

Genetic Diversity and Transmission Characteristics of Beijing Family Strains of *Mycobacterium tuberculosis* in Peru

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Abstract

Beijing family strains of *Mycobacterium tuberculosis* have attracted worldwide attention because of their wide geographical distribution and global emergence. Peru, which has a historical relationship with East Asia, is considered to be a hotspot for Beijing family strains in South America. We aimed to unveil the genetic diversity and transmission characteristics of the Beijing strains in Peru. A total of 200 Beijing family strains were identified from 2140 *M. tuberculosis* isolates obtained in Lima, Peru, between December 2008 and January 2010. Of them, 198 strains were classified into sublineages, on the basis of 10 sets of single nucleotide polymorphisms (SNPs). They were also subjected to variable number tandem-repeat (VNTR) typing using an international standard set of 15 loci (15-MIRU-VNTR) plus 9 additional loci optimized for Beijing strains. An additional 70 Beijing family strains, isolated between 1999 and 2006 in Lima, were also analyzed in order to make a longitudinal comparison. The Beijing family was the third largest spoligotyping clade in Peru. Its population structure, by SNP typing, was characterized by a high frequency of Sequence Type 10 (ST10), which belongs to a modern subfamily of Beijing strains (178/198, 89.9%). Twelve strains belonged to the ancient subfamily (ST3 [n = 3], ST25 [n = 1], ST19 [n = 8]). Overall, the polymorphic information content for each of the 24 loci values was low. The 24 loci VNTR showed a high clustering rate (80.3%) and a high recent transmission index ($RTI_{n-1} = 0.707$). These strongly suggest the active and on-going transmission of Beijing family strains in the survey area. Notably, 1 VNTR genotype was found to account for 43.9% of the strains. Comparisons with data from East Asia suggested the genotype emerged as a uniquely endemic clone in Peru. A longitudinal comparison revealed the genotype was present in Lima by 1999.

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Introduction

Strains of the Beijing family of *Mycobacterium tuberculosis* (*M. tuberculosis*), first described in 1995 [1], have attracted worldwide attention because of their wide geographical distribution and global emergence. These strains have also been shown to have an endemic prevalence in certain regions, including Asia, except for the Indian subcontinent; northern Eurasia; and South Africa [2,3,4,5,6,7]. The lineage has been reported to have caused major outbreaks worldwide, some involving drug-resistant variants [2,8,9]. These characteristics suggest that strains belonging to this family might have selective advantages (higher virulence or transmissibility) over other *M. tuberculosis* strains [10,11,12,13]. Therefore, a better understanding of the contribution of the

Beijing family to the tuberculosis (TB) pandemic is vital to improve global TB control.

The Beijing family is reported to be phylogenetically divisible into 2 main subfamilies: the modern (typical) and ancient (atypical) subfamilies, [14,15]. The modern subfamily is highly prevalent in China, western Russia, South Africa, and Thailand [3,15,16,17], whereas the ancient subfamily, with a deleted region of difference (RD) RD181[-] (late ancient type), is endemic in Japan and the ancient subfamily with RD181[+] (early ancient type) is endemic in Korea [4,18,19,20,21,22,23,24]. Although the reasons for the phylogeographical differences remain elusive, the above trends can be used to assess the influence of the Beijing family strains from East Asian countries, where the prevalence is very high [2,3], on the prevalence of Beijing family strains in other regions. In fact, a

large number of Chinese and Japanese immigrants settled in Peru in 19th century. Korea also has a history of migration to Peru, but this migration has occurred more recently. This historical relationship with East Asia encouraged us to characterize the Beijing family strains in Peru.

Unlike in other South American countries, the proportion of Beijing family strains of *M. tuberculosis* in Peru is exceptionally high [25]. In Brazil, Colombia, Paraguay, Venezuela, Argentina, Chile, and Ecuador the prevalence of Beijing family strains was reported to be less than 1% [25,26,27,28,29]. However, the prevalence in Peru was 5.9% (11/185) in 1999 [25] and 9.3% (30/323) for samples obtained between 2004 and 2006 [30]. Ritacco et al. [25] speculated that the Beijing family strains were first introduced into Peru, and eventually into other South American countries, when Peru received a significant number of Chinese immigrants in the mid-19th century. This same study also showed considerable diversity in the insertion sequence *IS6110* restriction fragment length polymorphism (RFLP) patterns, supporting the concept of earlier introduction(s) of different ancestral strains during the past 150 years. In addition to the importation of Beijing family strains from Asia to Peru, there is also evidence that the Beijing strains were imported into Europe through a South American route, specifically through Peru [31,32]. Therefore, Peru can be considered as the South American country that has been most strongly affected by the introduction of Beijing family and is also most commonly associated with the spread of these strains to other South American and European countries.

An in-depth analysis of Beijing family strains in Peru may have a significant impact on the understanding of global epidemics involving the *M. tuberculosis* Beijing family strains. The aims of the current study were to unveil the genetic diversity and transmission characteristics of Beijing family strains of *M. tuberculosis* in Lima, Peru, and to elucidate the probable impact of past immigration from East Asian countries.

Materials and Methods

Ethics Statement

Prior to the start of the study, ethical approval was obtained from both Universidad Peruana Cayetano Heredia and Imperial College London and institutional approval was obtained from the Peruvian Ministry of Health. Samples for this study were anonymized.

Study Samples

The Beijing family strains used in this study were identified on the basis of deletion of the spacers 1–34 assessed by the spoligotyping assay [33]. In order to negate the inclusion of “Pseudo-Beijing strains” [34], all of the strains classified as the early ancient type of Beijing family (RD 181[+]) were subjected to the RD 105 analysis [35].

A total of 200 Beijing family strains were identified from 2140 *M. tuberculosis* isolates obtained through a population-level implementation of a new diagnostic test (MODS, Microscopic Observation Drug Susceptibility) [36,37], which was conducted in Callao and South Lima between December 2008 and January 2010. Strains from all culture-positive patients with respiratory symptoms in the study area were included and none of the samples were duplicated from a single patient. Of the 200 Beijing family strains, 2 were excluded from this study because of insufficient DNA samples. For the remaining 198 Beijing strains, detailed information is provided in Tables 1 and S1.

An additional 70 Beijing family strains isolated in Lima between 1999 and 2006 were also analyzed in order to make a

longitudinal comparison. In detail, 26 were obtained from a previous study in North Lima between 2004 and 2006 [30] and 44 were obtained from 4 distinct areas in Lima (North, South, East, and Central Lima) through the hospital based studies between 1999 and 2004. The details of the strains are described in Tables 1 and S2.

