BK modulation as a therapeutic target for Cystic Fibrosis

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Introduction

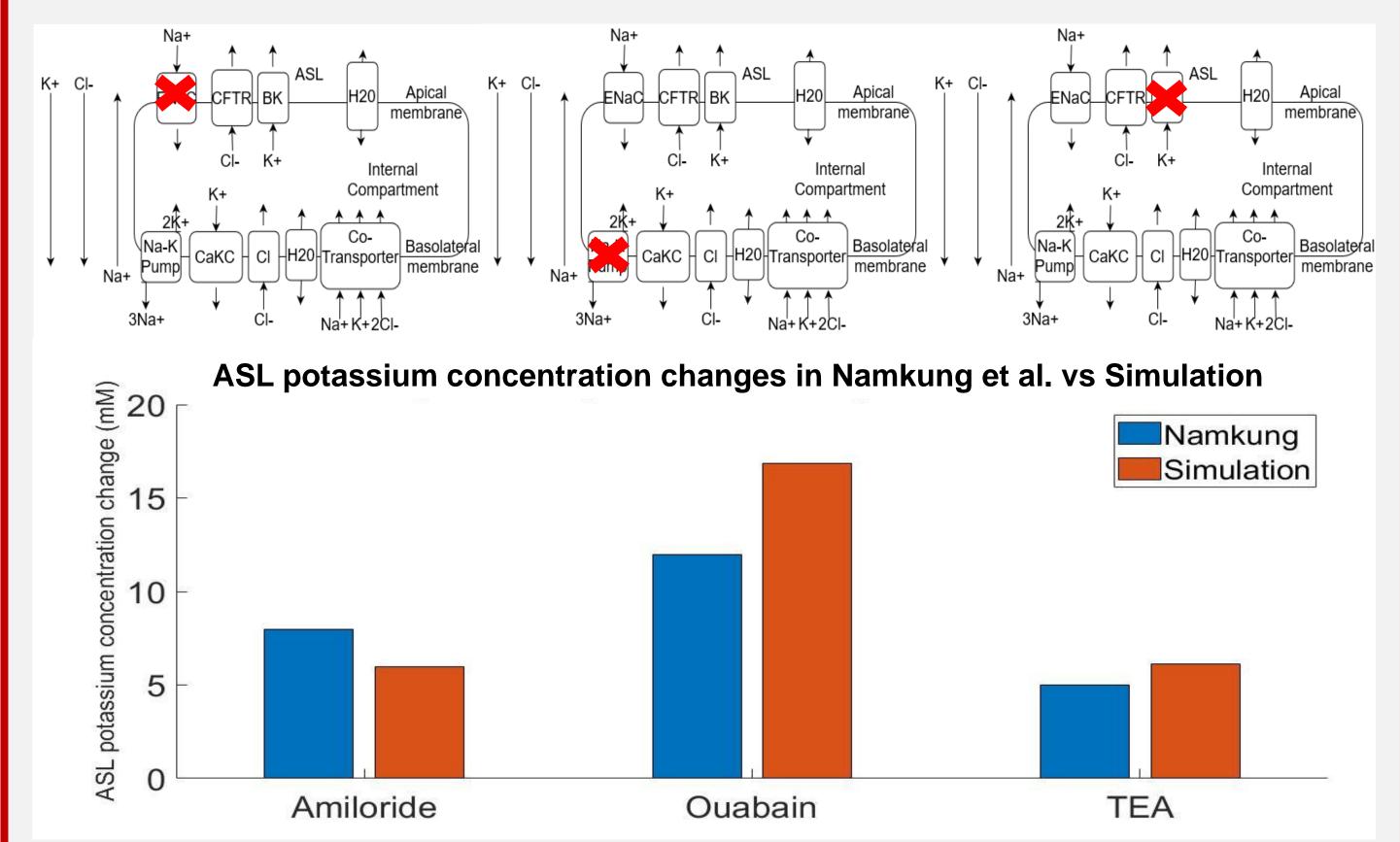
Large conductance calcium-activated potassium channels (BK channels) are vital for maintaining proper hydration of the airway surface liquid (ASL) [1]. We have employed in silico (Figures 1, 3 & 4) and in vitro methods to examine the potential of BK channel modulators to improve ASL hydration in cystic fibrosis (CF).

