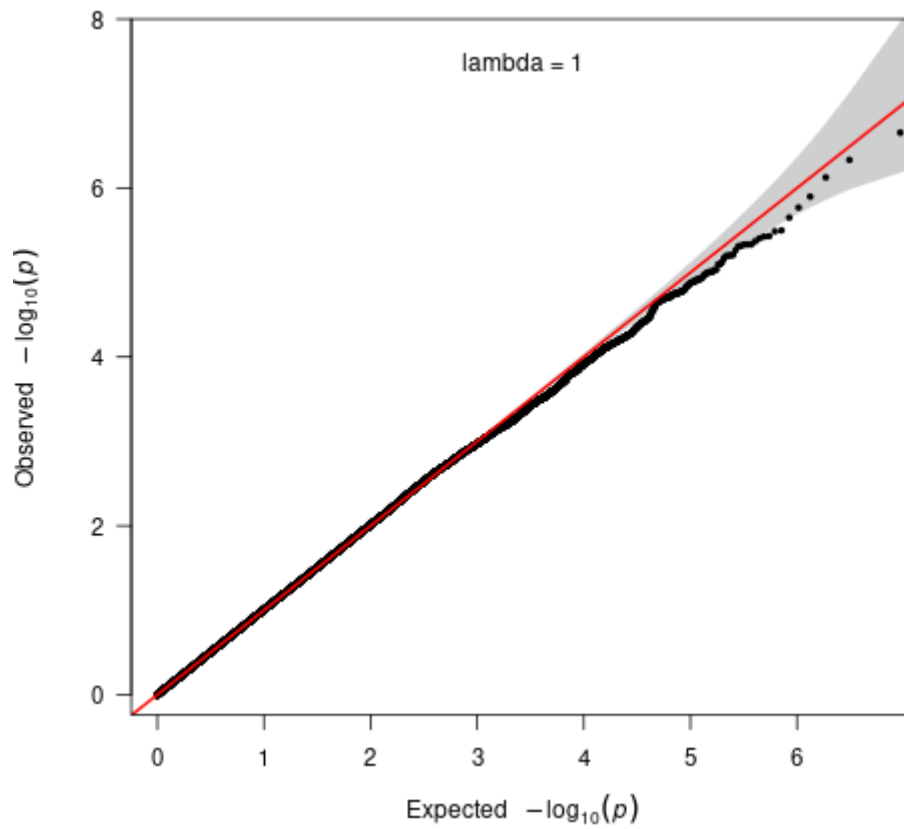
4.5 Supplementary Table 5 EPSE Medications Codes used to Select Cases

Medication Name	Medication Type	Medication Code
Akineton	akineton	1140872522
Artane	artane	1140872378
Benzatropine	benzatropine	1140909818
Benzhexol	benzhexol	1140883510
Biperiden	biperiden	1140872520
Cogentin	cogentin	1140872460
Kemadrin	kemadrin	1140872542
Orphenadrine	orphenadrine	1140883560
Procyclidine	procyclidine	1140883476
Tetrabenazine	tetrabenazine	1140872556
Tetrabenazine Product	tetrabenazine_product	1141157336
Trihexyphenidyl	trihexyphenidyl	1140909816
Xenazine	xenazine	1141171726