VNTR Typing

Genotypic data for the 24 loci that comprised the international standard set of 15 loci of variable number of tandem repeat(s) of mycobacterial interspersed repetitive units (15-MIRU-VNTR) [38], and 9 additional loci (2074, 2372, 3155, 3336, 3232, 3820, 4120, QUB11a, QUB18) were analyzed. The 9 additional loci were selected as Beijing-type optimized loci because of their highly discriminatory values in different studies focused on Beijing family strains [4,24,39,40,41,42,43]. This 24-loci VNTR was called the 24_{Beijing}-VNTR. The polymerase chain reaction (PCR) primers and the number of repeats for each locus, based on the *M. tuberculosis* H37Rv strain, are described in Table S3; the PCR conditions were as described previously [4]. The amplicon samples were diluted 20-fold with ultrapure water and analyzed on an AB3500 genetic analyzer system (Applied Biosystems, Foster City, CA) at a constant room temperature of 25°C following the manufacturers’ instruction, i.e., capillary temperature, 60°C; electrophoresis voltage, 8.5 kV; and separation time, 5800 s. A GeneScan 1200 LIZ Size Standard (Applied Biosystems) was used to provide internal size markers. Fragment sizes were measured using GeneMapper Ver. 4 (Applied Biosystems). The numbers of repeats at each locus were calculated using the offset values of the size, which correct differences in relative migration between the size standard and the amplicons depending on the locus. The reproducibility and accuracy of size calling and the size offsets were checked by including *M. tuberculosis* H37Rv and 1 Beijing family strain (reference for quality control) into every batch of the analysis (one 96-well plate was used as a batch). For the large alleles (specifically, for the one larger than 1000 bases) of a locus, we used stutter peak counting, as shown in Figure S1, to obtain unambiguous results with high reproducibility [4]. Moreover, we confirmed the reproducibility of our assay by blindly re-testing 22 selected samples (Table S1). The allelic diversity of each VNTR locus was evaluated using Nei’s diversity index [44], i.e., the polymorphic information content (PIC) (Table S4). Genotypic discrimination of the 198 Beijing strains, based on 15-MIRU-VNTR and 24_{Beijing}-VNTR, were calculated using the Hunter-Gaston discriminatory index (HGDI) [45]. A recent transmission index (RTI_{n-1}) [46,47] was also calculated using the VNTR profiles. A minimum spanning tree (MST), based on VNTR types, was constructed using Bionumerics software (Bionumerics ver. 4.2; Applied Math., Sint-Martens-Latem, Belgium), as previously described [23].

Single Nucleotide Polymorphisms at 10 Loci

The sequence types (ST) were determined based on the 10 synonymous SNPs, which were sufficient to divide the Beijing strains obtained from the global population [48]. Each chromosomal position in the whole-genome sequence of H37Rv [49] was as follows: 797736, 909166, 1477596, 1548149, 1692069, 1892017, 2376135, 2532616, 2825581, and 4137829. The SNP at position 1477596 is the same as *ogt12*, which discriminates between the ancient and modern type Beijing strains [16]. The SNP at 2532616 is the same as *adhE2* (codon 124), which further discriminates the modern type [50]. SNPs at 797736 and 2825581

Table 1. Patient demographics for the Beijing family strains in this study.

| Characteristics | No. (%) of isolates | |
|-----------------------------------|-----------------------|-----------------------|
| | Between 2008 and 2010 | Between 1999 and 2006 |
| Total | 198 (100) | 70 (100) |
| Sex | | |
| Male | 133 (67) | 45 (64) |
| Female | 63 (32) | 25 (36) |
| Unknown | 2 (1) | 0 (0) |
| Age group | | |
| <25 | 85 (43) | 19 (27) |
| 25–34 | 45 (23) | 17 (24) |
| 35–44 | 32 (16) | 8 (11) |
| 45–54 | 10 (5) | 3 (4) |
| 55–64 | 5 (3) | 0 (0) |
| 65+ | 8 (4) | 2 (3) |
| Unknown | 13 (7) | 21 (30) |
| Previous TB | | |
| Yes | 55 (28) | ND |
| No | 141 (71) | ND |
| Unknown | 2 (1) | 70 (100) |
| HIV status | | |
| Positive | 9 (5) | 9 (13) |
| Negative | 188 (95) | 35 (50) |
| Unknown | 1 (1) | 26 (37) |
| <i>M. tuberculosis</i> resistance | | |
| MDR | 17 (9) | 10 (14) |
| not MDR | 177 (89) | 47 (67) |
| Unknown | 4 (2) | 13 (19) |

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can discriminate between the early ancient (RD181[+]) and late ancient (RD181[-]) Beijing strains [19]. Polymorphic nucleotides of the respective isolates were determined according to Hanekom et al. [17]. STs were designated according to Filliol et al. [48] and Iwamoto et al. [19].

Data Retrieved from Previous Reports

15-MIRU-VNTR data and the results of subfamily classifications were retrieved from previous publications for the isolates in China [50], Japan [18], and Korea [20]. In order to compare the population structure, based on Filliol's STs, further sublineage classifications for the Chinese and Korean isolates were determined according to the phylogenetically informative VNTR loci data proposed by Wada and Iwamoto [51]. The VNTR data from Peru and these 3 East Asian countries were compared by constructing MST, based on 15-VNTR-MIRU, as previously described [23].

Statistical Analysis

Fisher's exact test (SPSS 17.0; IBM, NY, USA) was used to determine the association of large cluster-forming isolates with patient gender, history of TB treatment, HIV status, and strain drug susceptibility; the association with the median age of patients was evaluated by Mann-Whitney *U*-test using PASW statistics 18 (IBM).

Results

Proportion of Beijing Family Strains in Peru

The Beijing family strain, with a prevalence of 9.3% (200/2140), was the third largest spoligotyping clade after the H3 and T1 clades in the study population sampled between 2008 and 2010. This ratio is the same as for the previous study (9.3%, 30/323) conducted in 2004 and 2006 [30] but higher than in 1999 (5.9%, 11/185) [25]. In another study setting, which collected Beijing strains between 1999 and 2004 from 4 distinct areas in Lima, the ratio was 5% (46/912, 2 Beijing strains were excluded in this study because of the lack of DNA). This would imply an increasing trend of Beijing family strains in Lima, Peru. We negated the existence of "Pseudo-Beijing strains" [34] in our sample set by the detection of RD 105 [-] [35] for all of the strains classified as early ancient type by SNP typing.

Classification of Beijing Family Strains by SNP Typing and MIRU-VNTR

Further subdivision of the Beijing family strains in Peru, according to the 10 loci SNP panel [17,19,48], revealed similar population structures between the 2 sample sets (2008–2010 and 1999–2006) (Table 2). The high prevalence of isolates associated with the modern subfamily in Peru is consistent with the worldwide trend, except in the case of Japan and Korea where

the ancient subfamily predominates [20,50,51,52,53,54]. When we compared the population structure of the Beijing family strains in Peru with China, Japan, and Korea (Table 2), the characteristics of the Peruvian population structure highlighted a high frequency of the ST10 sublineage. ST19/ST25, which evolved before the modern subfamily [16,51], predominated in the ancient subfamily, as has been seen in China. Recently, Wada et al. proposed the use of SNP at 1576481 instead of 909166 due to the probable homoplastic behavior of SNP 909166 [55]. This is especially critical for discriminating between STK and ST3, which are highly prevalent in Japan. We, therefore, applied it for 4 ST3 strains in our sample set and confirmed them as true ST3 sublineage. Interestingly, the phylogenetic informativity of certain VNTR loci reported in Asian strains [16,51] was also retained for the strains in Peru (Table 3). Specifically, VNTR 4156, 1955, and 3155 demonstrated high sensitivity and specificity for the sublineages classification.

