

# Pathogenesis of Tuberculous Meningitis

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## **Declaration**

I Angharad Grace Davis confirm that the work presented in this thesis is my own.

Where information has been derived from other sources, I confirm that this has been indicated in the thesis.

## Abstract

Tuberculous Meningitis (TBM) is the most severe form of tuberculosis, affecting >100,000 people/year (Wilkinson et al., 2017, Marais et al., 2011). TBM arises when *Mycobacterium tuberculosis* crosses the blood-brain barrier, causing severe inflammation and tissue damage (Davis et al., 2019b). Inflammation also initiates metabolic derangement, mediating neuronal injury despite treatment (Davis et al., 2019b, Rohlwink et al., 2019, Rohlwink et al., 2017). In HIV-1 coinfection, TBM mortality is reported to be as high as 50% (Marais et al., 2011). Half of TBM survivors are affected by neurological disability (Thwaites et al., 2003). Thus, improved therapeutic strategies are needed, including targeted and nuanced modification of the injurious host inflammatory response. To develop these, we must understand the immune induced tissue damaging responses and metabolic disturbances contributing to brain damage which drive acute and long-term neurological complications.

The goals of this PhD thesis were to investigate the safety of a novel drug regimen in HIV-associated TBM and through a series of nested sub-studies, understand pathogenic mechanisms of acute and long term neurological sequelae in TBM.

Specifically, this thesis presents:

- i) A study protocol and results from a phase 2A randomised controlled trial of high dose rifampicin and adjunctive linezolid with and without aspirin in HIV-associated TBM
- ii) Results from a case-control study of cognitive and functional outcomes in HIV-associated TBM
- iii) Results from a study investigating *in vivo* markers of brain injury in HIV-associated TBM via magnetic resonance spectroscopy

iv) Results from a study investigating *ex vivo* markers of poor outcome via Luminex multiplex analysis of blood and cerebrospinal fluid.

Given the timing of this PhD, this thesis also contains results of an observational case-control study to understand neurological complications of COVID-19 via systematic analysis of cerebrospinal fluid of patients presenting with neurological symptoms during the first wave of the COVID-19 pandemic in South Africa. These results are included within the appendix of the thesis.

## Impact Statement

Tuberculous meningitis (TBM) arises when *Mycobacterium tuberculosis* crosses the blood-brain barrier and is the most lethal and disabling form of tuberculosis. In some contexts, including HIV co-infection, TBM mortality can be as high as 50% despite therapy, and long-term disability is common amongst survivors. These injuries are induced by tissue damaging immune responses and metabolic disturbances contributing to neurotoxic and degenerative neurological damage. Poor outcomes are thought in part due to i) inadequate antibiotic penetration into the central nervous system, ii) poor understanding of the host inflammatory response in the acute and longer term phases of illness and iii) a lack of adequate therapies which dampen a counterproductive host inflammatory response. This PhD contributes knowledge to these areas.

A phase 2 randomised controlled trial (RCT) demonstrated that i) high-dose rifampicin and adjunctive linezolid can safely be added to standard of care and ii) high-dose aspirin is safe when used in combination with intensified antibiotics in HIV-associated TBM. Results of this trial, now published in *Clinical Infectious Diseases* (Davis et al., 2022b), will inform the design of phase 3 RCT in TBM.

A case-control study in TBM shows that cognitive impairment occurs in half of patients at 6 months post diagnosis and that impairment is above and beyond that attributable to HIV or the systemic effects of TB alone. These results, now published in *Clinical Infectious Diseases* (Davis et al., 2022a), provide new knowledge on neurological complications in TBM. They also provide rationale for new studies to evaluate outcomes at longer-term timepoints and explore the relationship between cognitive performance, functional status and treatment adherence. The results also provide new

methods for clinical correlation of emerging knowledge in TBM pathogenesis which suggests that key pathways share similar mechanistic characteristics to neurodegenerative conditions.

*Ex vivo* analysis of blood and CSF in TBM provides insight into potential inflammatory mediators driving the immune response. The work highlights mediators which may serve as biomarkers of poor outcome or druggable targets in the development of host directed therapies; findings around IL-1 $\beta$  and inflammasome driven innate immune responses provide further rationale for the consideration of drugs which target these pathways. Studies using larger cohorts and including longer term timepoints are now needed to further investigate and validate these findings.

A pilot study evaluated a novel method for *in vivo* measurement of brain injury markers in TBM. It showed that single voxel magnetic resonance spectroscopy can feasibly measure glutamate and GABA within the brains of patients with TBM, providing methodology for future studies. Although results need validation in a larger cohort, they demonstrate that glutamate is significantly raised in the acute phase of TBM compared to healthy controls. This work provides a platform for further development of this technique which is important given the emerging importance of glutamate driven excitotoxicity in TBM.

Alongside this work, a case-control study evaluating the spectrum of neurological manifestations of COVID-19 provides knowledge on the neurotropism of SARS-CoV2, demonstrating that penetration of virus to the CNS is uncommon even in those with COVID-19 and neurological symptoms.

## Acknowledgement

I would firstly like to thank my primary supervisor Professor Robert J Wilkinson from whom I have received unwavering support and guidance over the last 5 years. Thank you trusting me to lead a clinical trial, for your diligence in your role as my supervisor, and for your honest and invaluable mentorship. These have been the most rewarding years of my professional life and I will always be deeply grateful for the opportunities I have been given. Thanks also to Professor Anne O'Garra for your teaching and guidance; it has been a privilege to learn from you. Likewise to Professor Kaila Srail for your guidance in navigating the administration of a PhD at University College London and for your always enthusiastic support.

I would like to express my upmost respect and gratitude for the clinical trial teams at CIDRI Africa who work tirelessly to recruit and care for study participants despite challenging settings. In particular thank you to my close colleagues Professor Sean Wasserman, Mpumi Maxebengula and Dr Cari Stek; your support, friendship and guidance has been invaluable. I am also grateful for the opportunity to have worked with Anna Dreyer, A/Professor Sam Nightingale, Dr Frances Robertson and Professor John Joska at the University of Cape Town. I have learnt a huge amount through this work with you and developed a new research interest which I endeavour to pursue far beyond this PhD. Thank you for sharing your expertise and passion with me. And of course thank you to the participants who agreed to be part of numerous clinical studies within this PhD; without your time and trust none of this would be possible.

Finally I would like to thank my family. My parents for teaching me to believe in myself when things don't go to plan. My husband Dave whose commitment to helping me

realise my career goals is limitless; thank you for always sharing the load and for your constant love. And lastly to my two children, Rowan and Cerys, both born during the course of this PhD. Thank you for your sacrifice, patience and for the joy you bring me every single day.



# Table of Contents

Abstract .....	3
Impact Statement .....	5
Acknowledgement .....	7
Table of Contents .....	9
List of Figures .....	11
List of Tables .....	13
Thesis outline	15
Author Contributions .....	17
Funding	20
<b>Chapter 1. Introduction</b> .....	<b>21</b>
<b>1.1 Background and PhD goals</b> .....	<b>21</b>
<b>1.2 Current treatment options in Tuberculous Meningitis</b> .....	<b>22</b>
1.2.1 Antibiotics .....	23
1.2.2 Host directed therapies .....	29
1.2.3 Potential Pathways for Future Host Directed Therapies for Tuberculous Meningitis .....	34
1.2.4 Supportive therapies .....	39
1.2.5 Conclusions and research priorities .....	44
<b>1.3 Pathogenesis of Tuberculous Meningitis</b> .....	<b>45</b>
1.3.1 From primary infection to the central nervous system .....	45
1.3.2 Pathogenic and pathophysiological mechanisms within the brain .....	48
1.3.3 Metabolic factors in the host .....	53
1.3.4 Host genetic factors .....	55
1.3.5 Pathogen virulence factors and their effect on pathogenesis .....	58
1.3.6 Differences in the intracerebral immune response in HIV .....	60
1.3.7 Macroscopic manifestations of the disease in relation to the immune response .....	63
1.3.8 Research gaps and the path forward .....	66
<b>Chapter 2. Study protocol for A phase 2A trial of the safety and tolerability of                 increased dose rifampicin and adjunctive linezolid, with or without                 aspirin, for HIV-associated tuberculous meningitis (The LASER-                 TBM trial)</b> .....	<b>77</b>
<b>2.1 Background</b> .....	<b>77</b>
<b>2.2 Methods and Study Design</b> .....	<b>81</b>
<b>2.3 Recruitment, randomisation, retention and withdrawal</b> .....	<b>85</b>
<b>2.4 Interventions</b> .....	<b>86</b>
<b>2.5 Study procedures, schedule and clinical assessments</b> .....	<b>89</b>
<b>2.6 Statistical considerations</b> .....	<b>94</b>
<b>2.7 Adverse events</b> .....	<b>96</b>
<b>2.8 Safety monitoring</b> .....	<b>97</b>
<b>2.9 Data access and handling</b> .....	<b>98</b>
<b>2.10 Data collection, management and storage</b> .....	<b>99</b>
<b>2.11 Trial committees, ethical procedures and sponsorship</b> .....	<b>100</b>
<b>2.12 Version control and protocol amendment policy</b> .....	<b>102</b>

<b>Chapter 3. Results from a phase 2A trial of the safety and tolerability of increased dose rifampicin and adjunctive linezolid, with or without aspirin, for HIV-associated tuberculous meningitis (The LASER-TBM Trial)</b>	<b>113</b>
3.1 Introduction .....	113
3.2 Methods .....	113
3.3 Results.....	115
3.4 Discussion .....	117
<b>Chapter 4. Cognitive Impairment in Tuberculous Meningitis .....</b>	<b>136</b>
4.1 Introduction .....	136
4.2 Methods .....	138
4.3 Results.....	143
4.4 Discussion .....	147
<b>Chapter 5. Luminex Multiplex analysis in blood and CSF of patients with HIV-associated TBM .....</b>	<b>159</b>
5.1 Introduction .....	159
5.2 Methods .....	161
5.3 Results.....	165
5.4 Discussion .....	168
5.5 Conclusions and future research.....	176
<b>Chapter 6. Magnetic resonance spectroscopy to detect GABA and Glutamate in HIV-associated Tuberculous Meningitis.....</b>	<b>204</b>
6.1 Introduction .....	204
6.2 Methods .....	207
6.3 Results.....	210
6.4 Discussion .....	212
<b>Chapter 7. Discussion .....</b>	<b>226</b>
<b>Appendix: Spectrum of neurological manifestations and systematic evaluation of cerebrospinal fluid for SARS-CoV2 in patients admitted to hospital during the COVID-19 epidemic in South Africa (The HIATUS-3 Study) .....</b>	<b>231</b>
<b>Reference List .....</b>	<b>253</b>

## List of Figures

Figure 1.1 Computerised tomography of the head in a patient with TBM .....	69
Figure 1.2 Treatment algorithm for the management of TBM.....	70
Figure 1.3 Summary of the pathogenesis of TBM .....	71
Figure 1.4 <i>M.tb</i> and vitamin D impact on tryptophan metabolism .....	73
Figure 1.5 Radiological features of TBM.....	74
Figure 1.6 Basal cisterns and pituitary anatomy.....	75
Figure 1.7 Lenticulostriate arteries.....	76
Figure 2.1 LASER-TBM Study Design .....	104
Figure 3.1 CONSORT diagram for LASER-TBM.....	122
Figure 3.2 Kaplan-Meier analysis of time to worst grade adverse events of special interest or death.....	123
Figure 3.3 Kaplan-Meier analysis time to death .....	124
Figure 3.4 Kaplan-Meier analysis time to worst grade adverse event of special interest .....	125
Figure 3.5 Functional neurological outcome at day 56 as defined by modified Rankin scale.....	126
Figure 3.6 Change in CSF parameters over time.....	127
Figure 4.1 CONSORT diagram for study of cognitive Impairment in TBM.....	156
Figure 4.2 Global T Scores across groups.....	157
Figure 5.1 Study sampling schedule for LASER-TBM.....	177
Figure 5.2 CONSORT diagram for Luminex analysis study .....	178
Figure 5.3 Summary Venn diagram to describe mediator findings across all analysis groups .....	179

Figure 5.4 Scatter graphs plotting individual mediator values in cases with HIV associated TBM compared to non-infectious comparator group.....	180
Figure 5.5 Day 3 CSF immune mediators in patients with HIV associated TBM compared to non-infectious comparator group.....	181
Figure 5.6 Day 3 CSF mediators in those with microbiologically confirmed HIV associated TBM versus those without.....	182
Figure 5.7 Comparison of baseline mediators in matched CSF and blood timepoint (day 3), CSF to blood ratio ranked by log fold increase.....	183
Figure 5.8 Day 3 CSF mediator concentrations with MRS outcome at D56.....	184
Figure 5.9 Day 3 CSF markers demonstrating difference ( $p < 0.05$ ) with good (MRS 0-3) versus poor (MRS 4-6) outcome .....	185
Figure 5.10 Day 3 Blood markers demonstrating difference ( $p < 0.05$ ) with good (MRS 0-3) versus poor (MRS 4-6) outcome.....	186
Figure 5.11 Day 3 CSF:Blood ratios in mediators demonstrating difference ( $p < 0.05$ ) with good (MRS 0-3) versus poor (MRS 4-6) outcome.....	187
Figure 5.12 Longitudinal change in selected CSF parameters .....	188
Figure 5.13 Longitudinal change in selected blood parameters.....	189
Figure 5.14 Longitudinal change in selected blood markers grouped by survival .....	190
Figure 6.1 Magnetic resonance spectrum of normal brain.....	218
Figure 6.2 Voxel placement .....	219
Figure 6.3 Study CONSORT for magnetic resonance spectroscopy study.....	220
Figure 6.4 Glutamate/glutamine (Glx) concentrations in participants with TBM versus healthy controls .....	221
Figure 6.5 GABA concentrations in participants with TBM versus healthy controls ...	222
Figure 6.6 Glutamate:GABA ratios in participants with TBM versus healthy controls	223
Figure 6.7 Baseline glutamate concentrations and glutamate:GABA concentrations in relation to cognitive outcomes.....	224

## List of Tables

Table 1.1 Currently available antibiotics for treatment of TBM .....	67
Table 1.2 Clinical rating scores used within surgical management of TBM .....	68
Table 2.1 Solicited treatment related adverse events, objective measures for assessment and management plan in each setting.....	109
Table 2.2 Details and dosing of study drug regimen provided for 56 days post randomisation .....	110
Table 2.3 Planned study assessments and procedure per study date.....	111
Table 3.1 Adverse events of special interest (AESI) assessed in LASER-TBM.....	128
Table 3.2 Baseline demographics and clinical characteristics in LASER-TBM .....	129
Table 3.3 AESI stratified by treatment arm.....	131
Table 3.4 Details of AESI by event .....	132
Table 3.5 Timing and cause of death prior to day 56 .....	134
Table 3.6 Reasons for screening exclusion.....	135
Table 3.7 Reasons for study withdrawal prior to day 56.....	135
Table 4.1 Parent Studies included within the study of cognitive impairment in TBM..	153
Table 4.2 Baseline demographics and clinical characteristics of participants included within the study of cognitive impairment in TBM .....	154
Table 4.3 (A) Comparison of Domain specific T scores in TBM cases vs comparator group 1 (PLWH, no history of TB) and (B) Comparison of Domain specific T scores in TBM cases vs comparator group 2 (PLWH non-CNS TB).....	155
Table 5.1 Baseline demographics and clinical characteristics of participants included within the Luminex study analysis.....	191
Table 5.2 Baseline (Day 3) CSF mediator concentrations in patients with TBM (n=14) and non-infectious comparators (n=22).....	192

Table 5.3 CSF mediator concentrations in those with microbiologically confirmed HIV associated TBM vs those without.....	193
Table 5.4 Baseline (day 3) mediators in HIV associated TBM, a comparison of blood and CSF concentrations .....	196
Table 5.5 Baseline cerebrospinal fluid (CSF) and blood (BI) mediator concentrations, and CSF:Blood ratios in patients with HIV associated TBM in those with good (MRS 0-3) vs poor (MRS 4-6) outcome.....	199
Table 6.1 Baseline demographics and clinical characteristics of participants included with the study of magnetic resonance spectroscopy in TBM .....	225

## Thesis outline

**Chapter 1** provides an introduction to the burden of Tuberculous Meningitis globally and sets out the goals of this PhD. Thereafter it provides a more detailed overview on i) current treatment options for TBM and its neurological sequelae, ii) potential novel targets for host directed therapies and iii) pathogenic mechanisms in TBM. These insights are largely taken from three review papers published throughout the course of my PhD (Davis et al., 2018, Davis et al., 2019b, Davis et al., 2020). The scientific rationale for the research conducted during the PhD is explained.

**Chapter 2** is a published research paper outlining the study protocol for the phase 2 randomised controlled trial evaluating the safety of high dose rifampicin, adjunctive linezolid with and without high dose aspirin in HIV-associated TBM ('The LASER-TBM Trial') (Davis et al., 2021).

**Chapter 3** is a research paper describing the results from the LASER-TBM Trial (published in *Clinical Infectious Diseases* at time of thesis submission (Davis et al., 2022b)).

**Chapter 4** is a case-control study describing cognitive and functional outcomes in HIV associated TBM (published in *Clinical Infectious Disease* at time of thesis submission (Davis et al., 2022a)).

**Chapter 5** describes results from *ex vivo* immunological analysis of blood and CSF arising from the LASER-TBM trial using Luminex platform technology.

**Chapter 6** describes results from a pilot study using magnetic resonance spectroscopy to measure *in vivo* concentrations of GABA and glutamate in patients with HIV-associated TBM recruited to the LASER-TBM trial.

**Chapter 7** provides a brief discussion of key findings from the thesis and describes next steps in relation to this work.

The appendix includes a case-control study describing the spectrum of neurological manifestations in SARS-CoV2 infection (available in medarchive at time of thesis submission).



## Author Contributions

The work undertaken within this PhD included several collaborative projects. Outlined below are details on the contributions to each chapter made by myself (AGD) and research collaborators.

**Chapter 1:** This chapter brings together three review papers published at the start of my PhD, which have been updated in line with the current literature. For the original publications, AGD was involved in paper conceptualisation, and produced the first written draft. She was the first and corresponding author on each submission.

Radiological images included within this chapter were kindly provided by colleagues at University of Cape Town, Professors Graeme Meintjes and Tony Figaji.

**Chapter 2:** Within this study protocol AGD was involved in study conceptualisation and design. She produced the first written draft of the study protocol and undertook subsequent edits following comments from listed co-authors. She also developed or oversaw the development of all study documentation (informed consent forms, standard operating procedures, case report forms, ethical and regulatory applications) relating to the trial. Professors Sean Wasserman and Graeme Meintjes at The University of Cape Town and my primary supervisor Robert J Wilkinson oversaw study conceptualisation and design and provided regular review of drafts of the protocol.

**Chapter 3:** AGD wrote the statistical analysis plan, with supervision from the trial statistician related to this analysis (published as supplementary material in (Davis et al., 2021)). AGD cleaned the data, undertook the analysis, interpreted the results and produced the first draft of the manuscript. Professors Sean Wasserman, Graeme

Meintjes and Robert J Wilkinson, as well as the trial statistician (Jason C Liang) provided close supervision via data presentations at interim points throughout the analysis.

**Chapter 4:** AGD conceived the idea for this study and designed the sub-study written into the LASER-TBM protocol. Colleagues at The University of Cape Town Anna Dreyer, A/Professor Sam Nightingale and Professor John Joska contributed to the study design. Data within this chapter is presented from cognitive test battery data arising from the LASER-TBM sub-study, but also includes;

i) cognitive test battery data arising from an unpublished data set submitted as part of an MSc thesis project by Dr Christine Albertyn at the University of Cape Town (additional TBM cases, and a population of patients with non-CNS TB) – included to increase statistical power and as a comparator group;

ii) a subset of cognitive test battery data arising from an unpublished prospective study of cognitive impairment in HIV (the CONNECT study) – included as a comparator group of people living with HIV.

With permission, AGD combined the raw data from these additional studies to that arising from LASER-TBM. AGD cleaned the data, undertook the analysis, interpreted the results and produced the first draft of the manuscript. This work was supervised by Anna Dreyer and A/Professor Sam Nightingale.

**Chapter 5:** AGD conceived the idea for the study, cleaned the data, analysed the results and wrote the chapter. The laboratory work (processing of CSF by Luminex Multiplex commercial kit) was performed by a University of Cape Town collaborator (Dr

Muki Shey) as travel to South Africa at the time of this work was prohibited due to the COVID-19 pandemic. Supervision and guidance in interpretation of the results and chapter draft review was provided by Professors Anne O'Garra and Robert J Wilkinson.

**Chapter 6:** AGD designed the study and wrote documentation related to the study (consent forms, standard operating procedures, ethical and regulatory approvals). The imaging protocol was developed in collaboration with colleagues at University of Cape Town (Dr Frances Robertson and Professor Ernesta Meinjtes). AGD cleaned the data, analysed the results, interpreted the findings, and wrote the chapter. Supervision and guidance in interpretation of the results and chapter draft review was provided by Dr Frances Robertson and Professor Robert J Wilkinson.

In the study of neurological manifestations of COVID-19 included within the appendix of this thesis AGD conceived the idea for the study and wrote the study protocol as well as documentation related to the study (standard operating procedures, case report forms, ethical approvals). AGD cleaned the data, analysed the results and produced the first draft of the manuscript. The laboratory work related to i) Luminex analysis of CSF and ii) detection of SARS-CoV2 in CSF were performed by University of Cape Town collaborators (Dr Muki Shey and Dr Georgia Schafer) as travel to South African at the time of this work was prohibited due to the COVID-19 pandemic. Oversight and supervision was provided by Professor Robert J Wilkinson.

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# Chapter 1. Introduction

## 1.1 Background and PhD goals

Tuberculosis remains a major global health problem. In 2015, an estimated 10.4 million new cases of TB occurred worldwide. The World Health Organization's 'End TB Strategy' calls for a 90% reduction in TB related deaths and 80% reduction in TB incidence rate by 2030, 15 years on from its declaration. In 2015 the rate of reduction in yearly incidence was 1.5%, which falls below the required target of 4-5%. These figures reflect the ongoing evolving challenges faced in the prevention and treatment of tuberculosis<sup>1</sup>.

Tuberculous meningitis (TBM) is the most lethal and disabling form of tuberculosis (TB), affecting >100,000 people per year (Wilkinson et al., 2017). TBM arises when *Mycobacterium tuberculosis (Mtb)* crosses the blood-brain barrier, causing severe inflammation and tissue damage. Inflammation also initiates metabolic derangement, contributing to ongoing neuronal injury despite effective antibiotics and anti-inflammatory therapy (Davis et al., 2019b). In HIV co-infection, TBM mortality approaches 50% while on therapy, and long-term disability is very common amongst survivors (Marais et al., 2011). The central hypothesis driving work undertaken within my PhD is that poor outcome in TBM and HIV-TBM is due to treatment which fails to provide adequate antibiotic penetration into central nervous system whilst simultaneously dampening counterproductive host responses.

The goal of my PhD was to address these failures by:

- i) evaluating the safety of a novel drug regimen which aims to increase bactericidal activity within the CNS in a phase 2A randomised controlled trial
- ii) embedding pathogenesis studies within the clinical trial to enable *in vivo* and *ex vivo* analysis of pathogenic mechanisms at play in HIV-associated TBM
- iii) developing robust quantitative clinical endpoints for TBM studies to better understand neurological outcomes, in particular cognitive and functional outcomes, and in turn inform studies of pathogenesis

This work was extended to include a case-control study of neurological complications related to SARS-CoV2 infection, which contributed knowledge to this field whilst serving as a control group for *ex vivo* analysis.

This introduction aims to cover two key areas, namely;

- i) current treatment options in TBM
- ii) current understanding of pathogenic mechanisms in TBM

## **1.2 Current treatment options in Tuberculous Meningitis**

In 1948 the modern era of tuberculosis treatment saw the first evidence of therapeutic response to streptomycin in pulmonary TB (Streptomycin in Tuberculosis Trials Committee, 1948). Isoniazid followed in 1952 with a key trial demonstrating improved efficacy when added to streptomycin (Anonymous, 1952) and in 1971 the addition of rifampicin and pyrazinamide led to reduction in treatment duration from 2 years to 6 months (Nitti, 1971). However, unlike in pulmonary TB where decades of clinical trials have instructed and refined treatment regimens in drug-sensitive and more recently drug-resistant TB, comparatively little evidence exists to guide optimal treatment in tuberculous meningitis. Here I evaluate evidence guiding the treatment of TBM in adults considering three aspects to successful management; i) effective antimicrobial treatments ii) controlling the host inflammatory response and iii) supportive

interventions to reduce raised intracranial pressure. I discuss TBM complicated by HIV co-infection, in particular timing of antiretroviral therapy and review the evidence for emerging therapies.

### **1.2.1 Antibiotics**

Current WHO guidelines for TBM are based on those developed to treat PTB and suggest treatment with two months of rifampicin, isoniazid, pyrazinamide and ethambutol followed by up to ten months of rifampicin and isoniazid for all patients (Anonymous, 2010). Although initiation of this regimen before the onset of coma is the strongest predictor of survival from TBM (Prasad et al., 2016), this regimen does not take into account the differential ability of antituberculosis drugs to penetrate the brain (Donald, 2010).

Introduced in 1952, isoniazid made immediate impact on mortality in all forms of tuberculosis. This drug penetrates the CNS freely (Ellard et al., 1993), and is a key chemotherapeutic agent in TBM with proven potent bactericidal activity within the first 2 days of treatment (Mitchison, 2000). Isoniazid metabolism occurs via the genetically polymorphic NAT2, with both fast and slow metabolisers described (Weber and Hein, 1979). In pulmonary TB studies have demonstrated an association between phenotype, exposure and clinical outcome (Weiner et al., 2003). One recent pharmacokinetic study has evaluated this association in TBM and found, using pharmacodynamic modelling that isoniazid exposure was associated with outcome, with a higher mortality rate in fast metabolizers with lower isoniazid exposures (Ding et al., 2020). These findings provide rationale to investigate higher doses of isoniazid in TBM, particularly in fast acetylators; a NAT2 stratified, randomized controlled trial in TBM is currently underway (NCT03787940).

Rifampicin does not penetrate the blood brain barrier as well with concentrations in CSF only 10-20% of that reached in plasma (Donald, 2010). Several studies have now investigated the efficacy of higher dose rifampicin, with varying results. In 2013 an open-labelled randomized phase 2 study in 60 Indonesian adults with TBM showed a 50% reduction in mortality with high dose rifampicin (600 mg, about 13mg/kg intravenously) compared to standard dose rifampicin (450mg, about 10mg/kg orally) (Ruslami et al., 2013b). A larger randomized placebo-controlled trial in Vietnam recruiting between 2011 and 2014 tested a higher dose of oral rifampicin (15mg/kg orally) alongside levofloxacin against standard therapy and did not show a mortality benefit (Heemskerk et al., 2016). A recent pharmacokinetic (PK) study in Uganda suggested that doses higher than 15mg/kg may improve outcomes by demonstrating ~8 and ~6 fold higher CSF exposures with 35mg/kg (oral) and 20mg/kg (IV) doses respectively compared to standard oral dose (10mg/kg) (Cresswell et al., 2021). Alongside this work, studies in PTB have shown that doses up to 35mg/kg are safe and well tolerated in the first two weeks of therapy (Boeree et al., 2015). Within this study, the highest doses of 30 and 35mg/kg showed highest early bacteriocidal activity measured by fall in colony forming units (CFU) and time to positivity. At two weeks 8 out of 14 patients taking 35mg/kg of rifampicin were culture negative compared 5 of 14 taking 20mg/kg, 0 of 15 taking 25mg/kg, 2 of 15 taking 30/kg and 3 of 8 controls (Boeree et al., 2015). This data may suggest that 15mg/kg within the aforementioned phase 3 RCT in TBM (Heemskerk et al., 2016) was not high enough and provides rationale for evaluating doses up to 35mg/kg in TBM. Several ongoing studies are now evaluating either the pharmacokinetics and/or efficacy of high dose rifampicin in adult and paediatric TBM (SIMPLE; NCT03537495; HARVEST; ISRCTN 15668391; SURE; ISRCTN40829906; ALTER; NCT04021121; INTENSE-TBM; NCT04145258). The safety of high dose rifampicin, in the context of HIV-associated TBM and when used in conjunction with



other adjunctive antibiotics and/or host directed therapies however is not well described.

Pyrazinamide was first introduced as an adjunctive agent to rifampicin to shorten treatment regimens in pulmonary TB (East African/British Medical Research Councils, 1972), and remains a pivotal drug in TB. Pyrazinamide has good CSF penetration with measured concentrations in CSF close to that of serum (Donald, 2010); however its dose in TBM has not been optimised. Though pyrazinamide has poor bactericidal activity in the first 2-4 days of treatment, studies in pulmonary TB have shown that thereafter (i.e. days 4-14) its activity matches that of isoniazid and rifampicin (Jindani et al., 1980, Botha et al., 1996). A recent pharmacokinetic study demonstrated good CSF penetration as well as high correlation between dose, plasma exposure and CSF concentration (Stemkens et al., 2019). The effect of higher CSF exposure on clinical outcome however is not known; studies to evaluate this whilst considering possible increased risk of hepatotoxicity and neurotoxicity (Pasipanodya and Gumbo, 2010, Torok et al., 2018) are needed. This is interesting given that pyrazinamide may also possess anti-inflammatory properties, via reduction of pro-inflammatory cytokine production (Manca et al., 2013).

Of the current recommended first line drugs, penetration of ethambutol is the poorest (Donald, 2010); highlighting whether ethambutol has a role within the standard regimen in TBM. This is also important in considering drug resistance in TBM, for which mortality rate approaches 100% (Tho et al., 2012, Patel et al., 2004, Vinnard et al., 2011). WHO guidelines for the treatment of rifampicin resistant- and multi-drug resistant TBM state that at least five effective drugs should be used initially including a fluoroquinolone and an injectable second line agent, and treatment should last 18-24 months (Falzon et al., 2017). Recommended core second line agents and their CSF penetrance are listed in Table 1.1.

Of the fluoroquinolones, ofloxacin was the first to be recognized as a potential effective treatment for tuberculosis (Tsukamura et al., 1985). In 2011 a randomized study in Vietnam investigated the pharmacokinetics and exposure-response relationships of three fluoroquinolones (ciprofloxacin, levofloxacin or gatifloxacin) in TBM (Thwaites et al., 2011). Population pharmacokinetic models describing the disposition of fluoroquinolones in CSF and plasma were used to determine exposure-response relationships through univariable analysis of clinical outcomes. Significant higher proportions of death and disability were observed for patients with lower or higher plasma and CSF fluoroquinolone exposures than for patients with intermediate exposures; a finding which may be explained by the increased permeability of the BBB in severe meningeal disease and/or reduced creatinine clearance in those with more severe systemic involvement and therefore those more likely to die. Nonetheless the study demonstrated improved clinical outcomes measured by survival, burden of disability and incidence disease relapse of fluoroquinolones when used prior to the onset of coma and informed dose finding for future studies (Thwaites et al., 2011). Subsequently two randomized controlled trials have evaluated the safety and efficacy of adjunctive fluoroquinolones in adult TBM with or without high dose rifampicin. In a phase 3 RCT 817 adults with TBM received either standard therapy or an intensified regimen including a higher dose of rifampicin (15mg/kg) as well as levofloxacin (20mg/kg); in this study levofloxacin did not improve outcomes (Heemskerk et al., 2016). The second, phase 2 RCT in Indonesia randomized 60 adults with TBM to receive standard dose or high dose rifampicin with either moxifloxacin (400mg) or ethambutol (750mg) and demonstrated no relationship between exposure to moxifloxacin in plasma and CSF and survival (Ruslami et al., 2013b). In a recently published RCT, children with TBM received either high dose rifampicin with or without levofloxacin (Paradkar et al., 2022). Although high dose rifampicin improved neurocognitive outcomes, this was not

statistically significant in the arm that included levofloxacin. It is possible that in this study levofloxacin negated in part the benefit of high dose rifampicin, or that the small sample size precluded a thorough assessment of potential benefit of adjunctive levofloxacin (van der Laan et al., 2016). This remains an area for consideration in future research trials, with no current RCT planned to evaluate adjunctive fluoroquinolones in either adult or paediatric TBM.

Linezolid a synthetic antimicrobial and the first agent of the oxazolidinone class, was licensed in 2000 for treatment of nosocomial pneumonia and skin infections caused by select gram positive bacteria (Ford et al., 2001, Brickner et al., 2008). The role of linezolid in tuberculosis was first investigated in the context of MDR tuberculosis. Early studies reported rapid sterilization of *M. tuberculosis* cultures following the administration of linezolid 600mg BD in addition to standard treatment (Fortun et al., 2005, von der Lippe et al., 2006) with subsequent studies demonstrating a role for linezolid as an effective treatment in drug resistant tuberculosis (Sotgiu et al., 2012). Broad tissue penetration of linezolid, including into the CNS (Nau et al., 2010) makes linezolid a favourable drug in TBM; however evidence to support the use in this context is limited. An observational study by Li et al demonstrated favourable clinical outcomes and a non-significant difference in adverse events in children with drug sensitive TBM treated with linezolid compared to control (Li et al., 2016). However, the study was a retrospective observational analysis with unblinded assessment of clinical outcomes. In adult TBM a retrospective cohort study of 33 adults with severe TBM demonstrated more rapid improvement in CSF parameters, recovery of consciousness and reduction of fever with adjunctive linezolid (Sun et al., 2014).

There has however been concern regarding safety of linezolid. The most common adverse events associated with LZD use in TB treatment are haematological toxicity

(mainly dose-dependent, occurring in up to 25%) and peripheral neuropathy (mainly duration-dependent, occurring in up to 31%) (Zhang et al., 2015). In one systematic review, haematological toxicity appeared to be dose related, with a significantly higher prevalence of anaemia (31.3% vs 13.6%,  $p=0.007$ ) and a lower prevalence of peripheral neuropathy (23.5% vs 37.2%,  $p=0.018$ ) in those taking a dose > 600 mg/day, compared to those taking < 600 mg/day (Zhang et al., 2015). Since LZD toxicity is thought to be related to trough concentrations once daily dosing is likely to result in less toxicity than twice daily dosing. This was demonstrated in a PK-toxicity analysis of a randomized controlled trial of LZD for TB, which demonstrated an inverse correlation between LZD trough concentrations and mitochondrial function. Furthermore, higher LZD trough concentrations correlated with the development of mitochondrial toxicity-related AE (Song et al., 2015). These findings were replicated in a hollow-fibre infection model demonstrating a favourable toxicity profile with once daily LZD dosing (Brown et al., 2015). The onset of symptoms related to linezolid toxicity are well described. Typically, anaemia usually occurs within 2 months, whereas peripheral neuropathy occurs between 2-4 months of therapy (Tang et al., 2015). Two recent studies however have shown that toxicity associated with linezolid in MDR-TB is rarely severe (Wasserman et al., 2022) and largely reversible (Conradie et al., 2020) and therefore may be acceptable in a disease where mortality is so high. Several ongoing and planned studies, besides LASER-TBM are evaluating either the pharmacokinetics or efficacy of linezolid in adult and paediatric TBM (SIMPLE; NCT03537495; ALTER; NCT04021121 and INTENSE-TBM NCT04145258).

### 1.2.2 Host directed therapies

In TBM adjunctive host-directed immune interventions which either enhance protective immunity or regulate pathological tissue-damaging immunity are needed (Davis et al., 2020).

Corticosteroids are the most widely used host directed therapy. The first randomised controlled trial of corticosteroids in TBM was conducted in 1969 and demonstrated a non-significant reduction in mortality (RR 0.53, 95% CI 0.39 to 1.37;  $n = 23$ ) (O'Toole et al., 1969). Since then, a further 6 published RCT have investigated efficacy of corticosteroids in reducing mortality associated with TBM (Girgis et al., 1991, Kumarvelu et al., 1994, Chotmongkol et al., 1996, Malhotra et al., 2009, Schoeman et al., 1997, Thwaites et al., 2004). A recent Cochrane review found that in a pooled analysis of these 7 trials alongside data from two unpublished trials there were 25% fewer deaths with corticosteroids (RR 0.75, 95% CI 0.65 to 0.87; 1337 participants) (Prasad et al., 2016). The largest of these was a randomised, double-blind placebo-controlled trial conducted in Vietnam in 545 patients with TBM (Thwaites et al., 2004). In this trial treatment with dexamethasone was associated with a reduction in risk of death (RR 0.69, 95% CI 0.52 to 0.92;  $p = 0.01$ ). Of the 9 trials included within the Cochrane review, 8 reported on neurological disability at 2 to 24 months' post diagnosis (Prasad et al., 2016). Pooled analysis of data from 1314 participants demonstrated no effect of corticosteroids between the two groups (RR 0.92, 95% CI 0.71 to 1.20). In the aforementioned Vietnamese study, there was also no significant reduction in the proportion of severely disabled patients (34 of 187 (18.2%) among survivors in the dexamethasone group vs 22 of 159 patients (13.8%) in the placebo group,  $p=0.27$ ) or on the combined outcome of death and severe disability at 9 months (OR 0.81, 95% CI 0.58 to 1.13;  $p = 0.22$ ). At 5 years there was no significant

association between dexamethasone treatment and disability status ( $p=0.32$ ) (Thwaites et al., 2004). This trial, as well as the analysis from this recent Cochrane review highlight the lack of evidence to support the efficacy of corticosteroids to reduce disability in TBM. Further work is required to investigate this.

There is a lack of efficacy data to inform the use of corticosteroids in HIV-associated TBM. Of the nine RCT taking place since 1969, only the 2004 Vietnamese study enrolled patients with HIV-1 co-infection (Thwaites et al., 2004). In this study, 98 of the 545 patients were co-infected with HIV. Although not powered to address the question of the efficacy of dexamethasone in HIV-associated TBM, there was no significant effect of dexamethasone on the combined endpoint of death and disability or on death alone (stratified relative risk of death 0.78; 95% CI 0.59 to 1.04;  $p=0.08$ ). A larger study to evaluate the role of dexamethasone in HIV-associated TBM has been completed and results are awaited (NCT03092817). The role of corticosteroids in the treatment of TBM-associated immune reconstitution inflammatory syndrome (IRIS) is unclear. Although in other forms of TB-IRIS use of prednisolone improves outcomes (Meintjes et al., 2010), there is a lack of evidence to support their use in TBM-IRIS. In a study to describe pathogenesis of TBM-IRIS, the use of corticosteroids often failed to prevent the onset of IRIS in HIV-1 infected patients with TBM (Marais et al., 2017).

A focus of recent research is the identification of genetic polymorphisms in immune response genes, in particular a single polymorphism in the leukotriene A4 hydrolase (LTA4H) promotor which plays a role in the balance of proinflammatory and anti-inflammatory eicosanoids thereby influencing expression of TNF alpha (Tobin et al., 2010). Studies in zebrafish and subsequently in humans have shown that expression of LTA4H can determine susceptibility to disease as well as response to corticosteroids (Tobin et al., 2012). In a retrospective analysis of patients enrolled to a trial of

adjunctive dexamethasone in TBM, survival benefit was restricted to homozygotes with a TT genotype of the LTA4H (hyperinflammatory) in contrast to CC (hypoinflammatory) genotypes where dexamethasone was associated with harm (Tobin et al., 2012). More recently in an analysis of patients enrolled to a study of intensified antituberculous regimens and adjunctive dexamethasone, LTA4H genotype predicted survival in HIV-1-uninfected patients with the TT genotype patients significantly more likely to survive than those with the CC genotype. In this study patients with the LTA4H TT genotype had high pro-inflammatory cytokine concentrations (IL-1 $\beta$ , IL-2, and IL-6). However, those with CT and CC genotypes had intermediate or lower concentrations respectively (Thuong et al., 2017). This may suggest that the suppression of inflammation by dexamethasone leads to survival benefit in patients with the TT genotype, however, it may be non-beneficial or even harmful in those with CT or CC genotypes. This highlights the potential role for individualized immunotherapy where adjunctive corticosteroids are given on the basis of pre-treatment genotyping and provides rationale for the LTA4H genotype stratified, randomised placebo-controlled phase III noninferiority trial evaluating adjunctive dexamethasone currently ongoing in Vietnam (NCT03100786).

Improved understanding of immunopathogenesis in TBM has led to discovery of target sites for immunotherapies. The cytokine TNF alpha has been a target in both animal and human studies. Thalidomide has a complex mechanism of action, including but not exclusively via inhibition of TNF-alpha. In a rabbit model of TBM, thalidomide was associated with survival benefit (Tsenova et al., 1999). In a safety and tolerability study using thalidomide at escalating doses, thalidomide was safe and well tolerated as an adjunctive therapy to treat children with stage 2 TBM (Schoeman et al., 2000). Clinical and radiological data also suggested improved outcome. The results of this study

supported a phase 3 randomised controlled trial in paediatric TBM to test thalidomide against placebo in stage 2 and 3 disease (Schoeman et al., 2004). Thalidomide was given at a dose of 28mg/kg/day for the first 28 days of treatment. Forty-seven children were enrolled, of which 30 received thalidomide. This study was terminated early as all adverse events and deaths occurred in the thalidomide arm. Debate around the influence of the high dose and late stage of disease on the adverse outcomes remains. Subsequent studies have suggested that thalidomide may still have a role in tuberculous mass lesions where treatment with corticosteroids has failed (van Toorn et al., 2015) and in children with CNS TB-related complications (van Toorn et al., 2021).

Early (within 2-4 weeks of commencing antitubercular therapy) antiretroviral (ART) therapy of HIV-associated tuberculosis is associated with survival benefit in patients with low CD4 counts (Abdool Karim et al., 2010, Blanc et al., 2011, Havlir et al., 2011). A metanalysis including 8 randomised controlled trials in pulmonary TB compared survival in patients in whom antiretroviral therapy was started within 1-4 weeks vs 8-12 weeks. Results demonstrated a survival benefit in patients newly diagnosed with tuberculosis and a CD4 count of less than 50/mm<sup>3</sup> where antiretroviral therapy was commenced within 1-4 weeks of diagnosis (Uthman et al., 2015). However, initiating such otherwise life-preserving therapy early during TB treatment may be complicated by more frequent immune reconstitution inflammatory syndrome (IRIS). In the CNS TBM-IRIS itself is associated with increased mortality (Marais et al., 2013). Early initiation of ART in TBM does not improve outcomes but rather increases the chance of grade 4 adverse events (Torok et al., 2011a); this data therefore supports the current recommendations stating that initiation of ART should be deferred until 4-6 weeks of anti-TB therapy has been completed.



Aspirin acts by irreversibly inhibiting the cyclooxygenase pathway of arachidonic acid metabolism, and thus reducing the downstream production of prostanoids (Vane, 1971). At low doses aspirin prevents ischaemic infarction through inhibitory effect on platelet and thrombus formation (Richman and Owens, 2017); which at high doses may be further augmented by production of 15-epi-lipoxins, 17R-resolvins and protectins known to contribute to resolution of inflammation (Spite and Serhan, 2010, Tobin et al., 2012). This potential to prevent infarction, as well as hasten resolution of cerebral inflammation, has led to its evaluation for use as an HDT in TBM. Three RCT have thus far investigated its potential. The first demonstrated a trend towards lower 3-month mortality and incidence of stroke in adults with TBM treated with 150mg OD of aspirin (Misra et al., 2010). The second showed a significant reduction in the incidence of new hemiplegia in children receiving high dose (1000mg OD) vs low dose (75mg OD) aspirin (Schoeman et al., 2011). The third demonstrated a potential reduction in new infarcts and deaths by day 60 in the aspirin-treated adults with microbiologically confirmed TBM (11/32 (34.4%) events in placebo vs. 4/27 (14.8%) in aspirin 81 mg vs. 3/28 (10.7%) in aspirin 1000 mg;  $p=0.06$ ) (Mai et al., 2018). In the latter, planned CSF analysis demonstrated aspirin dose-dependent inhibition of thromboxane A<sub>2</sub> and upregulation of pro-resolving CSF protectins. Although no increase in adverse events with high doses have been observed in these referenced studies, the safety of aspirin in combination with intensified antibiotics and in the context of HIV co-infection is not known.

Infliximab is a TNF-alpha inhibitor which is FDA approved for use in inflammatory bowel disease, rheumatoid arthritis and some seronegative arthropathies. Although most reports of infliximab in TB relate to the reactivation of latent tuberculosis, there are several case reports where corticosteroids have failed to control inflammation yet subsequent reintroduction of infliximab has led to a near complete resolution of

symptoms (Marais et al., 2021, Molton et al., 2015, Lee et al., 2012a). Other therapies to consider include interleukin receptor 1 inhibitors anakinra (IL-1 $\alpha$  and  $\beta$ ) (Keeley et al., 2020) and canakinumab (IL-1 $\beta$  only).

### **1.2.3 Potential Pathways for Future Host Directed Therapies for Tuberculous Meningitis**

Although host directed therapies are in use, they are limited in either efficacy or availability; in particular in HIV associated TBM. Therefore, the quest for more effective therapeutics is ongoing. Here I discuss potential therapies which target pathways highlighted in recent pathogenesis studies or draw on insights from other forms of TB or inflammatory conditions with shared mechanisms of pathogenesis.

#### *Statin Therapy Pathways*

HMG-CoA reductase inhibitors ('statins') are ubiquitously used in prevention and treatment of cardiovascular disease, but are also known to have immunomodulatory, anti-inflammatory and anti-oxidative properties. Several *in vitro* studies have demonstrated that statins enhance anti-inflammatory and inhibit pro-inflammatory functions in microglial cells and inhibit mechanisms involved in neurodegeneration (Kata et al., 2016, Churchward and Todd, 2014, Cordle and Landreth, 2005, McFarland et al., 2018). Anti-inflammatory properties may be due to modulation of isoprenylation (Waiczies et al., 2008) with downstream effects on inhibitory and stimulatory transcription pathways, or via allosteric inhibition of leucocyte function antigen (LFA)-1 integrin (Weitz-Schmidt et al., 2001) which is involved in the transmigration of activated T cells through the blood brain barrier. Neuroprotective effects may be due to

modulation of excitotoxicity, vascular function, angiogenesis, and/or reduced oxidative damage through nitric oxide stimulus (Bosel et al., 2005, Ponce et al., 2008).

Importantly, some studies have shown increased neuronal death with higher concentrations of statins (Michikawa and Yanagisawa, 1999, Tanaka et al., 2000, Schulz et al., 2004).

The potential of statins to effect CNS inflammation and neurodegeneration in other conditions are of interest given the shared mechanistic pathways in TBM. For example, animal models of multiple sclerosis (MS) show that statins skew immune responses towards an anti-inflammatory T-helper cell 2 response, inhibiting pro-inflammatory cytokines IL-2, IL-12 and IFN- $\gamma$  (Youssef et al., 2002). Patients with secondary progressive MS benefited from statin therapy (Chataway et al., 2014) with a phase 3 trial underway (NCT03387670). In a mouse model of traumatic brain injury, atorvastatin led to profound attenuation of T cell, neutrophil and natural killer cell invasion into the CNS, and reduction in production of pro-inflammatory cytokines (IFN- $\gamma$  and IL-6) and chemokines (CCL5 and CXCL10) (Xu et al., 2017). In a double-blind randomised trial involving 36 patients with traumatic brain injury, rosuvastatin given for 10 days in the acute phase of injury significantly reduced TNF- $\alpha$  which correlated with a reduction in disability scores (Sanchez-Aguilar et al., 2013). Other conditions where the role of statins has been explored include Alzheimer's disease (Jick et al., 2000), and Parkinson's disease (Yan et al., 2019). Further, statins may be associated with reduced risk of tuberculosis (Lai et al., 2016). In a TB murine model, adjunctive simvastatin shortened time to culture clearance by 1 month, enhanced bacterial killing, and decreased culture-positive relapse and enhance bacterial killing (Dutta et al., 2016, Skerry et al., 2014, Parihar et al., 2014). Clinical trials (NCT03456102, NCT04147286) will investigate the efficacy of statins in pulmonary tuberculosis. Given their potential use as an adjunctive TB therapy, their lipophilic properties allowing good penetration to

the CNS, as well as their potential as an anti-inflammatory and neuroprotective agent, statins may have a role as a HDT in TBM; trials to explore this hypothesis are needed.

### *Glutamate 'grabbing' drugs*

Excessive glutamate and neuro-excitotoxicity are thought to contribute to brain injury and cell death in TBM. In one study, RNA sequencing of whole blood and CSF from children with TBM demonstrated significant enrichment of transcripts associated with neural excitotoxicity predominantly driven by glutamate release, NMDA receptor binding and uptake (Rohlwink et al., 2019). This mechanism is thought to contribute to brain injury and cell death in other neurological conditions such as stroke, epilepsy, traumatic brain injury, Alzheimer's and Huntington's disease (Meldrum, 2000, Wang and Reddy, 2017). Therapeutics which aim to reduce glutamate excitotoxicity either by i) modulating the downstream effects of glutamate via NMDA receptor binding or ii) reducing extracellular glutamate (e.g. glutamate 'grabbing') may have a role in the treatment of TBM. In acute stroke, a similar approach was taken however although animal studies were promising, randomised trials in humans assessing efficacy of NMDA antagonists largely failed (Jia et al., 2015, Kalia et al., 2008, Grupke et al., 2015). Therapeutics have been designed to reduce glutamate induced excitotoxicity by lowering blood glutamate concentration thus leading to a larger natural glutamate gradient between the brain and blood thereby facilitating the efflux of extracellular brain glutamate into the blood (Castillo et al., 2016). In an animal study riboflavin (vitamin B<sub>2</sub>), selected for its ability to interact with Glutamate-Oxaloacetate transaminase (GOT) to significantly reduced blood glutamate levels compared to placebo (da Silva-Candal et al., 2018). In a randomised trial, riboflavin was correlated with improvement of disability when given intravenously in adults with acute stroke (da Silva-Candal et al., 2018). A number of studies have explored the neuroprotective properties of riboflavin including in conditions such as migraine and Parkinson's disease (Marashly and

Bohlega, 2017). It is unclear whether drugs such as riboflavin, or others which reduce glutamate neuro-excitotoxicity, have a role as an adjunctive therapy to promote neuroprotection in TBM; however, given the emerging body of evidence which suggest involvement of the glutamate-glutamine pathway, this is a potential area of interest for future studies.

#### *Tryptophan Pathway Drug Targets*

Tryptophan is an essential amino acid which can either be converted to serotonin or oxidized kynurenines via indoleamine 2,3-dioxygenase (IDO1). Further oxidization occurs to convert kynurenine to kynurenic acid, which has neuroprotective properties. Prior studies have shown that *M.tb* induces marked upregulation of IDO-1 expression in both human and murine macrophages *in vitro* (Blumenthal et al., 2012); and that blockade of IDO activity reduces both clinical manifestations of TB as well as microbial and pathological correlates of the human TB syndrome in macaques.(Gautam et al., 2018). In an observational cohort study of TBM, low CSF tryptophan levels were found in those who survived, compared to non-survivors or controls (van Laarhoven et al., 2018). It is therefore unclear in TBM whether drugs which block IDO-1 such as indoximod, an immunometabolic adjuvant that is current under investigation in cancer therapy (Fox et al., 2018), would cause benefit or harm. It is plausible that improved survival seen in those with low CSF tryptophan is due to increased availability of kynurenic acid which has neuroprotective action via glutamate receptors and reactive oxygen species. Further investigation into the influence of tryptophan and its downstream metabolites on pathogenesis in TBM is required in order to establish suitable targets along this pathway for HDT.

#### *Eicosanoid Modulating Drugs*

Eicosanoids are arachidonic acid derived lipid mediators that trigger pro-and anti-inflammatory responses and include prostaglandins, resolvins, lipoxins, and leukotrienes which serve as signalling molecules, modulating inflammation and cell death in TB (Ricciotti and FitzGerald, 2011). A delicate balance in eicosanoid levels is crucial for *M.tb* control and regulating the production of pro-inflammatory cytokines (Young et al., 2020).

Non-steroidal inflammatory drugs (NSAID), which exert their effects by inhibiting cyclooxygenase (COX) activity may lead to reduction of excessive inflammation in TBM. As discussed, aspirin, a non-selective COX inhibitor has been investigated in three trials in TBM with variable outcomes (Mai et al., 2018, Schoeman et al., 2011, Misra et al., 2010). New generation NSAID with more selective inhibition of COX2 may have more favourable safety profiles. Phase 1 trials to assess the safety and bactericidal activity of celecoxib and etoricoxib in healthy volunteers with a view to developing these agents as HDTs for drug sensitive TB are currently underway (NCT02602509; NCT02503839). Although trials to further investigate the role of aspirin in TBM are underway, future research should consider the potential contribution of newer more selective COX2 inhibitors in TBM.

#### *Phosphodiesterase Inhibitors*

Phosphodiesterase inhibitors (PDE-i) are small-molecule inhibitors that reduce inflammation by increasing intracellular cyclic adenosine monophosphate and cyclic guanine monophosphate (Page and Spina, 2011). Phosphodiesterase 4 (PDE-4) inhibitors such as roflumilast have shown to be effective in the treatment of numerous inflammatory conditions including chronic obstructive inflammatory disease (Calverley et al., 2009). PDE-4 is expressed within the cortex and hippocampus and animal models suggest that inhibition of PDE-4 may have a beneficial role in CNS conditions

where inflammation plays a role in pathogenesis (Schaal et al., 2012, Atkins et al., 2007, Gong et al., 2004, Gonzalez-Garcia et al., 2013, Wu et al., 2017). In animal models of pulmonary TB, inhibition of PDE-3 (cilostazol), PDE-4 (roflumilast) and PDE-5 (sildenafil) have all increased bacterial clearance and reduced pro-inflammatory cytokines which contributed to a reduction in neutrophil infiltration and lung pathology (Subbian et al., 2011, Maiga et al., 2015, Maiga et al., 2012, Konrad et al., 2015). The role of phosphodiesterase inhibitors has not been studied in TBM but the properties above make them intriguing candidates for adjunctive therapy in TBM.

#### **1.2.4 Supportive therapies**

Rich and McCordock were the first to describe the pathogenic mechanisms which lead to central nervous system tuberculosis (Rich, 1933). Research since then has enabled better understanding of the natural history including the neurological sequelae such as hydrocephalus, vasculitis leading to cerebral infarction and metabolic abnormalities especially hyponatremia. Early recognition and management of these phenomena remains integral to the treatment of patients with TBM.

- Hydrocephalus and raised intracranial pressure

The inflammatory infiltrate within the subarachnoid space or the ventricular pathways may lead to disruption of CSF flow resulting in hydrocephalus. Hydrocephalus can be communicating (caused by abnormal flow through the basal cisterns), or non-communicating (usually a later complication due to obstruction at the level of the fourth ventricle). Communicating hydrocephalus is more common and can be managed medically however may require intervention if progressing. Non-communicating hydrocephalus requires rapid intervention. CSF diversion techniques such as

ventriculoperitoneal shunts (VPS) and endoscopic third ventriculostomy are the mainstay of surgical treatment for hydrocephalus (reviewed in (Rajshekhar, 2015). Evidence as to which technique is most effective is lacking.

A systematic review of 1038 adults and children with TBM and hydrocephalus demonstrated good outcome, defined as Glasgow Outcome Scale 4 or 5 (Table 1.2) in 58.2% of patients. Good outcomes were observed in more patients with less severe disease specifically those found to be Grade I (78.57%) and II (65%) compared to those with more severe (Grade IV disease) where only 31.5% survived (Table 1.2). Subgroup analysis demonstrated that good outcomes occurred in significantly fewer patients with HIV-1 associated TBM with only 25% patients of patients achieving a good outcome compared to 61% of HIV negative patients (Rizvi et al., 2017). In a study of 30 participants with HIV-associated TBM and hydrocephalus, participants underwent VP shunt placement and outcomes were compared to age and gender matched HIV negative controls. Patients were followed up at two time points; discharge (short term) and three months after VP shunt insertion (long term). Although short term outcomes were only marginally better in the HIV negative group, long term outcomes differed significantly with 66.7% mortality and 76.2% poor outcome in HIV positive patients compared to 30.8% mortality and 34.6% poor outcome in the HIV negative controls. Their study demonstrated that HIV seropositivity is an independent predictor of poor outcome, although they did identify that in patients with less severe disease at presentation, 80% had good outcomes. By contrast to previous studies these results suggest a role of VP shunting in HIV-associated TBM in patients with less severe disease (Sharma et al., 2015). In paediatric TBM, a recent study showed that there is an association between the severity of hydrocephalus and CSF immune biomarkers GFAP and S100B (Rohlwink et al., 2017). It remains unclear as to whether this is due to the secondary compressive effect on brain parenchyma or whether these inflammatory mediators are involved in the



pathogenesis of hydrocephalus. Further research is required to establish best evidence-based practice for the treatment of this common complication in TBM in particular for HIV associated disease.

Although hydrocephalus is the most common cause of raised ICP, elevated ICP can also be caused by other pathological processes within the CNS. In TBM, meningeal pathology may extend into the parenchyma and lead to encephalitis, whilst obliterative vasculitis within the vessels leads to infarction. These processes may result in cytotoxic and vasogenic oedema. The presence of parenchymal pathology may lead to failure of cerebral vascular autoregulation. Metabolic abnormalities such as hyponatraemia, hyperthermia and hypercapnia can cause further dysregulation. Thus, clinical management should be directed at the frequent monitoring and correction of abnormalities in gas exchange and tissue oxygenation, through mechanical ventilation (if necessary), meticulous fluid and electrolyte management, monitoring and intervention to treat raised intracranial pressure where appropriate as well as adequate temperature control. When there is no surgical intervention indicated, yet ICP remains high hyperosmolar agents, most commonly mannitol, may be effective yet a randomised control trial to test this hypothesis is required (Oddo et al., 2009, Francony et al., 2008).

- Hyponatraemia

Hyponatraemia defined as a plasma sodium level  $<135\text{mmol/L}$  occurs in 40-50% of patients with TBM (Misra et al., 2016). Several mechanisms exist. Cerebral salt wasting (CSW) is characterised by natriuresis, hyponatremia and volume contraction in response to brain injury (Sterns and Silver, 2008). The syndrome of inappropriate anti-diuretic hormone (SIADH) is also associated with brain injury and occurs due to excessive release of antidiuretic hormone from the posterior pituitary gland resulting in

inappropriate, continued secretion or action of the antidiuretic hormone arginine vasopressin (AVP) despite normal or increased plasma volume leading to hyponatraemia (Moller et al., 2001). In a prospective hospital-based study conducted in India, of 76 patients with TBM, 34 (44.7%) had hyponatremia due to CSW in 17, SIADH in 3 and miscellaneous causes in 14 (Misra et al., 2016). Distinguishing between CSW and SIADH is critical: their presentations are similar, but management is different. By convention, SIADH is managed by fluid restriction and cerebral salt wasting by fluid administration. Some suggest that both conditions can be treated with hypertonic saline (Sterns and Silver, 2008), whereas others state that fluid restriction, the traditional treatment for SIADH, has had little benefit in meningitis and might result in worsening hypovolaemia and harm (Moller et al., 2001). This complex and often overlooked complication in TBM should be further investigated to define optimal investigation and management.

- Tuberculomas

Tuberculomas can occur together with or independently of TBM. Clinical presentation depends on site and includes seizures, focal neurological weakness or symptoms of raised intracranial pressure due to hydrocephalus or mass effect. Tuberculomas commonly present as a feature of paradoxical worsening in patients treated for TB or in HIV-1 infected patients starting ART. In a randomized study to assess effect of dexamethasone on TBM related cerebral MRI changes in Vietnam, 43 patients receiving either dexamethasone (n=24) or placebo (n=19) underwent serial MRI scans. The number of patients with one or more tuberculomas rose from 64% (14 of 22) before treatment to 74% (20 of 27) after 60 days. There was no effect of dexamethasone on incidence of tuberculoma formation or on resolution of tuberculomas (Thwaites et al., 2007). The mainstay of treatment remains antituberculous therapy, the duration of which

is debated due to a lack of evidence in this area. It is unknown as to whether persistent radiological enhancement of intracranial tuberculomas following completion of 9-12 months of anti-TB therapy represents ongoing active TB, an inflammatory response in a lesion without active TB, or revascularisation. A recent consensus report suggested that ongoing enhancement does not represent treatment failure, however studies to evaluate optimal duration of treatment for CNS tuberculomas are required (Marais et al., 2019).

In some cases, where there is diagnostic doubt or where the size and anatomical location of the tuberculoma is causing clinical worsening, surgical excision may be required. Stereotactic craniotomy and excision of superficial small tuberculomas and microsurgery are procedures now used. In cases where there is no response to dexamethasone alternative anti-inflammatory agents have been tried, particularly when the tuberculoma involves the optic chiasm and threatens vision including thalidomide or infliximab (Schoeman et al., 2006, de la Riva et al., 2013, Roberts et al., 2003, Molton et al., 2015, Lee et al., 2012a).

- Vasculitis and stroke

Stroke in TBM occurs in 15-57% of patients with TBM depending on the imaging modality used: CT reveals stroke in 13-35% and MRI in ~60% (Thomas et al., 1977, Shukla et al., 2008, Kalita et al., 2009). They are usually multiple, bilateral and occur most commonly in deep grey matter structures including the caudate, anterior thalamus, anterior and genu of the internal capsule, namely the 'tubercular zone (Misra et al., 2011) (Figure 1.1). The macroscopic pathological appearance in the brain vasculature is that of gelatinous fibrocellular leptomeningeal infiltrates initially enveloping the vessels including the carotid arteries, middle cerebral arteries and their branches. Vasculitis within the affected vessels may occur with intimal proliferation.

These process with or without superadded thrombosis likely leads to cerebral infarction(Thomas et al., 1977). There is no established prevention or treatment for stroke in TBM. There is no evidence to support corticosteroids in the prevention of stroke (Thwaites et al., 2004). Aspirin discussed earlier within this chapter has antiplatelet, anti-aggregant, anti-inflammatory and antioxidant properties, and in some cases has been shown to prevent stroke(Schoeman et al., 2011, Mai et al., 2018). It is unknown however whether other antiplatelet therapies such as clopidogrel may have a role, or whether treatment of acute stroke occurring within a 4 hour window should consider thrombolysis. Further research in the area of stroke prevention and treatment in TBM is required.

### **1.2.5 Conclusions and research priorities**

- The neurological presentations of tuberculosis are the most lethal and under-researched manifestations of TB which remains a major global health problem.
- Current anti-tuberculous therapy regimens for TBM are based on those which are efficacious in PTB but do not consider the differing efficacy of drugs across the BBB.
- Further research is required to investigate the safety and efficacy of intensified therapy regimens and newer anti-tuberculous agents to treat CNS tuberculosis.
- Corticosteroids have proven mortality benefit except in HIV-associated TBM where, as yet, no sufficiently powered study has been able to prove benefit or lack thereof. More research is required to develop and evaluate novel host directed therapies. Immune response phenotypes and genetic polymorphisms may direct individualized immune therapies and mediators of the innate immune response may provide targets for the development novel therapies.
- Stroke is a major cause of morbidity and mortality in TBM. Recent studies have shown a potential benefit of aspirin in the prevention of stroke as well as in the modulation of the host immune response in TBM.
- There is currently no significant evidence base to guide management of hydrocephalus in HIV-1- infected TBM. A large randomized clinical trial is required to investigate outcomes comparing available CSF diversion techniques in this particularly vulnerable subgroup of patients.
- A treatment algorithm (Figure 1.2) gives a practical holistic approach to the management of patients with tuberculous meningitis

## 1.3 Pathogenesis of Tuberculous Meningitis

### 1.3.1 From primary infection to the central nervous system

#### *The systemic immune response to tuberculous infection*

Tuberculosis (TB) infection occurs through the inhalation of infectious droplets of aerosolised *Mycobacterium tuberculosis* (*M.tb*), which cross the lung epithelium and infect lung alveolar macrophages, neutrophils and dendritic cells (DC). This leads to the secretion of antimicrobial peptides, cytokines (including interleukin-1 $\alpha$  and  $\beta$ , tumour necrosis factor alpha (TNF- $\alpha$ ), interleukin (IL)-6 and -12, chemokines, lipoxins that may stimulate necrosis and contribute to immune protection, and prostaglandins that may induce apoptosis (O'Garra et al., 2013). Under the influence of IL-12 and chemokines CCL-19 and -21, infected DC migrate to the local draining lymph nodes to stimulate the differentiation of T-helper I (Th1) cells. Th1 cell release of interferon gamma (IFN- $\gamma$ ) and TNF- $\alpha$  at the site of infection activates macrophages and DC to produce cytokines and antimicrobial factors that contribute to containment of the TB bacillus (O'Garra et al., 2013). This inflammatory process results in the formation of a granuloma, which encapsulates the infected cells and retards the replication of TB bacilli and can lead to latent infection. However, in the elderly, immunocompromised or very young in particular, the ongoing immune response may progress to active primary TB disease associated with tissue destruction in the lung and dissemination of the TB bacillus to other organ systems (Kumar and Kumar, 2010, Coico and Sunshine, 2009).

#### *Dissemination to the brain*

Dissemination of TB involves seeding of *M.tb* to other sites including the central nervous system (CNS). Various mechanisms by which the bacilli migrate into the lymphatic system or blood stream have been suggested (Krishnan et al., 2010, Jain et

al., 2006, Nguyen and Pieters, 2005). Bacterial proteins- early secretory antigenic target 6kDa (ESAT-6) and culture filtrate protein 10kDa (CFP-10) are involved in cell lysis, while heparin binding haemagglutinin adhesion (HBHA) aids *M.tb* translocation across the epithelium without lysis (Krishnan et al., 2010). *M.tb* may also invade and traverse vascular endothelial cells (Jain et al., 2006), replicate in lymphatic endothelial cells (Lerner et al., 2016), and be trafficked to distant locations in phagocytes (Krishnan et al., 2010). Furthermore, mycobacteria are able to survive and replicate in infected macrophages and lymphatic endothelial cells (LEC) surrounding granulomas in the lymph nodes. Research on LEC demonstrates that although *M.tb* bacilli are initially phagocytosed upon infecting the cell, through their genetic locus termed the region of difference 1 (RD1), the bacilli are able to escape from the phagosomes into the cytosol, where they more readily replicate. For those bacteria remaining in the phagosome they are able to prevent fusion of lysosomes with the phagosome, also in an RD1 dependent manner, thereby allowing bacterial replication in the phagosome and contributing to lymphatic tuberculosis (Lerner et al., 2016). The activation of LEC by IFN- $\gamma$  is key to restricting these RD1 mechanisms of replication (Lerner et al., 2016). Additionally, host immunity and *M.tb* strain variation may play a role; polymorphisms in the genes encoding for antigen recognition and macrophage activation (Hawn et al., 2006) or impaired pro-inflammatory cytokine release may influence the ability of the initial innate response to control infection (Krishnan et al., 2010). TB strains assigned greater virulence, like the lineage 2 strain, which is postulated to subvert the innate immune response, promoting its survival and replication and thereby more severe disease (Caws et al., 2008, Fernando et al., 2007, Caws et al., 2006).

The CNS is protected from influx of potentially harmful blood-borne bacteria by 2 vascular barriers; the blood-brain barrier (BBB) and the blood–cerebrospinal fluid barrier (BCSFB) (Pulzova et al., 2009) (Figure 1.3). The BBB is mainly formed by brain

microvascular endothelial cells that are characterised by intercellular tight junctions and a paucity of endocytic vesicles and fenestrae, and exhibit dedicated transport mechanisms to regulate influx and efflux across the CNS and blood compartments (Be et al., 2009, Pulzova et al., 2009). Pericytes, embedded within a basement membrane, and astrocytes' end-feet support the endothelial cells and also make an indispensable contribution to BBB integrity. In contrast, the BCSFB is composed of choroid plexus epithelial cells joined together by tight junctions and the arachnoid membrane. However, despite these protective mechanisms, *M.tb* bacilli migrate across these barriers. *In vitro* and animal models demonstrate that *M.tb* invades and traverses brain endothelial cells in the microvasculature through rearrangement of their actin (Jain et al., 2006, Be et al., 2009). Further, the *M.tb* gene *Rv0931c* (pknD) has been identified as a potential virulence factor promoting CNS infection in certain TB strains, as it enables the bacilli to interact with extracellular factors on the brain endothelium facilitating bacillary endothelial adhesion (Be et al., 2009). Another potential route of entry is the 'Trojan horse' mechanism by which *M.tb* are trafficked in infected macrophages and neutrophils across the BBB (Nguyen and Pieters, 2005).

Once the TB bacilli gain access to the brain, limited local innate immunity allows their survival and replication and the development of silent tuberculous lesions. Based on postmortem studies, Rich and McCordock suggested that TBM is initiated by the rupture of one of these lesions, the Rich focus, located under the cortical pia or adjacent to the meninges or ventricles, which releases *M.tb* bacilli into the sub-arachnoid space causing a granulomatous infection of the meninges (Rock et al., 2008, Donald et al., 2005, Rich A, 1933, Dastur et al., 1995) and the subsequent induction of inflammation. Recently the relationship between the Rich focus and the onset of miliary tuberculosis has been reviewed. Rich and McCordock did not acknowledge a role of miliary tuberculosis in the pathogenesis of TBM, however, several studies since have

suggested that the bacteraemia seen in these cases increased the likelihood that a meningeal or sub-cortical focus is established with subsequent rupture giving rise to TBM (reviewed in (Donald et al., 2005)).

### **1.3.2 Pathogenic and pathophysiological mechanisms within the brain**

#### *The host immune response to tuberculosis in the brain*

Microglia within the cerebral parenchyma are the principal CNS cells infected by *M.tb* (Peterson et al., 1995, Rock et al., 2005) and are involved in immune regulation. Other CNS cells that have potential roles in this process are astrocytes and neurons (Rock et al., 2005, Randall et al., 2014) (Figure 1.3). Randall *et al* have demonstrated direct infection of neurons with *M.tb*, however, the effect on neuronal function and implications for intercellular interactions is not clear (Randall et al., 2014). Although not as prominent as microglia in their role, astrocytes have also been implicated with a study by Rock and colleagues demonstrating 15% of astrocytes having cell associated bacilli (average of 1.3 bacilli per cell) compared to 76% of microglia (average of 4.2 bacilli/cell) under the same conditions (Rock et al., 2005).

*M.tb* is recognized by microglial cells via innate immune and neuro-specific receptors, including pattern recognition receptors. The toll-like receptors (TLR), a family of ten pattern recognition molecules, play a crucial role in innate immunity. Internalisation of *M.tb* by human microglia is dependent on CD14, a monocyte differentiation antigen, which binds to lipopolysaccharide with TLR-4 (Wright et al., 1990). This was demonstrated in a study in which uptake of non-opsonized tubercle bacilli by microglia was reduced by 64% and 62% in the presence of anti-CD14 monoclonal antibodies and soluble CD14 ligand respectively. This is in contrast to peripheral mononuclear phagocytes where CD14 neutralising anti-CD14 antibodies did not affect bacillary



uptake (Shams et al., 2003), but interestingly the presence of CD14 led to upregulation of CD14 expression on these cells perhaps facilitating pathogenic immune responses. This receptor, along with the  $\beta$ 2-integrin CD18 and TNF- $\alpha$ , is also involved in the formation of histologically characteristic multinucleated giant cells seen at autopsy and experimentally identified in porcine microglia infected with *Mycobacterium bovis* (Peterson et al., 1996).

Activation of microglia leads to secretion of a number of cytokines (Figure 1.3). In murine microglia the intracellular signalling pathways leading to cytokine release are *M.tb* inducible leading to a pro-inflammatory response through NADPH oxidase-dependant reactive oxygen species (ROS) generation (Yang et al., 2007). Although cytokines play a critical role in the host defence to infection with *M.tb*, they can also mediate inflammation. Tumour Necrosis Factor (TNF) is central to the pathogenesis of central nervous system tuberculosis (Mastroianni et al., 1997, Tobin et al., 2012, Tsenova et al., 1999). It has a protective role in the immune response to mycobacteria (Kaplan and Freedman, 1996); but is also associated with pathology *in vivo* via induction of fever, activation of the hypothalamo-adrenal axis and by triggering the release of other cytokines (Hashimoto et al., 1991, de Vries et al., 1997, Ramilo et al., 1990). Local TNF- $\alpha$  production in the CNS also increases permeability of the BBB and thus influx of other immune mediators to the CNS (de Vries et al., 1997). In a murine CNS TB model, Tsenova *et al* demonstrated a correlation between levels of TNF- $\alpha$  and the extent of cerebral pathology as measured by CSF leukocytosis, protein accumulation, meningeal inflammation, persistence of bacillary load and clinical deterioration (Tsenova et al., 1999). TNF- $\alpha$  antagonists such as thalidomide (Tsenova et al., 1998) and analogues thereof (Tsenova et al., 2002), used in rabbit models of TBM, downregulated the production of TNF- $\alpha$  and subsequently improved survival. This finding was not replicated in human studies where a clinical trial of thalidomide

used in conjunction with standard antituberculous therapy and corticosteroids in children with TBM was stopped early due to adverse events associated with thalidomide use (Schoeman et al., 2004). Since then, there has been some suggestion of benefit with the use of thalidomide in the context of tuberculous mass lesions (van Toorn et al., 2015).

CSF concentrations of IL-6 independently associate with more severe presentation of TBM (Simmons et al., 2006). In this context it is unclear whether this is due to its pro-inflammatory or anti-inflammatory effects. In murine models of TB, IL-6 has been implicated in the stimulation of IFN- $\gamma$  production but not necessarily essential for the protective immunity to *M.tb* (Saunders et al., 2000). It may also have an anti-inflammatory role by suppressing gene expression of pro-inflammatory cytokines (Xing et al., 1998). In a study by Rock *et al*, pro-inflammatory cytokines other than TNF- $\alpha$ , IFN- $\gamma$  and IL-6 found to be secreted by microglia in response to TBM included IL-1 $\beta$ , CCL2, CCL5 and CXCL-10. In contrast to microglia, astrocytes produced only moderate amounts of CXCL10 (Rock et al., 2005). Other cytokines, confirmed experimentally, to be secreted by microglia following *M.tb* stimulation include: IL-1 $\alpha$ , IL-10, IL-12p40, G-CSF, and GM-CSF (Curto et al., 2004, Yang et al., 2007).

