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# HEART AND CIRCULATORY PHYSIOLOGY.

**REVIEW** 

Impact of Aging on the Cardiovascular System

# Arterial pulse wave modeling and analysis for vascular-age studies: a review from VascAgeNet

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#### **Abstract**

Arterial pulse waves (PWs) such as blood pressure and photoplethysmogram (PPG) signals contain a wealth of information on the cardiovascular (CV) system that can be exploited to assess vascular age and identify individuals at elevated CV risk. We review the possibilities, limitations, complementarity, and differences of reduced-order, biophysical models of arterial PW propagation, as well as theoretical and empirical methods for analyzing PW signals and extracting clinically relevant information for vascular age assessment. We provide detailed mathematical derivations of these models and theoretical methods, showing how they are related to each other. Finally, we outline directions for future research to realize the potential of modeling and analysis of PW signals for accurate assessment of vascular age in both the clinic and in daily life.

aging; arteriosclerosis; atherosclerosis; hemodynamics; pulse wave

# 1. INTRODUCTION

Pulse wave (PW) signals are produced by the pumping heart and its interaction with the blood and the distensible arterial walls. Cardiac ejection increases blood pressure and distends the wall of the aorta, generating a compression/distension wave that propagates along the aorta and other

conduit arteries. Toward the end of systole, a decline in cardiac ejection rate creates a decompression/relaxation wave that manifests as a decline in pressure and a reduction in aortic diameter. Together, these and other (transmitted and reflected) waves create a waveform that is called the pulse wave. The pulse wave leads to a rhythmical expansion and relaxation of all arteries that follows the heartbeat; e.g.,





producing the pulse that can be felt in the radial artery of the wrist, despite the wrist being about a meter away from the heart. The pulsatile movement of the arterial wall is accompanied by changes, over time and space, in blood pressure, blood flow velocity, and blood volume flow rate throughout the arterial system, called, respectively, pressure, flow velocity, and flow rate PWs.

PW signals can be measured in vivo using a variety of (invasive and noninvasive) devices and are influenced by the heart and the vasculature, making them a rich source of information on cardiovascular (CV) health. In particular, the morphology of PW signals is affected by changes in the mechanical and structural properties of the vascular wall produced by vascular aging or disease, and their impact on cardiac mechanics and structure. Vascular aging is a complex biological process that involves the deterioration in structure and function of blood vessels over time and may occur at a different rate than chronological aging (1). It is a critical component of overall aging that entails an increase in arterial wall stiffness (arteriosclerosis) and the accumulation of atheroma that results in progressive narrowing of the arterial lumen (atherosclerosis) (2, 3). Initially, vascular deterioration is usually an asymptomatic process that eventually can cause damage to the heart, brain, kidneys, and other organs. Measures of vascular age encompass the cumulative effect of all CV risk factors on the arterial wall throughout life (3). Therefore, assessment of vascular age by PW analysis may help identify individuals with early vascular aging (4), and, hence, at elevated CV risk, at an early stage of disease progression.

Arterial PW modeling and analysis aims to unravel the functioning of the CV system through the measurement, mathematical analysis, and computational and experimental simulation of pulsatile hemodynamics (i.e., the dynamics of pulsatile blood flow). In addition to the widely used values of systolic, diastolic, and mean arterial pressure, other clinically relevant information for vascular age assessment can be derived from the morphology of PW signals. As shown in this review, several PW analysis techniques can provide multiple hemodynamic measures and indices that vary with aging and disease, suggesting that they may constitute relevant indicators of age-related CV risk. Models for simulating PWs can be used to investigate the accuracy of these techniques, provide mechanistical insights, and understand the physiological basis underlying measured hemodynamics phenomena. However, PW models should be developed further to better capture the diversity of PW measurements observed in vivo and combined with artificial intelligence (AI) for an improved assessment of vascular aging in daily

This article reviews the possibilities, limitations, complementarity, and differences of reduced-order, arterial PW models (Section 2) and analysis methods (Section 3) for CV assessment, with a focus on vascular age assessment. It aims to provide a comprehensive overview of models and analytical techniques to help someone new in the field get started, including engineers, mathematicians, and physicians, as well as to be a convenient compendium for established researchers. Directions for future research in the field are also provided (Section 4). This article is free of mathematical derivations and equations to make it accessible to readers with a limited mathematical background. It is accompanied by a Technical Supplement (https://doi.org/10.6084/m9.figshare.21758012.v3) containing technical details and mathematical derivations of all the biophysical models and hemodynamics-based analysis techniques covered in the article. All derivations start from the well-known Navier-Stokes equations, to show how different types of models and analysis techniques are related to each other.

#### 2. PULSE WAVE MODELS

Arterial hemodynamics obeys physical laws and principles (i.e., conservation of mass, momentum, and energy) that can be used to mathematically describe (i.e., model) arterial PW signals. There are three main physics-based modeling approaches—zero-dimensional (0-D), one-dimensional (1-D), and three-dimensional (3-D) models—that are illustrated in Fig. 1 and compared in Table 1 in terms of their spatial accuracy, computation time, and advantages and limitations to study vascular aging. Large-scale network simulations of PW signals often require simplification of the 3-D formulation to reduce the computation time while maintaining reasonable accuracy. This can be achieved using the reduced-order 1-D (Section 2.1) and 0-D (Section 2.2) models. There are only a few commercial software programs for PW modeling. One such program is called the "Aplysia CardioVascular Lab" (Aplysia Medical AB, Stockholm, Sweden), which features a basic 0-D model arterial network that enables users to simulate both central and peripheral PWs (5).

#### 2.1. 1-D Models

One-dimensional models of the arterial tree are considered a good compromise between accuracy and computational cost for simulating arterial PW signals. The inviscid 1-D governing equations of conservation of mass and momentum were derived by Leonhard Euler in 1755 (6). Other historical figures that made important contributions to the field of 1-D blood flow modeling include W. Weber, T. Young, J. L. Poiseuille, B. Riemann, and J. R. Womersley. For a historical overview see Parker (7), the PhD theses of Westerhof (8) and Hughes (9), and the introductions of the articles by Hughes and Lubliner (10), and van de Vosse and Stergiopulos (11). The 1-D model formulation is described in Section 2.1.1, followed by an overview of how to calibrate model parameters (Section 2.1.2), verify the accuracy of simulated PW signals (Section 2.1.3), and use 1-D models to study clinically relevant problems for vascular aging (Section 2.1.4).

#### 2.1.1. Formulation.

In 1-D modeling, the arterial network is described as a set of arterial segments interconnected at nodes (Fig. 1B). Within each segment, blood pressure, blood flow velocity, and luminal cross-sectional area vary with time and distance along the axis of the vessel, governed by a system of partial differential equations. The governing equations ensure that the 1) physical principles of conservation of mass and linear momentum for blood flow are satisfied in each arterial segment and 2) interaction between blood flow and vessel wall deformations is accounted for.



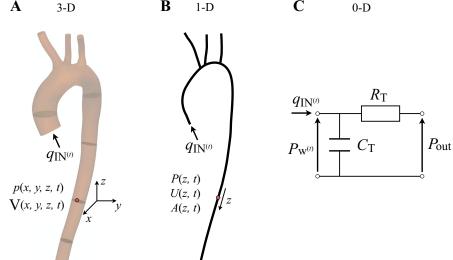


Figure 1. The main arterial blood flow modeling approaches illustrated for the upper aorta. A: threedimensional (3-D) models simulate blood pressure (p), blood flow velocity (V), and wall displacement (not shown) as a function of time (t) and three spatial dimensions (e.g., x, y, and z in Cartesian coordinates). B: one-dimensional (1-D) models describe blood pressure (P), blood flow velocity (U), and luminal area (A) with time and axial direction of the vessel (z). C: zero-dimensional (0-D) models can calculate a space-independent blood pressure  $(p_w)$  for the whole 3-D or 1-D arterial tree as a function of the aortic inflow  $(q_{IN})$ , total compliance  $(C_T)$  and resistance  $(R_T)$ , and outflow pressure ( $P_{out}$ ) at each terminal segment of 3-D and 1-D models (see Eq. 80 in Technical Supplement).

Technical Supplemental Section 2.1.1 provides a detailed derivation of the 1-D governing equations starting from the 3-D Navier-Stokes equations in cylindrical coordinates, based on the work of Barnard et al. (12), and involving the solid mechanics theory of thin-walled pressure vessels. Radial and azimuthal variations in blood pressure and flow velocity, which are considered in 3-D modeling, are neglected in 1-D modeling to reduce complexity and computational cost. This is achieved by 1) assuming cylindrical symmetry to eliminate azimuthal variations Eqs. 2, 11, and  $12^1$ , 2) assuming that axial blood flow velocities are much larger than radial velocities to eliminate secondary terms in the equations (i.e., considering  $\varepsilon \ll 1$  in Eqs. 15 and 16), and 3) integrating over the luminal cross section to eliminate radial variations (Eqs. 3 and 22). The second point follows from the long-wave approximation: arterial pulse wavelengths are much longer (of the order of meters) than vessel wall displacements in the radial direction ( $\ll$ 1 cm) (Eq. 17). Additional assumptions include fixed-length and longitudinally tethered vessels, incompressible and Newtonian fluid<sup>2</sup>, and fully developed laminar<sup>3</sup> flow.

Arterial wall models in 1-D modeling describe the relation between pressure and cross-sectional area. They are referred to as tube laws. These range from purely elastic laws in which the vessel wall elasticity (which decreases with vascular aging) is described by Young's modulus, to more complex laws that account for nonlinear elastic behavior (16), stress relaxation (17–19), wall viscosity (20–22), and wall inertia (16, 23). Technical Supplemental 2.1.1.4 shows how to derive an elastic tube law and extend this law to include wall viscosity.

At the arterial junctions of the arterial network, a junction problem needs to be solved, usually by enforcing the conservation of mass and energy (24), although more complex approaches that account for pressure losses at junctions are also available (25). In addition, appropriate boundary conditions need to be prescribed at the inflow and outflow arterial segments. At the inflow (usually the aortic root), the

flow waveform is often enforced (see, e.g., Refs. 26-28). Alternatively, 0-D models of cardiac contraction can be coupled to the aortic root if the 1-D model network starts there (see Section 2.2.3). Any 1-D model network has to be truncated after a few generations of bifurcations. Indeed, care should be taken when simulating blood vessels with diameters smaller than 1 mm since the assumptions of blood being a continuum and Newtonian fluid start failing as the relative size of red blood cells to vessel diameter increases. Terminal 1-D model branches are often coupled to 0-D models relating the flow to pressure at the branch's endpoint and accounting for physical properties of the downstream vasculature (see, e.g., Refs. 29-31). More sophisticated terminal models include single tapering vessels (32), structured-tree networks (27, 33-35), and open-loop or closed-loop 0-D compartmental models. Structured-tree models can be used to investigate the effects of small-vessel vascular disease, such as stiffening and rarefaction (36), and to predict flow and pressure profiles in the microvasculature. Compartmental 0-D models can describe the peripheral circulation, venous return, pulmonary circulation, and heart chambers; therefore bridging the inflow and outflow boundaries of the 1-D model arterial network and simulating the entire circulation as a closed-loop computational domain (31, 37–41).