The 15-MIRU-VNTR data from the modern subfamily of Beijing strains in China, Korea, and Japan were retrieved from previous reports [18,20,50] and compared with the data of the population-based study in Peru (2008–2010) (Fig. 1). Two extremely large clusters were found in the MST, one ($n = 103$) composed of almost exclusively Peruvian isolates (101 Peruvian isolates and 2 Japanese isolates) (C1 in Fig. 1) and the other ($n = 88$) composed of isolates from all of the 4 countries (C2 in Fig. 1). None of the large clusters were composed exclusively of Japanese, Chinese, or Korean isolates. The biggest cluster, C1, did not show a star-like network with the others; rather, it had a nearly terminal topological position. In addition, the VNTR profile could not be found in the MIRU-VNTR_{plus} database [56] or in other reports from Russia and China [57,58]. Together, these observations suggested that this genotype emerged as a uniquely endemic clone in Peru. The other large cluster (C2) was found at the core position and was connected with many isolates, regardless of their geographical origin. Because the Peruvian isolates belonging to the C2 cluster were further subdivided into many different genotypes by the 24_{Beijing}-VNTR analysis (data not shown), it is clear that the large cluster was formed because of the convergence of VNTR profiles. Other than these 2 large clusters, all of the other isolates dispersed equally (Fig. 1), i.e., no other branches consisted solely of Peruvian isolates.

Allelic Diversity of 24 VNTR Loci in Peru

The allelic diversity of each of the 24 VNTR loci is listed in Table S4, including the data from a previous report for Russia, Japan, and China [43]. Overall, the allelic diversity of the VNTR loci in Peru was much lower than in China or Japan. This low diversity was similar to that observed in Russia, where the Beijing family strains are considered to have recently emerged [57,59]. Only 11 out of the 24 loci had a PIC value greater than 0.1. Of these loci, 7 were in the 9 additional VNTR loci described in the Methods section. The PIC for the 3 hypervariable loci [4] were high: VNTR 3232 (0.710), VNTR 3820 (0.592), and VNTR 4120 (0.495). These results clearly supported the necessity of the use of the 9 additional loci for improving the discriminating power of VNTR genotyping for these strains. As expected, the use of 24_{Beijing}-VNTR improved discrimination compared to that of 15-MIRU-VNTR (Table 4).

Transmission Characteristics of Beijing Family Strains

Although 24_{Beijing}-VNTR improved the power of strain discrimination, the results showed high levels of clustering (80.3%) and RTI_{n-1} (0.707) (Table 4). This strongly suggests the active and on-going transmission of Beijing family strains in the survey area. Notably, a large size clustering (43.9%, 87/198) was identified, which was named Peru Cluster Type 001 (PCT001) (Fig. 2). The PCT001 strains belong to the largest cluster (C1) in 15 MIRU-VNTR (Fig. 1), therefore, they are considered a singular genotype in Peru. In the MST (Fig. 2), it formed a star-like VNTR-based network with the probable derivatives from it, mainly resulting from single locus changes. The results suggest the continuous evolution of the clone through active transmission and thus being a “currently successful clone” of the Beijing family strains in Peru. The second and third largest clusters were much smaller than PCT001: 15 isolates (7.6%) and 10 isolates (5.1%), respectively. Although the number of multidrug-resistant *M. tuberculosis* (MDR-TB) belonging to the PCT001 cluster is small at the moment ($n = 6$), this should be addressed as a potential public health threat. The comparison between patients harboring the PCT001 strains and those of other patients harboring Beijing strains revealed that only gender is significantly different ($P = 0.02$) (Table 5).

To ascertain whether the PCT001 genotype strains were present before the surveillance periods (in 2008–2010), an MST

Table 2. Distribution of Beijing sublineage strains.

| SNP type | No. (%) of isolates | | | | Definition ² | |
|----------|---------------------|--------------------|--------------------|--------------------|-------------------------|--------------------|
| | Peru | | Japan | China ¹ | | Korea ¹ |
| | 2008–2010 (n = 198) | 1999–2006 (n = 70) | ref [18] (n = 714) | ref [50] (n = 187) | | ref [20] (n = 62) |
| ST11 | 0 (0) | 0 (0) | 3 (0.4) | 9 (4.8) | 29 (46.8) | Early ancient |
| ST26 | 0 (0) | 4 (5.7) | 50 (7.0) | | | |
| STK | 0 (0) | 0 (0) | 111 (15.5) | 0 | 2 (3.2) | Late ancient |
| ST3 | 3 (1.5) | 1 (1.4) | 182 (25.5) | 3 (1.6) | 3 (4.8) | |
| ST25 | 1 (0.5) | 0 (0) | 6 (0.8) | 52 (27.8) | 10 (16.1) | |
| ST19 | 8 (4.0) | 5 (7.1) | 195 (27.3) | | | |
| ST10 | 178 (89.9) | 53 (75.7) | 135 (18.9) | 93 (49.7) | 18 (29.0) | Modern |
| ST22 | 8 (4.0) | 7 (10.0) | 32 (4.5) | 30 (16.0) | | |

¹Two isolates in Korea and 4 isolates in China could not be assigned sublineages and excluded from the analysis.

²Early ancient, RD181 [+]; Late ancient, RD181 [–].

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Table 3. Specific VNTR allele for Beijing sublineages *M. tuberculosis* in Peru (n = 268).

| VNTR locus | Specific allele | Corresponding sublineage(s) | No. of isolates | Sensitivity (%) | Specificity (%) |
|------------|-----------------|-----------------------------|-----------------|-----------------|-----------------|
| 4156 | 3 | Modern | 246 | 246/246 (100) | 246/248 (99.2) |
| | 4 | Early Ancient | 4 | 4/4 (100) | 4/4 (100) |
| | 5 | Late Ancient | 16 | 16/16 (100) | 16/18 (88.9) |
| 1955 | 4 | Modern | 246 | 235/246 (95.5) | 235/235 (100) |
| 3155 | 2 | Early Ancient | 4 | 4/4 (100) | 4/4 (100) |

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was constructed based on the 24_{Beijing}-VNTR profiles of the 268 Beijing family strains, comprised of the 198 strains (2008–2010) and the additional 70 strains isolated between 1999 and 2006 (Fig. 3). Twelve strains, which formed the second largest cluster among the 70 Beijing family strains, belonged to PCT001 (1 in 1999, 2 in 2000, 4 in 2001, 2 in 2004, and 3 in 2005), indicating that the genotype had existed since at least 1999. A striking difference between the 2 sample sets was found in the ratio of PCT001 genotype strains to total Beijing strains, i.e., a larger cluster (13 strains) than PCT001 was found in the 70 Beijing strains (PCT002 in Fig. 3). When we just focused on the strains isolated in South Lima (24/70 [34.3%]), the overlapping area of these 2 sample sets, PCT002 was still identified as the largest

cluster (7 strains) followed by PCT001 (6 strains). These results suggested that the prevalence of PCT001 in the survey area increased over a short period.