Non-CF Airway **CF** Airway

Simulations

Model Validation

We developed our model as an extension of O'Donoghue et al. (2013) [4] with a variable ASL volume and ion concentrations. We validated our model against the results of Namkung et al. (2009) [5], who measured changes in ASL potassium ion concentrations following application of various blockers (Figure 3)



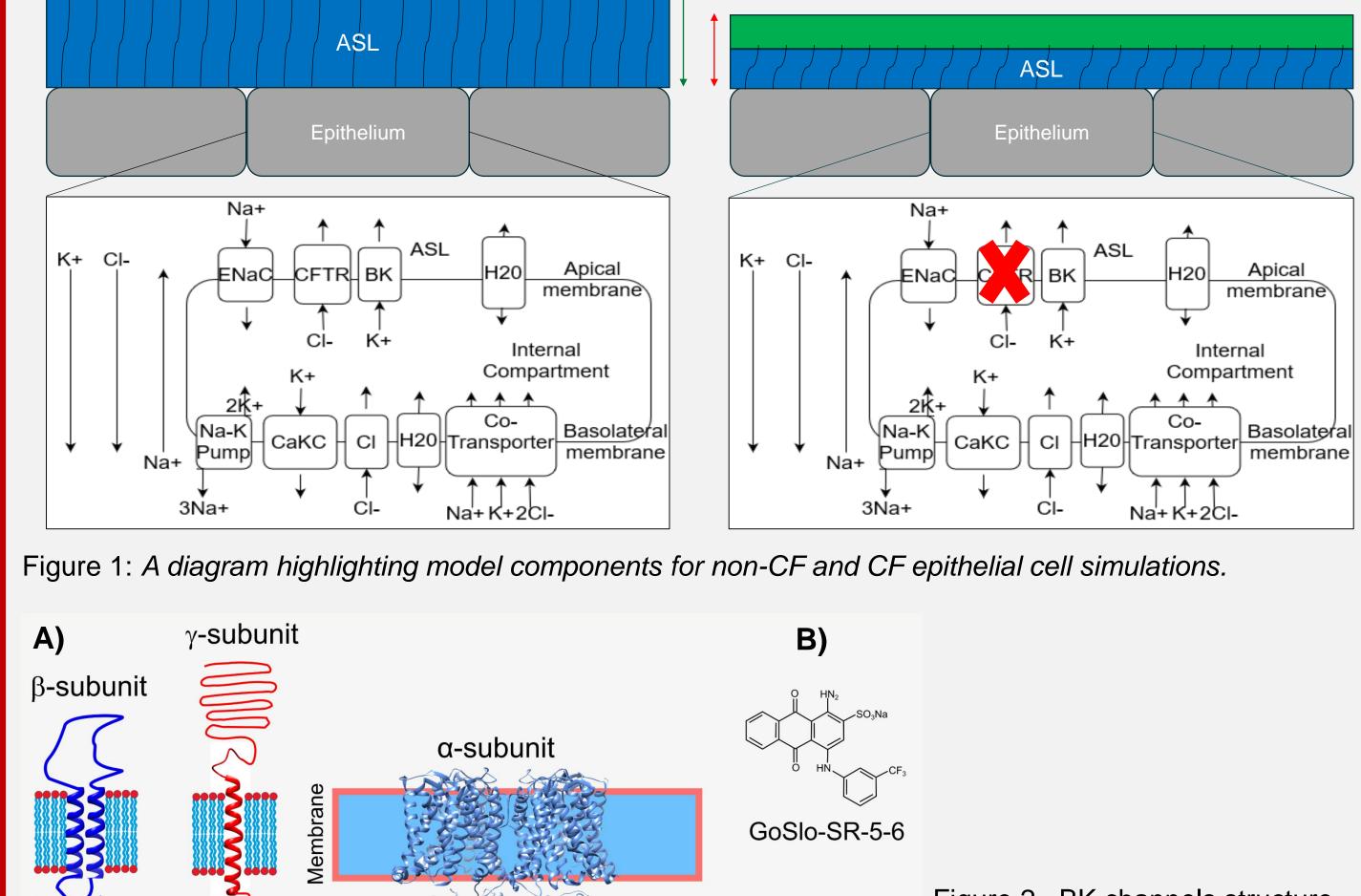


Figure 2. BK channels structure. A) BK channels must contain an α subunit but may also contain auxiliary β or γ subunits. B) Structure of the BK channel activator GoSlo.