Recent focus has turned to the pathogenic role of inflammatory mediators such as DAMPs (Damage Associated Molecular Patterns) and PAMPs (Pathogen Associated Molecular Patterns) (Chen et al., 2016), their validity as biomarkers of cerebral injury (Rohlwink et al., 2017), and as potential targets for novel host directed therapies in TBM (Berger et al., 2018). PAMPs are by-products released from pathogens that are recognised by host cell receptors subsequently leading to activation of the innate immune response. DAMPs, which are released by damaged host immune cells, interact with PAMPs leading to an accelerated cycle of cell death and injury. Host

poly(ADP-ribose) polymerase 1 (PARP1; also known as ARTD1) is an ADP-ribosylating enzyme essential for initiating various forms of DNA repair (Ko and Ren, 2012). Recent studies have suggested a role for PARP1 in the pathogenesis of TBM via its potential to modulate the release and activation of DAMPs. This includes high mobility group box-1 (HMGB1), a non-histone nuclear DNA binding protein expressed in all mammalian cells, and S100B, a protein synthesised by astrocytes, oligodendrocytes and Schwann cells known to be involved in cell-to-cell communication, cell growth, and intracellular signal transduction, as well as the development and maintenance of the CNS (Heizmann et al., 2002). These biomarkers of cerebral injury are known to increase in TBM (Rohlwink et al., 2017), and therefore suggest that PARP1 may be a potential new target in the development of host directed therapies (Mahon and Hafner, 2015). S100A8/9, also from the S100 family has a role in neutrophil chemoattraction and stimulation (Ryckman et al., 2003) and is implicated in the pathogenesis of tuberculosis in pulmonary disease. In HIV-1 uninfected patients raised S100A8/9 in serum correlated with increased radiographic disease severity (Pechkovsky et al., 2000, Gopal et al., 2013). In TBM Marais *et al* demonstrated that in patients with TBM and HIV infection, levels of S100A8/9 were significantly elevated two weeks after the initiation of antiretroviral therapy in those who developed Immune Reconstitution Inflammatory Syndrome (IRIS) defined as a paradoxical worsening of infection despite adequate treatment following the initiation of antiretroviral drugs, compared to those who did not. This observation may explain in part the ongoing paradoxical inflammation observed in IRIS (Marais et al., 2014).

In addition to the inflammatory milieu described above, there are several other factors implicated in the pathogenesis of TBM, in particular the subsequent increasing permeability of the BBB and influx of inflammatory mediators and cells (Figure 1.3). Vascular Endothelial Growth factor (VEGF) is a potent endothelial growth factor playing

diverse roles in vasculogenesis and angiogenesis. In tuberculosis it is now considered a useful biomarker of disease, where it may be used as an indicator of active vs latent disease activity or a marker of extrapulmonary vs primary lung disease. In several types of cancer, VEGF inhibitors such as Bevacizumab are well established as an effective therapeutic approach. In ischaemic conditions of the brain, VEGF has a pathological and protective role depending on pathogenic stage due to either effect on permeability of the microvasculature or subsequent reparative angiogenesis. In age related neurological conditions such as Alzheimer's disease, Parkinson's disease and motor neuron disease however, VEGF is thought to be pathogenic due to its effect on BBB dysfunction (Lange et al., 2016). In these conditions the effect of VEGF on increasing endothelial permeability is clear, however the mechanism by which this happens remains less understood, with possibilities including its effect on cell-cell junctions including tight junctions and adherens junctions, as well as on transcytosis (Ayata and Ropper, 2002). In TBM, VEGF disrupts the permeability of the BBB (van der Flier et al., 2004, Kim et al., 2008), which has been proposed as a mechanism by which dexamethasone exerts efficacy as a host directed therapy in TBM. Also interesting are its neuroprotective effects which have been more thoroughly explored in the context of amyotrophic lateral sclerosis where low VEGF has been reported with BBB dysfunction and the therapeutic use of a VEGF analogue (VEGF-A<sub>165</sub>) is currently under investigation in a clinical trial (NCT02269436). The release of intercellular and vascular adhesion molecules (ICAM and VCAM) as well as matrix metalloproteinases (MMP) from inflammatory cells within the CNS have also been shown to increase the permeability of the BBB (Rai et al., 2014).

### 1.3.3 Metabolic factors in the host

Neuroendocrine-associated metabolic abnormalities are common in TBM. In an observational study of patients newly diagnosed with TBM, neuroendocrine dysfunction occurred in half (More et al., 2017). This is likely due to the tendency for TBM to affect the basal structures such as the interpeduncular fossa, cisterna ambiens and cisterna pontis, which surround the pituitary gland, pituitary stalk and hypothalamus (Figure 1.6). Exudates here lead to oedema, perivascular infiltration and subsequent microglial reactions known collectively as 'borderzone encephalitis'. Subsequent metabolic abnormalities include gonadotropin deficiency, hyperprolactinemia, thyrotropin deficiency, corticotropin deficiency and somatotrophic hormone deficiency (More et al., 2017). Hyponatraemia is also common, discussed earlier within this chapter. Corticotropin deficiency may be modulated by treatment with dexamethasone; pituitary dysfunction and its role in the pathogenesis of TBM remains an area of interest, particularly given the observations of worsening outcomes and possible benefit of cortisol replacement.

Glucose and its metabolic effects are also of interest. In a pivotal study where adjunctive dexamethasone was found to decrease short term mortality in HIV-1 uninfected patients with TBM, low glucose at presentation predicted poor outcome (Thwaites et al., 2004). In more recent studies, including those in children, CSF glucose, lactate and protein levels have been linked to poor outcome (Bang et al., 2016, Simmons et al., 2006). If these metabolic markers are simply markers of disease activity as in most forms of neurological infection, then this finding is unsurprising. Mason *et al* demonstrated that the increase in lactate levels commonly observed in the CSF of patients who go on to develop poor outcomes is of the L-form and therefore solely a response of the host to infection, rather than being of microbial origin (Mason

et al., 2016). This finding contributes to our research group's hypothesis that in the context of neuroinflammatory-inducing infection, energy flow in brain metabolism is shifted away from the neurons and shunted towards the microglia. They theorise that this leads to lactic acid production by glycolysis in astrocytes, which subsequently participate in the activated immune response by contributing to oxidative phosphorylation and hence production of high levels of adenosine triphosphate (ATP) and forms of ROS required for degradation of the invading pathogen.

The metabolism of tryptophan is also of interest in TBM. Tryptophan is a key amino acid required for protein biosynthesis, and a precursor for various metabolites, including serotonin and melatonin (serotonin pathway) and kynurenine and quinolinic acid (kynurenine pathway) (Lesniak et al., 2013) (Figure 1.4). Pro-inflammatory cytokines such as IL-6, TNF- $\alpha$  and IFN- $\gamma$  are known to trigger the kynurenine pathway by stimulating indoleamine 2, 3-dioxygenase (IDO) (Campbell et al., 2014, O'Connor et al., 2009). Once IDO is activated, the kynurenine pathway is promoted at the expense of the serotonin pathway. Microglia and astrocytes then secrete quinolinic acid and kynurenic acid, respectively. Both products have opposite roles, while kynurenic acid acts as an antagonist of the glutamate N-methyl-D-aspartate receptor (NMDAr), quinolinic acid acts as its agonist and leads to neurotoxicity (Campbell et al., 2014). Activation of the kynurenine pathway is also known to exacerbate progression of neurodegenerative diseases and has been described in HIV-associated dementia (Majlath et al., 2013).

Recently, Van Laarhoven *et al* conducted a metabolomic study examining serum and CSF of TBM patients. Their results showed that while most metabolites demonstrated elevated concentrations in TBM patients compared to controls, concentrations of tryptophan were low in TBM and further reduced in patients who survived. They further

demonstrated upregulation of the gene *IDO1* and identified 11 trait loci that correlated with tryptophan concentrations and were prognostic of survival when combined with sex and age. The prognostic potential of these genetic correlates was demonstrated in a validation cohort. These data suggest that tryptophan metabolism may play an important role in TBM outcome, and that further investigation into this metabolic pathway is warranted (van Laarhoven et al., 2018).

While *M.tb* may directly stimulate of IDO at the site of infection (Weiner et al., 2012), vitamin D on the other hand is an essential cofactor of the tryptophan hydrolase (TPH) and promotes serotonin production and thus neurotransmission. Furthermore, Vitamin D alone or bound to the vitamin D receptor (VDR) expressed on various cell types, including astrocytes and microglia, leads to a decrease in inflammatory response (Wobke et al., 2014). Thus tryptophan metabolism could be a modality by which the vitamin D status of an individual modulates both susceptibility to *M.tb* infection and TBM pathogenesis. By restoring a non-inflammatory environment and restricting *M.tb* replication, vitamin D could promote the serotonin pathway at the expense of the kynurenine one and protect the brain from neurotoxicity.

#### **1.3.4 Host genetic factors**

A number of immune response genes encoding the pathways described are under genetic influence. For instance, mutations and polymorphisms within the genes involved in the TLR pathway have been associated with susceptibility to infection in tuberculosis. A recent study of patients with TBM in Vietnam was the first to demonstrate an association between single nucleotide polymorphisms (SNP) in the TLR9 gene region and TBM (Graustein et al., 2015). Further, a Vietnamese study found an association between TLR2 SNP T597C and the development of TBM and millary TB, suggesting that TLR2 influences the dissemination of *M.tb* (Thuong et al.,

2007). Toll Interleukin 1 receptor domain containing adaptor protein (TIRAP) mediates signals from TLR1, -2, -4 and -6 to activate macrophages and dendritic cells. Hawn *et al*/ investigated the association of the TIRAP SNP C558T with i) susceptibility to TB (odds ratio, 2.25,  $p < 0.001$ ) and ii) susceptibility to meningeal TB (OR, 3.02;  $p < 0.001$ ) vs pulmonary TB (OR, 1.55;  $p = 0.22$ ). They also demonstrated that compared to the to the 558CC genotype, the 558TT genotype was associated with decreased whole-blood interleukin-6 production, suggesting that TIRAP influences disease susceptibility by modulating the inflammatory response (Hawn *et al.*, 2006).

Another gene of interest is leukotriene A4 hydrolase (LTA4H). LTA4H influences the balance of pro- and anti- inflammatory eicosanoids and subsequent TNF- $\alpha$  regulation through either reduced inflammation due to excess lipoxins or augmented inflammation due to excess leukotriene B4. In a study of zebrafish and humans, mutations in the gene encoding LTA4H led to immunosuppressive Lipoxin A4 (LXA4) accumulation and increased susceptibility to mycobacteria (Tobin *et al.*, 2012). In this study, heterozygosity at several LTA4H SNPs was associated with protection against meningeal tuberculosis (TBM) (Tobin *et al.*, 2012). These findings supported a hypothesis that excess LTA4H activity leads to a 'hyperinflammatory state', whereas lack of LTA4H activity leads to an inadequate host response (Tobin *et al.*, 2010). In a later study by the same group the LTA4H promoter region SNP rs17525495, defining 3 genotypes—TT, CT, and CC— was identified as a likely molecular determinant of genetic susceptibility. In this study lymphoblastoid cell lines with SNP genotype CC conferred a hypoinflammatory and TT a hyperinflammatory phenotype (Tobin *et al.*, 2012). It was also demonstrated that i) genotype correlated with pre-treatment CSF leukocytosis and survival and ii) those benefiting from adjunctive dexamethasone were carriers of the 'hyperinflammatory' TT genotype (Tobin *et al.*, 2012). Several follow-up studies to further investigate the relevance of this finding have tended to reproduce an



association with susceptibility to disease, but not necessarily outcome (Yang et al., 2014a, Dunstan et al., 2015, van Laarhoven et al., 2017, Thuong et al., 2017). The two largest of these both published in 2017 investigated the association in Vietnamese (Thuong et al., 2017) and Indonesian (van Laarhoven et al., 2017) populations. The first showed that individuals with the LTA4H CC genotype had a higher risk of early death, whereas the second did not find an association between genotype and mortality. Possible explanations may include differences in linkage disequilibrium as well as the observed overall differences in patient characteristics such as mortality (40.7% in Indonesia vs 18.9% in Vietnam), frequency of culture confirmed diagnosis (55.3% vs 42.8% in Vietnam), severity of disease (BMC grade I severity in Vietnam 37% vs 11% in Indonesia) and median age of patients (41 years vs 29 years in Vietnam) (reviewed in (Fava and Schurr, 2017)). Nonetheless this remains an area of great interest and a clinical trial is under way to determine whether LTA4H genotype, defined at randomisation, influences dexamethasone's clinical efficacy when added to the first 6-8 weeks of anti-tuberculosis treatment of TBM (NCT03100786).

There are a number of other polymorphisms documented in the literature that relate to the pathogenesis of TBM. Polymorphisms in CD43 encoding a surface glycoprotein involved in *M.tb* adhesion and pro-inflammatory cytokine induction has been associated with decreased survival from TBM (Campo et al., 2015). The mannose binding protein (MBP) binds mycobacteria and acts as an opsonin *in vitro*. Although MBP plays a role in the first line of defence against *M.tb*, it may also facilitate the spread of the intracellular pathogen (Hoppe et al., 1997). A hypothesis was therefore advanced that phenotypes in which levels of MBP are low may result in protection from TBM. In a study to test this, a mutation in the MBP B allele (G54D), which leads to disruption of the collagen region of the MBP protein, was found to be associated with reduced dissemination to the brain and suggested protection against TBM (Hoal-Van

Helden et al., 1999). Polymorphism in the PKP3-SIGIRR-TMEM16J gene region encoding a negative regulator of toll-like receptor/IL-1R signaling has been associated with reduced survival in both pulmonary TB (PTB) and TBM (Horne et al., 2012). Lastly, Vitamin D Receptor gene polymorphisms with heterozygous (TC) and mutant (CC) genotypes of Taq1 VDR SNP associate with TBM (Rizvi et al., 2016), although further research is required to investigate whether this is universal across all populations (Areeshi et al., 2017).

### **1.3.5 Pathogen virulence factors and their effect on pathogenesis**

*M.tb* was formerly regarded as an organism exhibiting little genetic variation. More recent studies using genetic typing techniques to analyse *M.tb* isolates from diverse geographic populations have revealed a cladal phylogeographic distribution with variation between different lineages (Gagneux and Small, 2007, Gagneux et al., 2006). Seven lineages are now identified classified as “ancient” (lineages 1, 5, 6 and 7) or “modern” (2, 3 and 4) (Comas et al., 2013). One lineage of particular interest has been lineage 2, which is highly prevalent in East Asia (and therefore known as ‘Beijing’) and has been ascribed hypervirulence in a rabbit model (Tsenova et al., 2005) as well as some human studies demonstrating increased risk of disseminated disease (Kong et al., 2007). This has been attributed to an intact polyketide synthase (*pks 15/1*) gene and the production of a phenolic glycoprotein (PGL), which is thought to attenuate the early host immune response leading to a reduced production of pro-inflammatory cytokines (Dormans et al., 2004). A later study by Caws *et al* comparing bacterial and host genotype across two groups of Vietnamese adults with pulmonary or meningeal TB found that disease caused by the Euro-American lineage 4 was significantly more likely in pulmonary disease, however, by contrast found no association between the lineage 2 and disease phenotype (Caws et al., 2008).

Epidemiological studies have reported several differences in disease phenotype between *M.tb* lineages in terms of pathogen virulence (Guerra-Assuncao et al., 2015b, Reed et al., 2004), transmission of disease (Coscolla and Gagneux, 2014, Gagneux et al., 2006, Guerra-Assuncao et al., 2015a), progression from latent to active disease (de Jong et al., 2008) and in response to treatment (van Crevel et al., 2001, Parwati et al., 2010). *In vitro* studies have explored whether the host immune response is specific to genotype. A study in South Africa found differential mycobacterial growth and levels of early pro-inflammatory cytokines including TNF and IL-12p40 between three modern lineages (Sarkar et al., 2012). Others have found differences in human macrophage responses between lineages and have hypothesised that the lack of early pro-inflammatory response observed with modern lineages may contribute to more rapid disease progression and transmission and therefore confers survival advantage for these strains of *M.tb* (Portevin et al., 2011). In Madagascar differences between ancient and modern lineages were characterised by contrasting IFN- $\gamma$  responses (Rakotosamimanana et al., 2010). Specifically, comparison of the IFN- $\gamma$  responses with the spoligotype of the infecting clinical strains showed that modern *M.tb* strains tended to induce lower IFN-gamma responses than ancient strains in index cases and their household contacts (Rakotosamimanana et al., 2010). The aforementioned study by Caws *et al* was the first to demonstrate an interaction between pathogen and host genetic factors as a predictor of disease phenotype by showing that individuals with the C allele of the TLR02-T597C genotype were more likely to have tuberculosis caused by the Beijing genotype (OR 1.57; 95% CI 1.15 – 2.15) (Caws et al., 2008). The most recent study in this field performed whole genome sequencing of *M.tb* strains from 322 HIV-1 uninfected patients with TBM (n=106) and PTB (n=216). Unlike the previous studies (Caws et al., 2008) (Kong et al., 2007) there was no association with disease phenotype and lineage, however using a homoplasy based association analysis they

identified three *M.tb* genes associated with disease phenotype. This included Rv0218, a secretome gene encoding a protein that influences pathogen recognition and host-pathogen interaction. They hypothesise that a SNP in the region of Rv0218 would alter the appearance of the surface of *M.tb* therefore allowing it to evade recognition and the immune response, enabling dissemination to extrapulmonary sites.

### **1.3.6 Differences in the intracerebral immune response in HIV**

There is a relative paucity of data directly comparing the intracerebral immune response to TBM in HIV-1 infected and uninfected hosts. The impact of HIV-1 infection on the clinical presentation of TBM is debated, some studies report that HIV-1 infection does not alter the cerebral features of TBM (Katrak et al., 2000, Karande et al., 2005), while others suggest a higher rate of extra-meningeal and extra-pulmonary disease associated with HIV-1 infection (Thwaites et al., 2005, Karstaedt et al., 1998, Azuaje et al., 2006). These studies consistently report that HIV-1 infection associates with death (Katrak et al., 2000) and decreased efficacy of treatment against *M.tb* (Karstaedt et al., 1998). A better characterisation of the effect of HIV-1 in the pathogenesis of tuberculous meningitis is important.

A study conducted in Vietnam demonstrated a small reduction in IFN- $\gamma$  yet more significant reduction in the anti-inflammatory cytokine IL-10 in HIV-1 infected patients, the latter being 6-fold decreased in the context of HIV. This skewed the CSF IFN- $\gamma$ : IL-10 ratio towards excess IFN- $\gamma$  with subsequent analysis that was independently associated with HIV co-infection (odds ratio (95% CI), 2.47 (1.25 – 4.88),  $p=0.009$ ).

Torok *et al* (Torok et al., 2008) described a higher percentage of CSF neutrophils and suggested this may be due to prior HIV-1 infection. Indeed, this elevated percentage of

CSF neutrophils in HIV-1 infected patients may be caused by the microglial response to HIV-1 infection and HIV-1 proteins gp120, Tat (Trans-activator of transcription), Nef (Negative regulatory factor) and Vpr (Viral protein R). Atluri *et al* demonstrated that these four proteins disrupt the BBB integrity by triggering the decline of tight proteins expression on brain microvascular endothelial cells, MMP expression and apoptosis, and leading to infiltration of leukocytes (Atluri *et al.*, 2015). Moreover, it has been reported that microglia *in vitro* secrete numerous pro-inflammatory cytokines and chemokines when infected by HIV-1, including the pro-inflammatory neutrophil chemotactic factor IL-8 (Rock *et al.*, 2005). Those cells also secrete TNF, a cytokine implicated in BBB permeability and whose role in pathogenesis of *M.tb* is critical, indicating the influence of HIV-1 in the promotion of TBM and *M.tb* spread in the CNS.

Further insight into the mechanisms underlying immunopathogenesis of TBM have been observed through the study of paradoxical immune reactions. A paradoxical reaction in tuberculosis is characterised by worsening of pre-existing tuberculous lesions, or the appearance of new tuberculous lesions despite effective anti-tuberculosis treatment in patients who have demonstrated initial improvement to therapy. In HIV-1 co-infection, paradoxical worsening often occurs following the introduction of antiretroviral therapy (ART): a phenomenon known as immune reconstitution inflammatory syndrome (IRIS). Although initiation of ART during treatment of TBM is associated with a reduced six month mortality (HR 0.3, 95% CI=0.08–0.82) (Marais *et al.*, 2011) IRIS in this context carries a poor prognosis (Marais *et al.*, 2013). The role of inflammasomes, immune complexes of receptors and sensors that mediate innate immune responses and lead to inflammation, have been described in this setting (Lai *et al.*, 2015, Tan *et al.*, 2015, Tan *et al.*, 2016).

In TBM specifically, IRIS is characterised by high CSF neutrophil counts and *M.tb* positivity at presentation (Marais et al., 2013). In those who develop IRIS, CSF TNF was found to be high whereas CSF IFN $\gamma$  was low (Marais et al., 2013). Further analysis of predictive factors has demonstrated the role of neutrophil mediators, in particular S100A8/9, which two weeks after the initiation of ART was found to be significantly higher in the CSF of those who develop TBM-IRIS compared to those who do not (Marais et al., 2014). Using longitudinal microarray analysis of blood from patients with HIV and TBM, Marais *et al* have also demonstrated that neutrophil abundant transcripts were significantly more copious in those who develop IRIS from before ART initiation until the onset of TBM-IRIS (Marais et al., 2017). After initiation of ART a significantly higher number of transcripts associated with canonical and non-canonical inflammasome activation were found in patients who went on to develop IRIS than in those who did not (Marais et al., 2017). These observations together with the finding that inflammasome activation can contribute to pyroptosis (i.e. cell death triggered by proinflammatory signals and associated with inflammation\_) (Bergsbaken et al., 2009) suggest that tissue injury observed in TBM may be partly induced by inflammasome-mediated pyroptosis. It is unclear as to whether similar mechanisms exist in the absence of HIV.

HIV-1 infiltrates the CNS soon after systemic infection and around half of HIV-1 infected people develop HIV-associated neurocognitive disorders (Tozzi et al., 2005). It has been reported that IRIS may worsen established HIV related cognitive impairment (Miller et al., 2004). Little is known of the pathological mechanisms which lead to worsening cognition in the context of IRIS but some have postulated a role of the HIV-1 Trans-Activator of Transcription (Tat) protein (Johnson et al., 2013). They described the presence of Tat in the CSF of the virologically controlled ART-treated population

and proposed that Tat would contribute to neuroinflammation and IL-17 production by infiltrating Th17 cells. Given that an increase of IL-17 secretion predisposes to IRIS in HIV-1 infected individuals in the context of cryptococcal meningitis (Boulware et al., 2010), it is possible that Tat is a key player in the pathogenesis of HIV related cognitive impairment in the context of IRIS.

### **1.3.7 Macroscopic manifestations of the disease in relation to the immune response**

The characteristic feature of TBM in post-mortem studies is the presence of a thick, gelatinous inflammatory exudate in the basal cisterns and subarachnoid spaces of the brain (Figure 1.5), which may extend into the spinal canal (Dastur et al., 1995, Daniel, 1949, Shinoyama et al., 2012). The predominantly basal location has important implications; firstly, the major cerebral vessels originating from the base of the brain become encased with exudate; secondly, exudate blocks the circulation of CSF; and thirdly, it envelopes and compresses the local cranial nerves resulting in cranial nerve palsies (Dastur et al., 1995).

#### *Vasculitis*

As the exudate spreads along the cisternal extensions from the base of the brain it coats the large arteries and their small perforators, although it has a predilection for extension into the Sylvian fissures. Therefore, the middle cerebral artery (MCA) and its perforators around the floor of the 3<sup>rd</sup> ventricle are most commonly involved, resulting in ischemic damage to the sub-thalamic nuclei and lower internal capsule (Dastur et al., 1995) (Figure 1.7). Involvement of small vessels supplying the mid-brain may lead to infarction (Daniel, 1949) and exudate in the interpeduncular cistern can compromise the vessels supplying the hypothalamic region and precipitate hypothalamic symptoms

in these patients (Daniel, 1949). Vascular pathology includes infiltration of inflammation from the adventitia of arteries and veins inwards, resulting in peri- or pan-arteritis that involves segments or the full thickness of the vascular wall tissue (Lammie et al., 2009). Evolution of vascular inflammation may involve thickening of the vessel intima resulting in vessel stenosis or occlusion (Lammie et al., 2009). Both cerebral and spinal vessels may be affected by any or all of these processes and this may reflect the duration of illness (Lammie et al., 2009). Infarcts (Figure 1.5) and changes on angiography are reported in the majority of TBM patients (Rohlwink et al., 2016) and are associated with post-mortem cerebral and vascular pathology (Dastur et al., 1995).

#### *Hydrocephalus*

Extension of exudative material into the basal cisterns results in a collar of exudate around the midbrain and a blockage in CSF flow around the upper brain stem. Exudate may also collect around the cerebral aqueduct of the 3<sup>rd</sup> and 4<sup>th</sup> ventricles hindering the flow of CSF through the ventricular system (Figure 1.5). Consequently, hydrocephalus is common, occurring in 75-80% of TBM patients (Rohlwink et al., 2016, van Well et al., 2009, Thwaites et al., 2007). The pressure of expanding ventricles and the opposing pressure of brain oedema due to pathological processes can negatively impact the grey and white matter, leading to pallor and diffuse loss of myelin (Dastur et al., 1995), and the raised intracranial pressure can severely compromise cerebral blood flow.

#### *Tuberculomas and Tuberculous brain abscess*

Tuberculomas may coexist with TBM or occur independently and are thought to originate from expanding tubercles in the parenchyma (Figure 1.5). These lesions are typically granulomatous consisting of a necrotic centre surrounded by epithelioid cells (which may merge to form Langhans giant cells), lymphocytes and a border of astrocytes, and are associated with peripheral oedema and vascular proliferation (Rock



et al., 2008, Dastur et al., 1995). With time the tubercles become discretely organised and bound by a rim of connective tissue of reticulin fibres, which enhances on contrasted brain imaging. They may occur throughout the brain in the cerebrum, cerebellum and brainstem (Dastur et al., 1995, Rohlwink et al., 2016).

TB brain abscesses may follow TBM or develop independently (Schoeman et al., 2006, Kumar R, 2004), however, they occur infrequently (Kumar et al., 2002). Abscesses manifest as encapsulated collections of pus that contain tubercular bacilli in the absence of typical TB granuloma features including epithelioid and giant cells and mononuclear cell infiltrates (Kumar et al., 2002). These abscesses have been reported in the cerebrum (Schoeman et al., 2006, Kumar et al., 2002) and posterior fossa (Schoeman et al., 2006, Schoeman et al., 1998, Saini et al., 2011), occur infrequently in the brainstem (Kumar R, 2004), and may be singular or multiple (Rock et al., 2008).

### *Spinal TB*

Spinal TB may develop as a primary tuberculous lesion, from the downward extension of intracranial TBM, or secondary to vertebral TB (Dastur et al., 1995, Moghtaderi and Alavi Naini, 2003, Hernandez Pando et al., 2010). It involves the cord, meninges and nerve roots and manifests as spinal arachnoiditis, intradural (extramedullary) tuberculomas or rarely as intramedullary tuberculomas (Rohlwink et al., 2016, Srivastava and Kochar, 2003, Skendros et al., 2003, Lim et al., 2013). Spinal nerve roots may become entrapped and matted in exudate filling the subarachnoid space, and complete obliteration of the thecal space can occur in severe cases (Rohlwink et al., 2016). Exudate involvement of the lower lumbar segments is associated with difficulty in performing lumbar punctures and a high CSF protein (Dastur et al., 1995, Rohlwink et al., 2016). The microscopic appearance of spinal exudate is identical to intracranial exudate, consisting of giant cells and caseous areas, and causing vasculitis

of the spinal veins and arteries, which may lead to cord ischemia and infarction (Dastur et al., 1995, Kato et al., 1997).

### **1.3.8 Research gaps and the path forward**

Intracerebral and spinal pathology in TBM is mediated by a dysregulated inflammatory response that contributes to meningitis, tuberculoma formation, arteritis, obstruction of cerebrospinal fluid (CSF) flow, and vascular complications including stroke. The development of animal or *in vitro* cellular models could aid understanding of the underlying immune mechanisms and point the way to adjunctive anti-inflammatory and improved antibiotic therapies. In human studies high-resolution MRI or CT imaging to assess TBM disease activity could provide a more detailed clinical phenotype for TBM than is currently available. Unbiased application of 'omics' technologies (principally transcriptomics); in particular, analysis of cells and fluid arising from the site of disease is likely to be particularly valuable. Genes encoding proteins involved in metabolism, cell growth, transport, immune response, cell communication and signal transduction are particularly of interest. Longitudinal sampling of blood or CSF during observational studies or randomized controlled trials is highly informative regarding the dynamics of the disease, such as disease progression and drug response.

**Table 1.1 Currently available antibiotics for treatment of TBM**

Drug	WHO recommended daily dose	WHO recommended duration	CSF penetrance (CSF:plasma concentration)
<b>First-line drugs for treatment of drug sensitive TBM in adults</b>			
Rifampicin	10 mg/kg (range 8–12 mg/kg); max 600 mg	12 months	10–20%
Isoniazid	5 mg/kg (range 4–6 mg/kg); max 300 mg	12 months	80–90%
Pyrazinamide	25 mg/kg (range 20–30 mg/kg)	2 months	90–100%
Ethambutol	15 mg/kg (range 15–20 mg/kg)	2 months	20–30%
<b>Second-line drugs for treatment of TBM in adults</b>			
Levofloxacin	10–15 mg/kg	Throughout treatment	70–80%
Moxifloxacin	400 mg	Throughout treatment	70–80%
Amikacin	15 mg/kg; max 1g. IV or IM.	Intensive phase only	10–20%
Kanamycin	15 mg/kg; max 1g. IV or IM.	Intensive phase only	10–20%
Capreomycin	15 mg/kg; max 1g. IV or IM.	Intensive phase only	No data (probably very low)
Ethionamide or Prothionamide	15–20 mg/kg; max 1 g.	Throughout treatment	80–90%
Cycloserine	10–15 mg/kg; max 1 g	Throughout treatment	80–90%
Linezolid	600 mg	Throughout treatment	30–70%
<b>Other drugs used in treatment of multi-drug resistant TB but of uncertain benefit in TBM</b>			
Clofazimine	1 mg/kg	No recommended duration	Limited data (probably low)
p-Aminosalicylic acid	200–300 mg/kg	No recommended duration	No data (probably very low)
Bedaquiline	Not determined	New drug. Limited availability.	More recent study demonstrating equivalent concentrations to plasma
Delamanid	Not determined	New drug. Limited availability.	Animal data only, however this suggests adequate penetration to the CSF

**Table 1.2 Clinical rating scores used within surgical management of TBM**

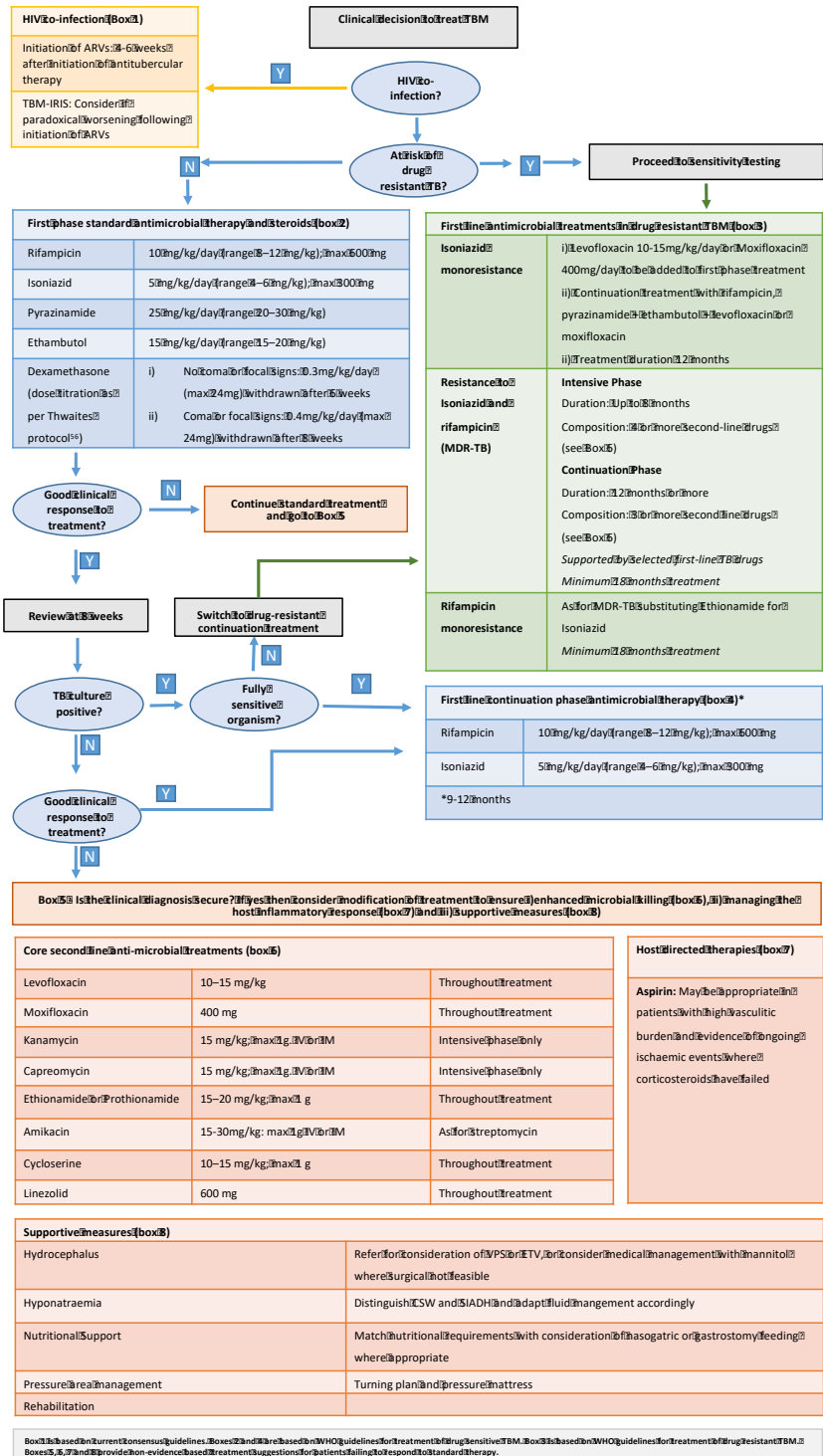
Glasgow outcome scale <sup>97</sup>		
1	Death	Severe injury or death without recovery of consciousness
2	Persistent vegetative state	Severe damage with prolonged state of unresponsiveness and a lack of higher mental functions
3	Severe disability	Severe injury with permanent need for help with daily living
4	Moderate disability	No need for assistance in everyday life, employment is possible but may require special equipment
5	Low disability	Light damage with minor neurological and psychological deficits
Modified vellore grading scale for TBM-induced hydrocephalus <sup>98</sup>		
Grade	Glasgow Coma Scale	Clinical features
I	15	Headache, vomiting +/- neck stiffness No neurological deficit
II	15	Neurological deficit present
III	9-14	Neurological deficit may or may not be present
IV	3-8	Neurological deficit may or may not be present

Figure 1.1 Computerised tomography of the head in a patient with TBM



Evidence of left MCA territory infarct within the 'tubercular zone' likely due to compromise of lenticulostriate perforator arteries

**Figure 1.2 Treatment algorithm for the management of TBM**



Abbreviations: ART; antiretroviral therapy; VPS; ventriculoperitoneal shunt; ETV; endoscopic third ventriculostomy; CSW; cerebral salt wasting; SIADH; syndrome of inappropriate antidiuretic hormone release

**Figure 1.3 Summary of the pathogenesis of TBM**

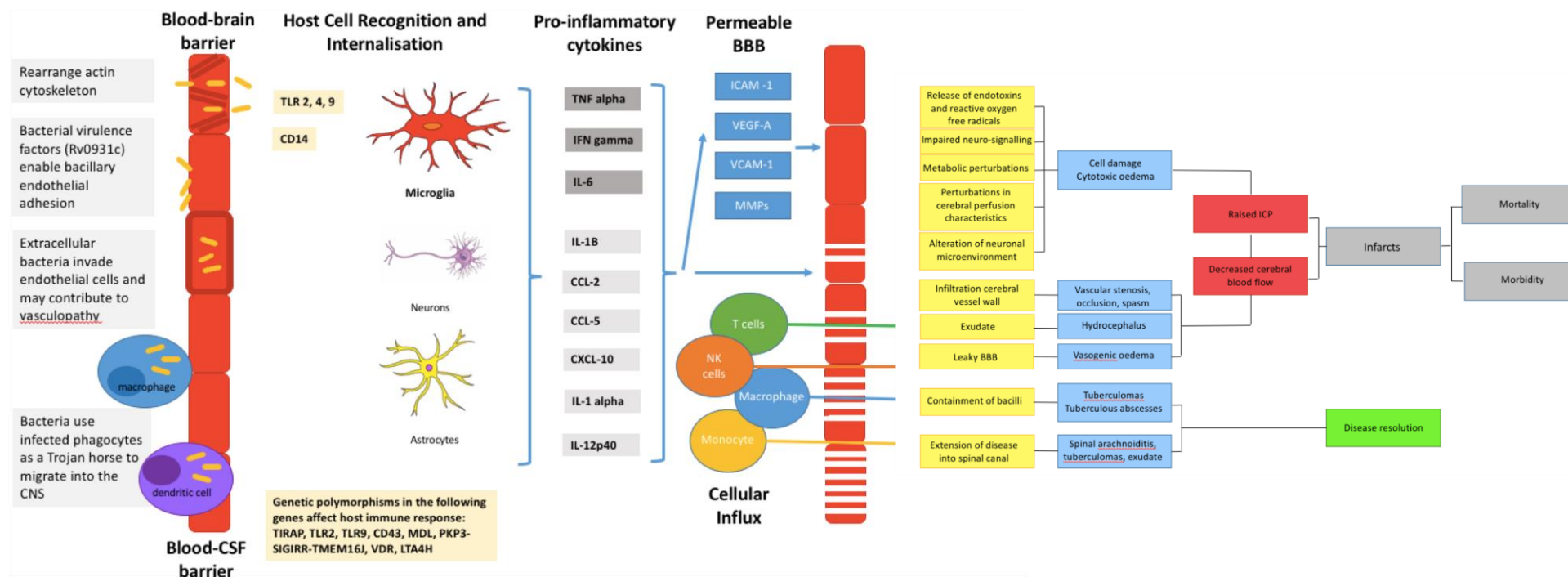
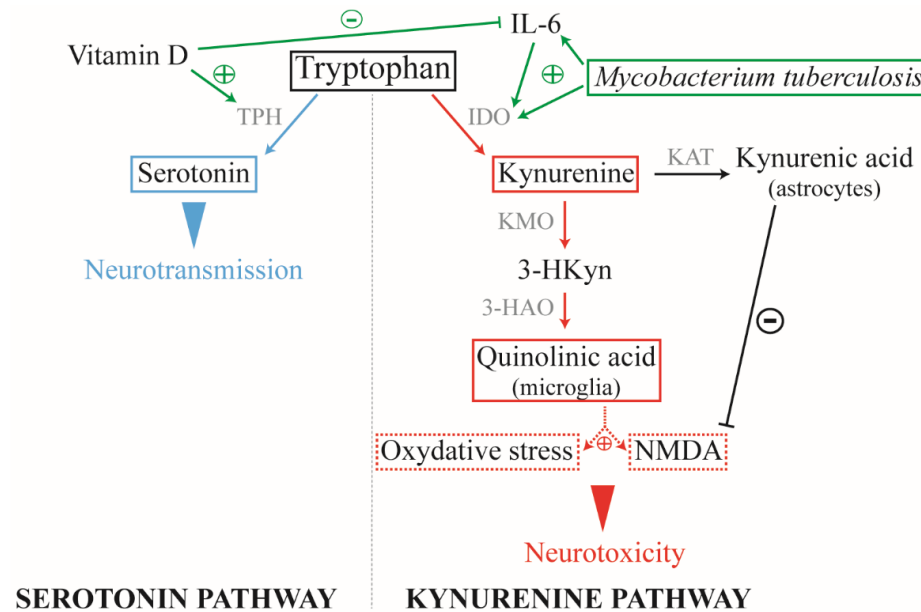


Figure 1.3 description: A: Mycobacterium tuberculosis bacilli (M.tb) disseminate from the primary site of infection in the lung to seed the brain. The bacilli are able to traverse the blood brain barrier (BBB) and blood cerebrospinal fluid barrier (BCSFB) through various virulence factors that enable the invasion of and migration through cerebral vascular endothelial cells, or are carried into the central nervous system (CNS) by infected peripheral innate immune cells. B: In the CNS antigen recognition and internalisation by microglia, neurons and astrocytes occurs, mediated by numerous host genetic factors. C: The resulting immune response stimulates the release of pro-inflammatory cytokines and chemokines and other immune mediators that contribute to the breakdown of the BBB and the influx of

innate and adaptive immune cells from the periphery. D: A prolific inflammatory response ensues. The inflammatory exudate in the basal cisterns contributes to cerebral vascular pathology and the development of hydrocephalus and raised intracranial pressure. Vasogenic oedema due to an influx of proteins through the leaky BBB, and cytotoxic oedema as a result of cellular damage contribute to the raised pressure. The overall decrease in cerebral blood flow puts the brain at risk of ischaemia, infarction and poor patient outcomes. In some cases the infection is controlled in discrete tuberculomas or abscesses, which resolve with treatment and time. Extension of the disease into the spinal canal manifests as spinal arachnoiditis, tuberculomas, or collection of exudate.

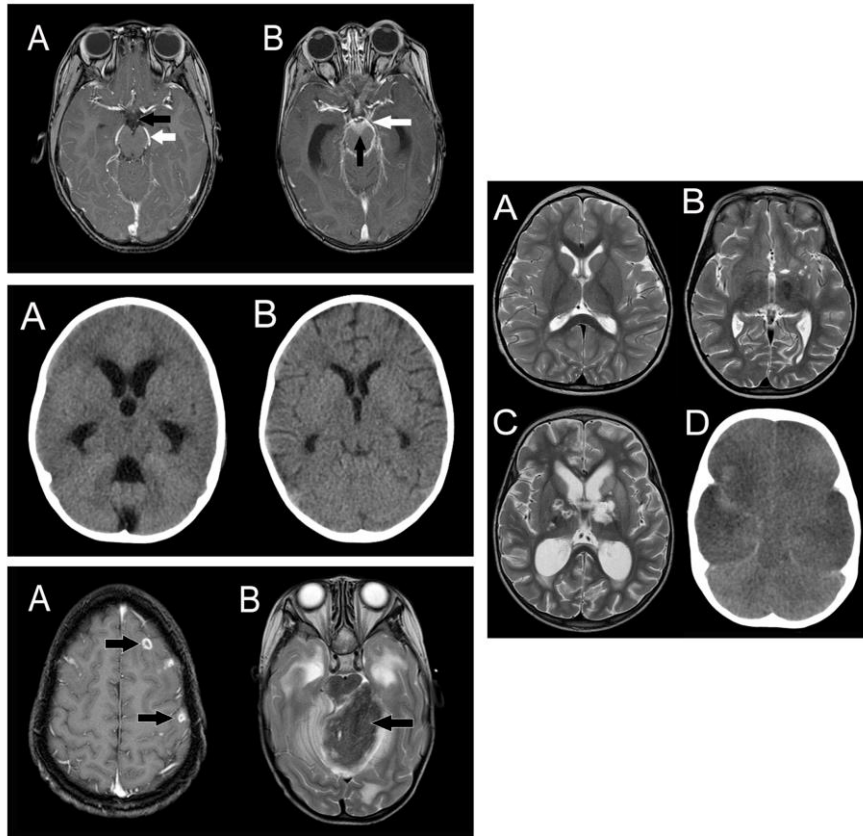


**Figure 1.4 *M.tb* and vitamin D impact on tryptophan metabolism**



*M.tb* infection activates the inflammatory response resulting in pro-inflammatory cytokine secretion. Along with *M.tb*, these cytokines can trigger the kynurenine pathway by stimulating IDO. Once the pathway is activated, astrocytes and microglia respectively produce kynurenic acid and quinolinic acid, the latter is neurotoxic. Thus, an imbalance in the synthesis of these two products may result in neurotoxicity. Vitamin D, cofactor of TPH, promotes serotonin production and neurotransmission. Alone or when bound with one of its receptors, VDR, vitamin D attenuates the inflammatory response. *M.tb*: *Mycobacterium tuberculosis*; IDO: indoleamine 2, 3-dioxygenase; TPH: tryptophan hydrolase; KAT: kynurenine aminotransferase; KMO: kynurenine-3-monooxygenase; 3-HAO: 3-hydroxyanthranilic acid dioxygenase; NMDA: *N*-methyl-d-aspartate; VDR: vitamin D receptor. Blue: neuroprotection; red: neurotoxicity; green: influential factors.

**Figure 1.5 Radiological features of TBM**



**Top left: Exudate** Contrast-enhanced T1-weighted MRI scan images: A: Normal scan showing cerebrospinal fluid in the cisterns (interpeduncular cistern in front of the midbrain, black arrow) and vessels at the base of the brain in normal cisterns (posterior cerebral artery, white arrow); B: Scan of a patient with TBM showing exudate in the basal cisterns of the brain (interpeduncular cistern, anterior to the brainstem and beneath the hypothalamus, black arrow) and vessels coursing through the exudate in the cisterns (posterior cerebral artery, white arrow).

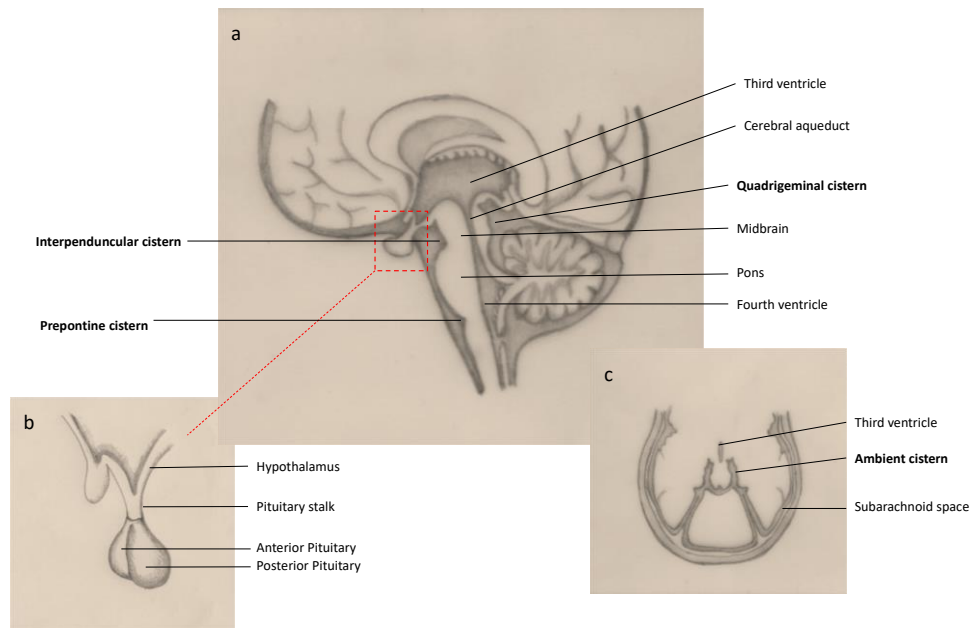
**Middle left: Hydrocephalus** A: Initial head CT scan images of a patient with TBM showing acute hydrocephalus with dilated ventricles and a compressed brain; B: Head CT scan of the same patient after 3 weeks of medical therapy showing resolution of the hydrocephalus.

**Lower left: Tuberculomas** MRI scans demonstrating different patterns and imaging characteristics of brain tuberculomas: A: Contrast-enhanced T1-weighted MRI scan showing multiple ring-enhancing small tuberculomas (arrowed); B: T2-weighted MRI scan showing a large tuberculoma in the cerebellum compressing the brainstem, surrounding oedema, and hydrocephalus from obstruction of the cerebral aqueduct.

**Lower Right: Infarcts** Patterns of infarction in TBM, A: T2-weighted MRI scan showing normal ventricular size and no infarcts; B: T2-weighted axial MRI scan of a patient with TBM showing discrete unilateral small perforator vessel infarcts; C: T2-weighted MRI scan showing more extensive infarcts in the thalami and basal ganglia; D: Head CT scan of a patient with TBM showing global infarction with hypodense hemispheres bilaterally and a swollen brain.

1 **Figure 1.6 Basal cisterns and pituitary anatomy**

2



3

4 **A:** Basal cisterns affected in TBM are represented here in a sagittal view of the brain.  
5 Note the quadrigeminal cistern which extends laterally to become a thin sheet like  
6 cistern surrounding the midbrain and posterior thalamus, named the ambient cistern  
7 shown in **C**. **B:** Anatomy of the pituitary gland and surrounding structures.

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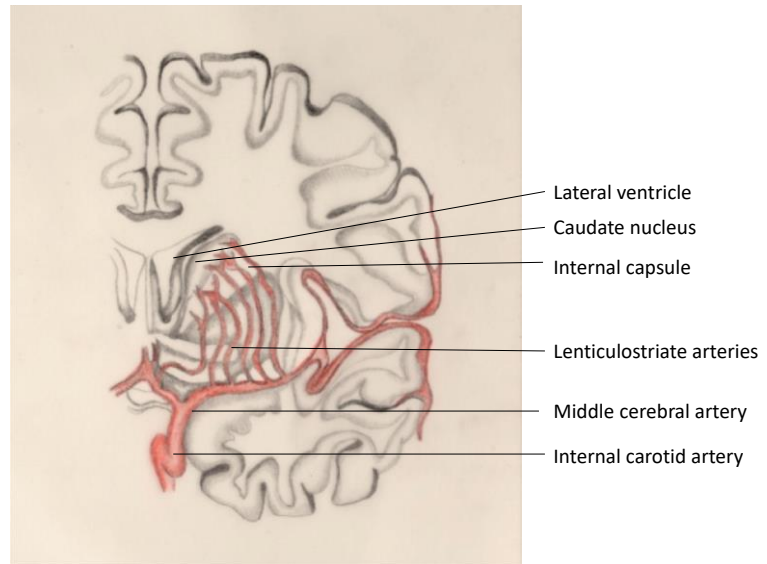
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21 **Figure 1.7 Lenticulostriate arteries**

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25 Lenticulostriate arteries branching from the M1 segment of the middle cerebral artery  
26 supply the basal ganglia and internal capsule.

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40 **Chapter 2. Study protocol for A phase 2A trial of the**  
41 **safety and tolerability of increased dose rifampicin**  
42 **and adjunctive linezolid, with or without aspirin, for**  
43 **HIV-associated tuberculous meningitis (The LASER-**  
44 **TBM trial)**

45

46 **2.1 Background**

47 In Africa, HIV-1 associated tuberculous meningitis (TBM) has a 2-month mortality  
48 approaching 50% (Marais et al., 2011). Although early antiretroviral therapy (ART) is of  
49 proven benefit in other forms of tuberculosis (TB) (Blanc et al., 2011) this has not been  
50 demonstrated for TBM (Torok et al., 2011b), a finding potentially contributed to by  
51 exacerbated immunopathology in the confined space of the central nervous system  
52 (CNS) (Marais et al., 2013). Furthermore, only a few clinical trials have addressed the  
53 recognized poor penetration of several antibiotics into cerebrospinal fluid (CSF) and  
54 adjunctive corticosteroids have not shown unequivocal benefit for HIV-1 co-infected  
55 patients in clinical trials (Thwaites et al., 2004). The recommended management of  
56 patients with TBM has remained unchanged for decades. The need to develop an  
57 effective drug treatment regimen combining agents to ensure effective bacterial killing,  
58 as well as therapies to control the host immune response is urgent.

59

60 *Linezolid*

61 Linezolid (LZD) is currently used as an effective rescue therapy in extensively drug  
62 resistant TB (Sotgiu et al., 2012, Schechter et al., 2010, Migliori et al., 2009, Anger et

63 al., 2010, Lee et al., 2012b, Singla et al., 2012). Its use is also established for the  
64 treatment of gram positive infections including pyogenic brain abscesses where  
65 patients receive 1200mg for four weeks (Nau et al., 2010). LZD is an attractive agent  
66 for the treatment of TBM due to its potent efficacy against *Mycobacterium tuberculosis*  
67 (*M.tb*) as well as its excellent CNS penetration (Nau et al., 2010). Two published  
68 studies have investigated its potential role in TBM; the first, an observational study  
69 demonstrated favorable clinical outcomes and a non-significant difference in adverse  
70 events in children with drug sensitive TBM treated with LZD compared to control (Li et  
71 al., 2016); the second, a retrospective cohort study of 33 adults with TBM the addition  
72 of LZD to a standard regimen led to more rapid improvement in CSF parameters,  
73 recovery of consciousness and reduction of fever (Sun et al., 2014).

74

75 LZD toxicity has however limited its use. The most common adverse events (AE)  
76 associated with LZD use in TB treatment are haematological toxicity (mainly dose-  
77 related) and peripheral neuropathy (mainly duration-dependent) (Zhang et al., 2015).  
78 These are usually mild and are reversible with dose reduction or treatment interruption  
79 if identified early. In a systematic review, AE related to LZD use at doses > 600 mg/day  
80 occurred at a median of 252 days (IQR 120 – 540) of LZD exposure (Ahuja et al.). In  
81 the *NiX-TB* trial where LZD was given at a total dose of 1200mg per day for 6 months,  
82 peripheral neuropathy occurred in 81% of cases with the majority of these occurring  
83 after 3 months of treatment (Conradie et al., 2020). Median time to return to a normal  
84 or mild neuropathy score was 3 months. Myelosuppression was the second most  
85 common AE occurring in 48% of cases. Although these AE led to frequent treatment  
86 interruption (66% of cases had treatment interruption) all 109 participants completed 26  
87 weeks of treatment. In the context of TBM, where morbidity and mortality is high, the  
88 risk-benefit of this potent antituberculous agent with good CNS penetration requires  
89 further evaluation in the context of a phase II safety trial.

90

91 *Aspirin*

92 Aspirin (ASA) is a safe, widely available and inexpensive drug with effects on the  
93 pathogenic processes recognised as integral to the pathogenesis of TBM and its  
94 complications (Bousser, 2009, Hovens et al., 2008). In a placebo-controlled trial of ASA  
95 in 118 adult patients with TBM in India, 150mg daily ASA was associated with a  
96 significantly lower 3-month mortality and a lower incidence of stroke albeit not  
97 significant (Misra et al., 2010). Following this a similar study in South Africa  
98 randomized 146 children with TBM to receive low dose (75mg/24hours) (n=47) or high  
99 dose (1000mg/24 hours). In this trial there was no significant effect of ASA on mortality  
100 however there was a significant reduction on incidence of new hemiplegia in those  
101 receiving high dose ASA (Schoeman et al., 2011). In a recent study in Vietnam, HIV-1  
102 uninfected individuals with TBM received ASA in addition to standard care. Patients  
103 were randomized to receive either placebo, ASA 81mg OD or ASA 1000mg OD for 56  
104 days. The pre-specified sub-analysis of results demonstrated a potential reduction in  
105 new infarcts and deaths by day 60 in patients with microbiologically confirmed TBM  
106 receiving 1000mg OD of ASA (Mai et al., 2018). Its safety for use in HIV-1 infected  
107 individuals with TBM, particularly when used in combination with an intensified  
108 antituberculous regimen, has yet to be investigated.

109

110 *High dose rifampicin*

111 Rifampicin (RIF), one of the four first line treatments for TBM demonstrates poor  
112 penetration of the blood brain barrier (BBB) with total concentrations in CSF only 10-  
113 20% of that reached in plasma (Donald, 2010). *In vitro*, animal and early bactericidal  
114 activity studies suggest that the standard 600mg once daily dose is at the lower end of  
115 the dose response curve (van Ingen et al., 2011). This has prompted a series of  
116 studies in both pulmonary and extrapulmonary tuberculosis investigating the safety and

117 efficacy treatment regimens containing higher doses of RIF (Heemskerk et al., 2016,  
118 van Toorn et al., 2014, Ruslami et al., 2013b, Boeree et al., 2015, Boeree et al., 2017,  
119 Aarnoutse et al., 2017, de Steenwinkel et al., 2013, Steingart et al., 2011). None of  
120 these studies have detected a significant safety signal thereby supporting the safety of  
121 RIF up to doses of 35mg/kg. Similarly, they provide evidence to suggest superior  
122 efficacy when used at a dose of 35 mg/kg compared to 20 mg/kg (Boeree et al., 2015,  
123 Boeree et al., 2017, de Steenwinkel et al., 2013, Steingart et al., 2011).

124

125 In TBM, the use of higher RIF doses is appealing given its incomplete penetration into  
126 the central nervous system. In 2013 an open-labelled randomized phase 2 study in 60  
127 Indonesian adults with TBM showed a 50% reduction in mortality with higher dose  
128 intravenous RIF (13 mg/kg, which equates to an approximate oral dose of 20mg/kg)  
129 compared with standard dose oral therapy (Ruslami et al., 2013a). This intensified  
130 treatment did not result in increased toxicity and was associated with a substantially  
131 lower 6-month mortality. A subsequent large randomized placebo-controlled trial in  
132 Vietnam evaluated a combined regimen of oral RIF 15 mg/kg plus levofloxacin. Unlike  
133 the previous trial using intravenous therapy (at higher equivalent oral doses of 20  
134 mg/kg) there was no effect of mortality (Heemskerk et al., 2016). These results, plus  
135 evidence from pre-clinical studies and pulmonary TB, provide adequate justification to  
136 systematically assess the effect on outcomes in TBM with RIF doses > 20 mg/kg.

137

138 The proposed trial combines three drugs for which there is sufficient evidence to  
139 suggest adequate safety profiles and potential benefit in a condition in which there is  
140 high mortality and inadequate treatment. Their safety in combination and in the context  
141 of HIV-1 co-infection requires careful evaluation in a controlled Phase II trial, before  
142 this regimen can be tested in the context of a phase III clinical trial.

143



144 **Our primary hypothesis is that increased dose RIF plus LZD and ASA can be**  
145 **safely added to standard therapy for HIV-1-associated TBM.**

146

## 147 **2.2 Methods and Study Design**

148 Study aims

149

150 **The primary aim** is to investigate the safety of enhanced antimicrobial therapy  
151 including increased dose RIF and LZD with or without adjunctive ASA added to  
152 standard therapy for TBM in HIV-1 infected adults.

153

154 The **secondary aims** are:

- 155 a. To determine CSF *M.tb* culture positivity and Gene Xpert® Ultra positivity at  
156 baseline and at 3 and 28 days post treatment by allocation.
- 157 b. To evaluate the effect of ASA and enhanced TB treatment on the incidence of  
158 Immune Reconstitution Inflammatory Syndrome (IRIS) in participants starting  
159 ART
- 160 c. To evaluate the effect of high dose RIF and LZD with and without ASA on CNS  
161 imaging (CT, MRI and MR Spectroscopy) in conjunction with clinical,  
162 immunological and transcriptional profiling.
- 163 d. To determine i) whether host genotype, including leukotriene A4 hydrolase  
164 (LTA4H) genotype, influences therapeutic effect of ASA in HIV-TBM and ii) the  
165 pharmacogenetic influence on RIF and LZD exposures and toxicity.

166

167 Three sub studies will recruit all consenting participants with the following aims:

168

169 **Sub study 1 (Pharmacokinetic-Pharmacodynamic):**

- 170 1. To describe the plasma and CSF PK of LZD and high dose RIF.
- 171 2. To evaluate the relationship between drug exposures, toxicity and efficacy.
- 172 3. To compare exposures between intravenous and oral RIF administration.
- 173 4. To investigate the impact of high dose RIF on LZD availability.

174

175 **Sub study 2 (Pathogenesis):**

- 176 1. To evaluate the effect of high dose RIF and LZD, with and without ASA on the
- 177 transcriptional signature derived from whole blood and CSF RNA sequencing, as well
- 178 as the metabolomic and proteomic profiles, in TBM.

179

180 **Sub study 3 (Cognitive and functional outcomes):**

- 181 1. To describe the frequency and characteristics of cognitive impairment following HIV-
- 182 associated TBM
- 183 2. To compare cognitive outcomes with: i) presence and location of structural
- 184 abnormalities on magnetic resonance imaging, ii) radiological markers of metabolic
- 185 dysfunction on magnetic resonance spectroscopy, iii) *in vivo* markers of
- 186 neurodegeneration and brain injury within the central nervous system
- 187 3. To quantify the functional impairment (including effect on quality of life) of TBM
- 188 associated cognitive impairment

189

190 A strategic aim of LASER-TBM is to serve as a planning study to generate data which  
191 will inform a planned phase 3 RCT of intensified treatment in TBM (INTENSE-TBM).

192 Data from LASER in particular i) pharmacokinetic data on exposure in intravenous  
193 versus high oral dose rifampicin and ii) safety data to exclude any signal which would  
194 preclude commencement of INTENSE-TBM will in part dictate the resulting sample  
195 size. If no safety signal is detected, and PK endpoints meet with adequate power then

196 LASER-TBM recruitment may cease prior to the maximum sample size of 100  
197 participants to allow timely commencement of INTENSE-TBM.

198

### 199 **Study design, recruitment and duration**

200 LASER-TBM is a parallel group, randomized, multi-arm Phase 2A trial evaluating the  
201 safety of increased dose RIF plus LZD, with or without ASA, for the treatment of HIV-  
202 infected adults with TBM (Figure 2.1). HIV-1 infected adults with newly-diagnosed  
203 TBM (up to n = 100) will be recruited from five public-sector hospitals across South  
204 Africa. Participants will be randomised in a 1.4:1:1 ratio across two experimental (n =  
205 30 each) and one standard of care (n = 40) arms (Figure 2.1). Relatively more  
206 participants will be randomised to the control group to account for the higher mortality  
207 anticipated in the standard of care arm. Treatment will be provided in all arms for 56  
208 days, after which participants will be referred back to public sector facilities to complete  
209 standard therapy for HIV-associated TBM. All participants will receive antitubercular  
210 chemotherapy as well as corticosteroids as per standard practice. Participants  
211 allocated to experimental arms 2 and 3 will receive additional RIF (total oral dose 35  
212 mg/kg/day) for 56 days plus oral LZD 1200mg daily for the first 28 days, reduced to  
213 600 mg daily for the next 28 days. Those randomized to experimental arm 3 will also  
214 receive oral aspirin 1000 mg daily for 56 days. A second randomization will take place  
215 before receipt of study drug for participants in the experimental arms (n = 60) to receive  
216 either oral (35 mg/kg) or intravenous (20 mg/kg) RIF. This will be continued for 3 days,  
217 after which all participants will receive oral RIF for the remainder of the intervention  
218 period (53 days). There are six scheduled study visits, which will occur at study sites  
219 or affiliated stepdown facilities. Visits will involve interviews, clinical examination,  
220 phlebotomy, lumbar puncture and brain imaging at the timepoints shown in Figure 2.1.  
221 In those who consent, intensive PK sampling will take place at day 3. Trial participation  
222 will be for 180 days post-randomization: primary safety endpoints and secondary

223 efficacy endpoints will be evaluated at day 56; additional secondary endpoints will be  
224 evaluated at day 180 through record review.

225

226 Endpoints

227 **The primary endpoint of the study is:** The incidence of solicited treatment-related AE  
228 (see Table 2.1) and death at 56 days associated with increased dose RIF plus LZD  
229 with or without adjunctive ASA, when administered alongside standard antitubercular  
230 therapy.

231

232 **Secondary study endpoints are:**

- 233 • Death and severe disability (Modified Rankin Scale Grade 5) at 56 days (Box 2.3).
- 234 • Death at 56 and 180 days.
- 235 • Disability at 56 and 180 days, stratified by baseline MRC grade.
  
- 236 • Grade 3 or 4 AE.
  
- 237 • Permanent discontinuation of experimental drugs.
  
- 238 • Severity and frequency of haematologic and neurologic AE related to LZD use.
  
- 239 • Severity and frequency of major bleeding (gastrointestinal and intracerebral) related  
240 to ASA use.
  
- 241 • *M. tb* culture status and time to positivity in automated liquid culture (MGIT) and  
242 Gene Xpert® Ultra cycle threshold (C<sub>t</sub>) values at days 28 and 56.
  
- 243 • The occurrence of TBM-IRIS assessed by the modified International Network for  
244 the Study of HIV-associated IRIS (INSHI) criteria (Meintjes et al., 2008).
  
- 245 • MRI and CT changes at day 56.

246

247 Study participants for LASER TBM must be adults (aged 18 or over), with proven HIV-1  
248 seropositivity, and a diagnosis of TBM meeting criteria for 'possible', 'probable' or  
249 'definite' as per the published consensus definition (Marais et al., 2010).

250

251 Potential participants will be excluded if they meet any of the exclusion criteria outlined  
252 in Box 2.1. The published uniform case definition criteria used to define 'definite',  
253 'probable' and 'possible' TBM in described in Box 2.2.

254

255

### 256 **2.3 Recruitment, randomisation, retention and withdrawal**

257

258 Recruitment will be from inpatients at the participating hospital sites in South Africa  
259 (Groote Schuur Hospital, Mitchells Plain Hospital, New Somerset Hospital and  
260 Livingstone Hospital). Suitable patients will be identified by attending ward doctors and  
261 co-investigators at each site and referred to the study staff for screening.

262

263 Participant identification numbers (PID), assigned at the screening visit, will be used  
264 throughout the study. After signing the informed consent document; eligible participants  
265 will be randomized to one of the treatment arms using an electronic randomization tool.  
266 The randomisation list will be generated and updated by the trial pharmacist who will  
267 have no direct contact with trial participants or involvement with the assessment for  
268 eligibility in the trial. The second randomisation to IV or oral RIF will take place  
269 immediately, prior to receipt of study drug, for all participants allocated to experimental  
270 arms. The trial is open-label, and regimens will not be masked.

271 All trial procedures will take place in hospital during the admission period. The  
272 decision to discharge trial participants will be made by clinical, and not trial, staff. Site-  
273 specific standard operation procedures (SOP) will be developed for trial follow up visits  
274 following discharge or referral to a stepdown facility.

275

276 A participant will be withdrawn from the study if:

277

- 278 • The initial *M.tb* strain is found to be RIF-resistant on confirmatory testing;
- 279 • HIV-1 result is found to be negative on confirmatory testing;
- 280 • An alternative diagnosis is established within 5 days of antitubercular treatment  
281 initiation which leads the treating physician to discontinue antitubercular therapy;
- 282 • Withdrawal of informed consent.

283

284 Participants who withdraw consent prior to completion of the study will not undergo any  
285 further study procedures or data collection. In such cases, consent for the use of data  
286 collected prior to withdrawal of consent will be sought from the withdrawing  
287 participants. There will be no replacement for participants withdrawn from the trial.

288

## 289 **2.4 Interventions**

290 Study drug regimens

291 Participants enrolled to the study will receive study drug regimens as outlined in Table  
292 2.2. Dosing of the RHZE fixed dose combination (FDC) will be according to WHO  
293 weight bands. Study drugs will be given orally, either as tablets/capsules or crushed,  
294 depending on the clinical circumstances. Half of participants in experimental arms will  
295 be randomised to receive IV RIF for the first 3 days of therapy and switched to the oral  
296 formulation thereafter. Study drugs will be prescribed by trial doctors, packaged and  
297 distributed by trial pharmacists.

298

299 *Oral RIF Dosing Bands*

300 Simulations were performed to determine the dose of RIF required to achieve the most  
301 equitable drug exposures across the weight range, 30 to 100 kg. Demographic data of  
302 a reference cohort of TB patients (n = 1225), with or without HIV-1 coinfection,  
303 recruited in clinical trials conducted in West Africa and South Africa were used for the  
304 simulations (Chigutsa et al., Lawn et al., Diacon et al., McIlleron et al.). An additional  
305 12 250 virtual patients were generated using the weight and height distributions of the  
306 1225 patients to increase the number of patients with a weight close to the boundaries  
307 of the weight range. Parameter estimates of the population PK model for RIF were  
308 used to simulate (100 replicates) RIF exposures (Chirehwa et al.). Four dosing  
309 scenarios were evaluated using the weight-band based dosing with 4-drug FDC tablets  
310 and extra RIF tablets with each tablet containing 150 mg or 600 mg RIF. The FDC  
311 tablets were assumed to have 20% reduced bioavailability based on data from a  
312 clinical trial where the same formulation was used (Court et al.).

313

#### 314 *Intravenous RIF*

315 Participants allocated to experimental arms will be randomised (1:1) to receive either  
316 oral RIF 35 mg/kg or IV RIF 20 mg/kg for the first 3 days of therapy (in addition to HZE  
317 and LZD with or without ASA, according to the experimental arm). Those randomised  
318 to IV RIF will receive the full RIF dose intravenously, plus additional antitubercular  
319 drugs as individual tablets (at standard doses). IV RIF will be administered as an  
320 infusion as per the package insert and according to a detailed SOP.

321

#### 322 Concomitant medications

##### 323 *Corticosteroids*

324 All participants will receive corticosteroids for the first 8 weeks of TBM treatment as  
325 used in a randomized controlled trial demonstrating mortality benefit in patients TBM  
326 (Thwaites et al., 2004).

327

328 *Antiretroviral Therapy (ART)*

329 ART will be commenced by treating clinicians after 4-6 weeks of antitubercular therapy  
330 in all participants based on the single randomized controlled trial of ART timing in TBM,  
331 which showed no benefit for earlier ART (Torok et al., 2011a). If available, a  
332 dolutegravir-based regimen will be used in accordance with international (Gunthard et  
333 al.) and local guidelines (Meintjes G).

334

335 *Gastric protection*

336 Participants can be prescribed omeprazole 40mg daily. A higher starting dose of 40mg  
337 OD was selected to account for the interaction between proton pump inhibitors (PPI)  
338 with rifampicin via CYP2C19 and CYP3A4 leading to reduction in levels of the PPI. In  
339 participants with persistent symptoms the dose will be titrated to 80mg daily and  
340 gastroscopy considered. Although the study initially planned to use ranitidine for this  
341 indication, this was withdrawn as a concomitant medication due to concerns over a  
342 potential contamination with the probable carcinogen N-nitrosodimethylamine  
343 (NDMA)(Mahase, 2019), making the medication unavailable for use in South Africa.

344

345 *Pyridoxine*

346 Participants will receive pyridoxine supplementation as per the South African  
347 guidelines for prevention of anti-tuberculosis drug-related peripheral neuropathy(South  
348 Africa Department of Health, 2014).

349

350 *Disallowed medications*

351 The medicines listed in Box 2.4 have been shown to interact with study drugs and are  
352 therefore contraindicated for concomitant use during the study

353



## 354 **2.5 Study procedures, schedule and clinical assessments**

355

356 Participants will undergo 6 scheduled study visits after screening, plus ascertainment  
357 of vital status and disability assessment at 6 months. Table 2.3 describes planned  
358 investigations at each study visit

359

### 360 *Clinical assessment*

361 Clinical assessment will include clinical history, conscious level by GCS, presence of  
362 new or ongoing focal neurological deficit, all adverse events, new medications started  
363 and adherence to drugs. The neurological examination at D180 is extended to assess  
364 for such as language, visuospatial deficit, visual agnosia and praxis: focal cognitive  
365 deficits which may be present in people with TBM.

366

### 367 *Modified Brief Peripheral Neuropathy Score*

368 This purely clinical early screening tool was adapted from the subjective peripheral  
369 neuropathy score (SPNS) validated for the assessment of HIV associated distal  
370 sensory polyneuropathy (DSP) (McArthur, 1998) and used previously in trials to assess  
371 LZD toxicity.

372

### 373 *Modified Total Neuropathy Score*

374 This screening tool, initially developed for the assessment of chemotherapy induced  
375 neuropathy (Cornblath et al., 1999), has been modified for use in the HIV research  
376 setting where it has shown acceptable sensitivity and specificity (85% and 80%  
377 respectively) (Evans et al., 2008). Prior studies have used a simplified 16-point  
378 (Robinson-Papp et al., 2009) or 20-point clinical scoring system (Maritz et al., 2010) as  
379 markers of HIV-associated DSP severity.

380

381 *Insomnia Severity Index*

382 All participants will complete the Insomnia Severity Index (ISI) at the Day 28 and 56  
383 visits. The ISI is a brief screening measure of insomnia which has been validated for  
384 use in in insomnia research (Bastien et al., 2001). Dolutegravir has been associated  
385 with neurotoxicity presenting with neuropsychiatric symptoms such as insomnia  
386 (Hoffmann et al., 2017), and this will be assessed as part of the PK/PD assessment.

387

388 *Measures of cognitive function*

389 These measures where possible will be done in the participants preferred language.

390 *i) Montreal Cognitive Assessment*

391 The Montreal Cognitive Assessment (MoCA) assesses six broad domains of  
392 ability and cognitive function(Nasreddine et al., 2005) and has been used to  
393 screen for cognitive impairment in previous studies within South Africa (Robbins  
394 et al., 2013). This will be carried out at day 56 and day 180.

395 *ii) Cognitive Assessment Tool-Rapid Version (CAT-Rapid)*. This instrument  
396 includes four questions about subjective cognitive complaints, as well as tasks  
397 assessing learning and memory and cognitive sequencing. CAT-rapid was  
398 developed in South Africa (Joska et al., 2016), in response to the need to  
399 develop a brief screening tool that includes functional symptom questions and a  
400 measure of executive function. The CAT-rapid incorporates aspects of the  
401 International HIV Dementia Scale and includes four symptom questions, as well  
402 as tasks assessing learning and memory and cognitive sequencing.

403 *iii) Brief neuropsychological battery*

404 Neuropsychological testing will be carried out by a trained neuropsychometric  
405 tester and clinical research worker at day 180 and will include a cognitive test  
406 battery and assessment of contributing mental health symptoms. The cognitive

407 battery comprised 12 standardized tests, each of which assessed performance  
408 in one of six cognitive domains commonly affected by TBM. The domains, tests,  
409 and outcome variables were: (1) Executive Functioning: Color Trails Test 2  
410 (CTT2) - completion time; Wisconsin Card Sorting Test (WCST) - perseverative  
411 errors; (2) Learning and Memory: Hopkins Verbal Learning Test-Revised  
412 (HVLTR) - total learning total and delayed recall total; Brief Visuospatial  
413 Memory Test-Revised (BVMTR) - total learning total and delayed recall total;  
414 (3) Generativity/fluency: category fluency - total number of animals / total  
415 number of fruits and vegetables named in 1 minute; (4) Attention/Working  
416 Memory: Wechsler Adult Intelligence Scale-Third Edition (WAIS-III) Digit Span -  
417 total score; (5) Processing Speed: CTT1 - completion time; Wechsler Adult  
418 Intelligence Scale-Third Edition (WAIS-III) Digit Symbol Coding - total score;  
419 WAIS-III Symbol Search- total score; (6) Motor Function: Grooved Pegboard  
420 Test (GPT) dominant (DH) and nondominant hand (NDH) - completion time;  
421 Finger Tapping Test (DH and NDH) - completion time. Tests were administered  
422 in either English or Xhosa depending on the participant's preference. Mental  
423 Health measures were the Centre for Epidemiological Studies-Depression  
424 (CES-D), State Trait Anxiety Inventory-trait (STAI-trait), Alcohol Use Disorders  
425 Identification Test (AUDIT) and Drug Use Disorders Identification Test (DUDIT).