Table 2 compares the main characteristics of existing 1-D models for simulating PW signals. These range from single-vessel models (e.g., of the aorta) to closed-loop models of the entire circulation, including the four heart chambers. The aorta and other larger arteries of the head, thorax, abdomen, and upper and lower limbs are often included, and a few models also account for the larger arteries of the pulmonary, coronary, and/or cerebral circulations. Earlier models focused on simulating a few arteries of systemic circulation, with special attention paid to the cerebral arteries. Existing 1-D formulations may differ on the tube law used and the way velocity profiles, convective accelerations, and distal vasculatures are simulated.

<sup>&</sup>lt;sup>1</sup>All the equation labels in the main text refer to equations found in Technical Supplement.

<sup>&</sup>lt;sup>2</sup>A reasonable assumption in large blood vessels (13). Non-Newtonian effects were studied by Robertson and Zakaria (14).

<sup>&</sup>lt;sup>3</sup>Reynolds' numbers based on mean velocities are well below 2,000 in normal conditions (15).



**Table 1.** Comparison of the main arterial blood flow modeling approaches

Model	Spatial Accuracy	Computation Time	Advantages	Limitations
0-D	Space-independent blood pressure, blood flow, and luminal volume	Seconds	Computationally inexpensive descriptions of 1) global blood flow features altered by VA (e.g., systemic arterial compliance) in the whole cardiovascular system and 2) boundary conditions for 1-D and 3-D models	Inability to describe 1) high-frequency PW features, 2) PW propagation phenomena, and 3) spatial variations in vessel geometric and material properties altered by VA
1-D	Crossed-sectionally averaged blood pressure and blood flow velocity, and luminal area along the vessel's axis	Seconds to minutes	Good trade-off between accuracy and computation time to 1) describe PW signals in large-scale networks, accounting for wave reflection and transmission effects and spatial variations in vessel geometric and material properties altered by VA and 2) improve boundary conditions for 3-D models	Inability to describe 1) complex blood flow phenomena with non-negligible radial and circumferential flows due to VA-related structural changes (e.g., aneurysms and stenoses), 2) the mechanical stresses these flows produce on the arterial wall, and 3) blood flow in the microcirculation
3-D	Blood pressure, blood flow velocity, and arterial wall displacements in three dimensions	Hours to days	Description of complex local blood flow phenomena and the me- chanical stresses they produce on the arterial wall, with a high level of geometrical, structural, and biophysical detail	Computationally expensive and reliance on detailed input data that can be challenging or even impossible to acquire (e.g., regional stiffness)

Zero-dimensional (0-D), one-dimensional (1-D), and three-dimensional (3-D) models can all describe time-varying pulse wave (PW) signals. However, they have different degrees of spatial accuracy and computation time, which determine their advantages and limitations to study vascular aging (VA).

Recently, the photoplethysmogram (PPG) signal [an optical measure of the arterial PW that can be measured in daily life (61)] has been simulated using 1-D modeling; either calculated 1) as being proportional to arterial blood volume in a vascular bed (28) or luminal area (62) or 2) from the simulated pressure wave using a transfer function (63).

#### 2.1.2. Calibration.

Arterial PW models are composed of sets of equations, each of which comprises a number of parameters that need to be specified. This is often referred to as calibration. A particular set of parameters allows for the simulation of physiologically realistic PWs representative of a particular subject or pathology. Baseline 1-D models have been calibrated to simulate PWs representative of young, healthy (21, 39) and male subjects (28). These baseline models have then been personalized to simulate PWs for specific subjects (64-66) and adapted to model changes that occur with aging (28, 60, 67-70) (summarized in Table 3), hypertension (24) and its treatment (80), aneurysms (81, 82), stenoses (83, 84), and variability within a cohort (28, 85). The following model parameters are typically calibrated, whereas others are held constant as they have less effect on PWs (28): arterial geometry (length and diameter of each arterial segment), arterial stiffness, flow from the left ventricle into the aorta, and microvascular properties (resistance, compliance, and outflow pressure). The methods used to calibrate models are described in Technical Supplemental Section 2.1.2.

Recently, 1-D models have been used to simulate PWs for a set of virtual subjects representative of a population sample. For instance, in Refs. 28 and 85, PWs were simulated to mimic those that would be measured from samples of healthy adults of different ages. This was performed in three steps: 1) suitable values for parameters were identified from the literature, including mean values for each age group, and

ranges of variability within each age group; 2) these values were converted into model parameters where necessary (such as converting reported PW velocities into Young's moduli); and 3) model parameters were adjusted where necessary to provide more realistic PWs. As illustrated in Fig. 2, this approach allows different types of PWs to be simulated, at a range of anatomical sites, for subjects with different CV properties, and different ages. The morphology of these waves matches in vivo data showing, for instance, 1) a similarity among pressure, luminal area and PPG signals, and between flow velocity and flow rate signals (Fig. 2A); 2) pulse pressure (PP, the amplitude of the pressure PW) amplification from central to peripheral anatomical sites (Fig. 2B); 3) an earlier arrival time of the diastolic peak in the PPG signal with increased arterial stiffness (Fig. 2C), and 4) increases in PP with aging (Fig. 2D).

## 2.1.3. Verification.

Arterial PW models can be verified by comparing the model outputs with reference in silico, in vitro, and in vivo data (see Table 4). Simulated PWs have been compared with reference PWs by qualitatively assessing their shapes and PWderived indices such as mean blood pressures and flow rates (21, 28, 39, 64, 65, 84, 95, 97–100). Error statistics have been used to quantify the performance of models: statistics such as the (relative) root mean square error (RMSE; see, for example, Refs. 64 and 90) and relative (or percentage) error (90) quantify the overall performance of model simulations (see Technical Supplemental Section 2.1.3). Quantitative comparisons have shown relative RMSEs between 1-D model and reference PWs of as little as 1.2% for pressure, 2.1% for the flow, and 2.6% for the luminal cross-sectional area (see Table 4). In these studies, reference PWs included those measured in well-defined CV simulation rigs made of flexible tubes (18, 92-95) and those computed using 3-D fluid-structure



Table 2. Main characteristics of existing 1-D models

References		Closed- Loop	Heart Model	Systemic Circulation	Pulmonary Circulation	Coronary Circulation	Cerebral Circulation	Tube Law	Velocity Profile	Convective Acceleration	Distal Vasculature Models
Streeter et al.	(42)	_	_	– (aorta)	_	_	_	NLE	– P	_	R
Schaaf and Abbrecht	(43)	_	_	+	_	_	_	LE	– P	_	R
Wemple and Mockros	(44)	_	_	+	_	_	_	NLE	+ W	+	3Wk
Raines et al.	(45)	_	_	<ul><li>(leg)</li></ul>	_	_	_	NLE	– P	_	3Wk
Avolio	(46)	_	_	+	_	_	+	VE	+ W	_	R
Stettler et al.	(47, 48)	_	_	+	_	+	_	NLE	– P	_	R
Kufahl and Clark	(49)	_	_	<ul><li>(cerebral)</li></ul>	_	_	+	NLE	– р	+	3Wk
Hillen et al.	(50)	_	_	<ul><li>(cerebral)</li></ul>	_	_	+	LE	– P	_	R
Papapanayotou et al.	(51)	_	_	<ul><li>(cerebral)</li></ul>	_	_	+	LE	– P	_	R
Fitchett	(52)	_	+1C	+	_	_	+	VE	+ W	_	R
Stergiopulos et al.	(29)	_	_	+	_	_	_	NLE	– P	+	3Wk
Cassot and Zagzoule	(53)	_	_	<ul><li>(cerebral)</li></ul>	_	_	+	LE	– P	_	R
Olufsen	(33)	_	_	+	_	_	_	LE	+ BL	_	ST
Wan et al.	(54)	_	_	+	_	_	_	LE	– P	+	R
Sherwin et al.	(55)	_	_	+	_	_	_	LE	– F	_	R
Wang and Parker	(56)	_	_	+	_	_	_	LE	– F	_	R
Formaggia et al.	(30)	_	+1C	+	_	_	_	LE	– P	_	3Wk
Azer and Peskin	(34)	_	_	+	_	_	_	LE	+ W	+	ST
Bessems et al.	(57)	_	_	<ul><li>(aorta and coronary)</li></ul>	_	_	_	LE	+ BL	+	3Wk
Huo and Kassab	(35)	_	_	<ul><li>(coronary)</li></ul>	_	+	_	LE	– P	+	ST
Liang et al.	(38)	+	+4C	+	+	_	_	LE	– р	_	Wk +
Reymond et al.	(21)	_	+1C	+	_	+	+	VE	+ W	+	3Wk
Blanco et al.	(26)	_	_	+	_	+	+	VE <sup>1</sup>	– P	_	3Wk
Müller and Toro	(31)	+	+4C	+	_	_	+	LE	– P	_	Wk +
Qureshi et al.	(27)	_	_	_	+	_	_	LE	+ BL	_	ST
Mynard and Smolich	(39)	+	+4C	+	+	+	+	VE <sup>3</sup>	– р	_	$Wk + \frac{2}{3}$
Acosta et al.	(58)	+	+4C	+	+	_	_	LE	– р	_	Wk + 2
Carson et al.	(59)	+	+4C	+	+	+	_	VE	+ BL	_	3Wk +
Charlton et al.	(28)	_	_	+	_	_	+	VE	– р	_	3Wk
Gallo et al.	(41)	+	+4C	+	_	_	+	VE	+ BL	_	3Wk +
Westerhof et al.	(60)	_	_	+	_	_	+	VE	– P	_	3Wk

Closed-loop: a closed-loop model of the circulation is (+) or not (-) included. Heart model: a heart model is (+) or not (-) coupled to the one-dimensional (1-D) model vessels, with the number of heart chambers (C) indicated. Systemic circulation: the larger systemic arteries are simulated using 1-D modeling (+), as opposed to none or a few arteries as indicated (-). Pulmonary circulation: the larger pulmonary arteries are simulated using 1-D modeling (+), as opposed to none or a few arteries (-). Coronary circulation: the larger coronary arteries are simulated using 1-D modeling (+), as opposed to none or a few arteries (-). Cerebral circulation: the larger cerebral arteries, including the circle of Willis, are simulated using 1-D modeling (+), as opposed to none or a few arteries (-). Tube law: Arterial wall modeled as a linear (LE) or nonlinear (NLE) purely elastic material, or as a viscoelastic (VE) material. Velocity profile: profile calculated (+) using Womersley flow (W) or a boundary layer method (BL), or prescribed (-) using Poiseuille flow (P), a higher-order polynomial (p), a boundary layer method (BL), or a flat profile (F). Convective acceleration: Full term simulated (+) or either simplified by assuming a flat velocity profile or completely neglected (-). Distal vasculature models: single resistance (R), three-element Windkessel (3Wk), Windkessel with more than three elements (Wk+), structured-tree (ST). <sup>1</sup>Elastin, collagen, smooth muscle contributions accounted for; <sup>2</sup>Nonlinear 0-D models, with specific models for the hepatic and coronary beds; <sup>3</sup>Nonlinear elastic term using a power law and Voigt-type viscous term.

interaction models with identical boundary conditions and compatible geometrical and material properties (32, 88, 89, 92). A few comparisons in diseased vasculature with stenosis and aneurysms have also been carried out (84, 92, 98). All these studies show that 1-D modeling can simulate PWs in large arteries, in steady state, supine conditions, and over one cardiac cycle, with a reasonable computational cost and with accuracies comparable with those obtained by 3-D models.