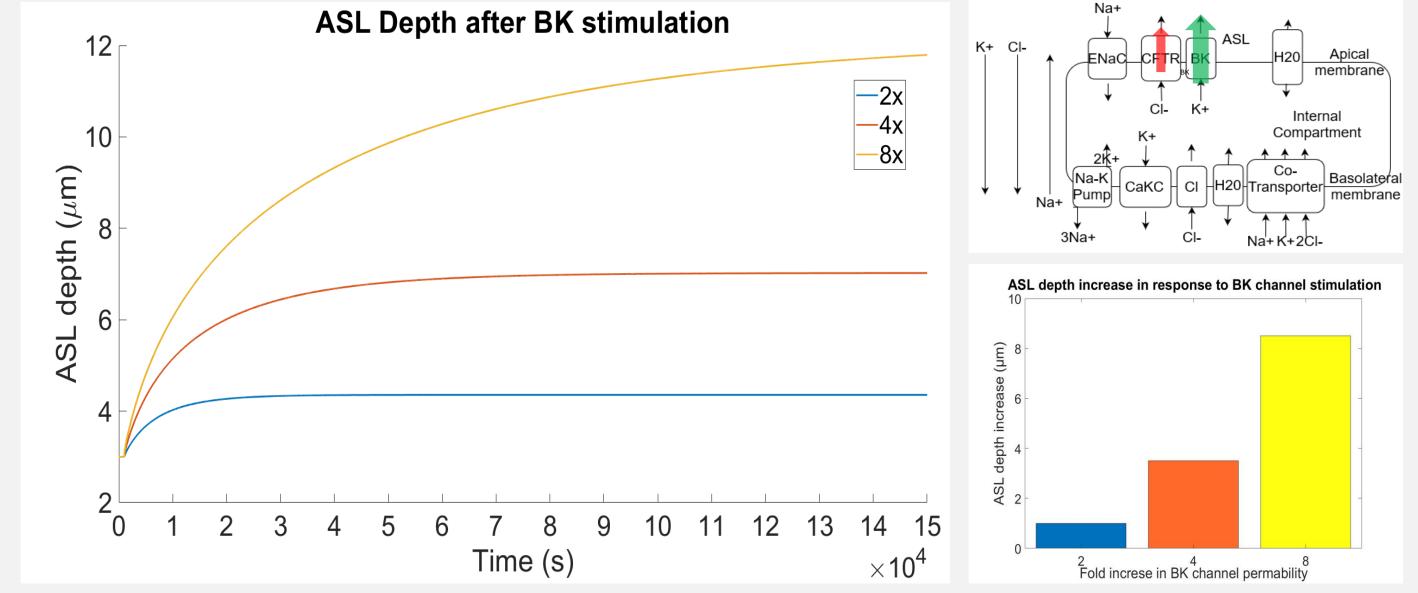
BK channels

BK channels are comprised of four α subunits and may co-assemble with β subunits (β 1-4) and γ subunits (γ 1-4) (Figure 2). The subunit combination present in the airway epithelium is unclear.

Figure 3: A bar chart comparing the simulation results to that of Namkung et al. (2009) with corresponding schematic diagrams of the simulation carried out in the model

Model Prediction

We used our model to simulate the effect of BK channel stimulation on ASL depth. The model predicts that the BK channel activation can significantly increase ASL depth when there is residual anion permeability was reduced by 75% to replicate the effect of the R334W mutation [7] (Figure 4).