426

427 *Measures of functional status*

428

429 *i) Modified Rankin Score*

430 The MRS a commonly used clinical outcome measure for patients suffering  
431 from stroke (Rankin, 1957), has demonstrated good inter-rater reliability (van  
432 Swieten et al., 1988) and is the most commonly used outcome measure to  
433 assess neurological disability in TBM (Saver et al., 2010).

434 *ii) Modified Patients Assessment of Own Functioning Inventory (PAOFI)*  
435 *This patient reported outcome measure* is designed to evaluate a patient's  
436 sense of his or her functional capacity in everyday activities concerning  
437 memory, language and communication, use of hands, sensor perception, higher  
438 level cognitive and intellectual functions, and work/recreation(Chelune).  
439 *iii) Lawton-SA* is designed to assess independent living skills, considered more  
440 complex than basic activities of daily living. This was developed in 1969(Lawton  
441 and Brody, 1969), but since modified for use in the South African context(Joska  
442 et al., 2010).

443

444

#### 445 *Blood*

446 Testing will be done at specified timepoints (as per table 2.3), and may be repeated to  
447 follow-up on abnormal results, for example after occurrence of an AE. Samples for  
448 haematology and biochemistry and HIV testing will be processed in National Health  
449 Laboratory Service (NHLS) laboratory according to local SOPs. Samples for non-  
450 clinical assays (immune markers, RNA, metabolomics, proteomics) and future use will  
451 be collected and transported to the Institute of Infectious Disease and Molecular  
452 Medicine at the University of Cape Town (UCT) for processing and storage. PK  
453 samples will be centrifuged *within an hour* of being taken at 1500 x g at room  
454 temperature for 10 minutes, the plasma aliquoted into cryovial tubes, stored at -80°C  
455 and transported to UCT Clinical Pharmacology laboratory for storage and processing.

456

#### 457 *Urine*

458 Bedside pregnancy testing will be done on urine at screening. Urine will be sent to  
459 chemistry laboratories for osmolality and electrolyte testing in the context of

460 hyponatraemia at the discretion of the investigator. Further urine will be collected for  
461 biobank storage.

462

#### 463 *Cerebrospinal fluid analysis*

464 CSF will be obtained via lumbar puncture at Days 3 and 28, in accordance with a  
465 detailed SOP. CSF collected for diagnosis in routine care (baseline) will be retrieved  
466 where possible. Routine microbiology, cell count, and biochemistry will be done in  
467 NHLS laboratory according to local SOPs. Samples for non-clinical testing (RNA  
468 sequencing, metabolomic and proteomic analysis and immunological assays) will be  
469 collected and transported to the IDM at UCT for processing and storage. CSF to  
470 determine drug concentrations for the PK study will be frozen at -80 degrees  
471 immediately following collection and transported to UCT Clinical Pharmacology  
472 laboratory for storage and processing.

473

#### 474 *Magnetic Resonance Imaging (MRI)*

475 MRI scans will be performed in all participants who can tolerate or access the  
476 investigation at baseline and day 56. A 3-Tesla (3T) MRI scanner located at Groote  
477 Schuur Hospital will be used for all imaging in the Cape Town area, whilst a 1.5T  
478 scanner located at Livingstone Hospital will be used for participants recruited in the  
479 Port Elizabeth Area. Gadolinium enhanced imaging will be performed on participants  
480 with eGFR > 30mL/min/1.73m<sup>2</sup>.

481

482 Image sequences will include the following:

- 483 • T1 weighted sequences with or without gadolinium
- 484 • T2 weighted sequences
- 485 • Diffusion weighted images (DWI)

- 486 • Susceptibility weighted images (SWI)  
487 • T2 Fluid-attenuated inversion recovery (FLAIR)  
488 • Point resolved spectroscopy (PRESS/MEGA-PRESS) to estimate brain metabolite  
489 changes

490

#### 491 *Computed Tomography (CT)*

492 If participants are unable to tolerate or access MRI, CT will be used as an alternative  
493 imaging method at the same time points as stated for MRI. Participants with eGFR >  
494 30 mL/min/1.73m<sup>2</sup> will have contrast enhanced imaging. Pre- and post- contrast  
495 sequences will be available for analysis. A standardised reporting form including  
496 positive and negative radiological findings will be used.

497

498

## 499 **2.6 Statistical considerations**

500 Sample size

501 Arm 1 - control - (standard-of-care): 40

502 Arm 2 - experimental - (standard-of care + high dose rifampicin + linezolid): 30

503 Arm 3 - experimental - (standard-of care + high dose rifampicin + linezolid + aspirin):

504 30

505

506 Total participants required for primary safety analysis = 100

507

#### 508 *Sample size justification*

509 This Phase IIA trial will focus on evaluating adverse events in the experimental arm

510 relative to the standard of care arm. Solicited treatment related AE (Table 2.1) plus

511 deaths will be evaluated, and the Data Safety Monitoring Board (DSMB) will provide  
512 recommendations accordingly. The DSMB will review all safety events and approve  
513 the ongoing conduct of the trial. Analyses that will aid their decision-making will be  
514 based on several sources:

515

516 First, a test of proportions will compare the AE rates between the standard-of-care arm  
517 and the experimental arms. Concerns about a worse safety profile will be flagged using  
518 a two-sided type I error rate of 0.1. Consider a scenario in which there are 10 out of 20  
519 AE in the standard-of-care arm and 14 out of 20 in the experimental arms. This  
520 corresponds to a two- sided p-value of 0.053 using Boschloo's test and would be  
521 reason for the DSMB to consider stopping the trial.

522

523 Similarly, a Bayesian posterior probability (with an uninformative prior) of the probability  
524 that the AE rate in the experimental arm is worse than that in the control arm. This will  
525 provide an additional means of interpreting the relative results. If this probability is  
526 high, the DSMB may recommend stopping the study. For example, in the 10/20 and  
527 14/20 scenario the posterior probability that the experimental arm has a worse rate of  
528 AEs is 94%. If the split was 10/20 versus 14/20, this probability would be 89%. The  
529 DSMB will be unblinded to safety data after every 15 patients recruited (5 per arm). At  
530 each point, absence of a significant safety signal (as outlined in the DSMB charter) will  
531 permit ongoing recruitment.

532

### 533 *Statistical analysis plan*

534 A detailed statistical analysis plan for LASER-TBM detailing the planned analysis for  
535 the primary and PK endpoints, co-authored and authorised by the trial statistician (JL)  
536 is included within the supplementary material of this manuscript.

537

538 **2.7 Adverse events**

539

540 *Assessment of AE*

541 Study participants will be monitored and assessed for new AE at all scheduled study  
542 visits (as shown in Table 2.3). At each visit, trial staff will also assess the evolution and  
543 outcome of previously recorded AE. Safety monitoring of the study will be performed by  
544 a DSMB as described below.

545

546 *Severity of AE*

547 All AE will be assessed for severity by study clinical investigators and graded as per  
548 the Division of AIDS (DAIDS) criteria(U.S. Department of Health and Human Services).

549 Each AE will be assigned a grade 1 to 5. For events not included in the protocol-  
550 defined grading systems, the following general definitions from grades 1 to 4 will be  
551 applied to classify event severity:

552

553 Changes in the severity of an AE should be documented to allow an assessment of the  
554 duration of the event at each level of intensity to be performed. AE characterised as  
555 intermittent require documentation of onset and duration of each episode.

556

557

558 *Solicited treatment related AE*

559 Table 2.1 lists AE of special interest which are considered 'solicited treatment related  
560 AE' and therefore comprise primary safety endpoints of this study. These AE are  
561 reported regardless of causal relationship to study drugs. For each AE there is a  
562 specific objective measure incorporating the DAIDS grading criteria and other  
563 parameters of clinical significance. The management of each AE is summarised.



564

565 Management of adverse events

566 Treatment must be discontinued for safety reasons if any clinical AE, laboratory  
567 abnormality, intercurrent illness, or other medical condition or situation occurs such that  
568 continued exposure to treatment would not be in the best interest of the participant.

569 Detailed guidance for management of AE is provided in the manual of operating  
570 procedures.

571

## 572 **2.8 Safety monitoring**

573 Safety oversight

574 Safety oversight will be under the direction of an independent DSMB. Comprised of  
575 independent internationally-recognized HIV-TB researchers and an independent  
576 statistician, the DSMB will meet after each 15 participants enrolled. The DSMB may  
577 also decide to convene an unscheduled review if warranted by safety or data quality  
578 concerns. The data for review will be prepared by an independent statistician.

579

580 The task of the DSMB will thus be to review study recruitment, data quality and trial  
581 drug safety and advise the sponsor of major safety issues and data quality issues. The  
582 DSMB may advise that trial enrolment should be paused or stopped entirely based on  
583 the decisions regarding the frequency and severity of solicited treatment related AE as  
584 outlined in Table 2.1 ('Solicited Treatment Related Adverse Events').

585

586 Pausing and stopping rules

587 Halting rules for safety reasons will be detailed in the DSMB charter. In the event of  
588 serious safety concerns, the DSMB chair will consult the full DSMB by email or

589 teleconference. Pending the DSMB response, the chair may use his/her discretion to  
590 recommend one or more if the following: Halt in study (arm) enrolment; halt in study  
591 (arm) dosing; provision of additional intervention; no action. After review, the DSMB  
592 will issue a recommendation to the trial steering committee to continue, modify (one or  
593 more arms) or terminate the trial.  
594

## 595 **2.9 Data access and handling**

596 Source documents

597 Source data are original records of clinical findings, observations, or other activities  
598 necessary for the evaluation of the trial. Examples of these original documents and  
599 data records include, but are not limited to: hospital records, laboratory reports, and  
600 radiological images. CRF may also be acceptable source documents. A complete list  
601 of source documents will be created prior to trial initiation.

602

603 The following individuals and groups will have access to study records:

- 604 - Members of the study team
- 605 - Relevant institutional review board (IRB)
- 606 - Regulatory agencies (South African Health Products Regulatory Authority -  
607 SAHPRA)
- 608 - Study Monitor
- 609 -

610 All site staff, the sponsor, and any sponsor representatives will preserve the  
611 confidentiality of all participants taking part in the study in accordance with ICH GCP,  
612 applicable South African national and local regulations and (to the extent applicable)  
613 the U.S. Health Insurance Portability and Accountability Act of 1996 (HIPAA). Subject

614 to the requirement for source data verification by the study personnel by reference to  
615 the participant's notes confidentiality of all participant identities will be maintained.  
616 Only participant study number and initials will be used on the CRF and in all study  
617 correspondence, as permitted. No material bearing a participant's name will be kept  
618 on file by the Sponsor. The written informed consent will contain a clause granting  
619 permission for review of the participants' source data.

## 620 **2.10 Data collection, management and storage**

621 Procedures to ensure data quality will be detailed in a Data Management plan. Data  
622 will be collected and captured onto hardcopy CRF on site and then entered into an  
623 electronic database. Clinical data will be entered onto paper CRF directly from the  
624 source documents on site. Other study data, such as laboratory reports and telephonic  
625 interviews will either be recorded on a specific study form before data entry or entered  
626 directly into the electronic database. CRF will be cross-checked for accuracy,  
627 authenticity and completeness at the site by study staff; checks for consistency will be  
628 implemented at the data entry level on site and centrally after data entry.

629

630 The data will be managed and stored using a GCP-compliant web-based REDCap®  
631 database custom-designed for the study. The REDCap® data entry and user  
632 permission structures provide auditing trails in line with international requirements.  
633 Access to the database is password controlled and will be limited to delegated trial staff  
634 with data entry and data management responsibilities.

635

636 Publication of research findings

637 The definition of publication for this purpose is any public presentation of the data  
638 emerging from this study. All unpublished information given to the investigator by the

639 Sponsor shall not be published or disclosed to a third party other than to the  
640 responsible IRB, within the understanding of the confidentiality of their nature, without  
641 the prior written consent of the Sponsor. Results of this research will be submitted for  
642 publication as soon as feasible upon completion of the study in the form of a joint  
643 publication(s) between the Sponsor and investigator(s), including site clinical and  
644 laboratory investigators, as appropriate.

645

646

## 647 **2.11 Trial committees, ethical procedures and sponsorship**

648

649 A Trial Management Group (TMG) responsible for the day-to-day management of the  
650 trial at the UCT CRC includes; National Trial Coordinator (Ms Mpumi Maxebengula),  
651 Lead Clinician (Dr Angharad Davis), Research Medical Officers (Dr Cari Stek, Dr Remy  
652 Daroowala, Dr Marise Bremer, Dr Stephani Botha, Dr Saalika Aziz), Project Manager  
653 (Ms Rene Goliath), Pharmacists (Ms Sonya Koekemoer, Mr Yakub Kadernani). The  
654 group will communicate weekly to discuss trial progress.

655

656 The Trial Steering Committee (TSC) is composed of Professor Guy Thwaites (chair,  
657 Infectious Disease Physician, University of Oxford), Professor Graeme Meintjes (site  
658 principal investigator), Dr Sean Wasserman (site principal investigator), Dr John Black  
659 (site principal investigator), Professor Robert J Wilkinson (National Principal  
660 Investigator) and Dr Angharad Davis (lead investigator). The role of the TSC is to  
661 provide overall supervision for the trial and provide advice through its independent  
662 chair. The ultimate decision for the continuation of the trial lies with the TSC.

663

664 The Data Safety and Management Board (DSMB) is composed of Professor David  
665 Laloo (chair, Director of the Liverpool School for Tropical Medicine and a Professor of  
666 Tropical Medicine), Dr David Meya (Infectious Diseases clinician, Senior Lecturer at the  
667 College of Health Sciences at Makerere University and Adjunct Associate Professor in  
668 the Division of Infectious Diseases and International Medicine at the University of  
669 Minnesota), Dr Evelyne Kestelyn (Head of the Clinical Trials Unit at the Centre for  
670 Tropical Medicine and Global Health, University of Oxford), Dr Maryline Bonnet  
671 (Medical Epidemiologist Institute of Research for Development and Epicentre), Dr  
672 Angela Crook, (Trial Statistician). The role of the DSMB is to protect and serve LASER-  
673 TBM trial patients and to assist and advise the Principal Investigators, so as to protect  
674 the validity and credibility of the trial.

675

#### 676 **ETHICS**

677

678 The trial has ethics approval from the University of Cape Town Human Research  
679 Ethics Committee (293/2018), Walter Sisulu University Human Research Committee  
680 (Ref 012/2019) and the South African Health Products Regulatory Authority (reference  
681 number 20180622). The trial is registered on the South African National Clinical Trials  
682 Register (DOH-27-0319-6230) and Pan African National Clinical Trials Register  
683 (PACTR201902921101705).

684

#### 685 **TRIAL SPONSOR**

686

687 University of Cape Town (Clinical Research Centre)

688 Delva Shamley

689 L51 Old Main Building

690 Groote Schuur Hospital  
691 Observatory, Cape Town  
692 Tel: 021 650 1975  
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694

## 695 **STUDY RECRUITMENT SITES**

696

### 697 **CAPE TOWN**

698 Groote Schuur Hospital , Main Road, Observatory, Cape Town, 7925, Republic of  
699 South Africa

700 Mitchells Plain Hospital, 8 A Z Berman Drive, Lentegeur, Cape Town, 7786, Republic  
701 of South Africa

702 New Somerset Hospital, Bay Court, Portwood Road, Green Point, Cape Town, 8001,  
703 Republic of South Africa

704

### 705 **PORT ELIZABETH**

706 Livingstone Hospital, Standford Road, Korsten, Port Elizabeth, 6020, Republic of South  
707 Africa

708

## 709 **2.12 Version control and protocol amendment policy**

710

711 Submitted version of the protocol: V6 (dated 11 May 2020)

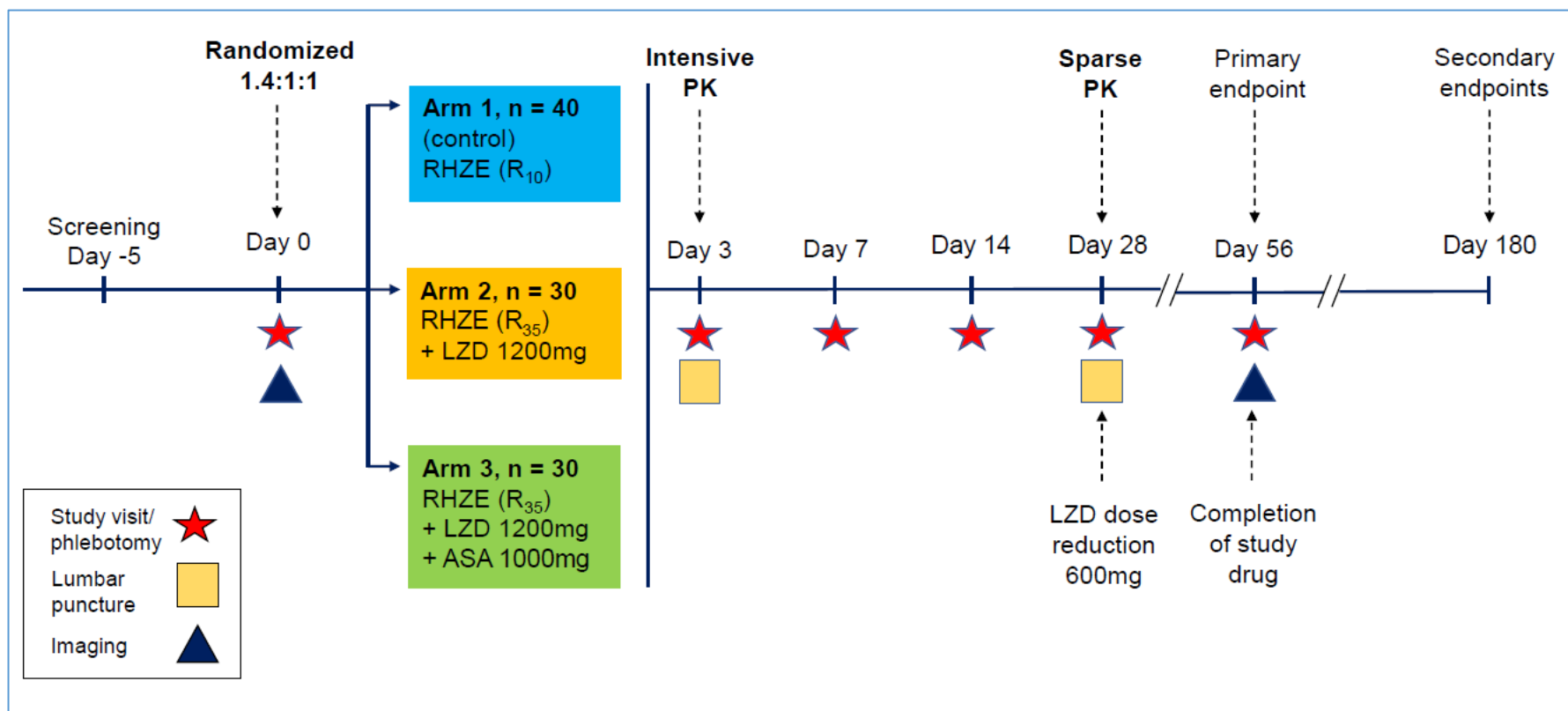
712

713 Any change to the protocol will be affected by means of a protocol amendment. The

714 PI, HREC, and Sponsor must agree on all amendments. No amendment will be

715 implemented until approved by the relevant authorities and signed by all required  
716 parties. Exceptions to this are when the PI considers that the participant's safety is  
717 compromised. No deviations from or changes to the protocol should be initiated  
718 without prior written approval from the IRB and regulatory authority. The PI, or  
719 designated site staff, is responsible for documenting and explaining any deviations  
720 from the protocol. Protocol deviations must be sent to the Sponsor and IRB in  
721 accordance with standard procedures.

722 **Figure 2.1 LASER-TBM Study Design**



723

724 Study design schematic describing randomisation to study arms, treatment intervention per arm, visit schedule, overview of clinical  
 725 procedures and timepoints relating to primary and secondary endpoint data collection. Abbreviations: RHZE: Rifampicin, Isoniazid,  
 726 Pyrazinamide, Ethambutol; R10: Rifampicin 10mg/kg/day; R35: Rifampicin 35mg/kg/day; LZD: Linezolid; ASA; Aspirin;



727 **Box 2.1 LASER-TBM eligibility criteria**

728	<b>Inclusion criteria</b>
729	<ul style="list-style-type: none"><li>• Age &gt;18 years</li></ul>
730	<ul style="list-style-type: none"><li>• proven HIV-1 seropositivity</li></ul>
731	<ul style="list-style-type: none"><li>• Diagnosis of 'possible', 'probable' or 'definite' TBM</li></ul>
732	<b>Exclusion criteria</b>
733	<ul style="list-style-type: none"><li>• Rifampicin-resistant <i>M.tb</i> detected on any clinical specimen;</li></ul>
734	<ul style="list-style-type: none"><li>• History of allergy or hypersensitivity to RIF, isoniazid, ethambutol,</li></ul>
735	<ul style="list-style-type: none"><li>pyrazinamide, LZD or ASA;</li></ul>
736	<ul style="list-style-type: none"><li>• Received more than 5 days of antitubercular therapy in the 30 days</li></ul>
737	<ul style="list-style-type: none"><li>prior to screening;</li></ul>
738	<ul style="list-style-type: none"><li>• Receipt of regular daily ASA or NSAID prior to TBM diagnosis</li></ul>
739	<ul style="list-style-type: none"><li>• CSF unobtainable by lumbar puncture or another procedure;</li></ul>
740	<ul style="list-style-type: none"><li>• Evidence of bacterial or cryptococcal meningitis;</li></ul>
741	<ul style="list-style-type: none"><li>• Severe concurrent uncontrolled opportunistic infection including, but</li></ul>
742	<ul style="list-style-type: none"><li>not limited to, active cytomegalovirus-associated disease, Kaposi</li></ul>
743	<ul style="list-style-type: none"><li>sarcoma, <i>Pneumocystis jirovecii</i> pneumonia, HIV related or</li></ul>
744	<ul style="list-style-type: none"><li>unrelated malignancy, or gastrointestinal bleeding;</li></ul>
745	<ul style="list-style-type: none"><li>• Any other form of immunosuppressive therapy, including</li></ul>
746	<ul style="list-style-type: none"><li>antineoplastic and biologic agents, apart from corticosteroids;</li></ul>
747	<ul style="list-style-type: none"><li>• More than 17 weeks pregnant at baseline;</li></ul>
748	<ul style="list-style-type: none"><li>• Peripheral neuropathy scoring Grade 3 or above on the BPNS;</li></ul>
749	<ul style="list-style-type: none"><li>• Any disease or condition in which the use of the standard anti-TB</li></ul>
750	<ul style="list-style-type: none"><li>drugs (or any of their components) are contraindicated. This</li></ul>
751	<ul style="list-style-type: none"><li>includes, but is not limited to, allergy to any TB drug or their</li></ul>
752	<ul style="list-style-type: none"><li>components;</li></ul>
753	<ul style="list-style-type: none"><li>• The presence of one or more of the following:</li></ul>
754	<ul style="list-style-type: none"><li>- Estimated glomerular filtration rate (eGFR) &lt; 20ml/min/1.73 m<sup>2</sup>*</li></ul>
755	<ul style="list-style-type: none"><li>- INR &gt; 1.4 and/or clinical evidence of liver failure or decompensated</li></ul>
756	<ul style="list-style-type: none"><li>cirrhosis;</li></ul>
757	<ul style="list-style-type: none"><li>- Haemoglobin &lt; 8.0 g/dL;</li></ul>
758	<ul style="list-style-type: none"><li>- Platelets &lt; 50 x10<sup>9</sup> /L;</li></ul>
759	<ul style="list-style-type: none"><li>- Neutrophils &lt; 0.5 x 10<sup>9</sup> cells/L;</li></ul>
760	<ul style="list-style-type: none"><li>• Any disease or condition in which any of the medicinal products</li></ul>
761	<ul style="list-style-type: none"><li>listed in the section pertaining to prohibited medication (See Box</li></ul>
762	<ul style="list-style-type: none"><li>2.4) is used and cannot be safely stopped;</li></ul>
763	<ul style="list-style-type: none"><li>• Known or suspected history of drug abuse or any other reason that</li></ul>
764	<ul style="list-style-type: none"><li>is, in the opinion of investigators, sufficient to compromise the safety</li></ul>
765	<ul style="list-style-type: none"><li>or cooperation of the participant.</li></ul>
766	

767 \*Calculated using the Cockcroft-Gault equation; INR: International  
768 normalised ration; BPNS: Brief Peripheral Neuropathy Score; NSAID: Non  
769 Steroidal Anti Inflammatory Drug;  
770  
771

772 **Box 2.2 Summary of uniform case definition criteria for tuberculous meningitis used to define ‘definite’, ‘probable’ and ‘possible TBM in**

773 **LASER-TBM (Marais et al., 2010)**

All patients require symptoms and signs suggestive of meningitis including one of more of the following; headache, irritability, vomiting, fever, neck stiffness, convulsions, focal neurological deficits, altered consciousness or lethargy. An alternative diagnosis must be excluded. In this context they should then be subsequently assessed on the following criteria.

Criteria	Score
<b>Clinical (maximum category score = 6)</b>	
Symptom duration of > 5 days	4
Weight loss, night sweats or persistent cough for > 2 weeks)	2
Focal neurological deficit (excluding cranial nerve palsy)	1
Cranial nerve palsy	1
Altered consciousness	1
<b>CSF (maximum category score = 4)</b>	
Clear appearance	1
Cells 10-500/ $\mu$ L	1
Lymphocytic predominance > 50%	1
Protein concentration greater than 1g/L	1
CSF to plasma glucose concentration of less than 50% or an absolute CSF glucose concentration less than 2.2mmol/L	1
<b>Cerebral Imaging criteria (max score 6)</b>	
Hydrocephalus	1
Basal meningeal enhancement	2
Tuberculoma	2
Infarcts	1
Pre-contrast basal hyperdensity	2
<b>Evidence of TB elsewhere (max score 4)</b>	

CXR suggestive of active TB	4 (miliary) 2 (other active TB)
CT/MRI/Ultrasound showing evidence for TB outside the CNS	2
AFB identified or M.tb cultured from another source or positive Urine LAM	4
<b>Scoring – scores should be added up and the one of the following diagnostic categories assigned</b>	
<b>Definite</b>	
Acid fast bacilli seen in CSF <i>and/or</i> M.tb culture from CSF	
<b>Probable</b>	
Total diagnostic score of 12 (cerebral imaging available) or 10 (cerebral imaging unavailable). At least 2 points should come from CSF or cerebral imaging criteria.	
<b>Possible</b>	
Total diagnostic score of 6-11 (cerebral imaging available) or 6-9 (cerebral imaging unavailable).	

775 **Box 2.3 Modified Rankin Score**

776

SCORE	DESCRIPTION
0	No symptoms
1	No significant disability. Able to carry out usual activities, despite some symptoms.
2	Slight disability. Able to look after own affairs without assistance, but unable to carry out all previous activities.
3	Moderate disability. Requires some help, but able to walk unassisted.
4	Moderately severe disability. Unable to attend to own bodily needs without assistance, and unable to walk unassisted.
5	Severe disability. Requires constant nursing care and attention, bedridden, incontinent.
6	Dead

777

778 **Box 2.4 LASER-TBM contraindicated medications**

779

780 **Tricyclic antidepressants:** Amitriptyline, Amoxapine, Clomipramine, Desipramine, Doxepin, Imipramine, Nortriptyline, Protriptyline, Trimipramine.  
 781 **Selective Serotonin Re-uptake Inhibitors (SSRI's):** Citalopram, Escitalopram, Fluoxetine, Fluvoxamine, Paroxetine, Sertraline. Serotonin-  
 782 Noradrenaline Re-uptake Inhibitors (SNRI's) Venlafaxine, Duloxetine, Levomilnacipran, Milnacipran. **Serotonin Receptor Agonist,** Buspirone,  
 783 **Mono-amine Oxidase Inhibitors (MOAIs),** Isocarboxazid, Nialamide, Phenelzine, Tranylcypromine, Selegiline, Rasagaline, Toloxatone. **Reversible**  
 784 **MOA Inhibitors (RIMAs):** Moclobemide, Pirlindole. **Migraine medications:** Triptans. **Macrolide antibiotics:** Clarithromycin, Erythromycin,  
 785 Troleandomycin. **Opiate analgesics:** Methadone, Tramadol, Pentazocine. **Stimulants:** MDMA (ecstasy), Cocaine, Methamphetamine **Hormonal**  
 786 **treatment:** Gestodene, Testosterone. **Other medications:** Antiretrovirals – Atazanavir, Anti-arrhythmic – Quinidine, Anti-malarial – Quinine,  
 787 Chemotherapy – Doxorubicin, Asthma –Furafylline, Hypertension – Hydracarbazine, Antifungal – Ketokonazole, Amino-acid - Tyramine

788

789 **Bold** represents class of drug. Underlined medications represent commonly used medications in South Africa.

790

791 **Table 2.1 Solicited treatment related adverse events, objective measures for assessment and management plan in each setting**

792

Adverse Event (responsible study drug)	Objective measure	Parameter	Management
Gastrointestinal haemorrhage (ASA)	Clinical and laboratory measures suggesting GI haemorrhage.	i) Vomiting fresh or changed blood of any volume, ii) Melena, iii) Unexplained drop in Hb concentration of >2g/L or iv) > 5mls of fresh or changed blood aspirated from nasogastric tube.	Discontinue study drug permanently.
Intracerebral haemorrhage (ASA)	Radiological evidence of haemorrhage.	CT or MRI finding	Discontinue study drug permanently.
Transaminitis (RIF)	alanine transferase (ALT), bilirubin	New Grade 3 or above	Discontinue study drug (and other potentially hepatotoxic agents). Place on alternative treatment for TBM if background regimen affected. Re-test every 2 days. Resume study drug with an escalating dose rechallenge once ALT < 100 IU and total bilirubin within normal range.
Anaemia (LZD)	Hemoglobin (Hb)	New Grade 3	Discontinue study drug (plus any other myelosuppressive drugs as appropriate). Consider transfusion with packed cells or erythropoietin therapy. Monitor Hb twice weekly. Restart at 50% dose once Hb ≥ 8 mg/dL
		New Grade 4	Discontinue study drug permanently. Consider hospital admission and/or transfusion with packed cells or erythropoietin therapy. Re-test every 2 days.
Neutropenia (LZD)	Neutrophils	New Grade 3	Discontinue study drug. Monitor white cell count (WCC) every week. Restart at 50% dose once neutrophil count $0.5 \times 10^9$ cells/L.
		New Grade 4	Discontinue study drug permanently. Consider therapy with GM-CSF. Monitor WCC every 1 – 2 days.
Thrombocytopenia (LZD)	Platelet (Plt) count	New Grade 3	Discontinue study drug (plus any other myelosuppressive drugs as appropriate). Monitor Plt count twice weekly. Restart at 50% dose once Plt count $> 50 \times 10^9$ cells/L.
		New Grade 4	Discontinue study drug permanently. Consider hospital admission and/or transfusion pooled Plts. Re-test every 1 - 2 days.

Peripheral Neuropathy (LZD)	Full neurological history and examination, Brief Peripheral Neuropathy Score (BPNS) and Modified Total Neuropathy Score (mTNS)	Change in clinical history of examination resulting in: i) 1 grade increase in BPNS ii) 2 grade change in any modality on mTNS	Review with a view to discontinuing study drug (plus any other neuropathic drugs like isoniazid). Consider restarting at 50% dose once completely resolved.
Optic Neuropathy (LZD)	14-plate Ishihara Test, visual acuity measured by logMAR chart	Change in score of 2 on 14-plate Ishihara Colour Test or new or worse logMAR score of 0.2	Stop study drug and EMB and refer for formal ophthalmological assessment. If assessment consistent with optic neuritis do not restart drug.

793  
794  
795  
796

Grade relates to DAIDS criteria(Services, Corrected Version 2.1 July 2017).

797 **Table 2.2 Details and dosing of study drug regimen provided for 56 days post randomisation**

798

Drug						
Arm	Rifampicin	Isoniazid	Ethambutol	Pyrazinamide	Linezolid	ASA
1	10 mg/kg O.D.	5 mg/kg O.D.	15 mg/kg O.D.	25 mg/kg O.D.		
2	35 mg/kg O.D.	5 mg/kg O.D.	15 mg/kg O.D.	25 mg/kg O.D.	1200 mg O.D. (28 days) then 600 mg O.D. (28 days)	
3	35 mg/kg O.D.	5 mg/kg O.D.	15 mg/kg O.D.	25 mg/kg O.D.	1200 mg O.D. (28 days) then 600 mg O.D. (28 days)	1000 mg O.D.

799  
800  
801  
802  
803  
804  
805

806 **Table 2.3 Planned study assessments and procedure per study date**

807

Visit (window in days)	SCR	ENR	Day 3 (+/- 1)	Day 7 (+/- 2)	Day 14 (+/- 2)	Day 28 (+/- 3)	Day 56 (+/- 4)	Day 180
<b>Bedside</b>								
Study Informed Consent	x	x						
Vital Signs	x	x	x	x	x	x	x	
Medical History	x	x	x	x	x	x	x	x
Physical Examination	x	x	x	x	x	x	x	
BPNS and mTNS	x							
Modified Rankin Score		x		x	x	x	x	x
Insomnia Questionnaire						x	x	
MOCA, IHDS, EQ5d5L							x	
Neurocognitive mini-battery								x
AE/SAE, Adherence Monitoring		x	x	x	x	x	x	
Randomisation and treatment assignment		x						
<b>Blood</b>								
Weight		x	x	x	x	x	x	
Haematology: FBC and white cell differential								
Biochemistry: Creatinine, eGFR, electrolytes, LFTs	x		x	x	x	x	x	
INR	x		x			x		
HIV-1 ELISA +/- HIV Rapid Test (x2) if required	x							
CD4+ count, HIV Viral Load		x						
Plasma for PK sub-study (sparse sampling)						x	x	
Plasma for PK sub-study (intensive sampling – if consented)			x					
Stored plasma for immunological, proteomic and metabolomic profiling		x	x	x	x	x	x	
PBMC for storage		x	x			x		
Whole blood for RNA extraction		x	x	x	x	x	X	
Whole blood for DNA extraction (if consented)		x						
<b>Urine</b>								
Urine for pregnancy test	x							
Urine for storage		x			x		x	
<b>Cerebrospinal Fluid Analysis</b>								
Cell count, MC+S, TB culture, GeneXpert Ultra (inc Rif resistance)			x			x		
Biochemistry: protein and glucose			x			x		

Stored CSF for immunological, cellular, proteomic and metabolomic profiling;			x			x		
CSF for RNA extraction								
CSF for PK sub-study			x			x		
<b>Imaging</b>								
MRI Head, or CT Head if MRI not tolerated (+/- 5 days)		x					x	

808 Table Abbreviations: SCR: Screening; ENR: Enrolment; AE: Adverse Event; BPNS: Brief Peripheral Neuropathy Score; CSF:  
809 Cerebrospinal Fluid; CT: Computerised Tomography; FBC: Full Blood Count; LFT: Liver Function Tests; IHDS: International HIV  
810 Dementia Score; MC+S: microscopy, culture and sensitivity; MOCA: Montreal Cognitive Assessment; mTNS: modified Total Neuropathy  
811 Score; MRI: Magnetic Resonance Imaging; PAOFI: Patients Assessment of Own Functioning Inventory; PBMC: Peripheral Blood  
812 Mononuclear Cells; PK: Pharmacokinetic;  
813  
814



815 **Chapter 3. Results from a phase 2A trial of the**  
816 **safety and tolerability of increased dose**  
817 **rifampicin and adjunctive linezolid, with or**  
818 **without aspirin, for HIV-associated tuberculous**  
819 **meningitis (The LASER-TBM Trial)**

820

821 **3.1 Introduction**

822 The background to the clinical trial is described within chapter 2 of the thesis.

823

824 **3.2 Methods**

825 The study methods are described in detail in chapter 2 of the thesis. The  
826 following information adds further detail, not included within chapter 2 on the final  
827 sample size and statistical analysis.

828

829 **Sample size**

830

831 No formal statistical power calculation was performed. Even as single adjunctive  
832 therapies, there was limited available data on the use of these drugs in TBM to  
833 predict likely rate of AESI (table 3.1) and/or death. Given this would be further  
834 complicated when considering likely event rate when the drugs were combined,  
835 it was felt a more pragmatic approach was to create a recruitment target of 100  
836 participants with frequent blinded review of cumulative safety events by an

837 independent DSMB. A secondary aim of LASER-TBM was also to serve as a  
838 planning study to generate PK and safety data to inform a phase 3 RCT of  
839 intensified treatment in TBM (NCT04145258), which in part would influence  
840 resulting sample size of that study. The decision to stop recruitment prior to 100  
841 participants enrolled was made as a result of the following:

- 842 - The rate of recruitment was slower than anticipated due to the COVID-19  
843 pandemic
- 844 - Funding for the trial was due to cease in March 2021 and therefore  
845 recruitment could only take place up until January 2021
- 846 - DSMB review of safety data (both during recruitment to the trial and  
847 following enrolment of last recruit) had revealed no reason for the  
848 planned RCT not to go ahead.

849

## 850 **Statistical analysis**

851

852 Analysis was performed in GraphPad Prism v.9.0 and R v.3.6.0. The primary  
853 analysis was performed in the modified intent-to-treat population (those who  
854 receive any dose of the study drug). A sensitivity analysis was planned for the  
855 per-protocol population (those who completed treatment as specified in the  
856 protocol). However given the small sample size and since these populations  
857 were similar, here we report the most conservative analysis (modified intention to  
858 treat).

859

860 The primary endpoint, frequency of AESI or death (where data is censored at the  
861 first event prior to day 56) was summarized and compared across arms using a  
862 chi-squared test. A time to event analysis was performed for worst grade (in  
863 each individual participant) AESI or death; comparisons between study groups  
864 were made using the log-rank test. Neurological disability (measured by Modified  
865 Rankin Score), as well as radiological outcomes at day 56 were compared  
866 across treatment arms using chi-squared test. We used spaghetti plots to  
867 visually represent longitudinal CSF parameters (lymphocytes,

868 polymorphonuclear cells, protein and glucose) over time and used t tests to  
869 compare longitudinal summaries (mean and SD) of each individual trajectory  
870 across treatment arms.

871

872 Details of further analysis can be found in the full statistical analysis plan  
873 published alongside the study protocol (Davis et al., 2021).

874

875

### 876 **3.3 Results**

877

878 98 patients were screened and 52 were randomised between June 2019 and  
879 January 2021 (Figure 3.1). Reasons for screening exclusion are summarised in  
880 Table 3.6. One participant was randomised but excluded prior to any study IP  
881 being dispensed due to emergence of an exclusion criterion (eGFR <20) on a  
882 hospital blood test performed prior to randomisation. Another participant was  
883 excluded from the modified intention to treat analysis as they died prior to  
884 receiving any dose of study drug. Six participants discontinued the study prior to  
885 day 56, and a further four participants discontinued between day 56 and day 180  
886 (Table 3.7).

887

888 The baseline characteristics of the participants stratified by treatment arm are  
889 described in Table 3.2. Most participants were male (71%) and the median age  
890 was 39 (34-46). Most participants had mild disease (MRC Grade 1 59%; Grade 2  
891 39%; Grade 3 2%). A third (33%) of participants had microbiologically confirmed  
892 TBM at baseline, with the remaining participants defined as possible (41%) or  
893 probable TBM (25%) as per the uniform TBM case definition (Marais et al., 2010).

894

895 The primary endpoint analysis was performed in the modified intention to treat  
896 population (n=50; arm 1, 20; arm 2, 14; arm 3, 16). The composite primary  
897 endpoint of AESI or death occurred in 6/20 in arm 1, 4/14 in arm 2, and 10/16  
898 participants in arm 3, ( $p=0.083$ ). The occurrence of each category of AESI  
899 stratified by treatment arm are summarised in table 3.3 with further detail on  
900 timing and outcome of each of these events listed in table 3.4. Frequency of  
901 death prior to day 56 was similar across arms (n=7; arm 1, 3; arm 2, 1; arm 3, 3;  
902  $p=0.649$ ) and in no case was cause of death related to study investigational  
903 product (table 3.5). Grade 3 or 4 AE (grade 3: arm 1, 7 vs arm 2, 7 vs ar, 3, 9,  
904  $p=0.44$ ; grade 4: arm 1, 2 vs arm 2, 4 vs arm 3, 4,  $p=0.38$ ) or serious adverse  
905 events for reasons other than death (arm 1, 6 vs arm 2, 8 vs arm 3, 7,  $p=0.37$ )  
906 were similar across treatment arms.

907

908 The cumulative incidence of the composite endpoint of worst grade AESI or  
909 death at day 56 demonstrated worse outcomes when comparing arm 3 vs arm 1  
910 ( $p=0.043$ ), with similar proportions observed in other pre-specified analysis (arm  
911 2 vs arm 1 ( $p=0.3$ ), arm 2+3 combined vs arm 1 ( $p=0.5$ )) (Figure 3.2, log rank  
912 test). Similarly, analysis for death alone demonstrated no difference between  
913 arms (Figure 3.3). The cumulative incidence of AESI events was greater in arm 3  
914 vs arm 1 ( $p=0.02$ ), however, when arms 2 and 3 were combined and compared  
915 to arm 1 this difference was less marked ( $p=0.18$ ) (Figure 3.4).

916

917 The frequency of grade 5 MRS (severe disability) or death was 4 (arm 1) vs 3  
918 (arm 2) vs 5 (arm 3),  $p=0.774$ . The frequency of good (defined as MRS grade 0-  
919 3), and bad outcomes (MRS Grade 4-6) were similar across arms ( $p=0.616$ )  
920 (Figure 3.5). *Post hoc* analysis of change in neurological function (as measured  
921 by MRS) found similar changes of MRS from baseline to day 56 between the

922 three arms (Figure 3.4a). Few IRIS events occurred (arm 1, 2; arm 2, 2; arm 3,  
923 3), of which 4/7 were defined as neurological IRIS. Within the first 56 days of  
924 treatment, four participants developed new onset lower limb weakness (TB  
925 myelopathy 2; TB radiculomyelopathy/arachnoiditis 1; other (no cause found  
926 prior to death) 1); three participants developed a new onset hemiplegia; one  
927 patient developed a new onset isolated cranial nerve palsy (lower motor neuron  
928 VII). Thirteen participants presented with new onset seizures at TBM diagnosis.  
929 A further nine participants had new onset seizures within the first 2 months of  
930 follow up (arm 1, 5; arm 2; 2; arm 3, 2;  $p=0.54$ ). Baseline and follow up imaging  
931 was performed in only 9 patients at the timepoints pre-specified within the  
932 protocol. Follow up imaging demonstrated new or worsening leptomeningeal  
933 enhancement in 2/9 participants (arm 1 and arm 2), new evidence of infarction in  
934 2/9 participants (arm 1 and arm 2), new or worsening tuberculomas in 2/9  
935 participants (arm 1 and arm 2) which was associated with worsening sulcal  
936 effacement in 1/9 participant (arm 1).

937

938 Spaghetti plot analysis of longitudinal CSF parameters (lymphocyte and  
939 polymorphonuclear cell count, protein and glucose) over time demonstrated  
940 downward trend of parameters across all three treatment arms (figure 3.6).  
941 Individual values are plotted and the superimposed line represents the mean  
942 values at each timepoint in each treatment arm. T tests comparing mean and  
943 variance at each time point demonstrated no difference between arms.

944

945

### 946 **3.4 Discussion**

947

948 The LASER-TBM study was a phase 2a RCT which evaluated the safety of high-  
949 dose rifampicin (35mg/kg daily), adjunctive linezolid (1200mg reducing to 600mg  
950 after 28 days) and adjunctive aspirin (1000mg daily) for the first 56 days of  
951 treatment in HIV-associated TBM. Primary endpoint analysis showed no  
952 significant difference in the incidence of AESI or death between treatment arms,  
953 although there was a trend towards an increase in events in arm 3. There was  
954 no difference in death or disability at day 56 across arms; and a similar  
955 frequency of clinical or radiological events occurred in each arm. Exploratory  
956 analysis found no difference in change in CSF parameters over time by arm.

957

958 Although secondary analysis revealed a significantly higher number of events  
959 (AESI or death) in arm 3 vs arm 1 ( $p=0.04$ ), it is reassuring that no deaths were  
960 attributed to aspirin. Only one bleeding event occurred, after minimal exposure to  
961 high-dose aspirin (1 dose), resolved immediately following discontinuation of the  
962 drug, and was not associated with any laboratory markers to suggest significant  
963 gastrointestinal bleeding. Toxicity attributable to linezolid was similarly mild; of  
964 seven events potentially attributable to linezolid occurring in participants  
965 randomised to experimental arms, 3/7 were due to an alternative cause and 2/7  
966 had recovered prior to the subsequent study visit. No patient was formally  
967 diagnosed with peripheral neuropathy, which is expected given the most recent  
968 studies showing a median time to onset of neuropathy occurs after 10 weeks of  
969 treatment (Wasserman et al., 2022, Conradie et al., 2020). Only one participant  
970 developed a change in visual acuity, which may have been due to linezolid,  
971 although on review by an ophthalmologist was assessed as more likely due to  
972 ethambutol. The number of participants in whom potential abnormalities were  
973 detected using the LogMAR and tumbling E assessments, compared to the  
974 confirmed number of cases of optic neuropathy calls into question the specificity  
975 of these outcome measures. Given that linezolid has potential to treat TBM as

976 well as drug resistant TB, we must consider whether better outcome measures  
977 can be developed to reliably detect abnormalities attributable to these drugs in  
978 both the clinical and research setting in order to prevent overestimation of  
979 toxicity, particularly when used for a short duration. Toxicity due to rifampicin  
980 was similarly infrequent with only two participants developing clinically significant  
981 transaminitis. In both cases the transaminitis recovered with treatment  
982 interruption. These results suggest that toxicity associated with the enhanced  
983 antitubercular regimen (rifampicin 35mg/kg and adjunctive linezolid) is not  
984 common when used in combination for two months to treat HIV-associated TBM.  
985 This is encouraging in the context of a disease where no specific evidenced-  
986 based antitubercular regimen exists and provides rationale for the ongoing  
987 phase 3 RCT (NCT04145258) where participants are randomised to receive both  
988 high-dose rifampicin and linezolid at doses identical to that used in this study.  
989  
990 There are several limitations to this study. Although no formal power calculation  
991 took place, final sample size was substantially smaller than the target of 100  
992 participants. The COVID-19 pandemic adversely affected recruitment to the  
993 study, and in January 2021 a decision was made to stop recruitment to enable  
994 commencement of a similar phase 3 study which was ready to start resulting in a  
995 lower than proposed sample size. It is unknown whether the significantly higher  
996 number of AESI or death in arm 3 vs arm 1 demonstrated within the secondary  
997 analysis reflects a true safety risk of the regimen containing aspirin, or is due to  
998 chance given the lower than anticipated numbers of participants recruited.  
999 Secondly, the majority participants recruited had mild TBM. The reasons for this  
1000 is likely multifactorial including i) patients dying prior to screening given that up to  
1001 5 days of TB treatment was allowed prior to enrolment and ii) patients with  
1002 decreased levels of consciousness arriving at hospital alone and therefore not  
1003 having available next of kin available for proxy consent. In the latter case a

1004 protocol amendment was approved to allow deferred consent in these patients,  
1005 however, is likely to explain in part the higher rate of mild disease in our cohort.  
1006 The mild level of disease within our patient cohort likely explains the low level of  
1007 mortality; 16% 2-month mortality contrasted the oft quoted 50% mortality within  
1008 the literature (Marais et al., 2011). The primary endpoint of AESI *or* death was  
1009 designed with the assumption that observed mortality would be near or  
1010 approaching 50%. The relatively few numbers of deaths led to a greater  
1011 proportion of AESI in the composite endpoint of AESI *or* death. Given that all of  
1012 the listed AESI were proportionally more likely to occur in the experimental arm  
1013 3, it is unsurprising that the number of events within the composite endpoint of  
1014 AESI *or* death occurred in arm 3 where the greatest number of interventions was  
1015 given. This is supported by the observation that when considering AESI alone,  
1016 the cumulative incidence of events was significantly greater in arm 3, suggesting  
1017 that the composite endpoint of AESI *or* death was driven by the higher rate of  
1018 AESI in arm 3. The bias towards milder disease may also have affected the  
1019 efficacy analysis. Given the nature of TBM and clinical trial research, it is  
1020 challenging to ensure inclusion of those with severe TBM, in particular around  
1021 gaining proxy or deferred consent in unconscious patients and ensuring early  
1022 referrals to include those who are most likely to die early within the disease  
1023 course. Future trials especially phase 3 RCT must endeavour to overcome these  
1024 hurdles and include such patients to ensure generalisability of results.

1025

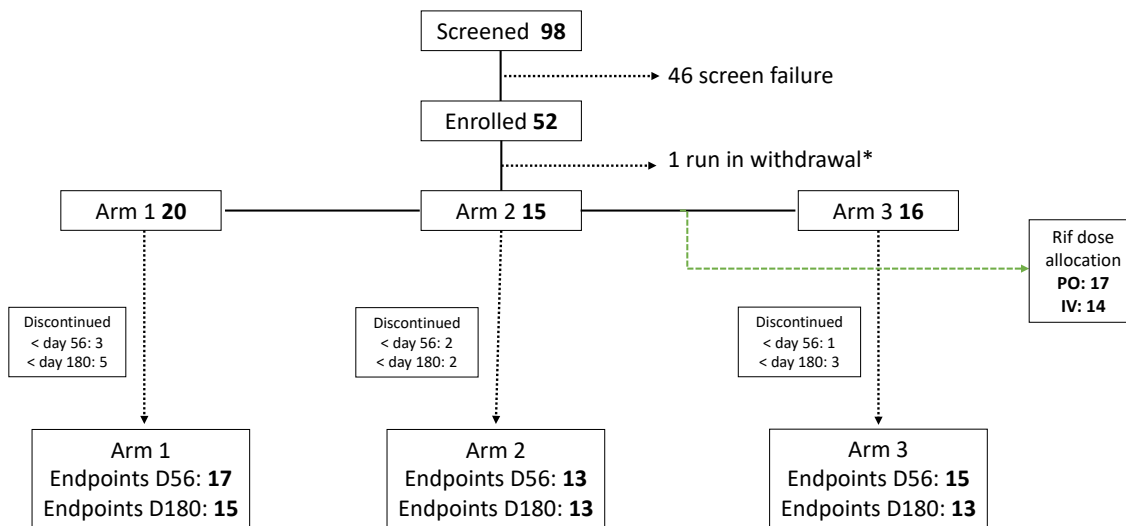
1026 Our study is the first RCT to evaluate linezolid in TBM and demonstrates that this  
1027 important drug can be safely added to standard of care to treat HIV-associated  
1028 TBM. It is also the first study to date to systematically evaluate the safety of a  
1029 novel drug regimen containing enhanced antitubercular treatments alongside a  
1030 host directed therapy in TBM, demonstrating that this approach can be safe. Our  
1031 results reassure that high-dose rifampicin and linezolid may be safely combined



1032 in HIV-associated TBM and supports evaluation of the efficacy of these drugs  
1033 either alone or in combination in phase 3 trials. Within our study we did not see  
1034 any significant bleeding events with the use of high-dose aspirin: a larger study  
1035 is now required to see if potential harm is offset by a morbidity and mortality  
1036 benefit.

1037 **Figure 3.1 CONSORT diagram for LASER-TBM**

1038



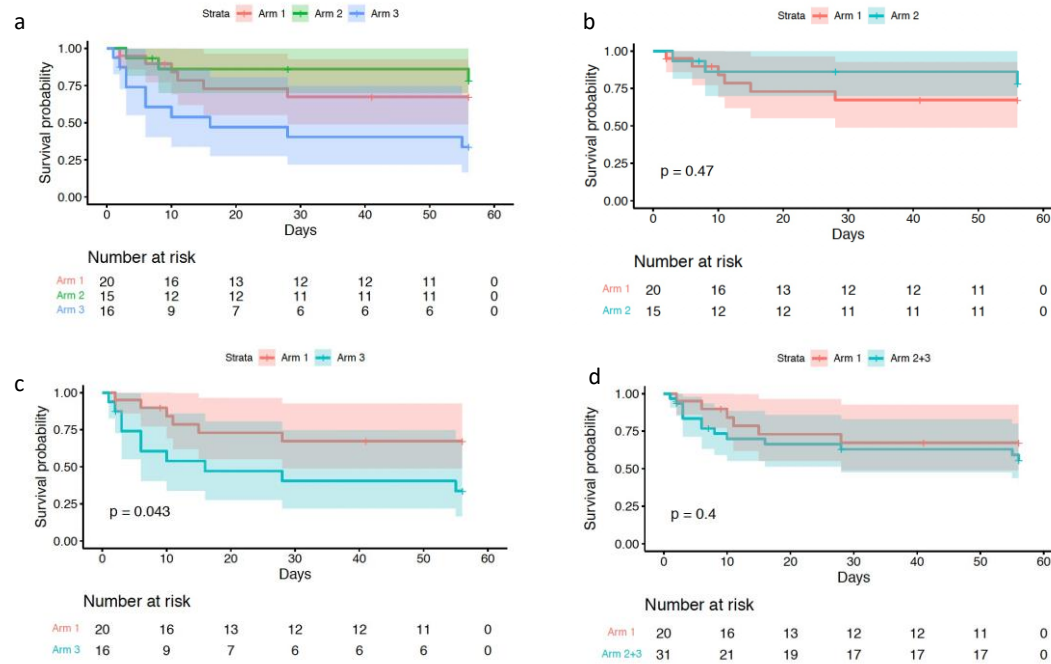
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1040

1041 CONSORT diagram to describe recruitment and arm allocation. Reasons for screening exclusions and early study withdrawals are listed  
1042 in Tables 3.6 and 3.7; Rif, rifampicin; \*patient randomized but withdrawn prior to receiving study IP due to emergence of exclusion criteria  
1043

1044 **Figure 3.2 Kaplan-Meier analysis of time to worst grade adverse events of special interest or death**

1045



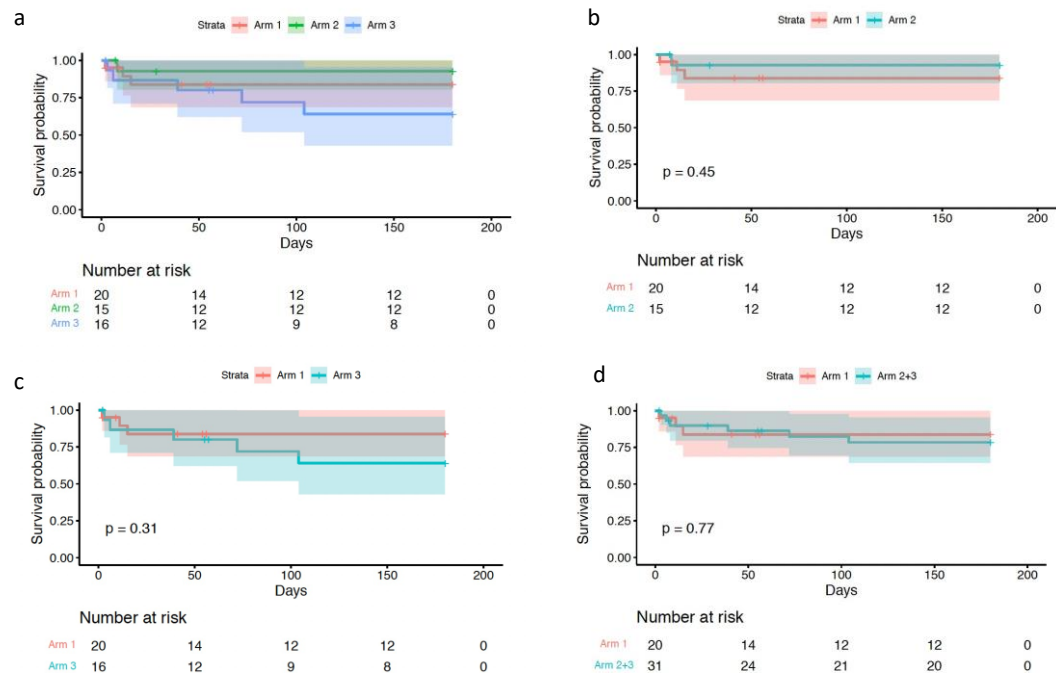
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1047

1048 Kaplan-Meier analysis of time to worse grade AEI or death, comparing arm 1, 2 and 3 (a), arm 2 vs arm 1 (b), arm 3 vs arm 1 (c) and  
 1049 arm 2 and 3 combined vs arm 1 (d).  
 1050

1051 **Figure 3.3 Kaplan-Meier analysis time to death**

1052



1053

1054

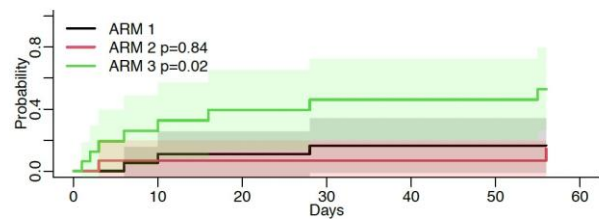
1055 Kaplan-Meier analysis of time to death, comparing arm 1, 2 and 3 (a), arm 2 vs arm 1 (b), arm 3 vs arm 1 (c) and arm 2 and 3 combined  
 1056 vs arm 1 (d).

1057

1058 **Figure 3.4 Kaplan-Meier analysis time to worst grade adverse event of special interest**

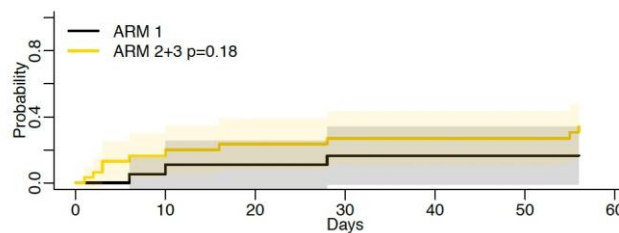
1059

a



	0	10	20	30	40	50	60
ARM 1	20	16	13	12	12	11	0
ARM 2	15	12	12	11	11	11	0
ARM 3	16	9	7	6	6	6	0

b



	0	10	20	30	40	50	60
ARM 1	20	16	13	12	12	11	0
ARM 2+3	31	21	19	17	17	17	0

1060

1061 Kaplan-Meier analysis of time to adverse event of special interest (AESI), comparing arm 1, 2 and 3 (a), and arm 2 and 3 combined vs  
1062 arm 1 (b).

1063

1064

1065

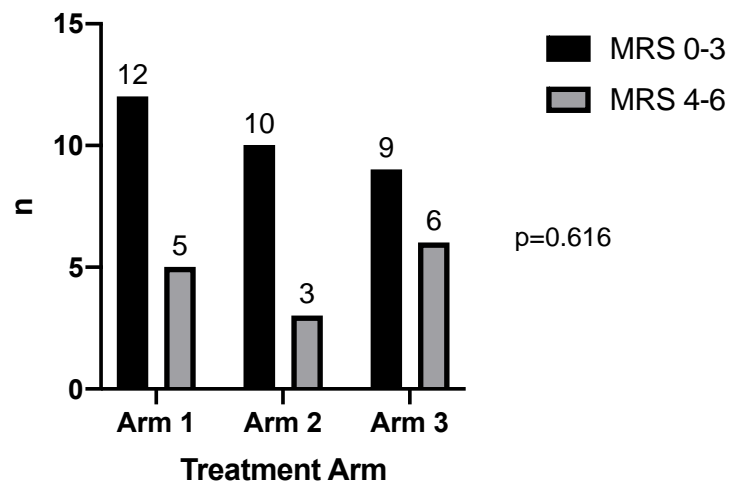
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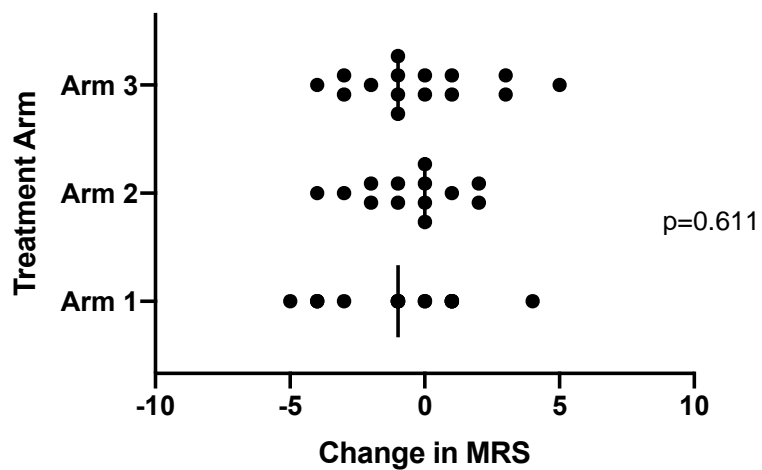
1068 **Figure 3.5 Functional neurological outcome at day 56 as defined by modified Rankin scale**

1069

1070 a



b



1071

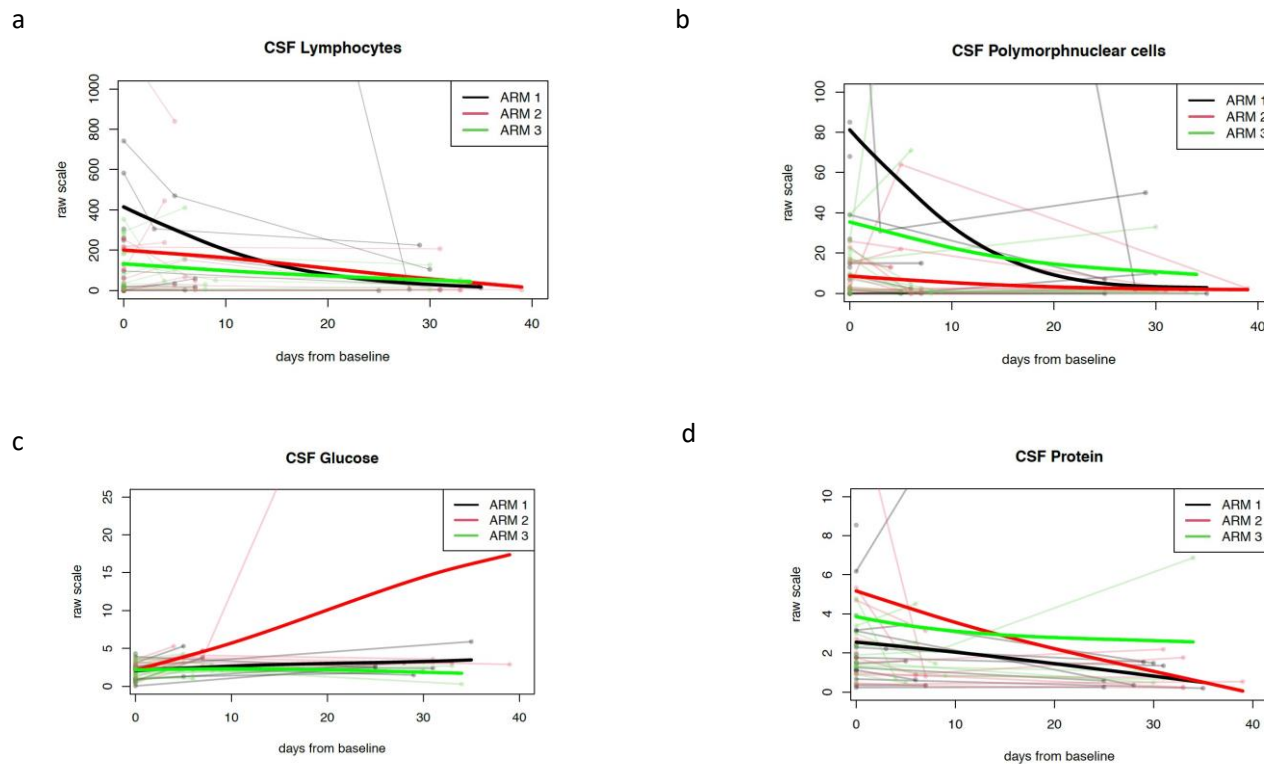
1072 A comparison between good outcome (MRS 0-3) vs bad outcome stratified by arm at day 56; 'b' compares change in MRS between  
1073 enrolment and day 56, across treatment arms.

1074

1075

1076 **Figure 3.6 Change in CSF parameters over time**

1077



1078

1079 Spaghetti plots for CSF parameters (lymphocyte count (a), polymorphonuclear cells (b), glucose (c), protein (d)) plotted as individual  
1080 values over time (faint lines), with mean values for each treatment arm represented by superimposed line (bold lines).

1081 **Table 3.1 Adverse events of special interest (AESI) assessed in**  
 1082 **LASER-TBM**  
 1083

<b>AESI</b>	<b>Investigational product</b>	<b>Objective measure</b>
Gastrointestinal haemorrhage	Aspirin	Clinical and laboratory measures to suggest haemorrhage
Intracerebral haemorrhage	Aspirin	Radiological evidence of haemorrhage
Transaminitis	Rifampicin	ALT, bilirubin (DAIDS criteria, Grade 3 and 4)
Anaemia	Linezolid	Hb (DAIDS criteria, Grade 3 and 4)
Neutropenia	Linezolid	Neutrophils (DAIDS criteria, Grade 3 and 4)
Thrombocytopenia	Linezolid	Plt count (DAIDS criteria, Grade 3 and 4)
Peripheral Neuropathy	Linezolid	1 grade increase on the BPNS and/or a 2 grade change in any modality on the mTNS
Change in LogMAR score (visual acuity)	Linezolid	Change of 0.2 on the LogMAR/Tumbling E Chart

1084  
 1085 ALT, alanine transaminase; DAIDS, Division of AIDS; BPNS, Brief  
 1086 Peripheral Neuropathy Score; mTNS, modified Total Neuropathy Score.  
 1087  
 1088  
 1089



**Table 3.2 Baseline demographics and clinical characteristics in LASER-TBM**

	Arm 1 (n=20)	Arm 2 (n=15)	Arm 3 (n=16)
Age, median (IQR) years	39.5 (34-48.5)	37 (34.5–42.5)	41.5 (31.8-46)
Gender, male n (%)	10 (50)	10 (66.7)	16 (62.5)
Uniform case definition, n (%)			
- Definite	8 (40)	3 (20)	6 (37.5)
- Probable	5 (25)	4 (26.7)	4 (25.0)
- Possible	7 (35)	8 (53.3)	6 (37.5)
BMRC TBM Grade, n (%)			
- Grade 1	11 (55)	11 (73.3)	8 (50)
- Grade 2	8 (40)	4 (26.7)	8 (50)
- Grade 3	1 (5)	0 (0)	0 (0)
CD4 T-cell count, median (IQR), cells/μL	116.5 (58.6-283)	131 (82.5-186)	158.5 (85.5-331.5)
HIV viral load, median (IQR), copies/mL	89,150 (1000-203,711)	37,960 (2428 – 394,839)	2686 (1361-777,620)
ART status, n (%)			
- On ART	6 (30)	5 (33)	5 (31)
- Previous ART	3 (15)	6 (40)	5 (31)
- ART naive	11 (55)	4 (27)	6 (38)
Of those on ART, duration in weeks, median (range)	288.9 (22.4-459.3)	23.7 (0.4-83.6)	355 (2.9-879.1)
CSF cell count/biochemical data available (n)	17	14	13
Polymorphonuclear cells, median (IQR), cells/ μL	13 (0-85)	4 (2-16)	16 (3-22)
Lymphocytes, median (IQR), cells/ μL	63 (10-259)	79 (11-218)	82 (28-278)
Protein, mg/dL	1.78 (1.13-3.13)	1.89 (0.95-4.2)	1.9 (1.32-2.99)
CSF Glucose, mg/dL	2.2 (0.9-2.5)	2.4 (1.9-2.9)	1.7 (1.2-3.3)
Baseline radiology available (n)	16	12	11

Hydrocephalus n (%)	1 (6.3)	1 (8.3)	1 (9.1)
Meningeal enhancement n, (%)	4 (25)	2 (16.7)	6 (54.5)
Tuberculoma(s) n, (%)	1 (6.3)	2 (16.7)	2 (18.2)
Infarct(s) n. (%)	4 (25)	1 (8.3)	3 (27.3)

Abbreviations: IQR, interquartile range; ART, antiretroviral therapy; BMRC, British Medical Research

**Table 3.3 AESI stratified by treatment arm**

	<b>Arm 1 (n=20)</b>	<b>Arm 2 (n=14)</b>	<b>Arm 3 (n=16)</b>	<b><i>p value</i> **</b>
Bleeding, n* (%)	0 (0)	0 (0)	1 (6)	0.338
Transaminitis, n (%)	0 (0)	0 (0)	2 (13)	0.109
Hematological, n (%)	2 (10)	0 (0)	1 (6)	0.481
Peripheral Neuropathy, n (%)	2 (10)	2 (14)	4 (25)	0.46
Change in LogMAR score, n (%)	0 (0)	2 (14)	2 (13)	0.231

\*individuals with an event

\*\* arm 3 vs arm 1

**Table 3.4 Details of AESI by event**

AE name	Treatment arm	Days of treatment	DAIDS grade	Pre-existing	Outcome
Melaena	3	1	1	No	Two episodes of black stool. No associated change in Hb or urea. Aspirin stopped and not restarted as per protocol. No further events.
Transaminitis	3	16	3	No	Improved to grade 2 but not restarted on high-dose rifampicin at discretion of site PI.
Transaminitis	3	6	4	No	Improved, successfully rechallenged with rifafour FDC. High-dose rifampicin not restarted per protocol.
Neutropenia	1	13	3	Yes	No change in study medication (arm 1)
Neutropenia	3	28	3	No	Linezolid stopped, resolved. Not restarted.
Anemia	1	28	3	No	No change in study medication (arm 1)
Neurosensory symptoms	1	10	2	No	No change in study medication (arm 1). Normal at subsequent visit.
Neurosensory symptoms	1	6	2	No	No change in study medication (arm 1). Normal at subsequent visit.
Neurosensory symptoms	2	3	1	No	Linezolid stopped. MRI show anterior cord changes (possible ischaemic or inflammatory aetiology). Not restarted on linezolid although felt clinically not to be consistent with peripheral neuropathy.
Neurosensory symptoms	2	7	1	No	All study medication stopped due to relocation of participant and therefore withdrawal from study. No follow up BPNS performed.
Bilateral lower limb weakness	3	18	2	No	Linezolid stopped. MRI showed changes consistent with TB radiculopathy//arachnoiditis. Linezolid not restarted at discretion of site PI , although clinically unlikely peripheral neuropathy.
Paresthesia left leg	3	18	1	No	
Neurosensory symptoms	3	3	1	No	Linezolid stopped. Normal at subsequent visit although linezolid not restarted at discretion of site PI.
Neurosensory symptoms	3	10	1	No	Linezolid stopped. Participant subsequently died, cause of death not related to linezolid.

Asymptomatic increase in BPNS	3	13	1	No	Linezolid stopped. Normal at subsequent visit. Linezolid restarted at 600mg as per protocol.
Increase in LogMAR	2	56	1	No	Noted on day 56 visit, therefore no change in study medication. No follow up notes.
Increase in LogMAR	2	55	1	No	Optic neuropathy ruled out by ophthalmology. Linezolid restarted.
Change in visual acuity with related change in LogMAR	3	42	4	No	Seen by ophthalmology. Diagnosis: Parietal stroke +/- ethambutol related optic neuropathy.
Increase in LogMAR	3	55	1	No	Noted on day 56 visit, therefore no change in study medication. No follow up notes.

Abbreviations: Hb; haemoglobin; FDC, fixed dose combination; BPNS, Brief Peripheral Neuropathy Score; LogMAR, Logarithm of the Minimum Angle of Resolution.; MRI, Magnetic Resonance Imaging; PI, principal investigator.

**Table 3.5 Timing and cause of death prior to day 56**

Cause of death	Treatment Arm	Days of IP
Renal Failure	1	11
TB Meningitis	1	15
TB Meningitis	1	2
TB Meningitis	2	8
TB Meningitis	2	0*
Pulmonary embolism	3	39
TB Meningitis	3	6
TB Meningitis	3	3

\*death prior to receiving study IP and therefore excluded from modified intention to treat population analysis

**Table 3.6 Reasons for screening exclusion**

Reason*	n	%
Chronic aspirin or NSAID use	9	13.2
HIV uninfected	8	11.8
Investigator discretion	8	11.8
Received more than 5 days of anti-TB treatment in 30 days prior to enrolment	7	10.3
Not 'possible', 'probable' or 'definite' TBM	6	8.8
Haemoglobin < 8 g/dL	5	7.4
INR > 1.4	4	5.9
No consent for enrolment given by the patient	3	4.4
Peripheral neuropathy scoring Grade 3 or above on modified BPNS	3	4.4
Standard TB treatment contraindicated	2	2.9
Known rifampicin resistance during this episode	2	2.9
Previous drug resistant TB	2	2.9
Uses a 'disallowed medication' that cannot safely be stopped	2	2.9
Pregnant (>17 weeks at baseline)	1	1.5
Allergy to RHZE, LZD, aspirin	1	1.5
Died before enrolment	1	1.5
Evidence of bacterial or cryptococcal meningitis	1	1.5
eGFR < 20	1	1.5
Platelet count < 50 10 <sup>9</sup> /L	1	1.5
Relocation prior to enrolment	1	1.5

Abbreviations: NSAID, non-steroidal anti-inflammatory drug; BPNS; brief peripheral neuropathy score; Rif, rifampicin; RHZE; rifamur; LZD, linezolid; ASA, aspirin

\*more than one reason can be assign to each participant

**Table 3.7 Reasons for study withdrawal prior to day 56**

Withdrawal	Reason
1	Participant relocated to Malawi and therefore unable to attend study follow up visits
2	Participant relocated to Kwazulu-Natal and therefore unable to attend study follow up visits
3	Participant withdrew consent
4	Participant withdrew consent
5	Participant developed acute psychosis and was unsafe to follow up
6	Participant lost to follow up

# Chapter 4. Cognitive Impairment in Tuberculous Meningitis

## 4.1 Introduction

Tuberculous meningitis (TBM) affects approximately 100,000 people per year (Wilkinson et al., 2017). Cognitive impairment is often referred to as a common complication in TBM, however only four studies have reported its frequency. Two of these used brief screening tests rather than comprehensive cognitive batteries (Ranjan et al., 2003, Kalita et al., 2007), and another gathered information on cognition from the clinical history alone (Anderson et al., 2010). The only study to undertake more comprehensive cognitive testing assessed a small group of 17 TBM participants (Chen et al., 2015). No studies have systematically evaluated the frequency of cognitive impairment following TBM including affected cognitive domains or described the effect of cognitive performance on functional outcome. Moreover, no published studies have compared findings to an appropriate control group, considered the contribution of HIV co-infection or non-central nervous system (CNS) TB disease, or been undertaken in an African setting where TBM is endemic.