The ability of 1-D models to accurately simulate PWs and precisely mimic changes in PWs under changing CV conditions are both of interest. Statistical relationships between continuous measures derived from PW models are commonly examined using correlation, under the assumption of linear (Pearson's) or monotonic (Spearman's) relationships between variables to produce quantitative estimates of dependency. Information theory-based metrics, such as mutual information

that quantifies all the dependencies between two variables (not just linear or rank dependencies) (101), have also been used in studies of PWs (102, 103) but seem not to have been used as a measure of fidelity. In Refs. 66 and 104, the accuracy and precision of simulated blood pressures were assessed using the bias (i.e., mean error) and limits of agreement (i.e., range around the bias within which 95% of errors are expected to fall), respectively: this separation allows assessment of the suitability of a model for simulating PWs for an individual at a given time and simulating changes in PWs either between individuals or within an individual over time.

#### 2.1.4. Applications.

Arterial network models provide high-resolution arterial pressure and flow waveforms throughout the arterial domain in a fully defined setting. The use of models to simulate PWs



Table 3. Changes in mechanical and structural properties of the cardiovascular system with chronological age

Parameter	Age Variation	References
Heart rate	Nonlinear change in M/F	(71)
Stroke volume	Dec. in M/F	(72)
Cardiac output	Dec. by 24% (M) and 7% (F) between 20 and 69 yr old	(73)
Left ventricular ejection time	No change	(74)
End-systolic elastance	Inc. by $51\%$ between 20 and 80 yr old to normalize left ventricular stress (F $>$ M)	(70, 75)
End-diastolic elastance	Inc. by 51% between 20 and 80 yr old to normalize left ventricular stress $(F > M)$	(70, 75)
Arterial ventricular coupling	Dec. slightly in F; no change in M	(75)
Arterial length	Inc. in proximal aorta length; no change in the lengths of other arterial segments	(76)
Arterial diameter	Inc. in aortic and carotid diameters; no change in the diameters of other arterial segments	(76, 77)
Arterial stiffness	Nonlinear inc.	(78)
Arterial tree compliance	Dec.	(79)
Peripheral vascular resistance	Inc. or no change	(28)

Variations are given for adult males (M) and females (F) and in childhood if available. dec., decrease; inc., increase.

is complementary to clinical studies, and offers many advantages: data can be obtained under a wide range of simulated CV conditions; they allow for studying the effect of changes in the model parameters on the wave shape; they are free of measurement error; they can be obtained simultaneously at all measurement sites; they are relatively inexpensive to obtain; and the reference physiological parameters can be specified precisely (105). Consequently, 1-D models have found several applications in CV research. They are valuable tools with which to 1) study the impact of aging on aortic hemodynamics and wave dynamics (100, 106), 2) provide mechanistic insights into arterial physiology, pathophysiology, and hemodynamic phenotypes (see Refs. 28, 107-109 for some examples), and 3) assess the validity of methods and medical devices for the assessment and treatment of vascular aging.

One-dimensional models have been used to assess the validity of methods to estimate arterial system properties (250, 335), methods for pulse wave velocity (PWV) estimation (85, 112-114), estimation of cardiac output (28, 115, 116), estimation of central blood pressure from peripheral pressure (104, 117–119), to detect aneurysms (81, 82, 120) or stenoses (120), or to estimate ventricular contractility (121). Arterial network models have also provided important insights into the performance of loop-based methods for estimating local PWV, demonstrating their susceptibility to the presence of wave reflections (122, 123). The models have been at the basis of a debate on the accuracy of the Arteriograph, a device intended to estimate aortic PWV from a brachial cuff recording inflated to suprasystolic pressures. Model simulations indicated that the device measures brachioaxillary PWV, rather than aortic PWV, because of reflections and rereflections in the brachioaxillary arterial segment (124). The models have also been instrumental in the debate on the reservoir-wave concept (125), demonstrating inconsistencies in the original formulation of the paradigm leading to spurious interpretation of wave dynamics (and, hence, arterial physiology) confirmed by in vivo experiments (126).

One-dimensional arterial network models may also be particularly suitable to assess the impact of vascular surgical or transcatheter interventions if variables of interest are pressure and flow. Models have, for instance, been used to assess the impact of lower-limb bypass surgery (97, 127) and the creation of a forearm vascular access (arteriovenous shunt) for dialysis on arterial hemodynamics (95). The

forearm model was further extended to account for vascular remodeling and was validated in patients, demonstrating its ability to successfully predict maturation of the arteriovenous fistula in patients (128). Another application of 1-D models is the hemodynamic impact of (aortic) (stent) grafts in the arterial tree, which can be described by changing the local stiffness parameter of the desired section in the arterial tree (129). One-dimensional models can only assess the impact of a stent graft on pressure and flow dynamics, and cannot provide any information on the impact of a stent graft on arterial wall stresses or the local flow field.

Nonetheless, the utility and validity of 1-D models should not be overstretched, as computational models have inherent simplifications and assumptions (see Section 2.1.1). Most models lack important physiological feedback and control mechanisms, and model results depend on the particular topological network that is being simulated, its boundary conditions, and solution methods. It is not guaranteed that, because a method works on simulated data, it will be applicable in any in vivo setting, where measurement error and biological and physiological variability apply. Conversely, it is reasonable to assume that a method that does not perform well on synthetic data would not perform well in vivo.

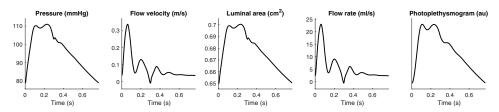
#### 2.2. 0-D Models

Zero-dimensional models further reduce reality to mathematical descriptions without spacial dimensions. Often these are called lumped parameter models since distributed CV parameters are grouped into single parameters; e.g., distributed vessel elasticity is lumped into vessel compliance, which in turn can be described as a single compliance for the whole arterial tree. For a historical overview on 0-D models see Parker et al. (7) and Westerhof et al. (130). This section focuses on the 0-D formulation (Section 2.2.1), the Windkessel (Section 2.2.2), and 0-D heart (Section 2.2.3) models, and their applications (Section 2.2.4).

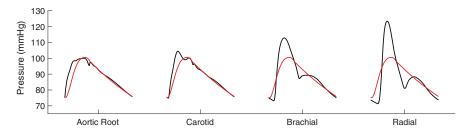
#### 2.2.1. Formulation.

Zero-dimensional models are described by ordinary differential equations, with time as the only independent variable. The linear 0-D equations for 1) blood flow in a blood vessel, 2) the entire arterial tree, or 3) a portion of it can be obtained from the nonlinear 1-D equations, as described in Technical Supplemental Section 2.2.1. Linearization of the

# A Different types of pulse waves



# **B** Pressure pulse waves at different anatomical sites



# C PPG pulse waves in subjects with different cardiovascular properties

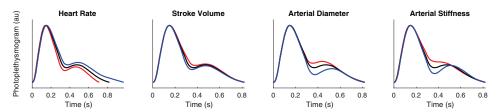
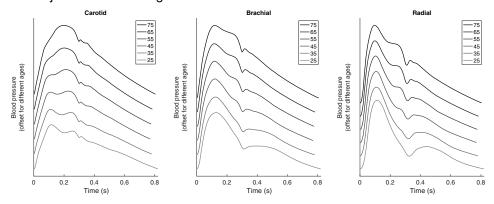


Figure 2. One-dimensional (1-D) blood flow modeling used to simulate arterial pulse waves (PWs). A: simulation of different types of PWs at the carotid artery. B: simulation of pressure PWs at different anatomical sites together with the analytical zero-dimensional (0-D) pressure (red) given by Eq. 80 (Technical Supplement). C: simulation of photoplethysmogram (PPG) PWs at the wrist for subjects with different cardiovascular properties (black, baseline; red, increase; blue, decrease). D: simulation of pressure PWs for subjects of different ages. Source: data were obtained from the Pulse Wave Database (28, 86).

# **D** Subjects of different ages



1-D equations (see Technical Supplemental Section 2.2.1.1) and integration over the vessel length (so that the axial coordinate is eliminated; see Technical Supplemental Section 2.2.1.2) yield the 0-D equations (*Eq. 63*) for blood flow in a vessel segment. These equations are analogous to the transmission line equations and, hence, 0-D models are usually represented by electrical circuits (130) (Fig. 3B). Blood flow and pressure are analogous to electric current and potential, respectively. The compliance of the vessel is equivalent to a capacitance; the inertia of blood is comparable with an inductance; and the resistance to blood flow is matched by a resistor. Electrical analog models have been created to simulate blood flow in the systemic arterial tree (132, 133).