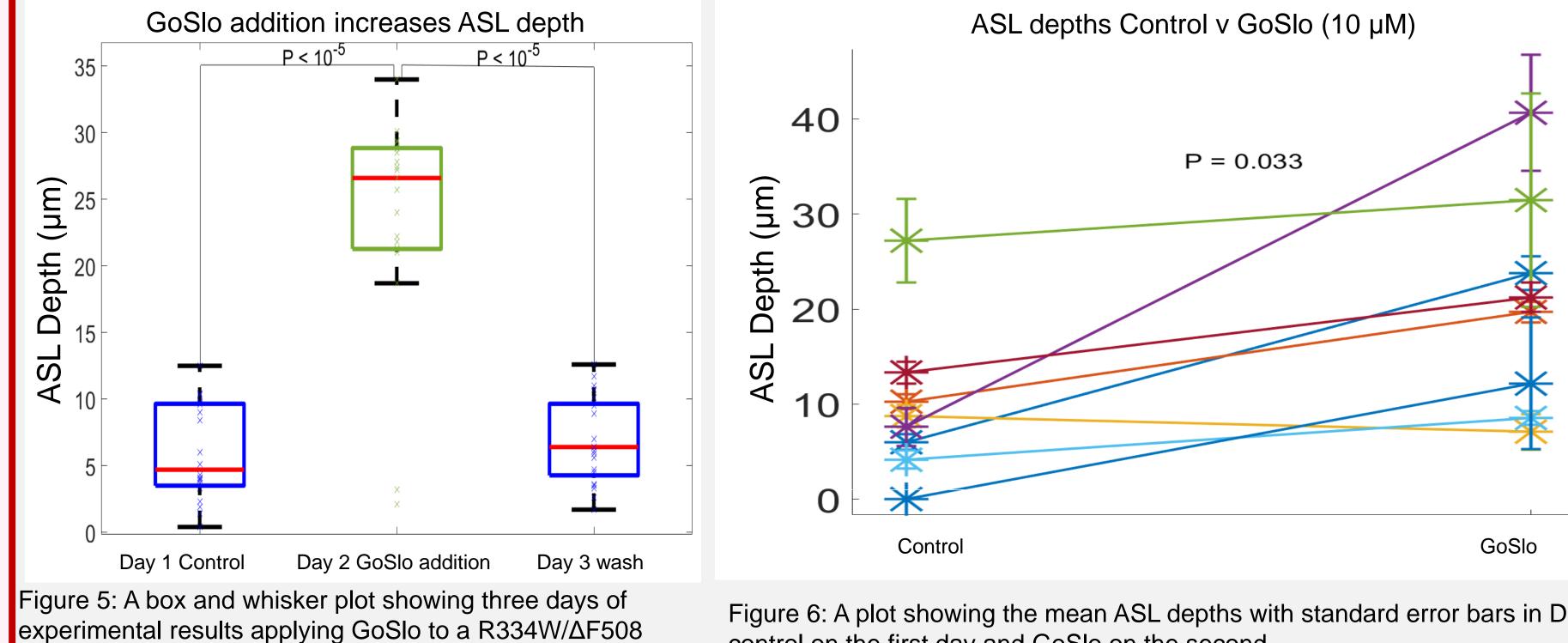


Methods

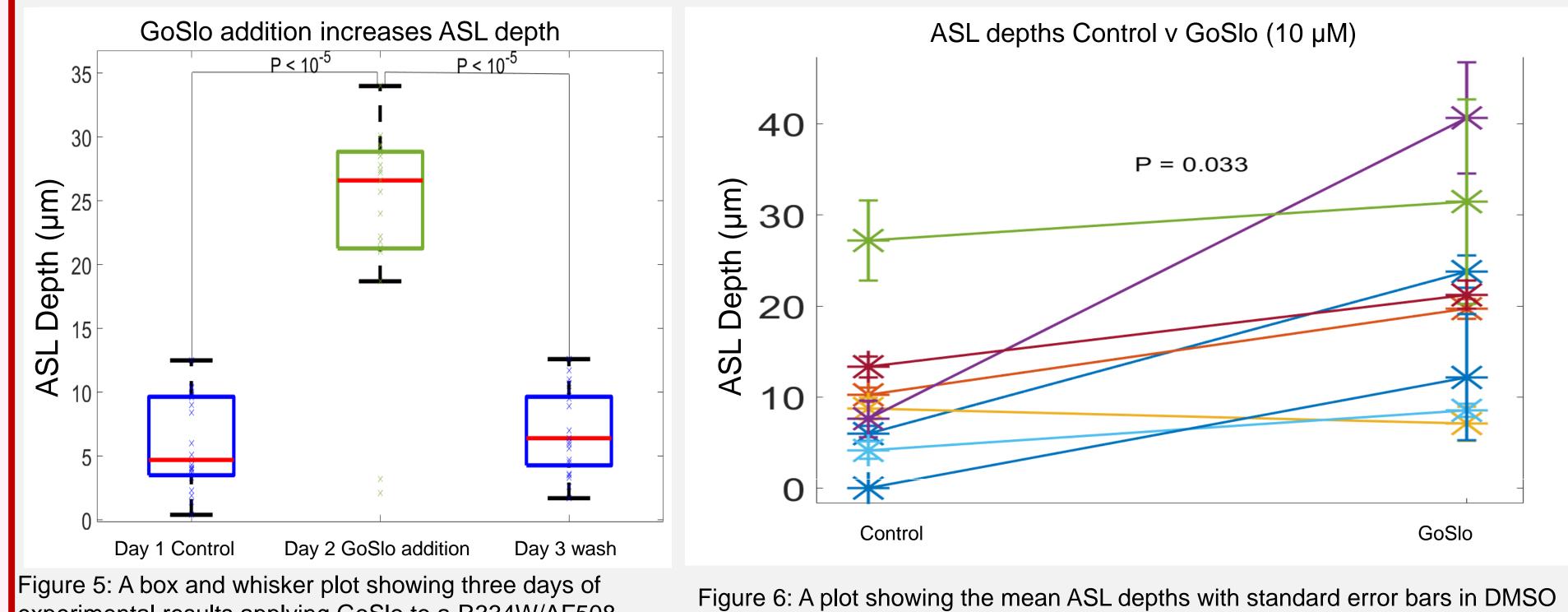
Human bronchial epithelial (HBE) cells, expressing R334W/ΔF508 mutations were grown in air liquid interface (ALI) culture using PneumaCult[™]-Ex Plus Medium and PneumaCult[™]-ALI Medium. Cultures were grown in the absence of antifungals, and antibiotics. ASL depth measurements were made using scanning ion conductance microscopy [2]. During measurements an environmental chamber was used to maintain the cultures at 37 °C, 5 % CO_2 and 99% humidity. The BK channel modulator GoSlo-SR-5-6 (GoSlo) was chosen as it is effective for all channel subunit combinations [3].