Better understanding of cognitive impairment in TBM is crucial for several reasons. Firstly, objective measures of cognition are a quantitative measure of clinical outcome which are infrequently used in clinical trials in adult TBM. By contrast, paediatric studies routinely consider these as part of a neurodevelopmental outcome assessment (Davis et al., 2019a).

Developing an accessible battery of cognitive tests which taps into



cognitive domains known to be impaired in TBM, for use in TBM endemic settings, would improve the precision of measurable outcomes for TBM studies. Secondly, recent TBM pathogenesis studies have unveiled mechanisms of brain injury such as the upregulation of neuroexcitatory pathways (Rohlwink et al., 2019) and release of damage associated proteins also seen in neurodegenerative conditions (Herbst et al., 2019). These findings must encourage us to better understand whether cognitive impairment leads to longer-term disability in TBM. In particular, whether cognitive impairment is focal, attributable to discrete structural abnormalities in the brain e.g. stroke or tuberculomas and/or whether there is a clinical presentation in keeping with a diffuse cortical or subcortical process at play. Most importantly however, understanding cognitive and functional impairment in TBM, particularly its effect on treatment adherence, will improve the long-term care of patients with this condition; including the provision of appropriate resources to aid recovery of these individuals following TBM.

In a case-control study of HIV-associated TBM we aimed to:

- i) Evaluate the frequency and nature of cognitive impairment in HIV-associated TBM using formal cognitive testing alongside physician assessment;
- ii) Assess the pattern of impairment, and correlate with radiological and neurological measures, to understand whether cognitive impairment relates to focal brain injury, diffuse inflammation, or a combination of these;
- iii) Assess the suitability of currently available screening tools to identify cognitive impairment;

iv) Measure the impact of impaired cognitive performance on functional outcomes, including treatment adherence.

## **4.2 Methods**

### **Participants**

We drew participants from three parent studies that took place in Cape Town between 2015 and 2020 (Table 4.1). All studies were performed in similar adult populations in a low-income, peri-urban area of Cape Town, South Africa, with high HIV and TB prevalence.

Participants formed three groups:

- i) HIV-associated TBM cases (from the LASER-TBM and Albertyn studies).
- ii) Comparator group 1: PLWH, no history of TB (from the CONNECT study).
- iii) Comparator group 2: PLWH, non-CNS TB (from the Albertyn study).

Normative cognitive data was drawn from HIV-negative individuals selected to be demographically similar to PLWH in the CONNECT study as described in the statistical methods below.

The studies were each approved by the University of Cape Town's Faculty of Health Sciences Research Ethical Committee (Table 4.1).

Sample size was pragmatic bringing together two unpublished data sets of HIV-associated TBM patients (cases), and non-CNS TB patients

(comparator group 2). The decision to include PLWH (comparator group 1) at a 1:2 ratio was made to increase statistical power for this analysis.

### **Outcome measures and procedure**

Assessment took place at 6 months following diagnosis of TBM or non-CNS TB, or at the time of enrolment in comparator group 1. Timing of follow up was dictated by the umbrella studies from which these participants were recruited.

### **Baseline assessments**

We graded severity of TBM at baseline using the modified British Medical Research Council (BMRC) scale (Thwaites and Tran, 2005), and classified TBM as 'definite', 'probable' or 'possible' as per the uniform case definition for research (Marais et al., 2010). We collected data on educational history, drug and alcohol use; in the LASER and CONNECT cohorts the Alcohol Use Disorders Identification Test (AUDIT) (Barbor, 2001) (cut-off  $\geq 20$ , 'high risk') and Drug Use Disorders Identification Test (DUDIT) (Berman and <https://doi.org/10.1037/t02890-000>) (cut-off  $\geq 6$  (men),  $\geq 2$  (women)) questionnaires were used to ascertain alcohol and drug use respectively. Within the Albertyn cohort, active alcohol or illicit drug use was an exclusion criterion.

### **Cognitive testing**

Cognitive testing was performed in the participants' first language in all cases. An identical cognitive test battery was administered across the three studies to assess 10 measures in 7 cognitive domains: motor skills, processing speed, attention and working memory, fluency, audioverbal and visuospatial learning and memory and executive function (see

Supplementary Material Box 1 published alongside (Davis et al., 2022a)). This is based on a battery with established psychometric validity in South Africa (Nyamayaro et al., 2019). Participants from LASER-TBM also underwent a comprehensive neurological examination, performed by a trained physician, to identify focal cortical syndromes. This included assessment for motor, sensory and cranial nerve abnormalities, language deficits, apraxia, visual agnosia, and right hemisphere dysfunction (visuospatial deficit, anosognosia, sensory neglect). The physician assessment was performed in English with where necessary an isiXhosa translator.

### **Imaging**

Within the LASER cohort we correlated results from cognitive assessment with computed tomography (CT) brain scans ( $\leq 2$  months since diagnosis) reported by an independent blinded neuroradiologist.

### **Mental health assessment**

All participants completed a Beck Depression Inventory ((BDI) (Upton, 2013) (cut-off 'depression'  $>17$ ) or CES-D (Radloff, 1977) (cut-off 'depression'  $\geq 16$ ), to assess mood.

### **Functional measures**

We administered the Patient's Assessment of Own Functioning Inventory (PAOFI) (Chelune et al., 1986) in all participants recruited to LASER-TBM. We calculated total score, and total number of responses with 'affirmative responses' as previously described (Woods et al., 2004, Bell et al., 2013). Lower total PAOFI scores indicate higher levels of functioning;  $\geq 3$  'affirmative' responses have been previously described as a cut-off for functional impairment. Within LASER-TBM we collected data on treatment adherence for the first 56 days of treatment. Self-reported adherence was assessed by asking participants at each visit if they had missed any doses

since their last visit. Observed adherence was assessed by totalling number of missed doses noted on pill count at each study visit.

### **Cognitive screening measures**

Two screening measures i) Montreal Cognitive Assessment (MoCA) (Nasreddine et al., 2005) (cut-off indicating impairment  $\leq 26$ ) and ii) CatRAPID v2.0 (Joska et al., 2016) (cut-off indicating impairment  $\leq 16$ ) were performed in participants recruited to LASER-TBM.

### **Statistical analysis**

We used R v.4.1.2(2021-11-01), RStudio v.2021.09.0 and GraphPad Prism v.9.3.1, to complete all analyses, with the threshold for statistical significance set at  $\alpha=0.05$ .

First, we processed and standardized the cognitive test battery data. Normative scores for the cognitive tests were based on healthy control data collected by the CONNECT study. These data were collected between 2018 and 2020 from healthy HIV-negative community-dwelling individuals who presented to the same community health clinics in Gugulethu from which the PLWH comparator group was recruited, within the same area of Cape Town where TBM and TB participant recruitment took place. They therefore shared key demographics (age, ethnicity, language, education), as well as psychosocial and socioeconomic characteristics. We used the control data to calculate demographically corrected z-scores (mean [M] = 0, standard deviation [SD] = 1), using standard regression-based norming processes. The z-scores were then converted to demographically corrected T-scores (M=50, SD=10). If participants had z-scores  $>5$  SD below the mean, the conversion to a T-score resulted in a negative T-score. In these

cases, we assigned a score of 0, the lowest possible T-score to maintain the clinical significance of the low performance. Cognitive performance data were summarised into domain and global T-scores by taking the average of T-scores within each domain and then across domain T-scores. T-scores were converted to deficit scores (See Supplementary Material Box 2 published alongside (Davis et al., 2022a)). The overall global deficit score (GDS) was calculated by averaging deficit scores. A cut-off GDS of  $\geq 0.5$  has been considered consistent with 'cognitive impairment' on cognitive test performance (Carey et al., 2004); for the purposes of this study we termed this group as having 'low performance on cognitive testing' whilst the clinical significance and functional impact in TBM is further explored, aligned with recent trends in the HIV literature (Nightingale et al., 2021)

Second, we used the Matchit package in R (<https://cran.r-project.org/web/packages/MatchIt/vignettes/MatchIt.html>) to select comparator group 1 (PLWH, no history of TB) from the parent study CONNECT with the ratio 1:2 with matched co-variants (age, gender, education) to TBM cases so distributions of covariates in the two groups were approximately equal. We subsequently processed the cognitive test data for this group as described above.

We checked normality of distribution of data with a Shapiro-Wilk test. We determined between group differences in baseline and clinical variables using either parametric T tests or Mann-Whitney (for continuous variables where data was normally or not normally distributed, respectively), or chi-squared tests (for dichotomous variables). We performed a primary comparison of cognitive test performance between HIV-associated TBM cases and comparator group 1, and subsequently HIV-associated TBM

cases and comparator group 2; we did not perform an ANOVA given that our primary aim was to evaluate differences between TBM cases and each comparator group individually, rather than differences between the comparator groups or across the three groups. We compared total PAOFI scores using a Mann-Whitney test of significance in HIV-associated TBM participants with low cognitive performance vs those without. We compared adherence between those with and without low cognitive performance by i) self-report of missing  $\geq 1$  dose with Fishers Exact, and ii) observed adherence from number of missed doses on pill count within the first 56 days of treatment with a Mann-Whitney test. We used a 2x2 table of agreement to create a Cohen's Kappa value for agreement between the screening tests (cut-offs: CatRAPID  $\leq 16$ ; MoCA  $\geq 26$ ) and GDS  $\geq 0.5$  on the cognitive test battery and assigned levels of agreement as per a published scale (Landis and Koch, 1977).

### **4.3 Results**

We included 34 participants with HIV-associated TBM (cases), 66 participants with HIV and no history of TB (comparator group 1), and 26 participants with HIV and non-CNS TB (comparator group 2) (Figure 4.1). Age, gender and years of education were similar across groups (Table 4.2). IsiXhosa was the first spoken language in 119/126 (94%); in 2/126 (1.6%) it was English and in 3/126 (2.4%) it was Shona. This information was missing in 2 cases. All HIV-associated TBM cases were either mild (BMRC 1) (18/34, 53%) or moderate (BMRC 2) (16/34, 47%) at presentation. Within LASER-TBM, 2/19 participants reported head injury resulting in loss

of consciousness, 2 participants had 'high risk' alcohol use, and 0 participants had 'high risk' drug use. These were exclusions in the CONNECT and Albertyn studies.

In those with HIV-associated TBM, 16/34 (47%) of participants had low performance on cognitive testing consistent with cognitive impairment ( $GDS \geq 0.5$ ). When compared to comparator group 1 (PLWH, no history of TB), those with TBM had worse global T scores (mean score 41 vs 48,  $p < 0.0001$ ), with a greater proportion of those within the TBM group meeting the GDS cut-off  $\geq 0.5$  (16/34 (47%) vs 17/66 (26%),  $p = 0.032$ ). Domain specific T scores for all cognitive domains were significantly worse in the TBM group, with the exception of attention and working memory (Table 4.3A). When compared to comparator group 2 (PLWH, non-CNS TB), global T scores were also worse in those with TBM (mean score 41 vs 46,  $p = 0.016$ ). Although a greater proportion of those with TBM met the GDS cut-off for cognitive impairment ( $\geq 0.5$ ) (16/34 (47%) vs 8/26 (31%)), this was not statistically significant ( $p = 0.201$ ) (Table 4.3B). Global T scores were worst in those with HIV-associated TBM (cases), better in those with HIV and non-CNS TB (comparator group 2), and best in those with HIV alone (comparator group 1) (Figure 4.2). Radiological assessment (either CT, MRI, or both) was performed in 16/19 participants included in LASER-TBM who had imaging performed either as part of the study or for a clinical indication at presentation. 7/16 (43%) of participants had abnormal imaging; 7/16 (43%) had meningeal enhancement and 4/16 had stroke (33%). 1 participant had multiple calcified granulomas, and another hydrocephalus. In the 4 participants with stroke, 3 had a GDS consistent with low cognitive performance. In these participants the individual domain T scores were reviewed by a clinical neuropsychologist (ADr) and



neurologist (AD) who concluded that the profile of abnormality across the T scores could not be explained by the anatomical location alone. Although we believe that in these cases the infarcts are likely to be contributing to the burden of cognitive impairment, it was felt that the scores represented a more global picture in each of the cases. There was no statistically significant difference in the number of those found to have abnormal imaging in those with low cognitive performance versus those without (6/10 vs 2/6,  $p=0.6084$ ). Physician assessment did not reveal any clear cases where focal motor and sensory deficits correlated to impairment in a discrete corresponding cognitive domain e.g. due to focal stroke or tuberculomas.

The mean score for CATRAPID v2.0 and MoCA screens were performed in 19 TBM participants (the LASER-TBM cohort) and the means (SD) were 16 (3.15) and 21 (3.70) respectively. Using a cut-off of  $\leq 16$  for CATRAPID and  $\leq 26$  for MoCA, 7 (34%) and 17 (89%) participants respectively would have been flagged as having mild cognitive impairment using these screening tests. A Kappa value of 0.242 (95% CI -0.179-0.661) for CatRAPID and 0.137 (95% CI -0.115-0.389) for MoCA equated to 'fair' and 'slight' agreement respectively when comparing these measures to a  $GDS \geq 0.5$  on the cognitive test battery.

Proportionally more participants had depression in the TBM group than the comparator group 1 (8/34 vs 6/66,  $p=0.049$ ); whereas no difference was found between the TBM cases and comparator group 2 (8/34 vs 5/26,  $p=0.76$ ). In the sub-group of participants with TBM where PAOFI was completed ( $n=19$ ) we found that lower cognitive performance in TBM cases was associated with better functional status (mean (SD) of PAOFI score in

patients with low cognitive performance vs those without, 23.5 (14.8) vs 40.6 (19.1),  $p=0.042$ ). When comparing a cut-off of 3 or more affirmative responses corresponding to 'functional impairment', the difference was not statistically significant (5/11 (45%) 'functionally impaired' in group with low cognitive performance, vs 6/8 (75%) 'functionally impaired' in group without low cognitive performance,  $p=0.352$ ). Given the unexpected finding of worse functional outcomes in those normal cognitive performance, we further explored the cases individually. Although PAOFI is a measure of functional status, the questionnaire centres around self-reporting of cognitive symptoms; it is therefore plausible that low mood, lack of insight and premorbid status may affect reporting of cognitive functioning. We identified one case where performance on the cognitive test battery was severely impaired (GDS 2.85), yet PAOFI scores were low (affirmative score 0, total score 18) suggesting lack of insight and underreporting of impairment. We identified two cases where the BDI suggested clinical depression and PAOFI scores were high (i.e. high burden of cognitive symptoms), yet performance on the cognitive test battery was within normal limits suggesting over-reporting of symptoms associated with low mood. In another case, PAOFI scores were high (affirmative 12, total 71), yet GDS suggested cognitive performance within the low-normal range (GDS 0.428). This individual however had high pre-morbid functioning (completed 12 grades education) and therefore a drop in cognitive functioning may not have been identified using a GDS cut-off of  $\geq 0.5$ .

Within the LASER-TBM cohort, 6/11 (55%) of those with low cognitive performance reported missing medication doses compared to 3/8 (38%) of those with normal cognitive performance. Mean number of missed doses was 2.72 in those with low cognitive performance compared to 0.37 in

those normal cognitive performance. Neither difference was statistically significant.

## **4.4 Discussion**

In this case-control study of HIV-associated TBM we evaluated the frequency and nature of cognitive impairment and functional outcomes, including treatment adherence. Within our study, almost half of participants with HIV-associated TBM demonstrated low performance on cognitive testing. This was a significantly greater proportion of individuals when compared to an age, gender and education matched comparator group of participants with HIV alone; demonstrating that impairment seen in HIV-associated TBM is in addition to that attributable to HIV alone. We also showed that participants with HIV-associated TBM perform worse than those with HIV and non-CNS TB suggesting that low cognitive performance in TBM is due to CNS-specific pathology in addition to any other mechanisms at play in non-CNS TB disease such as systemic inflammation and polypharmacy (Kass and Shandera, 2010, Annane and Sharshar, 2015).

Low cognitive performance was seen across all cognitive domains in participants with TBM when compared to those with HIV only, with the exception of attention and working memory. Within our cohort we did not see cases where motor/sensory deficits on neurological examination and radiological findings correlated with a single focal cognitive deficit. These findings begin to characterise low cognitive performance in TBM as generalised, affecting multiple cognitive domains which are, at least in our

cohort, beyond what is attributable alone to structural deficits e.g. stroke. The predominant motor impairment suggests subcortical damage, and the relative sparing of attention and working memory suggests that ongoing delirium is unlikely to explain our findings. These observations also highlight the limitations of CT as an imaging modality in identifying changes such as cortical and subcortical inflammation and microvascular damage which may present clinically with generalised cognitive deficits. These are important findings when considering i) pathogenic mechanisms in TBM and ii) suitable imaging techniques for identifying those at risk of impairment.

In the subgroup where PAOFI was administered, the finding of better self-reported functional status in those with low cognitive performance compared to those with normal cognition was unexpected. Assessment of individual cases suggested examples where functioning may have been under-reported due to lack of insight, over-reported due to low mood, or not reflected in the GDS cut-off due to higher pre-morbid functioning. These examples illustrate the complexity of measuring cognitive and functional performance in diverse populations, and the potential limitations of dichotomised cut-offs for impairment based on cognitive performance alone. More work is needed to explore the association of cognitive performance with clinical indicators of cognitive impairment, functional outcomes and measures of brain injury; an area currently being explored in the field of cognitive impairment in PLWH (Nightingale et al., 2021). Future studies may consider including i) observer accounts of functional status (eg DECO (Ritchie and Fuhrer, 1996)) to add clarity where other factors e.g. depression may bias self-reporting of functional status and ii) more familiar functional outcome measures in this setting (e.g. Modified Rankin Score) in

order to contextualise the results of more comprehensive measures for the treating physician.

We found no statistically significant difference in self-report or objective measures of treatment adherence between those with and without low cognitive performance, which may be due to the small number of participants where this data was available (n=19). Medication non-adherence is the major cause of poor outcome in TBM, hence this potential association should be further explored in larger cohorts as it may provide new avenues to address adherence and improve outcomes in this group.

There are limitations to this study. Firstly, although all cognitive test batteries were administered by trained individuals, these were different across the three studies. Although training and alignment was overseen by the same neuropsychologist (ADr) to minimise interrater variability we acknowledge the possibility that subtle differences in administration may have influenced outcomes. Along these lines, in 4 of the 7 domains only one measure was used across all three studies and therefore included within the analysis, when ideally >1 measure should be used for each domain. This may explain the unexpected finding of preserved attention and working memory despite this being an early indicator of pathology in many sub-cortical dementias. Secondly, there were differences in the timing of investigations related to HIV status across the three studies; specifically, CD4 counts were collected at different time points (within a range of 12 months) in relation to the cognitive testing making them incomparable between groups. Given the nature of recruitment (inpatient recruitment for unwell TBM subjects, vs outpatient recruitment for well PLWH comparator group) it is highly likely that HIV disease was better

controlled within comparator group 1 than in those with TBM. Nonetheless this does not explain the finding of ~50% low cognitive performance within the TBM group (with no difference in HIV control between those with low cognitive performance and those without) and it is unlikely to entirely explain the large differences we demonstrated between the TBM and PLWH groups. Also, given that CD4 count was on average higher in those with TBM, compared to those with non-CNS TB who performed better on cognitive testing, it is unlikely that between group differences in cognitive performance is likely explained by HIV control alone. Such differences, together with differences in cognitive impairment prior to the development of TBM could be addressed within a prospective study design which should be considered in future research within this field. Thirdly, the lack of TBM cases with severe (BMRC Grade 3) disease may suggest that the frequency of low cognitive performance is underestimated. In LASER-TBM recruitment of stage 3 disease was infrequent, and none of the participants followed up with full cognitive testing had stage 3 disease at baseline. Similarly, no participant included within the Albertyn study had stage 3 disease. Inclusion of participants fortunate enough to survive stage 3 disease should be included within future studies to ensure generalisability of results. Finally, our findings of low cognitive performance at 6 months following TBM diagnosis should not be interpreted as a finding of long-term disability in this population. Longer-term follow up studies are needed to understand whether cognitive performance improves or worsens in the years following active illness. These studies must incorporate detailed assessment of neurobehavioural functioning to understand whether low cognitive performance translates to clinically apparent cognitive impairment following TBM. This is particularly timely given emerging data from *ex-vivo* studies implicating pathogenic mechanisms such as neuroexcitotoxicity

leading to neuronal injury (Rohlwink et al., 2019); mechanisms also described in neurodegenerative conditions (Wang and Reddy, 2017, Barkhoudarian et al., 2016).

Although this study provides important characterisation of neurological sequelae in TBM, it may not be feasible to adopt full cognitive testing in TBM endemic settings. We found only 'fair' or 'slight' agreement between CatRAPID and MoCA screens and the cognitive test battery when using standard cut-offs. The MoCA characterised most participants as cognitively impaired and may not be appropriate for this setting; this tool was developed and normed for a North American population, whereas in a low-income peri-urban South African population one study found the mean score in cognitively unimpaired, healthy controls was 21.7/30 (Robbins et al., 2013). A Receiver Operating Characteristic analysis is the best method to establish disease-specific cut-offs however we were underpowered to perform this. Given the high frequency of low cognitive performance within our TBM cohort, a larger study to validate and/or adapt cut-offs for impairment in existing screening tools within the TBM context is required, so that potential impairment can be identified where resources are limited. Where resources are available, our results highlight the value of including detailed cognitive and functional outcome assessments as quantitative measures of clinical outcomes within clinical trials in adult TBM.

In summary, our study demonstrates for the first time in adults that low cognitive performance in TBM occurs in approximately half of participants and is characterised by generalised impairment, affecting multiple cognitive domains. We also demonstrate that low cognitive performance in TBM is independent of and additional to the effects of HIV and non-CNS TB

disease. TBM occurring in people without HIV co-infection may have different cognitive sequelae. TBM has different clinical and neuropathological characteristics in PLWH compared to those without HIV (Marais et al., 2011, Cecchini et al., 2009). In addition, PLWH may have underlying HIV-associated brain injury which could decrease cognitive reserve and increase vulnerability to cognitive impairment from TBM neuropathology. A study of TBM cognitive outcomes in those without HIV co-infection would provide further insight into the burden of cognitive impairment in this disease in a different context and would be an important comparison for future studies. Future work is now needed to evaluate outcomes at longer-term timepoints, describe the relationship between cognitive performance, functional status and treatment adherence and validate sensitive context-specific screening tools to identify individuals at risk, in order to improve outcomes for patients with TBM.



**Table 4.1 Parent Studies included within the study of cognitive impairment in TBM**

	<b>LASER-TBM</b>	<b>Albertyn Study</b>	<b>CONNECT</b>
Study design	Phase 2a randomized, open label clinical trial of intensified antibiotics and high dose aspirin in HIV-associated TBM(Davis et al., 2021)	Prospective case control study evaluating cognitive and functional impairment in HIV-TBM (control group of other forms [non-CNS] TB)	Prospective case control study evaluating cognition, neuropsychiatric symptoms and neuroinflammation in PLWH switching from efavirenz to dolutegravir
Ethical approval	UCT HREC 293/2018	UCT HREC 565/2014	UCT HREC 017/2019
N recruited	52 enrolled	27 TBM 25 controls (non-CNS TB)	180 HIV +ve participants 60 HIV –ve controls
Setting	Inpatient	Inpatient	Outpatient
Inclusion criteria	Adults with TBM (definite, probable, possible) Confirmed diagnosis of HIV	Adults with TBM (definite, probable) Confirmed diagnosis of HIV	180 PLWH studied before and after switch from efavirenz to dolutegravir
Exclusion criteria	Many exclusions related to RCT (see (Davis et al., 2021)) None related to prior head injury, neurological disease or drug/alcohol dependence	Significant prior CNS disease (stroke, opportunistic CNS infection, significant head injury, dementia) Active alcohol or substance abuse/dependence Poor socio-economic support	Excessive drug or alcohol misuse History of CNS infection (inc meningitis) Previous stroke Major head injury (loss of consciousness >30 minutes)
Follow up timepoint	6 months	6 months	Baseline and after switch assessments (6-12 months)

**Table 4.2 Baseline demographics and clinical characteristics of participants included within the study of cognitive impairment in TBM**

	HIV-associated TBM (n= 34)	PLWH (n=66)	Non-CNS TB (n=26)
Age in years (mean, SD)	36.4 (8.9)	37.4 (7.5)	35.3 (8.0)
Gender (% male)	47	41	58
Years of Education (mean, SD)	9.6 (3.1)	10.4 (1.5)	10.2 (1.6)
First language spoken: n (%)			
isiXhosa	34 (100)	59 (89)	26 (100)
English	0	2 (3)	0
Shona	0	3 (5)	0
Missing data	0	2 (3)	0
Baseline BMRC Grade			
Grade 1	18 (53)	n/a	n/a
Grade 2	16 (47)		
Grade 3	0		
Uniform Case Definition TBM Category			
Possible	7 (21)	n/a	n/a
Probable	13 (38)		
Definite	14 (41)		
HIV details (n) available in CD4 count (cells/mm <sup>3</sup> ) (mean, SD)	34* 177 (183)	41** 533 (293)	26* 88.6 (90.4)

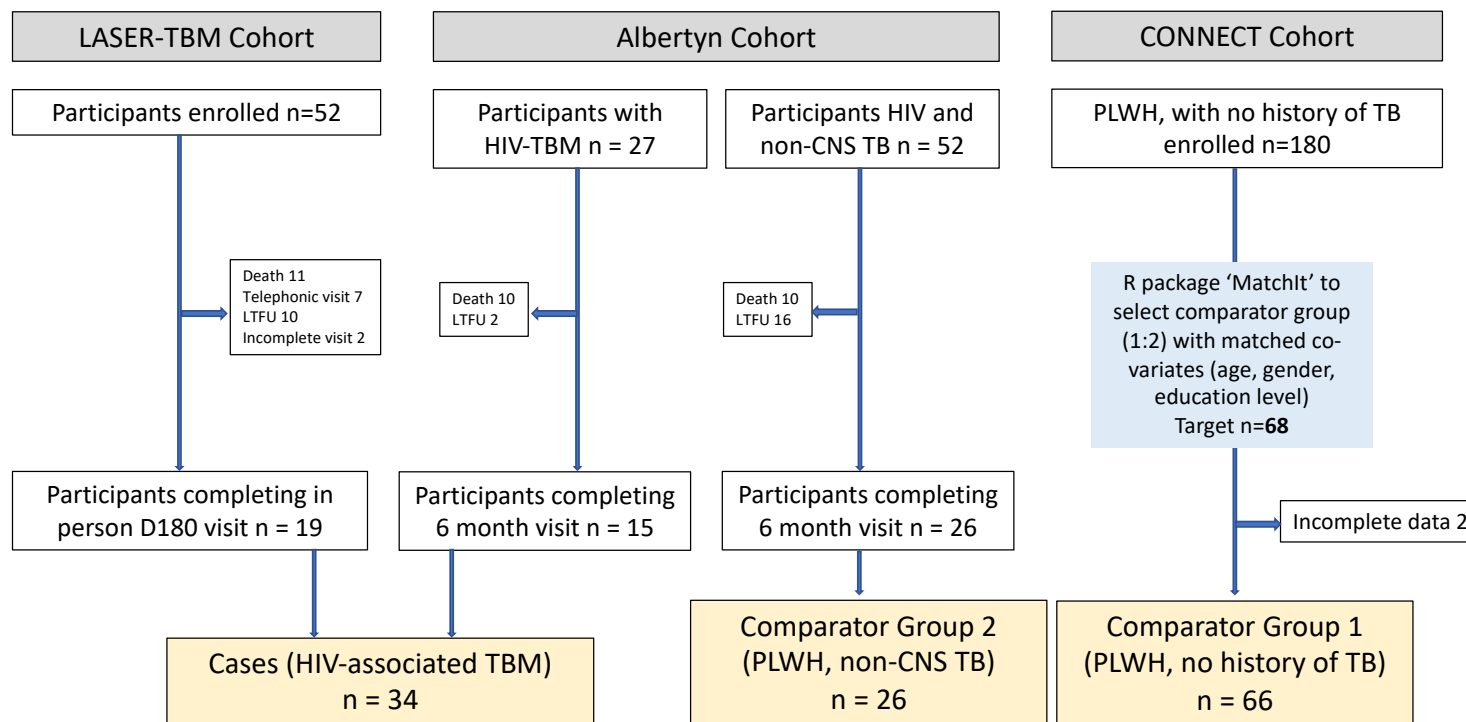
**Table 4.3 (A) Comparison of Domain specific T scores in TBM cases vs comparator group 1 (PLWH, no history of TB) and (B)**

**Comparison of Domain specific T scores in TBM cases vs comparator group 2 (PLWH non-CNS TB)**

<b>A</b>	<b>HIV-associated TBM cases (n=34)</b>	<b>Comparator group 1 (PLWH) (n=66)</b>	<b>p value</b>
<b>Domain T Score (Mean, SD)</b>			
Motor skills	38 (14)	46 (11)	<0.05*
Processing speed	36 (15)	47 (7)	<0.0001***
Attention and working memory	47 (10)	49 (9)	0.243
Fluency	45 (10)	49 (7)	0.029*
Audioverbal learning and memory	39 (15)	49 (9)	0.0004***
Visuospatial learning and memory	40 (8)	48 (10)	0.0004***
Executive function	41 (14)	47 (10)	0.036*
Global T score (Mean, SD)	41 (9)	48 (6)	<0.0001***
GDS suggesting CI (frequency, %)	16 (47)	17 (26)	0.032

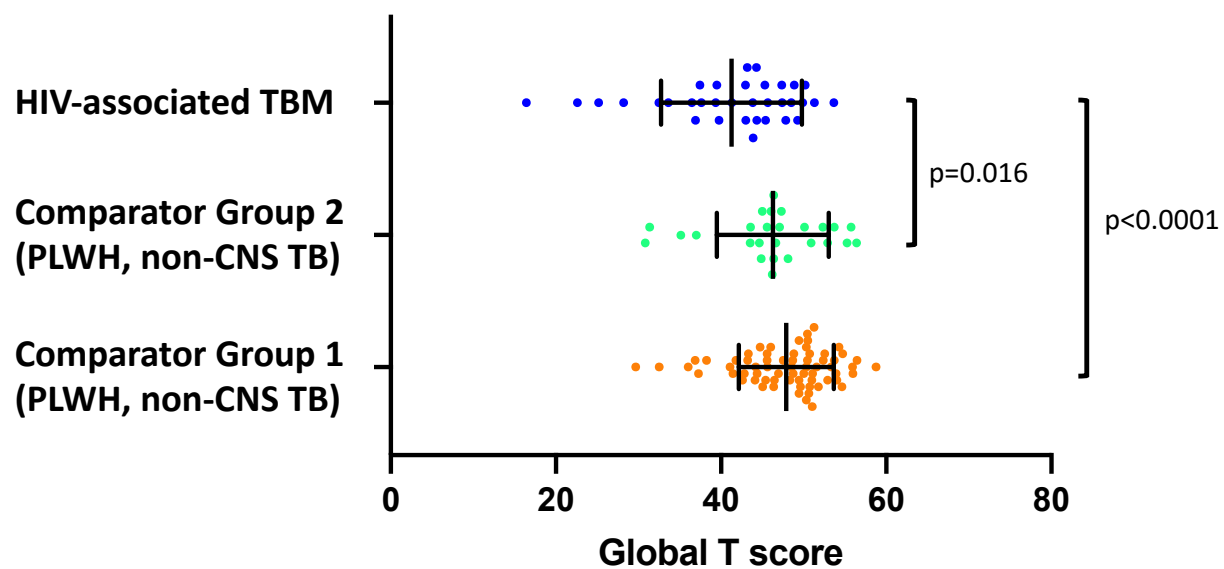
<b>B</b>	<b>HIV-associated TBM cases (n=34)</b>	<b>Comparator group 2 (PLWH, non-CNS TB) (n=26)</b>	<b>P value</b>
<b>Domain T Score (Mean, SD)</b>			
Motor skills	38 (14)	44 (8)	0.108
Processing speed	36 (15)	43 (9)	0.096
Attention and working memory	47 (10)	52 (9)	0.067
Fluency	45 (10)	48 (9)	0.205
Audioverbal learning and memory	39 (15)	43 (15)	0.273
Visuospatial learning and memory	40 (8)	46 (12)	0.039*
Executive function	41 (14)	48 (10)	0.156
Global T score (Mean, SD)	41 (9)	46 (7)	0.016*
GDS suggesting CI (frequency, %)	16 (47)	8 (31)	0.201

**Figure 4.1 CONSORT diagram for study of cognitive Impairment in TBM**



Consort diagram to describe enrolments across three parent studies

Figure 4.2 Global T Scores across groups



Scatter plot graph with mean and SD as well as individual values plotted in i) participants with HIV-associated TBM, ii) participants with HIV and non-CNS TB (comparator group 2), vs iii) participants with HIV only (comparator group 1).



## **Chapter 5. Luminex Multiplex analysis in blood and CSF of patients with HIV-associated TBM**

### **5.1 Introduction**

The use of host biomarker immunoassays, including the fluorochrome (Luminex) technique are well established in the field of tuberculous meningitis (TBM). Luminex uses a bead-based immunoassay which combines enzyme-linked immunosorbent assay (ELISA) with flow cytometry to simultaneously detect and quantify multiple proteins including cytokines, chemokines and growth factors within a single assay (Faresjo, 2014). The beads (microspheres), which have monoclonal antibodies directed against the proteins of interest, can detect up to 100 analytes simultaneously using a dual laser flow analyser (Faresjo, 2014). Given that the technology measures multiple analytes within a single well only a small sample volume (<25uL) is required. Together with time and cost saving, these features make Luminex preferable to running simple analyte ELISA to detect a large range of inflammatory mediators.

Several studies have harnessed Luminex multiplex platform technology to describe the profile of cytokine responses in the context of neurological sequelae or discover potential diagnostic biomarkers in TBM. In a study of TBM related immune reconstitution inflammatory syndrome (IRIS) all analysed mediators were upregulated in patients going on to develop TBM-IRIS compared to those who did not; implicating both innate and adaptive

immune cellular responses in TBM-IRIS (Marais et al., 2014). Further analysis of selected neutrophil-associated mediators including IL17-A and S100A8/A9 suggested that neutrophil associated inflammation may be a key driver of pathology in TBM-IRIS (Marais et al., 2014). In paediatric TBM, multiplex platform analysis identified a 3 biomarker signature (VEGF, IFN $\gamma$  and MPO) which showed a sensitivity and specificity of up to 91.3% and up to 100% in the diagnosis of TBM (Manyelo et al., 2019a, Visser et al., 2015); which has since been reproduced in a larger validation cohort (Manyelo et al., 2022). Further sub-group analysis of this cohort identified significantly higher levels of sVCAM-1, MMP-1, sRAGE, and IP-10/CXCL10 in the CSF of children with stroke compared to those without, providing potential insight into the biology of stroke in TBM, as well as identifying potential biomarkers who may require preventative management (Manyelo et al., 2021). Although a highly compartmentalised immune response is a feature in studies using matched CSF and plasma samples (Marais et al., 2014, Yang et al., 2014b), obtaining cerebrospinal fluid is not without complication. One study has demonstrated that a host serum 3 biomarker signature (adipsin (complement factor D), A $\beta$ 42, and IL-10) showed potential in the diagnosis of childhood TBM, with a sensitivity and specificity of 82.6% and 75.0% however larger studies are required to validate this finding (Manyelo et al., 2019b).

These studies support the rationale to use Luminex multiplex technologies to uncover pathogenic mechanisms in TBM, including HIV associated TBM. They also demonstrate the value of matched timepoint CSF and blood analysis with the goal to develop blood-based biomarkers given the complexity of obtaining samples from the site of disease in neurological conditions. Although these studies may have considered mechanistic



drivers of neurological sequelae such as HIV associated TBM-IRIS or stroke, no studies to date have applied this technology to consider mechanistic drivers of all cause poor outcome in adult HIV-associated TBM.

Using a Luminex multiplex assay we:

1. Described immune profiles in the blood and CSF of patients with HIV associated TBM and compare this to a group of patients with neurological presentations of a non-infectious aetiology
2. Identified markers associated with microbiologically confirmed disease and with poor outcome in the blood and CSF of patients with HIV associated TBM
3. Described longitudinal changes in blood and CSF over time and in relation to outcome in HIV associated TBM

## **5.2 Methods**

### **Setting and Participants**

We included within this analysis

- i) participants with HIV-associated TBM recruited to LASER-TBM (see chapter 2, LASER TBM study protocol) as our 'cases' group
- ii) participants enrolled to a study of neurological complications of COVID-19 (HIATUS-3) (see appendix HIATUS-3 results). We selected from this cohort participants where the discharge documentation diagnosis was due to a non-infectious cause (confirmed with a negative CSF cryptococcal latex agglutination test (CLAT), Gene Xpert Ultra, Gram Stain and bacterial culture) and included these participants as a 'non-infectious comparator' group.

Both studies (LASER-TBM and HIATUS-3) were approved by the University of Cape Town Human Research Ethical Committee (293/2018)

and 207/2020) and the Walter Sisulu University Faculty of Health Sciences Research Ethical Committee (012/2019 and 031/2020) respectively.

### **Procedure**

In LASER-TBM lumbar puncture was performed at study visits 3 and 28 days following enrolment (Figure 5.1). A total of 3ml of CSF was collected for immunological, cellular, proteomic and metabolomic analysis where feasible. Samples were spun at 200 x g for 10 mins at 4°C, and CSF supernatant stored separately in aliquots at -80C. By contrast, venepuncture was performed at each study visit. Where feasible up to 6ml of blood was collected for immunological, proteomic and metabolomic analysis within an EDTA tube. Within 4 hours of collection samples were spun at 500 g for 10 mins at room temperature, and the plasma aliquoted for storage at -80°C. Within the stored CSF supernatant, samples were prioritised for proteomic and metabolomic analysis, with Luminex Multiplex panel analysis undertaken where at least 200uL of CSF was remaining. In participants with available CSF, plasma from matched timepoints (day 3 and/or day 28), as well as baseline (enrolment) and latest timepoint (day 56) were selected for Luminex Multiplex panel analysis. Clinical, laboratory and radiological data was collected at study visits as described in chapter 2. Clinical outcome was assessed via the Modified Rankin Scale at the day 56 visit. We pre-specified a good and poor outcome (see statistical analysis plan, supplementary appendix) as an MRS between 0 and 3, and 4 and 6 respectively.

In HIATUS-3 lumbar puncture was performed between 3 and 7 days of hospital admission. A total of 2ml of CSF was collected for immunological, cellular, proteomic and metabolomic analysis where feasible. Samples

were frozen within 4 hours of the procedure at -80°C. At the time of this analysis, samples were thawed, spun at 200 g for 10 mins at 4°C and 200uL supernatant used for Luminex Multiplex panel analysis. Clinical, laboratory and radiological data was collected at study visits as described within the HIATUS study (see appendix). No matched timepoint plasma samples were available for analysis from this study.

### **Luminex Multiplex panel assays**

A total of 200uL of CSF supernatant collected as part of LASER-TBM was thawed, filtered, and subsequently analysed using a 65 Plex Human ProcartaPlex™ Panel (catalogue number EPX650-100650-901) as per the manufacturers guidance. Concentrations of cytokines IL-1 $\alpha$ , IL-1 $\beta$ , IL-2, IL-2R, IL-3, IL-4, IL-5, IL-6, IL-7, IL-9, IL-10, IL-12p70, IL-13, IL-15, IL-16, IL-17A, IL-18, IL-20, IL-21, IL-22, IL-23, IL-27, IL-31, IFN $\alpha$ , IFN $\gamma$ , TNF $\beta$ , MIF, TSLP, LIF, NGFF $\beta$ , GM-CSF, G-CSF and M-CSF; chemokines GRO  $\alpha$  (CXCL1), CXCL5, IL-8 (CXCL8), MIG (CXCL9), IP-10 (CXCL10), I-TAC (CXCL11), BLC (CXCL13), MCP-1 (CCL2), MIP-A  $\alpha$  (CCL3), MIP-1  $\beta$  (CCL4), MCP-3 (CCL7), MCP-2 (CCL8), EOTAXIN (CCL11), MIP-3  $\alpha$  (CCCL20), MDC, EOTAXIN-2 (CCL24), EOTAXIN-3 (CCL26), FRACTALKINE (CX3CL1); mediators of the TNF superfamily TNF  $\alpha$ , TNF RII, TWEAK, CD30, APRIL, TRAIL, BAFF; and other factors VEGF-A, HGF, MMP-1, SCF, SDF-1  $\alpha$ , FGF-2, CD40L were obtained.

Samples obtained from HIATUS-3 were processed in the same way using an 18-plex Human ProcartaPlex™ Panel (catalogue number EPX180-15837-901). Only the concentrations of the common mediators obtained for both LASER and HIATUS panels were selected for this analysis, namely:

G-CSF, IL12p70, IL-10, IL-1  $\beta$ , IL-6, IP10 (CXCL10), IL-18, IL-17A, MCP1 (CCL2).

## Statistical analysis

### We compared:

- i) Baseline (day 3) mediator concentrations in CSF of patients with HIV associated TBM and compared these with non-infectious comparators;
- ii) Baseline (day 3) mediators concentrations in Blood of patients with HIV associated TBM
- iii) Baseline mediator concentrations in CSF to blood in a matched timepoint (day 3) and calculated median CSF: blood ratios for each mediator in all patients with HIV associated TBM;
- iv) Baseline (day 3) mediator concentrations in blood and CSF in i) those microbiologically confirmed disease (either positive CSF Gene Xpert Ultra, positive CSF TB culture or both) to ii) those without microbiologically confirmed disease
- v) Baseline (day 3) mediator concentrations in blood and CSF in i) those with good outcome (MRS 0-3) versus those with poor outcome (MRS 4-6) and ii) those who survived versus those who died (within 2 months of diagnosis).
- vi) Baseline (day 3) CSF: Blood ratios in those with good versus those with poor outcome;
- vii) Longitudinal change in CSF (day 3 and day 28) and blood (days 1, 3, 28 and 56) mediators in all TBM patients and those with good versus those with poor outcome;

Statistical analysis was performed using GraphPad Prism (version 9.3.1) software. We compared variables between groups using unpaired T Tests (parametric data) or Mann Whitney Tests of significance (non-parametric data). Throughout the analysis we used an unadjusted p value of <0.05 as a nominal threshold for statistical significance. Given the explorative nature of the analysis, and the small sample size we did not account for multiple testing.

## 5.3 Results

### Baseline demographic and clinical results

A total of 18 of the 52 participants recruited to LASER-TBM had sufficient stored CSF to be included within this analysis. Of these, 14 participants had CSF at day 3 timepoint only, 4 at day 3 and day 28 timepoints, and 4 at day 28 timepoint only. All 18 participants had plasma collected at the enrolment visit, and 12 of 18 participants had plasma available from the day 56 timepoint. We analysed 14 and 8 plasma samples at respective day 3 and day 28 timepoints (Figure 5.2). 39 participants provided consent for enrolment to HIATUS-3. Of these 39 participants, CSF from 38 participants underwent Luminex analysis. Of 38 participants, data on 16 participants was not included as final diagnosis was due to an infectious aetiology, or not confirmed as non-infectious aetiology. 22 participants were included in the 'non-infectious' comparator group (Figure 5.2). Baseline demographic and clinical characteristics are outline in Table 5.1.

A summary of mediators found to have statistically significant differences in each of the comparisons listed below are highlighted in figure 5.3. Of note, only IL1beta was highlighted in more than one comparison other than in the comparison of those with microbiologically confirmed disease versus those without. Specifically, IL1 $\beta$  was i) significantly raised in the CSF of those with TBM compared to non-infectious comparators and ii) significantly raised in the blood of those with poor outcome. These are discussed in greater detail in the paragraphs below.

### **CSF mediator concentrations in patients with TBM and non-infectious comparators**

Concentrations of mediators IL-1 $\beta$ , IL10, IL12p70, IL-17A, IL-18, IP-10 (CXCL10), and MCP1 (CCL2) were significantly raised in the CSF of patients with TBM at baseline (day 3) compared to non-infectious comparators (Table 5.2, Figures 5.3, 5.4 and 5.5). IL-6, an interleukin which acts as both a pro-inflammatory cytokine was non-significantly higher in non-infectious comparators compared to patients with TBM (median concentration 20.3 vs 5.79,  $p=0.14$ ). Similarly, concentrations G-CSF, a haemopoietic growth factor were not significantly different between the two groups (median concentration in TBM vs non-infectious comparators, 14.8 vs 18.7,  $p=0.61$ ).

### **CSF mediator concentrations in patients with microbiologically confirmed TBM vs those without**

Of 14 patients with HIV associated TBM included in baseline analysis, 5 had microbiologically confirmed TBM (TB culture positive and/or Gene Xpert Ultra (GXP Ultra) positive CSF). Concentrations of 38/65 mediators measured were found to be significantly different in concentration in those with microbiologically confirmed TBM compared to those without (Table 5.3, Figure 5.6).

### **Highly compartmentalised inflammatory responses in TBM despite clinical outcome**

Within the TBM group, at the day 3 timepoint concentrations of 55 of the 65 mediators measured were significantly higher in CSF when compared to matched timepoint plasma samples (Table 5.4). Figure 5.7 plots median

fold increase in concentration calculated by CSF:Blood ratio demonstrating mediators which highest and least degree of compartmentalisation.

**Biomarkers of poor outcome in CSF and blood and effect of outcome on CSF: blood concentration ratio and degree of compartmentalisation.**

In the CSF we found 7 mediators (IL-8 (CXCL1), MIP A $\alpha$  (CCL3), APRIL, TRAIL, MIF, SCF and SDF-1 $\alpha$  where concentrations were significantly raised at baseline in the CSF in those with poor versus those with good outcome (Table 5.5, Figures 5.8 and 5.9). A statistically significant difference in the concentration of G-CSF was also noted with outcome although given this was not significantly raised in CSF of patients with TBM compared to non-infectious comparators this was not felt to be a clinically significant finding. In the blood 8 mediators (IL-1  $\beta$ , IL-27, LIF, IFN  $\gamma$ , BAFF) were significantly raised in those with poor compared to those with good outcome (Table 5.5, Figure 5.10). On comparing CSF: blood ratios in those with differing outcomes, we found 5 mediators where CSF: blood ratios were significantly higher in those with poor outcome (IP-10, CXCL-5, MCP 2, BLC, MDC) (Table 5.5, Figure 5.11). Conversely median CSF: blood ratios of IL-27 were significantly lower in those with poor versus those with good outcome (2.39 vs 20.94,  $p=0.0008$ ). Table 5.5 also shows that in those with poor outcome a fewer number of mediators demonstrated significantly higher concentrations in the CSF compared to blood, than in those with good outcomes (11/65 vs 49/65,  $p<0.0001$ ).

**Change in CSF and Blood mediators over time dependant on outcome**

We selected mediators for longitudinal comparison that had demonstrated a significant relationship to outcome (i.e. IL-1  $\beta$ , IL-27, LIF, IFN $\gamma$ , BAFF in

blood, and IL-8 (CXCL8), MIP-1 $\alpha$  (CCL3), APRIL, TRAIL, MIF, SCF and SDF-1 $\alpha$  in CSF). Only 4 participants (3 with good outcome, 1 with poor outcome) had CSF at both baseline and follow up timepoints and therefore it was difficult to draw conclusions from longitudinal CSF markers (Figure 5.12). Within the blood IL-1 $\beta$ , IFN $\gamma$ , IL-4, IL-15, IL-27 and BAFF appeared to increase in participants who died, however in the one participant who survived despite severe disability (MRS 5) mediator concentrations remained at similar levels to those with 'good' outcome (Figure 5.13). When comparing means (and standard error) of mediator concentrations in those who survived vs those who didn't, concentrations appeared to increase prior to death in the two cases who died (Figure 5.14).

## 5.4 Discussion

We described immune profiles in blood and CSF of patients with HIV associated TBM and compared these to a group of patients with non-infectious aetiology, using a comprehensive immunoassay panel. We have shown, in keeping with the literature, that the immune response in HIV associated TBM is highly compartmentalised and that inflammatory mediators appear to increase prior to death in those with HIV associated TBM. Although small, our study identifies mediators which are associated with microbiologically confirmed disease, and with poor clinical outcome including death. To our knowledge this is the first study to consider how immune profiles change in those with poor outcome in HIV associated TBM (death or disability due to any cause) and how the degree of compartmentalisation differs with clinical outcome, an important



consideration in the search for suitable blood-based biomarkers of poor outcome.

### **Heterogeneity and microbiologically confirmed disease**

We identified 7 mediators in which a significant difference in concentration was found in those with HIV associated TBM compared to a comparator group of patients where neurological infections (bacterial, tuberculous and cryptococcal) had been ruled out. Mediators known to play a role in the innate immune response such as IL-1 $\beta$ , IL-10, IL-17 $\alpha$ , as well as those thought to bridge a link between innate and adaptive response such as IL-18, IL-12p70 and IP-10 were raised; in keeping with published data where upregulation of a diverse set of mediators suggests that dissemination of TB to the central nervous system leads to both innate and adaptive immune responses. In those with microbiologically confirmed disease IL-6 and IFN $\gamma$  previously described in immunopathological pathways in TBM were also upregulated (Saghazadeh and Rezaei, 2022). Also present in higher concentrations in those with microbiologically confirmed disease was IL-16 a pleiotropic cytokine that functions as a chemoattractant, modulator of T cell activation and inhibitor of HIV replication which to our knowledge has not previously been described in TBM (Cruikshank et al., 2000). IP-10 was also present in higher concentrations within this group. This finding is in keeping with the literature where IP-10 concentration is widely quoted as a marker of TB disease activity (Hong et al., 2014, Azzurri et al., 2005). It has also been found in higher concentrations in the CSF of patients with TBM, alongside MIG (Yang et al., 2014b), which was also raised within our microbiologically confirmed TBM participants. These findings need validation in a larger cohort but may provide rationale to investigate IP-10

and MIG as dynamic biomarkers to monitor the efficacy of anti-TB drugs in TBM.

Overall however, although all participants included within the TBM case group were defined as having either possible, probable or definite TBM (Marais et al., 2010), there was significant heterogeneity in the immune mediator profiles between cases, including where TBM was microbiologically confirmed versus those where it was not. This finding echoes the heterogeneity in the clinical presentation of TBM, where a wide spectrum of clinical presentations and neurological sequelae exist; leading to the recognised difficulties when diagnosing TBM, and in part subsequent delays in management. However it also highlights the limitations of the now widely used case definition criteria, in particular when including those with 'possible' TBM where an alternative diagnosis may be mimic TBM thus leading to the subsequent complexities of clinical diagnosis and the need for improved diagnostic tools. Given the size of this study, it would not have been possible to exclude those with possible TBM, however in larger studies the inclusion of only those with microbiologically confirmed or 'probable' TBM may lead to less heterogeneity in cases, and thus more obvious insights into key pathogenic pathways. Comparing definite and probable cases with possible cases within this cohort may also shed light on this issue.

### **Variation in the compartmentalisation of the host inflammatory response in TBM**

Our results align with prior studies which demonstrate a highly compartmentalised inflammatory response within the CSF. We found a

significant ( $p < 0.05$ ) difference in the concentration of 55 of the 65 mediators we measured when comparing matched CSF and blood timepoints with median fold increase in concentration up to 250.83. In general, cytokines appeared more likely to be present in high concentrations in CSF compared to chemokines, where often greater concentrations were found in blood compared to CSF. This is an important observation when considering both pathogenic mechanisms but also potential blood biomarkers of disease. Moreover, the number of mediators with significantly different concentrations in CSF compared to blood changed depending on the clinical outcome. Specifically, in those with poor outcome the extent of compartmentalisation appeared to reduce, with only 11/65 mediators found at significantly higher concentrations in CSF compared to blood; compared to 49/65 mediators in those with good outcome. This finding is most likely to be due to the small number of participants included who suffered poor outcome ( $n=3$ ), but could also reflect blood brain barrier integrity and degree of systemic inflammation in those with severe disease. This observation however provides rationale that blood based biomarkers in TBM may still hold promise as potential diagnostic and management tools, particularly in those with severe disease.

### **Innate vs adaptive immune responses in TBM**

The innate immune response, particularly mechanisms driving neutrophil activation is an important contributor to paradoxical inflammation occurring in HIV-associated TB including TBM (Lai et al., 2013, Lowe et al., 2012, Marais et al., 2017, Marais et al., 2014). The results of our study highlight the role of the innate immune response in TBM, with higher concentrations of IL-8, MIP-1  $\alpha$  and MIF in the CSF and IL-1  $\beta$  in the blood, in those with

poor clinical outcomes. However, we also demonstrate that in those with poor outcomes, concentrations of mediators known to be involved in T cell regulation and adaptive immune responses, such as TRAIL in CSF as well as BAFF and IL-27 in blood are also upregulated, suggesting that the T cell compartment may also play an important role in severe cases of TBM. This may have important implications for considering biomarkers but also treatment approaches in those with severe disease.

### **IL-1 $\beta$ , a potential biomarker of poor outcome and target for host directed therapies?**

In IRIS occurring as a complication of HIV-associated TBM, inflammasome activation leads to cleavage and secretion of IL-1 $\beta$  a pro-inflammatory cytokine (Marais et al., 2017). In herpes simplex encephalitis, CSF IL-1 $\beta$  is associated with clinical severity, blood brain barrier permeability and disease outcome (Michael et al., 2016). A recent study in TBM however showed that although CSF IL-1 $\beta$  levels were raised in patients with TBM, this mediator did not associate with mortality (Koeken et al., 2021). Similar to this, our study demonstrated a raised level of IL-1 $\beta$  in the CSF of patients with TBM compared to non-infectious comparator group, although this did not predict poor outcome. However, we did show that IL-1 $\beta$  was raised in the blood of those who went on to have poor clinical outcomes. Moreover, our longitudinal analysis demonstrated that blood IL-1 $\beta$  levels, increased following initiation of treatment reaching highest concentrations prior to death in the two participants who died. These results provide further rationale for the validation of these findings in a larger cohort, particularly given anakinra, a drug which targets the IL-1 $\beta$  pathway has been reported

to be effective in treating paradoxical inflammation in TBM (Keeley et al., 2020).

Our findings around IL-1 $\beta$  may also suggest that existing blood-based biomarkers of inflammation may be effectively utilised; C-reactive protein an acute phase protein is released by hepatocytes in response to IL-6 with its synthesis enhanced by IL-1 $\beta$  (Mackiewicz et al., 1991). One study showed that a raised serum C-reactive protein was helpful in discriminating TBM from bacterial meningitis at disease onset (Ersoy et al., 2012). In another study, high CSF C-reactive protein predicted mortality with a relative risk of 2.92 ( $p=0.027$ ) however authors discuss how methods for measurement of serum C-reactive protein would need optimisation given that CSF concentrations are significantly lower than in serum (Ratinam et al., 2020). Nonetheless, in a condition where no blood biomarkers or poor outcome exist, the possibility that longitudinal measurements of IL-1 $\beta$  or C-reactive protein as a proxy for IL-1 $\beta$  may predict clinical worsening is an interesting area for future research.

### **Novel sites for biomarker measurement in TBM**

Our study only investigated for potential biomarkers in the blood and cerebrospinal fluid of patients with TBM. In other forms of TB urine biomarkers have been considered. Lipoarabinomannan (LAM) a mycobacterial antigen is commercially available for TB diagnostics, however its use is largely limited to patients with HIV where CD4 counts are low (Songkhla et al., 2019). Recent studies have also highlighted host urine immunological biomarkers for the diagnosis of pulmonary TB (Petroni et al., 2016, Eribo et al., 2020). In other neurological disease, such as multiple

sclerosis urinary biomarkers have been evaluated (Dobson et al., 2013); to our knowledge this has not been explored in TBM. Along these lines, detection of host based biomarkers in saliva have shown promise for the diagnosis of tuberculosis in children (Khambati et al., 2021). It remains unknown whether these assays have a role in extra-pulmonary TB such as TBM. Given that the WHO has prioritised the development of non-sputum-based point-of-care tests on easily obtainable samples like blood, urine, stool, and saliva for tuberculosis (Denkinger et al., 2015), future studies within the TBM field should also consider these novel sites for biomarker development.

### **Study limitations**

There are several limitations to this study. Firstly, the number of participants included was small and therefore the study was underpowered, particularly in the longitudinal analysis and the comparison of good to poor outcomes where in the latter only 3 participants with poor outcome were included. Due to the small numbers it was also not feasible to account for multiple comparisons within our analysis, and therefore it is possible that a proportion of our significant findings may be due to chance. Larger studies are now required to validate these findings, with a continued effort to acquire matched longitudinal timepoint samples particularly in the CSF. Secondly the heterogeneity within the group may in part be due to the inclusion of patients in whom an alternative diagnosis is more likely. For example, one of the two participants who died repeatedly appeared as an outlier within the analysis. Delving further into this case revealed that this participant, although clearly fulfilling criteria for 'possible TBM' using the case definition criteria had a dual diagnosis of neurosyphilis. In cases such

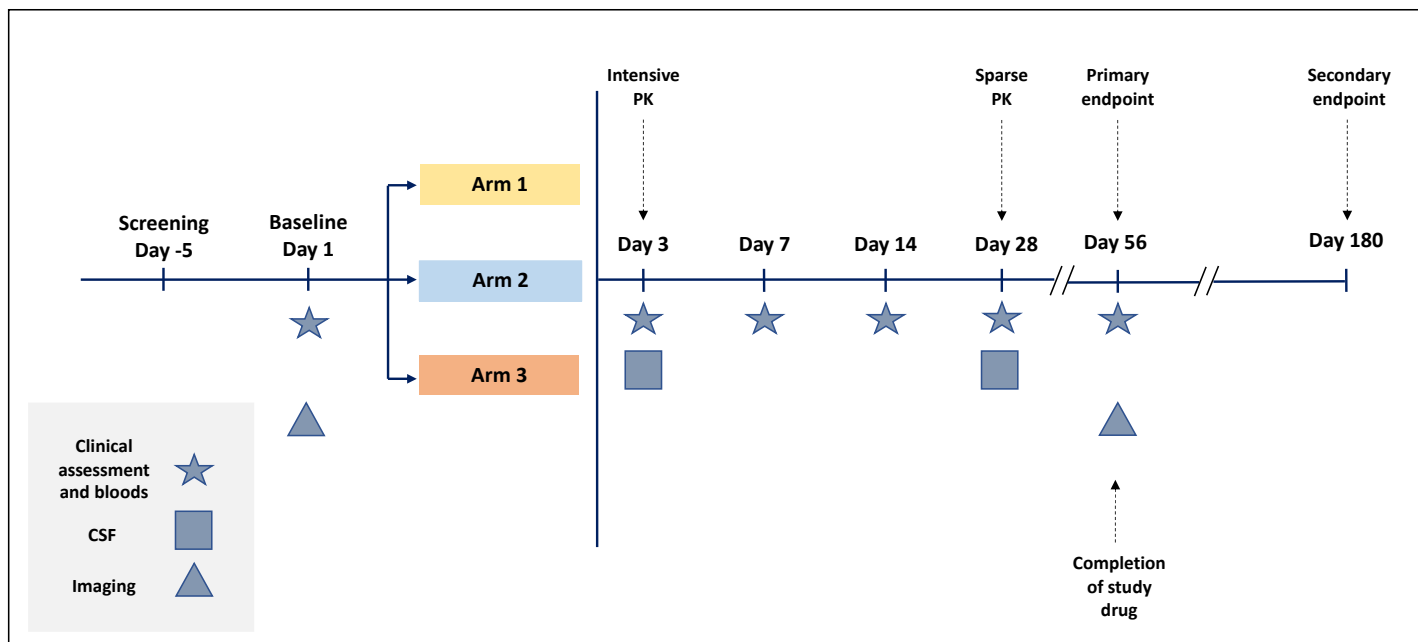
as this the contribution of co-infection, and/or the possibility that TBM was after all an incorrect diagnosis highlights the complexity of pathogenesis research in a condition where no highly sensitive, highly specific diagnostic test exists. Ultimately the design of studies on a scale which allows such cases to be excluded from the analysis will lead to a less heterogeneous clinical group and therefore more robust insights into pathogenic mechanisms of disease. A principle component analysis in future analysis would also help to identify outliers. Along these lines, the inclusion of a non-infectious comparator group was pragmatic and therefore, although bacterial, mycobacterial and cryptococcal infectious aetiology had been ruled out in this group the spectrum of final diagnosis was highly heterogeneous. Although acquiring CSF from healthy controls is difficult, only a comparison with such a group can allow us to reliably conclude the full spectrum of raised inflammatory mediators in TBM. Finally, it was interesting to note that in the limited longitudinal CSF data available despite 4 weeks of treatment, multiple inflammatory mediators remain raised. Longer term follow up timepoints would allow us to glean important information as to which inflammatory markers remain raised. In particular, future studies should consider the inclusion of mediators known to be markers of brain injury (eg NSE, S100B, tau) to understand whether the inflammatory mediators we have seen raised in those with poor outcomes continue to rise and/or initiate a cascade of brain injury processes previously described in paediatric TBM, as well as in conditions where there is a recognised interplay between neuroinflammation and neurodegeneration.

## 5.5 Conclusions and future research

Despite these limitations, these analyses allow us to glean insight into pathogenic mechanisms in TBM, including those who have microbiologically confirmed disease. They highlight potential mediators for further investigation which may serve as useful biomarkers of poor outcome including within the blood and/or serve as useful druggable targets in the development of host directed therapies. In particular, the findings around IL-1 $\beta$  and inflammasome driven innate immune responses provide further rationale for the consideration of drugs which target these pathways. Further studies using larger cohorts and including longer term timepoints, ideally with a healthy control group are now needed to further investigate and validate these findings. Moreover, integration of these results with transcriptomic, metabolomic and proteomic analysis within the same cohort will add depth of knowledge and clarity to pathways of interest. This analysis is planned, funded and will be undertaken by myself as part of the Wilkinson Group at Crick beyond the course of my PhD.

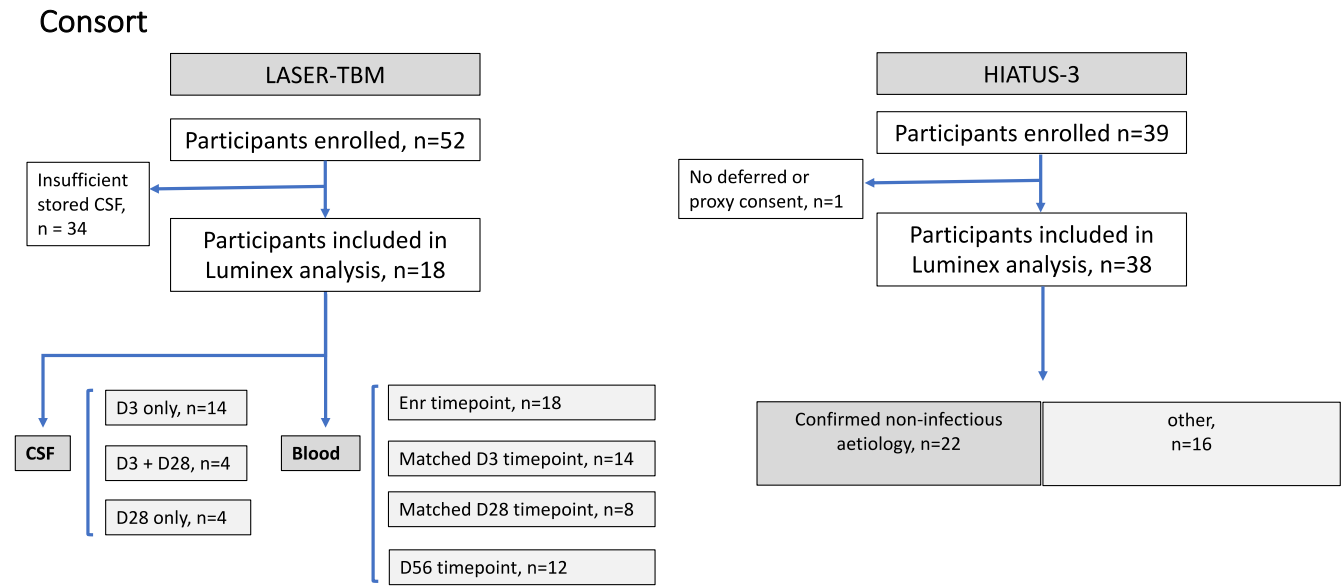


**Figure 5.1 Study sampling schedule for LASER-TBM**



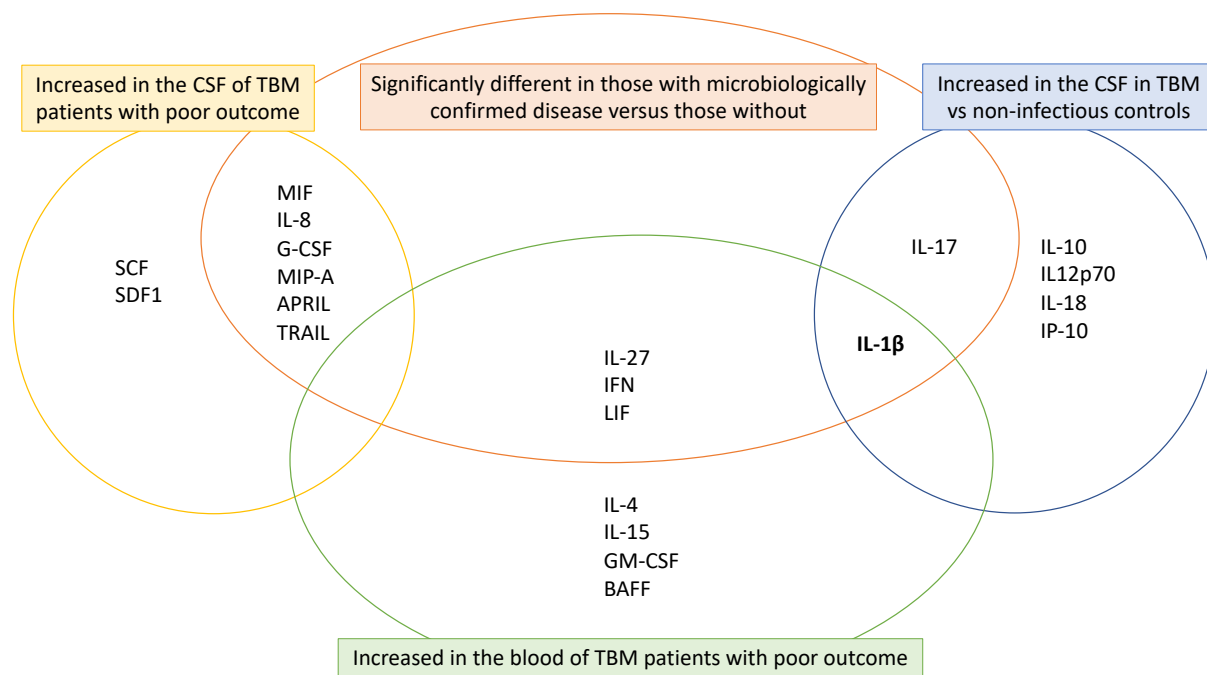
Study participants enrolled to LASER-TBM were seen at 6 timepoints between day 1 (enrolment) and day 56. Blood for immunological analysis was collected at each of these timepoints. Where possible lumbar puncture was performed at day 3 and day 56

**Figure 5.2 CONSORT diagram for Luminex analysis study**



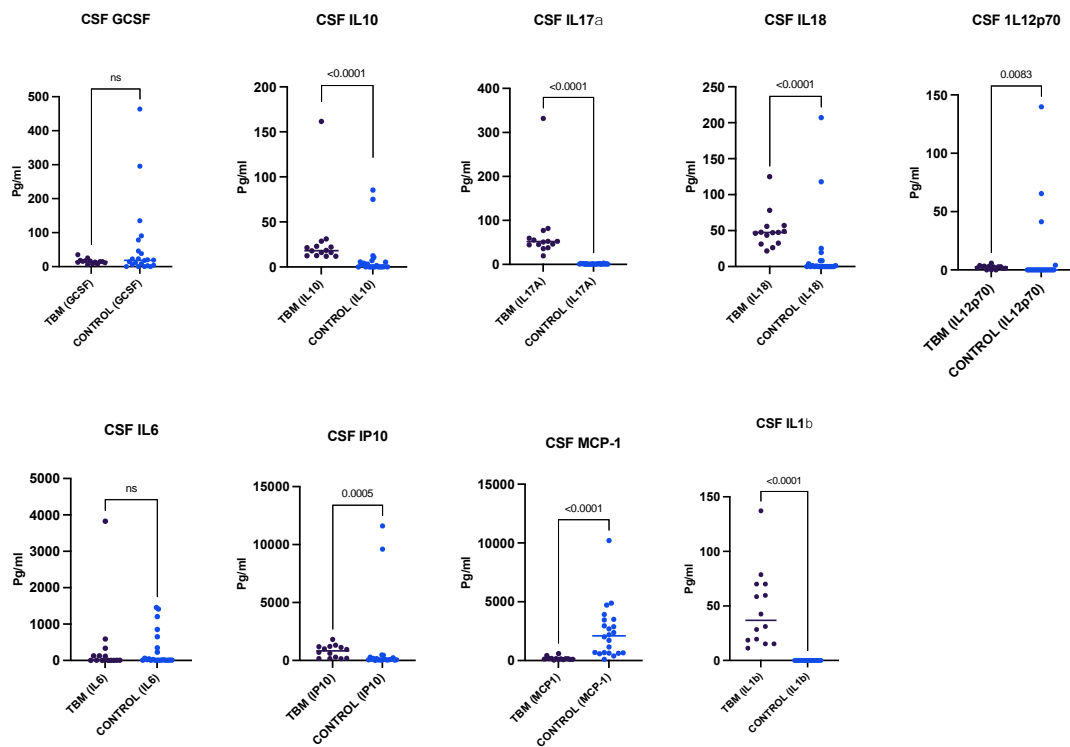
Participants were included as per the consort diagram from the parent studies LASER-TBM and HIATUS-3. Enr; enrolment

**Figure 5.3 Summary Venn diagram to describe mediator findings across all analysis groups**



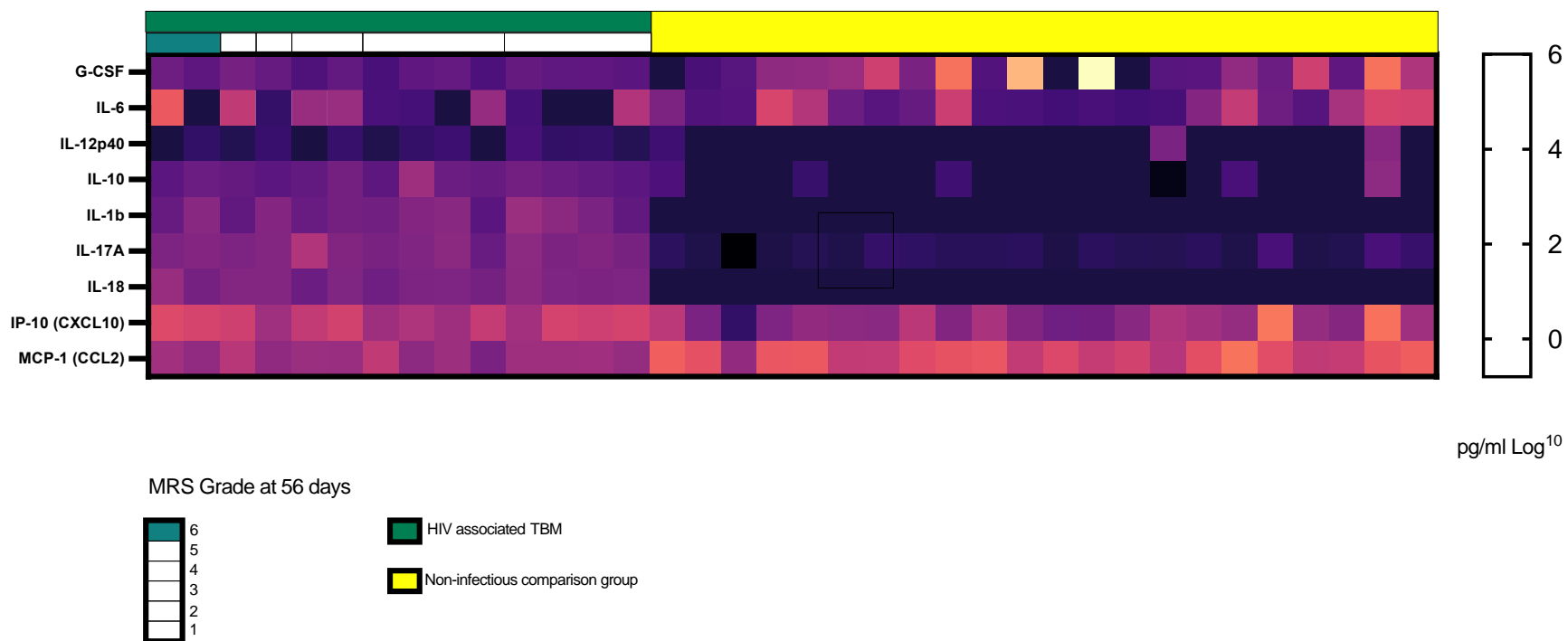
This Venn diagram highlights mediators in which a significant difference ( $p < 0.05$ ) was found in each of the three comparisons i) comparison of CSF mediators in those HIV associated TBM with good versus poor outcome (yellow), ii) blood mediators in those with HIV associated TBM in good versus poor outcome (green) and iii) CSF mediators in those with HIV associated TBM versus non-infectious comparators (blue). Those overlapping with the red circle are those which were found to be significantly different ( $p < 0.05$ ) in those with microbiologically confirmed disease versus those without

**Figure 5.4 Scatter graphs plotting individual mediator values in cases with HIV associated TBM compared to non-infectious comparator group**



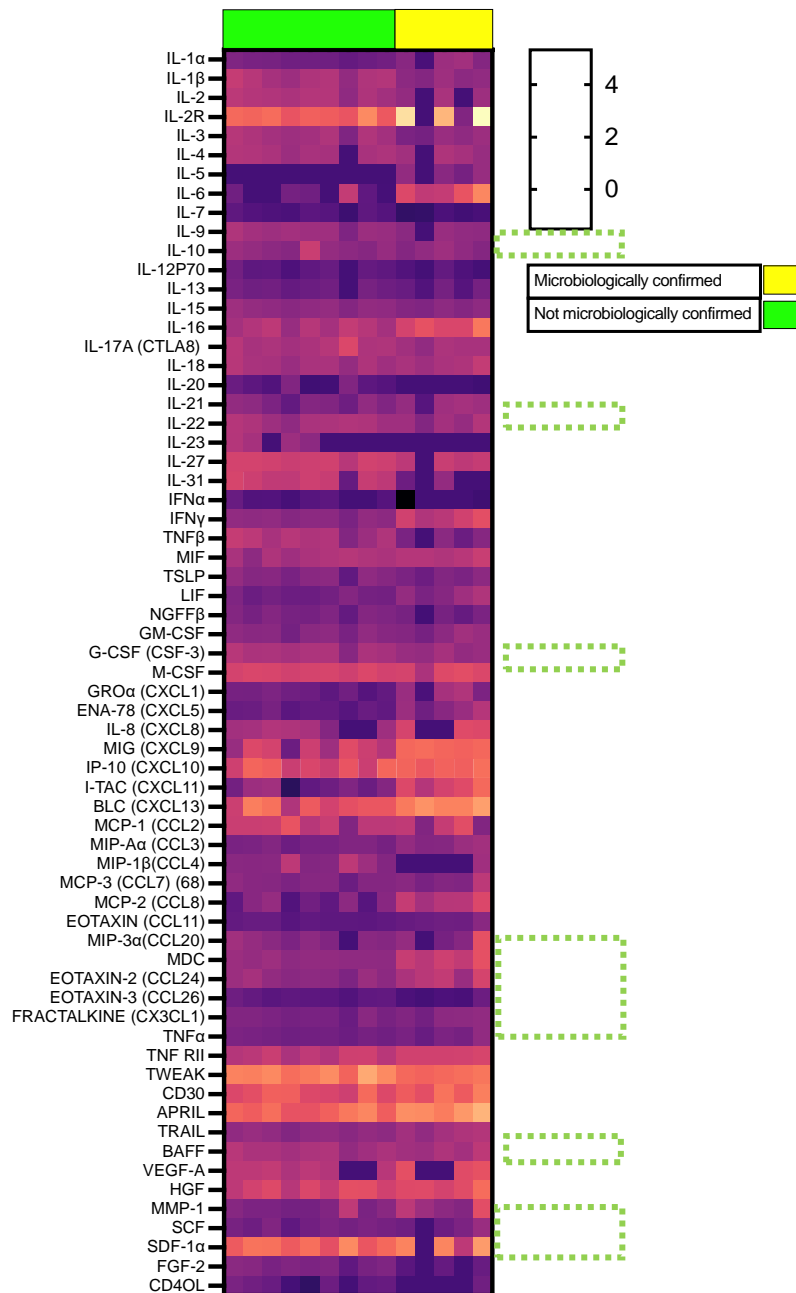
Scatter graphs plot individual mediator concentrations in cases (HIV associated TBM) and comparators (non-infectious aetiology)

**Figure 5.5 Day 3 CSF immune mediators in patients with HIV associated TBM compared to non-infectious comparator group**



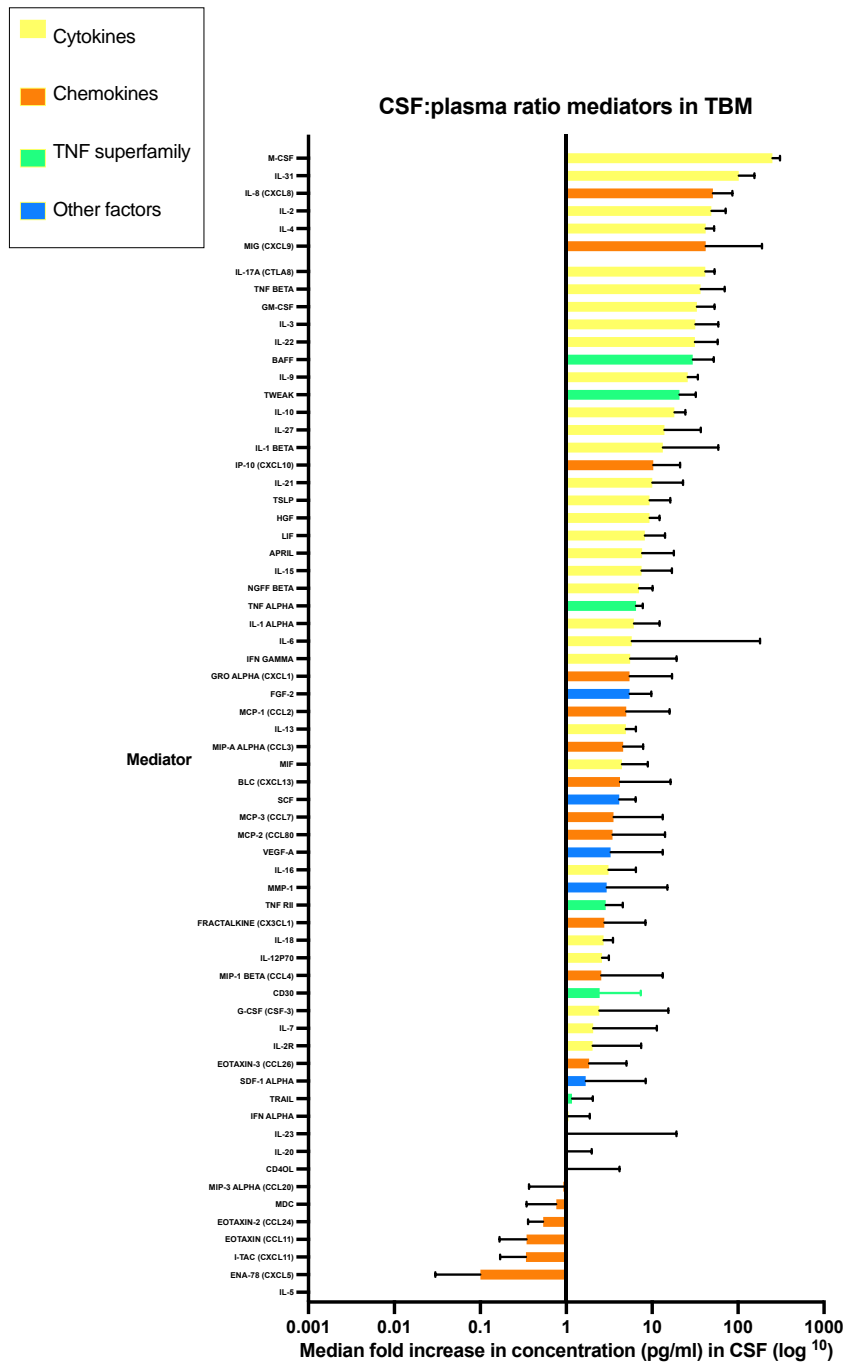
Heatmap plotting individual participants as columns with HIV associated TBM cases on the left (green) and non-infectious comparators on right (yellow). HIV associated TBM cases are ordered from most severe 2 month outcome (MRS 6) on left to best outcome (MRS 0) on right.