By combining the 0-D model equations for each segment of the arterial tree we can obtain a differential equation (see Technical Supplemental Section 2.2.1.3, *Eq. 76*) relating arterial blood pressure in the entire arterial tree to the following CV parameters that are affected by vascular aging (as described in Table 3): time-varying aortic inflow, outflows to the microcirculation, outflow pressure, and distributed physical properties of the vasculature (length, diameter, and stiffness for each arterial segment, and peripheral compliances and resistances). This equation can be solved analytically for blood pressure as a time-varying, space-independent analytical function (*Eq. 80*). It shows the ability of 0-D modeling to approximate distributed 1-D model pressures, particularly during diastole (Fig. 2*B*),



Table 4. Review of studies assessing the accuracy of 1-D model pulse waveforms

References		Test Data	Test Data Simulated Arteries		ε <sub>Q</sub>	٤٥	$\epsilon_{\mathbf{A}}$
Mynard et al.	(32)	3-D data	Carotid bifurcation	-	-	*	
Reymond et al.	(87)	3-D data	Upper Ao and supra Ao arteries	*	*	-	-
Grinberg et al.	(88)	3-D data	50 larger intracraneal arteries	*	*	-	-
Xiao et al.	(89)	3-D data	CCA, thoracic Ao, aortic bif.	1.4	2.1	-	2.6
Xiao et al.	(89)	3-D data	20 larger sys. arteries	2.1	4.9	-	-
Boileau et al.	(90)	3-D data	CCA, thoracic Ao, aortic bif.	1.2	2.6	-	4.3
Alastruey et al.	(91)	3-D data	Upper Ao and supra Ao arteries	2.0	5.0	-	3.0
Jin and Alastruey	(92)	3-D data	Abdominal Ao, carotid and iliac§	5.4	7.3	-	-
Bessems et al.	(18)	in vitro	Ao†	*	*	-	-
Alastruey et al.	(93)	in vitro	37 larger sys. arteries	2.5	10.8	-	-
Saito et al.	(94)	in vitro	9 larger sys. arteries	10.0	*	-	-
Huberts et al.	(95)	in vitro	Upper-limb arteries	*	*	-	-
Boileau et al.	(90)	in vitro	37 larger sys. arteries	4.0	25.6	-	-
Jin and Alastruey	(92)	in vitro	Ao	5.0	-	-	-
Avolio	(46)	human	128 larger sys. arteries	-	-	*	-
Stettler et al.	(47, 48)	human	Ao and lower limb arteries	*	*	-	-
Olufsen et al.	(96)	human	29 larger sys. arteries	-	*	-	-
Reymond et al.	(21)	human	103 larger sys. arteries	*	*	-	-
Reymond et al.	(64)	human	94 larger sys. arteries	6.0‡	11.0	-	-
Willemet et al.	(97)	human	Lower-limb arteries	9.6	-	16.0	-
Guala et al.	(65)	human	Larger sys. arteries	13.0	-	-	-
Mynard and Smolich	(32)	human	Larger sys. and pul. arteries	*	*	*	-
Alastruey et al.	(91)	human	Upper Ao and supra Ao arteries	10.0	7.0	-	8.0
Strocchi et al.	(84)	human	55 larger sys. arteries	*	*	-	-
Charlton et al.	(28)	human	116 larger sys. arteries	*	*	-	-
Steele et al.	(98)	animal	Aortic bypass	-	4.2	-	-
Mynard et al.	(99)	animal	Left conduit coronary arteries	-	16.7	-	-

The third column shows the type of reference data used in each study. Upper bounds for relative errors (in percentage) for pressure  $(\epsilon_P)$ , flow rate  $(\epsilon_O)$ , flow velocity  $(\epsilon_U)$  and cross-sectional area  $(\epsilon_A)$  wave morphology, calculated as described in the corresponding article, are shown when available. (Adapted from Ref. 92). Ao, aorta; bif., bifurcation; CCA, common carotid artery; pul., pulmonary; sys., systemic. -, no comparison made. \*Qualitative comparison; †according to the dimensions shown in Fig. 4 of (18); †except at the abdominal aorta, where root mean square error is 21%; §for stenosis and aneurysm sizes of up to 85% and 400%, respectively.

and identifies three key factors that describe arterial blood flow: total vascular resistance, total arterial compliance, and outflow pressure. Furthermore, changes in pressure can be assumed to occur synchronously throughout the arterial tree during diastole, with fluid inertia having a negligible effect compared with compliance and resistance (134).

# 2.2.2. The Windkessel model.

The 0-D model for the arterial tree described by Eq. 76 leads to the well-known Frank's two-element Windkessel model (Eq. 79) (135) when all peripheral compliances are neglected. The Windkessel model describes the whole arterial tree as a reservoir of constant compliance into which blood flows from the left ventricle (Fig. 3A). The time-varying pressure in the reservoir encounters a constant peripheral vascular resistance and flows out into the vascular beds that are at a constant pressure (usually assumed to be right atrial pressure or zero). Despite its simplicity, this model is able to predict the exponential decay of pressure in diastole and the increases in mean arterial pressure and PP with, respectively, increasing resistance and decreasing compliance (Fig. 3C); both characteristics of vascular aging. The model predicts a pressure decay with a time constant given by the product of the total resistance and compliance of the arterial network (Eq. 83). Hence, it can describe the steeper diastolic pressure decay observed with vascular aging as a result of the smaller exponential time constant produced by the reduction in arterial compliance (Fig. 2D). Windkessel models are common choices for outflow boundary conditions in 1-D (see Table 2) and 3-D modeling. They can contain more than two elements and physically exist as bench hydraulic models (Fig. 3B) (136).

#### 2.2.3. Heart models and elastance.

There are several possibilities to describe the filing and contraction of the heart in 1-D and 0-D modeling. The simplest approach is to simulate the left ventricle as a "pressure source" and prescribe the pressure PW at the aortic root independently of vascular load. Then, pressure and load together determine blood flow (see Eq. 82). Otherwise, the left ventricle can be modeled as a "flow source:" blood flow is forced into the vascular system and the pressure build-up has no impact on the flow. In reality, the heart is neither a pressure source, nor a flow source; i.e., the pressure that the ventricle experiences while ejecting has an effect on the flow it outputs, and vice versa. However, it has been proposed that a hypertrophied heart resembles a flow source; i.e., it can generate output even if the afterload pressure is high. And a failing heart is closer to a pressure source; i.e., pressure may still be maintained but the flow becomes lessened by the load (137).

More physiologically accurate heart models are based on the pressure-volume description of cardiac function (138-140). Ventricular pressure (P) plotted versus ventricular volume (V) for a complete cardiac cycle produces the so-called PV loop (Fig. 3E) (141). This has four phases: 1) filling phase in diastole (V increases with little P elevation), 2) isovolumic contraction phase (no changes in V, steep P increase), 3) ejection phase (V decreased by stroke volume, relatively moderate P alterations), and 4) isovolumic relaxation phase (no V

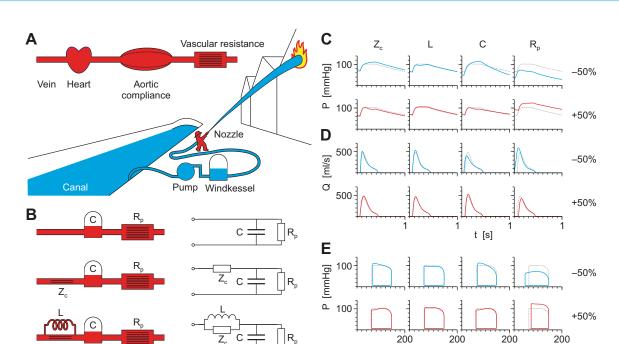


Figure 3. Zero-dimensional Windkessel models of the systemic circulation: fire engine analogy (A), hydraulic (I) and electrical circuit (I) analogies (B); blood pressure (P; C), and blood flow (Q; D) with time, and pressure-volume (P-V; E) loops simulated using the four-element model (131, 144) with independent increases (red) or decreases (blue) by  $\pm 50\%$  in characteristic impedance ( $Z_c$ ), inductance (L), compliance (C), and peripheral resistance ( $R_p$ ), from the baseline model (gray). Changes in  $Z_c$  alter pressure wave shape, with decreased  $Z_c$  causing late systolic peaking. Variations in L have limited impact, with a slight flattening of systolic P observed when L is low. Changes in C affect pulse pressure and systolic peak timing, whereas decreased  $R_p$  lowers overall P and causes earlier ejection. Higher pressures result in lower flows, and changes in P vs. Q appear as "mirrored" alterations. The alterations in P, such as changes in pulse pressure and early or late systolic peaks, can also be seen in P-V loops.

changes, steep P drop). During these phases, the ventricle changes from a high-compliance chamber, receiving blood volume with limited P increase, to a chamber in which P is increased to the extent that it becomes higher than aortic pressure and ejection starts. These phases are conveniently described by the slope of the line defined by a point on the PV loop and a fixed point on the volume axis (i.e., the hypothetical V when P = 0). This slope is referred to as elastance. It measures the rate of change in *P* with the change in *V* and is therefore the reciprocal of compliance (Section 3.2.4). It is low in diastole, increases during contraction, and decreases again with relaxation. The end-systolic elastance ( $E_{es}$ ) is a measure of ventricular contractility, whereas the end-diastolic elastance is a measure of diastolic myocardial stiffness (70). The slope of the line joining the end-systolic and end-diastolic points in the PV loop, called effective arterial elastance ( $E_a$ ), is a measure of arterial load (142). The ratio  $E_a/E_{\rm es}$  is a measure of arterial ventricular coupling (75). Table 3 shows the effect of aging on elastance properties. Zero-dimensional heart models often use a time-dependent elastance curve, which is similar for healthy hearts and several heart diseases when normalized by height and peak onset time. As a result, the same curve shape can be used by adjusting the peak based on heart function and heart rate (139, 140).

#### 2.2.4. Applications.

Zero-dimensional models have been used, e.g., to study the load on the heart (afterload) (130), provide mechanistic

insights into arterial physiology, pathophysiology, and hemodynamic phenotypes (143–146), estimate central blood pressure from aortic flow (104), and estimate cardiac output (147, 148).

The Windkessel model allows us to describe the Windkessel effect, an important vascular function whereby the pulsatile nature of blood flow is smoothed by the elasticity of the arterial wall. The Windkessel<sup>4</sup> was used to "store" pressure in a fire engine (Fig. 3A). By pumping water into the engine at a higher rate than the flow leaves the spout (where the resistance is located), the pressure in the Windkessel (the compliant chamber) increases, compressing the air inside. As a result, a relatively constant pressure can build up, maintaining flow between the strokes of the pump. This results in less wastage of water that would otherwise drop to the ground between the strokes. In a similar way, the beating heart pumps blood into the compliant large arteries, and since the outflow is restricted by vascular resistance (Fig. 3A), blood pressure is built to mean arterial pressure over several cycles. Blood pressure still fluctuates between diastolic and systolic values (Fig. 3C), but not between systolic and nearly zero pressure, as it does within the ventricle (Fig. 3E). Thus, the pressure drop in diastole is limited, the PP is reduced, and the flow is continued between heartbeats. When the arteries become stiffer with aging and disease, the Windkessel function is decreased because of the decrease in compliance and, hence, PP increases (Fig. 3C), in agreement with in vivo measurements (149, 150).