Discussion

The high prevalence of Beijing family strains in Peru (5.9%, 11/185) compared to other South American countries was first reported as the result of a survey of 7 countries on the continent [25]. Very recently, Taype et al. [30] reported that the proportion of TB patients with the Beijing family strains in Peru was 9.3% (30/323). The report nicely described the genetic diversity of all the *M. tuberculosis* in Peru. However, the population size was too

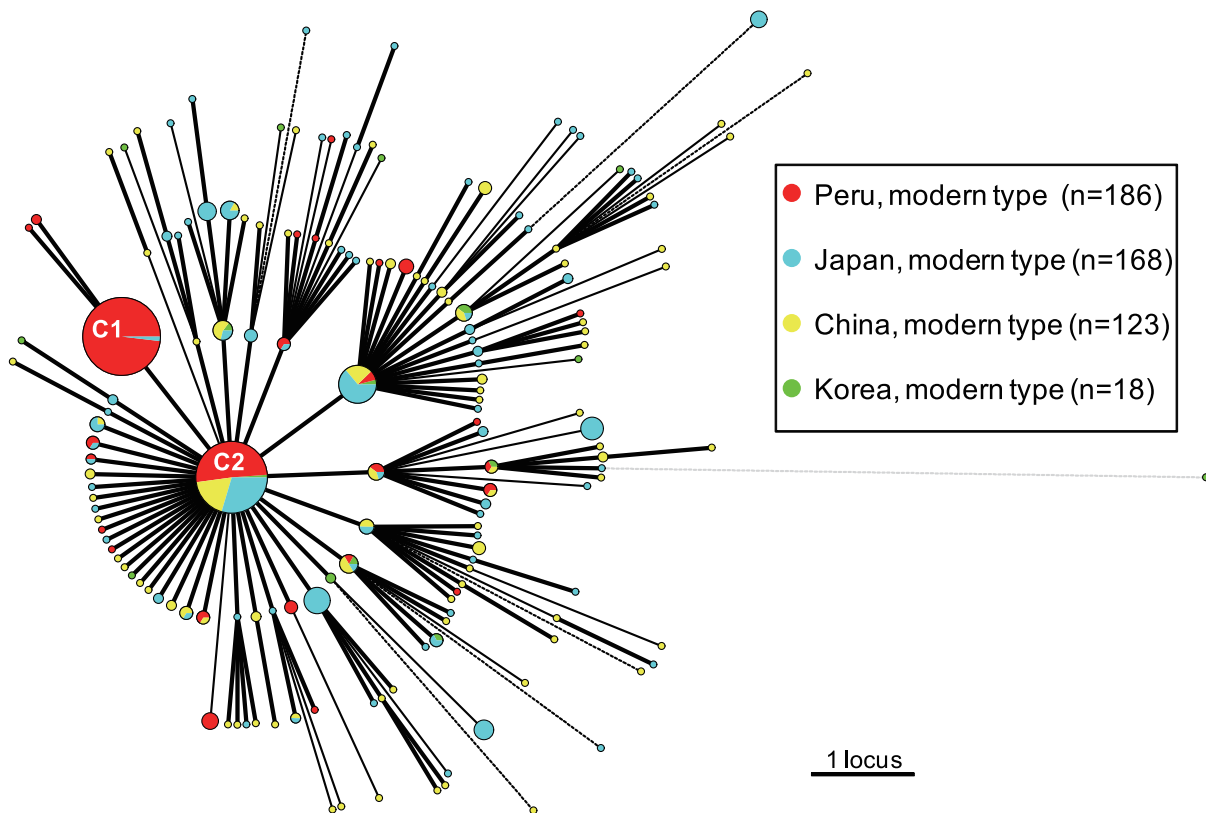


Figure 1. A minimum spanning tree based on 15 loci of a variable number of tandem repeat(s) of mycobacterial interspersed repetitive units (15-MIRU-VNTR) genotyping of the modern subfamily of *M. tuberculosis* Beijing strains from Peru (n = 186), Japan (n = 168), China (n = 123), and Korea (n = 18). Circles correspond to the different types discriminated by 15-MIRU-VNTR genotypes. Their sizes are proportional to the numbers of isolates sharing an identical pattern. The origin of each isolate is represented by different colors. Heavy lines connecting 2 types denote single-locus variants; thin lines connect double-locus variants; and dotted lines (black), triple-locus variants. The gray dotted lines indicate the most likely connection between 2 types differing by more than 3 VNTR loci.

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Table 4. Clustering analysis of 198 Beijing family strains (2008–2010) in Peru.

| | No. of patterns | No. of clusters | No. clustered isolates | Clustering rate (%) | RTI _{n-1} | HGDI |
|---------|-----------------|-----------------|------------------------|---------------------|--------------------|-------|
| 15 VNTR | 31 | 14 | 181 | 91.4 | 0.843 | 0.688 |
| 24 VNTR | 58 | 19 | 159 | 80.3 | 0.707 | 0.797 |

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small to investigate the Beijing family strains in detail. In the current study, with a much larger population size and a well-designed sample collection scheme between 2008 and 2010, the genetic diversity and transmission dynamics of Beijing strains in Peru was clarified. The results highlighted the following: 1) an increasing prevalence of Beijing family strains during a past decade in Peru, 2) high clonality of the Beijing family strains, suggesting active and ongoing transmission, 3) the successful clone “PCT001 genotype,” which has existed since at least 1999 as a singular clone in Peru, and 4) China as the greatest contributor of imported Beijing family strains into Peru. This study is limited by being unable to determine whether the highly active, ongoing transmission of Beijing family strains is specific for the family or a common trend for all strains in Peru. However, we could find a similarly high clustering rate of Beijing family strains (24/30 [80%]), compared with that of the non-Beijing family strains (174/293 [59.4%]) in the previous study using 12-MIRU-VNTR and spoligotyping [30]. This higher clustering in Beijing family strains implies a high transmission of it in Peru (more successful than non-Beijing strains). This trend could be confirmed later on by further VNTR analysis of all of the 2140 isolates.