**Figure 5.6 Day 3 CSF mediators in those with microbiologically confirmed HIV associated TBM versus those without**



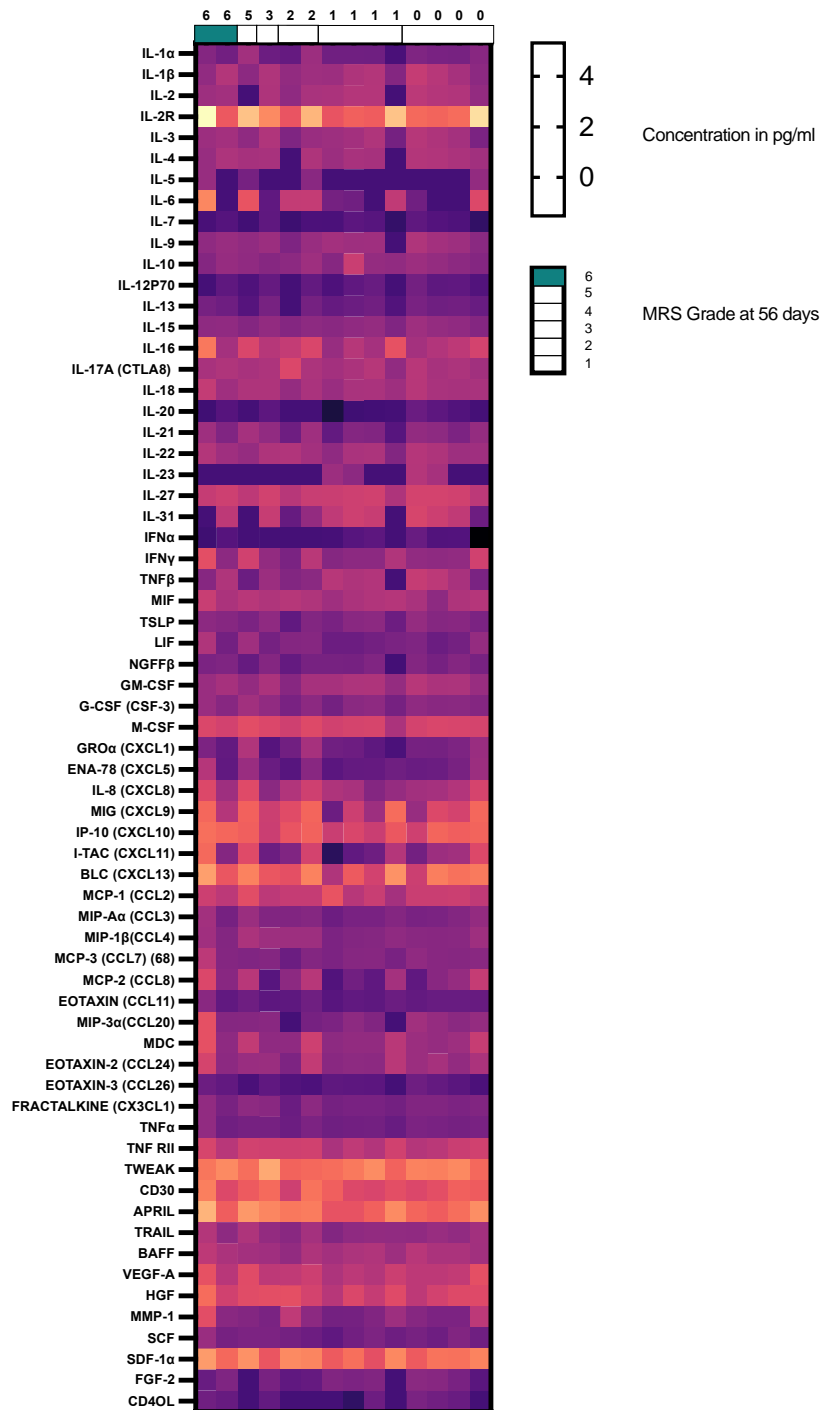
Heatmap plotting individual participants with HIV associated TBM as columns with participants with microbiologically confirmed disease on the right (yellow) and those without microbiologically confirmed disease on the left (green). Highlighted in green dashed boxes are mediators in which obvious differences in concentrations are apparent between the groups.

**Figure 5.7 Comparison of baseline mediators in matched CSF and blood timepoint (day 3), CSF to blood ratio ranked by log fold increase**



Median fold change in CSF:Blood ratio (pg/ml) (CSF concentration/Blood concentration), converted to a log<sub>10</sub> scale, are plotted here for each of the CSF mediators measured in those with HIV associated TBM cases. Mediators are colour coded as per their type (see key).

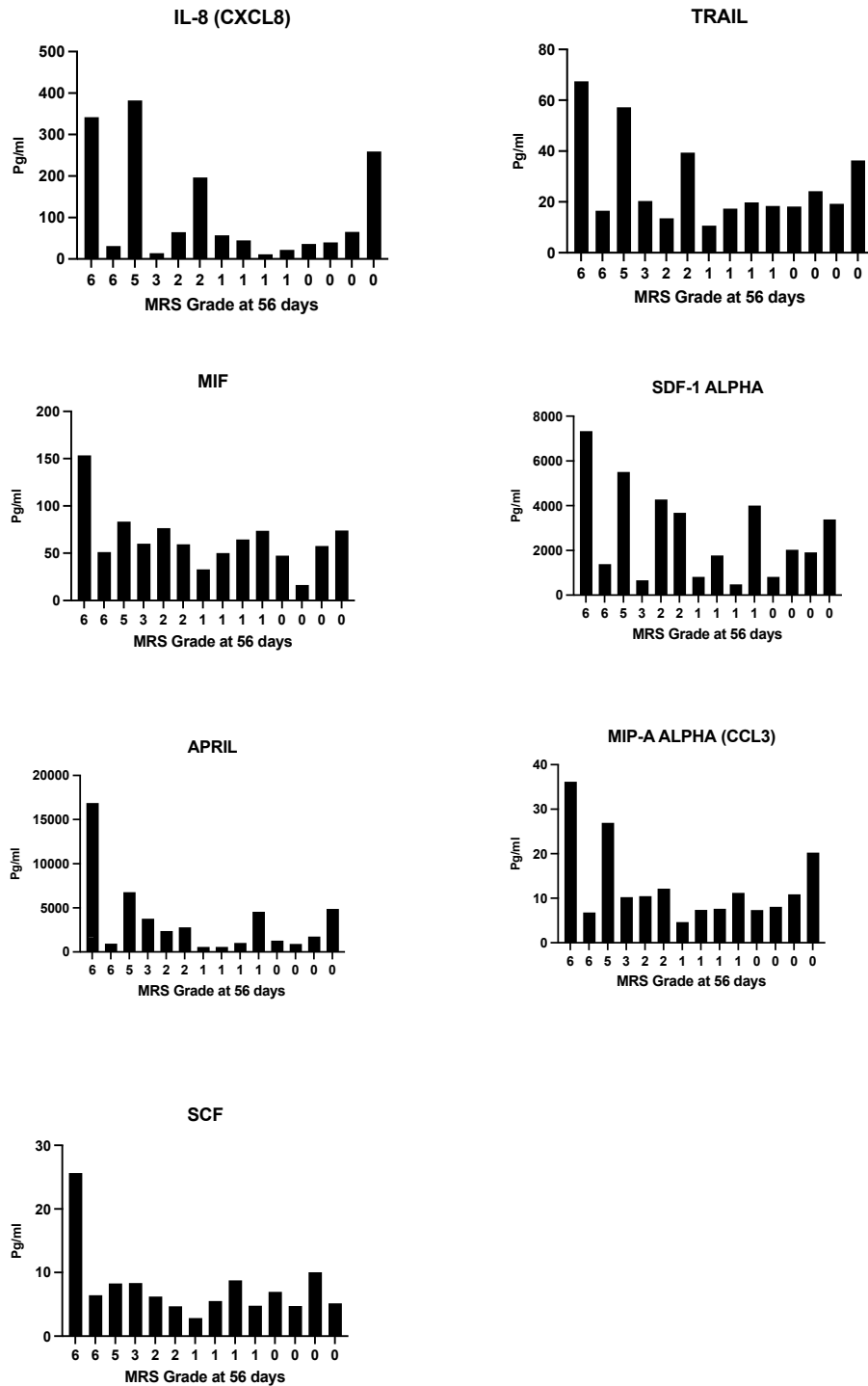
**Figure 5.8 Day 3 CSF mediator concentrations with MRS outcome at D56**



Heatmap in which individual participants with HIV associated TBM are represented as columns with those with most severe outcomes (MRS 6) on left, and those with best outcomes (MRS 0) at day 56 on right.

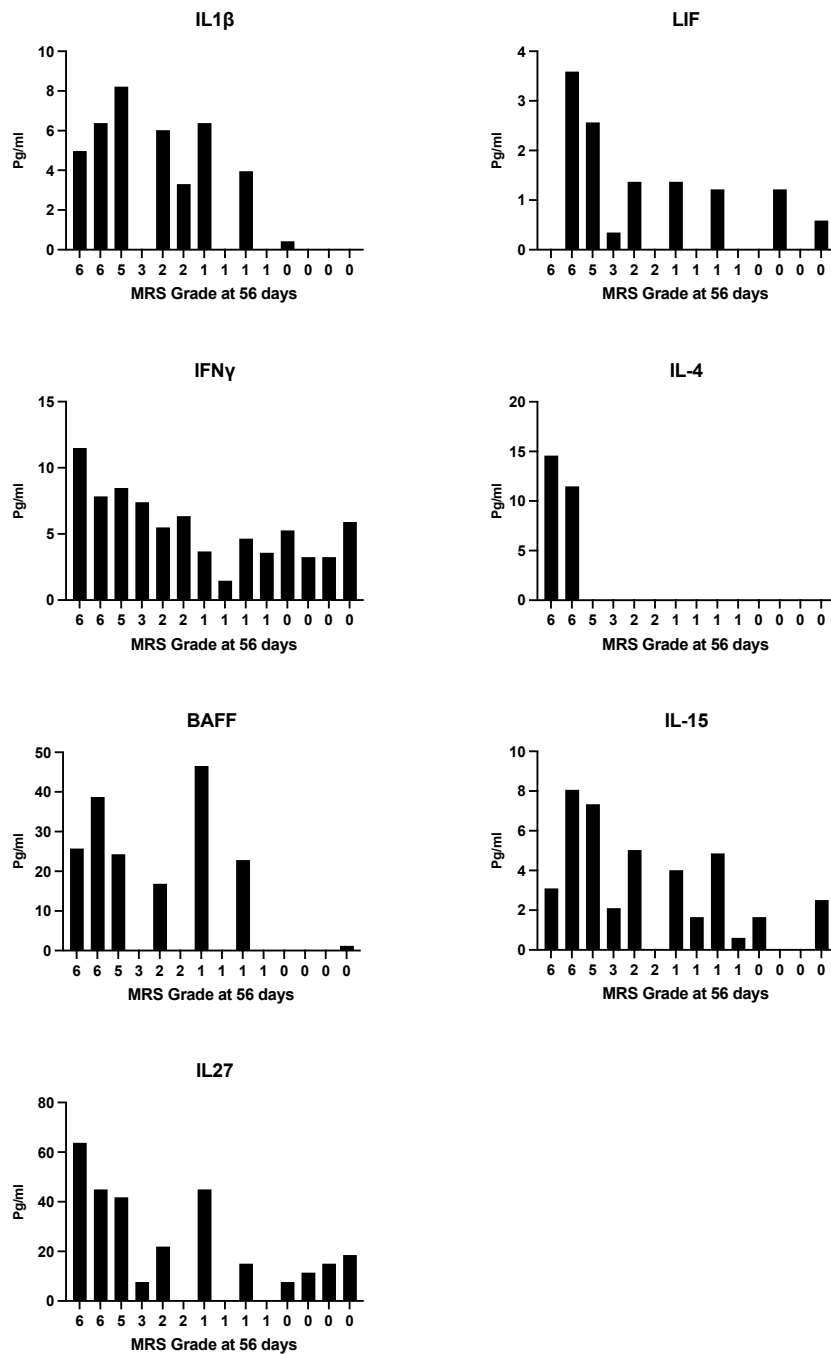


**Figure 5.9 Day 3 CSF markers demonstrating difference ( $p < 0.05$ ) with good (MRS 0-3) versus poor (MRS 4-6) outcome**



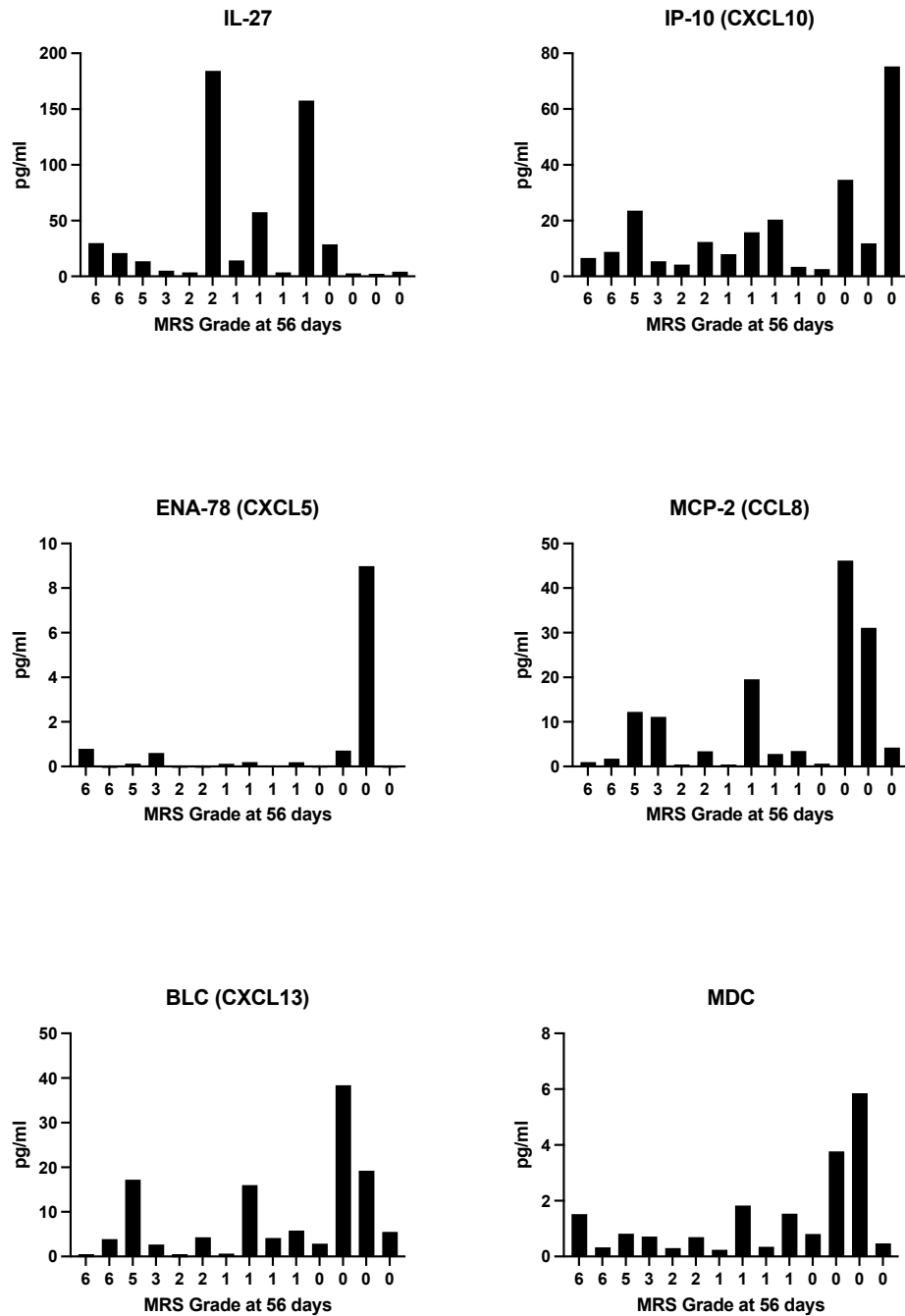
CSF mediators in which a significant difference ( $p < 0.05$ ) in concentration between those with poor versus those with good outcome were selected. Individual participants are represented by individual columns with those with most severe outcomes (MRS 6) on left, and those with best outcomes MRS 0 on right.

**Figure 5.10 Day 3 Blood markers demonstrating difference ( $p < 0.05$ ) with good (MRS 0-3) versus poor (MRS 4-6) outcome**



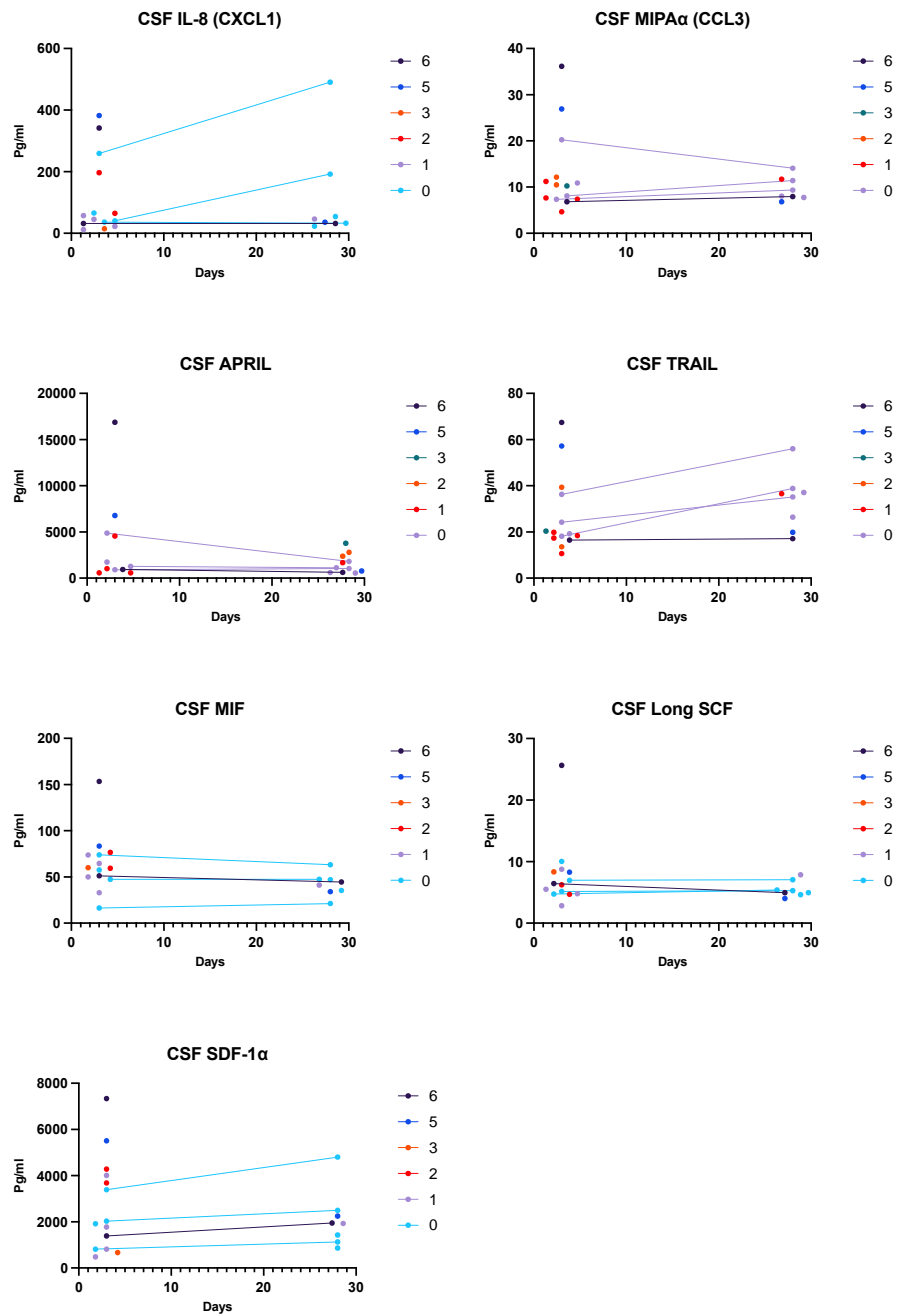
Blood mediators in which a significant difference ( $p < 0.05$ ) in concentration between those with poor versus those with good outcome were selected. Individual participants are represented by individual columns with those with most severe outcomes (MRS 6) on left, and those with best outcomes MRS 0 on right.

**Figure 5.11 Day 3 CSF:Blood ratios in mediators demonstrating difference ( $p < 0.05$ ) with good (MRS 0-3) versus poor (MRS 4-6) outcome**



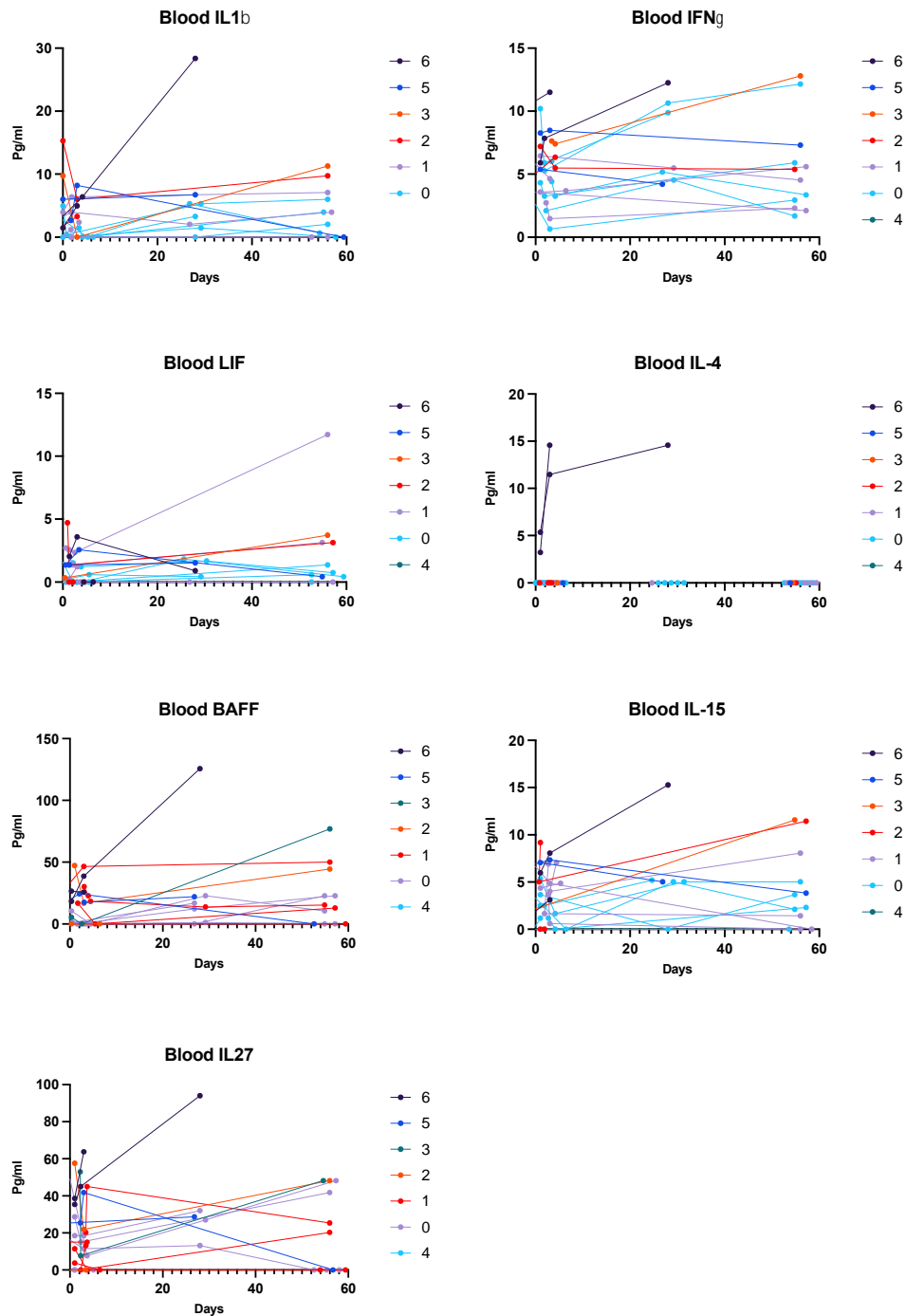
CSF:Blood ratios in mediators where a significant difference in the ratio was found in those with good versus those with poor outcome. In all mediators except IL-27 the trend is towards greater CSF:Blood ratio (ie higher degree of CNS compartmentalisation) in those with milder disease (MRS 0) compared to those with severe disease (MRS 6).

**Figure 5.12 Longitudinal change in selected CSF parameters**



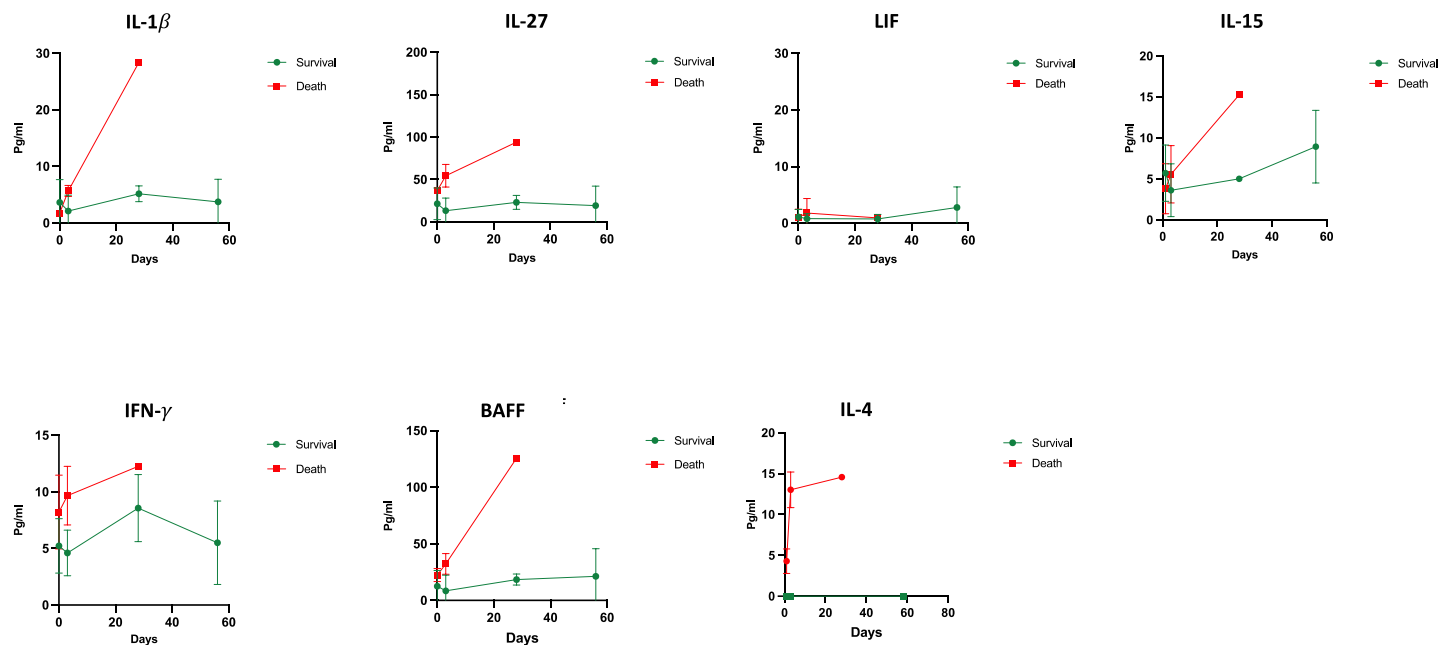
CSF mediators shown to be significantly different in concentration ( $p < 0.05$ ) in those with good versus those with poor outcome were selected. Baseline (day 3) and follow up (day 28) were plotted. Participants were colour coded as per the day 56 MRS outcome (0-6). Those with paired samples (ie both a day 3 and day 28 samples) are linked by a single line.

**Figure 5.13 Longitudinal change in selected blood parameters**



Blood mediators shown to be significantly different in concentration ( $p < 0.05$ ) in those with good versus those with poor outcome were selected. The following timepoints were plotted: day 1, day 3, day 28 and day 56 where available, and when from the same participant linked with a single line. Participants were colour coded as per the day 56 MRS outcome (0-6)

**Figure 5.14 Longitudinal change in selected blood markers grouped by survival**



Blood mediators shown to be significantly different in concentration ( $p < 0.05$ ) in those with good versus those with poor outcome were selected. Participants were grouped into those who survived (green) and those who died ( $n=2$ ) by day 56 and the median (IQR) of mediator concentrations at the following timepoints were plotted: day 1, day 3, day 28 and day 56 where available.

**Table 5.1 Baseline demographics and clinical characteristics of participants included within the Luminex study analysis**

	HIV associated TBM (all)	HIV associated TBM Good Outcome (MRS 0-3)	HIV associated TBM Poor Outcome (MRS 4-6)	Non-infectious comparison group
<b>n</b>	18	14	4	22
<b>Age (years) (median, IQR)</b>	38.5	38.5 (28.0-38.5)	40.5 (32.5-44.8)	47.5 (36.8-66.0)
<b>Gender (% female)</b>	39	50	0 (0)	13 (59)
<b>Blood</b>				
<b>WCC</b>	6.07 (3.84-8.68)	5.80 (3.84-9.02)	6.07 (3.72 – 8.01)	8.25 (6.0-15.3)
<b>CD4 count, cells/uL</b>	168 (86.8-383)	168 (122-383)	193 (17.8-749)	n/a
<b>HIV viral load, log<sub>10</sub></b>	60,807 (1,812-219,620)	68,497 (5,608 – 378,736)	60,807 (1,812-151,261)	n/a
<b>Cerebrospinal fluid</b>				
<b>Lymphocyte cell count x10<sup>6</sup>/L</b>	5.5 (1.0-23.0)	5.5 (1.75-23.0)	8.0 (0.25-33.0)	0.0 (0.0-1.25)
<b>Polymorphonuclear cell count x10<sup>6</sup>/L</b>	19.0 (3.75-308)	27.5 (7.75-411)	5.0 (0.0-64.0)	0.0 (0.0-0.25)
<b>Protein, g/L</b>	2.31 (1.1-3.72)	2.03 (0.865-3.2)	4.25 (1.74-12.8)	0.36 (0.21-0.55)
<b>Glucose, mmol/L</b>	2.2 (0.9-2.95)	2.30 (1.2-3.1)	0.95 (0.625-2.03)	3.70 (2.6-5.73)
<b>GXPU positive, n (%)</b>	5 (27.7)	3 (21.4)	2 (50)	0

**Table 5.2 Baseline (Day 3) CSF mediator concentrations in patients with TBM (n=14) and non-infectious comparators (n=22)**

	HIV associated TBM (n=14)		Non-Infectious comparator (n=22)		HIV associated TBM vs Non-Infectious Comparators	Direction in HIV associated TBM group compared to non-infectious comparators
Mediator	CSF		CSF			
pg/ml	median	(IQR)	median	(IQR)	p-value	
<b>Mediators measured across both groups (HIV associated TBM and non-infectious comparators)</b>						
<b>Cytokines</b>						
G-CSF	14.8	10.4-17.6	18.7	3.97-54.2	0.6135	↓
IL-6	5.79	0.77-181	20.2	6.8-422	0.1391	↓
IL-12P70	2.61	1.01-3.13	<	0.0-0.0	0.0083	↑
IL-10	18.2	12.6-24.4	0.935	0.0-6.03	<0.0001	↑
IL-1 $\beta$	36.9	17.8-70.1	<	0-0	<0.0001	↑
IL-18	47.4	32.3-56.3	<	0-8.21	<0.0001	↑
IL-17A	51.8	42.8-63.9	0.82	0.5-1.66	<0.0001	↑
<b>Chemokines</b>						
IP-10 (CXCL10)	834	186-1198	91.8	50.6-259	0.0005	↑
MCP1 (CCL2)	146	95.3-175	2104	651-3466	<0.0001	↓



**Table 5.3 CSF mediator concentrations in those with microbiologically confirmed HIV associated TBM vs those without**

Mediator pg/ml *	Microbiologically confirmed		Not microbiologically confirmed		P value	P value summary	Direction (micro confirmed disease vs not)
	CSF median	IQR	CSF median	IQR			
<b>Cytokines – raised in microbiologically confirmed disease</b>							
IL-16	310	271- 1462	66.8	39.4- 89.4	0.0010	***	↑
IL-5	13.6	3.35- 21.9	0	0-0	0.0050	**	↑
IL-6	337	122- 2208	2.77	0- 5.79	0.0015	**	↑
IFN $\gamma$	207	75.4- 341	15.8	13.8- 19.1	0.0029	**	↑
LIF	21.3	10.1- 46.5	5.99	4.44- 7.91	0.0035	**	↑
IL-1 $\alpha$	13.1	6.35- 33.1	5.26	4.86- 7.16	0.0207	*	↑
IL-21	31.4	12- 36.2	10.2	6.68- 17.6	0.0233	*	↑
MIF	74	66.5- 118	51.3	40.1- 62.3	0.0217	*	↑
G-CSF	15.8	9.07- 30.4	14.8	10.2- 17.1	0.0186	*	↑
IL-2R	18353	5- 137668	1051	715- 1525	0.6064	ns	↑
M-CSF	322	150- 404	251	217- 293	0.5712	ns	↑
IL-18	49	39.4- 90.6	46.4	29.5- 52.5	0.3636	ns	↑
<b>Cytokines – lower in microbiologically confirmed disease</b>							
IL-1 $\beta$	15.3	13.3- 24.9	59.6	35.5- 74.4	0.0035	**	↓
IL-2	20.4	0- 38.9	63.8	43.6- 72.5	0.0062	**	↓
IL-7	0.87	0.51- 1.2	1.81	1.35- 2.49	0.0087	**	↓
IL-20	0	0- 0.415	2.43	1.28- 7.04	0.0015	**	↓
IL-31	0	0- 12.6	143	102- 183	0.0018	**	↓
TNF $\beta$	7.92	2.16- 13.6	60	36.7- 86.4	0.0062	**	↓
IL-3	17.3	7.2- 27.9	39.6	34.3- 60.6	0.0138	*	↓
IL-9	17.3	7.32- 21.8	31.9	25.9- 37.5	0.0271	*	↓
IL-17A	45.3	27.6- 49.4	55.4	47.7- 79.5	0.0460	*	↓
IL-27	100	47- 148	204	173- 226	0.0120	*	↓
IFN $\alpha$	0.03	0- 0.865	1.7	0.53- 2.16	0.0476	*	↓
IL-4	36	11.8- 50.4	46.2	34.6- 59.1	0.3169	ns	↓
IL-10	17.9	11.9- 24.8	21.4	14.3- 25.8	0.4166	ns	↓
IL-12P70	1.46	0- 2.36	2.83	1.97- 3.54	0.0867	ns	↓

IL-13	3.7	1.73- 6.57	5.09	3.81- 6.27	0.5025	ns	↓
IL-15	11.8	10.6- 16.9	18.1	15.7- 21.4	0.0565	ns	↓
IL-22	33	16.5- 53.6	53.5	31.6- 61.5	0.2246	ns	↓
IL-23	0	0-0	0	0-36.2	0.1983	ns	↓
TSLP	8.16	6.43- 13.2	13.4	9.85- 16.7	0.2370	ns	↓
NGFF $\beta$	7.04	2.22- 7.57	9.49	6.67- 11.1	0.0684	ns	↓
GM-CSF	24.9	21.1- 33.2	51.9	42.2- 55.6	0.2204	ns	↓
Chemokines – raised in microbiologically confirmed disease							
MIG (CXCL9)	1350	1178- 1471	173	27- 291	<0.0001	****	↑
I-TAC (CXCL11)	352	154- 910	5.81	3.62- 20.6	0.0010	***	↑
MCP2 (CCL8)	81.3	60- 232	5.4	2.33- 14.7	0.0010	***	↑
MDC	144	103- 357	19.8	17.3- 25.2	0.0005	***	↑
ENA-78 (CXCL5)	26.4	9.21- 50.9	3.28	2.62- 4.19	0.0015	**	↑
BLC (CXCL13)	3299	2917- 7045	681	199- 1354	0.0012	**	↑
MIP A $\alpha$ (CCL3)	20.3	11.7- 31.5	7.66	7.08- 10.4	0.0027	**	↑
EOTAXIN (CCL11)	4.9	3.97- 9.54	2.96	2.53- 3.43	0.0035	**	↑
EOTAXIN-2 (CCL24)	88.1	37.5- 186	16.6	15- 27.4	0.0070	**	↑
IL-8 (CXCL8)	259	0- 362	36.2	5.76- 51.1	0.0186	*	↑
FRACTALKINE (CX3CL1)	15.9	8.04- 17.7	7.55	6.79- 10.1	0.0370	*	↑
GRO $\alpha$ (CXCL1)	26	4.78- 52.3	5.34	2.87- 6.4	0.1119	ns	↑
IP-10 (CXCL10)	1136	857- 1503	298	165- 1087	0.0829	ns	↑
Chemokines – lower in microbiologically confirmed disease							
MIP 1 $\beta$ (CCL4)	0	0- 16.7	14.1	11.6- 65.2	0.0340	*	↓
MCP1 (CCL2)	110	10- 284	155	86.4- 168	0.5160	ns	↓
MCP3 (CCL7)	11.1	8.53- 54.1	12.4	10- 12.9	0.9640	ns	↓
MIP 3 $\alpha$ (CCL20)	12.6	3.23- 274	13.8	9.27- 19.3	0.9276	ns	↓
EOTAXIN-3 (CCL26)	1.26	0.58- 2.89	2.68	2.25- 3.11	0.0599	ns	↓
TNF superfamily - raised in microbiologically confirmed disease							
TNF RII	214	208- 249	89	68.9- 178	0.0016	**	↑
APRIL	4879	3671- 11825	1023	741- 2059	0.0020	**	↑
TRAIL	39.4	27.4- 62.3	18.2	15- 20.1	0.0016	**	↑
TNF $\alpha$	7.53	5.28- 13.7	6.01	5.21- 7.05	0.1898	ns	↑
CD30	883	638- 2516	348	307- 1008	0.0583	ns	↑

TNF superfamily - lower in microbiologically confirmed disease							
Tweak/TNFSF12	1419	1267-1971	3297	2106- 4426	0.1058	ns	↓
BAFF	37.3	33- 79.2	50.1	35.2- 58.2	0.7840	ns	↓
Other factors – raised in microbiologically confirmed disease							
VEGF-A	401	0- 523	82.9	27.5- 102	0.0299	*	↑
MMP1	30.6	13.2- 274	8.74	7.05- 13	0.0290	*	↑
HGF	385	308- 1004	224	119- 417	0.1119	ns	↑
SDF-1 $\alpha$	3389	50- 5509	1389	742- 1974	0.2561	ns	↑
Other factors – lower in microbiologically confirmed disease							
FGF-2	2.25	0- 3.61	9.57	7.4- 11.8	0.0008	***	↓
SCF	5.15	2.34- 17	6.44	5.13- 8.54	0.6064	ns	↓
CD40L	0	0- 2.65	3.64	0.23- 5	0.1559	ns	↓

**Table 5.4 Baseline (day 3) mediators in HIV associated TBM, a comparison of blood and CSF concentrations**

Mediator	TBM (n=14)						
	Blood		CSF		CSF:Blood ratio		CSF:Blood p value
	median	IQR	median	IQR	median	IQR	
<b>Cytokines – higher in CSF compared to blood</b>							
IL-1 $\alpha$	0.0	0.0-1.2	6.34	5.01-11.9	6.14	2.98-12.20	<0.0001
IL-1 $\beta$	1.86	0.0-6.11	36.9	17.8-70.1	13.3	4.28-58.86	<0.0001
IL-2	0.0	0.0-0.0	48.9	19.7-71.6	48.88	19.68-71.61	<0.0001
IL-3	0.0	0.0-0.0	34.3	15.4-58.7	31.86	15.38-58.69	<0.0001
IL-4	0.0	0.0-0.0	44.3	26.5-55.8	42.02	3.99-52.58	<0.0001
IL-5	0.0	0.0-0.0	0.0	0.0-8.42	0.00	0.0-8.42	0.0978
IL-6	0.0	0.0-0.0	5.79	0.77-181	5.79	1.0-180.87	<0.0001
IL-7	0.28	0.07-1.17	1.37	0.84-2.05	2.06	1.38-11.34	0.0022
IL-9	0.0	0.0-0.16	25.9	16.6-34.0	25.94	11.93-34.04	<0.0001
IL-10	0.0	0.0-0.0	18.2	12.6-24.4	18.16	12.59-24.38	<0.0001
IL-12P70	0.0	0.0-0.0	2.61	1.01-3.13	2.61	1.26-3.13	<0.0001
IL-13	0.0	0.0-0.0	4.96	2.99-6.48	4.96	2.99-6.48	<0.0001
IL-15	2.31	0.46-4.90	17.5	11.8-20.2	7.52	3.11-16.97	<0.0001
IL-16	49.9	26.4-101	89.4	40.0-308	3.11	0.88-6.47	0.0571
IL-17A	0.0	0.0-4.58	51.8	42.8-63.9	41.58	12.51-53.32	<0.0001
IL-18	18.7	13.2-22.1	47.4	32.3-56.3	2.72	1.63-3.50	<0.0001
IL-20	0.0	0.0-0.0	0.83	0.0-1.97	1.00	0.67-1.97	0.0103
IL-21	0.0	0.0-4.29	14.1	7.57-24.4	10.06	1.59-22.87	0.0022
IL-22	0.0	0.0-0.85	42.8	28.1-60.0	31.55	14.53-57.76	<0.0001
IL-23	0.0	0.0-0.0	0.0	0.0-19.2	1.00	1.0-19.18	0.0978
IL-27	15.0	5.78-42.6	173	98.5-215	13.88	3.6-36.88	<0.0001
IL-31	0.0	0.0-0.0	102	2.65-155	102.16	2.9-155.40	<0.0001
IFN $\alpha$	0.0	0.0-0.0	1.01	0.0-1.88	1.03	0.91-1.88	0.0009
IFN $\gamma$	5.38	3.5-7.52	19.1	15.4-118	5.56	3.14-19.21	<0.0001
TNF $\beta$	0.0	0.0-0.0	36.7	9.99-69.8	36.68	9.99-69.78	<0.0001

MIF	12.2	7.94-42.4	59.7	49.4-74.6	4.46	1.98-8.91	0.0025
TSLP	0.385	0.0-4.18	12.3	8.0-15.9	9.35	1.83-16.27	<0.0001
LIF	0.47	0.0-1.37	7.1	5.6-14.6	8.22	4.21-14.11	<0.0001
NGFF $\beta$	0.0	0.0-0.0	7.04	5.78-10.1	7.04	5.78-10.08	<0.0001
GM-CSF	0.0	0.0-9.23	42.2	24.1-53.1	33.23	3.91-53.08	<0.0001
G-CSF	5.86	0.0-14.9	14.8	10.4-17.6	2.43	0.80-15.4	0.0165
M-CSF	0.0	0.0-0.0	258	221-323	250.83	204.05-307.81	<0.0001
<b>Cytokines – lower in CSF compared to blood</b>							
IL-2R	1605	449-4096	1525	885-28,338	2.02	0.67-7.45	0.2649
<b>Chemokines – higher in CSF compared to blood</b>							
GRO $\alpha$ (CXCL1)	0.0	0.0-4.32	5.89	2.99-13.1	5.48	0.63-17.05	0.0026
IL-8 (CXCL8)	0.0	0.0-0.39	51.1	29.2-212	51.15	21.01-85.70	<0.0001
MIG (CXCL9)	6.53	0.0-123	291	62.2-1278	42.0	2.51-189.95	0.0003
IP-10 (CXCL10)	39.5	27.7-143	834	186-1198	10.31	5.14-21.19	<0.0001
BLC (CXCL13)	291	125-484	1354	431-3281	4.22	2.13-16.34	0.0067
MCP1 (CCL2)	23.8	15.2-38.5	146	95.3-175	5.00	3.29-15.90	<0.0001
MIP A $\alpha$ (CCL3)	3.05	0.0-6.20	10.4	7.37-14.2	4.59	2.03-7.85	0.0006
MIP 1 $\beta$ (CCL4)	6.6	0.0-16.0	15.7	11.8-30.9	2.56	1.41-13.29	0.0117
MCP3 (CCL7)	3.12	0.0-10.8	11.9	9.43-13.2	3.56	0.90-13.23	0.0010
MCP2 (CCL8)	4.32	1.97-8.24	14.7	2.78-80.7	3.47	0.88-14.09	0.0357
EOTAXIN-3 (CCL26)	0.41	0.0-2.50	2.42	1.29-3.07	1.84	0.77-5.02	0.0451
FRACTALKINE (CX3CL1)	3.17	0.0-10.7	9.21	6.99-14.7	2.79	0.75-8.35	0.0278
<b>Chemokines – lower in CSF compared to blood</b>							
ENA-78 (CXCL5)	69.6	31.3-112	4.2	3.13-16.6	0.10	0.03-0.64	<0.0001
EOTAXIN (CCL11)	11.0	6.32-25.5	3.37	2.55-4.47	0.34	0.17-0.47	0.0009
I-TAC (CXCL11)	40.6	24.4-78.1	20.6	4.76-265	0.34	0.17-3.22	0.3346
MIP 3 $\alpha$ (CCL20)	14.2	1.94-28.9	13.2	8.06-21.5	0.93	0.37-7.58	0.9729
MDC (CCL22)	48.8	31.0-83.5	25.2	17.7-126	0.76	0.35-1.61	0.2408
EOTAXIN-2 (CCL24)	27.7	14.1-199	26.2	15.8-59.4	0.54	0.36-1.23	0.7097
<b>TNF superfamily – higher in CSF compared to blood</b>							
TNF $\alpha$	0.0	0.0-0.0	6.44	5.28-7.75	6.52	5.50-7.75	<0.0001

TNF RII	44.5	27.7-88.6	178	80.9-211	2.88	1.51-4.56	0.0013
Tweak/TNFSF12	155	73.4-358	2379	1413-4128	20.78	7.11-32.23	<0.0001
CD30	249	111-405	645	339-1146	2.45	1.60-7.38	0.0067
APRIL	197	91.8-436	2059	932-4635	7.66	5.79-18.79	<0.0001
BAFF	0.615	0.0-24.7	50.1	34.6-57.5	29.73	1.47-52.04	<0.0001
<b>TNF superfamily – lower in CSF compared to blood</b>							
TRAIL	23.7	11.5-38.2	19.5	17.1-37.1	1.17	0.51-2.04	0.9100
<b>Other factors – higher in CSF compared to blood</b>							
VEGF-A	45.5	23.1-119	109	93.4-235	3.29	0.78-13.26	0.0067
HGF	35.4	20.4-59.8	338	196-431	9.32	4.32-12.18	<0.0001
MMP1	5.32	3.03-11.4	11.6	8.15-46.5	2.96	1.27-15.01	0.0091
SCF	1.72	1.08-2.34	6.34	4.78-8.44	4.15	2.61-6.45	<0.0001
SDF-1 $\alpha$	709	344-1743	1974	819-4076	1.69	1.16-8.42	0.0141
FGF-2	0.0	0.0-0.0	7.40	2.68-10.4	5.47	2.68-9.78	<0.0001
CD40L	0.0	0.0-2.16	1.81	0.0-4.81	1.00	0.48-4.15	0.1789

**Table 5.5 Baseline cerebrospinal fluid (CSF) and blood (BI) mediator concentrations, and CSF:Blood ratios in patients with HIV associated TBM in those with good (MRS 0-3) vs poor (MRS 4-6) outcome**

Mediator	HIV associated TBM Good Outcome (MRS 0-3), n=11							HIV associated TBM Poor Outcome (MRS 4-6), n=3							Good outcome vs Poor Outcome*		
	Blood		CSF		CSF vs Blood	CSF: Blood ratio		Blood		CSF		CSF vs Blood	CSF:Blood ratio		Blood	CSF	CSF:Blood ratio
	pg/ml *	median	(IQR)	median	(IQR)	p value	median	IQR	median	(IQR)	median	(IQR)	p value	median	IQR	Direction and p-value	Direction and p-value*
<b>Cytokines</b>																	
IL-1 $\beta$	0	0-3.95	<b>42.6</b>	<b>19.5-70.1</b>	<b>&lt;0.0001</b>	17.74	9.44-17.74	6.38	4.97-8.21	<b>18.7</b>	<b>15.3-70.1</b>	0.1876	3.76	1.87-3.76	<b>↑0.0137</b>	0.5181	0.0604
IL-4	0	0-0	<b>46.2</b>	<b>27.5-58.6</b>	<b>0.0002</b>	46.21	46.21	11.5	0-14.6	<b>42.3</b>	<b>23.6-54.9</b>	<b>0.0354</b>	4.78	1.62-4.78	<b>↑0.0330</b>	0.9771	0.1389
IL-15	1.65	0-3.01	<b>17.8</b>	<b>11.6-21.1</b>	<b>&lt;0.0001</b>	10.81	4.33-10.81	7.34	3.1-8.06	<b>17.1</b>	<b>11.8-18.1</b>	<b>0.0186</b>	2.24	1.51-2.24	<b>↑0.0092</b>	0.6288	0.0733
IL-27	11.4	0-18.5	<b>184</b>	<b>94.1-221</b>	<b>&lt;0.0001</b>	20.94	5.07-20.94	45.0	41.8-63.8	<b>138</b>	<b>100-190</b>	<b>0.0266</b>	2.39	2.16-2.39	<b>↑0.0008</b>	0.5429	<b>↓0.0220</b>
IFN $\gamma$	4.64	3.26-5.91	<b>19.1</b>	<b>15.4-62.6</b>	<b>&lt;0.0001</b>	5.26	3.30-5.26	8.48	7.84-11.5	<b>207</b>	<b>15.8-468</b>	0.1671	24.38	2.02-24.38	<b>↑0.0014</b>	0.1978	0.1324
LIF	0.35	0-1.22	<b>6.40</b>	<b>4.44-11.9</b>	<b>&lt;0.0001</b>	7.79	4.44-7.79	2.57	0-3.59	<b>32.7</b>	<b>5.99-60.3</b>	0.1203	12.72	1.67-12.72	<b>↑0.0311</b>	0.1786	0.6593
GM-CSF	0	0-0	<b>51.4</b>	<b>24.9-54.1</b>	<b>&lt;0.0001</b>	51.43	21.67-51.43	20	0-20	<b>25.4</b>	<b>20.6-42.9</b>	0.1625	2.14	1.03-2.14	<b>↑0.0385</b>	0.2261	0.0606
MIF	13.8	9.15-37.8	<b>59.4</b>	<b>47.4-73.7</b>	<b>0.0233</b>	3.59	0.93-3.59	4.46	1.98-56.1	<b>83.4</b>	<b>51.3-153</b>	0.0975	11.51	2.74-11.51	0.2912	<b>↑0.0413</b>	0.1621
G-CSF	5.66	0-11.4	<b>14.8</b>	<b>7.51-16.9</b>	<b>0.0240</b>	2.44	0.67-2.44	14.2	0-21.7	<b>25.5</b>	<b>12.8-35.3</b>	0.2396	1.63	0.90-1.63	0.3926	<b>↑0.0137</b>	0.7473
IL-1 $\alpha$	0	0-0.37	<b>5.26</b>	<b>4.55-9.46</b>	<b>0.0001</b>	5.26	3.55-5.26	0	0.711	<b>11.53</b>	<b>0.80-11.53</b>	0.1854	0.95	0.78-0.95	0.6291	0.1648	0.5549
IL-2	0	0-0	<b>58.4</b>	<b>20.4-71.9</b>	<b>&lt;0.0001</b>	58.38	20.44-58.38	0	0-5.6	<b>30.4</b>	<b>0-36.8</b>	0.1488	30.42	0.18-30.42	0.2143	0.1231	0.1205

IL-2R	2758	445-7598	<b>1437</b>	<b>919-18353</b>	0.6063	1.90	0.50-1.90	1258	450-1943	<b>28125</b>	<b>780-203599</b>	0.2962	22.37	1.73-22.37	0.5549	<b>0.4560</b>	0.1703
IL-3	0	0-0	<b>38.3</b>	<b>9.54-59.4</b>	<0.0001	39.32	9.54-38.42	0	0-1.15	<b>29.6</b>	<b>17.3-37.7</b>	<b>0.0094</b>	29.61	17.32-29.61	0.3956	<b>0.5238</b>	0.5009
IL-5	0	0-0	<b>0</b>	<b>0-0</b>	0.4762	0.0	0.0-8.42	0	0-0	<b>6.69</b>	<b>0-23.9</b>	0.2255	0.0	0.0-0.0	n/a	<b>0.1209</b>	0.1209
IL-6	0	0-0	<b>5.19</b>	<b>0-127</b>	<b>0.0010</b>	5.19	1.00-5.19	0	0-2.77	<b>591</b>	<b>1.03-3825</b>	0.2834	591.31	0.37-591.31	0.2143	<b>0.2170</b>	0.4341
IL-7	0.22	0.09-1.18	<b>1.46</b>	<b>0.74-2.34</b>	<b>0.0077</b>	5.44	1.46-5.44	0.53	0-1.16	<b>1.12</b>	<b>0.87-1.81</b>	0.1832	1.56	1.12-1.56	0.9038	<b>0.5657</b>	0.2253
IL-9	0	0-0	<b>30.3</b>	<b>14.7-37.4</b>	<b>0.0003</b>	30.25	14.65-30.25	0	0-6.43	<b>19.3</b>	<b>17.3-24.3</b>	<b>0.0037</b>	17.28	3.78-17.28	0.9167	<b>0.4498</b>	0.2119
IL-10	0	0-0	<b>18.4</b>	<b>12.7-28.7</b>	<0.0001	18.42	12.65-18.42	0	0-0	<b>17.9</b>	<b>11.9-23.0</b>	<b>0.0054</b>	17.89	11.88-17.89	n/a	<b>0.6868</b>	0.6868
IL-12P70	0	0-0	<b>2.93</b>	<b>1.34-3.31</b>	<b>0.0002</b>	2.83	1.34-2.83	0	0-0	<b>1.46</b>	<b>0-2.63</b>	0.1476	1.46	0.0-1.46	n/a	<b>0.3161</b>	0.1817
IL-13	0	0-0	<b>5.09</b>	<b>3.36-6.38</b>	<0.0001	5.09	3.36-5.09	0	0-0	<b>4.83</b>	<b>1.87-6.76</b>	<b>0.0343</b>	4.83	1.87-4.83	n/a	<b>0.9322</b>	0.9322
IL-16	55.9	36.7-102	<b>77.1</b>	<b>40.1-234</b>	0.2169	2.99	0.54-2.99	27.4	23.6-85.7	<b>308</b>	<b>39.2-2371</b>	0.3078	11.22	1.66-11.22	0.4287	<b>0.5549</b>	0.1264
IL-17A	0.00	0-3.19	<b>52.6</b>	<b>37.9-77.2</b>	<0.0001	37.86	13.94-37.86	0.00	0-19.8	<b>46.1</b>	<b>45.3-59.4</b>	0.100	45.29	3.0-45.29	0.7527	<b>0.9780</b>	0.6273
IL-18	17.4	13.2-23.7	<b>47.3</b>	<b>31.4-49.0</b>	<0.0001	2.66	1.65-2.66	20.1	17.4-20.6	<b>56.0</b>	<b>32.6-125</b>	0.1356	2.79	1.58-2.79	0.8887	<b>0.1232</b>	0.3294
IL-20	0	0-0	<b>0.83</b>	<b>0-2.43</b>	<b>0.0039</b>	1.0	0.83-1.0	0	0-7.39	<b>0.83</b>	<b>0-1.82</b>	0.5645	1.0	0.11-1.0	0.2143	<b>0.9121</b>	0.5619
IL-21	0	0-0	<b>11.7</b>	<b>4.92-20.0</b>	<b>0.0080</b>	8.45	1.93-8.45	0	0-17.1	<b>31.4</b>	<b>9.62-39.9</b>	0.1172	31.42	0.56-32.52	>0.999	<b>0.0780</b>	0.1341
IL-22	0	0-3.39	<b>48.1</b>	<b>30.2-58.4</b>	<b>0.0006</b>	30.15	10.80-30.15	0	0-0	<b>33.0</b>	<b>22.1-69.6</b>	<b>0.0444</b>	32.95	22.11-32.95	0.5467	<b>0.8500</b>	0.5241
IL-23	0	0-0	<b>0</b>	<b>0-30</b>	0.0902	1.0	1.0-1.0	0	0-0	<b>0</b>	<b>0-0</b>	n/a	1.0	1.0-1.0	n/a	<b>0.5055</b>	0.5055
IL-31	0	0-0	<b>110</b>	<b>4.71-178</b>	<0.0001	110.19	4.71-110.19	0	0-0	<b>0</b>	<b>0-97.4</b>	0.3739	1.0	1.0-1.0	n/a	<b>0.2073</b>	0.2098
IFN $\alpha$	0	0-0	<b>1.06</b>	<b>0-1.91</b>	<b>0.0039</b>	1.06	1.0-1.06	0	0-0	<b>0.95</b>	<b>0.78-1.87</b>	<b>0.0239</b>	1.0	1.0-1.0	n/a	<b>0.9401</b>	0.7556



TNF $\beta$	0	0-0	<b>45.6</b>	<b>10.7-78.3</b>	<b>&lt;0.0001</b>	45.57	10.68-45.57	0	0-0	<b>12.4</b>	4.31-12.4	0.2150	12.41	4.31-12.41	n/a	0.3923	0.3923
TSLP	0	0-2.71	<b>12.2</b>	<b>7.51-16.0</b>	<b>&lt;0.0001</b>	10.53	4.70-10.53	4.32	0-8.49	<b>12.5</b>	8.16-15.9	0.0758	1.89	1.47-1.89	0.0669	0.8576	0.4074
NGFF $\beta$	0	0-0	<b>7.04</b>	<b>6.54-10.9</b>	<b>&lt;0.0001</b>	7.04	6.54-7.04	0	0-0	<b>8.10</b>	3.49-9.49	0.0179	8.10	3.49-8.10	n/a	0.8330	>0.9999
M-CSF	0	0-0	<b>251</b>	<b>207-303</b>	<b>&lt;0.0001</b>	250.83	196.50-250.83	0	0-0	<b>322</b>	226-448	<b>0.0066</b>	321.78	226.43-321.78	>0.999	0.1679	0.1252
<b>Chemokines</b>																	
IL-8 (CXCL8)	0	0-0	<b>45.1</b>	<b>22.2-65.7</b>	<b>&lt;0.0001</b>	45.06	17.43-45.06	0	0-5.3	<b>342</b>	<b>31.6-382</b>	0.0869	64.50	31.58-64.50	0.69	<b><math>\uparrow</math>0.0251</b>	0.4560
IP-10 (CXCL10)	40.6	29.4-139	<b>617</b>	<b>167-1136</b>	<b>0.0010</b>	7.96	4.24-7.96	28.9	17.5-153	<b>1318</b>	<b>1002-1814</b>	0.0055	34.65	11.85-34.65	0.7266	<b><math>\uparrow</math>0.0385</b>	<b><math>\uparrow</math>0.0076</b>
MIP A $\alpha$ (CCL3)	1.35	0-4.9	<b>10.2</b>	<b>7.38-11.2</b>	<b>0.0029</b>	7.38	2.00-7.38	5.39	2.42-8.63	<b>26.9</b>	<b>6.82-36.2</b>	0.1142	4.19	2.82-4.19	0.2720	<b><math>\uparrow</math>0.0147</b>	0.5290
ENA-78 (CXCL5)	71.5	33.6-120	<b>4.0</b>	<b>3.18-7.19</b>	<b>&lt;0.0001</b>	0.1	0.03-0.64	36.5	8.21-110	<b>26.4</b>	<b>2.96-73.8</b>	0.6662	0.72	0.03-0.72	0.5549	<b>0.3544</b>	<b><math>\uparrow</math>0.0413</b>
BLC (CXCL13)	347	180-568	<b>830</b>	<b>226-2926</b>	0.0759	3.88	0.51-3.88	127	85.3-444	<b>3275</b>	<b>698-8532</b>	0.1622	19.24	5.51-19.24	0.3196	<b>0.1213</b>	<b><math>\uparrow</math>0.0159</b>
MCP2 (CCL8)	3.16	1.76-10.5	<b>13.3</b>	<b>2.73-39.5</b>	0.2228	2.81	0.51-2.81	2.93	0-6.44	<b>81.3</b>	<b>13.5-327</b>	0.2286	31.11	4.26-31.11	0.7692	<b>0.1264</b>	<b><math>\uparrow</math>0.0070</b>
MDC	50.4	28.4-81.3	22.7	17.7-86.5	0.1461	0.72	0.33-0.72	13.8	9.15-37.8	120	16.9-528	0.3429	3.77	0.47-3.77	0.8846	<b>0.5220</b>	<b><math>\uparrow</math>0.0078</b>
GRO $\alpha$ (CXCL1)	0	0-2.86	<b>5.41</b>	<b>2.62-8.75</b>	<b>0.0059</b>	4.59	0.64-4.59	0.59	0-12	<b>8.31</b>	<b>3.11-62.4</b>	0.3508	14.08	0.26-14.08	0.4286	<b>0.4560</b>	0.6593
MIG (CXCL9)	2.23	0-130	<b>236</b>	<b>30.3-1253</b>	<b>0.0023</b>	10.43	2.04-10.43	10.8	0-121	<b>1103</b>	<b>330-1289</b>	0.1122	72.87	11.12-72.87	>0.999	<b>0.2642</b>	0.8846
I-TAC (CXCL11)	58.4	16.4-87.9	<b>9.79</b>	<b>4.3-72.8</b>	0.1330	0.34	0.17-3.22	35.9	27.1-41.1	<b>385</b>	<b>11.9-1435</b>	0.2477	14.20	0.33-14.2	0.3655	<b>0.0879</b>	0.0879
MCP1 (CCL2)	23.4	16.3-37.7	<b>138</b>	<b>94.2-163</b>	<b>&lt;0.0001</b>	5.28	2.69-5.28	4.88	2.02-7.48	<b>178</b>	<b>95.7-434</b>	0.1086	4.73	4.09-4.73	0.9301	<b>0.3681</b>	0.7692
MIP 1 $\beta$ (CCL4)	5.52	0-19.4	<b>14.2</b>	<b>12-30.5</b>	0.0876	2.50	1.27-2.50	7.67	0-12.7	<b>33.4</b>	<b>11.1-51.9</b>	0.1094	2.62	1.45-2.62	0.6003	<b>0.1229</b>	0.2026
MCP3 (CCL7)	2.93	0-6.44	<b>12.4</b>	<b>8.58-31.1</b>	<b>0.0003</b>	3.73	1.18-3.73	10.8	0-12.7	<b>11.4</b>	<b>9.71-94.6</b>	0.3379	0.90	0.90-0.90	0.2361	<b>0.8077</b>	0.8626

EOTAXIN (CCL11)	11.3	7.14-25.5	<b>3.13</b>	<b>2.55-3.78</b>	<b>0.0008</b>	0.33	0.16-0.33	8.44	3.67-35.9	<b>4.90</b>	<b>2.96-14</b>	0.4582	0.39	0.35-0.39	0.8110	<b>0.1813</b>	0.1703
MIP 3 $\alpha$ (CCL20)	7.9	0-19.4	<b>13.9</b>	<b>6.47-21.2</b>	0.6622	1.0	0.34-1.0	31.4	14.5-36.9	<b>12.6</b>	<b>11.9-527</b>	0.4144	0.87	0.38-0.87	0.1236	<b>0.5330</b>	>0.9999
EOTAXIN-2 (CCL24)	32.6	14.3-184	<b>27.1</b>	<b>15.8-49.8</b>	0.4887	0.44	0.30-0.44	19.9	0.68-416	<b>25.3</b>	<b>15.6-250</b>	0.7714	0.79	0.60-0.79	0.8846	<b>0.8846</b>	0.2253
EOTAXIN-3 (CCL26)	0.5	0-2.35	<b>2.35</b>	<b>1.3-2.78</b>	0.1115	2.16	0.61-2.16	0.28	0-2.94	<b>3.03</b>	<b>1.16-4.49</b>	0.2471	1.53	1.16-1.53	0.8049	<b>0.4508</b>	0.6593
FRACTALKINE (CX3CL1)	1.95	0-9.65	<b>8.39</b>	<b>6.4-10.4</b>	<b>0.0327</b>	3.28	0.76-3.28	9.87	1.95-14.3	<b>16.3</b>	<b>7.18-19.2</b>	0.3434	1.34	0.73-1.34	0.2198	<b>0.0821</b>	0.6593
<b>TNF superfamily related</b>																	
BAFF	0.00	0-16.8	<b>50.1</b>	<b>33-55</b>	<b>&lt;0.0001</b>	33.02	2.75-33.02	25.8	24.3-38.7	<b>50.1</b>	<b>37.3-103</b>	0.1751	1.54	1.29-1.54	<b><math>\uparrow</math>0.0192</b>	<b>0.2453</b>	0.0544
APRIL	267	96.9-411	<b>1749</b>	<b>908-3772</b>	<b>&lt;0.0001</b>	7.19	5.46-7.19	120	32.5-563	<b>6779</b>	<b>940-16871</b>	0.1626	29.99	7.81-29.99	0.8564	<b><math>\uparrow</math>0.0253</b>	0.0879
TRAIL	20.6	10.9-39.7	<b>19.3</b>	<b>17.4-24.2</b>	0.6023	0.99	0.52-0.99	35.4	16-37.7	<b>57.2</b>	<b>16.5-67.4</b>	0.3649	1.90	0.44-1.90	0.6262	<b><math>\uparrow</math>0.0140</b>	0.3963
TNF $\alpha$	0	0-0	<b>6.27</b>	<b>5.08-7.53</b>	<b>&lt;0.0001</b>	6.27	5.08-6.27	0	0-0.83	<b>6.68</b>	<b>5.34-18.8</b>	0.0806	6.68	6.43-6.68	0.2143	<b>0.0969</b>	0.0635
TNF RII	49.1	30.7-72.4	<b>164</b>	<b>72.2-206</b>	<b>0.0065</b>	1.66	1.34-1.66	19.8	19.6-156	<b>221</b>	<b>83.8-276</b>	0.1535	4.27	1.77-4.27	0.3681	<b>0.3019</b>	0.1398
Tweak/TNFSF12	162	74.7-328	<b>2539</b>	<b>1394-4107</b>	<b>&lt;0.0001</b>	16.43	7.83-16.43	149	55.2-448	<b>220</b>	<b>1722-4193</b>	<b>0.0309</b>	28.12	4.96-28.12	>0.999	<b>0.7692</b>	0.8846
CD30	279	112-558	<b>473</b>	<b>325-1029</b>	<b>0.0473</b>	2.09	1.17-2.09	142	50.7-345	<b>818</b>	<b>344-2995</b>	0.2155	6.80	2.37-6.80	0.3681	<b>0.2546</b>	0.1703
<b>Other factors</b>																	
SCF	1.55	1.03-2.22	<b>5.51</b>	<b>4.74-8.33</b>	<b>&lt;0.0001</b>	3.77	2.32-3.77	2.38	1.83-3.12	<b>8.28</b>	<b>6.44-25.6</b>	0.1467	4.52	2.71-4.52	0.2745	<b><math>\uparrow</math>0.0365</b>	0.7692
SDF-1 $\alpha$	738	490-1405	<b>1919</b>	<b>817-3685</b>	0.0652	1.33	0.96-1.33	182	0-3784	<b>5511</b>	<b>1389-7334</b>	0.1861	7.64	1.94-7.64	0.5549	<b><math>\uparrow</math>0.0492</b>	0.1264
VEGF-A	47.3	21.1-165	<b>103</b>	<b>96.9-172</b>	0.0879	2.36	0.69-2.36	42.3	30.1-66.2	<b>401</b>	<b>82.9-560</b>	0.0984	13.23	1.25-13.23	0.4397	<b>0.3681</b>	0.4560

HGF	36.9	26.9-59.3	<b>314</b>	<b>139-385</b>	<b>&lt;0.0001</b>	8.48	3.33-8.48	21.6	16.7-140	<b>417</b>	<b>214-1590</b>	0.1889	11.32	9.94-11.32	0.7692	<b>0.0586</b>	0.2253
MMP1	4.86	2.84-11.9	<b>10.2</b>	<b>7.74-30.6</b>	<b>0.0192</b>	3.29	1.19-3.29	8.88	4.16-11.3	<b>13.6</b>	<b>10.9-453</b>	0.3632	2.62	1.53-2.62	0.5549	<b>0.2912</b>	0.3395
FGF-2	0	0-0	<b>7.69</b>	<b>2.82-10.5</b>	<b>&lt;0.0001</b>	7.10	2.82-7.10	0	0-0	<b>3.83</b>	<b>0-9.57</b>	0.1835	3.83	1.00-3.83	>0.999	<b>0.3785</b>	0.5169
CD40L	0	0-2.05	<b>0.46</b>	<b>0-4.64</b>	0.2192	1.0	0.46-1.0	0	0-7.51	<b>3.64</b>	<b>0-5.30</b>	0.8796	1.0	0.71-1.0	0.7527	<b>0.8242</b>	0.6257

\*If parametric, unpaired T Test; if non-parametric, Mann-Whitney

\*\* Arrow represents direction in those with poor versus good outcome i.e. if higher in those with poor outcome ↑; if lower in those with poor outcome ↓. Only indicated for those with statistically significant differences (p<0.05)

## Chapter 6. Magnetic resonance

### spectroscopy to detect GABA and Glutamate in HIV-associated Tuberculous Meningitis

#### 6.1 Introduction

Nuclear magnetic resonance (NMR) refers to the behaviour of atoms when subjected to a magnetic field and was first described in 1946 by Bloch and Purcell (Packard et al., 1946, Purcell et al., 1946). In biomedical research this principle can be applied via a range of techniques including structural magnetic resonance imaging (MRI) and magnetic resonance spectroscopy (MRS). The latter encompasses a diverse set of methods, including via *in vivo* clinical systems to measure metabolism of well-defined regions of the human body, including the brain. When combined with structural MRI in this context, the tool obtains functional metabolic information in addition to anatomical information within the area of interest.