<sup>&</sup>lt;sup>4</sup>Windkessel is German (hence the capital initial letter) for air chamber.

Closed-loop 0-D models have been created to describe the entire circulation using 1) lumped-parameter resistors, capacitors, and inductances to simulate blood flow in the arterial and venous vasculatures and 2) elastance functions to model the right and left heart chambers (151–153). The vasculature may be divided into several compartments representing, for example, thoracic, abdominal, and more distal vasculatures. Closed-loop models are used to study cardiac and vascular pathophysiology for the whole CV system in the neonate, children, and adult (5, 154, 155) and the effect of aging (156). Changes or redistribution of blood volume (e.g., by a changed unstressed volume of the veins) can be modeled in a straightforward manner, as well as the effects of autonomic control of the heart and vasculature (138, 151, 157).

#### 3. PULSE WAVE ANALYSIS METHODS

This section reviews methods for analyzing PW signals and extracting relevant information for vascular age assessment. It begins with considerations of how to obtain in vivo PW signals (Section 3.1) that are ready to be analyzed using theoreticalbased (Section 3.2) and empirical-based (Section 3.3) methods. Both types of analysis techniques can also be applied to in silico PW signals, which often do not require the preprocessing steps described in Section 3.1.2 since they are free of measurement errors and artifacts affecting their quality.

#### 3.1. In Vivo Pulse Wave Signals

The arterial PW can be represented by blood pressure, flow or velocity, arterial distension, and PPG signals. These are continuous signals, showing a typical repeating pattern with each heartbeat. The characteristics and morphologies of PW signals, which differ between different types, measurement sites, and ages, need to be considered when measuring (Section 3.1.1) and preprocessing (Section 3.1.2) them.

#### 3.1.1. Measurements.

PW signals can be probed centrally (e.g., heart, aorta, carotid arteries) or on peripherally accessible measuring sites (e.g., upper arm, wrist, groin, finger, toe, retina, and earlobe). Peripheral PWs differ from central PWs because of the effects of PW propagation and reflection along the arterial tree. Signals can be directly measured with methods such as pressure catheters, applanation tonometry, volume clamp method, oscillometric cuff (179, 180), (Doppler) ultrasound (336), and magnetic resonance imaging (MRI) (181), but are

also increasingly accessible to wearable technologies such as photoplethysmography (182), which is acquired by pulse oximeters and consumer devices (e.g., smartwatches and smartphones). These techniques are compared in Table 5 in terms of characteristics of the PW signals they measure and preferred features for measuring the PW in daily life. Photoplethysmography is widely used because of its ease of use; however, the PW signal it measures is not as well understood physiologically as the signals measured by the other techniques. Pressure catheters are the gold standard for measuring blood pressure and central hemodynamics; however, they are an invasive technique, making them unsuitable for large studies and used in apparently healthy individuals. Ultrasound and MRI provide noninvasive and accurate measurements of blood velocity, blood flow, and luminal diameter, tonometry is a practical technique for noninvasively measuring arterial pressure waveforms, and the use of cuffs to measure PW has become increasingly popular recently. These techniques have been used in many large epidemiological studies and in routine clinical practice, but they are generally not suitable for self-measurement since most require specialist equipment and trained operators.

Since arterial PW analysis depends on detailed features of PW morphology, the reliability of the measurements is crucial. Measurement methods are sensitive to technical errors [e.g., damping or ringing of the catheter-manometer system (183, 184)], artifacts [e.g., movement during MRI acquisition (185)], operator-dependent inaccuracies [e.g., an incorrect insonation angle for Doppler measurements (186)], and physiological effects [e.g., respiration induced changes (187)]. The frequencies of interest in the PW signal are below 20 Hz for adults (188, 189), which means that a sampling frequency of at least 40 Hz is needed according to the Nyquist theorem (190). In general, the greatest care should be taken to obtain good quality data, since in most cases it is impossible to correct measurements afterward.

#### 3.1.2. Preprocessing.

A PW signal is typically preprocessed before analysis to improve the reliability of the analysis. The process consists of several steps, which are summarized in Fig. 4. First, individual PWs are identified for analysis using a beat detection algorithm (193, 194). Second, any periods of signal that are incomplete, of low quality, or that contain data outside of the plausible measurement scope should be excluded. Techniques for assessing the quality of blood pressure and

**Table 5.** Comparison of pulse wave measurement techniques

	Pressure Catheter	Applanation Tonometry	Volume Clamp	Oscillometric Cuff	PPG	US	MRI
Signal type	Р	Р	Р	Р	PW	U, Q, D	U, Q, D
Central signal	+	+	_	_	_	+	+
Peripheral signal	+	+	+	+	+	+	+
Noninvasive	_	+	+	+	+	+	+
Continuous acquisition	+	_	+	_	+	_	_
No trained operator	_	_	_	+	+	_	_
Calibrated	+	_	+	+	_	+	+
Unobtrusive monitoring	_	_	_	+	+	_	_

The first three rows show characteristics of the pulse wave signals directly measured by each technique, and the remaining rows compare techniques in terms of preferred features for measuring the pulse wave in daily life. D, luminal diameter; MRI, magnetic resonance imaging; P, blood pressure; PPG, photoplethysmography; PW, pulse wave signal with arbitrary units; Q, blood flow rate; U, blood flow velocity; US, ultrasound.

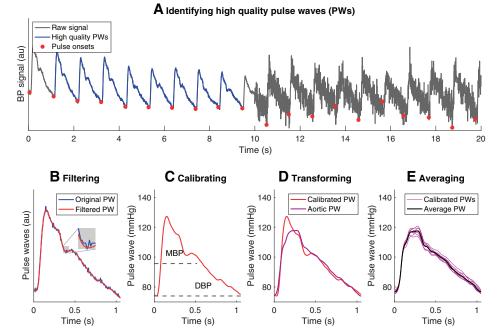


Figure 4. Preprocessing an arterial pulse wave (PW) signal, A: brachial applanation tonometry blood pressure (BP) signal was processed to identify high-quality PW data for analysis (blue, from 1 to 9 s). PW onsets are detected (indicated by red dots). B: individual PWs are filtered to eliminate high-frequency content. C: PWs are calibrated using independent mean and diastolic blood pressure measurements (MBP and DBP, respectively). D: brachial (peripheral) PWs are transformed to aortic (central) PWs E: PWs are ensemble averaged to produce a final PW for analysis. Sources: (data) Brachial data from the Asklepios data set, with artificial noise added (334); (processing) PulseAnalyse (28, 192).

PPG signals are reviewed in Refs. 195 and 196. Third, highfrequency noise (e.g., electrical interference) and low-frequency variations (e.g., due to respiration) can be eliminated through digital filtering (61, 196). The filter design is important: different filter cutoff frequencies may be required for different analyses (197); and zero-phase digital filtering should be used to prevent any phase shifting (196). After filtering, a sufficient number of harmonics should still be present in the signal since excessive filtering can lead to the loss of important information in the PW shape.

There are several additional preprocessing steps that can optionally be performed, as illustrated in Fig. 4: 1) calibrating blood pressure PWs acquired by applanation tonometry with an independent pressure measurement to convert the piezoelectric measurement (voltage) into a pressure signal (198); 2) transforming a peripheral PW to a central PW using a transfer function (117, 199); and 3) ensemble averaging PWs to create a single, averaged PW (169). Other considerations include 4) interpolating signals to increase their temporal resolution, which is particularly helpful when extracting the timings of PWs (such as for PWV estimation) (200); 5) calculating derivatives for PW analysis [e.g., using Savitzky-Golay filtering (201)]; and 6) when working with multiple simultaneous PW signals, ensuring that they are time aligned, such as by the times of systolic upstrokes (see Section 3.3.1), by waveform matching, or by cross correlation (202).

#### 3.2. Theoretical-Based Analysis Methods

The 1-D and 0-D blood flow models presented in Section 2 form the basis of several theoretical PW analysis methods used to assess vascular age. These methods offer valuable insights into the interpretation of PW morphology and its relationship to CV parameters associated with vascular aging and are subsequently reviewed. Table 6 shows how vascular age indices obtained using these methods vary with chronological age, accounting for sex differences if available.

#### 3.2.1. Forward- and backward-traveling waveforms.

Theoretical analysis of the 1-D model governing equations reveals the presence of PW motion in flexible blood vessels (see Technical Supplemental Section 3.2.1.1). PWs propagate in the forward direction (i.e., from the heart to the periphery) and interact with tapered vessels and bifurcations, producing reflected waves that travel back toward the heart, where they can be rereflected into forward-running waves (203). This analysis enables separation of the blood pressure, P. and blood flow velocity, U, waveforms measured at the same location into forward-traveling  $(P_f, U_f)$  and backward-traveling  $(P_b, U_b)$  waves (Fig. 5A); i.e.,  $P = P_f + P_b$  and  $U = U_f + U_b$ (204) (see Technical Supplemental Section 3.2.1.2). The amplitude of  $P_b$  and the ratio of the  $P_b$  amplitude to the  $P_f$  amplitude have been shown to be an independent predictor of CV events (205, 206). Changes in the amplitudes of  $P_f$  and  $P_b$ have been observed with aging (see Table 6).

The separation into forward- and backward-traveling waveforms can identify the direction (forward or backward) of the waves that make up *P* and *U* at a given time within the cardiac cycle. However, this method cannot provide the physical locations in the CV system where the waves originated (207, 208). Alternative separation techniques have been suggested to achieve this when analyzing in silico (208) and in vivo (125, 203) data, although their potential for assessing vascular age and disease has been questioned (209, 210), or needs to be investigated (203).