4.6 Supplementary Figure 1 The Manhattan Plot of EPSE GWAS Meta-analysis

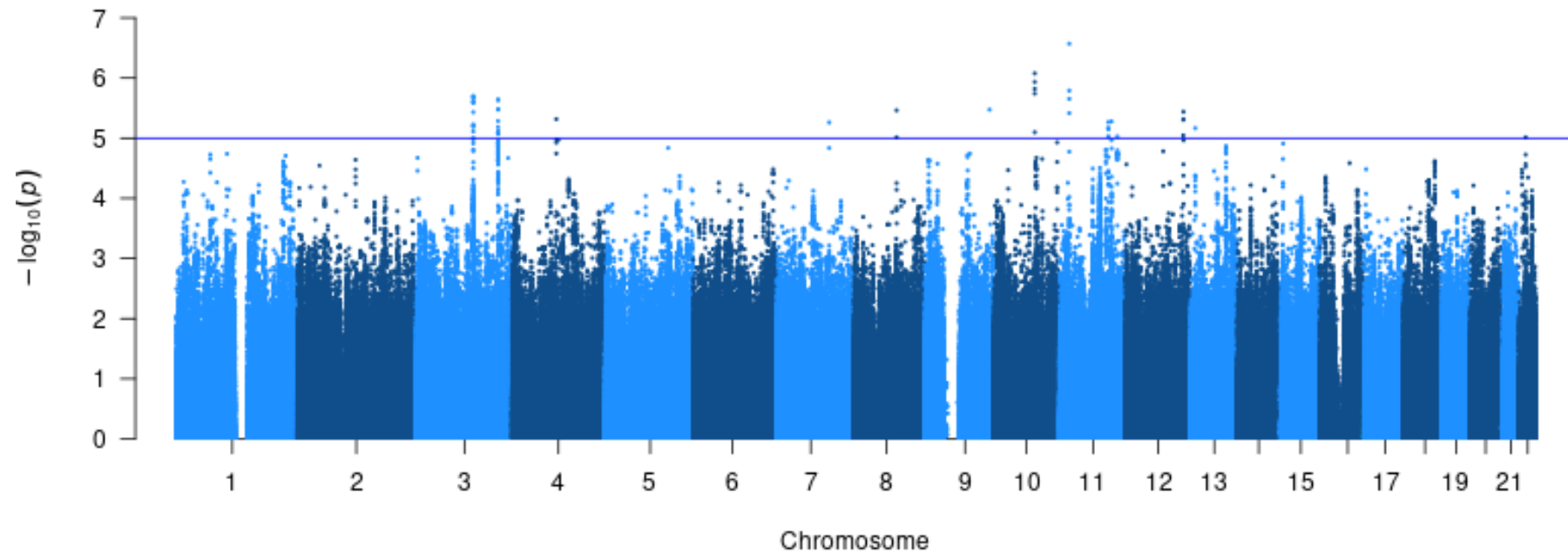


4.7 Supplementary Figure 2 The QQ Plot of EPSE GWAS Meta-analysis for FGA and SGA Combined

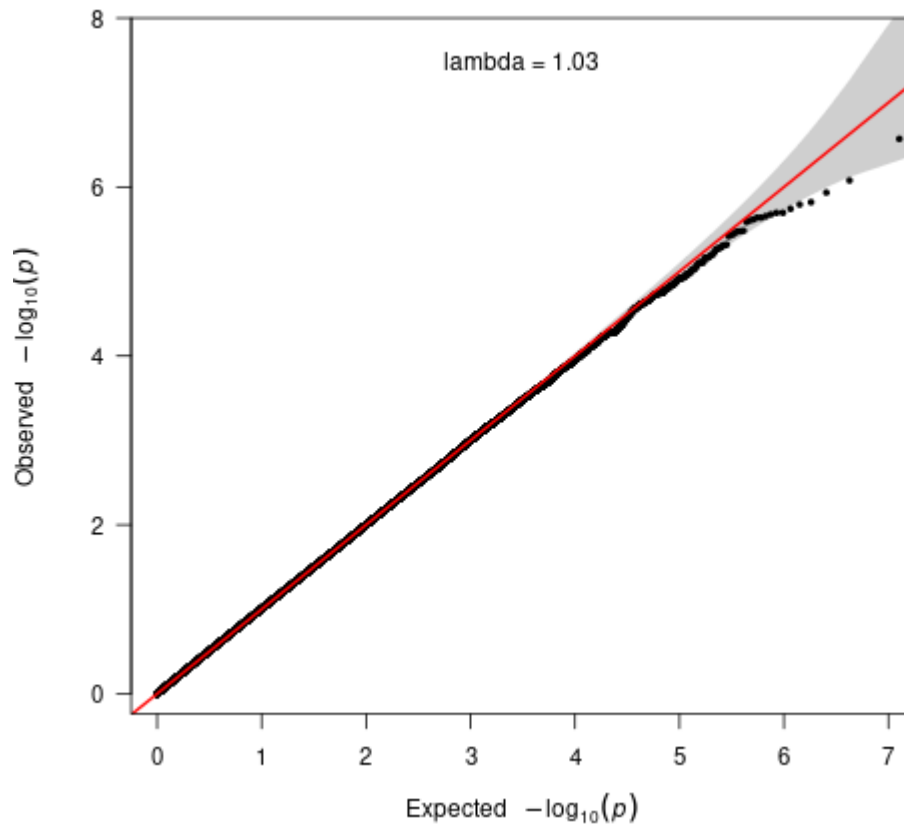


Notes. X-axis (Expected $-\log_{10}(p)$ -values)): Represents the theoretical quantiles under the null hypothesis, where p-values are uniformly distributed on a logarithmic scale; Y-axis (Observed $-\log_{10}(p)$ -values)): Shows the observed $-\log_{10}(p)$ -values from the GWAS.