The basic physical principles of MRS are complex, but the central underlying concept relates to 'nuclear spin' which can be understood as the nucleus of an atom spinning around its own axis. In a large magnetic field, nuclei with spin can undergo resonance and emit a characteristic electromagnetic signal. In  $^1\text{H}$  MRS, radiofrequency signals arising from the hydrogen atom within tissue metabolites are detected. These signals possess chemically specific frequencies which is determined by the environment (chemical shift) (Tognarelli et al., 2015). These signals are expressed as an MR spectrum i.e. peaks occurring at different frequencies; a plot of signal intensity, proportional to metabolite concentration, against

chemical shift (Figure 6.1). The most important visible peaks on the *in vivo* cerebral proton MR spectra using field strengths of 1.5 to 3.0 Tesla (T) are N-acetyl aspartate (NAA), choline (cho), creatine (Cr) myo-inositol (ml), the combined glutamine and glutamate peak (Glx) and lactate (Lac). More complex to acquire is information regarding concentration of GABA, however validated methods exist to isolate GABA signals from the spectra (Puts and Edden, 2012).

Glutamate is a neuroexcitatory neurotransmitter which acts on receptors classed as either ionotropic (i.e. ligand gated ion channels) or metabotropic (i.e. G protein coupled) on post-synaptic cells, astrocytes, oligodendrocytes and glial cells (Dingledine et al., 1999). Extracellular glutamate levels are regulated by the glutamate-glutamine cycle to prevent excessive glutamate and its associated toxicity (Lipton and Rosenberg, 1994). The balance of glutamate/glutamine and GABA, a neuroinhibitory neurotransmitter, is crucial in maintaining normal neurological function. Prolonged glutamate exposure results in neuronal injury and damage via influx of  $\text{Ca}^{2+}$  and  $\text{Na}^{+}$  ions across the membrane. This is worsened by the activation of GTP-binding protein-coupled glutamate receptors which stimulate  $\text{Ca}^{2+}$  release from the endoplasmic reticulum. When neurons are under oxidative or metabolic stress e.g. due to ischaemia, these glutamate-driven excitotoxic processes can occur even in the context of normal concentrations of glutamate (Gamir-Morralla et al., 2017).

Glutamate driven neuro-excitotoxicity is a key driver of pathogenesis in many neurological conditions including Alzheimer's Disease and traumatic brain injury (Butterfield and Pocernich, 2003, Chamoun et al., 2010). In TBM, a study of ventricular CSF in children with TBM found that transcripts

associated with glutamate release, post synaptic NMDA receptor binding and  $\text{Ca}^{2+}$  influx were enriched, suggesting that in acute TBM neuro-excitotoxicity may be driven by glutamate release (Rohlwink et al., 2019). A more recent study, also in paediatric TBM supported this hypothesis using  $^1\text{H}$  metabolomics demonstrating that glutamine and glutamate are elevated in the CSF at disease presentation (van Zyl et al., 2020). Although glutamate and GABA have been evaluated *in vivo* using MRS in many central nervous system diseases including neuro-oncology (Hazany et al., 2007), Alzheimer's disease (JM Scott et al., 2011), psychiatric diseases (Mason and Krystal, 2006), and multiple sclerosis (Cianfoni et al., 2007b), no studies have evaluated its potential in TBM. If feasible, this non-invasive technique would allow correlation of *ex vivo* findings and add depth of understanding relating to the glutamate-glutamine cycle and neuro-excitotoxicity in TBM. It could also, albeit in well-resourced settings, serve as a potentially useful biomarker of clinical outcome, as has been explored in traumatic brain injury (Shutter et al., 2004).

In a pilot study using  $^1\text{H}$ -MRS in adults with HIV-associated TBM we aimed to examine *in vivo* concentrations of glutamate/glutamine (hitherto referred to as Glx) and GABA within the brain and compare these to a group of healthy control subjects. Our hypotheses were:

- i. Magnetic Resonance Spectroscopy is a feasible mechanism to measure *in vivo* concentrations of Glx and GABA in HIV-associated TBM
- ii. Glx would be found in higher concentrations whereas GABA would be found in lower concentrations, in the acute phase of HIV-associated TBM compared to healthy controls

## **6.2 Methods**

This study was approved by the University of Cape Town Human Ethical Research Committee (HREC 293/2018), including the enrolment of up to 10 healthy controls for comparison analysis.

### **Participants**

Participants enrolled to the LASER-TBM study (see chapter 2) were consented for imaging including MRI and MRS at two timepoints; enrolment and day 56. This consent was part of the main study consent and therefore although all enrolled participants were eligible for inclusion, imaging only took place when participants were well enough to travel from the referring hospital site to the University of Cape Town's imaging facility. Participants were required to complete an MRI questionnaire prior to imaging being performed those with contra-indications for MRI were not imaged.

A total of 9 healthy controls provided written informed consent to be included in this study. The study was advertised on the University of Cape Town Health Sciences campus, and participants enrolled were staff and students at the university. Prior to inclusion in the study, any volunteers were required to complete the MRI safety questionnaires; those with contraindications were not included within the study. HC underwent a single timepoint scan.

### **Neuroimaging**

Participants were scanned on a 3T Siemens Skyra MRI scanner at CUBIC using a 32- channel head coil. No sedation was given. Two healthy control volunteers were scanned to evaluate feasibility prior to imaging patients with TBM. These data were analysed and the protocol refined prior to scanning of patients with TBM. Remaining healthy volunteer scans (n=7) were performed following completion of the LASER-TBM participant scans.

#### *Voxel placement*

Voxel placement was considered in the context of the common anatomical location of vascular injury in TBM. Specifically, infarcts within the tubercular zone account for around 60% of strokes in TBM (Misra et al., 2011). The tubercular zone comprises the caudate, anterior thalamus, anterior limb and genu of the internal capsule. Its vulnerability to microvascular damage is likely due to the exudate affecting the vessel wall architecture in the medial striate, thalamotuberal and thalamostriate arteries which supply this area. It is also an area vulnerable to further ischaemic injury in the hydrocephalic patients. Therefore, placement of the voxel within this location was thought most likely to yield interesting results (Figure 6.2).

#### *Imaging protocol*

An imaging protocol was developed in collaboration with an MRI physicist (Dr Frances Robertson), a physicist (Professor Ernesta Meintjes) and senior radiographer (Petronella Samuels). A structural high resolution T1-weighted magnetisation prepared rapid gradient echo acquisition was acquired for voxel placement with the following variables: Field of View (FOV) 230 mm × 230 mm, Repetition Time (TR) 1720 ms, Inversion Time (TI) 900 ms, Echo Time (TE) = 2.47 ms, flip angle 8 degrees, bandwidth



250 Hz/px, 176 slices, resolution 1 mm × 1.0 mm × 1.0 mm (duration 3 minutes 30s). This was followed by 2 single voxel point-resolved spectroscopy (PRESS) acquisitions from the same voxel with Chemical Shift Selective (CHESS) water suppression: TR=2000 ms, TE =30s, spectral bandwidth 2000Hz, vector size 1024, 64 averages, voxel size 3 x 3 x 3 cm<sup>3</sup>. A water reference (2 averages) was acquired in the same voxel without water suppression for subsequent analysis (total duration 2 and a half minutes). Single Voxel Spectroscopy (SVS) acquisitions were then obtained from a 3 x 3 x 3 cm voxel in the same region using Mescher-Garwood point-resolved spectroscopy (MEGAPRESS)(Mikkelsen et al., 2017): TR=2000ms, TE=80ms, 320 averages, bandwidth 2000 Hz, vector size 2048 with ON editing pulse at 1.9 ppm targeting GABA (duration 30 minutes)

### **Statistical Analysis**

We used MEGAPRESS (for GABA) and PRESS (for Glx) acquisitions and subsequently GANNET software(Edden et al., 2014) to estimate metabolite concentrations within the tubercular zone. Metabolite concentrations were measured in institutional units (IU). A combined Glutamine/Glutamate (Glx) concentration was used as a proxy for glutamate concentration, given that these are challenging to differentiate, consistent with previous studies in the literature. Concentrations were corrected to account for the proportion of grey and white matter, as well as CSF within the voxel using published methods(Harris et al., 2015). These ‘tissue-alpha corrected’ concentrations for both Glx and GABA were subsequently used for analysis. We compared:

- i) Glx and ii) GABA concentrations in patients with TBM at baseline (within 5 days of enrolment) with healthy controls
- i) Glx and ii) GABA concentrations in patients with TBM at baseline and follow up timepoints (day 56 +/- 5 days).
- Glx: GABA ratios in patients with TBM at baseline compared to healthy controls.

Statistical analysis was performed using GraphPad Prism (version 9.3.1) software. We compared variables between groups using unpaired t Tests (parametric data) or Mann Whitney Tests of significance (non-parametric data). Throughout the analysis we used an unadjusted p value of <0.05 as a nominal threshold for statistical significance.

## 6.3 Results

A total of 11 participants with HIV-associated TBM enrolled to LASER-TBM completed the baseline MRI/MRS imaging protocol (Figure 6.3). 7 of these 11 participants were available for follow up imaging. 4 baseline scans were excluded from the analysis; 2 due to a file formatting error, 2 due to lack of water reference data, which made the data uninterpretable. Of the follow up scans performed, 3 were excluded for similar reasons, leaving 4 for analysis. Baseline characteristics are outlined in table 6.1. Age was not similar across groups, however gender was. The majority of TBM participants had mild disease.

### Glx concentrations

We found significantly higher concentrations in those with HIV-associated TBM at baseline compared to healthy controls (median (IQR) concentration

(IU) 15.9 (13.4-16.2) vs 12.2 (11.7-15.5),  $p=0.0428$  (Figure 6.4(A)). When we compared concentrations at baseline and follow up, we saw a reduction in concentration over time, however this did not reach statistical significance (median (IQR) concentration (IU) 15.9 (13.4-16.2) vs 14.6 (13.8-15.6),  $p=0.4136$ ). Although concentrations of Glx reduced within the first 2 months of treatment, there remained a trend towards raised levels when compared to healthy controls (median (IQR) concentration (IU) 14.65 vs 12.79,  $p=0.2190$ ) (Figure 6.4(B)).

### **GABA concentrations**

There was no statistically significant difference between concentrations of GABA in patients with TBM at baseline compared to healthy controls (median (IQR) concentration (IU) 3.79 (3.34-4.5) vs 4.02 (3.09-4.69),  $p=0.69$ ) (Figure 6.5(A)). Similarly, there was no significant change in concentration from baseline to day 56 those with TBM (median (IQR) concentration (IU) 3.79 (3.34-4.5) vs 3.789 (2.72-4.27),  $p=0.65$ ) (Figure 6.5(B)).

### **Glx:GABA concentration ratios**

We compared ratios of Glx and GABA in those with TBM (at baseline) and healthy controls as a proxy for degree of neuroexcitotoxicity and found a non-significantly higher ratio in those with TBM compared to healthy controls (median (IQR) concentration 3.98 (3.56-4.74) vs 3.31 (2.53-4.19),  $p=0.17$ ) (Figure 6.6).

### **Glx concentrations and ratios in relation to cognitive outcomes**

In the 7 participants in whom baseline data was available, only 3 participants had completed a full cognitive and functional outcome assessment at day 180 as detailed in chapter 4. We compared global T score as a continuous variable indicating cognitive performance and i) glutamate/glutamine concentrations at baseline and ii) the ratio of Glx:GABA concentrations at baselines using simple linear regression. Contrary to our central hypothesis that glutamate driven excitotoxicity may be a driver of poor outcome, we found higher glutamate/glutamine: GABA ratios were associated with better neuropsychological performance ( $p=0.032$ ) (Figure 6.7). None of these three participants in whom cognitive data was available had paired timepoint MRS data for comparison of glutamate levels over time.

## 6.4 Discussion

Within this proof-of-concept study, we demonstrated that *in vivo* measurement of Glx and GABA using  $^1\text{H}$  MRS is feasible via placement of a single voxel within the deep grey matter of the brain. Within our small cohort, our results suggest that i) Glx concentrations and ii) Glx:GABA ratios are significantly elevated in those with HIV-associated TBM around the time of their diagnosis, when compared to a group of healthy controls. They also show that over a 2 month period, concentrations of glutamate/glutamine decrease, but remain raised compared to healthy controls. As glutamate/glutamine are neuroexcitatory neurotransmitters, and Glx:GABA ratios a proxy for neuroexcitation, this study begins to support findings within the literature (van Zyl et al., 2020, Rohlwick et al.,

2019) which suggest that glutamate driven neuroexcitotoxicity may be a pathogenic mechanism in the acute phase of HIV-associated TBM. Larger studies are now required to validate these findings.

Although our findings of increased glutamate are in keeping with the TBM literature on this topic, it is interesting to consider them within the context of other diseases in which MRS is widely used and in which glutaminergic dysfunction is thought to be a key pathogenic mechanism. For example, in Alzheimer's disease glutaminergic dysfunction is thought to occur via A $\beta$  and tau mediated disruption of glutamate receptor function leading to glutamate accumulation within the synaptic and extra-synaptic spaces (Yeung et al., 2021). Studies using MRS, however, have shown that glutamate is *reduced* in the brains of patients with Alzheimer's, a finding which is thought to represent neuronal loss and injury (Rupsingh et al., 2011). In contrast, a study of four patients with tumefactive demyelinating lesions, a highly active inflammatory form of multiple sclerosis, MRS demonstrated marked elevation of glutamate and glutamine peaks within focal lesions (Cianfoni et al., 2007a); however, in primary and secondary progressive forms of MS longitudinal studies using MRS demonstrate that glutamate and glutamine reduces over time (MacMillan et al., 2016). Our findings in the context of these studies support the hypothesis that in acute TBM, extracellular glutamate is present in excessive levels, potentially driving neuro-excitotoxicity. However, they also raise some important questions, namely what happens to glutamate levels following the acute phase of illness? Although we saw a reduction at 2 months within our study, would longer term imaging timepoints reveal lower levels of glutamate when compared to our healthy controls demonstrating neuronal loss and injury, as seen in neurodegenerative diseases? Given the role of

glutamate in learning and memory(Riedel et al., 2003), and its association with cognitive outcomes in conditions such as multiple sclerosis(Muhlert et al., 2014) and HIV(Mohamed et al., 2010), how would this correlate with cognitive function at longer term timepoints? Further exploration of these questions would allow us to consider whether MRS has a role as a predictive biomarker in TBM, as is the case in traumatic brain injury(Babikian et al., 2006, Ashwal et al., 2000, Eisele et al., 2020).

The number of participants in whom both MRS and detailed cognitive testing at 6 months was performed were too few to draw conclusions; nonetheless in these three participants our results were surprising. Namely we saw that baseline glutamate levels were lower in those with worse cognitive outcomes. Although this is most likely due to the small number of participants included within this analysis, it is also possible that in these three participants baseline cognitive function (which was not assessed) was worse due to poorer HIV control. To address this question, we looked at baseline CD4 count in these three participants. Although the participant with the lowest Glutamate:GABA ratio and the poorest global T score (participant G-2) had the lowest CD4 count (50); the other two participants did not follow this correlation (M-13, CD4 count 468; M-16, CD4 count 373) (see figure 6.7). CD4 count at time of TBM diagnosis is a poor proxy marker for overall HIV control, therefore this explanation may still be plausible. The usefulness of further analysis into only three participants to find explanations for this trend is questionable; however, the result highlights the complexity of interpretation of MRS results in this context given the broad spectrum of variables which may influence metabolite concentration besides TBM itself; namely HIV co-infection, previous traumatic brain injury, aging and co-morbidities such as other forms of

dementia. Further studies considering the utility of MRS must consider these variables, and control for them within the populations assessed.

Within this small study, our aim was to measure glutamate/glutamine and GABA concentrations within our participants. However, data on other metabolites measured within the spectrum may be interesting within TBM. In traumatic brain injury glutamate/choline ratios as well as glutamate levels have been used as a predictive marker of neurocognitive outcome (Shutter et al., 2004). Studies in HIV have shown creatine, choline, myo-inositol and N-acetyl-aspartate have roles in measurement of brain injury in HIV (reviewed in (Chaganti and Brew, 2021)). In multiple sclerosis, N-acetyl-aspartate is a marker of neuronal loss whilst elevated choline peaks represent increased cell membrane turnover and therefore interpreted as a marker of demyelination, inflammation and gliosis; in fact the complete spectrum of metabolite concentrations can be interpreted to establish prognosis, following disease evolution, understand pathogenesis and as a marker of therapeutic efficacy in MS (reviewed in (Narayana, 2005)). Further development of this tool within TBM, learning from other neuroinflammatory and neurodegenerative diseases, in settings where MRS is available, would allow us to harness the full capability of this tool to improve outcomes in TBM.

There are several limitations to this study. Firstly, only a small number of participants were included. This is in part due to the complexity of transporting participants from referring sites across Cape Town to the imaging facility, but also due to COVID-19 related restrictions making the ongoing inclusion of TBM participants impossible after March 2020. Moreover, given the novel nature of our imaging protocol, a number of

imaging sequences could not be included e.g. due to lack of water reference data. A larger study, following further refinement of the protocol to ensure all data collected is interpretable, is required to validate these findings. Secondly, the majority of participants enrolled had mild disease. All participants enrolled to LASER-TBM were eligible for inclusion; however, i) in general most LASER participants had mild or moderate disease and ii) those who were more unwell were in many cases not stable enough to be transported and/or imaged. It is plausible that in more severe disease, our findings may be more striking? However, future studies including patients with severe disease are required to test this hypothesis and to ensure generalisability of results. Thirdly, our healthy controls were younger than our TBM participants, and did not have a diagnosis of HIV. One study has demonstrated that glutamate concentrations within a similar region imaged within our study (the striatum) reduces with age(Zahr et al., 2008), therefore it is possible that differences in glutamate concentrations between our groups are unlikely to have been due to age and may well have been more marked in a better matched group. Other studies however have demonstrated that aging can be associated with different patterns of glutamate changes depending on whether white or grey matter is captured within the single voxel(Kaiser et al., 2005). Along these lines, our study results reflect changes within a single location within the brain where we know microvascular pathology is likely to occur. In TBM however, neurological sequelae can occur in many regions of the brain. Our results in chapter 4 suggest that cognitive impairment is diffuse, affecting multiple domains corresponding to multiple anatomical areas. Therefore, placement of the voxel within other areas, including cortex and subcortical white matter tracts, as well as within lesions such as infarcts or tuberculomas would provide a greater breadth of understanding of metabolic changes



occurring in TBM. Finally, it is unclear whether the changes seen are specific to TBM; comparison to a control group of other forms of meningitis would allow us to differentiate changes specific to TBM.

In conclusion, our study provides *in vivo* correlation of findings within the literature, which suggest that glutamate-driven excitotoxicity may have a role in the pathogenesis of TBM. A recent international survey found that although 76% of TBM settings surveyed did have access to MRI, this wasn't routinely used, likely due to higher burden on available resources (Tucker et al., 2020). This suggests that the framework for developing MRS exists however that in the short term at least its potential may be limited to understanding pathogenic mechanisms via studies undertaken in well-resourced research settings; this study proves that MRS can be developed as a useful tool in this context. Larger studies are now required to validate these findings, measure changes at longer term timepoints and correlate findings with neurological outcomes including cognitive impairment to improve our understanding of pathogenic mechanisms. In time, with greater evidence for its utility and greater resource and expertise in TBM endemic areas, MRS may have a wider role including as a predictive measure and monitoring tool in order to improve outcomes for patients with TBM.

Figure 6.1 Magnetic resonance spectrum of normal brain

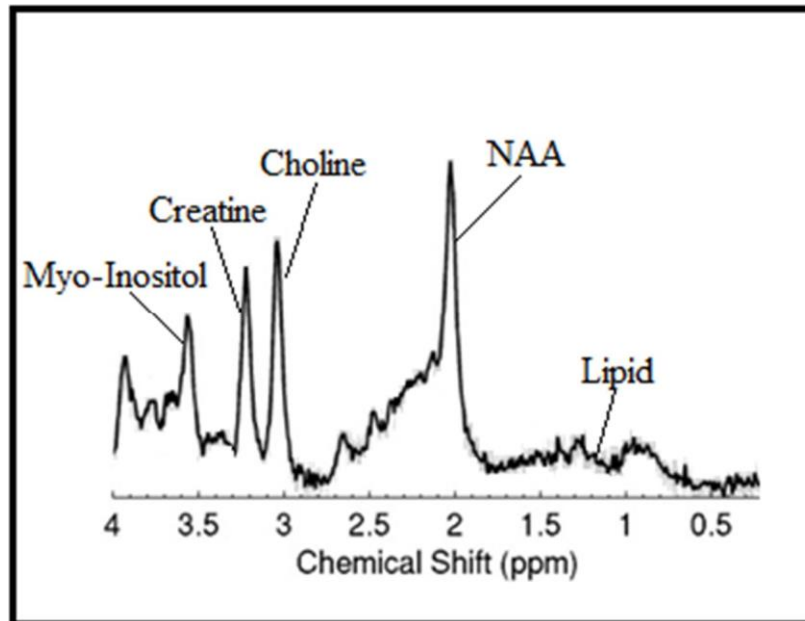
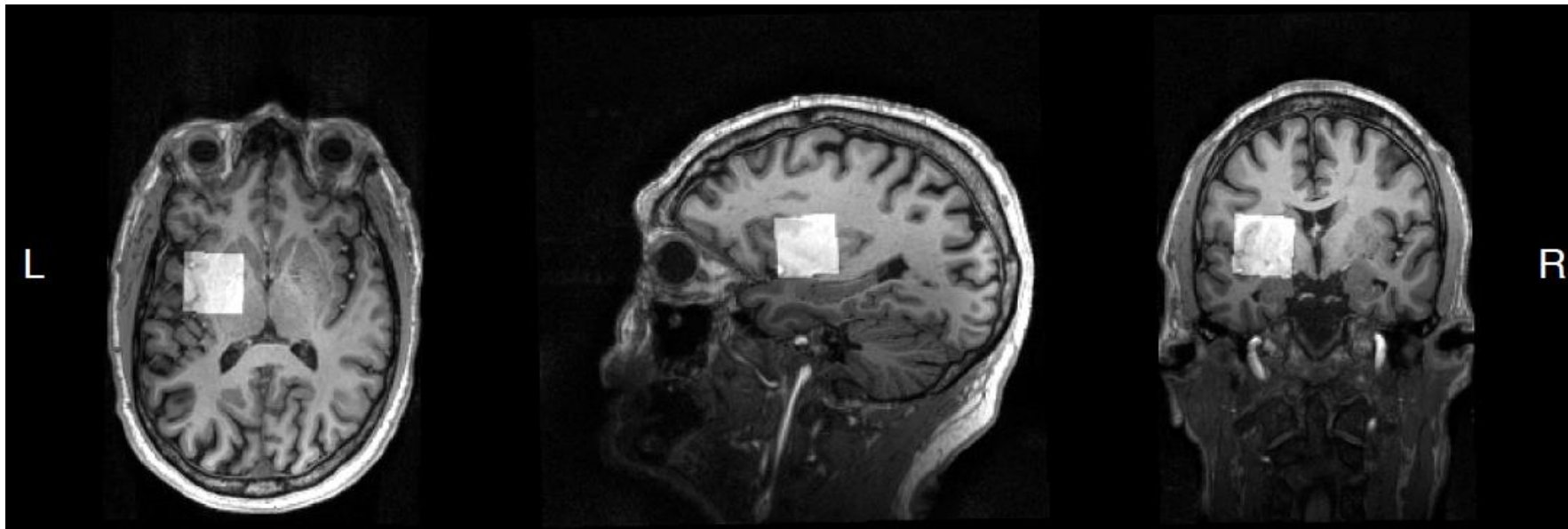


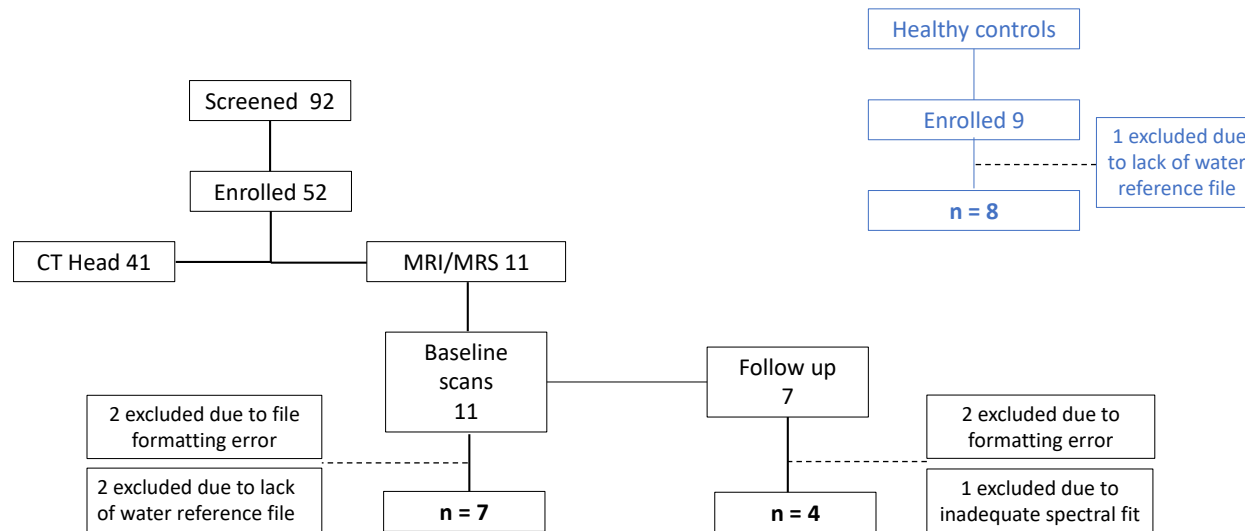
Figure taken from (Manias and Peet, 2018) showing normal MRS spectra. NAA, N-acetyl-aspartate.

Figure 6.2 Voxel placement



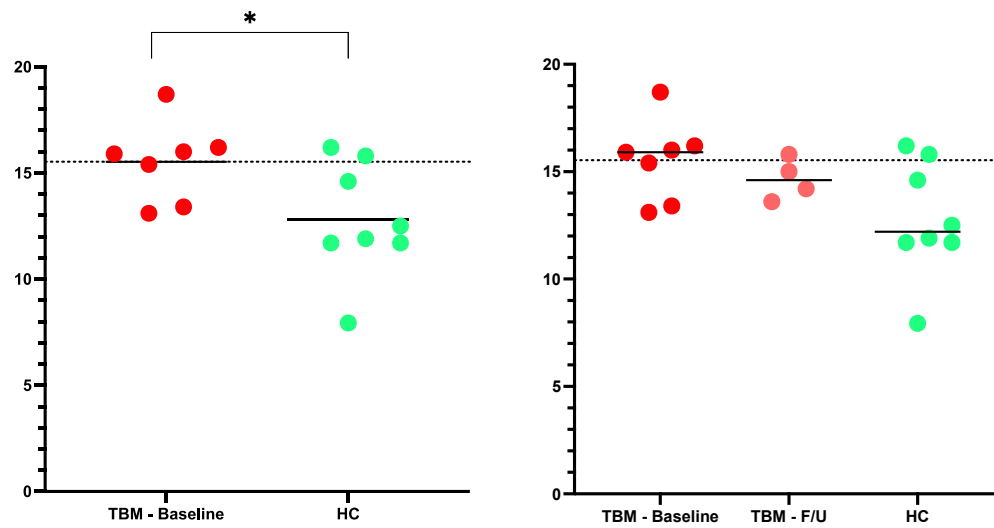
From left to right, axial, sagittal and coronal images demonstrating placement of the single voxel within the 'tubercular zone'

**Figure 6.3 Study CONSORT for magnetic resonance spectroscopy study**



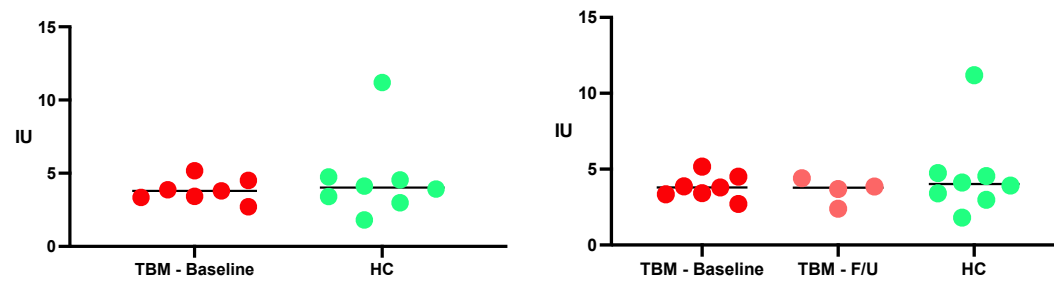
CONSORT diagram demonstrating i) imaging modalities used in LASER-TBM and inclusion of participants for magnetic resonance spectroscopy imaging and ii) recruitment of healthy controls for comparison

**Figure 6.4 Glutamate/glutamine (Glx) concentrations in participants with TBM versus healthy controls**



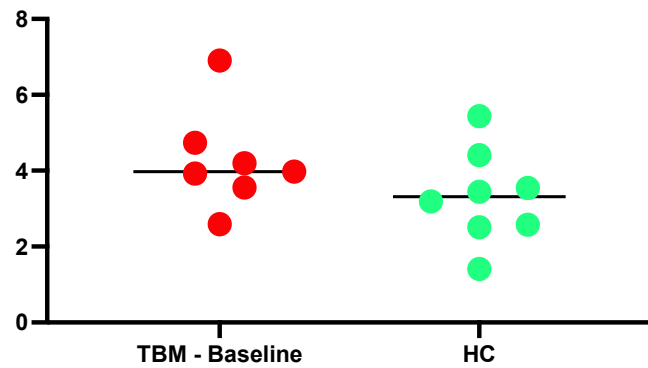
The graph on the left demonstrates that there is significant reduction of Glx concentrations at baseline in TBM participants compared to healthy controls ( $p=0.0428$ ). The graph on the right demonstrates that these concentrations reduce within the first 2 months of treatment but not to the level of healthy controls. X axis units IU.

**Figure 6.5 GABA concentrations in participants with TBM versus healthy controls**



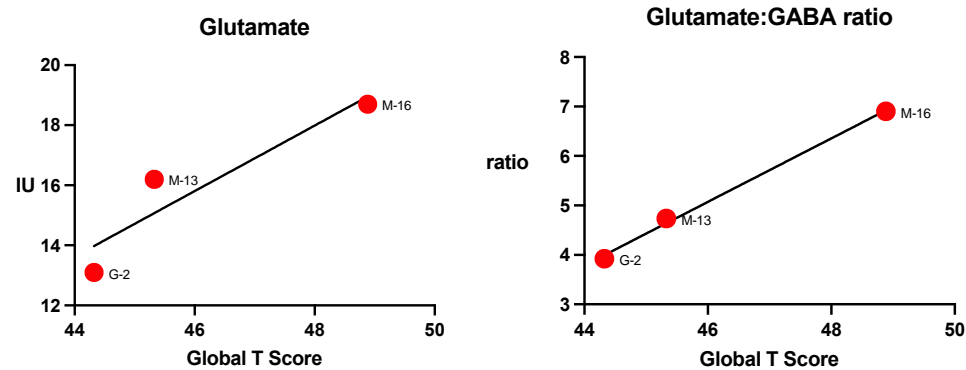
The graph on the left demonstrates that there is no difference in GABA concentrations at baseline in TBM participants compared to healthy controls. The graph on the right demonstrates that these concentrations do not change within the first 2 months of treatment.

Figure 6.6 Glutamate:GABA ratios in participants with TBM versus healthy controls



There is a non-significantly raised Glx:GABA ratio in TBM participants compared to healthy controls ( $p=0.173$ )

**Figure 6.7 Baseline glutamate concentrations and glutamate:GABA concentrations in relation to cognitive outcomes**



The graph on the left plots glutamate concentration at baseline scan to global T score on cognitive test performance at 6 months and shows a non-significant correlation by simple linear regression analysis ( $p=0.2382$ ). The graph on the right plots Glutamate:GABA ratio at baseline scan to global T score on cognitive test performance at 6 months and shows a significant correlation by simple linear regression analysis ( $p=0.0362$ ).



**Table 6.1 Baseline demographics and clinical characteristics of participants included with the study of magnetic resonance spectroscopy in TBM**

	TBM	Healthy controls	<i>p</i> value
<i>n</i>	7	8	
Age (median, IQR)	38 (34-43)	23 (22-28)	<i>p</i> =0.006
Gender			
Male	2	4	
Female	4	4	
Median days from enrolment to baseline scan	5		
MRC Grade			
Grade 1	7		
Grade 2	0		
Grade 3	0		
TBM case definition criteria			
Definite	1		
Probable	5		
Possible	1		

## Chapter 7. Discussion

My PhD set the ambitious goal of improving understanding of pathogenesis in TBM. To achieve this goal, under the supervision of my primary supervisor, I set out to design and complete a clinical trial, within which nested sub-studies would enable distinct yet interlinked studies of disease complications and pathogenic mechanisms in TBM. Within the chapters of this thesis, I have:

1. Given a comprehensive summary of current treatment strategies, potential novel drug targets and current understanding of pathogenic mechanisms in TBM
2. Described the design and results of a phase 2A randomised controlled trial evaluating a novel drug regimen in HIV-associated TB
3. Described cognitive and functional impairment in HIV associated TBM
4. Described immune profiles in HIV-associated TBM via ex vivo analysis of blood and CSF using Luminex platform technology
5. Developed a novel protocol to measure GABA and glutamate levels in the brain and compared levels to healthy controls via a pilot study using Magnetic Resonance Spectroscopy in TBM

The COVID-19 pandemic occurred during the course of this PhD, and as such a hiatus to this work enabled me to design a study of neurological manifestations of SARS-CoV2 infection within a wider platform of studies, namely: 'Health facility-based observational studies to investigate interaction and overlap between SARS- CoV2, HIV-1 and *M. tuberculosis* infections', the 'HIATUS' study. As well as describing neurological presentations to a tertiary COVID-19 referral centre during the first wave of

the pandemic in South Africa, the *ex vivo* analysis provided insights on neurotropism of SARS-CoV2. The cohort also provided a comparison group for the *ex vivo* analysis of TBM samples.

In each of the results chapters (chapters 3-6) I have discussed the main findings and limitations of each of these studies in turn. Within this chapter, I aim to summarise these and comment upon the implications of these results within the context of advancing our understanding of pathogenesis in TBM.

In chapter 3, we demonstrated that linezolid and high dose rifampicin can be safely combined in the treatment of HIV-associated TBM. We also showed that high-dose aspirin was safe when combined with intensified antibiotics. As well as providing timely safety data to support further evaluation of these important drugs in a phase 3 setting, the trial is the first to evaluate a treatment approach which simultaneously aims to ensure i) effective killing of Mycobacteria within the CNS and ii) control of the dysregulated immune response in TBM. Given the poor treatment outcomes in TBM and the lack of any proven effective host directed therapy in HIV co-infected patients, we believe that this approach is rational to ensure that drug development in this field moves forward at a pace which is timely and appropriate. The nested sub-studies within LASER-TBM have, in the context of my PhD, advanced our understanding of pathogenesis in TBM, which I expand upon in summaries below. However, beyond the scope of this PhD, samples arising from this trial will contribute to our understanding of pathogenic processes via ongoing and planned integrated multi-omic analysis of blood and CSF in patients with HIV-associated TBM.

In chapter 4, we demonstrated that cognitive impairment occurs in 50% of people with HIV-associated TBM at six months following diagnosis. Inclusion of comparator groups of people living with HIV and patients with TB occurring outside of the CNS allowed us to conclude that impairment was above that attributable to HIV, or systemic inflammation and polypharmacy in TB alone. This was, by far, the largest study to date within this field and describes for the first time, using validated and comprehensive neuropsychological assessments, the frequency and nature of this disabling complication in TBM. It also however highlights the value of using objective and quantitative measures of neurological outcome. As we endeavour to uncover drugs which improve survival in TBM, we must also think hard about better ways to evaluate morbidity in this disease. The Modified Rankin Scale has been adopted by the TBM community however is a crude marker of outcome in a condition where neurological complications are diverse and complex. Identifying outcome measures which have the potential to be adapted for both well and poorly resourced settings will not only provide more granular information on the clinical phenotype for pathogenesis studies of TBM, but also allow us to better understand the true impact of this disease for our patients with TBM.

In chapter 5, via a comprehensive immunoassay analysis of CSF and blood, we show that immune responses in TBM are highly compartmentalised, remain persistently raised at four weeks following diagnosis, despite treatment, with highest concentrations evident prior to death. In keeping with the literature, we highlight the role of the innate immune response, but also identify mediators known to be involved in adaptive immune responses in those with poor outcome. We identify IL-1 $\beta$  as a potential blood-based biomarker of poor outcome. These results

require validation in a larger cohort, however, they do contribute insight to pathogenic mechanisms and biomarkers of poor outcome in TBM.

In chapter 6, we demonstrate that MRS is a feasible mechanism to measure concentrations of glutamate and GABA in the brains of patients with TBM. Within this small pilot study, we show that concentrations of glutamate are significantly raised in the acute phase of TBM compared to healthy controls, and there is a trend towards persistently raised levels at 2 months despite treatment. In light of recent publications, which describe the potential contribution of glutamate driven excitotoxicity following TBM, this novel mechanism for validating these findings *in vivo* is exciting. Further work following this PhD now plans to use multi-omic analysis to understand whether pathways associated with glutamate and GABA are associated with poor outcome within the same cohort. Beyond this, MRS may have a broader role in understanding pathogenic mechanisms in TBM, particularly in relation to longer term sequelae such as cognitive impairment as it has done in other neurological diseases.

Within the study included within the appendix of this thesis, during the first wave of COVID-19 at a time when SARS-CoV2 was a new and poorly understood virus, we set out to evaluate the spectrum of neurological manifestations and the potential for this virus to penetrate the CNS. In this study, we described neurological presentations in a setting in where COVID-19 infections were high and, through systematic evaluation of the CSF, demonstrate that penetration of SARS-CoV2 to the CNS is uncommon, even in suspected cases of meningoencephalitis.

## Conclusions

Whilst I recognise limitations in my work, I believe that the research contained within this thesis contributes important information for the development of new and effective treatment regimens, the knowledge of neurological sequelae and understanding of pathogenic mechanisms in TBM. TBM is a disease which affects some of the world's most vulnerable people, is a disease of poverty and one with devastating outcomes.

Fortunately, senior academics within the field are collectively driving forward a diverse and energetic programme of research which address many of the unanswered questions within this field. The work undertaken within my PhD has provided me the opportunity to work with, learn from and be inspired by some of the most effective researchers across the world who share with me an ambition to improve outcomes for those with TBM. In particular, the work has presented to me many new questions around longer term neurological complications in TBM and the pathogenic mechanisms underlying these. I intend to remain active in the TBM field, building upon the skills I have acquired within this PhD to address these questions and others (whilst asking many more) throughout my research career.

# **Appendix: Spectrum of neurological manifestations and systematic evaluation of cerebrospinal fluid for SARS-CoV2 in patients admitted to hospital during the COVID-19 epidemic in South Africa (The HIATUS-3 Study)**

## **Introduction**

Although COVID-19, the disease caused by SARS-CoV2, is primarily a disease of the respiratory tract, coronaviruses (CoV) are known to be neurotropic with some strains leading to meningitis, encephalitis and cerebral vasculitis (Puelles et al., 2020). In the first case series of SARS-CoV2 conducted in Wuhan, China, it was found that in 62 patients, 21 (34%) reported headache (Xu et al., 2020). In a subsequent retrospective case series to investigate neurological presentations in 214 patients with COVID-19, 36% of patients were found to have neurological signs and symptoms, which were more frequent in severe compared to non-severe COVID-19 (45% vs 30.2%). Since then, a wealth of literature suggests that SARS-CoV2 has the potential to present with neurological manifestations, with or without pulmonary involvement. Reports include cases of meningitis/encephalitis, with (Moriguchi et al., 2020, Khodamoradi et al., 2020b) and without (Espinosa et al., 2020) detection of SARS-CoV2 within the cerebrospinal fluid (CSF), stroke (Beyrouiti et al., 2020), transverse myelitis (Kang Zhao, Sarma and Bilello, 2020, Valiuddin et al., 2020), acute disseminated encephalomyelitis (de Miranda Henriques-Souza et al., 2021) and inflammatory polyradiculopathies such as Guillain-Barre Syndrome

(Sedaghat and Karimi, 2020, El Otmani et al., 2020, Alberti et al., 2020, Toscano et al., 2020a). Whether the mechanism by which these phenomena occur are due to, either individually or in combination, direct neurotropic invasion, para- or post-infectious inflammation, a systemic coagulopathic processes or indeed due to separate and co-incidental pathogenic processes is not well understood.

The distinction between direct CNS invasion and other potential mechanisms is however important to make. Clinicians who assess patients presenting with neurological symptoms in the context of a COVID-19 epidemic must know when and how to investigate for neurological complications of SARS-CoV2 infection. Moreover, should SARS-CoV2 commonly lead to neurological symptoms via direct CNS invasion, particularly in the absence of pulmonary symptoms, then patients should be screened routinely for neurological involvement, and physicians must consider this unique potential of the virus. If direct SARS-CoV2 invasion to the CNS does not explain neurological presentations in this context, then greater focus must be towards understanding the nature and contribution of para- and post-infectious inflammatory phenomena and coagulopathy in order to develop effective therapeutics to treat these often severe complications of disease. This is particularly important in resource limited settings where testing for SARS-CoV2 is not always available and as yet, there are no bespoke methods to test for SARS-CoV2 in cerebrospinal fluid (CSF). In these settings, the precise incidence of SARS-CoV2 infection may not be known or be underestimated: in the Eastern Cape of South Africa for instance where this study took place, seroprevalence following the first wave of the epidemic has been found to be as high as 62.5% in adults under the age of 65 years which is 8-fold higher than the official case



count (Sykes et al., 2021), and has led to death in an estimated 1 in 300 people in the region (SAMRC data) (<https://www.samrc.ac.za/reports/report-weekly-deaths-south-africa?bc=254>). Such high rates of infection and subsequent mortality not only calls for better resource to manage the disease in these settings, but should also alert the clinician to the increased possibility of encountering less typical presentations of SARS-CoV2 infection.

To date no published studies have systematically investigated the presence of SARS-CoV2 within the CNS in patients presenting with neurological symptoms with and without pulmonary manifestations of COVID-19 in a context where COVID-19 infection is the most frequent reason for hospital admission. In a prospective cohort study in the Eastern Cape of South Africa, we described clinical and radiological features, and assessed for the presence of SARS-CoV2 within the CNS in those presenting to hospital with neurological symptoms during the first peak of the COVID-19 epidemic.

## **Methods**

### *Patient recruitment*

We undertook a prospective cohort study at Livingstone Hospital, Eastern Cape. The study was approved by the Faculty of Health Sciences Human Research Ethical Committee of the University of Cape Town (HREC 207/2020) and by the ethical review board at Livingstone Hospital. We sequentially enrolled adults (>18 years) presenting with neurological symptoms who at the discretion of the treating physician required inpatient investigation by lumbar puncture and cerebrospinal fluid analysis between

12<sup>th</sup> July and 20<sup>th</sup> October 2020. During this time Livingstone Hospital served as a COVID-19 referral centre in the Eastern Cape with an average admission rate of 40 confirmed COVID-19 cases per day. Written informed consent was taken from the patients where possible in those with capacity to consent. In those with decreased consciousness, patient relatives were approached for proxy consent. In those where no relative was contactable, permission was sought on an individual case basis by the Faculty of Health Sciences Human Research Ethical Committee of the University of Cape Town.

#### *Clinical and radiological data collection*

Clinical data was collected on symptoms and signs at presentation as well as relevant past medical history at two timepoints (baseline, and again between 3 and 7 days of enrolment). Computerised Tomography (CT) head images performed as part of standard of care included as routine sagittal and axial views. Two dimensional images were retrieved and independently viewed using a picture archive and communication system (PACS) by a blinded neuroradiologist using a standardised case report form. No specific study procedures took place, however at the time of diagnostic lumbar puncture a total of 6 ml of additional CSF was retrieved for study-specific analysis and biobanking. Similarly, venipuncture was not performed as a study procedure, however 24 ml of additional whole blood samples were collected for study specific blood work up and biobanking. Data on routine blood and CSF investigations performed as part of the diagnostic work up were collected from the National Health Laboratory Database and patient medical record. The outcome of admission (including death and final diagnosis) were recorded retrospectively.

### *Laboratory methods to detect SARS-CoV2 in CSF*

An in-house RT-PCR that compared well with the routine nationally employed test, the Multiplex TaqMan™2019-nCoV kit (Applied Biosystems, Waltham, Massachusetts, USA) was established and used to detect SARS-CoV2 in CSF targeting the E gene and subgenomic RNA (sgRNA). Viral subgenomic mRNA is transcribed only in infected cells and not packaged into virions and therefore a positive sample may indicate evidence of actively infected cells within the CSF (Wolfel et al., 2020). RNA was extracted from patient samples using the E.Z.N.A. Viral RNA kit (Omega Bio-tek), followed by reverse transcription and PCR-amplification of the SARS-CoV2-specific targets E and sgRNA (as well as a human RNA control, RNaseP (RP)) using the TaqPath™ 1-Step Master Mix kit (Thermo Fisher) on a QuantStudio 7 Real-Time PCR machine (Thermo Fisher). Primers and probes for SARS-CoV2 E gene and sgRNA readouts have been published elsewhere (Corman et al., 2020) and were synthesized by Inqaba Biotec (South Africa), while primers and probes for the RP control target were provided by the CDC 2019-Novel Coronavirus (2019-nCoV) Real-Time RT-PCR Diagnostic Panel. Each run included a positive control (PC), which for sgRNA runs included a sample from a previously positive patient, a human specimen extraction control (HSC; e.g. HeLa cell RNA) as well as a 'no template' control (NTC, i.e. water). All samples were amplified under the same conditions using 400nM concentrations of each of the primers, as well as 200nM of probe. Thermal cycling involved 10min at 53°C for reverse transcription, followed by 3min at 95°C to deactivate Reverse Transcriptase and Taq activation, and 45 cycles of 3s at 95°C and 30s at 57°C. A run was considered valid if the control samples yielded the following results with a Ct value < 40 being considered a positive signal: NTC negative for E, sgRNA and RP; HSC negative for E and sgRNA,

positive for RP; PC: positive for E, sgRNA and RP. When all controls exhibited the expected performance, an unknown patient sample was considered negative if the Ct values for E and sgRNA were  $> 37$  AND the Ct value for RP  $< 37$ . A specimen was considered positive if the Ct values for E and sgRNA were  $< 37$ . Runs with a CT between 37 and 40 were repeated.

### *Statistical analysis*

Data was analysed as an entire cohort with continuous characteristics described in terms of median values and interquartile ranges, and dichotomous variables as counts and percentages. Comparison between patients who tested positive for SARS-CoV2 on nasopharyngeal (NP) swab PCR, and those who did not was assessed using Wilcoxon rank-sum tests to compare continuous variables, and chi squared test for dichotomous variables. All analysis was performed within GraphPad Prism (version 9, Prism for MacOS) software.

## **Results**

A total of 40 participants (24 female, 16 male) were screened for inclusion in the study. One participant was not enrolled as neither deferred and proxy consent were available. Therefore, 39 participants were included within the analysis. The median age at enrollment was 44 years. Baseline characteristics are outlined in table A.1. At the time, routine testing for SARS-CoV2 by NP swab was not available for all inpatients due to limited resource; however, 31/39 participants included within this study underwent testing due to symptomatic presentation, or recent COVID-19 contact. 7/31 participants tested positive for SARS-CoV2 via NP swab. In tables A.1 and 5.2, demographics and clinical characteristics of the cohort are described,

as well as for those who tested positive and negative for SARS-CoV2 via NP swab.

Neurological complaints at baseline are described in table A.1. The most prevalent complaint was confusion (27/39). Less common complaints, occurring with or without confusion included: neck stiffness (14/39), headache (12/39), new onset or increasing frequency of seizures (7/39), acute psychotic symptoms (10/39). On neurological assessment, 5/39 had new onset hemiplegia and 1/39 had new onset bilateral lower limb weakness. In the absence of other motor or sensory disturbance, 1/39 had new onset lower motor neuron facial nerve (VII cranial nerve) weakness, and 1/39 had abducens (VI cranial nerve) weakness. Neurological complaints at presentation in those testing positive for SARS-CoV2 on NP swab included: confusion (4/7) of which 1/7 demonstrated acute psychotic symptoms, new onset seizures (1/7), headache and neck stiffness (1/7), acute onset right sided weakness (1/7) and bilateral lower limb weakness (1/7). Of these 7 participants, 3 had no classical symptoms of COVID-19 (cough, shortness of breath, fever, anosmia or dysgeusia). Between those with and without a positive NP swab for SARS-CoV2, there was no significant difference in the neurological complaints at baseline. In 7 patients with a confirmed diagnosis of COVID-19 (SARS-CoV2 positive PCR on NP swab); 2/7 were thought to have COVID-19 pneumonia with stroke (see table A.4), 1/7 presented with a clinical diagnosis of myelitis treated as possible viral or *M. tb* in aetiology, with the remaining 4/7 patients, all of which presented with confusion, thought to be due to delirium secondary to COVID-19 pneumonia. No further investigation was performed to formally assess for encephalopathy.

Baseline blood and CSF analysis are described in table A.2. This analysis was performed as part of routine care, where there was a clinical indication and therefore not all tests were performed on every participant. There was no difference in CSF markers to suggest an acute infective or inflammatory process in those who tested positive for SARS-CoV2 via NP swab, versus those who did not, this includes: lymphocyte count ( $p=0.62$ ), polymorphonuclear cells ( $p=0.39$ ) and protein ( $p=0.59$ ). CSF glucose was significantly higher in those with a diagnosis of COVID-19 (5.0 vs 3.0,  $p=0.01$ ); however, of note HbA1C (%) was significantly higher in patients with COVID-19 than in those without (13.0 vs 7.25,  $p=0.02$ ), reflecting the non-significant higher proportion of patients with pre-existing diabetes mellitus in those with a diagnosis of COVID-19 compared to those without (2/7 vs 4/32,  $p=0.29$ ).

Computerised Tomography (CT) scans of the brain were performed at baseline in 26/39 participants, of which 2 were performed in patients with a confirmed diagnosis of COVID-19. 3/26 scans were performed with contrast enhancement. Radiological findings at baseline are summarized in table A.3. In the two scans performed in patients with COVID-19, one demonstrated multi-focal subacute infarcts within the left middle cerebral artery (MCA) and anterior deep borderzone territories. In the second, imaging demonstrated multi-territory mature infarcts in in both cerebral hemispheres and in the cerebellum (figure A.1). In both patients, CSF findings were unremarkable (see table A.4), and a diagnosis of COVID-19 pneumonia with presumed diagnosis of stroke (clinical in the former, radiological in the latter) was made.

Multiplex PCR (targeting nucleocapsid (N), spike protein (S) and Orf1ab) runs revealed no evidence of SARS-CoV2 in any of the 39 samples. Using

the primer combinations for E gene readout and subgenomic readout repeatedly gave no or weak ( $Ct > 37$ ) signals that were interpreted as negative. Therefore, none of the 39 samples demonstrated SARS-CoV2 via any of the PCR primer combinations applied. Raw data is presented in table A.5.

## **Discussion**

We describe a cohort of patients presenting with clinical symptoms suggestive of possible neuroinfective or neuroinflammatory aetiology, with and without symptoms and a confirmatory diagnosis of COVID-19 during the first peak of the pandemic in a resource poor setting in South Africa. Examination of the CSF using PCR for multiple targets was negative in all cases therefore suggesting little evidence of direct neurotropic invasion of SARS-CoV2 in CNS.

Published reports provide rationale for direct neurotropic invasion of SARS-CoV2 including cases of meningitis and encephalitis where PCR for SARS-CoV2 was positive in CSF with and without classical symptoms of COVID-19 (Moriguchi et al., 2020) [(Khodamoradi et al., 2020a). At autopsy, SARS-CoV2 RNA has been detected in the brains of patients who have died due to COVID-19 albeit at titers lower than in other affected organs (Wichmann et al., 2020, Puelles et al., 2020). Evidence for direct neurotropic invasion is supported by findings from the 2002 SARS-CoV outbreaks where studies demonstrated the presence of coronavirus particles in the brain (Ding et al., 2004, Gu et al., 2005, Xu et al., 2005), with subsequent studies describing penetration of the CNS via the olfactory nerve (Netland et al., 2008). In SARS-CoV2, a case of olfactory gyrus intracerebral hemorrhage, an

uncommon location for spontaneous hemorrhage, as well as the high rates of anosmia, has highlighted whether SARS-CoV2 can invade neurological structures such as the olfactory bulb via nasal mucosa (Thu et al., 2020). Entry into the CNS via synaptic connections may also provide rationale to consider a centrally-driven contribution to cardiorespiratory dysfunction in coronaviruses, where acute onset respiratory failure leads to significant morbidity and mortality [36]. These observations have, during the course of the COVID-19 pandemic, raised questions as to whether SARS-CoV2 should be investigated as a causative organism, particularly in patients presenting with a meningitis or encephalitis in a setting where SARS-CoV2 infection rates are high, both in those with and without a confirmed diagnosis of COVID-19. In our cohort who were enrolled sequentially in a tertiary setting for COVID-19 care, despite detailed examination of the CSF for presence of SARS-CoV2 using PCR primers for genomic and subgenomic RNA, no patients were found to have evidence of direct neurotropic invasion to the CNS. This finding is important in shaping the direction of clinical care in the management of patients presenting with neurological symptoms in the COVID-19 era, particularly in resource limited settings where the prioritisation of investigations is an important consideration in clinical care.

In our study, one patient had a clinical presentation consistent with myelitis alongside a diagnosis of COVID-19, however in this case an explanation other than SARS-CoV2 infection was thought more likely to account for their clinical presentation and no evidence of direct neurotropic invasion of SARS-CoV2 was found. Acute myelitis (Kang Zhao) is one of many cases reported in the literature where the mechanism was thought due to a post- or para-infectious inflammatory response to SARS-CoV2. Other cases



which may suggest inflammatory sequelae occurring during or following SARS-CoV2 infection include: acute necrotising hemorrhagic encephalopathy (Rossi, 2008, Poyiadji et al., 2020), Guillain Barre Syndrome (Sedaghat and Karimi, 2020, El Otmani et al., 2020, Alberti et al., 2020, Toscano et al., 2020b) and Miller Fisher Syndrome (Gutierrez-Ortiz et al., 2020), and acute disseminated encephalomyelitis (Parsons et al., 2020, Novi et al., 2020), all of which occurred without evidence of SARS-CoV2 in the CSF. Given that within our cohort direct neurotropic invasion of the CNS was not found, these results might suggest that greater emphasis should now turn towards understanding the role of inflammation at the time of, or following, SARS-CoV2 infection in a subset of patients.

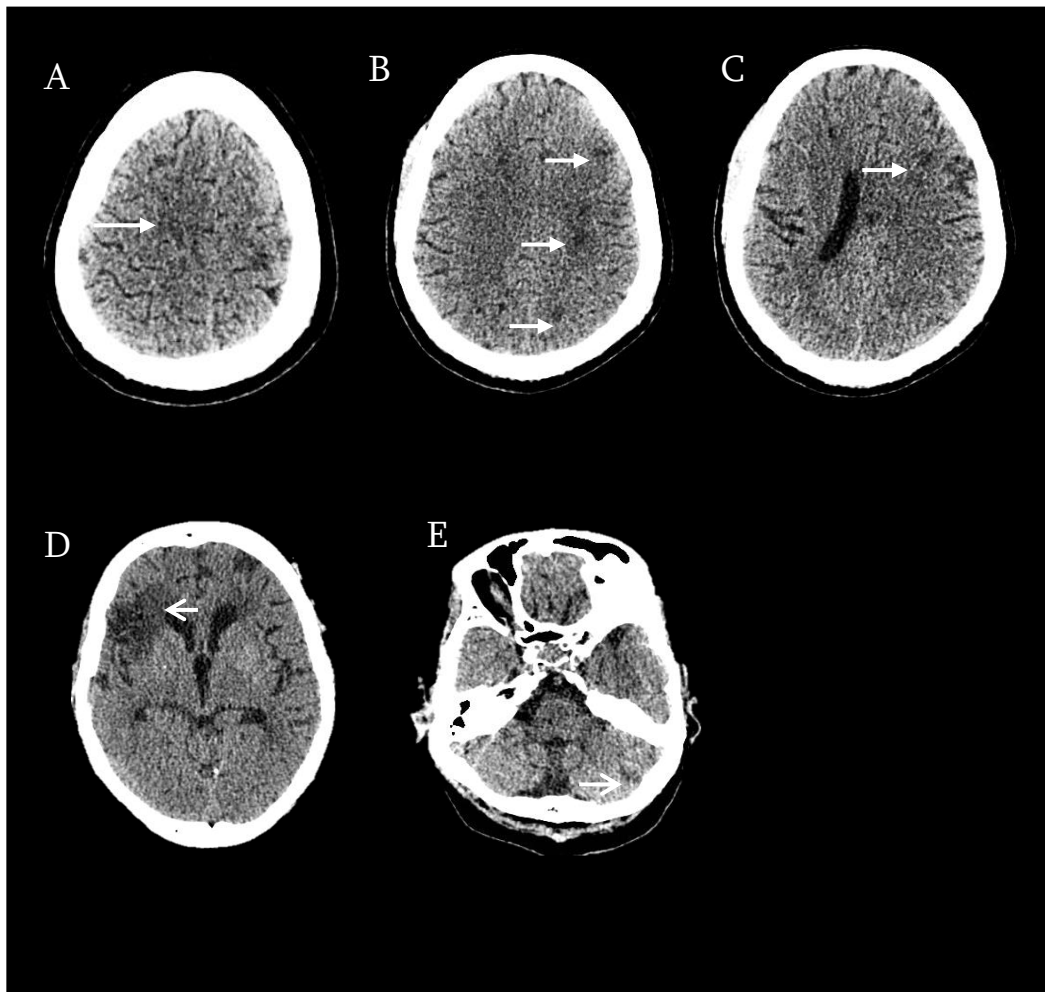
Moreover, the proportional contribution of the now well-described coagulopathy leading to endothelial dysfunction and eventual end organ damage is unknown (Hess et al., 2020). This is particularly important to understand in the context of stroke in patients with COVID-19; now frequently reported to occur where otherwise no clear vascular risk factors exist (Goldberg et al., 2020, Avula et al., 2020). In our cohort, two patients presented with presumed stroke alongside COVID-19 pneumonia. In both cases, vascular risk factors co-existed and may in part or completely explain the vascular complications. Further research is required to understand the interplay of the presumed coagulopathy both on pre-existing vascular risk factors such as hypertension and diabetes, and other stroke risk factors such as HIV, particularly within the South African context.

There were several limitations to this study. Given the pragmatic nature of its design, only data on investigations indicated as part of routine clinical care were available for analysis, resulting in an incomplete laboratory and

radiological data set. This includes cerebral imaging, which was not performed in all participants, and in instances where it was, contrast was not given in the majority (23/26) of cases. Factors related to coagulation, such as D-Dimer levels, would have provided interesting comparison of patients presenting with complications such as stroke with and without confirmed COVID-19. In contrast, laboratory procedures related to the discovery of SARS-CoV2 in CSF were thorough and robust, which is reassuring that despite multiple runs, the negative findings are reliable.

Although small, this pragmatic observational cohort study contributes knowledge to our increasing understanding of COVID-19 management. Through systematic analysis of CSF in patients presenting with neurological symptoms in a context where incidence of SARS-CoV2 infection is high, we have demonstrated that although cases within the literature exist, direct neurotropic invasion of the CNS is uncommon. This includes suspected cases of meningitis and encephalitis, syndromes most aligned to direct neurotropic mechanism. This considered, neurological presentations in cases of COVID-19 continue to be reported, and lead to morbidity and mortality in patients affected. The results from our study suggest that further emphasis must now turn towards understanding the role of inflammation and coagulopathy in the development of neurological syndromes. This includes studies to assess the efficacy of proven anti-inflammatory drugs such as corticosteroids and tocilizumab, and therapeutics to manage acute stroke in the treatment of patients who develop neurological symptoms due to SARS-CoV2 infection.

**Figure A.1: Neuroimaging in patients with confirmed diagnosis of COVID-19**



Axial unenhanced CT head imaging in two patients where a diagnosis of COVID-19 was confirmed (PCR for SARS-CoV2 positive on NP swab). Top row demonstrates poorly-defined multifocal cortical and subcortical hypodensities in keeping with subacute left middle cerebral artery territory (A,C) and anterior deep borderzone territory (B) infarcts. Bottom row demonstrates multi-territory mature infarcts in the right middle cerebral artery territory (D) and left posterior inferior cerebellar artery territory (E).

**Table A.1 Baseline demographics and clinical characteristics in HIATUS-3**

	All	SARS-CoV2 -ve on NP swab or unknown SARS-CoV2 status	SARS-CoV2 +ve on NP swab
n	39	32	7
Gender			
Male	15	11	4
Female	24	21	3
Age (years)			
Median (range)	44 (27-84)	42 (27-79)	47 (42-84)
HIV seropositivity			
HIV co-infected	22	19	3
On ART	11	8	3
ART naïve/defaulted	11	11	0
HIV uninfected	13	11	2
Not known	4	2	2
HIV-1 VL (copies/ml) (n=13, median ; IQR)	75831; 28,0141	253,344; 258,249	205; 206
CD4 Cell count (cells/mm3) (nr 332-1642) (n=16, median; IQR)	35.5; 186	32; 99	260; 69
Comorbidities			
Cardiovascular Disease	2	2	0
Hypertension	13	9	4
Diabetes Mellitus	4	4	2
Obesity	15	11	4
Underlying Respiratory Disease	5	5	0
Chronic Kidney Disease	1	0	1
Baseline Symptoms			
Cough	17	12	5
Fever	7	3	4
Dyspnoea	11	7	4
Lethargy	10	9	1
Nausea	3	3	0
Diarrhoea	11	10	1
Headache	12	11	1
Neck Stiffness	14	13	1
Seizures	7	6	1
Agitation/psychosis	10	9	1

Confusion	27	23	4
Anosmia/Dysgeusia	1	0	1
Days symptomatic median (IQR)	7 (13)	8 (13)	7 (5)
Baseline Neurological Examination			
Hemiplegia	5	4	1
Paraplegia	1	0	1
Cerebellar ataxia	0	0	0
Tremor	0	0	0
Seizures	1	1	0
Cranial Nerve Abnormality	4	4	0
Glasgow Coma Score (Median)	14	14	14

**Table A.2 Blood and CSF analysis**

	Value expressed as	Performed in (n)	Normal range	All participants	SARS-CoV2 -ve on NP swab or unknown SARS-CoV2 status	SARS-CoV2 +ve on NP swab	p values
<b>Blood</b>							
Hemoglobin (g/dL)		39	12-15	<b>11.5</b> ; 4	<b>11.1</b> ; 2	<b>12.3</b> ; 2	
Platelets (cells x10 <sup>9</sup> /L)		37	186-454	<b>281</b> ; 140	<b>281</b> ; 116	<b>271</b> ; 168	
White cells (cells x10 <sup>9</sup> /L)		39	3.9 – 12.6	<b>8.4</b> ; 9	<b>8.3</b> ; 7	<b>8.4</b> ; 9	
Neutrophils (cells x10 <sup>9</sup> /L)		15	1.6 – 8.3	<b>6.0</b> ; 0	<b>7.8</b> ; 7	<b>4.4</b> ; 1	
Lymphocytes (cells x10 <sup>9</sup> /L)		15	1.4 – 4.5	<b>1.12</b> ; 12	<b>(In 13/15), 1.12</b> ; 1	<b>(in 2/15), 8.56</b> ; 8	
Eosinophils (cells x10 <sup>9</sup> /L)		15	0.0 – 0.4	<b>0.02</b> ; 0	<b>0.02</b> ; 0	<b>0.01</b> ; 0	
C-Reactive Protein (mg/L)			<10	<b>68</b> ; n/a	<b>47</b> ; n/a	<b>110</b> ; n/a	<i>p=0.36</i>
Erythrocyte Sedimentation Rate (mm/hr)		5	0-10	<b>96</b> ; n/a	<b>96</b> ; 67	<b>n/a</b> ; n/a	
Ferritin (µg/L)	<b>Median</b> ; IQR	3	11-307	<b>83</b> ; n/a	<b>83</b> ; 492	<b>n/a</b> ; n/a	
D Dimer (ng/mL)		14	0 – 0.25	<b>1.25</b> ; 2	<b>1.32</b> ; 1	<b>0.66</b> ; 2	
ALT (IU/l)		32	19-25	<b>29.5</b> ; 15	<b>32</b> ; 27	<b>19</b> ; 16	
Total Billi (µmol/L)		32	5-21	<b>12</b> ; 8	<b>11</b> ; 12	<b>15</b> ; 6	
Na+ (mmol/L)		39	136-145	<b>136</b> ; 9	<b>135</b> ; 10	<b>137</b> ; 6	
K+ (mmol/L)		39	3.5-5.1	<b>3.9</b> ; 0	<b>4.0</b> ; 1	<b>3.8</b> ; 0	
Creatinine (µmol/L)		39	49-90	<b>79.0</b> ; 24	<b>82.5</b> ; 64	<b>72.0</b> ; 18	
Total Protein (g/L)		24	6-78	<b>75.0</b> ; 14	<b>75.0</b> ; 13	<b>76.5</b> ; 10	
HbA1C (%)		7	4-5.6	<b>7.9</b> ; 6	<b>7.25</b> ; 2	<b>13.0</b> ; 0	<i>p=0.02</i>
<b>Cerebrospinal Fluid</b>							
Polymorphonuclear cells (cells/µL)		38	<3	<b>0</b> ; 6; 0-118	<b>0</b> ; 1; 0-118	<b>0</b> ; 0; 0-0	<i>p=0.39</i>
Lymphocytes (cells/µL)		38	<3	<b>0</b> ; 6; 0-2073	<b>0</b> ; 9; 0-2073	<b>2</b> ; 4; 0-12	<i>p=0.62</i>
Erythrocytes (cells/µL)	<b>Median</b> ; IQR; range	38	<3	<b>5</b> ; 48; 3-349	<b>3</b> ; 17; 0-349	<b>62</b> ; 148; 0-311	<i>P=0.28</i>
Protein (g/L)		39	0.15-0.45	<b>0.39</b> ; 1; 0.13-4.82	<b>0</b> ; 1; 0.13 – 4.82	<b>0</b> ; 0; 0.17 – 2.19	<i>p=0.56</i>
Glucose (mmol/L)		39		<b>3</b> ; 3; 0.1-10.2	<b>3</b> ; 2; 0.1-7.0	<b>5</b> ; 5; 2.8 – 10.2	<i>p=0.01</i>
Bacterial Culture	n, (% positive)	39		5/39 (12.8)	4/32 (12.5)	1/7 (14.2)	<i>p=0.90</i>
CLAT	n, (% positive)	32		5/32 (15.6)	5/29 (17.2)	0/3 (0)	
GXPU	n, (% positive)	22		1/22 (4.5)	1/18 (5.5)	0/4 (0)	
PCR for SARS-CoV2 in CSF							

Positive CT value for E		39		0/39	0/32	0/7	
Positive CT value for sgRNA		39		0/39	0/32	0/7	

Na+: sodium; K+: potassium; CLAT: cryptococcal latex agglutination titre; GXPU: GeneXpert Ultra; CT (cycle threshold); sgRNA: subgenomic RN.