#### 3.2.2. Wave intensity analysis.

Wave intensity is the rate of energy flux per unit area carried by the PW, and is analogous to acoustic intensity (211). It can be calculated from simultaneous P and U measurements at any location in the arterial network using Eq. 101 (Fig. 5C) and separated into forward- and backward-traveling components using Eq. 107. As shown in Technical Supplemental Section 3.2.2.1, wave intensity measures the prominence of changes in P and U in the forward and backward directions



Table 6. Evolution of vascular age indices with chronological age

Index	Age Variation	Study Type	References
Forward pressure wave amplitude	Inc. in M/F	С	(158)
· ·	Dec. in M/F	L	(159)
Backward pressure wave amplitude	Inc. initially in M/F; flattening in M and falling in F	С	(158)
	Dec. in M/F	L	(159)
Wave intensity	Dec. in forward compression wave in M/F; inc. in backward compression wave in M/F; no change in forward decom- pression wave in M/F	С	(160–163)
Local pulse wave velocity	Inc. in M/F	С	(161, 162)
Regional pulse wave velocity	Exponential inc. in M/F; minor inc. in childhood	С	(3, 78, 164)
	Exponential inc. in M/F (F $>$ M)	L†	(3, 159)
Compliance	Inc. in M/F up to 30 yr old; dec. after 50 yr old ( $F > M$ )	С	(165, 166)
	Inc. in 35–40 yr old M (but not in F); minor dec. over 40–55 yr old in M/F	L	(159)
Aortic input impedance	Inc. in M/F; rightward shift of the minimum modulus	С	(166, 167)
	No significant changes over 10 yr in middle-aged M/F	L	(159)
Aortic characteristic impedance	No significant change	С	(168)
Pulse pressure	Exponential inc. in M/F; minor inc. (or dic.) in childhood	С	(3, 164)
	Exponential inc. in M/F; can dec. in young and elderly M	L	(3)
Pulse pressure amplification	Dec. in M/F	С	(164, 169)
Augmentation index	Inc. in M/F (M $>$ F); dec. in childhood	С	(3, 164)
Augmentation pressure	Linear inc. in M/F (F $>$ M); plateau in elderly M	С	(3, 164)
Ankle-brachial index	Inc. in M/F up to 60–69 yr old; dec. after (M $>$ F)	С	(170)
Ambulatory arterial stiffness index	Inc. in M/F (F $>$ M)	С	(171)
Cardio-ankle vascular index	Linear inc. in M/F (M $>$ F)	С	(172)
Flow augmentation index	Quadratic upwardly concave inc. in M/F	С	(173)
Reverse-to-forward flow index	Inc. in M/F (aorta); no significant change (femoral artery)	С	(174, 175)
Diastolic-to-systolic forward flow ratio	Dec. in M/F (femoral artery)	С	(174)
Photoplethysmogram second derivative	Linear inc. in $b/a$ and aging index in M/F; linear dec. in $c/a$ , $d/a$ and $e/a$ in M/F	С	(176–178)

These indices have been calculated using the theoretical-based and empirical-based methods described in Sections 3.2 and 3.3, respectively, in cross-sectional (C) or longitudinal (L) studies. Variations with chronological age are given for adult males (M) and females (F), and in childhood if available. dec., decrease; inc., increase. †Results found for carotid-femoral and brachial-ankle pulse wave velocity only.

at any time during the cardiac cycle (204, 211). It is particularly well suited to understanding the role of wave reflections on pressure and flow in systemic arteries (212-219), including coronary circulation (220-222). In vivo studies at the ascending aorta have shown that the magnitude and arrival time of the backward compression wave in mid-systole varies with age (see Table 6), disease, arterial compliance, and vascular tone (213, 223), and the magnitude of the forward compression wave is a predictor of cognitive decline (160).

The units of wave intensity vary between studies, which can limit comparison of data across different studies (see Technical Supplemental Section 3.2.2.2). In addition, several other factors affect absolute wave amplitude. Noise reduction, using techniques described in Section 3.1.2, is crucial in reducing the effect of noise when calculating wave intensity (224). Moreover, time alignment of P and U is critical for accurate calculation of wave intensity, since misalignment can lead to inaccurate wave profiles and artifactual waves. Finally, differentiating between "real" wave peaks and background noise can be challenging, but may be aided by a recently described maximum entropy technique (225).

In an attempt to use wave intensity analysis as a diagnostic tool, the equations can be rewritten using luminal diameter and flow velocity (see Technical Supplemental Section 3.2.2.3); both of which can be measured noninvasively (see Table 5). In healthy subjects, this noninvasive wave intensity has shown a decline in left ventricular early and late systolic functions with age (161) and a greater effect of the aging process on the carotid than the femoral artery (162).

Numerical studies using 1-D blood flow modeling have shown that wave intensity can identify the timing, direction, and magnitude of the predominant waves that shape aortic pressure and flow waveforms in systole. However, wave intensity fails to identify the important contribution of wave reflections during diastole and those arising from pulses in previous cardiac cycles (209, 226, 227). This occurs because wave intensity analysis tends to accentuate high-frequency waves, whereas repeated reflections in the arterial tree, along with wave dispersion, attenuate high frequencies and lead to low-frequency waves predominating during diastole (227, 228).

#### 3.2.3. Pulse-wave velocity.

PWV is the speed by which the PW travels in arteries. Several methods have been proposed to determine PWV, which can broadly be grouped under two categories based on 1) local and 2) regional measurements. Local methods use measurements at a single location of either pressure and velocity, diameter and velocity, flow rate and area, or pressure and diameter. The classical 19th-century Moens-Korteweg equation (Eq. 116) (229, 230) was originally introduced in the context of flows in thin elastic tubes. It relates local PWV to the geometrical and mechanical properties of the local arterial wall, showing an increase in PWV with increasing elastic modulus (i.e., wall stiffness) and wall thickness, and with decreasing luminal radius. Therefore, PWV quantifies arterial stiffness, which is of clinical interest for assessing the arteriosclerosis component of vascular aging (3). In the 20th century, the Bramwell-Hill equation was introduced, which describes the relationship

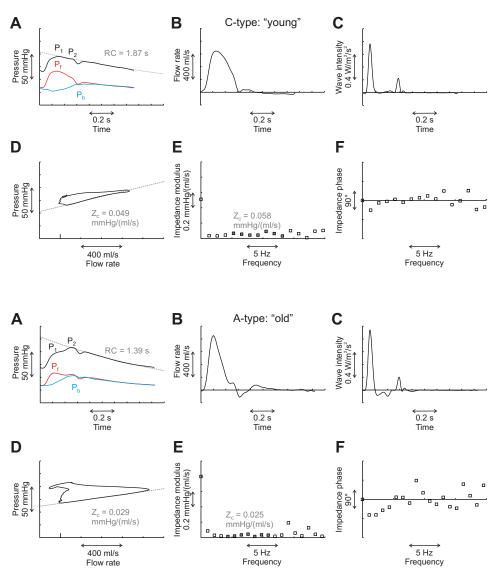


Figure 5. Theoretical-based methods of pulse wave (PW) analysis. The following methods are applied to ensemble averaged pressure and flow waveforms measured in the ascending aorta of young (top) and old (bottom) subjects (149). A: pressure wave separation into forward (Pf, red)- and backward ( $P_b$ , blue)-traveling components using the flow rate in B. RC is the time constant of the total pressure wave (black). P1 and P2 are the inflection points described in Section 3.3.1. C: wave intensity analysis. D: pressure-flow loop with the calculated characteristic impedance ( $Z_c$ , see Eq. 134) from the straight portion. E: impedance modulus. F: impedance phase. E shows  $Z_c$ calculated from the 4th to 10th harmonics (filled boxes).

between local PWV and local arterial wall distensibility (*Eq. 111*). In Technical Supplemental Section 3.2.3.1, the Bramwell–Hill equation is derived from the 1-D model governing equations. In Technical Supplemental Section 3.2.3.2, the Moens–Korteweg equation is derived from the Bramwell–Hill equation.

Early in the 21st century, a series of techniques for estimating local PWV were introduced. These can be classified as loop and sum-of-squares methods. Most of the loop methods rely on the existence of a reflection-free period within the cardiac cycle (231–234) (Fig. 5D) (see Technical Supplemental Section 3.2.3.3). The sum-of-squares technique (Technical Supplemental Section 3.2.3.4) was introduced to assess local PWV from simultaneous pressure and velocity measurements in the coronary arteries, where a reflection-free period cannot be safely assumed during the cardiac cycle (235). These methods have primarily been studied using in vitro and in silico data. Novel derivations of the sum-of-squares method for diameter and velocity, or flow rate and area are provided in Technical Supplemental Section 3.2.3.4.

Regional methods require pulse waveforms measured at two arterial sites; e.g., along the aorta (236, 237), at the carotid and

femoral arteries (238), or at the brachial and ankle arteries (239). Regional PWV is calculated as the ratio of the distance between the two measurement sites to the time delay for the wave to travel from one site to the other. Distances are typically measured from surface markings or intra-arterial distance, and time delays by identifying the feet of the two waves measured at the two locations or using cross-correlation methods (113, 240). Regional PWV measures increase with age starting in childhood (see Table 6). The effects of aging, however, are not uniform in systemic arteries: central arteries such as the aorta stiffen with age more than peripheral arteries in the arms and legs (241). Regional PWV measures have been combined with CV risk factors (242) and a measure of atherosclerosis, such as coronary artery calcification (243), intima-media thickness (244), to quantify an individual's vascular age. Alternatively, regional PWV alone may be able to quantify an individual's vascular age (243) and predict CV events and mortality (245, 246).

Computational blood flow modeling has been used to assess the performance of local (122, 226, 234, 247) and regional (85, 113, 124) measures of PWV. These studies have shown that *I*) methods using aortic PW data as well as the carotid-femoral



foot-to-foot method are accurate indicators of aortic stiffness, 2) other local and regional methods tend to over- or underestimate aortic PWV, and 3) large PW reflections have an adverse effect on the accuracy of PWV estimates.

#### 3.2.4. Compliance.

Compliance is the rate of change in blood volume with the change in blood pressure (often expressed in mL/mmHg or m<sup>3</sup>/Pa). It quantifies the buffer capacity or Windkessel effect of the vasculature (see Section 2.2.4) and is directly related to arterial size and inversely related to local PWV (see Technical Supplemental Section 3.2.4). Direct measurement of compliance is impossible, as it would require sealing off the arterial tree for a pressure inflation test. The most simple estimate of compliance is the ratio of stroke volume to PP (248), but this method does not account for arterial outflow in systole and overestimates compliance (249).

Other methods implicitly or explicitly rely on an assumed Windkessel model (Fig. 3B) (130). In the decay time method, an exponential is fitted to the diastolic part of the aortic pressure wave, with the time constant providing an estimate of the product of total peripheral resistance and compliance, also termed the decay time of the arterial system (250) (see Eq. 83 and Fig. 5A). A variation of this method is the area method (251), computing the decay time from the area under the diastolic pressure waveform, rather than fitting an exponential. Note that to obtain compliance using methods that estimate the decay time, resistance must be calculated from mean arterial pressure and flow (cardiac output). To eliminate the sensitivity of the area and decay time method to wave morphology (that may be far from exponential) or the selected diastolic segment, the PP method was introduced (111, 252): an iterative method estimating compliance through minimizing the difference between the measured PP and the pulse predicted by a two-element Windkessel model. The method is robust with results highly correlating with the ratio of stroke volume to PP (249). Virtually all other compliance estimation methods make use of more complex Windkessel models consisting of more elements (3-element or 4-element Windkessel models) (110, 251, 253) and nonlinear terms, including pressure-dependent (254) or frequency-dependent compliance (255) (see Technical Supplemental Section 3.2.4).