4.8 Supplementary Figure 3 The Manhattan Plot of EPSE GWAS Meta-analysis for any Exposure to FGA

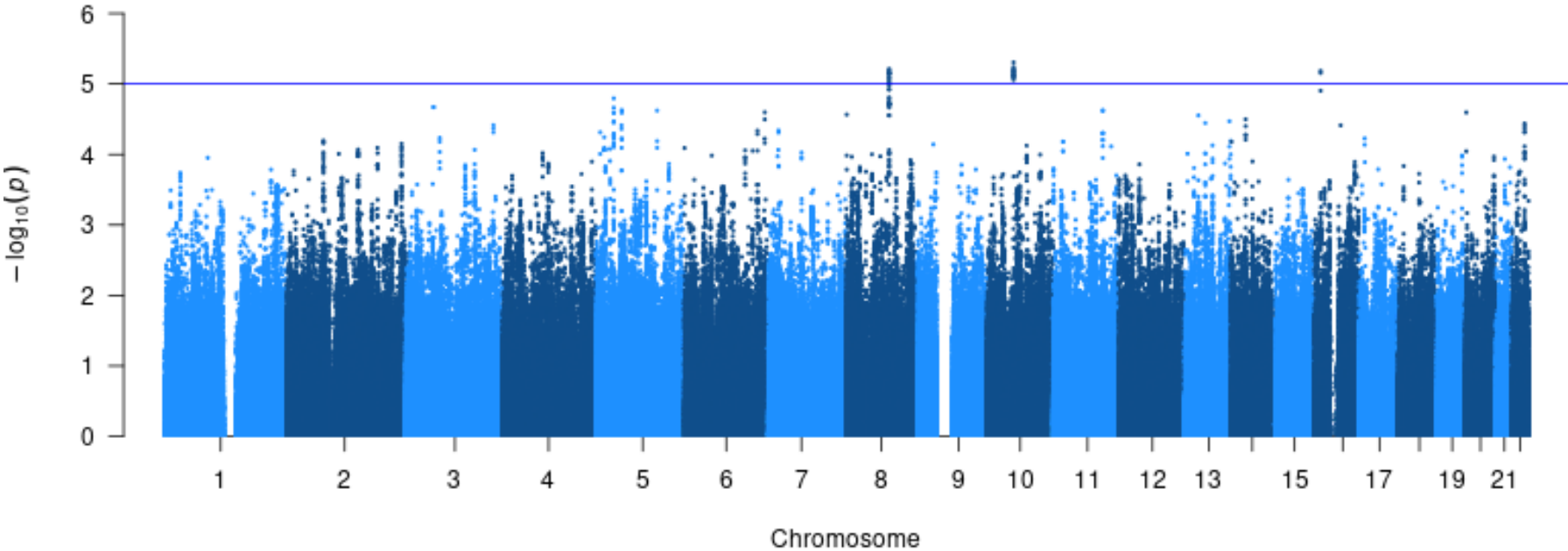


4.9 Supplementary Figure 4 The QQ Plot of EPSE GWAS Meta-analysis for any Exposure to FGA

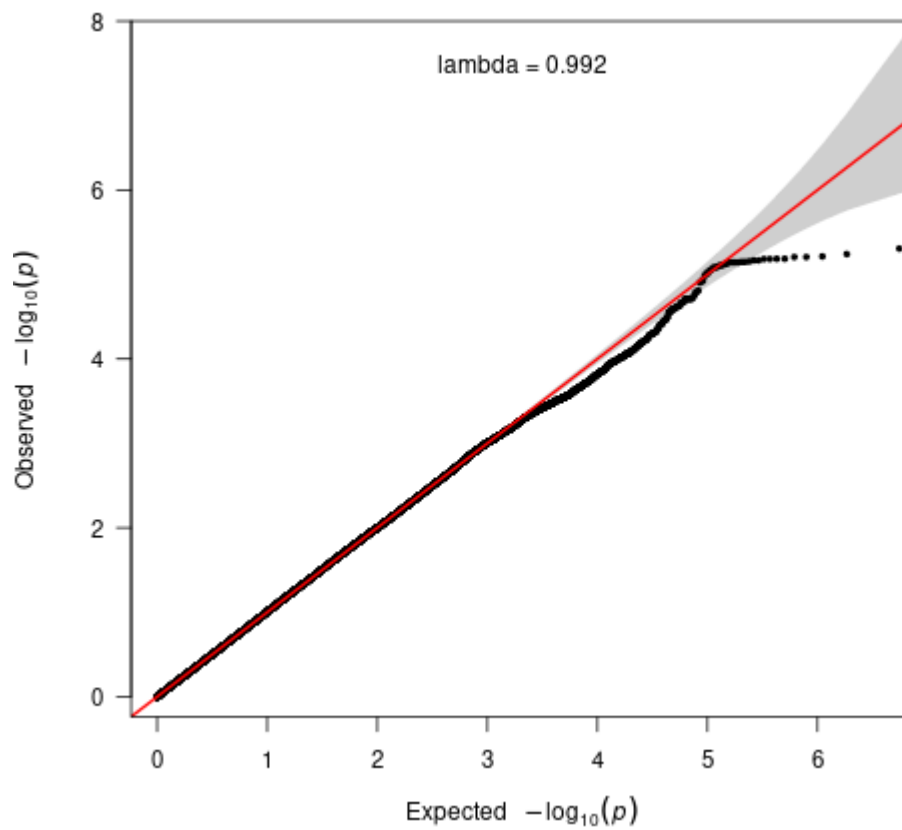


Notes. X-axis (Expected $-\log_{10}(\text{p-values})$): Represents the theoretical quantiles under the null hypothesis, where p-values are uniformly distributed on a logarithmic scale; Y-axis (Observed $-\log_{10}(\text{p-values})$): Shows the observed $-\log_{10}(\text{p-values})$ from the GWAS.

4.10 Supplementary Figure 5. The Manhattan Plot of EPSE GWAS Meta-analysis for only Exposure to SGA



4.11 Supplementary Figure 6 The QQ Plot of EPSE GWAS Meta-analysis for only Exposure to SGA



Notes. X-axis (Expected $-\log_{10}(\text{p-values})$): Represents the theoretical quantiles under the null hypothesis, where p-values are uniformly distributed on a logarithmic scale; Y-axis (Observed $-\log_{10}(\text{p-values})$): Shows the observed $-\log_{10}(\text{p-values})$ from the GWAS.