A longitudinal comparison with the 70-strain sample set collected between 1999 and 2006 suggested that the successful clone, PCT001, was already present in Lima, by 1999, but at a lower prevalence ($n = 12$ [17.1%]) (Fig. 3). This difference in the ratio of PCT001 genotype strains to the total Beijing strains between the 2 sample sets suggests that the increase in PCT001 prevalence occurred recently, over a relatively short period. HIV infection, one of the high-risk characteristics for a large-scale outbreak [2], did not explain this high prevalence of PCT001 strains in the survey area (Table 5). One of the possible explanations for the high prevalence of PCT001 strains could be that it is highly transmissible and/or has increased virulence. The highly prevalent strains from large clusters have been previously reported to be more virulent than the sporadic strains of lower prevalence [59,60]. An attractive hypothesis is that PCT001 strains gained a selective advantage that allowed them to have spread more easily between the 2 sample collection periods (1999–2006 and 2008–2010).

The 15-MIRU-VNTR data from the modern subfamily of Beijing strains in China, Japan, and Korea were retrieved from previous reports [18,20,50] for comparison with the Peruvian isolates. The topology of the MST, based on these data (Fig. 1),

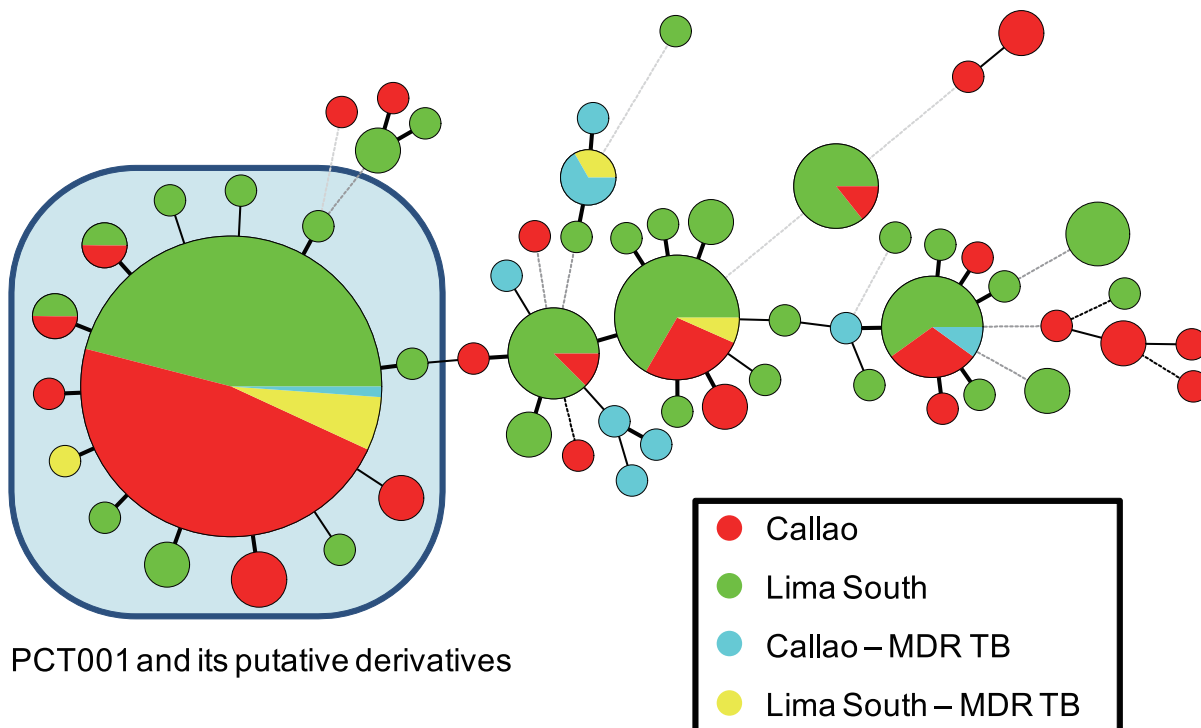


Figure 2. A minimum spanning tree of 198 Beijing family strains from Peru based on the 24-loci variable number of tandem repeats (VNTR). The colors of the circles represent the areas where the strain was isolated and its multidrug-resistant status. The designations for each circle and line in the tree are the same as in Fig. 1.

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Table 5. Demographics of successful clone “PCT001” and other strains.

| Variable | No. of strains (%) | | P value |
|--------------------|--------------------|------------------|---------|
| | PCT001 (n = 87) | Others (n = 111) | |
| Gender | | | |
| Male | 67 (77) | 66 (59) | 0.02 |
| Female | 20 (23) | 43 (39) | |
| Unknown | 0 (0) | 2 (2) | |
| Median age (range) | 23.5 (12–69) | 27 (12–83) | 0.07 |
| Unknown | 4 (5) | 9 (8) | |
| Previous TB | | | 1 |
| Yes | 24 (28) | 31 (28) | |
| No | 63 (72) | 78 (70) | |
| Unknown | 0 (0) | 2 (2) | |
| HIV status | | | 0.30 |
| Positive | 2 (2) | 7 (6) | |
| Negative | 85 (98) | 103 (93) | |
| Unknown | 0 (0) | 1 (1) | |
| MDR TB | | | 0.45 |
| Yes | 6 (7) | 11 (10) | |
| No | 81 (93) | 96 (86) | |
| Unknown | 0 (0) | 4 (4) | |

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suggested that the PCT001 genotype was a uniquely endemic clone in Peru. Except for the 2 largest clusters, the VNTR patterns from these countries were similarly diverse and none of the branches was composed exclusively of Peruvian isolates. This result would further support the earlier introduction of

different ancestral strains into Peru from Asia during the past 150 years, an idea that was proposed on the basis of the diversity in IS6110 RFLP patterns in a previous study [25].

The 10 loci SNPs could identify the modern Beijing lineage ST10 as the predominant sublineage in Peru. Very recently, this sublineage was reported as the most common in Taiwan and Thailand [16,61]. Allelic distribution of VNTR loci in each sequence type of Peruvian samples revealed the phylogenetic informativity of 3 MIRU-VNTR loci (4156, 1955, 3155) (Table 3). This is consistent with the results for East Asian strains [51] and could be an evidence for sharing the common ancestors of the Beijing family strains in Peru with those in East Asian countries. With the tremendous increase in whole-genome sequencing data, an increasing number of SNP typing systems have been developed [54,62,63]. The 10 loci SNPs typing is useful for a classification of Beijing family strains into sublineages level but apparently needed more discriminatory power to compensate for the VNTR homoplasmy effect (VNTR-based clusters with mixed SNP sublineages), which was demonstrated with optical sets of 8 SNPs in Shanghai [50]. Moreover, inclusion of the lineage specific SNPs for the clades other than Beijing family [62,63] would expand the potential for phylogenetic studies. Further elaboration and optimization of the SNP sets would facilitate future molecular epidemiology and phylogenetic studies on *M. tuberculosis* in Peru.