With compliance depending on both arterial stiffness (which is dependent on blood pressure) and arterial size, the relation with age is not straightforward. It has been reported to increase up to age 30, vary little in middle age, and decline rapidly above age 50 (165). Longitudinal analysis of data from round 1 of the Asklepios study in subjects aged 35-55 yr confirmed a relatively constant compliance in males, but a decrease in compliance in females (168), consistent with the study by Waddell et al. (166). Longitudinal data from the Asklepios population, with an effective age change of  $\sim$ 10 yr, showed an increase in compliance in the younger men (35-40 yr at baseline) but not in females. Compliance remained fairly constant at the higher age categories, suggesting that the increase in stiffness (increase in PWV) is balanced by a change in aortic dimensions in this age range (159).

#### 3.2.5. Input impedance and characteristic impedance.

Although hydraulic resistance is calculated as the ratio of mean pressure drop (difference between inlet and outlet of the resistance) and mean flow, impedance can be defined as the ratio of the pulsatile components of pressure and flow. Rather than viscous friction, inertia of the blood and vessel stiffness are the determinants of impedance (256). The terminology is adopted from electrical engineering. Impedance is typically calculated in the frequency domain, after Fourier decomposition of pressure and flow into harmonics (see Technical Supplemental Section 3.2.5). It is a complex number, most often represented in a modulus (Fig. 5E) and phase (Fig. 5F) notation, where the modulus represents the ratio of the amplitude of corresponding pressure and flow harmonics, and the phase angle the phase delay between both (see Eq. 135). When calculated from ascending aorta pressure and flow, it is termed input impedance (often denoted as  $Z_{in}$ ) and constitutes a global systemic description that characterizes the cumulative effect of wave travel and reflection from the arterial tree and constitutes the afterload of the heart (257, 258).

A special case arises for a system that is free of reflections.  $Z_{in}$  is reduced to characteristic impedance ( $Z_c$ ), which can be shown to approximate the ratio of the product of blood density and local PWV to the vessel cross-sectional area (Eq. 137). Therefore,  $Z_c$  constitutes a local arterial parameter. Since, for high frequencies, the arterial system can be considered to be reflectionless because of destructive interference of reflected waves (259),  $Z_{\rm in}$  at high frequencies approaches  $Z_c$  (Eq. 146). Hence,  $Z_c$  can be calculated by averaging the modulus of  $Z_{\rm in}$  between the fourth and tenth harmonics (Fig. 5E) (260). Alternatively,  $Z_{\rm c}$  can also be estimated in the time domain from the ratio of changes in pressure and flow in early systole (Fig. 5D) (261) (see Technical Supplemental Section 3.2.5).

Arterial impedance has been used less often to assess vascular age. Aortic  $Z_{in}$  has been shown to increase with aging in healthy populations (167), suggesting that it may constitute a relevant indicator of age-related CV risk. A study involving over 2,000 healthy individuals aged 35 to 55 found  $Z_{\rm in}$  to evolve from a pattern indicative of wave transmission and reflection in the younger to a pattern more compatible with a Windkessel-like system in the elder. In women, but not in men, a decrease in total arterial compliance led to an increased  $Z_{\rm in}$  in the low-frequency range. Little to no changes with age were observed in  $Z_c$ , possibly because of compensatory effects of aortic dilatation and stiffening (168). Albeit, arterial impedance can provide major mechanistic insights into age-related changes in vascular function, other parameters (e.g., PWV) are required for a more complete interpretation and to disentangle the effects of changes in stiffness from changes in arterial dimensions.

## 3.3. Empirical-Based Analysis Methods

This section reviews indices of vascular age that are based on empirical analyses of PW morphology or on semiempirical analyses incorporating theoretical concepts described in Section 2. These indices require the identification of fiducial points on a PW (Section 3.3.1) from which indices of vascular age can be calculated (Section 3.3.2). Figure 6 shows examples of how the shapes of 1) carotid blood pressure and 2) finger PPG PWs change with age, allowing the effects of aging to be elucidated from the shapes of PWs.

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#### 3.3.1. Fiducial points.

Several fiducial points can be identified on a PW signal, as shown for the PPG PW in Fig. 7A. Accurate identification of these fiducial points is crucial for reliable analysis of PW shape. We now describe the methodology for identifying fiducial points that are common to all PW types, followed by the methodology for fiducial points that are specific to different types of PWs.

3.3.1.1. Systolic and diastolic phases. PWs can be separated into systolic and diastolic phases, where the systolic phase corresponds to the time during which blood is ejected from the left ventricle into the aorta, and the diastolic phase corresponds to the time during which no blood is ejected. When PWs are measured at sites close to the heart, the end of systole can be identified on blood pressure waves as the time of the dicrotic notch (as described later), and on flow velocity PWs as the time at which the flow velocity reduces to (close to) zero. Under normal physiological conditions, flow into the aorta is zero during diastole (Fig. 5B), but more distally a positive diastolic flow may be present (Fig. 8).

3.3.1.2. Pulse onset. The pulse onset is the local minimum at the start of each PW, often called the foot of the wave, which indicates the beginning of systole (see red dots in Fig. 4A). The simplest approach to identify the pulse onset is as the point corresponding to the minimum value on a PW. On pressure PWs this corresponds to the diastolic blood pressure (DBP). Several more complex approaches have been proposed to more accurately identify the pulse onset for use in pulse transit time measurement (113). For example, the intersecting tangents method fits a line to the upstroke and another line to the preceding diastolic decay (or simply a horizontal line to the diastolic minimum). The intersection of the two tangents is then used to define the onset of the

wave. Other approaches include using the maximum in the first or second derivative (202), or using a slope sum function to accentuate the systolic upstroke and therefore more clearly define the pulse onset (262). The choice of approach can influence the accuracy of parameters derived from the timing of the pulse onset, such as PWV (240, 263).

3.3.1.3. Systolic peak. The systolic peak is the highest point on a PW, separating the systolic upstroke from the systolic downstroke. On pressure PWs this corresponds to systolic blood pressure (SBP) and on flow velocity PWs it corresponds to systolic forward peak velocity (Fig. 8). PPG PWs exhibit a similar systolic peak (Fig. 7, top), although the timing of the systolic peak can vary greatly between measurement sites (such as finger and wrist) (264).

3.3.1.4. Dicrotic notch. In pressure PWs, the closure of the aortic valve causes a (relatively sharp) notch called an incisura. This may be seen at the end of systole in central pressure PWs, at approximately one-third of the heart period, and around one-third down the descending part of the wave (Fig. 5A and Fig. 6A; ages, 30-39 and 40-49 yr). The notch may be followed by a secondary dicrotic wave because when the aortic valve closes, the elastic recoil of the aorta can cause a small increase in blood pressure (265). As the wave travels away from the heart, the waveform changes with the incisura becoming less sharp, which is then called a dicrotic notch, and also sometimes followed by an increasing secondary dicrotic wave (266) (Fig. 6A; ages, 30–39 and 40– 49 yr). With the change in the waveform, the time relation of the dicrotic notch and the incisura as functional measures of end ejection may become disturbed.

A range of algorithms has been proposed to identify the dicrotic notch. The first approach is to analyze the original pressure PW, or its first or second derivatives (267). A challenge with this approach is to correctly identify dicrotic notches even

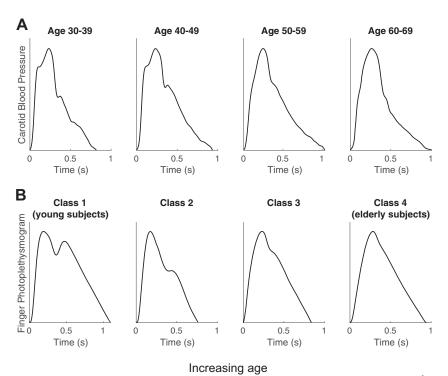


Figure 6. Changes in in vivo pulse waves (PWs) with age: changes in carotid pressure PWs (A) and changes in finger photoplethysmogram (PPG) PWs (B), labeled with classes as it is common for this type of PW (269). Sources: blood pressure data from the Asklepios data set (334); photoplethysmogram data from the VORTAL data set (182). Figure adapted from "Classes of photoplethysmogram (PPG) pulse wave shape," Wikimedia Commons, under CC-BY 4.0.

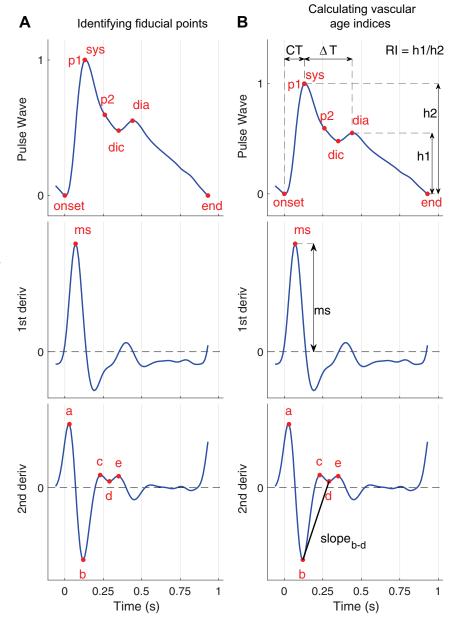


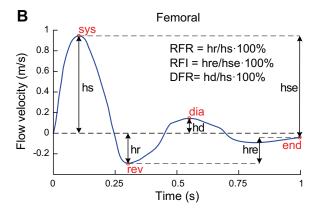
Figure 7. Extracting vascular age indices: vascular age indices can be obtained from a single photoplethysmogram (PPG) pulse wave (PW) in two steps. A: identifying fiducial points on the PW [systolic (sys) and diastolic (dia) peaks, dicrotic notch (dic), early and late systolic peaks (p1 and p2)], its first derivative [slope of the rising front (ms)], and its second derivative (a, c, e peaks and b and d troughs). B: calculating features from the amplitudes and timings of these points, such as the time from pulse onset to sys (CT), the time from sys to dia ( $\Delta T$ ), the reflection index (RI), the maximum upslope (ms), and the slope between b and d troughs (slopes b-d). Sources for A and B: Peter Charlton, Photoplethysmogram (PPG) pulse wave fiducial points (CC tBY 4.0; A) and Photoplethysmogram (PPG) pulse wave indices (CC BY 4.0; B) https://commons. wikimedia.org/wiki/File:Photoplethysmogram\_(PPG)\_ pulse\_wave\_indices.svg.

when they only manifest as an incisura rather than a notch. An alternative approach is to estimate an arterial flow signal from the pressure signal using a three-element Windkessel model, and then identify the dicrotic notch as the minimum of the first negative dip in the flow signal after the systolic peak (268). Figure 6A illustrates some of the difficulties in detecting the dicrotic notch. It is clearly visible on the downslopes of the pressure PWs for 30- to 39- and 40- to 49-yr-old subjects (at  $\sim$ 0.3 s). However, it manifests as a less clear incisura in the 60- to 69yr-old PW. Similarly, dicrotic notches can be less clear at more peripheral measurement sites. It has been suggested that the presence or absence of a clearly defined dicrotic notch in a peripheral pressure PW may be indicative of CV health or aging

The PPG signal can exhibit a trough at a similar time to the dicrotic notch on a pressure PW (e.g., the Class 1 wave in Fig. 6B). Although this may look similar to the dicrotic notch,

it is not yet clear how closely this corresponds to the end of systole (270).