4.12 Supplementary Table 6 EWAS Participants' Demographics and Clinical Characteristics concerning EPSE Presence

	N	Overall	EPSE Presence	Controls	p-values
UCL	379		n = 64 (17%)	n = 315 (83%)	
Age at assessment	362	38.24 (14.81)	36.90 (14.74)	44.48 (13.58)	<0.001^a
mAge (Horvath)	379	40.31 (12.46)	39.54 (12.41)	44.08 (12.13)	0.008^a
Sex					<0.001^b
Male		192 (51%)	52 (81%)	140 (44%)	
Female		187 (49%)	12 (19%)	175 (56%)	
Antipsychotics	64				
First generation			57 (89%)	/	
Second generation			50 (78%)	/	
Aberdeen	480		n = 47 (10%)	n = 433 (90%)	
mAge (Horvath)	480	53.16 (9.88)	54.29 (11.48)	53.04 (9.70)	0.473 ^a
Sex	480				
Male		352 (73%)	33 (70%)	319 (74%)	0.737 ^b
Female		128 (27%)	14 (30%)	114 (36%)	
Antipsychotics	47				
First generation			30 (64%)	/	
Second generation			23 (49%)	/	

Notes. EPSE, extrapyramidal side effects; SD, standard deviation

^a Two Simple t-test; mean (SD)