Experimental Validation

BK activation can increase ASL depth (Figure 5), when R334W CFTR provides residual anion activity. R334W has been reported to be resistant to modulators [6]



Looking across all measurements made going from vehicle (DMSO) to GoSlo shows an increase in the mean ASL depth for seven of eight cultures (Figure 6).



control on the first day and GoSlo on the second

Figure 4: A diagram showing simulation results of BK channel stimulation with corresponding schematic diagrams of the simulation carried out in the model

Conclusion

We have shown using in silico and in vitro methods that BK channels may be a good target for restoring airway surface liquid hydration in CF, provided there is some residual anion channel activity. We have been able to demonstrate this using R334W, which has been reported to be poorly responsive to current CFTR modulators. These findings suggest that BK channel modulation may provide a useful adjunct therapy for CF patients.



References:

culture

1] Manzanares, D., Gonzalez, C., Ivonnet, P., Chen, R.S., Valencia-Gattas, M., Conner, G.E., Larsson, H.P. and Salathe, M., 2011. Functional apical large conductance, Ca2+-activated, and voltage-dependent K+ channels are required for maintenance of airway surface liquid volume. Journal of Biological Chemistry, 286(22), pp.19830-19839. [2] Ivanova, R., Benton, D.C., Munye, M.M., Rangseesorranan, S., Hart, S.L. and Moss, G.W., 2019. A nanosensor toolbox for rapid, label-free measurement of airway surface liquid and epithelial cell function. ACS applied materials & interfaces, 11(9), pp.8731-8739. [3] Roy, S., Morayo Akande, A., Large, R.J., Webb, T.I., Camarasu, C., Sergeant, G.P., McHale, N.G., Thornbury, K.D. and Hollywood, M.A., 2012. Structure-Activity Relationships of a Novel Group of Large-Conductance Ca2+-Activated K+ (BK) Channel Modulators: The GoSlo-SR Family. ChemMedChem, 7(10), pp.1763-1769. [4] O'Donoghue, D.L., Dua, V., Moss, G.W. and Vergani, P., 2013. Increased apical Na+ permeability in cystic fibrosis is supported by a quantitative model of epithelial ion transport. *The Journal of physiology*, 591(15), pp.3681-3692. [5] Namkung, W., Song, Y., Mills, A.D., Padmawar, P., Finkbeiner, W.E. and Verkman, A.S., 2009. In situ measurement of airway surface liquid [K+] using a ratioable K+-sensitive fluorescent dye. *Journal of Biological Chemistry*, 284(23), pp.15916-15926. [6] Ciciriello, F., Bijvelds, M.J., Alghisi, F., Meijsen, K.F., Cristiani, L., Sorio, C., Melotti, P., Fiocchi, A.G., Lucidi, V. and De Jongé, H.R., 2022. Theratyping of the rare CFTR variants E193K and R334W in rectal organoid-derived epithelial monolayers. Journal of Personalized Medicine, 12(4), p.632. [7] Sheppard, D.N. and Welsh, M.J., 1999. Structure and function of the CFTR chloride channel. *Physiological reviews*, 79(1), pp.S23-S45.