**Table A.3 Radiological analysis**

Radiological Finding	All participants	SARS-CoV2 -ve on NP swab or unknown SARS-CoV2 status	SARS-CoV2 +ve on NP swab
Leptomeningeal Enhancement	1/26	1/24	0/2
Hydrocephalus	3/26	3/26	0/2
of which communicating	3/26	3/26	n/a
Radiological evidence of infarcts	14/26	13/26	1/2
of which new	3/14	3/13	0/1
single	10/14	9/13	1/1
multiple	4/14	4/13	0/1
Territory			
Middle cerebral artery territory	9/14	8/13	1/1
Anterior cerebral artery territory	0/14	0/13	0/1
Posterior cerebral artery territory	0/14	0/13	0/1
Ring enhancing lesions	0/26	0/24	0/2
Effacement	1/26	1/24	0/2
Of which local sulcal	1/26	1/24	0/2
Of which hemispheric sulcal	1/26	1/24	0/2
Of which global sulcal	0/26	0/24	0/2
Of which basal cistern	0/26	0/24	0/2



**Table A.4 Detailed clinical presentations, laboratory findings in patients presenting with COVID-19 and neurological symptoms**

Age range, Gender	HIV status, VL, CD4	Co-morbidities	Baseline symptoms and examination findings	Significant blood and CSF findings (Units as described in table 2)*	Radiological findings	Final diagnosis
50-50 M	Neg	Nil	2 day history of cough, fever, seizures (unclear if new onset) and confusion. GCS 14 on assessment	Blood: Hb 10.7, WCC 23 (eosinophils 291), CRP 110, Na 132. CSF: Lymphocytes 2, erythrocytes 13, protein 0.17, glucose 4.7, normal gram stain, negative bacterial culture	Not done	COVID pneumonia with confusion +/- new onset/worsening seizure frequency
40-50 F	UNK	Hypertension, DM, Obesity	2 day history of shortness of breath, anosmia, worsening confusion and agitation. GCS 7 on assessment with no focal neurological abnormalities	Blood: WCC 15, CRP 77, Na 147 CSF: Lymphocytes 1, Erythrocytes 62, protein 0.19, glucose 7.8, occasional lymphocytes on gram stain, negative bacterial culture	Not done	COVID pneumonia, with DKA/HHS
40-50 M	Pos, VL undetectable, CD4 329, unknown treatment status	Obesity	7 day history of cough, fever, sore throat, headache and neck stiffness. GCS 15 and unremarkable neurological examination.	Blood: WCC 5, CRP 74 CSF: Lymphocytes 2, erythrocytes 76, protein 0.42, glucose 5.5, negative gram stain and bacterial culture, GXPU trace on first LP, negative on second. TB culture negative.	Not done	COVID pneumonia, no cause for neurological symptoms identified
40-50 M	Neg	Hypertension	7 day history of cough, shortness of breath, sore throat and fever, new onset right sided weakness. Examination revealed reduced power (MRC 3/5 throughout) in right upper and lower limbs. GCS 15.	Blood: WCC 14, CRP 196. CSF: WCC 0, CSF protein 0.3, CSF glucose 3.0 (no blood glucose comparator), negative gram stain, no growth on bacterial culture, negative GXPU and CLAT.	Subacute left MCA territory infarct	COVID-19 pneumonia and stroke
60-70 F	Pos, unknown VL, CD4 191, on ART	Hypertension, DM, Obesity	7 day history of cough, shortness of breath and increasing confusion. GCS 14.	Blood: WCC 7, CRP 453, Na 130, D Dimer 4.94 CSF lymphocytes 4, erythrocytes 311, CSF protein 0.3, CSF glucose 10.2 (no blood glucose comparator), occasional lymphocytes seen on gram stain, negative bacterial culture, negative GXPU and CLAT.	Multi-territory mature infarcts in both cerebral hemispheres and in the cerebellum.	COVID-19 pneumonia and radiological diagnosis of multifocal infarcts

40-50 F	Pos, VL 411, CD4 unknown, on ART	Obesity	20 day history of GI symptoms with increasing weakness in both lower limbs. Pyramidal distribution weakness, hyperreflexia, T8 sensory level	CSF: Lymphocytes 12, erythrocytes 161, protein 2.19, glucose 2.8, Viral PCR positive for Epstein-Barr virus, HIV-1 viral load 838368 copies/mL	No brain imaging done  MRI spine reported as showing expanded poorly enhancing STIR/T2W hyperintense T7-T12 spinal cord associated with subtle enhancement of the adjacent leptomeninges. Arachnoiditis. Impression of cauda equina syndrome.	Viral myelitis with HIV CSF escape
80-90 F	Unknown,	Nil	7 day history of cough, fever, shortness of breath, lethargy, increasing confusion	Blood: Hb 10.0 CSF: WCC 0. protein 0.3, glucose 4.5, GS occasional lymphocytes, bacterial culture negative	No brain imaging done	COVID-19 pneumonia with new onset confusion

**Table A.5 Results of CSF PCR analysis for detection of SARS-CoV2**

Sample CODE	Primers	CT (run 22.1.21)	Freshly isolated RNA from 300ul CSF (run 1)	Freshly isolated RNA from 1ml CSF (run 2)
NTC	E/sgRNA/RP	U/U/U		
negative RNA	E/sgRNA/RP	U/U/19.64	Not run	Not run
positive RNA	E/sgRNA/RP	29.588/35.725/23.228		
A	E/sgRNA/RP	U/U/27.838	U/U/28.543	Not run
B	E/sgRNA/RP	U/U/32.167	U/U/31.723	Not run
C	E/sgRNA/RP	U/U/30.423	U/U/30.291	Not run
D	E/sgRNA/RP	U/U/33.769	U/U/34.833	Not run
E	E/sgRNA/RP	U/U/29.596	U/U/28.904	Not run
F	E/sgRNA/RP	U/U/29.508	U/U/29.433	Not run
G	E/sgRNA/RP	U/U/30.766	U/U/31.742	Not run
H	E/sgRNA/RP	U/U/28.1	U/U/27.882	Not run
J	E/sgRNA/RP	U/U/28.301	U/U/26.715	Not run
K	E/sgRNA/RP	U/U/30.967	U/U/31.610	Not run
L	E/sgRNA/RP	U/U/32.565	U/U/29.195	Not run
M	E/sgRNA/RP	U/U/29.434	U/U/31.586	Not run
N	E/sgRNA/RP	U/U/30.329	U/U/30.767	Not run
P	E/sgRNA/RP	U/U/32.947	U/U/32.885	Not run
Q	E/sgRNA/RP	U/U/32.693	U/U/33.181	Not run
R	E/sgRNA/RP	U/U/31.684	U/U/31.898	Not run

S	E/sgRNA/RP	U/U/31.920	U/U/32.882	Not run
T	E/sgRNA/RP	U/U/30.518	U/U/30.682	Not run
U	E/sgRNA/RP	U/U/21.694	U/U/21.572	Not run
V	E/sgRNA/RP	U/U/31.449	U/U/32.053	Not run
W	E/sgRNA/RP	U/U.27.471	U/U/26.683	Not run
X	E/sgRNA/RP	U/U/26.666	U/U/27.990	Not run
Y	E/sgRNA/RP	U/U/28.835	U/U/28.412	Not run
Z	E/sgRNA/RP	U/U/27.502	Not run	Not run
A2	E/sgRNA/RP	U/U/28.626	Not run	Not run
B2	E/sgRNA/RP	U/U/29.912	Not run	Not run
C2	E/sgRNA/RP	U/U/31.769	Not run	Not run
D2	E/sgRNA/RP	U/U/23.736	Not run	
E2	E/sgRNA/RP	37.867/39.461/30.956	Not run	U/U/29.642
F2	E/sgRNA/RP	U/U/24.674	Not run	U/U/30.549
G2	E/sgRNA/RP	U/38.837/31.717	Not run	U/U/30.549
H2	E/sgRNA/RP	40.430/U/27.301	Not run	U/U/24.015
J2	E/sgRNA/RP	U/U/26.379	Not run	Not run
K2	E/sgRNA/RP	U/U/29.545	Not run	Not run
L2	E/sgRNA/RP	U/38.438/23.510	Not run	U/U/21.109
M2	E/sgRNA/RP	U/U/29.039	Not run	Not run
N2	E/sgRNA/RP	U/U/19.058	Not run	Not run
P2	E/sgRNA/RP	U/38.087/27.014	Not run	U/U/29.229

Abbreviations: CT (cycle threshold), CSF (cerebrospinal fluid), NTC (no template control), RP (RNaseP), U (Undetermined)

## Reference List

- AARNOOTSE, R. E., KIBIKI, G. S., REITHER, K., SEMVUA, H. H., HARAHA, F., MTABHO, C. M., MPAGAMA, S. G., VAN DEN BOOGAARD, J., SUMARI-DE BOER, I. M., MAGIS-ESCURRA, C., WATTENBERG, M., LOGGER, J. G. M., TE BRAKE, L. H. M., HOELSCHER, M., GILLESPIE, S. H., COLBERS, A., PHILLIPS, P. P. J., PLEMPER VAN BALEN, G., BOEREE, M. J. & PAN, A. C. 2017. Pharmacokinetics, Tolerability, and Bacteriological Response of Rifampin Administered at 600, 900, and 1,200 Milligrams Daily in Patients with Pulmonary Tuberculosis. *Antimicrob Agents Chemother*, 61.
- ABDOOL KARIM, S. S., NAIDOO, K., GROBLER, A., PADAYATCHI, N., BAXTER, C., GRAY, A., GENGLIAH, T., NAIR, G., BAMBER, S., SINGH, A., KHAN, M., PIENAAR, J., EL-SADR, W., FRIEDLAND, G. & ABDOOL KARIM, Q. 2010. Timing of initiation of antiretroviral drugs during tuberculosis therapy. *N Engl J Med*, 362, 697-706.
- AHUJA, S. D., ASHKIN, D., AVENDANO, M., BANERJEE, R., BAUER, M., BAYONA, J. N., BECERRA, M. C., BENEDETTI, A., BURGOS, M., CENTIS, R., CHAN, E. D., CHIANG, C. Y., COX, H., D'AMBROSIO, L., DERIEMER, K., DUNG, N. H., ENARSON, D., FALZON, D., FLANAGAN, K., FLOOD, J., GARCIA-GARCIA, M. L., GANDHI, N., GRANICH, R. M., HOLLM-DELGADO, M. G., HOLTZ, T. H., ISEMAN, M. D., JARLSBERG, L. G., KESHAVJEE, S., KIM, H. R., KOH, W. J., LANCASTER, J., LANGE, C., DE LANGE, W. C., LEIMANE, V., LEUNG, C. C., LI, J., MENZIES, D., MIGLIORI, G. B., MISHUSTIN, S. P., MITNICK, C. D., NARITA, M., O'RIORDAN, P., PAI, M., PALMERO, D., PARK, S. K., PASVOL, G., PENA, J., PEREZ-GUZMAN, C., QUELAPIO, M. I., PONCE-DE-LEON, A., RIEKSTINA, V., ROBERT, J., ROYCE, S., SCHAAF, H. S., SEUNG, K. J., SHAH, L., SHIM, T. S., SHIN, S. S., SHIRAISHI, Y., SIFUENTES-OSORNIO, J., SOTGIU, G., STRAND, M. J., TABARSI, P., TUPASI, T. E., VAN ALTENA, R., VAN DER WALT, M., VAN DER WERF, T. S., VARGAS, M. H., VIKLEPP, P., WESTENHOUSE, J., YEW, W. W., YIM, J. J. & COLLABORATIVE GROUP FOR META-ANALYSIS OF INDIVIDUAL PATIENT DATA IN, M.-T. 2012. Multidrug resistant pulmonary tuberculosis treatment regimens and patient outcomes: an individual patient data meta-analysis of 9,153 patients. *PLoS Med*, 9, e1001300.
- ALBERTI, P., BERETTA, S., PIATTI, M., KARANTZOULIS, A., PIATTI, M. L., SANTORO, P., VIGANO, M., GIOVANNELLI, G., PIRRO, F., MONTISANO, D. A., APPOLLONIO, I. & FERRARESE, C. 2020. Guillain-Barre syndrome related to COVID-19 infection. *Neurol Neuroimmunol Neuroinflamm*, 7.
- ANDERSON, N. E., SOMARATNE, J., MASON, D. F., HOLLAND, D. & THOMAS, M. G. 2010. Neurological and systemic complications of tuberculous meningitis and its treatment at Auckland City Hospital, New Zealand. *J Clin Neurosci*, 17, 1114-8.

- ANGER, H. A., DWORKIN, F., SHARMA, S., MUNSIFF, S. S., NILSEN, D. M. & AHUJA, S. D. 2010. Linezolid use for treatment of multidrug-resistant and extensively drug-resistant tuberculosis, New York City, 2000-06. *J Antimicrob Chemother*, 65, 775-83.
- ANNANE, D. & SHARSHAR, T. 2015. Cognitive decline after sepsis. *Lancet Respir Med*, 3, 61-9.
- ANONYMOUS 1952. Treatment of pulmonary tuberculosis with isoniazid; an interim report to the Medical Research Council by their Tuberculosis Chemotherapy Trials Committee. *Br Med J*, 2, 735-46.
- ANONYMOUS 2010. *Treatment of Tuberculosis: Guidelines. 4th ed.* Geneva: World Health Organisation.
- AREESHI, M. Y., MANDAL, R. K., WAHID, M., DAR, S. A., JAWED, A., LOHANI, M., ABDALLAH, A. M. A., KHAN, S., PANDA, A. K., MISHRA, B. N. & HAQUE, S. 2017. Vitamin D Receptor Apal (rs7975232) Polymorphism Confers Decreased Risk of Pulmonary Tuberculosis in Overall and African Population, but not in Asians: Evidence from a Meta-analysis. *Ann Clin Lab Sci*, 47, 628-637.
- ASHWAL, S., HOLSHOUSER, B. A., SHU, S. K., SIMMONS, P. L., PERKIN, R. M., TOMASI, L. G., KNIERIM, D. S., SHERIDAN, C., CRAIG, K., ANDREWS, G. H. & HINSHAW, D. B. 2000. Predictive value of proton magnetic resonance spectroscopy in pediatric closed head injury. *Pediatr Neurol*, 23, 114-25.
- ATKINS, C. M., OLIVA, A. A., JR., ALONSO, O. F., PEARSE, D. D., BRAMLETT, H. M. & DIETRICH, W. D. 2007. Modulation of the cAMP signaling pathway after traumatic brain injury. *Exp Neurol*, 208, 145-58.
- ATLURI, V. S., HIDALGO, M., SAMIKKANNU, T., KURAPATI, K. R., JAYANT, R. D., SAGAR, V. & NAIR, M. P. 2015. Effect of human immunodeficiency virus on blood-brain barrier integrity and function: an update. *Front Cell Neurosci*, 9, 212.
- AVULA, A., NALLEBALLE, K., NARULA, N., SAPOZHNIKOV, S., DANDU, V., TOOM, S., GLASER, A. & ELSAYEGH, D. 2020. COVID-19 presenting as stroke. *Brain Behav Immun*.
- AYATA, C. & ROPPER, A. H. 2002. Ischaemic brain oedema. *J Clin Neurosci*, 9, 113-24.
- AZUAJE, C., FERNANDEZ HIDALGO, N., ALMIRANTE, B., MARTIN-CASABONA, N., RIBERA, E., DIAZ, M., PRATS, G. & PAHISSA, A. 2006. [Tuberculous meningitis: a comparative study in relation to concurrent human immunodeficiency virus infection]. *Enferm Infecc Microbiol Clin*, 24, 245-50.
- AZZURRI, A., SOW, O. Y., AMEDEI, A., BAH, B., DIALLO, S., PERI, G., BENAGIANO, M., D'ELIOS, M. M., MANTOVANI, A. & DEL PRETE, G. 2005. IFN-gamma-inducible protein 10 and pentraxin 3 plasma levels are tools for monitoring inflammation and disease activity in Mycobacterium tuberculosis infection. *Microbes Infect*, 7, 1-8.
- BABIKIAN, T., FREIER, M. C., ASHWAL, S., RIGGS, M. L., BURLEY, T. & HOLSHOUSER, B. A. 2006. MR spectroscopy: predicting long-term neuropsychological outcome following pediatric TBI. *J Magn Reson Imaging*, 24, 801-11.

- BANG, N. D., CAWS, M., TRUC, T. T., DUONG, T. N., DUNG, N. H., HA, D. T., THWAITES, G. E., HEEMSKERK, D., TARNING, J., MERSON, L., VAN TOI, P., FARRAR, J. J., WOLBERS, M., POUPLIN, T. & DAY, J. N. 2016. Clinical presentations, diagnosis, mortality and prognostic markers of tuberculous meningitis in Vietnamese children: a prospective descriptive study. *BMC Infect Dis*, 16, 573.
- BARBOR, T. F. H.-B., JOHN C.; SAUNDERS, JOHN B; MONTEIRO, MARISTELA G, WORLD HEALTH ORGANIZATION 2001. *AUDIT: the Alcohol Use Disorders Identification Test : guidelines for use in primary health care / Thomas F. Babor ... [et al.]*, 2nd ed. World Health Organization. .
- BARKHOUDARIAN, G., HOVDA, D. A. & GIZA, C. C. 2016. The Molecular Pathophysiology of Concussive Brain Injury - an Update. *Phys Med Rehabil Clin N Am*, 27, 373-93.
- BASTIEN, C. H., VALLIERES, A. & MORIN, C. M. 2001. Validation of the Insomnia Severity Index as an outcome measure for insomnia research. *Sleep Med*, 2, 297-307.
- BE, N. A., KIM, K. S., BISHAI, W. R. & JAIN, S. K. 2009. Pathogenesis of central nervous system tuberculosis. *Curr Mol Med*, 9, 94-9.
- BELL, M. J., TERHORST, L. & BENDER, C. M. 2013. Psychometric analysis of the Patient Assessment of Own Functioning Inventory in women with breast cancer. *J Nurs Meas*, 21, 320-34.
- BERGER, N. A., BESSON, V. C., BOULARES, A. H., BURKLE, A., CHIARUGI, A., CLARK, R. S., CURTIN, N. J., CUZZOCREA, S., DAWSON, T. M., DAWSON, V. L., HASKO, G., LIAUDET, L., MORONI, F., PACHER, P., RADERMACHER, P., SALZMAN, A. L., SNYDER, S. H., SORIANO, F. G., STROSZNAJDER, R. P., SUMEGI, B., SWANSON, R. A. & SZABO, C. 2018. Opportunities for the repurposing of PARP inhibitors for the therapy of non-oncological diseases. *Br J Pharmacol*, 175, 192-222.
- BERGSBAKEN, T., FINK, S. L. & COOKSON, B. T. 2009. Pyroptosis: host cell death and inflammation. *Nat Rev Microbiol*, 7, 99-109.
- BERMAN, A. H., BERGMAN, H., PALMSTIERN, T., & SCHLYTER, F. (2002). DRUG USE DISORDERS IDENTIFICATION TEST (DUDIT) [DATABASE RECORD]. APA PSYCTESTS. & [HTTPS://DOI.ORG/10.1037/T02890-000](https://doi.org/10.1037/T02890-000).
- BEYROUTI, R., ADAMS, M. E., BENJAMIN, L., COHEN, H., FARMER, S. F., GOH, Y. Y., HUMPHRIES, F., JAGER, H. R., LOSSEFF, N. A., PERRY, R. J., SHAH, S., SIMISTER, R. J., TURNER, D., CHANDRATHEVA, A. & WERRING, D. J. 2020. Characteristics of ischaemic stroke associated with COVID-19. *J Neurol Neurosurg Psychiatry*, 91, 889-891.
- BLANC, F. X., SOK, T., LAUREILLARD, D., BORAND, L., REKACEWICZ, C., NERRIENET, E., MADEC, Y., MARCY, O., CHAN, S., PRAK, N., KIM, C., LAK, K. K., HAK, C., DIM, B., SIN, C. I., SUN, S., GUILLARD, B., SAR, B., VONG, S., FERNANDEZ, M., FOX, L., DELFRAISSY, J. F., GOLDFELD, A. E. & TEAM, C. S. 2011. Earlier versus later start of antiretroviral therapy in HIV-infected adults with tuberculosis. *N Engl J Med*, 365, 1471-81.

- BLUMENTHAL, A., NAGALINGAM, G., HUCH, J. H., WALKER, L., GUILLEMIN, G. J., SMYTHE, G. A., EHRT, S., BRITTON, W. J. & SAUNDERS, B. M. 2012. M. tuberculosis induces potent activation of IDO-1, but this is not essential for the immunological control of infection. *PLoS One*, 7, e37314.
- BOEREE, M. J., DIACON, A. H., DAWSON, R., NARUNSKY, K., DU BOIS, J., VENTER, A., PHILLIPS, P. P., GILLESPIE, S. H., MCHUGH, T. D., HOELSCHER, M., HEINRICH, N., REHAL, S., VAN SOOLINGEN, D., VAN INGEN, J., MAGIS-ESCURRA, C., BURGER, D., PLEMPER VAN BALEN, G., AARNOUTSE, R. E. & PAN, A. C. 2015. A dose-ranging trial to optimize the dose of rifampin in the treatment of tuberculosis. *Am J Respir Crit Care Med*, 191, 1058-65.
- BOEREE, M. J., HEINRICH, N., AARNOUTSE, R., DIACON, A. H., DAWSON, R., REHAL, S., KIBIKI, G. S., CHURCHYARD, G., SANNE, I., NTINGINYA, N. E., MINJA, L. T., HUNT, R. D., CHARALAMBOUS, S., HANEKOM, M., SEMVUA, H. H., MPAGAMA, S. G., MANYAMA, C., MTAFYA, B., REITHER, K., WALLIS, R. S., VENTER, A., NARUNSKY, K., MEKOTA, A., HENNE, S., COLBERS, A., VAN BALEN, G. P., GILLESPIE, S. H., PHILLIPS, P. P. J., HOELSCHER, M. & PAN, A. C. 2017. High-dose rifampicin, moxifloxacin, and SQ109 for treating tuberculosis: a multi-arm, multi-stage randomised controlled trial. *Lancet Infect Dis*, 17, 39-49.
- BOSEL, J., GANDOR, F., HARMS, C., SYNOWITZ, M., HARMS, U., DJOUFACK, P. C., MEGOW, D., DIRNAGL, U., HORTNAGL, H., FINK, K. B. & ENDRES, M. 2005. Neuroprotective effects of atorvastatin against glutamate-induced excitotoxicity in primary cortical neurones. *J Neurochem*, 92, 1386-98.
- BOTHA, F. J., SIRGEL, F. A., PARKIN, D. P., VAN DE WAL, B. W., DONALD, P. R. & MITCHISON, D. A. 1996. Early bactericidal activity of ethambutol, pyrazinamide and the fixed combination of isoniazid, rifampicin and pyrazinamide (Rifater) in patients with pulmonary tuberculosis. *S Afr Med J*, 86, 155-8.
- BOULWARE, D. R., MEYA, D. B., BERGEMANN, T. L., WIESNER, D. L., RHEIN, J., MUSUBIRE, A., LEE, S. J., KAMBUGU, A., JANOFF, E. N. & BOHJANEN, P. R. 2010. Clinical features and serum biomarkers in HIV immune reconstitution inflammatory syndrome after cryptococcal meningitis: a prospective cohort study. *PLoS Med*, 7, e1000384.
- BOUSSER, M. G. 2009. Antithrombotic agents in the prevention of ischemic stroke. *Cerebrovasc Dis*, 27 Suppl 3, 12-9.
- BRICKNER, S. J., BARBACHYN, M. R., HUTCHINSON, D. K. & MANNINEN, P. R. 2008. Linezolid (ZYVOX), the first member of a completely new class of antibacterial agents for treatment of serious gram-positive infections. *J Med Chem*, 51, 1981-90.
- BROWN, A. N., DRUSANO, G. L., ADAMS, J. R., RODRIQUEZ, J. L., JAMBUNATHAN, K., BALUYA, D. L., BROWN, D. L., KWARA, A., MIRSALIS, J. C., HAFNER, R. & LOUIE, A. 2015. Preclinical Evaluations To Identify Optimal Linezolid Regimens for Tuberculosis Therapy. *MBio*, 6, e01741-15.



- BUTTERFIELD, D. A. & POCERNICH, C. B. 2003. The glutamatergic system and Alzheimer's disease. *CNS drugs*, 17, 641-652.
- CALVERLEY, P. M., RABE, K. F., GOEHRING, U. M., KRISTIANSEN, S., FABBRI, L. M., MARTINEZ, F. J., M & GROUPS, M. S. 2009. Roflumilast in symptomatic chronic obstructive pulmonary disease: two randomised clinical trials. *Lancet*, 374, 685-94.
- CAMPBELL, B. M., CHARYCH, E., LEE, A. W. & MOLLER, T. 2014. Kynurenines in CNS disease: regulation by inflammatory cytokines. *Front Neurosci*, 8, 12.
- CAMPO, M., RANDHAWA, A. K., DUNSTAN, S., FARRAR, J., CAWS, M., BANG, N. D., LAN, N. N., HONG CHAU, T. T., HORNE, D. J., THUONG, N. T., THWAITES, G. E. & HAWN, T. R. 2015. Common polymorphisms in the CD43 gene region are associated with tuberculosis disease and mortality. *Am J Respir Cell Mol Biol*, 52, 342-8.
- CAREY, C. L., WOODS, S. P., GONZALEZ, R., CONOVER, E., MARCOTTE, T. D., GRANT, I., HEATON, R. K. & GROUP, H. 2004. Predictive validity of global deficit scores in detecting neuropsychological impairment in HIV infection. *J Clin Exp Neuropsychol*, 26, 307-19.
- CASTILLO, J., LOZA, M. I., MIRELMAN, D., BREA, J., BLANCO, M., SOBRINO, T. & CAMPOS, F. 2016. A novel mechanism of neuroprotection: Blood glutamate grabber. *J Cereb Blood Flow Metab*, 36, 292-301.
- CAWS, M., THWAITES, G., DUNSTAN, S., HAWN, T. R., LAN, N. T., THUONG, N. T., STEPNIEWSKA, K., HUYEN, M. N., BANG, N. D., LOC, T. H., GAGNEUX, S., VAN SOOLINGEN, D., KREMER, K., VAN DER SANDE, M., SMALL, P., ANH, P. T., CHINH, N. T., QUY, H. T., DUYNEN, N. T., THO, D. Q., HIEU, N. T., TOROK, E., HIEN, T. T., DUNG, N. H., NHU, N. T., DUY, P. M., VAN VINH CHAU, N. & FARRAR, J. 2008. The influence of host and bacterial genotype on the development of disseminated disease with *Mycobacterium tuberculosis*. *PLoS Pathog*, 4, e1000034.
- CAWS, M., THWAITES, G., STEPNIEWSKA, K., NGUYEN, T. N., NGUYEN, T. H., NGUYEN, T. P., MAI, N. T., PHAN, M. D., TRAN, H. L., TRAN, T. H., VAN SOOLINGEN, D., KREMER, K., NGUYEN, V. V., NGUYEN, T. C. & FARRAR, J. 2006. Beijing genotype of *Mycobacterium tuberculosis* is significantly associated with human immunodeficiency virus infection and multidrug resistance in cases of tuberculous meningitis. *J Clin Microbiol*, 44, 3934-9.
- CECCHINI, D., AMBROSIONI, J., BREZZO, C., CORTI, M., RYBKO, A., PEREZ, M., POGGI, S. & AMBROGGI, M. 2009. Tuberculous meningitis in HIV-infected and non-infected patients: comparison of cerebrospinal fluid findings. *Int J Tuberc Lung Dis*, 13, 269-71.
- CHAGANTI, J. & BREW, B. J. 2021. MR spectroscopy in HIV associated neurocognitive disorder in the era of cART: a review. *AIDS Res Ther*, 18, 65.
- CHAMOUN, R., SUKI, D., GOPINATH, S. P., GOODMAN, J. C. & ROBERTSON, C. 2010. Role of extracellular glutamate measured by

- cerebral microdialysis in severe traumatic brain injury. *Journal of neurosurgery*, 113, 564-570.
- CHATAWAY, J., SCHUERER, N., ALSANOUSI, A., CHAN, D., MACMANUS, D., HUNTER, K., ANDERSON, V., BANGHAM, C. R., CLEGG, S., NIELSEN, C., FOX, N. C., WILKIE, D., NICHOLAS, J. M., CALDER, V. L., GREENWOOD, J., FROST, C. & NICHOLAS, R. 2014. Effect of high-dose simvastatin on brain atrophy and disability in secondary progressive multiple sclerosis (MS-STAT): a randomised, placebo-controlled, phase 2 trial. *Lancet*, 383, 2213-21.
- CHELUNE, G., HEATON, R. K. & LEHMAN, R. A. W. Neuropsychological and Personality Correlates of Patients' Complaints of Disability. 1986.
- CHELUNE, G. J., HEATON, R.K. AND LEHMAN, R.A., 1986. ADVANCES IN CLINICAL NEUROPSYCHOLOGY. US: SPRINGER, PP.95-126.
- CHEN, H. L., LU, C. H., CHANG, C. D., CHEN, P. C., CHEN, M. H., HSU, N. W., CHOU, K. H., LIN, W. M., LIN, C. P. & LIN, W. C. 2015. Structural deficits and cognitive impairment in tuberculous meningitis. *BMC Infect Dis*, 15, 279.
- CHEN, Y., ZHANG, J., WANG, X., WU, Y., ZHU, L., LU, L., SHEN, Q. & QIN, Y. 2016. HMGB1 level in cerebrospinal fluid as a complimentary biomarker for the diagnosis of tuberculous meningitis. *Springerplus*, 5, 1775.
- CHIGUTSA, E., VISSER, M. E., SWART, E. C., DENTI, P., PUSHPAKOM, S., EGAN, D., HOLFORD, N. H., SMITH, P. J., MAARTENS, G., OWEN, A. & MCILLERON, H. 2011. The SLCO1B1 rs4149032 polymorphism is highly prevalent in South Africans and is associated with reduced rifampin concentrations: dosing implications. *Antimicrob Agents Chemother*, 55, 4122-7.
- CHIREHWA, M. T., RUSTOMJEE, R., MTHIYANE, T., ONYEBUJOH, P., SMITH, P., MCILLERON, H. & DENTI, P. 2015. Model-Based Evaluation of Higher Doses of Rifampin Using a Semimechanistic Model Incorporating Autoinduction and Saturation of Hepatic Extraction. *Antimicrob Agents Chemother*, 60, 487-94.
- CHOTMONGKOL, V., JITPIMOLMARD, S. & THAVORNPITAK, Y. 1996. Corticosteroid in tuberculous meningitis. *J Med Assoc Thai*, 79, 83-90.
- CHURCHWARD, M. A. & TODD, K. G. 2014. Statin treatment affects cytokine release and phagocytic activity in primary cultured microglia through two separable mechanisms. *Mol Brain*, 7, 85.
- CIANFONI, A., NIKU, S. & IMBESI, S. G. 2007a. Metabolite Findings in Tumefactive Demyelinating Lesions Utilizing Short Echo Time Proton Magnetic Resonance Spectroscopy. *American Journal of Neuroradiology*, 28, 272-277.
- CIANFONI, A., NIKU, S. & IMBESI, S. G. 2007b. Metabolite findings in tumefactive demyelinating lesions utilizing short echo time proton magnetic resonance spectroscopy. *AJNR Am J Neuroradiol*, 28, 272-7.
- COICO, R. & SUNSHINE, G. 2009. *Immunology : a short course*, Hoboken, N.J, Wiley-Blackwell.
- COMAS, I., COSCOLLA, M., LUO, T., BORRELL, S., HOLT, K. E., KATOMAEDA, M., PARKHILL, J., MALLA, B., BERG, S., THWAITES, G., YEBOAH-MANU, D., BOTHAMLEY, G., MEI, J., WEI, L., BENTLEY, S.,

- HARRIS, S. R., NIEMANN, S., DIEHL, R., ASEFFA, A., GAO, Q., YOUNG, D. & GAGNEUX, S. 2013. Out-of-Africa migration and Neolithic coexpansion of *Mycobacterium tuberculosis* with modern humans. *Nat Genet*, 45, 1176-82.
- CONRADIE, F., DIACON, A. H., NGUBANE, N., HOWELL, P., EVERITT, D., CROOK, A. M., MENDEL, C. M., EGIZI, E., MOREIRA, J., TIMM, J., MCHUGH, T. D., WILLS, G. H., BATESON, A., HUNT, R., VAN NIEKERK, C., LI, M., OLUGBOSI, M., SPIGELMAN, M. & NIX, T. B. T. T. 2020. Treatment of Highly Drug-Resistant Pulmonary Tuberculosis. *N Engl J Med*, 382, 893-902.
- CORDLE, A. & LANDRETH, G. 2005. 3-Hydroxy-3-methylglutaryl-coenzyme A reductase inhibitors attenuate beta-amyloid-induced microglial inflammatory responses. *J Neurosci*, 25, 299-307.
- CORMAN, V. M., LANDT, O., KAISER, M., MOLENKAMP, R., MEIJER, A., CHU, D. K., BLEICKER, T., BRUNINK, S., SCHNEIDER, J., SCHMIDT, M. L., MULDER, D. G., HAAGMANS, B. L., VAN DER VEER, B., VAN DEN BRINK, S., WIJSMAN, L., GODERSKI, G., ROMETTE, J. L., ELLIS, J., ZAMBON, M., PEIRIS, M., GOOSSENS, H., REUSKEN, C., KOOPMANS, M. P. & DROSTEN, C. 2020. Detection of 2019 novel coronavirus (2019-nCoV) by real-time RT-PCR. *Euro Surveill*, 25.
- CORNBLATH, D. R., CHAUDHRY, V., CARTER, K., LEE, D., SEYSEDADR, M., MIERNICKI, M. & JOH, T. 1999. Total neuropathy score: validation and reliability study. *Neurology*, 53, 1660-4.
- COSCOLLA, M. & GAGNEUX, S. 2014. Consequences of genomic diversity in *Mycobacterium tuberculosis*. *Semin Immunol*, 26, 431-44.
- COURT, R., CHIREHWA, M. T., WIESNER, L., WRIGHT, B., SMYTHE, W., KRAMER, N. & MCILLERON, H. 2018. Quality assurance of rifampicin-containing fixed-drug combinations in South Africa: dosing implications. *Int J Tuberc Lung Dis*, 22, 537-543.
- CRESSWELL, F. V., MEYA, D. B., KAGIMU, E., GRINT, D., TE BRAKE, L., KASIBANTE, J., MARTYN, E., RUTAKINGIRWA, M., QUINN, C. M., OKIRWOTH, M., TUGUME, L., SSEMBAMBULIDDE, K., MUSUBIRE, A. K., BANGDIWALA, A. S., BUZIBYE, A., MUZOORA, C., SVENSSON, E. M., AARNOUTSE, R., BOULWARE, D. R. & ELLIOTT, A. M. 2021. High-Dose Oral and Intravenous Rifampicin for the Treatment of Tuberculous Meningitis in Predominantly Human Immunodeficiency Virus (HIV)-Positive Ugandan Adults: A Phase II Open-Label Randomized Controlled Trial. *Clin Infect Dis*, 73, 876-884.
- CRUIKSHANK, W. W., KORNFELD, H. & CENTER, D. M. 2000. Interleukin-16. *J Leukoc Biol*, 67, 757-66.
- CURTO, M., REALI, C., PALMIERI, G., SCINTU, F., SCHIVO, M. L., SOGOS, V., MARCIALIS, M. A., ENNAS, M. G., SCHWARZ, H., POZZI, G. & GREMO, F. 2004. Inhibition of cytokines expression in human microglia infected by virulent and non-virulent mycobacteria. *Neurochem Int*, 44, 381-92.
- DA SILVA-CANDAL, A., PEREZ-DIAZ, A., SANTAMARIA, M., CORREA-PAZ, C., RODRIGUEZ-YANEZ, M., ARDA, A., PEREZ-MATO, M., IGLESIAS-REY, R., BREA, J., AZUAJE, J., SOTELO, E., SOBRINO, T., LOZA, M.

- I., CASTILLO, J. & CAMPOS, F. 2018. Clinical validation of blood/brain glutamate grabbing in acute ischemic stroke. *Ann Neurol*, 84, 260-273.
- DANIEL, P. M. 1949. Gross morbid anatomy of the central nervous system of cases of tuberculous meningitis treated with streptomycin. *Proc R Soc Med*, 42, 169-74.
- DASTUR, D. K., MANGHANI, D. K. & UDANI, P. M. 1995. Pathology and pathogenetic mechanisms in neurotuberculosis. *Radiol Clin North Am*, 33, 733-52.
- DAVIS, A., MEINTJES, G. & WILKINSON, R. J. 2018. Treatment of Tuberculous Meningitis and Its Complications in Adults. *Curr Treat Options Neurol*, 20, 5.
- DAVIS, A. G., DONOVAN, J., BREMER, M., VAN TOORN, R., SCHOEMAN, J., DADABHOY, A., LAI, R. P. J., CRESSWELL, F. V., BOULWARE, D. R., WILKINSON, R. J., THUONG, N. T. T., THWAITES, G. E., BAHR, N. C. & TUBERCULOUS MENINGITIS INTERNATIONAL RESEARCH, C. 2020. Host Directed Therapies for Tuberculous Meningitis. *Wellcome Open Res*, 5, 292.
- DAVIS, A. G., DREYER, A. J., ALBERTYN, C., MAXEBENGULA, M., STEK, C., WASSERMAN, S., MARAIS, S., BATEMAN, K., SOLMS, M., JOSKA, J., WILKINSON, R. J. & NIGHTINGALE, S. 2022a. Cognitive Impairment in Tuberculous Meningitis. *Clin Infect Dis*.
- DAVIS, A. G., NIGHTINGALE, S., SPRINGER, P. E., SOLOMONS, R., ARENIVAS, A., WILKINSON, R. J., ANDERSON, S. T., CHOW, F. C. & TUBERCULOUS MENINGITIS INTERNATIONAL RESEARCH, C. 2019a. Neurocognitive and functional impairment in adult and paediatric tuberculous meningitis. *Wellcome Open Res*, 4, 178.
- DAVIS, A. G., ROHLWINK, U. K., PROUST, A., FIGAJI, A. A. & WILKINSON, R. J. 2019b. The pathogenesis of tuberculous meningitis. *J Leukoc Biol*, 105, 267-280.
- DAVIS, A. G., WASSERMAN, S., MAXEBENGULA, M., STEK, C., BREMER, M., DAROOWALA, R., AZIZ, S., GOLIATH, R., STEGMANN, S., KOEKEMOER, S., JACKSON, A., LAI SAI, L., KADERNANI, Y., SIHOYIYA, T., LIANG, C. J., DODD, L., DENTI, P., CREDE, T., NAUDE, J., SZYMANSKI, P., VALLIE, Y., BANDERKER, I., MOOSA, S., RAUBENHEIMER, P., LAI, R. P. J., JOSKA, J., NIGHTINGALE, S., DREYER, A., WAHL, G., OFFIAH, C., VORSTER, I., CANDY, S., ROBERTSON, F., MEINTJES, E., MAARTENS, G., BLACK, J., MEINTJES, G. & WILKINSON, R. J. 2021. Study protocol for a phase 2A trial of the safety and tolerability of increased dose rifampicin and adjunctive linezolid, with or without aspirin, for HIV-associated tuberculous meningitis [LASER-TBM]. *Wellcome Open Res*, 6, 136.
- DAVIS, A. G., WASSERMAN, S., STEK, C., MAXEBENGULA, M., LIANG, C. J., STEGMANN, S., KOEKEMOER, S., JACKSON, A., KADERNANI, Y., BREMER, M., DAROOWALA, R., AZIZ, S., GOLIATH, R., SAI, L. L., SIHOYIYA, T., DENTI, P., LAI, R. P., CREDE, T., NAUDE, J., SZYMANSKI, P., VALLIE, Y., BANDERKER, I. A., MOOSA, M. S., RAUBENHEIMER, P., CANDY, S., OFFIAH, C., WAHL, G., VORSTER, I., MAARTENS, G., BLACK, J., MEINTJES, G. & WILKINSON, R. J.

- 2022b. A phase 2A trial of the safety and tolerability of increased dose rifampicin and adjunctive linezolid, with or without aspirin, for HIV-associated tuberculous meningitis (The LASER-TBM Trial). *Clin Infect Dis*.
- DE JONG, B. C., HILL, P. C., AIKEN, A., AWINE, T., ANTONIO, M., ADETIFA, I. M., JACKSON-SILLAH, D. J., FOX, A., DERIEMER, K., GAGNEUX, S., BORGDORFF, M. W., MCADAM, K. P., CORRAH, T., SMALL, P. M. & ADEGBOLA, R. A. 2008. Progression to active tuberculosis, but not transmission, varies by *Mycobacterium tuberculosis* lineage in The Gambia. *J Infect Dis*, 198, 1037-43.
- DE LA RIVA, P., URTASUN, M., CASTILLO-TRIVINO, T., CAMINO, X., ARRUTI, M., MONDRAGON, E. & LOPEZ DE MUNAIN, A. 2013. Clinical response to thalidomide in the treatment of intracranial tuberculomas: case report. *Clin Neuropharmacol*, 36, 70-2.
- DE MIRANDA HENRIQUES-SOUZA, A. M., DE MELO, A., DE AGUIAR COELHO SILVA MADEIRO, B., FREITAS, L. F., SAMPAIO ROCHA-FILHO, P. A. & GONCALVES, F. G. 2021. Acute disseminated encephalomyelitis in a COVID-19 pediatric patient. *Neuroradiology*, 63, 141-145.
- DE STEENWINKEL, J. E., AARNOUTSE, R. E., DE KNEGT, G. J., TEN KATE, M. T., TEULEN, M., VERBRUGH, H. A., BOEREE, M. J., VAN SOOLINGEN, D. & BAKKER-WOUDENBERG, I. A. 2013. Optimization of the rifampin dosage to improve the therapeutic efficacy in tuberculosis treatment using a murine model. *Am J Respir Crit Care Med*, 187, 1127-34.
- DE VRIES, H. E., KUIPER, J., DE BOER, A. G., VAN BERKEL, T. J. & BREIMER, D. D. 1997. The blood-brain barrier in neuroinflammatory diseases. *Pharmacol Rev*, 49, 143-55.
- DENKINGER, C. M., KIK, S. V., CIRILLO, D. M., CASENGHI, M., SHINNICK, T., WEYER, K., GILPIN, C., BOEHME, C. C., SCHITO, M. & KIMERLING, M. 2015. Defining the needs for next generation assays for tuberculosis. *The Journal of infectious diseases*, 211, S29-S38.
- DIACON, A. H., PATIENTIA, R. F., VENTER, A., VAN HELDEN, P. D., SMITH, P. J., MCILLERON, H., MARITZ, J. S. & DONALD, P. R. 2007. Early bactericidal activity of high-dose rifampin in patients with pulmonary tuberculosis evidenced by positive sputum smears. *Antimicrob Agents Chemother*, 51, 2994-6.
- DING, J., THUY THUONG THUONG, N., PHAM, T. V., HEEMSKERK, D., POUPLIN, T., TRAN, C. T. H., NGUYEN, M. T. H., NGUYEN, P. H., PHAN, L. P., NGUYEN, C. V. V., THWAITES, G. & TARNING, J. 2020. Pharmacokinetics and Pharmacodynamics of Intensive Antituberculosis Treatment of Tuberculous Meningitis. *Clin Pharmacol Ther*, 107, 1023-1033.
- DING, Y., HE, L., ZHANG, Q., HUANG, Z., CHE, X., HOU, J., WANG, H., SHEN, H., QIU, L., LI, Z., GENG, J., CAI, J., HAN, H., LI, X., KANG, W., WENG, D., LIANG, P. & JIANG, S. 2004. Organ distribution of severe acute respiratory syndrome (SARS) associated coronavirus (SARS-CoV)

- in SARS patients: implications for pathogenesis and virus transmission pathways. *J Pathol*, 203, 622-30.
- DINGLELINE, R., BORGES, K., BOWIE, D. & TRAYNELIS, S. F. 1999. The glutamate receptor ion channels. *Pharmacol Rev*, 51, 7-61.
- DOBSON, R., TOPPING, J., DAVIS, A., THOMPSON, E. & GIOVANNONI, G. 2013. Cerebrospinal fluid and urinary biomarkers in multiple sclerosis. *Acta Neurol Scand*, 128, 321-7.
- DONALD, P. R. 2010. Cerebrospinal fluid concentrations of antituberculosis agents in adults and children. *Tuberculosis (Edinb)*, 90, 279-92.
- DONALD, P. R., SCHAAF, H. S. & SCHOEMAN, J. F. 2005. Tuberculous meningitis and miliary tuberculosis: the Rich focus revisited. *J Infect*, 50, 193-5.
- DORMANS, J., BURGER, M., AGUILAR, D., HERNANDEZ-PANDO, R., KREMER, K., ROHOLL, P., AREND, S. M. & VAN SOOLINGEN, D. 2004. Correlation of virulence, lung pathology, bacterial load and delayed type hypersensitivity responses after infection with different *Mycobacterium tuberculosis* genotypes in a BALB/c mouse model. *Clin Exp Immunol*, 137, 460-8.
- DUNSTAN, S. J., TRAM, T. T., THWAITES, G. E., CHAU, T. T., PHU, N. H., HIEN, T. T., FARRAR, J. J., WOLBERS, M. & MAI, N. T. 2015. LTA4H genotype is associated with susceptibility to bacterial meningitis but is not a critical determinant of outcome. *PLoS One*, 10, e0118789.
- DUTTA, N. K., BRUINERS, N., PINN, M. L., ZIMMERMAN, M. D., PRIDEAUX, B., DARTOIS, V., GENNARO, M. L. & KARAKOUSIS, P. C. 2016. Statin adjunctive therapy shortens the duration of TB treatment in mice. *J Antimicrob Chemother*, 71, 1570-7.
- EAST AFRICAN/BRITISH MEDICAL RESEARCH COUNCILS 1972. Controlled clinical trial of short-course (6-month) regimens of chemotherapy for treatment of pulmonary tuberculosis. *Lancet*, 1, 1079-85.
- EDDEN, R. A., PUTS, N. A., HARRIS, A. D., BARKER, P. B. & EVANS, C. J. 2014. Gannet: A batch-processing tool for the quantitative analysis of gamma-aminobutyric acid-edited MR spectroscopy spectra. *J Magn Reson Imaging*, 40, 1445-52.
- EISELE, A., HILL-STRATHY, M., MICHELS, L. & RAUEN, K. 2020. Magnetic Resonance Spectroscopy following Mild Traumatic Brain Injury: A Systematic Review and Meta-Analysis on the Potential to Detect Posttraumatic Neurodegeneration. *Neurodegener Dis*, 20, 2-11.
- EL OTMANI, H., EL MOUTAWAKIL, B., RAFAI, M. A., EL BENNA, N., EL KETTANI, C., SOUSSI, M., EL MDAGHRI, N., BARROU, H. & AFIF, H. 2020. Covid-19 and Guillain-Barre syndrome: More than a coincidence! *Rev Neurol (Paris)*, 176, 518-519.
- ELLARD, G. A., HUMPHRIES, M. J. & ALLEN, B. W. 1993. Cerebrospinal fluid drug concentrations and the treatment of tuberculous meningitis. *Am Rev Respir Dis*, 148, 650-5.
- ERIBO, O. A., LEQHEKA, M. S., MALHERBE, S. T., MCANDA, S., STANLEY, K., VAN DER SPUY, G. D., WALZL, G. & CHEGOU, N. N. 2020. Host urine immunological biomarkers as potential candidates for the diagnosis of tuberculosis. *International Journal of Infectious Diseases*, 99, 473-481.

- ERSOY, Y., YETKIN, F., BAYRAKTAR, M. R., ERSOY, Y. & YOLOGLU, S. 2012. A new diagnostic scoring for discrimination of tuberculous and bacterial meningitis on the basis of clinical and laboratory findings. *Med Princ Pract*, 21, 259-63.
- ESPINOSA, P. S., RIZVI, Z., SHARMA, P., HINDI, F. & FILATOV, A. 2020. Neurological Complications of Coronavirus Disease (COVID-19): Encephalopathy, MRI Brain and Cerebrospinal Fluid Findings: Case 2. *Cureus*, 12, e7930.
- EVANS, S. R., CLIFFORD, D. B., KITCH, D. W., GOODKIN, K., SCHIFITTO, G., MCARTHUR, J. C. & SIMPSON, D. M. 2008. Simplification of the research diagnosis of HIV-associated sensory neuropathy. *HIV Clin Trials*, 9, 434-9.
- FALZON, D., SCHUNEMANN, H. J., HARAUSZ, E., GONZALEZ-ANGULO, L., LIENHARDT, C., JARAMILLO, E. & WEYER, K. 2017. World Health Organization treatment guidelines for drug-resistant tuberculosis, 2016 update. *Eur Respir J*, 49.
- FARESJO, M. 2014. A useful guide for analysis of immune markers by fluorochrome (Luminex) technique. *Methods Mol Biol*, 1172, 87-96.
- FAVA, V. M. & SCHURR, E. 2017. Evaluating the Impact of LTA4H Genotype and Immune Status on Survival From Tuberculous Meningitis. *J Infect Dis*, 215, 1011-1013.
- FERNANDO, S. L., SAUNDERS, B. M., SLUYTER, R., SKARRATT, K. K., GOLDBERG, H., MARKS, G. B., WILEY, J. S. & BRITTON, W. J. 2007. A polymorphism in the P2X7 gene increases susceptibility to extrapulmonary tuberculosis. *Am J Respir Crit Care Med*, 175, 360-6.
- FORD, C. W., ZURENKO, G. E. & BARBACHYN, M. R. 2001. The discovery of linezolid, the first oxazolidinone antibacterial agent. *Curr Drug Targets Infect Disord*, 1, 181-99.
- FORTUN, J., MARTIN-DAVILA, P., NAVAS, E., PEREZ-ELIAS, M. J., COBO, J., TATO, M., DE LA PEDROSA, E. G., GOMEZ-MAMPASO, E. & MORENO, S. 2005. Linezolid for the treatment of multidrug-resistant tuberculosis. *J Antimicrob Chemother*, 56, 180-5.
- FOX, E., OLIVER, T., ROWE, M., THOMAS, S., ZAKHARIA, Y., GILMAN, P. B., MULLER, A. J. & PRENDERGAST, G. C. 2018. Indoximod: An Immunometabolic Adjuvant That Empowers T Cell Activity in Cancer. *Front Oncol*, 8, 370.
- FRANCONY, G., FAUVAGE, B., FALCON, D., CANET, C., DILOU, H., LAVAGNE, P., JACQUOT, C. & PAYEN, J. F. 2008. Equimolar doses of mannitol and hypertonic saline in the treatment of increased intracranial pressure. *Crit Care Med*, 36, 795-800.
- GAGNEUX, S., DERIEMER, K., VAN, T., KATO-MAEDA, M., DE JONG, B. C., NARAYANAN, S., NICOL, M., NIEMANN, S., KREMER, K., GUTIERREZ, M. C., HILTY, M., HOPEWELL, P. C. & SMALL, P. M. 2006. Variable host-pathogen compatibility in Mycobacterium tuberculosis. *Proc Natl Acad Sci U S A*, 103, 2869-73.
- GAGNEUX, S. & SMALL, P. M. 2007. Global phylogeography of Mycobacterium tuberculosis and implications for tuberculosis product development. *Lancet Infect Dis*, 7, 328-37.

- GAMIR-MORRALLA, A., LÓPEZ-MENÉNDEZ, C., MEDINA, M. & IGLESIAS, T. 2017. Chapter 7 - A Novel Neuroprotection Target With Distinct Regulation in Stroke and Alzheimer's Disease. *In: GOZES, I. (ed.) Neuroprotection in Alzheimer's Disease*. Academic Press.
- GAUTAM, U. S., FOREMAN, T. W., BUCSAN, A. N., VEATCH, A. V., ALVAREZ, X., ADEKAMBI, T., GOLDEN, N. A., GENTRY, K. M., DOYLE-MEYERS, L. A., RUSSELL-LODRIGUE, K. E., DIDIER, P. J., BLANCHARD, J. L., KOUSOULAS, K. G., LACKNER, A. A., KALMAN, D., RENGARAJAN, J., KHADER, S. A., KAUSHAL, D. & MEHRA, S. 2018. In vivo inhibition of tryptophan catabolism reorganizes the tuberculoma and augments immune-mediated control of Mycobacterium tuberculosis. *Proc Natl Acad Sci U S A*, 115, E62-E71.
- GIRGIS, N. I., FARID, Z., KILPATRICK, M. E., SULTAN, Y. & MIKHAIL, I. A. 1991. Dexamethasone adjunctive treatment for tuberculous meningitis. *Pediatr Infect Dis J*, 10, 179-83.
- GOLDBERG, M. F., GOLDBERG, M. F., CEREJO, R. & TAYAL, A. H. 2020. Cerebrovascular Disease in COVID-19. *AJNR Am J Neuroradiol*.
- GONG, B., VITOLO, O. V., TRINCHESE, F., LIU, S., SHELANSKI, M. & ARANCIO, O. 2004. Persistent improvement in synaptic and cognitive functions in an Alzheimer mouse model after rolipram treatment. *J Clin Invest*, 114, 1624-34.
- GONZALEZ-GARCIA, C., BRAVO, B., BALLESTER, A., GOMEZ-PEREZ, R., EGUILUZ, C., REDONDO, M., MARTINEZ, A., GIL, C. & BALLESTER, S. 2013. Comparative assessment of PDE 4 and 7 inhibitors as therapeutic agents in experimental autoimmune encephalomyelitis. *Br J Pharmacol*, 170, 602-13.
- GOPAL, R., MONIN, L., TORRES, D., SLIGHT, S., MEHRA, S., MCKENNA, K. C., FALLERT JUNECKO, B. A., REINHART, T. A., KOLLS, J., BAEZ-SALDANA, R., CRUZ-LAGUNAS, A., RODRIGUEZ-REYNA, T. S., KUMAR, N. P., TESSIER, P., ROTH, J., SELMAN, M., BECERRIL-VILLANUEVA, E., BAQUERA-HEREDIA, J., CUMMING, B., KASPROWICZ, V. O., STEYN, A. J., BABU, S., KAUSHAL, D., ZUNIGA, J., VOGL, T., RANGEL-MORENO, J. & KHADER, S. A. 2013. S100A8/A9 proteins mediate neutrophilic inflammation and lung pathology during tuberculosis. *Am J Respir Crit Care Med*, 188, 1137-46.
- GRAUSTEIN, A. D., HORNE, D. J., ARENTZ, M., BANG, N. D., CHAU, T. T., THWAITES, G. E., CAWS, M., THUONG, N. T., DUNSTAN, S. J. & HAWN, T. R. 2015. TLR9 gene region polymorphisms and susceptibility to tuberculosis in Vietnam. *Tuberculosis (Edinb)*, 95, 190-6.
- GRUPKE, S., HALL, J., DOBBS, M., BIX, G. J. & FRASER, J. F. 2015. Understanding history, and not repeating it. Neuroprotection for acute ischemic stroke: from review to preview. *Clin Neurol Neurosurg*, 129, 1-9.
- GU, J., GONG, E., ZHANG, B., ZHENG, J., GAO, Z., ZHONG, Y., ZOU, W., ZHAN, J., WANG, S., XIE, Z., ZHUANG, H., WU, B., ZHONG, H., SHAO, H., FANG, W., GAO, D., PEI, F., LI, X., HE, Z., XU, D., SHI, X., ANDERSON, V. M. & LEONG, A. S. 2005. Multiple organ infection and the pathogenesis of SARS. *J Exp Med*, 202, 415-24.



- GUERRA-ASSUNCAO, J. A., CRAMPIN, A. C., HOUBEN, R. M., MZEMBE, T., MALLARD, K., COLL, F., KHAN, P., BANDA, L., CHIWAYA, A., PEREIRA, R. P., MCNERNEY, R., FINE, P. E., PARKHILL, J., CLARK, T. G. & GLYNN, J. R. 2015a. Large-scale whole genome sequencing of *M. tuberculosis* provides insights into transmission in a high prevalence area. *Elife*, 4.
- GUERRA-ASSUNCAO, J. A., HOUBEN, R. M., CRAMPIN, A. C., MZEMBE, T., MALLARD, K., COLL, F., KHAN, P., BANDA, L., CHIWAYA, A., PEREIRA, R. P., MCNERNEY, R., HARRIS, D., PARKHILL, J., CLARK, T. G. & GLYNN, J. R. 2015b. Recurrence due to relapse or reinfection with *Mycobacterium tuberculosis*: a whole-genome sequencing approach in a large, population-based cohort with a high HIV infection prevalence and active follow-up. *J Infect Dis*, 211, 1154-63.
- GUNTARD, H. F., SAAG, M. S., BENSON, C. A., DEL RIO, C., ERON, J. J., GALLANT, J. E., HOY, J. F., MUGAVERO, M. J., SAX, P. E., THOMPSON, M. A., GANDHI, R. T., LANDOVITZ, R. J., SMITH, D. M., JACOBSEN, D. M. & VOLBERDING, P. A. 2016. Antiretroviral Drugs for Treatment and Prevention of HIV Infection in Adults: 2016 Recommendations of the International Antiviral Society-USA Panel. *Jama*, 316, 191-210.
- GUTIERREZ-ORTIZ, C., MENDEZ, A., RODRIGO-REY, S., SAN PEDRO-MURILLO, E., BERMEJO-GUERRERO, L., GORDO-MANAS, R., DE ARAGON-GOMEZ, F. & BENITO-LEON, J. 2020. Miller Fisher Syndrome and polyneuritis cranialis in COVID-19. *Neurology*.
- HARRIS, A. D., PUTS, N. A. & EDDEN, R. A. 2015. Tissue correction for GABA-edited MRS: Considerations of voxel composition, tissue segmentation, and tissue relaxations. *J Magn Reson Imaging*, 42, 1431-40.
- HASHIMOTO, M., ISHIKAWA, Y., YOKOTA, S., GOTO, F., BANDO, T., SAKAKIBARA, Y. & IRIKI, M. 1991. Action site of circulating interleukin-1 on the rabbit brain. *Brain Res*, 540, 217-23.
- HAVLIR, D. V., KENDALL, M. A., IVE, P., KUMWENDA, J., SWINDELLS, S., QASBA, S. S., LUETKEMEYER, A. F., HOGG, E., ROONEY, J. F., WU, X., HOSSEINIPOUR, M. C., LALLOO, U., VELOSO, V. G., SOME, F. F., KUMARASAMY, N., PADAYATCHI, N., SANTOS, B. R., REID, S., HAKIM, J., MOHAPI, L., MUGYENYI, P., SANCHEZ, J., LAMA, J. R., PAPE, J. W., SANCHEZ, A., ASMELASH, A., MOKO, E., SAWE, F., ANDERSEN, J., SANNE, I. & A, A. C. T. G. S. 2011. Timing of antiretroviral therapy for HIV-1 infection and tuberculosis. *N Engl J Med*, 365, 1482-91.
- HAWN, T. R., DUNSTAN, S. J., THWAITES, G. E., SIMMONS, C. P., THUONG, N. T., LAN, N. T. N., QUY, H. T., CHAU, T. T. H., HIEU, N. T., RODRIGUES, S., JANER, M., ZHAO, L. P., HIEN, T. T., FARRAR, J. J. & ADEREM, A. 2006. A polymorphism in Toll-interleukin 1 receptor domain containing adaptor protein is associated with susceptibility to meningeal tuberculosis. *J Infect Dis*, 194, 1127-1134.

- HAZANY, S., HESSELINK, J. R., HEALY, J. F. & IMBESI, S. G. 2007. Utilization of glutamate/creatine ratios for proton spectroscopic diagnosis of meningiomas. *Neuroradiology*, 49, 121-7.
- HEEMSKERK, A. D., BANG, N. D., MAI, N. T., CHAU, T. T., PHU, N. H., LOC, P. P., CHAU, N. V., HIEN, T. T., DUNG, N. H., LAN, N. T., LAN, N. H., LAN, N. N., PHONG LE, T., VIEN, N. N., HIEN, N. Q., YEN, N. T., HA, D. T., DAY, J. N., CAWS, M., MERSON, L., THINH, T. T., WOLBERS, M., THWAITES, G. E. & FARRAR, J. J. 2016. Intensified Antituberculosis Therapy in Adults with Tuberculous Meningitis. *N Engl J Med*, 374, 124-34.
- HEIZMANN, C. W., FRITZ, G. & SCHAFFER, B. W. 2002. S100 proteins: structure, functions and pathology. *Front Biosci*, 7, d1356-68.
- HERBST, S., MARAIS, S., GUTIERREZ, M. G., WADDELL, S. J., WILKINSON, R. J. & LAI, R. P. 2019. Elevated LRRK2 and  $\alpha$ -synuclein levels in CSF of infectious meningitis patients. *bioRxiv*, 599381.
- HERNANDEZ PANDO, R., AGUILAR, D., COHEN, I., GUERRERO, M., RIBON, W., ACOSTA, P., OROZCO, H., MARQUINA, B., SALINAS, C., REMBAO, D. & ESPITIA, C. 2010. Specific bacterial genotypes of Mycobacterium tuberculosis cause extensive dissemination and brain infection in an experimental model. *Tuberculosis (Edinb)*, 90, 268-77.
- HESS, D. C., ELDAHSHAN, W. & RUTKOWSKI, E. 2020. COVID-19-Related Stroke. *Transl Stroke Res*, 11, 322-325.
- HOAL-VAN HELDEN, E. G., EPSTEIN, J., VICTOR, T. C., HON, D., LEWIS, L. A., BEYERS, N., ZURAKOWSKI, D., EZEKOWITZ, A. B. & VAN HELDEN, P. D. 1999. Mannose-binding protein B allele confers protection against tuberculous meningitis. *Pediatr Res*, 45, 459-64.
- HOFFMANN, C., WELZ, T., SABRANSKI, M., KOLB, M., WOLF, E., STELLBRINK, H. J. & WYEN, C. 2017. Higher rates of neuropsychiatric adverse events leading to dolutegravir discontinuation in women and older patients. *HIV Med*, 18, 56-63.
- HONG, J. Y., LEE, H. J., KIM, S. Y., CHUNG, K. S., KIM, E. Y., JUNG, J. Y., PARK, M. S., KIM, Y. S., KIM, S. K., CHANG, J., CHO, S. N. & KANG, Y. A. 2014. Efficacy of IP-10 as a biomarker for monitoring tuberculosis treatment. *J Infect*, 68, 252-8.
- HOPPE, H. C., DE WET, B. J., CYWES, C., DAFTE, M. & EHLERS, M. R. 1997. Identification of phosphatidylinositol mannoside as a mycobacterial adhesin mediating both direct and opsonic binding to nonphagocytic mammalian cells. *Infect Immun*, 65, 3896-905.
- HORNE, D. J., RANDHAWA, A. K., CHAU, T. T., BANG, N. D., YEN, N. T., FARRAR, J. J., DUNSTAN, S. J. & HAWN, T. R. 2012. Common polymorphisms in the PKP3-SIGIRR-TMEM16J gene region are associated with susceptibility to tuberculosis. *J Infect Dis*, 205, 586-94.
- HOVENS, M. M., SNOEP, J. D., GROENEVELD, Y., FROLICH, M., TAMSMA, J. T. & HUISMAN, M. V. 2008. Effects of aspirin on serum C-reactive protein and interleukin-6 levels in patients with type 2 diabetes without cardiovascular disease: a randomized placebo-controlled crossover trial. *Diabetes Obes Metab*, 10, 668-74.

[HTTPS://WWW.SAMRC.AC.ZA/REPORTS/REPORT-WEEKLY-DEATHS-SOUTH-AFRICA?BC=254](https://www.samrc.ac.za/reports/report-weekly-deaths-south-africa?bc=254).

- JAIN, S. K., PAUL-SATYASEELA, M., LAMICHHANE, G., KIM, K. S. & BISHAI, W. R. 2006. Mycobacterium tuberculosis invasion and traversal across an in vitro human blood-brain barrier as a pathogenic mechanism for central nervous system tuberculosis. *J Infect Dis*, 193, 1287-95.
- JIA, M., NJAPO, S. A., RASTOGI, V. & HEDNA, V. S. 2015. Taming glutamate excitotoxicity: strategic pathway modulation for neuroprotection. *CNS Drugs*, 29, 153-62.
- JICK, H., ZORNBERG, G. L., JICK, S. S., SESHADRI, S. & DRACHMAN, D. A. 2000. Statins and the risk of dementia. *Lancet*, 356, 1627-31.
- JINDANI, A., ABER, V. R., EDWARDS, E. A. & MITCHISON, D. A. 1980. The early bactericidal activity of drugs in patients with pulmonary tuberculosis. *Am Rev Respir Dis*, 121, 939-49.
- JM SCOTT, C., M SZILAGYI, G., CHAVEZ, S., GANDA, A. & E BLACK, S. 2011. Applications of chemical shift imaging for AD. *Current Medical Imaging*, 7, 88-95.
- JOHNSON, T. P., PATEL, K., JOHNSON, K. R., MARIC, D., CALABRESI, P. A., HASBUN, R. & NATH, A. 2013. Induction of IL-17 and nonclassical T-cell activation by HIV-Tat protein. *Proc Natl Acad Sci U S A*, 110, 13588-93.
- JOSKA, J. A., GOUSE, H., PAUL, R. H., STEIN, D. J. & FLISHER, A. J. 2010. Does highly active antiretroviral therapy improve neurocognitive function? A systematic review. *J Neurovirol*, 16, 101-14.
- JOSKA, J. A., WITTEN, J., THOMAS, K. G., ROBERTSON, C., CASSON-CROOK, M., ROOSA, H., CREIGHTON, J., LYONS, J., MCARTHUR, J. & SACKTOR, N. C. 2016. A Comparison of Five Brief Screening Tools for HIV-Associated Neurocognitive Disorders in the USA and South Africa. *AIDS Behav*, 20, 1621-31.
- KAISER, L. G., SCHUFF, N., CASHDOLLAR, N. & WEINER, M. W. 2005. Age-related glutamate and glutamine concentration changes in normal human brain: 1H MR spectroscopy study at 4 T. *Neurobiol Aging*, 26, 665-72.
- KALIA, L. V., KALIA, S. K. & SALTER, M. W. 2008. NMDA receptors in clinical neurology: excitatory times ahead. *Lancet Neurol*, 7, 742-55.
- KALITA, J., MISRA, U. K. & NAIR, P. P. 2009. Predictors of stroke and its significance in the outcome of tuberculous meningitis. *J Stroke Cerebrovasc Dis*, 18, 251-8.
- KALITA, J., MISRA, U. K. & RANJAN, P. 2007. Predictors of long-term neurological sequelae of tuberculous meningitis: a multivariate analysis. *Eur J Neurol*, 14, 33-7.
- KANG ZHAO, J. H., DAN DAI, YUWEI FENG, LIMING LIU, SHUKE NIE Acute myelitis after SARS-CoV-2 infection: a case report. *medRxiv* 2020.03.16.20035105; .
- KANG ZHAO, J. H., DAN DAI, YUWEI FENG, LIMING LIU, SHUKE NIE, ACUTE MYELITIS AFTER SARS-COV-2 INFECTION: A CASE REPORT. *MEDRXIV* 2020.03.16.20035105.
- KAPLAN, G. & FREEDMAN, V. H. 1996. The role of cytokines in the immune response to tuberculosis. *Res Immunol*, 147, 565-72.

- KARANDE, S., GUPTA, V., KULKARNI, M., JOSHI, A. & RELE, M. 2005. Tuberculous meningitis and HIV. *Indian J Pediatr*, 72, 755-60.
- KARSTAEDT, A. S., VALTCHANOVA, S., BARRIERE, R. & CREWE-BROWN, H. H. 1998. Tuberculous meningitis in South African urban adults. *QJM*, 91, 743-7.
- KASS, J. S. & SHANDERA, W. X. 2010. Nervous system effects of antituberculosis therapy. *CNS Drugs*, 24, 655-67.
- KATA, D., FOLDESI, I., FEHER, L. Z., HACKLER, L., JR., PUSKAS, L. G. & GULYA, K. 2016. Rosuvastatin enhances anti-inflammatory and inhibits pro-inflammatory functions in cultured microglial cells. *Neuroscience*, 314, 47-63.
- KATO, M., MOCHIZUKI, T., NEGARO, K., FUKUSAKO, T., NOGAKI, H. & MORIMATSU, M. 1997. [Magnetic resonance imaging of a case of central nervous system tuberculosis with tuberculous arachnoiditis and multiple tuberculomas]. *Nihon Ronen Igakkai Zasshi*, 34, 818-24.
- KATRAK, S. M., SHEMBALKAR, P. K., BIJWE, S. R. & BHANDARKAR, L. D. 2000. The clinical, radiological and pathological profile of tuberculous meningitis in patients with and without human immunodeficiency virus infection. *J Neurol Sci*, 181, 118-26.
- KEELEY, A. J., PARKASH, V., TUNBRIDGE, A., GREIG, J., COLLINI, P., MCKANE, W. & TATTERSALL, R. S. 2020. Anakinra in the treatment of protracted paradoxical inflammatory reactions in HIV-associated tuberculosis in the United Kingdom: a report of two cases. *Int J STD AIDS*, 31, 808-812.
- KHAMBATI, N., OLBRICH, L., ELLNER, J., SALGAME, P., SONG, R. & BIJKER, E. M. 2021. Host-Based Biomarkers in Saliva for the Diagnosis of Pulmonary Tuberculosis in Children: A Mini-Review. *Front Pediatr*, 9, 756043.
- KHODAMORADI, Z., HOSSEINI, S. A., GHOLAMPOOR SAADI, M. H., MEHRABI, Z., SASANI, M. R. & YAGHOUBI, S. 2020a. COVID-19 meningitis without pulmonary involvement with positive cerebrospinal fluid PCR. *Eur J Neurol*.
- KHODAMORADI, Z., HOSSEINI, S. A., GHOLAMPOOR SAADI, M. H., MEHRABI, Z., SASANI, M. R. & YAGHOUBI, S. 2020b. COVID-19 meningitis without pulmonary involvement with positive cerebrospinal fluid PCR. *Eur J Neurol*, 27, 2668-2669.
- KIM, H., LEE, J. M., PARK, J. S., JO, S. A., KIM, Y. O., KIM, C. W. & JO, I. 2008. Dexamethasone coordinately regulates angiotensin-1 and VEGF: a mechanism of glucocorticoid-induced stabilization of blood-brain barrier. *Biochem Biophys Res Commun*, 372, 243-8.
- KO, H. L. & REN, E. C. 2012. Functional Aspects of PARP1 in DNA Repair and Transcription. *Biomolecules*, 2, 524-48.
- KOEKEN, V., GANIEM, A. R., DIAN, S., RUSLAMI, R., CHAIDIR, L., NETEA, M. G., KUMAR, V., ALISJAHBANA, B., VAN CREVEL, R. & VAN LAARHOVEN, A. 2021. Cerebrospinal fluid IL-1beta is elevated in tuberculous meningitis patients but not associated with mortality. *Tuberculosis (Edinb)*, 126, 102019.

- KONG, Y., CAVE, M. D., ZHANG, L., FOXMAN, B., MARRS, C. F., BATES, J. H. & YANG, Z. H. 2007. Association between Mycobacterium tuberculosis Beijing/W lineage strain infection and extrathoracic tuberculosis: Insights from epidemiologic and clinical characterization of the three principal genetic groups of M. tuberculosis clinical isolates. *J Clin Microbiol*, 45, 409-14.
- KONRAD, F. M., BURY, A., SCHICK, M. A., NGAMSRI, K. C. & REUTERSHAN, J. 2015. The unrecognized effects of phosphodiesterase 4 on epithelial cells in pulmonary inflammation. *PLoS One*, 10, e0121725.
- KRISHNAN, N., ROBERTSON, B. D. & THWAITES, G. 2010. The mechanisms and consequences of the extra-pulmonary dissemination of Mycobacterium tuberculosis. *Tuberculosis (Edinb)*, 90, 361-6.
- KUMAR, R., PANDEY, C. K., BOSE, N. & SAHAY, S. 2002. Tuberculous brain abscess: clinical presentation, pathophysiology and treatment (in children). *Childs Nerv Syst*, 18, 118-23.
- KUMAR R, S. V. 2004. Tuberculous brain stem abscesses in children. *Journal of Pediatric Neurology*, 2, 101-106.
- KUMAR, V. & KUMAR, V. 2010. *Robbins and Cotran's pathologic basis of disease*, Philadelphia, Pa. ; London, Saunders.
- KUMARVELU, S., PRASAD, K., KHOSLA, A., BEHARI, M. & AHUJA, G. K. 1994. Randomized controlled trial of dexamethasone in tuberculous meningitis. *Tuber Lung Dis*, 75, 203-7.
- LAI, C. C., LEE, M. T., LEE, S. H., HSU, W. T., CHANG, S. S., CHEN, S. C. & LEE, C. C. 2016. Statin treatment is associated with a decreased risk of active tuberculosis: an analysis of a nationally representative cohort. *Thorax*, 71, 646-51.
- LAI, R. P., MEINTJES, G., WILKINSON, K. A., GRAHAM, C. M., MARAIS, S., VAN DER PLAS, H., DEFFUR, A., SCHUTZ, C., BLOOM, C., MUNAGALA, I., ANGUIANO, E., GOLIATH, R., MAARTENS, G., BANCHEREAU, J., CHAUSSABEL, D., O'GARRA, A. & WILKINSON, R. J. 2015. HIV-tuberculosis-associated immune reconstitution inflammatory syndrome is characterized by Toll-like receptor and inflammasome signalling. *Nat Commun*, 6, 8451.
- LAI, R. P., NAKIWALA, J. K., MEINTJES, G. & WILKINSON, R. J. 2013. The immunopathogenesis of the HIV tuberculosis immune reconstitution inflammatory syndrome. *Eur J Immunol*, 43, 1995-2002.
- LAMMIE, G. A., HEWLETT, R. H., SCHOEMAN, J. F. & DONALD, P. R. 2009. Tuberculous cerebrovascular disease: a review. *J Infect*, 59, 156-66.
- LANDIS, J. R. & KOCH, G. G. 1977. The measurement of observer agreement for categorical data. *Biometrics*, 33, 159-74.
- LANGE, C., STORKEBAUM, E., DE ALMODOVAR, C. R., DEWERCHIN, M. & CARMELIET, P. 2016. Vascular endothelial growth factor: a neurovascular target in neurological diseases. *Nat Rev Neurol*, 12, 439-54.
- LAWN, S. D., WILKINSON, R. J., LIPMAN, M. C. & WOOD, R. 2008. Immune reconstitution and "unmasking" of tuberculosis during antiretroviral therapy. *Am J Respir Crit Care Med*, 177, 680-5.

- LAWTON, M. P. & BRODY, E. M. 1969. Assessment of older people: self-maintaining and instrumental activities of daily living. *Gerontologist*, 9, 179-86.
- LEE, J. Y., YIM, J. J. & YOON, B. W. 2012a. Adjuvant interferon-gamma treatment in two cases of refractory tuberculosis of the brain. *Clin Neurol Neurosurg*, 114, 732-4.
- LEE, M., LEE, J., CARROLL, M. W., CHOI, H., MIN, S., SONG, T., VIA, L. E., GOLDFEDER, L. C., KANG, E., JIN, B., PARK, H., KWAK, H., KIM, H., JEON, H. S., JEONG, I., JOH, J. S., CHEN, R. Y., OLIVIER, K. N., SHAW, P. A., FOLLMANN, D., SONG, S. D., LEE, J. K., LEE, D., KIM, C. T., DARTOIS, V., PARK, S. K., CHO, S. N. & BARRY, C. E., 3RD 2012b. Linezolid for treatment of chronic extensively drug-resistant tuberculosis. *N Engl J Med*, 367, 1508-18.
- LERNER, T. R., DE SOUZA CARVALHO-WODARZ, C., REPNIK, U., RUSSELL, M. R., BOREL, S., DIEDRICH, C. R., ROHDE, M., WAINWRIGHT, H., COLLINSON, L. M., WILKINSON, R. J., GRIFFITHS, G. & GUTIERREZ, M. G. 2016. Lymphatic endothelial cells are a replicative niche for Mycobacterium tuberculosis. *J Clin Invest*, 126, 1093-108.
- LESNIAK, W. G., JYOTI, A., MISHRA, M. K., LOUISSAINT, N., ROMERO, R., CHUGANI, D. C., KANNAN, S. & KANNAN, R. M. 2013. Concurrent quantification of tryptophan and its major metabolites. *Anal Biochem*, 443, 222-31.
- LI, H., LU, J., LIU, J., ZHAO, Y., NI, X. & ZHAO, S. 2016. Linezolid is Associated with Improved Early Outcomes of Childhood Tuberculous Meningitis. *Pediatr Infect Dis J*, 35, 607-10.
- LIM, Y. S., KIM, S. B., KIM, M. K. & LIM, Y. J. 2013. Disseminated tuberculosis of central nervous system : spinal intramedullary and intracranial tuberculomas. *J Korean Neurosurg Soc*, 54, 61-4.
- LIPTON, S. A. & ROSENBERG, P. A. 1994. Excitatory amino acids as a final common pathway for neurologic disorders. *N Engl J Med*, 330, 613-22.
- LOWE, D. M., REDFORD, P. S., WILKINSON, R. J., O'GARRA, A. & MARTINEAU, A. R. 2012. Neutrophils in tuberculosis: friend or foe? *Trends Immunol*, 33, 14-25.
- MACKIEWICZ, A., SPEROFF, T., GANAPATHI, M. K. & KUSHNER, I. 1991. Effects of cytokine combinations on acute phase protein production in two human hepatoma cell lines. *J Immunol*, 146, 3032-7.
- MACMILLAN, E. L., TAM, R., ZHAO, Y., VAVASOUR, I. M., LI, D. K., OGER, J., FREEDMAN, M. S., KOLIND, S. H. & TRABOULSEE, A. L. 2016. Progressive multiple sclerosis exhibits decreasing glutamate and glutamine over two years. *Mult Scler*, 22, 112-6.
- MAHASE, E. 2019. GSK recalls ranitidine products over potential carcinogen contamination. *BMJ*, 367, l5933.
- MAHON, R. N. & HAFNER, R. 2015. Immune Cell Regulatory Pathways Unexplored as Host-Directed Therapeutic Targets for Mycobacterium tuberculosis: An Opportunity to Apply Precision Medicine Innovations to Infectious Diseases. *Clin Infect Dis*, 61Suppl 3, S200-16.

- MAI, N. T., DOBBS, N., PHU, N. H., COLAS, R. A., THAO, L. T., THUONG, N. T., NGHIA, H. D., HANH, N. H., HANG, N. T., HEEMSKERK, A. D., DAY, J. N., LY, L., THU, D. D., MERSON, L., KESTELYN, E., WOLBERS, M., GESKUS, R., SUMMERS, D., CHAU, N. V., DALLI, J. & THWAITES, G. E. 2018. A randomised double blind placebo controlled phase 2 trial of adjunctive aspirin for tuberculous meningitis in HIV-uninfected adults. *Elife*, 7.
- MAIGA, M., AGARWAL, N., AMMERMAN, N. C., GUPTA, R., GUO, H., MAIGA, M. C., LUN, S. & BISHAI, W. R. 2012. Successful shortening of tuberculosis treatment using adjuvant host-directed therapy with FDA-approved phosphodiesterase inhibitors in the mouse model. *PLoS One*, 7, e30749.
- MAIGA, M. C., AHIDJO, B. A., MAIGA, M. & BISHAI, W. R. 2015. Roflumilast, a Type 4 Phosphodiesterase Inhibitor, Shows Promising Adjunctive, Host-Directed Therapeutic Activity in a Mouse Model of Tuberculosis. *Antimicrob Agents Chemother*, 59, 7888-90.
- MAJLATH, Z., TAJTI, J. & VECSEI, L. 2013. Kynurenines and other novel therapeutic strategies in the treatment of dementia. *Ther Adv Neurol Disord*, 6, 386-97.
- MALHOTRA, H. S., GARG, R. K., SINGH, M. K., AGARWAL, A. & VERMA, R. 2009. Corticosteroids (dexamethasone versus intravenous methylprednisolone) in patients with tuberculous meningitis. *Ann Trop Med Parasitol*, 103, 625-34.
- MANCA, C., KOO, M. S., PEIXOTO, B., FALLOWS, D., KAPLAN, G. & SUBBIAN, S. 2013. Host targeted activity of pyrazinamide in Mycobacterium tuberculosis infection. *PLoS One*, 8, e74082.
- MANIAS, K. A. & PEET, A. 2018. What is MR spectroscopy? *Arch Dis Child Educ Pract Ed*, 103, 213-216.
- MANYELO, C. M., CHEGOU, N. N., SEDDON, J. A., SNYDERS, C. I., MUTAVHATSINDI, H., MANNINGO, P. M., WALZL, G., STANLEY, K. & SOLOMONS, R. S. 2021. Serum and cerebrospinal fluid host proteins indicate stroke in children with tuberculous meningitis. *PLoS One*, 16, e0250944.
- MANYELO, C. M., SOLOMONS, R. S., SNYDERS, C. I., KIDD, M., KOOBLAL, Y., LEUKES, V. N., CLAASSEN, C., ROOS, K., STANLEY, K., WALZL, G., CHEGOU, N. N. & GROUP, T. S. 2022. Validation of host cerebrospinal fluid protein biomarkers for early diagnosis of tuberculous meningitis in children: a replication and new biosignature discovery study. *Biomarkers*, 1-13.
- MANYELO, C. M., SOLOMONS, R. S., SNYDERS, C. I., MANNINGO, P. M., MUTAVHATSINDI, H., KRIEL, B., STANLEY, K., WALZL, G. & CHEGOU, N. N. 2019a. Application of Cerebrospinal Fluid Host Protein Biosignatures in the Diagnosis of Tuberculous Meningitis in Children from a High Burden Setting. *Mediators Inflamm*, 2019, 7582948.
- MANYELO, C. M., SOLOMONS, R. S., SNYDERS, C. I., MUTAVHATSINDI, H., MANNINGO, P. M., STANLEY, K., WALZL, G. & CHEGOU, N. N. 2019b. Potential of Host Serum Protein Biomarkers in the Diagnosis of Tuberculous Meningitis in Children. *Front Pediatr*, 7, 376.