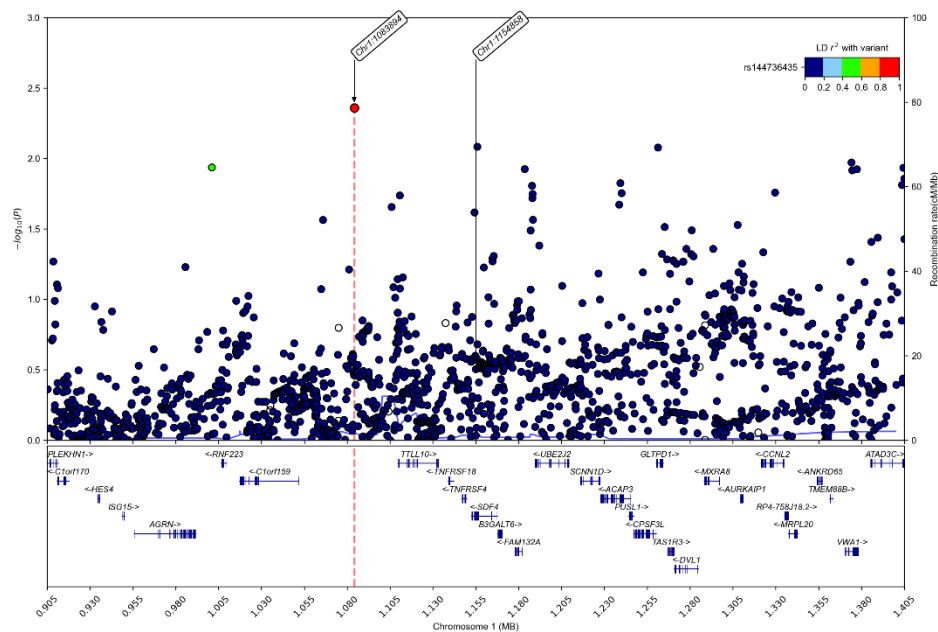
^b Pearson's Chi-squared test of independence; n (%)

In bold p passed significance threshold

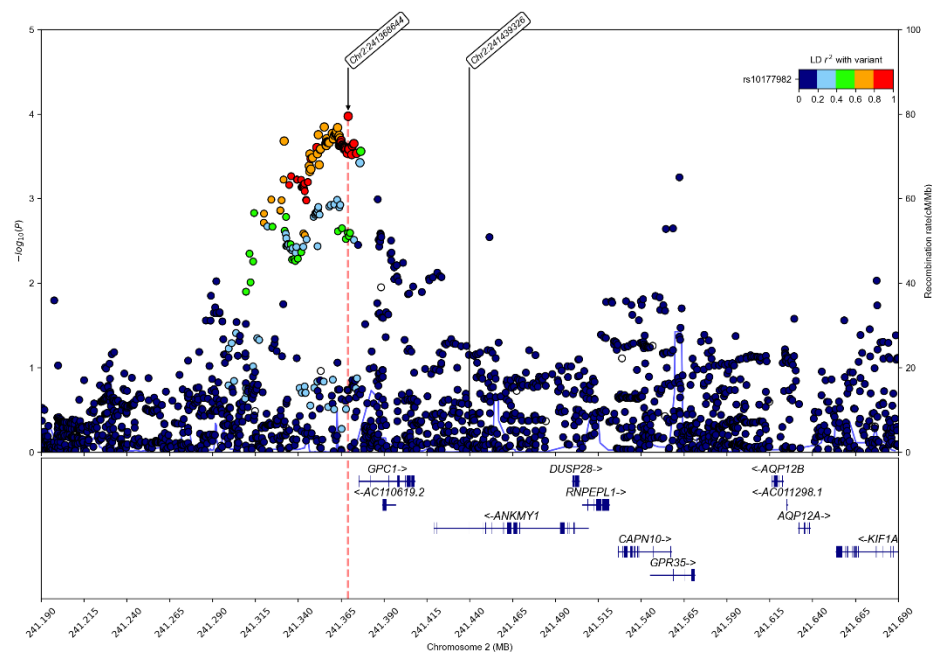
4.13 Supplementary Figures 7–15 The Mapping of CpG to the Schizophrenia GWAS Region

The figures illustrate the mapping of significant CpG sites to Schizophrenia GWAS within a 250kb window. Black line in the central mark the SNP closest to the CpG. Red dotted line marked the SNP in lead within the region and other nearby SNPs in linkage disequilibrium.