In conclusion, the current results revealed the predominance of the modern subfamily and active transmission of Beijing family strains within Peru. Moreover, the emergence of the highly prevalent strains with the PCT001 genotype was also detected. The importation of Beijing family strains into European countries from Peru has already been reported [31,32], raising concern over transnational transmission. Future trends regarding the prevalence of PCT001 strains and the changes in population structure need to be carefully monitored from both the local and global epidemiological standpoints.

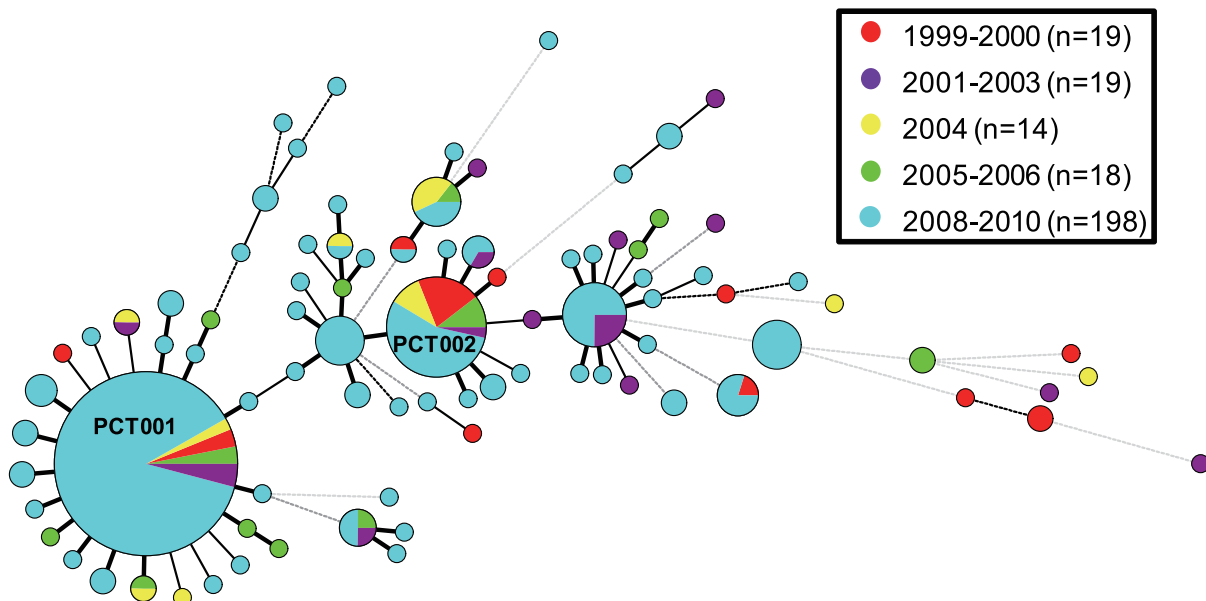


Figure 3. A minimum spanning tree of 268 Beijing family strains comprised of the 198 strains from the population-based study between 2008 and 2010 and an additional 70 strains isolated between 1999 and 2006. The colors of the circles represent the years of isolation. The designations for the circles and lines in the tree are the same as in Fig. 1.

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Supporting Information

Figure S1 Quality control approach for VNTR.
(PDF)

Table S1 Description of *M. tuberculosis* Beijing family strains obtained between December 2008 and January 2010 in Lima.
(XLS)

Table S2 Description of the additional 70 *M. tuberculosis* Beijing family strains in Lima, Peru.
(XLS)

Table S3 Locus designations and PCR primer sequences of the VNTR locus.

(XLS)

Table S4 Allelic diversity of VNTR loci.
(XLS)

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Author Contributions

Conceived and designed the experiments: TI LG DM RG. Performed the experiments: TI LG KA NN JC LC PS. Analyzed the data: TI LG KA NN RG. Contributed reagents/materials/analysis tools: JC LC PS TW CT MS DM RG. Wrote the paper: TI LG CT MS RG.