- MARAIS, B. J., CHEONG, E., FERNANDO, S., DANIEL, S., WATTS, M. R., BERGLUND, L. J., BARRY, S. E., KOTSIU, G., HEADLEY, A. P. & STAPLEDON, R. A. 2021. Use of Infliximab to Treat Paradoxical Tuberculous Meningitis Reactions. *Open Forum Infect Dis*, 8, ofaa604.
- MARAIS, S., LAI, R. P. J., WILKINSON, K. A., MEINTJES, G., O'GARRA, A. & WILKINSON, R. J. 2017. Inflammasome Activation Underlying Central Nervous System Deterioration in HIV-Associated Tuberculosis. *J Infect Dis*, 215, 677-686.
- MARAIS, S., MEINTJES, G., PEPPER, D. J., DODD, L. E., SCHUTZ, C., ISMAIL, Z., WILKINSON, K. A. & WILKINSON, R. J. 2013. Frequency, severity, and prediction of tuberculous meningitis immune reconstitution inflammatory syndrome. *Clin Infect Dis*, 56, 450-60.
- MARAIS, S., PEPPER, D. J., SCHUTZ, C., WILKINSON, R. J. & MEINTJES, G. 2011. Presentation and outcome of tuberculous meningitis in a high HIV prevalence setting. *PLoS One*, 6, e20077.
- MARAIS, S., THWAITES, G., SCHOEMAN, J. F., TOROK, M. E., MISRA, U. K., PRASAD, K., DONALD, P. R., WILKINSON, R. J. & MARAIS, B. J. 2010. Tuberculous meningitis: a uniform case definition for use in clinical research. *Lancet Infect Dis*, 10, 803-12.
- MARAIS, S., VAN TOORN, R., CHOW, F. C., MANESH, A., SIDDIQI, O. K., FIGAJI, A., SCHOEMAN, J. F., MEINTJES, G. & TUBERCULOUS MENINGITIS INTERNATIONAL RESEARCH, C. 2019. Management of intracranial tuberculous mass lesions: how long should we treat for? *Wellcome Open Res*, 4, 158.
- MARAIS, S., WILKINSON, K. A., LESOSKY, M., COUSSENS, A. K., DEFFUR, A., PEPPER, D. J., SCHUTZ, C., ISMAIL, Z., MEINTJES, G. & WILKINSON, R. J. 2014. Neutrophil-associated central nervous system inflammation in tuberculous meningitis immune reconstitution inflammatory syndrome. *Clin Infect Dis*, 59, 1638-47.
- MARASHLY, E. T. & BOHLEGA, S. A. 2017. Riboflavin Has Neuroprotective Potential: Focus on Parkinson's Disease and Migraine. *Front Neurol*, 8, 333.
- MARITZ, J., BENATAR, M., DAVE, J. A., HARRISON, T. B., BADRI, M., LEVITT, N. S. & HECKMANN, J. M. 2010. HIV neuropathy in South Africans: frequency, characteristics, and risk factors. *Muscle Nerve*, 41, 599-606.
- MASON, G. F. & KRYSTAL, J. H. 2006. MR spectroscopy: its potential role for drug development for the treatment of psychiatric diseases. *NMR Biomed*, 19, 690-701.
- MASON, S., REINECKE, C. J., KULIK, W., VAN CRUCHTEN, A., SOLOMONS, R. & VAN FURTH, A. M. 2016. Cerebrospinal fluid in tuberculous meningitis exhibits only the L-enantiomer of lactic acid. *BMC Infect Dis*, 16, 251.
- MASTROIANNI, C. M., PAOLETTI, F., LICHTNER, M., D'AGOSTINO, C., VULLO, V. & DELIA, S. 1997. Cerebrospinal fluid cytokines in patients with tuberculous meningitis. *Clin Immunol Immunopathol*, 84, 171-6.
- MARTHUR, J. H. 1998. The reliability and validity of the subjective peripheral neuropathy screen. *J Assoc Nurses AIDS Care*, 9, 84-94.



- MCFARLAND, A. J., DAVEY, A. K., MCDERMOTT, C. M., GRANT, G. D., LEWOHL, J. & ANOOPKUMAR-DUKIE, S. 2018. Differences in statin associated neuroprotection corresponds with either decreased production of IL-1beta or TNF-alpha in an in vitro model of neuroinflammation-induced neurodegeneration. *Toxicol Appl Pharmacol*, 344, 56-73.
- MCILLERON, H., RUSTOMJEE, R., VAHEDI, M., MTHIYANE, T., DENTI, P., CONNOLLY, C., RIDA, W., PYM, A., SMITH, P. J. & ONYEBUJOH, P. C. 2012. Reduced antituberculosis drug concentrations in HIV-infected patients who are men or have low weight: implications for international dosing guidelines. *Antimicrob Agents Chemother*, 56, 3232-8.
- MEINTJES G, MOORHOUSE MA, CARMONA S, ET AL 2017. Adult antiretroviral therapy guidelines 2017. *S Afr J HIV Med.*, 18, a776.
- MEINTJES, G., LAWN, S. D., SCANO, F., MAARTENS, G., FRENCH, M. A., WORODRIA, W., ELLIOTT, J. H., MURDOCH, D., WILKINSON, R. J., SEYLER, C., JOHN, L., VAN DER LOEFF, M. S., REISS, P., LYNEN, L., JANOFF, E. N., GILKS, C., COLEBUNDERS, R. & INTERNATIONAL NETWORK FOR THE STUDY OF, H. I. V. A. I. 2008. Tuberculosis-associated immune reconstitution inflammatory syndrome: case definitions for use in resource-limited settings. *Lancet Infect Dis*, 8, 516-23.
- MEINTJES, G., WILKINSON, R. J., MORRONI, C., PEPPER, D. J., REBE, K., RANGAKA, M. X., ONI, T. & MAARTENS, G. 2010. Randomized placebo-controlled trial of prednisone for paradoxical tuberculosis-associated immune reconstitution inflammatory syndrome. *AIDS*, 24, 2381-90.
- MELDRUM, B. S. 2000. Glutamate as a neurotransmitter in the brain: review of physiology and pathology. *J Nutr*, 130, 1007S-15S.
- MICHAEL, B. D., GRIFFITHS, M. J., GRANEROD, J., BROWN, D., KEIR, G., WNEK, M., COX, D. J., VIDYASAGAR, R., BORROW, R., PARKES, L. M. & SOLOMON, T. 2016. The Interleukin-1 Balance During Encephalitis Is Associated With Clinical Severity, Blood-Brain Barrier Permeability, Neuroimaging Changes, and Disease Outcome. *J Infect Dis*, 213, 1651-60.
- MICHIKAWA, M. & YANAGISAWA, K. 1999. Inhibition of cholesterol production but not of nonsterol isoprenoid products induces neuronal cell death. *J Neurochem*, 72, 2278-85.
- MIGLIORI, G. B., EKER, B., RICHARDSON, M. D., SOTGIU, G., ZELLWEGER, J. P., SKRAHINA, A., ORTMANN, J., GIRARDI, E., HOFFMANN, H., BESOZZI, G., BEVILACQUA, N., KIRSTEN, D., CENTIS, R., LANGE, C. & GROUP, T. S. 2009. A retrospective TBNET assessment of linezolid safety, tolerability and efficacy in multidrug-resistant tuberculosis. *Eur Respir J*, 34, 387-93.
- MIKKELSEN, M., BARKER, P. B., BHATTACHARYYA, P. K., BRIX, M. K., BUUR, P. F., CECIL, K. M., CHAN, K. L., CHEN, D. Y., CRAVEN, A. R., CUYPERS, K., DACKO, M., DUNCAN, N. W., DYDAK, U., EDMONDSON, D. A., ENDE, G., ERSLAND, L., GAO, F., GREENHOUSE, I., HARRIS, A. D., HE, N., HEBA, S., HOGGARD, N.,

- HSU, T. W., JANSEN, J. F. A., KANGARLU, A., LANGE, T., LABEL, R. M., LI, Y., LIN, C. E., LIOU, J. K., LIRNG, J. F., LIU, F., MA, R., MAES, C., MORENO-ORTEGA, M., MURRAY, S. O., NOAH, S., NOESKE, R., NOSEWORTHY, M. D., OELTZSCHNER, G., PRISCIANDARO, J. J., PUTS, N. A. J., ROBERTS, T. P. L., SACK, M., SAILASUTA, N., SALEH, M. G., SCHALLMO, M. P., SIMARD, N., SWINNEN, S. P., TEGENTHOFF, M., TRUONG, P., WANG, G., WILKINSON, I. D., WITTSACK, H. J., XU, H., YAN, F., ZHANG, C., ZIPUNNIKOV, V., ZOLLNER, H. J. & EDDEN, R. A. E. 2017. Big GABA: Edited MR spectroscopy at 24 research sites. *Neuroimage*, 159, 32-45.
- MILLER, R. F., ISAACSON, P. G., HALL-CRAGGS, M., LUCAS, S., GRAY, F., SCARAVILLI, F. & AN, S. F. 2004. Cerebral CD8+ lymphocytosis in HIV-1 infected patients with immune restoration induced by HAART. *Acta Neuropathol*, 108, 17-23.
- MISRA, U. K., KALITA, J., BHOI, S. K. & SINGH, R. K. 2016. A study of hyponatremia in tuberculous meningitis. *J Neurol Sci*, 367, 152-7.
- MISRA, U. K., KALITA, J. & MAURYA, P. K. 2011. Stroke in tuberculous meningitis. *J Neurol Sci*, 303, 22-30.
- MISRA, U. K., KALITA, J. & NAIR, P. P. 2010. Role of aspirin in tuberculous meningitis: a randomized open label placebo controlled trial. *J Neurol Sci*, 293, 12-7.
- MITCHISON, D. A. 2000. Role of individual drugs in the chemotherapy of tuberculosis. *Int J Tuberc Lung Dis*, 4, 796-806.
- MOGHTADERI, A. & ALAVI NAINI, R. 2003. Tuberculous radiculomyelitis: review and presentation of five patients. *Int J Tuberc Lung Dis*, 7, 1186-90.
- MOHAMED, M. A., BARKER, P. B., SKOLASKY, R. L., SELNES, O. A., MOXLEY, R. T., POMPER, M. G. & SACKTOR, N. C. 2010. Brain metabolism and cognitive impairment in HIV infection: a 3-T magnetic resonance spectroscopy study. *Magn Reson Imaging*, 28, 1251-7.
- MOLLER, K., LARSEN, F. S., BIE, P. & SKINHJØJ, P. 2001. The syndrome of inappropriate secretion of antidiuretic hormone and fluid restriction in meningitis--how strong is the evidence? *Scand J Infect Dis*, 33, 13-26.
- MOLTON, J. S., HUGGAN, P. J. & ARCHULETA, S. 2015. Infliximab therapy in two cases of severe neurotuberculosis paradoxical reaction. *Med J Aust*, 202, 156-7.
- MORE, A., VERMA, R., GARG, R. K., MALHOTRA, H. S., SHARMA, P. K., UNİYAL, R., PANDEY, S. & MITTAL, M. 2017. A study of neuroendocrine dysfunction in patients of tuberculous meningitis. *J Neurol Sci*, 379, 198-206.
- MORIGUCHI, T., HARI, N., GOTO, J., HARADA, D., SUGAWARA, H., TAKAMINO, J., UENO, M., SAKATA, H., KONDO, K., MYOSE, N., NAKAO, A., TAKEDA, M., HARO, H., INOUE, O., SUZUKI-INOUE, K., KUBOKAWA, K., OGIHARA, S., SASAKI, T., KINOUCI, H., KOJIN, H., ITO, M., ONISHI, H., SHIMIZU, T., SASAKI, Y., ENOMOTO, N., ISHIHARA, H., FURUYA, S., YAMAMOTO, T. & SHIMADA, S. 2020. A first case of meningitis/encephalitis associated with SARS-Coronavirus-2. *Int J Infect Dis*, 94, 55-58.

- MUHLERT, N., ATZORI, M., DE VITA, E., THOMAS, D. L., SAMSON, R. S., WHEELER-KINGSHOTT, C. A., GEURTS, J. J., MILLER, D. H., THOMPSON, A. J. & CICCARELLI, O. 2014. Memory in multiple sclerosis is linked to glutamate concentration in grey matter regions. *J Neurol Neurosurg Psychiatry*, 85, 833-9.
- NARAYANA, P. A. 2005. Magnetic resonance spectroscopy in the monitoring of multiple sclerosis. *J Neuroimaging*, 15, 46S-57S.
- NASREDDINE, Z. S., PHILLIPS, N. A., BEDIRIAN, V., CHARBONNEAU, S., WHITEHEAD, V., COLLIN, I., CUMMINGS, J. L. & CHERTKOW, H. 2005. The Montreal Cognitive Assessment, MoCA: a brief screening tool for mild cognitive impairment. *J Am Geriatr Soc*, 53, 695-9.
- NAU, R., SORGEL, F. & EIFFERT, H. 2010. Penetration of drugs through the blood-cerebrospinal fluid/blood-brain barrier for treatment of central nervous system infections. *Clin Microbiol Rev*, 23, 858-83.
- NETLAND, J., MEYERHOLZ, D. K., MOORE, S., CASSELL, M. & PERLMAN, S. 2008. Severe acute respiratory syndrome coronavirus infection causes neuronal death in the absence of encephalitis in mice transgenic for human ACE2. *J Virol*, 82, 7264-75.
- NGUYEN, L. & PIETERS, J. 2005. The Trojan horse: survival tactics of pathogenic mycobacteria in macrophages. *Trends Cell Biol*, 15, 269-76.
- NIGHTINGALE, S., DREYER, A. J., SAYLOR, D., GISSLEN, M., WINSTON, A. & JOSKA, J. A. 2021. Moving on From HAND: Why We Need New Criteria for Cognitive Impairment in Persons Living With Human Immunodeficiency Virus and a Proposed Way Forward. *Clin Infect Dis*, 73, 1113-1118.
- NITTI, V. 1971. Results and tolerance of prolonged rifampicin treatment in recent and chronic forms of pulmonary tuberculosis. *Respiration*, 28, Suppl:57-69.
- NOVI, G., ROSSI, T., PEDEMONTE, E., SAITTA, L., ROLLA, C., ROCCATAGLIATA, L., INGLESE, M. & FARININI, D. 2020. Acute disseminated encephalomyelitis after SARS-CoV-2 infection. *Neurol Neuroimmunol Neuroinflamm*, 7.
- NYAMAYARO, P., CHIBANDA, D., ROBBINS, R. N., HAKIM, J. & GOUSE, H. 2019. Assessment of neurocognitive deficits in people living with HIV in Sub Saharan Africa: A systematic review. *Clin Neuropsychol*, 33, 1-26.
- O'CONNOR, J. C., ANDRE, C., WANG, Y., LAWSON, M. A., SZEGEDI, S. S., LESTAGE, J., CASTANON, N., KELLEY, K. W. & DANTZER, R. 2009. Interferon-gamma and tumor necrosis factor-alpha mediate the upregulation of indoleamine 2,3-dioxygenase and the induction of depressive-like behavior in mice in response to bacillus Calmette-Guerin. *J Neurosci*, 29, 4200-9.
- O'GARRA, A., REDFORD, P. S., MCNAB, F. W., BLOOM, C. I., WILKINSON, R. J. & BERRY, M. P. 2013. The immune response in tuberculosis. *Annu Rev Immunol*, 31, 475-527.
- O'TOOLE, R. D., THORNTON, G. F., MUKHERJEE, M. K. & NATH, R. L. 1969. Dexamethasone in tuberculous meningitis. Relationship of cerebrospinal fluid effects to therapeutic efficacy. *Ann Intern Med*, 70, 39-48.

- ODDO, M., LEVINE, J. M., FRANGOS, S., CARRERA, E., MALONEY-WILENSKY, E., PASCUAL, J. L., KOFKE, W. A., MAYER, S. A. & LEROUX, P. D. 2009. Effect of mannitol and hypertonic saline on cerebral oxygenation in patients with severe traumatic brain injury and refractory intracranial hypertension. *J Neurol Neurosurg Psychiatry*, 80, 916-20.
- PACKARD, M., HANSEN, W. & BLOCH, F. Nuclear induction. *Physical Review*, 1946. AMERICAN PHYSICAL SOC ONE PHYSICS ELLIPSE, COLLEGE PK, MD 20740-3844 USA, 680-680.
- PAGE, C. P. & SPINA, D. 2011. Phosphodiesterase inhibitors in the treatment of inflammatory diseases. *Handb Exp Pharmacol*, 391-414.
- PARADKAR, M. S., DEVALEENAL, D. B., MVALO, T., ARENIVAS, A., THAKUR, K. T., WOLF, L., NIMKAR, S., INAMDAR, S., GIRIDHARAN, P., SELLADURAI, E., KINIKAR, A., VALVI, C., KHWAJA, S., GADAMA, D., BALAJI, S., KATTAGONI, K. Y., VENKATESAN, M., SAVIC, R., SWAMINATHAN, S., GUPTA, A., GUPTA, N., MAVE, V., DOOLEY, K. E. & TEAM, T.-K. S. 2022. Randomized Clinical Trial of High Dose Rifampicin with or without Levofloxacin versus Standard of Care for Paediatric Tuberculous Meningitis: The TBM-KIDS Trial. *Clin Infect Dis*.
- PARIHAR, S. P., GULER, R., KHUTLANG, R., LANG, D. M., HURDAYAL, R., MHLANGA, M. M., SUZUKI, H., MARAIS, A. D. & BROMBACHER, F. 2014. Statin therapy reduces the mycobacterium tuberculosis burden in human macrophages and in mice by enhancing autophagy and phagosome maturation. *J Infect Dis*, 209, 754-63.
- PARSONS, T., BANKS, S., BAE, C., GELBER, J., ALAHMADI, H. & TICHAUER, M. 2020. COVID-19-associated acute disseminated encephalomyelitis (ADEM). *J Neurol*.
- PARWATI, I., ALISJAHBANA, B., APRIANI, L., SOETIKNO, R. D., OTTENHOFF, T. H., VAN DER ZANDEN, A. G., VAN DER MEER, J., VAN SOOLINGEN, D. & VAN CREVEL, R. 2010. Mycobacterium tuberculosis Beijing genotype is an independent risk factor for tuberculosis treatment failure in Indonesia. *J Infect Dis*, 201, 553-7.
- PASIPANODYA, J. G. & GUMBO, T. 2010. Clinical and toxicodynamic evidence that high-dose pyrazinamide is not more hepatotoxic than the low doses currently used. *Antimicrob Agents Chemother*, 54, 2847-54.
- PATEL, V. B., PADAYATCHI, N., BHIGJEE, A. I., ALLEN, J., BHAGWAN, B., MOODLEY, A. & MTHIYANE, T. 2004. Multidrug-resistant tuberculous meningitis in KwaZulu-Natal, South Africa. *Clinical infectious diseases*, 38, 851-856.
- PECHKOVSKY, D. V., ZALUTSKAYA, O. M., IVANOV, G. I. & MISUNO, N. I. 2000. Calprotectin (MRP8/14 protein complex) release during mycobacterial infection in vitro and in vivo. *FEMS Immunol Med Microbiol*, 29, 27-33.
- PETERSON, P. K., GEKKER, G., HU, S., ANDERSON, W. B., TEICHERT, M., CHAO, C. C. & MOLITOR, T. W. 1996. Multinucleated giant cell formation of swine microglia induced by Mycobacterium bovis. *J Infect Dis*, 173, 1194-201.

- PETERSON, P. K., GEKKER, G., HU, S., SHENG, W. S., ANDERSON, W. R., ULEVITCH, R. J., TOBIAS, P. S., GUSTAFSON, K. V., MOLITOR, T. W. & CHAO, C. C. 1995. CD14 receptor-mediated uptake of nonopsonized *Mycobacterium tuberculosis* by human microglia. *Infect Immun*, 63, 1598-602.
- PETRONE, L., CANNAS, A., VANINI, V., CUZZI, G., ALOI, F., NSUBUGA, M., SSERUNKUMA, J., NAZZIWA, R., JUGHELI, L. & LUKINDO, T. 2016. Blood and urine inducible protein 10 as potential markers of disease activity. *The International Journal of Tuberculosis and Lung Disease*, 20, 1554-1561.
- PONCE, J., DE LA OSSA, N. P., HURTADO, O., MILLAN, M., ARENILLAS, J. F., DAVALOS, A. & GASULL, T. 2008. Simvastatin reduces the association of NMDA receptors to lipid rafts: a cholesterol-mediated effect in neuroprotection. *Stroke*, 39, 1269-75.
- PORTEVIN, D., GAGNEUX, S., COMAS, I. & YOUNG, D. 2011. Human macrophage responses to clinical isolates from the *Mycobacterium tuberculosis* complex discriminate between ancient and modern lineages. *PLoS Pathog*, 7, e1001307.
- POYIADJI, N., SHAHIN, G., NOUJAIM, D., STONE, M., PATEL, S. & GRIFFITH, B. 2020. COVID-19-associated Acute Hemorrhagic Necrotizing Encephalopathy: CT and MRI Features. *Radiology*, 201187.
- PRASAD, K., SINGH, M. B. & RYAN, H. 2016. Corticosteroids for managing tuberculous meningitis. *Cochrane Database Syst Rev*, 4, CD002244.
- PUELLES, V. G., LUTGEHETMANN, M., LINDENMEYER, M. T., SPERHAKE, J. P., WONG, M. N., ALLWEISS, L., CHILLA, S., HEINEMANN, A., WANNER, N., LIU, S., BRAUN, F., LU, S., PFEFFERLE, S., SCHRODER, A. S., EDLER, C., GROSS, O., GLATZEL, M., WICHMANN, D., WIECH, T., KLUGE, S., PUESCHEL, K., AEPFELBACHER, M. & HUBER, T. B. 2020. Multiorgan and Renal Tropism of SARS-CoV-2. *N Engl J Med*, 383, 590-592.
- PULZOVA, L., BHIDE, M. R. & ANDREJ, K. 2009. Pathogen translocation across the blood-brain barrier. *FEMS Immunol Med Microbiol*, 57, 203-13.
- PURCELL, E. M., TORREY, H. C. & POUND, R. V. 1946. Resonance absorption by nuclear magnetic moments in a solid. *Physical review*, 69, 37.
- PUTS, N. A. & EDDEN, R. A. 2012. In vivo magnetic resonance spectroscopy of GABA: a methodological review. *Prog Nucl Magn Reson Spectrosc*, 60, 29-41.
- RADLOFF, L. S. 1977. The CES-D Scale: A self-report depression scale for research in the general population. *Applied Psychological Measurement*, 1, 385-401.
- RAI, D., GARG, R. K., MAHDI, A. A., JAIN, A., VERMA, R., TRIPATHI, A. K., SINGH, M. K., MALHOTRA, H. S., SINGH, G. P. & AHMAD, M. K. 2014. Cerebrospinal fluid cytokines and matrix metalloproteinases in human immunodeficiency seropositive and seronegative patients of tuberculous meningitis. *Ann Indian Acad Neurol*, 17, 171-8.

- RAJSHEKHAR, V. 2015. Surgery for brain tuberculosis: a review. *Acta Neurochir (Wien)*, 157, 1665-78.
- RAKOTOSAMIMANANA, N., RAHARIMANGA, V., ANDRIAMANDIMBY, S. F., SOARES, J. L., DOHERTY, T. M., RATSITORAHINA, M., RAMAROKOTO, H., ZUMLA, A., HUGGETT, J., ROOK, G., RICHARD, V., GICQUEL, B., RASOLOFO-RAZANAMPARANY, V. & GROUP, V. V. S. 2010. Variation in gamma interferon responses to different infecting strains of *Mycobacterium tuberculosis* in acid-fast bacillus smear-positive patients and household contacts in Antananarivo, Madagascar. *Clin Vaccine Immunol*, 17, 1094-103.
- RAMILO, O., SAEZ-LLORENS, X., MERTSOLA, J., JAFARI, H., OLSEN, K. D., HANSEN, E. J., YOSHINAGA, M., OHKAWARA, S., NARIUCHI, H. & MCCRACKEN, G. H., JR. 1990. Tumor necrosis factor alpha/cachectin and interleukin 1 beta initiate meningeal inflammation. *J Exp Med*, 172, 497-507.
- RANDALL, P. J., HSU, N. J., LANG, D., COOPER, S., SEBESHO, B., ALLIE, N., KEETON, R., FRANCISCO, N. M., SALIE, S., LABUSCHAGNE, A., QUESNIAUX, V., RYFFEL, B., KELLAWAY, L. & JACOBS, M. 2014. Neurons are host cells for *Mycobacterium tuberculosis*. *Infect Immun*, 82, 1880-90.
- RANJAN, P., KALITA, J. & MISRA, U. K. 2003. Serial study of clinical and CT changes in tuberculous meningitis. *Neuroradiology*, 45, 277-82.
- RANKIN, J. 1957. Cerebral vascular accidents in patients over the age of 60. II. Prognosis. *Scott Med J*, 2, 200-15.
- RATINAM, J., MISHRA, A. K., MUTHURAM, A. J., MIRACLIN, A., CHANDY, G. M., VANJARE, H. A., TURAKA, V. P., JUDE, J., HANSDAK, S. G. & IYADURAI, R. 2020. Role of cerebrospinal fluid C-reactive protein in tuberculous meningitis. *Int J Mycobacteriol*, 9, 422-428.
- REED, M. B., DOMENECH, P., MANCA, C., SU, H., BARCZAK, A. K., KREISWIRTH, B. N., KAPLAN, G. & BARRY, C. E., 3RD 2004. A glycolipid of hypervirulent tuberculosis strains that inhibits the innate immune response. *Nature*, 431, 84-7.
- RICCIOTTI, E. & FITZGERALD, G. A. 2011. Prostaglandins and inflammation. *Arterioscler Thromb Vasc Biol*, 31, 986-1000.
- RICH A, M. H. 1933. The Pathogenesis of Tuberculous Meningitis. *Bull John Hopkins Hosp*, 52, 5-37.
- RICH, A. R. 1933. The pathogenesis of tuberculous meningitis. *Bull John Hopkins Hosp*, 52, 5.
- RICHMAN, I. B. & OWENS, D. K. 2017. Aspirin for Primary Prevention. *Med Clin North Am*, 101, 713-724.
- RIEDEL, G., PLATT, B. & MICHEAU, J. 2003. Glutamate receptor function in learning and memory. *Behav Brain Res*, 140, 1-47.
- RITCHIE, K. & FUHRER, R. 1996. The validation of an informant screening test for irreversible cognitive decline in the elderly: performance characteristics within a general population sample. *International journal of geriatric psychiatry*, 11, 149-156.
- RIZVI, I., GARG, R. K., JAIN, A., MALHOTRA, H. S., SINGH, A. K., PRAKASH, S., KUMAR, N., GARG, R., VERMA, R., MAHDI, A. A. & SHARMA, P. K.

2016. Vitamin D status, vitamin D receptor and toll like receptor-2 polymorphisms in tuberculous meningitis: a case-control study. *Infection*, 44, 633-40.
- RIZVI, I., GARG, R. K., MALHOTRA, H. S., KUMAR, N., SHARMA, E., SRIVASTAVA, C. & UNIYAL, R. 2017. Ventriculo-peritoneal shunt surgery for tuberculous meningitis: A systematic review. *J Neurol Sci*, 375, 255-263.
- ROBBINS, R. N., JOSKA, J. A., THOMAS, K. G., STEIN, D. J., LINDA, T., MELLINS, C. A. & REMIEN, R. H. 2013. Exploring the utility of the Montreal Cognitive Assessment to detect HIV-associated neurocognitive disorder: the challenge and need for culturally valid screening tests in South Africa. *Clin Neuropsychol*, 27, 437-54.
- ROBERTS, M. T., MENDELSON, M., MEYER, P., CARMICHAEL, A. & LEVER, A. M. 2003. The use of thalidomide in the treatment of intracranial tuberculomas in adults: two case reports. *J Infect*, 47, 251-5.
- ROBINSON-PAPP, J., GONZALEZ-DUARTE, A., SIMPSON, D. M., RIVERA-MINDT, M., MORGELLO, S. & MANHATTAN, H. I. V. B. B. 2009. The roles of ethnicity and antiretrovirals in HIV-associated polyneuropathy: a pilot study. *J Acquir Immune Defic Syndr*, 51, 569-73.
- ROCK, R. B., HU, S., GEKKER, G., SHENG, W. S., MAY, B., KAPUR, V. & PETERSON, P. K. 2005. Mycobacterium tuberculosis-induced cytokine and chemokine expression by human microglia and astrocytes: effects of dexamethasone. *J Infect Dis*, 192, 2054-8.
- ROCK, R. B., OLIN, M., BAKER, C. A., MOLITOR, T. W. & PETERSON, P. K. 2008. Central nervous system tuberculosis: pathogenesis and clinical aspects. *Clin Microbiol Rev*, 21, 243-61, table of contents.
- ROHLWINK, U. K., FIGAJI, A., WILKINSON, K. A., HORSWELL, S., SESAY, A. K., DEFFUR, A., ENSLIN, N., SOLOMONS, R., VAN TOORN, R., ELEY, B., LEVIN, M., WILKINSON, R. J. & LAI, R. P. J. 2019. Tuberculous meningitis in children is characterized by compartmentalized immune responses and neural excitotoxicity. *Nat Commun*, 10, 3767.
- ROHLWINK, U. K., KILBORN, T., WIESELTHALER, N., BANDERKER, E., ZWANE, E. & FIGAJI, A. A. 2016. Imaging Features of the Brain, Cerebral Vessels and Spine in Pediatric Tuberculous Meningitis With Associated Hydrocephalus. *Pediatr Infect Dis J*, 35, e301-10.
- ROHLWINK, U. K., MAUFF, K., WILKINSON, K. A., ENSLIN, N., WEGOYE, E., WILKINSON, R. J. & FIGAJI, A. A. 2017. Biomarkers of Cerebral Injury and Inflammation in Pediatric Tuberculous Meningitis. *Clin Infect Dis*, 65, 1298-1307.
- ROSSI, A. 2008. Imaging of acute disseminated encephalomyelitis. *Neuroimaging Clin N Am*, 18, 149-61; ix.
- RUPSINGH, R., BORRIE, M., SMITH, M., WELLS, J. & BARTHA, R. 2011. Reduced hippocampal glutamate in Alzheimer disease. *Neurobiology of aging*, 32, 802-810.
- RUSLAMI, R., GANIEM, A. R., AARNOUTSE, R. E., VAN CREVEL, R. & STUDY, T. 2013a. Rifampicin and moxifloxacin for tuberculous meningitis--authors' reply. *Lancet Infect Dis*, 13, 570.

- RUSLAMI, R., GANIEM, A. R., DIAN, S., APRIANI, L., ACHMAD, T. H., VAN DER VEN, A. J., BORM, G., AARNOUTSE, R. E. & VAN CREVEL, R. 2013b. Intensified regimen containing rifampicin and moxifloxacin for tuberculous meningitis: an open-label, randomised controlled phase 2 trial. *Lancet Infect Dis*, 13, 27-35.
- RYCKMAN, C., VANDAL, K., ROULEAU, P., TALBOT, M. & TESSIER, P. A. 2003. Proinflammatory activities of S100: proteins S100A8, S100A9, and S100A8/A9 induce neutrophil chemotaxis and adhesion. *J Immunol*, 170, 3233-42.
- SAGHAZADEH, A. & REZAEI, N. 2022. Central Inflammatory Cytokines in Tuberculous Meningitis: A Systematic Review and Meta-analysis. *J Interferon Cytokine Res*, 42, 95-107.
- SAINI, A. G., DOGRA, S., KUMAR, R., NADA, R. & SINGH, M. 2011. Primary tuberculous cerebellar abscess: case report. *Ann Trop Paediatr*, 31, 367-9.
- SANCHEZ-AGUILAR, M., TAPIA-PEREZ, J. H., SANCHEZ-RODRIGUEZ, J. J., VINAS-RIOS, J. M., MARTINEZ-PEREZ, P., DE LA CRUZ-MENDOZA, E., SANCHEZ-REYNA, M., TORRES-CORZO, J. G. & GORDILLO-MOSCOSO, A. 2013. Effect of rosuvastatin on cytokines after traumatic head injury. *J Neurosurg*, 118, 669-75.
- SARKAR, R., LENDERS, L., WILKINSON, K. A., WILKINSON, R. J. & NICOL, M. P. 2012. Modern lineages of Mycobacterium tuberculosis exhibit lineage-specific patterns of growth and cytokine induction in human monocyte-derived macrophages. *PLoS One*, 7, e43170.
- SARMA, D. & BILELLO, L. A. 2020. A Case Report of Acute Transverse Myelitis Following Novel Coronavirus Infection. *Clin Pract Cases Emerg Med*, 4, 321-323.
- SAUNDERS, B. M., FRANK, A. A., ORME, I. M. & COOPER, A. M. 2000. Interleukin-6 induces early gamma interferon production in the infected lung but is not required for generation of specific immunity to Mycobacterium tuberculosis infection. *Infect Immun*, 68, 3322-6.
- SAVER, J. L., FILIP, B., HAMILTON, S., YANES, A., CRAIG, S., CHO, M., CONWIT, R., STARKMAN, S., INVESTIGATORS, F.-M. & COORDINATORS 2010. Improving the reliability of stroke disability grading in clinical trials and clinical practice: the Rankin Focused Assessment (RFA). *Stroke*, 41, 992-5.
- SCHAAL, S. M., GARG, M. S., GHOSH, M., LOVERA, L., LOPEZ, M., PATEL, M., LOURO, J., PATEL, S., TUESTA, L., CHAN, W. M. & PEARSE, D. D. 2012. The therapeutic profile of rolipram, PDE target and mechanism of action as a neuroprotectant following spinal cord injury. *PLoS One*, 7, e43634.
- SCHECTER, G. F., SCOTT, C., TRUE, L., RAFTERY, A., FLOOD, J. & MASE, S. 2010. Linezolid in the treatment of multidrug-resistant tuberculosis. *Clin Infect Dis*, 50, 49-55.
- SCHOEMAN, J. F., FIEGGEN, G., SELLER, N., MENDELSON, M. & HARTZENBERG, B. 2006. Intractable intracranial tuberculous infection responsive to thalidomide: report of four cases. *J Child Neurol*, 21, 301-8.



- SCHOEMAN, J. F., JANSE VAN RENSBURG, A., LAUBSCHER, J. A. & SPRINGER, P. 2011. The role of aspirin in childhood tuberculous meningitis. *J Child Neurol*, 26, 956-62.
- SCHOEMAN, J. F., MORKEL, A., SEIFART, H. I., PARKIN, D. P., VAN HELDEN, P. D., HEWLETT, R. H. & DONALD, P. R. 1998. Massive posterior fossa tuberculous abscess developing in a young child treated for miliary tuberculosis. Possible role of very rapid acetylation of isoniazid. *Pediatr Neurosurg*, 29, 64-8.
- SCHOEMAN, J. F., SPRINGER, P., RAVENSCROFT, A., DONALD, P. R., BEKKER, L. G., VAN RENSBURG, A. J., HANEKOM, W. A., HASLETT, P. A. & KAPLAN, G. 2000. Adjunctive thalidomide therapy of childhood tuberculous meningitis: possible anti-inflammatory role. *J Child Neurol*, 15, 497-503.
- SCHOEMAN, J. F., SPRINGER, P., VAN RENSBURG, A. J., SWANEVELDER, S., HANEKOM, W. A., HASLETT, P. A. & KAPLAN, G. 2004. Adjunctive thalidomide therapy for childhood tuberculous meningitis: results of a randomized study. *J Child Neurol*, 19, 250-7.
- SCHOEMAN, J. F., VAN ZYL, L. E., LAUBSCHER, J. A. & DONALD, P. R. 1997. Effect of corticosteroids on intracranial pressure, computed tomographic findings, and clinical outcome in young children with tuberculous meningitis. *Pediatrics*, 99, 226-31.
- SCHULZ, J. G., BOSEL, J., STOECKEL, M., MEGOW, D., DIRNAGL, U. & ENDRES, M. 2004. HMG-CoA reductase inhibition causes neurite loss by interfering with geranylgeranylpyrophosphate synthesis. *J Neurochem*, 89, 24-32.
- SEDAGHAT, Z. & KARIMI, N. 2020. Guillain Barre syndrome associated with COVID-19 infection: A case report. *J Clin Neurosci*, 76, 233-235.
- SERVICES, D. O. A. N. I. O. A. A. I. D. N. I. O. H. U. D. O. H. A. H. Corrected Version 2.1 July 2017. Division of AIDS (DAIDS) Table for Grading the Severity of Adult and Pediatric Adverse Events.
- SHAMS, H., WIZEL, B., LAKEY, D. L., SAMTEN, B., VANKAYALAPATI, R., VALDIVIA, R. H., KITCHENS, R. L., GRIFFITH, D. E. & BARNES, P. F. 2003. The CD14 receptor does not mediate entry of Mycobacterium tuberculosis into human mononuclear phagocytes. *FEMS Immunol Med Microbiol*, 36, 63-9.
- SHARMA, R. M., PRUTHI, N., ARIMAPPAMAGAN, A., SOMANNA, S., DEVI, B. I. & PANDEY, P. 2015. Tubercular meningitis with hydrocephalus with HIV co-infection: role of cerebrospinal fluid diversion procedures. *J Neurosurg*, 122, 1087-95.
- SHINOYAMA, M., SUZUKI, M. & NOMURA, S. 2012. Fulminant tuberculous meningitis--autopsy case report. *Neurol Med Chir (Tokyo)*, 52, 761-4.
- SHUKLA, R., ABBAS, A., KUMAR, P., GUPTA, R. K., JHA, S. & PRASAD, K. N. 2008. Evaluation of cerebral infarction in tuberculous meningitis by diffusion weighted imaging. *J Infect*, 57, 298-306.
- SHUTTER, L., TONG, K. A. & HOLSHOUSER, B. A. 2004. Proton MRS in acute traumatic brain injury: role for glutamate/glutamine and choline for outcome prediction. *J Neurotrauma*, 21, 1693-705.

- SIMMONS, C. P., THWAITES, G. E., QUYEN, N. T., TOROK, E., HOANG, D. M., CHAU, T. T., MAI, P. P., LAN, N. T., DUNG, N. H., QUY, H. T., BANG, N. D., HIEN, T. T. & FARRAR, J. 2006. Pretreatment intracerebral and peripheral blood immune responses in Vietnamese adults with tuberculous meningitis: diagnostic value and relationship to disease severity and outcome. *J Immunol*, 176, 2007-14.
- SINGLA, R., CAMINERO, J. A., JAISWAL, A., SINGLA, N., GUPTA, S., BALI, R. K. & BEHERA, D. 2012. Linezolid: an effective, safe and cheap drug for patients failing multidrug-resistant tuberculosis treatment in India. *Eur Respir J*, 39, 956-62.
- SKENDROS, P., KAMARIA, F., KONTOPOULOS, V., TSITOURIDIS, I. & SIDIROPOULOS, L. 2003. Intradural, eextramedullary tuberculoma of the spinal cord as a complication of tuberculous meningitis. *Infection*, 31, 115-7.
- SKERRY, C., PINN, M. L., BRUINERS, N., PINE, R., GENNARO, M. L. & KARAKOUSIS, P. C. 2014. Simvastatin increases the in vivo activity of the first-line tuberculosis regimen. *J Antimicrob Chemother*, 69, 2453-7.
- SONG, T., LEE, M., JEON, H. S., PARK, Y., DODD, L. E., DARTOIS, V., FOLLMAN, D., WANG, J., CAI, Y., GOLDFEDER, L. C., OLIVIER, K. N., XIE, Y., VIA, L. E., CHO, S. N., BARRY, C. E., 3RD & CHEN, R. Y. 2015. Linezolid Trough Concentrations Correlate with Mitochondrial Toxicity-Related Adverse Events in the Treatment of Chronic Extensively Drug-Resistant Tuberculosis. *EBioMedicine*, 2, 1627-33.
- SONGKHLA, M. N., TANTIPONG, H., TONGSAI, S. & ANGKASEKWINAI, N. Lateral flow urine lipoarabinomannan assay for diagnosis of active tuberculosis in adults with human immunodeficiency virus infection: a prospective cohort study. *Open Forum Infectious Diseases*, 2019. Oxford University Press US, ofz132.
- SOTGIU, G., CENTIS, R., D'AMBROSIO, L., ALFFENAAR, J. W., ANGER, H. A., CAMINERO, J. A., CASTIGLIA, P., DE LORENZO, S., FERRARA, G., KOH, W. J., SCHECTER, G. F., SHIM, T. S., SINGLA, R., SKRAHINA, A., SPANEVELLO, A., UDWADIA, Z. F., VILLAR, M., ZAMPOGNA, E., ZELLWEGER, J. P., ZUMLA, A. & MIGLIORI, G. B. 2012. Efficacy, safety and tolerability of linezolid containing regimens in treating MDR-TB and XDR-TB: systematic review and meta-analysis. *Eur Respir J*, 40, 1430-42.
- SOUTH AFRICA DEPARTMENT OF HEALTH 2014. *National Tuberculosis Management Guidelines*.
- SPITE, M. & SERHAN, C. N. 2010. Novel lipid mediators promote resolution of acute inflammation: impact of aspirin and statins. *Circ Res*, 107, 1170-84.
- SRIVASTAVA, T. & KOCHAR, D. K. 2003. Asymptomatic spinal arachnoiditis in patients with tuberculous meningitis. *Neuroradiology*, 45, 727-9.
- STEINGART, K. R., JOTBLAD, S., ROBSKY, K., DECK, D., HOPEWELL, P. C., HUANG, D. & NAHID, P. 2011. Higher-dose rifampin for the treatment of pulmonary tuberculosis: a systematic review. *Int J Tuberc Lung Dis*, 15, 305-16.

- STEMKENS, R., LITJENS, C. H. C., DIAN, S., GANIEM, A. R., YUNIVITA, V., VAN CREVEL, R., TE BRAKE, L. H. M., RUSLAMI, R. & AARNOUTSE, R. E. 2019. Pharmacokinetics of pyrazinamide during the initial phase of tuberculous meningitis treatment. *Int J Antimicrob Agents*, 54, 371-374.
- STERNS, R. H. & SILVER, S. M. 2008. Cerebral salt wasting versus SIADH: what difference? *J Am Soc Nephrol*, 19, 194-6.
- STREPTOMYCIN IN TUBERCULOSIS TRIALS COMMITTEE, MEDICAL RESEARCH COUNCIL 1948. STREPTOMYCIN treatment of tuberculous meningitis. *Lancet*, 1, 582-96.
- SUBBIAN, S., TSENOVA, L., O'BRIEN, P., YANG, G., KOO, M. S., PEIXOTO, B., FALLOWS, D., ZELDIS, J. B., MULLER, G. & KAPLAN, G. 2011. Phosphodiesterase-4 inhibition combined with isoniazid treatment of rabbits with pulmonary tuberculosis reduces macrophage activation and lung pathology. *Am J Pathol*, 179, 289-301.
- SUN, F., RUAN, Q., WANG, J., CHEN, S., JIN, J., SHAO, L., ZHANG, Y. & ZHANG, W. 2014. Linezolid manifests a rapid and dramatic therapeutic effect for patients with life-threatening tuberculous meningitis. *Antimicrob Agents Chemother*, 58, 6297-301.
- SYKES, W., MHLANGA, L., SWANEVELDER, R., GLATT, T. N., GREBE, E., COLEMAN, C., PIETERSON, N., CABLE, R., WELTE, A., VAN DEN BERG, K. & VERMEULEN, M. 2021. Prevalence of anti-SARS-CoV-2 antibodies among blood donors in Northern Cape, KwaZulu-Natal, Eastern Cape, and Free State provinces of South Africa in January 2021. *Res Sq*.
- TAN, H. Y., YONG, Y. K., ANDRADE, B. B., SHANKAR, E. M., PONNAMPALAVANAR, S., OMAR, S. F., NARENDRAN, G., KAMARULZAMAN, A., SWAMINATHAN, S., SERETI, I., CROWE, S. M. & FRENCH, M. A. 2015. Plasma interleukin-18 levels are a biomarker of innate immune responses that predict and characterize tuberculosis-associated immune reconstitution inflammatory syndrome. *AIDS*, 29, 421-31.
- TAN, H. Y., YONG, Y. K., SHANKAR, E. M., PAUKOVICS, G., ELLEGARD, R., LARSSON, M., KAMARULZAMAN, A., FRENCH, M. A. & CROWE, S. M. 2016. Aberrant Inflammasome Activation Characterizes Tuberculosis-Associated Immune Reconstitution Inflammatory Syndrome. *J Immunol*, 196, 4052-63.
- TANAKA, T., TATSUNO, I., UCHIDA, D., MOROO, I., MORIO, H., NAKAMURA, S., NOGUCHI, Y., YASUDA, T., KITAGAWA, M., SAITO, Y. & HIRAI, A. 2000. Geranylgeranyl-pyrophosphate, an isoprenoid of mevalonate cascade, is a critical compound for rat primary cultured cortical neurons to protect the cell death induced by 3-hydroxy-3-methylglutaryl-CoA reductase inhibition. *J Neurosci*, 20, 2852-9.
- TANG, S., YAO, L., HAO, X., ZHANG, X., LIU, G., LIU, X., WU, M., ZEN, L., SUN, H., LIU, Y., GU, J., LIN, F., WANG, X. & ZHANG, Z. 2015. Efficacy, safety and tolerability of linezolid for the treatment of XDR-TB: a study in China. *Eur Respir J*, 45, 161-70.
- THO, D. Q., TÖRÖK, M. E., YEN, N. T. B., BANG, N. D., LAN, N. T. N., KIET, V. S., VAN VINH CHAU, N., DUNG, N. H., DAY, J. & FARRAR, J. 2012.

- Influence of antituberculosis drug resistance and Mycobacterium tuberculosis lineage on outcome in HIV-associated tuberculous meningitis. *Antimicrobial Agents and Chemotherapy*, 56, 3074-3079.
- THOMAS, M. D., CHOPRA, J. S. & WALIA, B. N. 1977. Tuberculous meningitis (T.B.M.)(a clinical study of 232 cases). *J Assoc Physicians India*, 25, 633-9.
- THU, S. S., MATIN, N. & LEVINE, S. R. 2020. Olfactory gyrus intracerebral hemorrhage in a patient with COVID-19 infection. *J Clin Neurosci*, 79, 275-276.
- THUONG, N. T., HAWN, T. R., THWAITES, G. E., CHAU, T. T., LAN, N. T., QUY, H. T., HIEU, N. T., ADEREM, A., HIEN, T. T., FARRAR, J. J. & DUNSTAN, S. J. 2007. A polymorphism in human TLR2 is associated with increased susceptibility to tuberculous meningitis. *Genes Immun*, 8, 422-8.
- THUONG, N. T. T., HEEMSKERK, D., TRAM, T. T. B., THAO, L. T. P., RAMAKRISHNAN, L., HA, V. T. N., BANG, N. D., CHAU, T. T. H., LAN, N. H., CAWS, M., DUNSTAN, S. J., CHAU, N. V. V., WOLBERS, M., MAI, N. T. H. & THWAITES, G. E. 2017. Leukotriene A4 Hydrolase Genotype and HIV Infection Influence Intracerebral Inflammation and Survival From Tuberculous Meningitis. *J Infect Dis*, 215, 1020-1028.
- THWAITES, G. E., BHAVNANI, S. M., CHAU, T. T., HAMMEL, J. P., TOROK, M. E., VAN WART, S. A., MAI, P. P., REYNOLDS, D. K., CAWS, M., DUNG, N. T., HIEN, T. T., KULAWY, R., FARRAR, J. & AMBROSE, P. G. 2011. Randomized pharmacokinetic and pharmacodynamic comparison of fluoroquinolones for tuberculous meningitis. *Antimicrob Agents Chemother*, 55, 3244-53.
- THWAITES, G. E., DUC BANG, N., HUY DUNG, N., THI QUY, H., THI TUONG OANH, D., THI CAM THOA, N., QUANG HIEN, N., TRI THUC, N., NGOC HAI, N., THI NGOC LAN, N., NGOC LAN, N., HONG DUC, N., NGOC TUAN, V., HUU HIEP, C., THI HONG CHAU, T., PHUONG MAI, P., THI DUNG, N., STEPNIIEWSKA, K., SIMMONS, C. P., WHITE, N. J., TINH HIEN, T. & FARRAR, J. J. 2005. The influence of HIV infection on clinical presentation, response to treatment, and outcome in adults with Tuberculous meningitis. *J Infect Dis*, 192, 2134-41.
- THWAITES, G. E., MACMULLEN-PRICE, J., TRAN, T. H., PHAM, P. M., NGUYEN, T. D., SIMMONS, C. P., WHITE, N. J., TRAN, T. H., SUMMERS, D. & FARRAR, J. J. 2007. Serial MRI to determine the effect of dexamethasone on the cerebral pathology of tuberculous meningitis: an observational study. *Lancet Neurol*, 6, 230-6.
- THWAITES, G. E., NGUYEN, D. B., NGUYEN, H. D., HOANG, T. Q., DO, T. T., NGUYEN, T. C., NGUYEN, Q. H., NGUYEN, T. T., NGUYEN, N. H., NGUYEN, T. N., NGUYEN, N. L., NGUYEN, H. D., VU, N. T., CAO, H. H., TRAN, T. H., PHAM, P. M., NGUYEN, T. D., STEPNIIEWSKA, K., WHITE, N. J., TRAN, T. H. & FARRAR, J. J. 2004. Dexamethasone for the treatment of tuberculous meningitis in adolescents and adults. *N Engl J Med*, 351, 1741-51.
- THWAITES, G. E., SIMMONS, C. P., THAN HA QUYEN, N., THI HONG CHAU, T., PHUONG MAI, P., THI DUNG, N., HOAN PHU, N., WHITE, N. P.,

- TINH HIEN, T. & FARRAR, J. J. 2003. Pathophysiology and prognosis in vietnamese adults with tuberculous meningitis. *J Infect Dis*, 188, 1105-15.
- THWAITES, G. E. & TRAN, T. H. 2005. Tuberculous meningitis: many questions, too few answers. *Lancet Neurol*, 4, 160-70.
- TOBIN, D. M., ROCA, F. J., OH, S. F., MCFARLAND, R., VICKERY, T. W., RAY, J. P., KO, D. C., ZOU, Y., BANG, N. D., CHAU, T. T., VARY, J. C., HAWN, T. R., DUNSTAN, S. J., FARRAR, J. J., THWAITES, G. E., KING, M. C., SERHAN, C. N. & RAMAKRISHNAN, L. 2012. Host genotype-specific therapies can optimize the inflammatory response to mycobacterial infections. *Cell*, 148, 434-46.
- TOBIN, D. M., VARY, J. C., JR., RAY, J. P., WALSH, G. S., DUNSTAN, S. J., BANG, N. D., HAGGE, D. A., KHADGE, S., KING, M. C., HAWN, T. R., MOENS, C. B. & RAMAKRISHNAN, L. 2010. The *Ita4h* locus modulates susceptibility to mycobacterial infection in zebrafish and humans. *Cell*, 140, 717-30.
- TOGNARELLI, J. M., DAWOOD, M., SHARIFF, M. I., GROVER, V. P., CROSSEY, M. M., COX, I. J., TAYLOR-ROBINSON, S. D. & MCPHAIL, M. J. 2015. Magnetic Resonance Spectroscopy: Principles and Techniques: Lessons for Clinicians. *J Clin Exp Hepatol*, 5, 320-8.
- TOROK, M. E., ALJAYYOSSI, G., WATERHOUSE, D., CHAU, T., MAI, N., PHU, N. H., HIEN, T. T., HOPE, W., FARRAR, J. J. & WARD, S. A. 2018. Suboptimal Exposure to Anti-TB Drugs in a TBM/HIV+ Population Is Not Related to Antiretroviral Therapy. *Clin Pharmacol Ther*, 103, 449-457.
- TOROK, M. E., CHAU, T. T., MAI, P. P., PHONG, N. D., DUNG, N. T., CHUONG, L. V., LEE, S. J., CAWS, M., DE JONG, M. D., HIEN, T. T. & FARRAR, J. J. 2008. Clinical and microbiological features of HIV-associated tuberculous meningitis in Vietnamese adults. *PLoS One*, 3, e1772.
- TOROK, M. E., YEN, N. T., CHAU, T. T., MAI, N. T., PHU, N. H., MAI, P. P., DUNG, N. T., CHAU, N. V., BANG, N. D., TIEN, N. A., MINH, N. H., HIEN, N. Q., THAI, P. V., DONG, D. T., ANH, D. T., THOA, N. T., HAI, N. N., LAN, N. N., LAN, N. T., QUY, H. T., DUNG, N. H., HIEN, T. T., CHINH, N. T., SIMMONS, C. P., DE JONG, M., WOLBERS, M. & FARRAR, J. J. 2011a. Timing of initiation of antiretroviral therapy in human immunodeficiency virus (HIV)--associated tuberculous meningitis. *Clin Infect Dis*, 52, 1374-83.
- TOROK, M. E., YEN, N. T., CHAU, T. T., MAI, N. T., PHU, N. H., MAI, P. P., DUNG, N. T., CHAU, N. V., BANG, N. D., TIEN, N. A., MINH, N. H., HIEN, N. Q., THAI, P. V., DONG, D. T., ANH, D. T., THOA, N. T., HAI, N. N., LAN, N. N., LAN, N. T., QUY, H. T., DUNG, N. H., HIEN, T. T., CHINH, N. T., SIMMONS, C. P., DE JONG, M., WOLBERS, M. & FARRAR, J. J. 2011b. Timing of initiation of antiretroviral therapy in human immunodeficiency virus (HIV)-associated tuberculous meningitis. *Clin Infect Dis*, 52, 1374-83.
- TOSCANO, G., PALMERINI, F., RAVAGLIA, S., RUIZ, L., INVERNIZZI, P., CUZZONI, M. G., FRANCIOTTA, D., BALDANTI, F., DATURI, R.,

- POSTORINO, P., CAVALLINI, A. & MICIELI, G. 2020a. Guillain-Barre Syndrome Associated with SARS-CoV-2. *N Engl J Med*, 382, 2574-2576.
- TOSCANO, G., PALMERINI, F., RAVAGLIA, S., RUIZ, L., INVERNIZZI, P., CUZZONI, M. G., FRANCIOTTA, D., BALDANTI, F., DATURI, R., POSTORINO, P., CAVALLINI, A. & MICIELI, G. 2020b. Guillain-Barre Syndrome Associated with SARS-CoV-2. *N Engl J Med*.
- TOZZI, V., BALESTRA, P., LORENZINI, P., BELLAGAMBA, R., GALGANI, S., CORPOLONGO, A., VLASSI, C., LARUSSA, D., ZACCARELLI, M., NOTO, P., VISCO-COMANDINI, U., GIULIANELLI, M., IPPOLITO, G., ANTINORI, A. & NARCISO, P. 2005. Prevalence and risk factors for human immunodeficiency virus-associated neurocognitive impairment, 1996 to 2002: results from an urban observational cohort. *J Neurovirol*, 11, 265-73.
- TSENOVA, L., BERGTOLD, A., FREEDMAN, V. H., YOUNG, R. A. & KAPLAN, G. 1999. Tumor necrosis factor alpha is a determinant of pathogenesis and disease progression in mycobacterial infection in the central nervous system. *Proc Natl Acad Sci U S A*, 96, 5657-62.
- TSENOVA, L., ELLISON, E., HARBACHEUSKI, R., MOREIRA, A. L., KUREPINA, N., REED, M. B., MATHEMA, B., BARRY, C. E., 3RD & KAPLAN, G. 2005. Virulence of selected Mycobacterium tuberculosis clinical isolates in the rabbit model of meningitis is dependent on phenolic glycolipid produced by the bacilli. *J Infect Dis*, 192, 98-106.
- TSENOVA, L., MANGALISO, B., MULLER, G., CHEN, Y., FREEDMAN, V. H., STIRLING, D. & KAPLAN, G. 2002. Use of IMiD3, a thalidomide analog, as an adjunct to therapy for experimental tuberculous meningitis. *Antimicrob Agents Chemother*, 46, 1887-95.
- TSENOVA, L., SOKOL, K., FREEDMAN, V. H. & KAPLAN, G. 1998. A combination of thalidomide plus antibiotics protects rabbits from mycobacterial meningitis-associated death. *J Infect Dis*, 177, 1563-72.
- TSUKAMURA, M., NAKAMURA, E., YOSHII, S. & AMANO, H. 1985. Therapeutic effect of a new antibacterial substance ofloxacin (DL8280) on pulmonary tuberculosis. *Am Rev Respir Dis*, 131, 352-6.
- TUCKER, E. W., MARAIS, S., SEDDON, J. A., VAN CREVEL, R., GANIEM, A. R., RUSLAMI, R., ZHANG, W., SUN, F., ZHOU, X., SOLOMONS, R. S., CRESSWELL, F. V., WILMSHURST, J. & ROHLWINK, U. 2020. International Survey Reveals Opportunities to Improve Tuberculous Meningitis Management and the Need for Standardized Guidelines. *Open Forum Infect Dis*, 7, ofaa445.
- U.S. DEPARTMENT OF HEALTH AND HUMAN SERVICES, N. I. O. H., NATIONAL INSTITUTE OF ALLERGY AND INFECTIOUS DISEASES, DIVISION OF AIDS. DIVISION OF AIDS (DAIDS) TABLE FOR GRADING THE SEVERITY OF ADULT AND PEDIATRIC ADVERSE EVENTS, CORRECTED VERSION 2.1. [JULY 2017].
- UPTON, J. 2013. Beck Depression Inventory (BDI). In: GELLMAN, M. D. & TURNER, J. R. (eds.) *Encyclopedia of Behavioral Medicine*. New York, NY: Springer New York.
- UTHMAN, O. A., OKWUNDU, C., GBENGA, K., VOLMINK, J., DOWDY, D., ZUMLA, A. & NACHEGA, J. B. 2015. Optimal Timing of Antiretroviral

- Therapy Initiation for HIV-Infected Adults With Newly Diagnosed Pulmonary Tuberculosis: A Systematic Review and Meta-analysis. *Ann Intern Med*, 163, 32-9.
- VALIUDDIN, H., SKWIRSK, B. & PAZ-ARABO, P. 2020. Acute transverse myelitis associated with SARS-CoV-2: A Case-Report. *Brain Behav Immun Health*, 5, 100091.
- VAN CREVEL, R., NELWAN, R. H., DE LENNE, W., VEERARAGU, Y., VAN DER ZANDEN, A. G., AMIN, Z., VAN DER MEER, J. W. & VAN SOOLINGEN, D. 2001. Mycobacterium tuberculosis Beijing genotype strains associated with febrile response to treatment. *Emerg Infect Dis*, 7, 880-3.
- VAN DER FLIER, M., HOPPENREIJS, S., VAN RENSBURG, A. J., RUYKEN, M., KOLK, A. H., SPRINGER, P., HOEPELMAN, A. I., GEELLEN, S. P., KIMPEN, J. L. & SCHOEMAN, J. F. 2004. Vascular endothelial growth factor and blood-brain barrier disruption in tuberculous meningitis. *Pediatr Infect Dis J*, 23, 608-13.
- VAN DER LAAN, L. E., SCHAAF, H. S., SOLOMONS, R., WILLEMSE, M., MOHAMED, N., BABOOLAL, S. O., HESSELING, A. C., VAN TOORN, R. & GARCIA-PRATS, A. J. 2016. Probable Levofloxacin-associated Secondary Intracranial Hypertension in a Child With Multidrug-resistant Tuberculosis. *Pediatr Infect Dis J*, 35, 706-8.
- VAN INGEN, J., AARNOUTSE, R. E., DONALD, P. R., DIACON, A. H., DAWSON, R., PLEMPER VAN BALEN, G., GILLESPIE, S. H. & BOEREE, M. J. 2011. Why Do We Use 600 mg of Rifampicin in Tuberculosis Treatment? *Clin Infect Dis*, 52, e194-9.
- VAN LAARHOVEN, A., DIAN, S., AGUIRRE-GAMBOA, R., AVILA-PACHECO, J., RICANO-PONCE, I., RUESEN, C., ANNISA, J., KOEKEN, V., CHAIDIR, L., LI, Y., ACHMAD, T. H., JOOSTEN, L. A. B., NOTEBAART, R. A., RUSLAMI, R., NETEA, M. G., VERBEEK, M. M., ALISJAHBANA, B., KUMAR, V., CLISH, C. B., GANIEM, A. R. & VAN CREVEL, R. 2018. Cerebral tryptophan metabolism and outcome of tuberculous meningitis: an observational cohort study. *Lancet Infect Dis*, 18, 526-535.
- VAN LAARHOVEN, A., DIAN, S., RUESEN, C., HAYATI, E., DAMEN, M., ANNISA, J., CHAIDIR, L., RUSLAMI, R., ACHMAD, T. H., NETEA, M. G., ALISJAHBANA, B., RIZAL GANIEM, A. & VAN CREVEL, R. 2017. Clinical Parameters, Routine Inflammatory Markers, and LTA4H Genotype as Predictors of Mortality Among 608 Patients With Tuberculous Meningitis in Indonesia. *J Infect Dis*, 215, 1029-1039.
- VAN SWIETEN, J. C., KOUDSTAAL, P. J., VISSER, M. C., SCHOUTEN, H. J. & VAN GIJN, J. 1988. Interobserver agreement for the assessment of handicap in stroke patients. *Stroke*, 19, 604-7.
- VAN TOORN, R., DU PLESSIS, A. M., SCHAAF, H. S., BUYS, H., HEWLETT, R. H. & SCHOEMAN, J. F. 2015. Clinikoradiologic response of neurologic tuberculous mass lesions in children treated with thalidomide. *Pediatr Infect Dis J*, 34, 214-8.
- VAN TOORN, R., SCHAAF, H. S., LAUBSCHER, J. A., VAN ELSLAND, S. L., DONALD, P. R. & SCHOEMAN, J. F. 2014. Short intensified treatment in

- children with drug-susceptible tuberculous meningitis. *Pediatr Infect Dis J*, 33, 248-52.
- VAN TOORN, R., SOLOMONS, R. S., SEDDON, J. A. & SCHOEMAN, J. F. 2021. Thalidomide Use for Complicated Central Nervous System Tuberculosis in Children: Insights From an Observational Cohort. *Clin Infect Dis*, 72, e136-e145.
- VAN WELL, G. T., PAES, B. F., TERWEE, C. B., SPRINGER, P., ROORD, J. J., DONALD, P. R., VAN FURTH, A. M. & SCHOEMAN, J. F. 2009. Twenty years of pediatric tuberculous meningitis: a retrospective cohort study in the western cape of South Africa. *Pediatrics*, 123, e1-8.
- VAN ZYL, C. W., LOOTS, D. T., SOLOMONS, R., VAN REENEN, M. & MASON, S. 2020. Metabolic characterization of tuberculous meningitis in a South African paediatric population using (1)H NMR metabolomics. *J Infect*, 81, 743-752.
- VANE, J. R. 1971. Inhibition of prostaglandin synthesis as a mechanism of action for aspirin-like drugs. *Nat New Biol*, 231, 232-5.
- VINNARD, C., WINSTON, C. A., WILEYTO, E. P., MACGREGOR, R. R. & BISSON, G. P. 2011. Multidrug resistant tuberculous meningitis in the United States, 1993–2005. *Journal of Infection*, 63, 240-242.
- VISSER, D. H., SOLOMONS, R. S., RONACHER, K., VAN WELL, G. T., HEYMANS, M. W., WALZL, G., CHEGOU, N. N., SCHOEMAN, J. F. & VAN FURTH, A. M. 2015. Host immune response to tuberculous meningitis. *Clin Infect Dis*, 60, 177-87.
- VON DER LIPPE, B., SANDVEN, P. & BRUBAKK, O. 2006. Efficacy and safety of linezolid in multidrug resistant tuberculosis (MDR-TB)--a report of ten cases. *J Infect*, 52, 92-6.
- WAICZIES, S., BENDIX, I. & ZIPP, F. 2008. Geranylgeranylation but not GTP-loading of Rho GTPases determines T cell function. *Sci Signal*, 1, pt3.
- WANG, R. & REDDY, P. H. 2017. Role of Glutamate and NMDA Receptors in Alzheimer's Disease. *J Alzheimers Dis*, 57, 1041-1048.
- WASSERMAN, S., BRUST, J. C. M., ABDELWAHAB, M. T., LITTLE, F., DENTI, P., WIESNER, L., GANDHI, N. R., MEINTJES, G. & MAARTENS, G. 2022. Linezolid toxicity in patients with drug-resistant tuberculosis: a prospective cohort study. *J Antimicrob Chemother*, 77, 1146-1154.
- WEBER, W. W. & HEIN, D. W. 1979. Clinical pharmacokinetics of isoniazid. *Clin Pharmacokinet*, 4, 401-22.
- WEINER, J., 3RD, PARIDA, S. K., MAERTZDORF, J., BLACK, G. F., REPSILBER, D., TELAAR, A., MOHNEY, R. P., ARNDT-SULLIVAN, C., GANOZA, C. A., FAE, K. C., WALZL, G. & KAUFMANN, S. H. 2012. Biomarkers of inflammation, immunosuppression and stress with active disease are revealed by metabolomic profiling of tuberculosis patients. *PLoS One*, 7, e40221.
- WEINER, M., BURMAN, W., VERNON, A., BENATOR, D., PELOQUIN, C. A., KHAN, A., WEIS, S., KING, B., SHAH, N., HODGE, T. & TUBERCULOSIS TRIALS, C. 2003. Low isoniazid concentrations and outcome of tuberculosis treatment with once-weekly isoniazid and rifapentine. *Am J Respir Crit Care Med*, 167, 1341-7.



- WEITZ-SCHMIDT, G., WELZENBACH, K., BRINKMANN, V., KAMATA, T., KALLEN, J., BRUNS, C., COTTENS, S., TAKADA, Y. & HOMMEL, U. 2001. Statins selectively inhibit leukocyte function antigen-1 by binding to a novel regulatory integrin site. *Nat Med*, 7, 687-92.
- WICHMANN, D., SPERHAKE, J. P., LUTGEHETMANN, M., STEURER, S., EDLER, C., HEINEMANN, A., HEINRICH, F., MUSHUMBA, H., KNIEP, I., SCHRODER, A. S., BURDELSKI, C., DE HEER, G., NIERHAUS, A., FRINGS, D., PFEFFERLE, S., BECKER, H., BREDEREKE-WIEDLING, H., DE WEERTH, A., PASCHEN, H. R., SHEIKHZADEH-EGGERS, S., STANG, A., SCHMIEDEL, S., BOKEMEYER, C., ADDO, M. M., AEPFELBACHER, M., PUSCHEL, K. & KLUGE, S. 2020. Autopsy Findings and Venous Thromboembolism in Patients With COVID-19: A Prospective Cohort Study. *Ann Intern Med*, 173, 268-277.
- WILKINSON, R. J., ROHLWINK, U., MISRA, U. K., VAN CREVEL, R., MAI, N. T. H., DOOLEY, K. E., CAWS, M., FIGAJI, A., SAVIC, R., SOLOMONS, R., THWAITES, G. E. & TUBERCULOUS MENINGITIS INTERNATIONAL RESEARCH, C. 2017. Tuberculous meningitis. *Nat Rev Neurol*, 13, 581-598.
- WOBKE, T. K., SORG, B. L. & STEINHILBER, D. 2014. Vitamin D in inflammatory diseases. *Front Physiol*, 5, 244.
- WOLFEL, R., CORMAN, V. M., GUGGEMOS, W., SEILMAIER, M., ZANGE, S., MULLER, M. A., NIEMEYER, D., JONES, T. C., VOLLMAR, P., ROTHE, C., HOELSCHER, M., BLEICKER, T., BRUNINK, S., SCHNEIDER, J., EHMANN, R., ZWIRGLMAIER, K., DROSTEN, C. & WENDTNER, C. 2020. Virological assessment of hospitalized patients with COVID-2019. *Nature*, 581, 465-469.
- WOODS, S. P., RIPPETH, J. D., FROL, A. B., LEVY, J. K., RYAN, E., SOUKUP, V. M., HINKIN, C. H., LAZZARETTO, D., CHERNER, M., MARCOTTE, T. D., GELMAN, B. B., MORGELLO, S., SINGER, E. J., GRANT, I. & HEATON, R. K. 2004. Interrater reliability of clinical ratings and neurocognitive diagnoses in HIV. *J Clin Exp Neuropsychol*, 26, 759-78.
- WRIGHT, S. D., RAMOS, R. A., TOBIAS, P. S., ULEVITCH, R. J. & MATHISON, J. C. 1990. CD14, a receptor for complexes of lipopolysaccharide (LPS) and LPS binding protein. *Science*, 249, 1431-3.
- WU, Q., QI, L., LI, H., MAO, L., YANG, M., XIE, R., YANG, X., WANG, J., ZHANG, Z., KONG, J. & SUN, B. 2017. Roflumilast Reduces Cerebral Inflammation in a Rat Model of Experimental Subarachnoid Hemorrhage. *Inflammation*, 40, 1245-1253.
- XING, Z., GAULDIE, J., COX, G., BAUMANN, H., JORDANA, M., LEI, X. F. & ACHONG, M. K. 1998. IL-6 is an antiinflammatory cytokine required for controlling local or systemic acute inflammatory responses. *J Clin Invest*, 101, 311-20.
- XU, J., ZHONG, S., LIU, J., LI, L., LI, Y., WU, X., LI, Z., DENG, P., ZHANG, J., ZHONG, N., DING, Y. & JIANG, Y. 2005. Detection of severe acute respiratory syndrome coronavirus in the brain: potential role of the chemokine mig in pathogenesis. *Clin Infect Dis*, 41, 1089-96.

- XU, X., GAO, W., CHENG, S., YIN, D., LI, F., WU, Y., SUN, D., ZHOU, S., WANG, D., ZHANG, Y., JIANG, R. & ZHANG, J. 2017. Anti-inflammatory and immunomodulatory mechanisms of atorvastatin in a murine model of traumatic brain injury. *J Neuroinflammation*, 14, 167.
- XU, X. W., WU, X. X., JIANG, X. G., XU, K. J., YING, L. J., MA, C. L., LI, S. B., WANG, H. Y., ZHANG, S., GAO, H. N., SHENG, J. F., CAI, H. L., QIU, Y. Q. & LI, L. J. 2020. Clinical findings in a group of patients infected with the 2019 novel coronavirus (SARS-Cov-2) outside of Wuhan, China: retrospective case series. *BMJ*, 368, m606.
- YAN, J., QIAO, L., TIAN, J., LIU, A., WU, J., HUANG, J., SHEN, M. & LAI, X. 2019. Effect of statins on Parkinson's disease: A systematic review and meta-analysis. *Medicine (Baltimore)*, 98, e14852.
- YANG, C. S., LEE, H. M., LEE, J. Y., KIM, J. A., LEE, S. J., SHIN, D. M., LEE, Y. H., LEE, D. S., EL-BENNA, J. & JO, E. K. 2007. Reactive oxygen species and p47phox activation are essential for the Mycobacterium tuberculosis-induced pro-inflammatory response in murine microglia. *J Neuroinflammation*, 4, 27.
- YANG, J., CHEN, J., YUE, J., LIU, L., HAN, M. & WANG, H. 2014a. Relationship between human LTA4H polymorphisms and extra-pulmonary tuberculosis in an ethnic Han Chinese population in Eastern China. *Tuberculosis (Edinb)*, 94, 657-63.
- YANG, Q., CAI, Y., ZHAO, W., WU, F., ZHANG, M., LUO, K., ZHANG, Y., LIU, H., ZHOU, B., KORNFELD, H. & CHEN, X. 2014b. IP-10 and MIG are compartmentalized at the site of disease during pleural and meningeal tuberculosis and are decreased after antituberculosis treatment. *Clin Vaccine Immunol*, 21, 1635-44.
- YEUNG, J. H. Y., WALBY, J. L., PALPAGAMA, T. H., TURNER, C., WALDVOGEL, H. J., FAULL, R. L. M. & KWAKOWSKY, A. 2021. Glutamatergic receptor expression changes in the Alzheimer's disease hippocampus and entorhinal cortex. *Brain Pathol*, 31, e13005.
- YOUNG, C., WALZL, G. & DU PLESSIS, N. 2020. Therapeutic host-directed strategies to improve outcome in tuberculosis. *Mucosal Immunol*, 13, 190-204.
- YOUSSEF, S., STUVE, O., PATARROYO, J. C., RUIZ, P. J., RADOSEVICH, J. L., HUR, E. M., BRAVO, M., MITCHELL, D. J., SOBEL, R. A., STEINMAN, L. & ZAMVIL, S. S. 2002. The HMG-CoA reductase inhibitor, atorvastatin, promotes a Th2 bias and reverses paralysis in central nervous system autoimmune disease. *Nature*, 420, 78-84.
- ZAHR, N. M., MAYER, D., PFEFFERBAUM, A. & SULLIVAN, E. V. 2008. Low Striatal Glutamate Levels Underlie Cognitive Decline in the Elderly: Evidence from In Vivo Molecular Spectroscopy. *Cerebral Cortex*, 18, 2241-2250.
- ZHANG, X., FALAGAS, M. E., VARDAKAS, K. Z., WANG, R., QIN, R., WANG, J. & LIU, Y. 2015. Systematic review and meta-analysis of the efficacy and safety of therapy with linezolid containing regimens in the treatment of multidrug-resistant and extensively drug-resistant tuberculosis. *J Thorac Dis*, 7, 603-15.