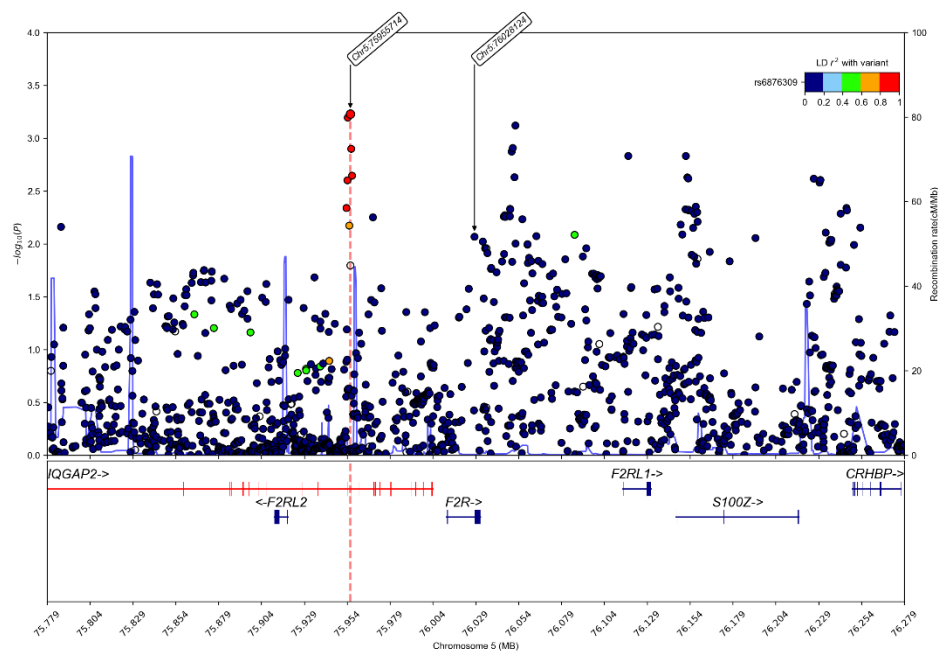
Supplementary Figure 7. The Mapping of CpG cg14531564 to the Schizophrenia GWAS Region



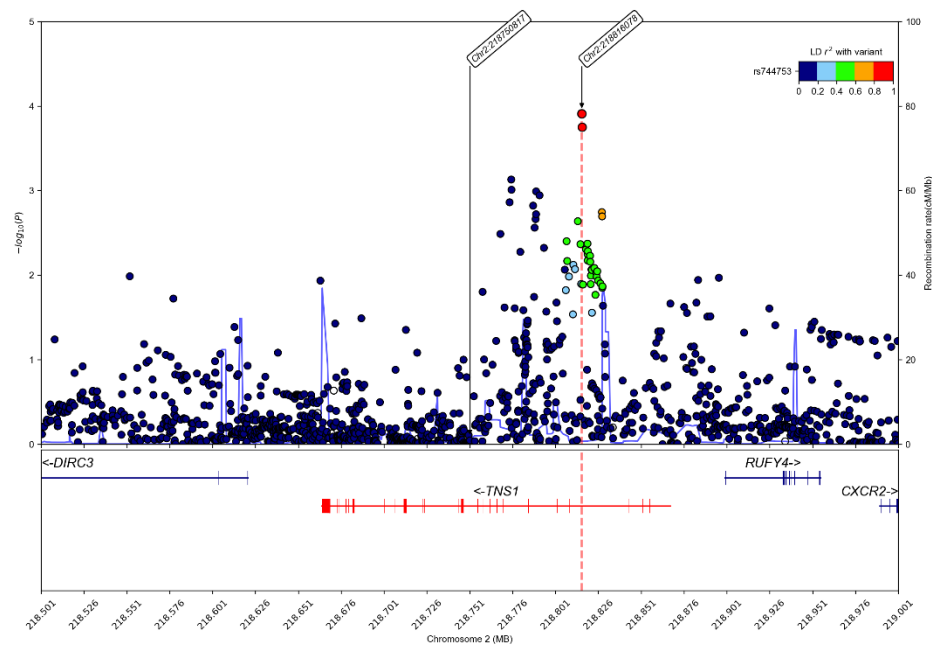
Supplementary Figure 8. The Mapping of CpG cg20647656 to the Schizophrenia GWAS Region