References

- van Soolingen D, Qian L, de Haas PE, Douglas JT, Traore H, et al. (1995) Predominance of a single genotype of *Mycobacterium tuberculosis* in countries of east Asia. *J Clin Microbiol* 33: 3234–3238.
- Bifani PJ, Mathema B, Kurepina NE, Kreiswirth BN (2002) Global dissemination of the *Mycobacterium tuberculosis* W-Beijing family strains. *Trends Microbiol* 10: 45–52.
- Glynn JR, Whiteley J, Bifani PJ, Kremer K, van Soolingen D (2002) Worldwide occurrence of Beijing/W strains of *Mycobacterium tuberculosis*: a systematic review. *Emerg Infect Dis* 8: 843–849.
- Iwamoto T, Yoshida S, Suzuki K, Tomita M, Fujiyama R, et al. (2007) Hypervariable loci that enhance the discriminatory ability of newly proposed 15-loci and 24-loci variable-number tandem repeat typing method on *Mycobacterium tuberculosis* strains predominated by the Beijing family. *FEMS Microbiol Lett* 270: 67–74.
- Kremer K, Glynn JR, Lillebaek T, Niemann S, Kurepina NE, et al. (2004) Definition of the Beijing/W lineage of *Mycobacterium tuberculosis* on the basis of genetic markers. *J Clin Microbiol* 42: 4040–4049.
- Mokrousov I (2008) Genetic geography of *Mycobacterium tuberculosis* Beijing genotype: a multifacet mirror of human history? *Infect Genet Evol* 8: 777–785.
- Wan K, Liu J, Hauck Y, Zhang Y, Zhao X, et al. (2011) Investigation on *Mycobacterium tuberculosis* diversity in China and the origin of the Beijing clade. *PLoS One* 6: e29190.
- Devaux I, Kremer K, Heersma H, Van Soolingen D (2009) Clusters of multidrug-resistant *Mycobacterium tuberculosis* cases, Europe. *Emerg Infect Dis* 15: 1052–1060.
- Afolabi D, Faihun F, Sanoussi N, Anyo G, Shamputa IC, et al. (2009) Possible outbreak of streptomycin-resistant *Mycobacterium tuberculosis* Beijing in Benin. *Emerg Infect Dis* 15: 1123–1125.
- Parwati I, van Crevel R, van Soolingen D (2010) Possible underlying mechanisms for successful emergence of the *Mycobacterium tuberculosis* Beijing genotype strains. *Lancet Infect Dis* 10: 103–111.
- Parwati I, Alisjahbana B, Apriani L, Soetkno RD, Ottenhoff TH, et al. (2010) *Mycobacterium tuberculosis* Beijing genotype is an independent risk factor for tuberculosis treatment failure in Indonesia. *J Infect Dis* 201: 553–557.
- Gagneux S, Small PM (2007) Global phylogeography of *Mycobacterium tuberculosis* and implications for tuberculosis product development. *Lancet Infect Dis* 7: 328–337.
- Reed MB, Gagneux S, Deriemer K, Small PM, Barry CE 3rd (2007) The W-Beijing lineage of *Mycobacterium tuberculosis* overproduces triglycerides and has the DosR dormancy regulon constitutively upregulated. *J Bacteriol* 189: 2583–2589.
- Mokrousov I, Ly HM, Otten T, Lan NN, Vyshnevskiy B, et al. (2005) Origin and primary dispersal of the *Mycobacterium tuberculosis* Beijing genotype: clues from human phylogeography. *Genome Res* 15: 1357–1364.
- Mokrousov I, Narvskaya O, Otten T, Vyazovaya A, Limeschenko E, et al. (2002) Phylogenetic reconstruction within *Mycobacterium tuberculosis* Beijing genotype in northwestern Russia. *Res Microbiol* 153: 629–637.
- Fakri K, Drobniewski F, Nikolayevskiy V, Brown T, Prammananan T, et al. (2011) Genetic diversity of the *Mycobacterium tuberculosis* Beijing family based on IS6110, SNP, LSP and VNTR profiles from Thailand. *Infect Genet Evol* 11: 1142–1149.
- Hanekom M, van der Spuy GD, Streicher E, Ndabambi SL, McEvoy CR, et al. (2007) A recently evolved sublineage of the *Mycobacterium tuberculosis* Beijing strain family is associated with an increased ability to spread and cause disease. *J Clin Microbiol* 45: 1483–1490.
- Iwamoto T, Fujiyama R, Yoshida S, Wada T, Shirai C, et al. (2009) Population structure dynamics of *Mycobacterium tuberculosis* Beijing strains during past decades in Japan. *J Clin Microbiol* 47: 3340–3343.
- Iwamoto T, Yoshida S, Suzuki K, Wada T (2008) Population structure analysis of the *Mycobacterium tuberculosis* Beijing family indicates an association between certain sublineages and multidrug resistance. *Antimicrob Agents Chemother* 52: 3805–3809.
- Kang HY, Wada T, Iwamoto T, Maeda S, Murase Y, et al. (2010) Phylogeographical particularity of the *Mycobacterium tuberculosis* Beijing family in South Korea based on international comparison with surrounding countries. *J Med Microbiol* 59: 1191–1197.
- Maeda S, Wada T, Iwamoto T, Murase Y, Mitarai S, et al. (2010) Beijing family *Mycobacterium tuberculosis* isolated from throughout Japan: phylogeny and genetic features. *Int J Tuberc Lung Dis* 14: 1201–1204.
- Millet J, Miyagi-Shiohira C, Yamane N, Mokrousov I, Rastogi N (2012) High-resolution MIRU-VNTRs typing reveals the unique nature of *Mycobacterium tuberculosis* Beijing genotype in Okinawa, Japan. *Infect Genet Evol* 12: 637–641.
- Wada T, Iwamoto T, Maeda S (2009) Genetic diversity of the *Mycobacterium tuberculosis* Beijing family in East Asia revealed through refined population structure analysis. *FEMS Microbiol Lett* 291: 35–43.
- Yokoyama E, Hachisu Y, Hashimoto R, Kishida K (2012) Population genetic analysis of *Mycobacterium tuberculosis* Beijing subgroup strains. *Infect Genet Evol* 12: 630–636.
- Ritacco V, Lopez B, Cafrune PI, Ferrazoli L, Suffys PN, et al. (2008) *Mycobacterium tuberculosis* strains of the Beijing genotype are rarely observed in tuberculosis patients in South America. *Mem Inst Oswaldo Cruz* 103: 489–492.
- Gomes HM, Elias AR, Oelemann MA, Pereira MA, Montes FF, et al. (2012) Spoligotypes of *Mycobacterium tuberculosis* complex isolates from patients residents of 11 states of Brazil. *Infect Genet Evol* 12: 649–656.
- Cerezo J, Jimenez Y, Hernandez J, Zozio T, Murcia MI, et al. (2012) A first insight on the population structure of *Mycobacterium tuberculosis* complex as studied by spoligotyping and MIRU-VNTRs in Bogota, Colombia. *Infect Genet Evol* 12: 657–663.
- Candia N, Lopez B, Zozio T, Carrivale M, Diaz C, et al. (2007) First insight into *Mycobacterium tuberculosis* genetic diversity in Paraguay. *BMC Microbiol* 7: 75.
- Abadia E, Sequera M, Ortega D, Mendez MV, Escalona A, et al. (2009) *Mycobacterium tuberculosis* ecology in Venezuela: epidemiologic correlates of common spoligotypes and a large clonal cluster defined by MIRU-VNTR-24. *BMC Infect Dis* 9: 122.
- Taype CA, Agapito JC, Accinelli RA, Espinoza JR, Godreuil S, et al. (2012) Genetic diversity, population structure and drug resistance of *Mycobacterium tuberculosis* in Peru. *Infect Genet Evol* 12: 577–585.
- García de Viedma D, Chaves F, Inigo J (2006) New route of importation of *Mycobacterium tuberculosis* Beijing genotype. *Emerg Infect Dis* 12: 169–170.
- Lari N, Rindi L, Bonanni D, Rastogi N, Sola C, et al. (2007) Three-year longitudinal study of genotypes of *Mycobacterium tuberculosis* isolates in Tuscany, Italy. *J Clin Microbiol* 45: 1851–1857.
- Kamerbeek J, Schouls L, Kolk A, van Agterveld M, van Soolingen D, et al. (1997) Simultaneous detection and strain differentiation of *Mycobacterium tuberculosis* for diagnosis and epidemiology. *J Clin Microbiol* 35: 907–914.
- Fenner L, Malla B, Ninet B, Dubuis O, Stucki D, et al. (2011) “Pseudo-Beijing”: evidence for convergent evolution in the direct repeat region of *Mycobacterium tuberculosis*. *PLoS One* 6: e24737.
- Tsolaki AG, Hirsh AE, DeRiemer K, Enciso JA, Wong MZ, et al. (2004) Functional and evolutionary genomics of *Mycobacterium tuberculosis*: insights from genomic deletions in 100 strains. *Proc Natl Acad Sci U S A* 101: 4865–4870.
- Caviedes L, Lee TS, Gilman RH, Sheen P, Spellman E, et al. (2000) Rapid, efficient detection and drug susceptibility testing of *Mycobacterium tuberculosis* in sputum by microscopic observation of broth cultures. The Tuberculosis Working Group in Peru. *J Clin Microbiol* 38: 1203–1208.
- Moore DA, Mendoza D, Gilman RH, Evans CA, Holm Delgado MG, et al. (2004) Microscopic observation drug susceptibility assay, a rapid, reliable diagnostic test for multidrug-resistant tuberculosis suitable for use in resource-poor settings. *J Clin Microbiol* 42: 4432–4437.
- Supply P, Allix C, Lesjean S, Cardoso-Oelemann M, Rusch-Gerdes S, et al. (2006) Proposal for standardization of optimized mycobacterial interspersed repetitive unit-variable-number tandem repeat typing of *Mycobacterium tuberculosis*. *J Clin Microbiol* 44: 4498–4510.