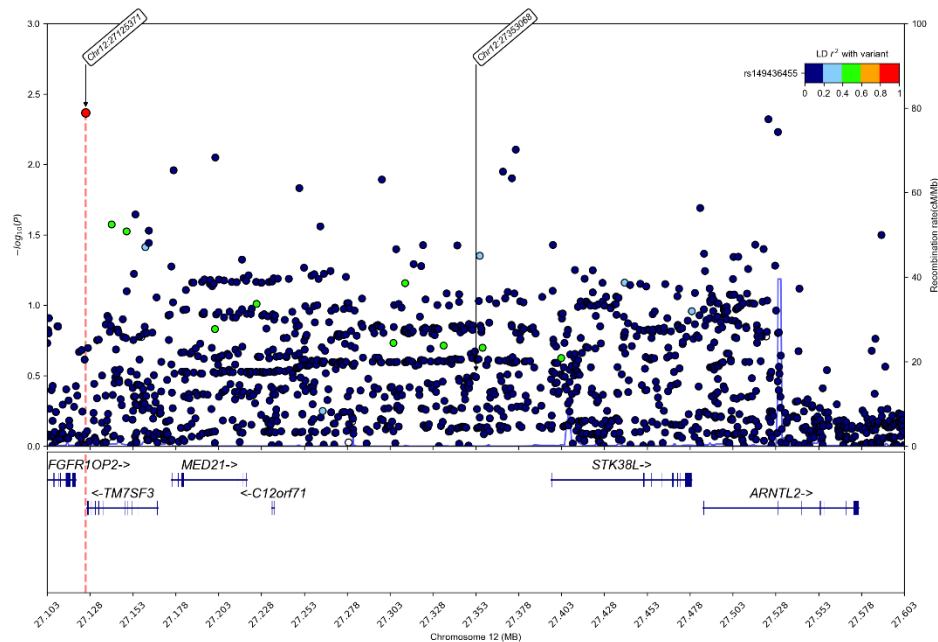
Supplementary Figure 9. The Mapping of CpG cg12524168 to the Schizophrenia GWAS Region



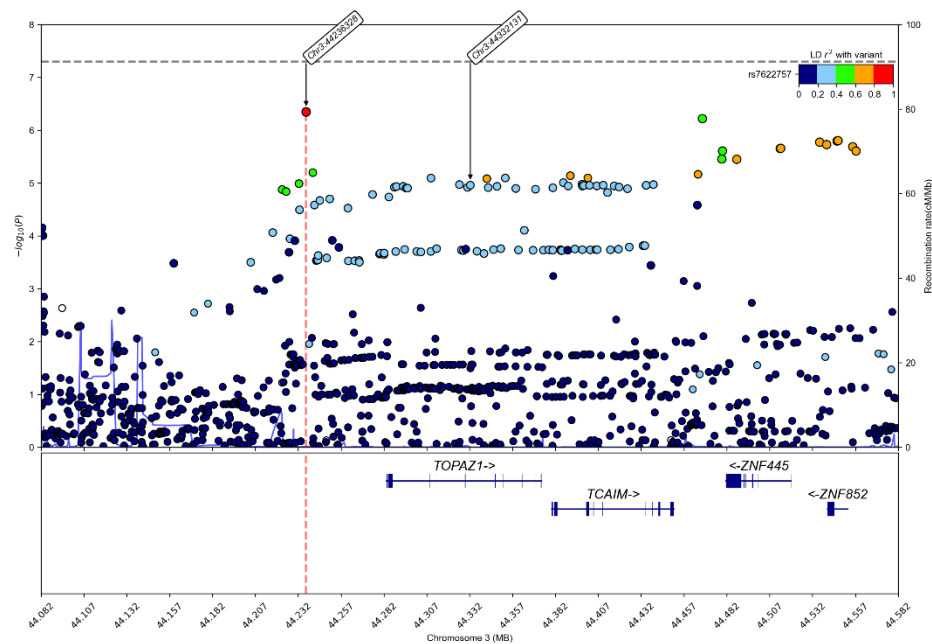
Supplementary Figure 10. The Mapping of CpG cg12004641 to the Schizophrenia GWAS Region