39. Murase Y, Mitarai S, Sugawara I, Kato S, Maeda S (2008) Promising loci of variable numbers of tandem repeats for typing Beijing family *Mycobacterium tuberculosis*. *J Med Microbiol* 57: 873–880.
40. Zhang L, Chen J, Shen X, Gui X, Mei J, et al. (2008) Highly polymorphic variable-number tandem repeats loci for differentiating Beijing genotype strains of *Mycobacterium tuberculosis* in Shanghai, China. *FEMS Microbiol Lett* 282: 22–31.
41. Velji P, Nikolayevskyy V, Brown T, Drobniewski F (2009) Discriminatory ability of hypervariable variable number tandem repeat loci in population-based analysis of *Mycobacterium tuberculosis* strains, London, UK. *Emerg Infect Dis* 15: 1609–1616.
42. Smittipat N, Billamas P, Palittapongarnpim M, Thong-On A, Temu MM, et al. (2005) Polymorphism of variable-number tandem repeats at multiple loci in *Mycobacterium tuberculosis*. *J Clin Microbiol* 43: 5034–5043.
43. Wang J, Liu Y, Zhang CL, Ji BY, Zhang LZ, et al. (2011) Genotypes and characteristics of clustering and drug susceptibility of *Mycobacterium tuberculosis* isolates collected in Heilongjiang Province, China. *J Clin Microbiol* 49: 1354–1362.
44. Keim P, Price LB, Klevytska AM, Smith KL, Schupp JM, et al. (2000) Multiple-locus variable-number tandem repeat analysis reveals genetic relationships within *Bacillus anthracis*. *J Bacteriol* 182: 2928–2936.
45. Hunter PR, Gaston MA (1988) Numerical index of the discriminatory ability of typing systems: an application of Simpson's index of diversity. *J Clin Microbiol* 26: 2465–2466.
46. Small PM, Hopewell PC, Singh SP, Paz A, Parsonnet J, et al. (1994) The epidemiology of tuberculosis in San Francisco. A population-based study using conventional and molecular methods. *N Engl J Med* 330: 1703–1709.
47. Durmaz R, Zozio T, Gunal S, Allix C, Fauville-Dufaux M, et al. (2007) Population-based molecular epidemiological study of tuberculosis in Malatya, Turkey. *J Clin Microbiol* 45: 4027–4035.
48. Filliol I, Motiwala AS, Cavatore M, Qi W, Hazbon MH, et al. (2006) Global phylogeny of *Mycobacterium tuberculosis* based on single nucleotide polymorphism (SNP) analysis: insights into tuberculosis evolution, phylogenetic accuracy of other DNA fingerprinting systems, and recommendations for a minimal standard SNP set. *J Bacteriol* 188: 759–772.
49. Cole ST, Brosch R, Parkhill J, Garnier T, Churcher C, et al. (1998) Deciphering the biology of *Mycobacterium tuberculosis* from the complete genome sequence. *Nature* 393: 537–544.
50. Luo T, Yang C, Gagneux S, Gicquel B, Mei J, et al. (2012) Combination of single nucleotide polymorphism and variable-number tandem repeats for genotyping a homogenous population of *Mycobacterium tuberculosis* Beijing strains in China. *J Clin Microbiol* 50: 633–639.
51. Wada T, Iwamoto T (2009) Allelic diversity of variable number of tandem repeats provides phylogenetic clues regarding the *Mycobacterium tuberculosis* Beijing family. *Infect Genet Evol* 9: 921–926.
52. Mokrousov I, Jiao WW, Sun GZ, Liu JW, Valcheva V, et al. (2006) Evolution of drug resistance in different sublineages of *Mycobacterium tuberculosis* Beijing genotype. *Antimicrob Agents Chemother* 50: 2820–2823.
53. Dou HY, Tseng FC, Lu JJ, Jou R, Tsai SF, et al. (2008) Associations of *Mycobacterium tuberculosis* genotypes with different ethnic and migratory populations in Taiwan. *Infect Genet Evol* 8: 323–330.
54. Mestre O, Luo T, Dos Vultos T, Kremer K, Murray A, et al. (2011) Phylogeny of *Mycobacterium tuberculosis* Beijing strains constructed from polymorphisms in genes involved in DNA replication, recombination and repair. *PLoS One* 6: e16020.
55. Wada T, Iwamoto T, Hase A, Maeda S (2012) Scanning of genetic diversity of evolutionarily sequential *Mycobacterium tuberculosis* Beijing family strains based on genome wide analysis. *Infect Genet Evol* 12: 1392–1396.
56. Weniger T, Krawczyk J, Supply P, Niemann S, Harmsen D (2010) MIRU-VNTRplus: a web tool for polyphasic genotyping of *Mycobacterium tuberculosis* complex bacteria. *Nucleic Acids Res* 38: W326–331.
57. Mokrousov I, Narvskaya O, Vyazovaya A, Millet J, Otten T, et al. (2008) *Mycobacterium tuberculosis* Beijing genotype in Russia: in search of informative variable-number tandem-repeat loci. *J Clin Microbiol* 46: 3576–3584.
58. Jiao WW, Mokrousov I, Sun GZ, Guo YJ, Vyazovaya A, et al. (2008) Evaluation of new variable-number tandem-repeat systems for typing *Mycobacterium tuberculosis* with Beijing genotype isolates from Beijing, China. *J Clin Microbiol* 46: 1045–1049.
59. Lasunskaja E, Ribeiro SC, Manicheva O, Gomes LL, Suffys PN, et al. (2010) Emerging multidrug resistant *Mycobacterium tuberculosis* strains of the Beijing genotype circulating in Russia express a pattern of biological properties associated with enhanced virulence. *Microbes Infect* 12: 467–475.
60. Hernandez-Pando R, Marquina-Castillo B, Barrios-Payan J, Mata-Espinosa D (2012) Use of mouse models to study the variability in virulence associated with specific genotypic lineages of *Mycobacterium tuberculosis*. *Infect Genet Evol* 12: 725–731.
61. Chen YY, Chang JR, Huang WF, Kuo SC, Su IJ, et al. (2012) Genetic diversity of the *Mycobacterium tuberculosis* Beijing family based on SNP and VNTR typing profiles in Asian countries. *PLoS One* 7: e39792.
62. Homolka S, Projahn M, Feuerriegel S, Ubben T, Diel R, et al. (2012) High resolution discrimination of clinical *Mycobacterium tuberculosis* complex strains based on single nucleotide polymorphisms. *PLoS One* 7: e39855.
63. Stucki D, Malla B, Hostettler S, Huna T, Feldmann J, et al. (2012) Two New Rapid SNP-Typing Methods for Classifying *Mycobacterium tuberculosis* Complex into the Main Phylogenetic Lineages. *PLoS One* 7: e41253.