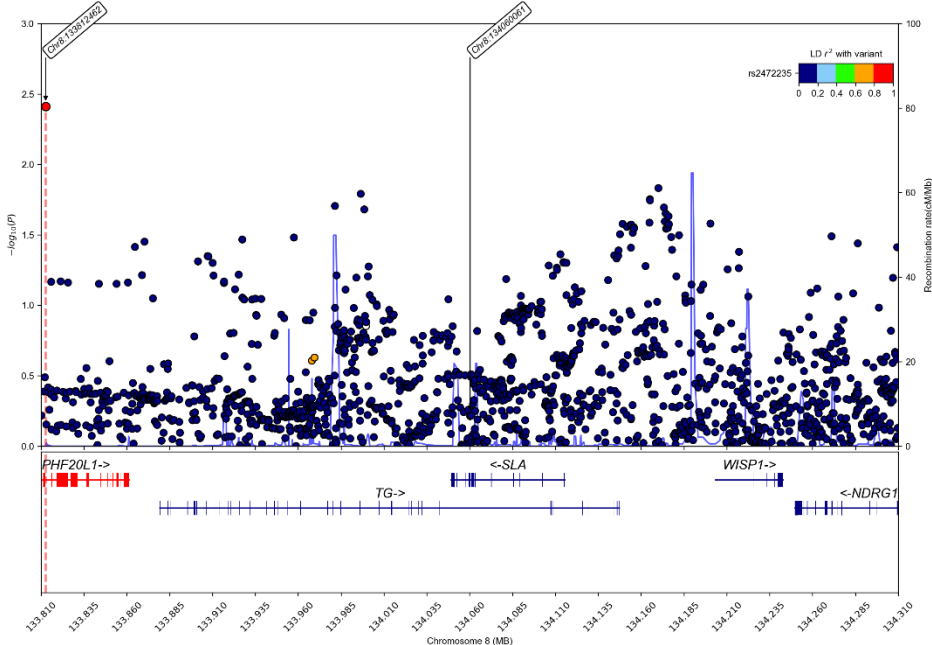
Supplementary Figure 11. The Mapping of CpG cg05419385 to the Schizophrenia GWAS Region



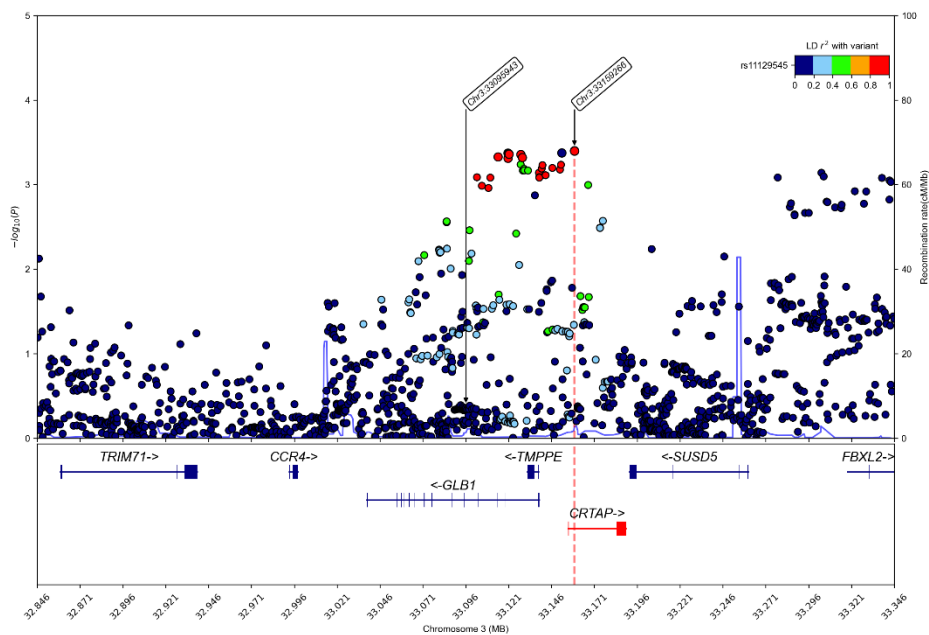
Supplementary Figure 12. The Mapping of CpG cg22583147 to the Schizophrenia GWAS Region



Supplementary Figure 13. The Mapping of CpG cg22845912 to the Schizophrenia GWAS Region



Supplementary Figure 14. The Mapping of CpG cg20730966 to the Schizophrenia GWAS Region



Supplementary Figure 15. The Mapping of CpG cg12044923 to the Schizophrenia GWAS Region