3.3.1.5. Fiducial points on pressure PWs. The shape of the pressure PW is determined by the shapes of the forward and reflected waves (Fig. 5A). The addition of reflected waves to the forward wave may result in an inflection point during systole or anacrotic notches (i.e., notches in the systolic upstroke). An inflection point occurs where the slope of a curve changes from becoming less steep to becoming steeper again (formally: the curvature changes from "concave downward" to "concave upward"). Inflection points are typically identified from higher-order derivatives of the curve. When the inflection point occurs before the systolic peak [an Atype PW as shown in Fig. 5A, bottom (149)], the pressure at the inflection point is termed P1, and the systolic pressure is termed P2 (271). In this case, P1 can be defined as coincident with the inflection point (identified using the 4<sup>th</sup> derivative) Time (s)



**Figure 8.** Definition of fiducial points and indices on flow velocity pulse waves (PWs). *A*: flow augmentation index (Alx) can be obtained, e.g., in the carotid artery, using fiducial points of early (sys1) and late (sys2) systolic peak velocities, and end-diastolic (end) velocity. *B*: on bidirectional PWs, e.g., in the femoral artery, reverse-to-forward flow ratio (RFR), reverse-to-forward flow index (RFI), and diastolic-to-systolic forward flow ratio (DFR) can be obtained using fiducial points of systolic (sys) and diastolic (dia) forward peak velocities, and reverse (rev) peak velocity. Sources: flow data taken from Refs. 173 and 174.

(272), or alternatively defined as the "shoulder point" just before the inflection point (identified using the  $2^{nd}$  derivative) (273). When the inflection point occurs after the systolic peak [a C-type PW as shown in Fig. 5*A*, *top* (149)], the systolic pressure becomes P1 and the pressure at the inflection point becomes P2. In this case, P2 can be identified using the  $2^{nd}$  derivative (273).

The wave shape changes when traveling from the heart to the periphery and with it, the timing and height of the inflection points and anacrotic notches. The A-type wave commonly seen in the aorta transforms to a C-type in the more distal vessels. A central A-type with a small augmentation (i.e., difference between P1 and P2) is related to a low postsystolic inflection point in the periphery, whereas a high A-type augmentation corresponds to a high peripheral inflection point (271). Since it thus seems that the same information may be obtained from a peripherally and centrally measured wave, it has been suggested that preprocessing by using a transfer function to reconstruct the corresponding central wave is not necessary (274). Extensive derivative-based analysis techniques for peripherally measured waves have been developed, see for instance Ref. 176.

3.3.1.6. Fiducial points on PPG PWs. The finger PPG signal is typically characterized by multiple points (61). The PW can exhibit a diastolic peak, indicated by "dia" in Fig. 7, top. It has been hypothesized that this peak is caused by wave reflections, which is why it is more prominent in younger subjects (Fig. 6B) (275). The first derivative is dominated by a point of maximum slope, indicated by "ms" in Fig. 7, middle. The second derivative is typically described using five fiducial points, named a, b, c, d, and e, whose amplitudes vary with age (176) (Fig. 7, bottom). Algorithms have been proposed to identify these fiducial points (276, 277). Much of the literature on these fiducial points is based on the analysis of finger PPG PWs. Further research is required to determine whether they can be accurately identified at other anatomical sites such as the wrist (264).

3.3.1.7. Fiducial points on flow PWs. In cases where flow augmentation is present, such as in a common carotid artery, the systolic phase of the flow velocity PW may contain not a single systolic peak, but early and late systolic

peaks, which can also be called shoulders (Fig. 8A) (173). A bidirectional flow velocity PW, which can be seen, e.g., in the femoral artery (Fig. 8B) or the distal aorta, exhibits the reverse peak velocity (174, 175). Since the volume flow rate is assessed from the cross-sectional area of and blood flow velocity at the artery of interest, the flow rate and flow velocity waves exhibit similar morphology (assuming that the influence of arterial diameter is negligibly small).

#### 3.3.2. Indices of vascular age.

Several indices of vascular age can be calculated from fiducial points identified on pressure, flow, and PPG PWs. Table 6 shows the evolution of these indices with aging.

3.3.2.1. Pressure indices. Pulse pressure (PP), the difference between systolic and diastolic pressures, is an easily accessible measure of vascular age (278) which is associated with unfavorable CV outcomes (279). The increase in PP with aging has several consequences for CV health, including 1) increased left ventricular afterload leading to left ventricular remodeling, dysfunction, and failure (280, 281) and 2) microvascular lesions in high-flow/low-resistance organs, such as the brain and kidneys, by increased transmission of pulsatile pressure and flow to the microvasculature (282, 283).

Pulse pressure amplification (PPA) describes the increase in PP along the arterial tree. It is primarily quantified as the ratio of distal to proximal PP (284), although other definitions have been proposed (285, 286). Amplification decreases with age (164, 169), increases with heart rate (287), and is different for men and women (288). A lower PPA is usually associated with increased CV risk (205, 289) and vascular aging (290).

The difference in pressure between P1 and P2 is called augmentation pressure (or  $\Delta P$  in the original publication) (149).  $\Delta P$  indicates the rise in pressure because of reflected waves. It is often expressed as the augmentation index, AIx =  $\Delta P/PP$ . When the inflection point occurs after the systolic peak (C-type waves shown in Fig. 5A, top), P1 > P2, and therefore  $\Delta P < 0$  and AIx < 0: this does not imply that the reflected pressure wave is negative, but that it starts contributing at a late point in time.  $\Delta P$  and AIx were introduced as measures of wave reflection (238), but the relation between the two is



not straightforward (see also Refs. 105 and 108). In general,  $\Delta P$  and AIx increase with age (see the A-type waves in Fig. 6) (168, 291). Alx has been proposed as a surrogate for PWV. However, the association between AIx and PWV is limited (292), or even nonexistent when using invasive measurements (293). Although arterial stiffness continues to increase with age, the rise in AIx levels off around the age of 60 yr (164, 294). The increase with age of AIx is related to the decrease in PP amplification (169). Despite the interpretation of AIx not being straightforward, several significant associations have been established between AIx and CV disorders (295-298).

Since ventricular ejection patterns influence the shape of the forward wave (and thus, also of the reflected wave) (203), heart function has an impact on the inflection points from which  $\Delta P$  and AIx are calculated (299). This at least in part explains the obfuscated relation between AIx and timing of the reflected wave (191). Recently, the notion that inflection points are not purely markers of the vascular status, but also of heart function, has been gaining momentum (300–303). For instance, the need to correct the AIx for different heart rates (304) points to the influence of heart function on inflection points.

Interestingly, the time at which the inflection point is found on the A-type wave hardly changes with age (305). This opposed the general view that reflections occur at one or two distinct distal reflection sites, and that with agerelated increases in arterial stiffness and thus PWV, the reflected wave would return earlier. The notion of a few distinct reflection sites is probably an oversimplification; a more comprehensive analysis of wave reflection using 1-D models reconciles the findings of increasing AIx with limited changes in "reflection time" (306, 307). It is not yet clear how the arrival time of reflected waves from pressure PWs can be best determined (273, 308). Indeed, detailed model-based analysis suggests that neither the inflection point nor the shoulder point can be directly related to the return of the reflected wave (309, 310).

The ankle-brachial index (ABI) is an easily accessible and well-known indicator of the atherosclerosis component of vascular aging. It is calculated as the ratio between ankle and brachial systolic blood pressure (311). A more novel index of atherosclerosis is the ambulatory arterial stiffness index (AASI). It is assessed by a 24-h ambulatory blood pressure measurement and therefore loses the disadvantages of a snapshot measurement. AASI describes the linear relationship between systolic and diastolic blood pressure (312), however, is not so much a measure of arterial stiffness, but more a measure of ventriculo-arterial coupling determined by heart rate and vascular resistance (313, 314). The cardio-ankle vascular index (CAVI) is an advanced index reflecting the stiffness of the arterial tree. It uses PWV to deduce the parameter  $\beta$  (315), which was developed as a pressure-independent measure of stiffness (316). However, the pressure independence has been challenged and an improved parameter has been provided (317).

3.3.2.2. Flow indices. The flow AIx is defined similarly as pressure AIx and is calculated as the ratio between the late and early systolic velocity wave heights (Fig. 8A); thus, flow AIx is related to the amplitude and timing of wave

reflection (318). It has been shown that carotid flow AIx is more closely associated with age, arterial stiffness parameters (such as a ortic PWV, compliance, and elastic/muscular PWV ratio), and microvascular damage in brain than aortic pressure AIx (173). The aortic reverse-to-forward flow ratio has been found to be independently associated with aortic PWV and characteristic impedance, supporting the hypotheses that a rtic stiffness determines the extent of flow reversal from the descending aorta to the aortic arch (175). Furthermore, aortic arteriosclerosis (assessed as reduced PPA, increased aortic PWV, and pressure augmentation) affects femoral flow wave morphology by decreasing femoral reverse-to-forward flow index and diastolic-tosystolic forward flow ratio (Fig. 8B) (174).

3.3.2.3. PPG indices. Several indices of vascular age can be derived from the PPG PW, as illustrated in Fig. 7B. These are typically calculated from the time delay or difference in amplitudes between two fiducial points. For instance,  $\Delta T$  is the time delay between systolic and diastolic peaks (top), and the aging index is calculated from the amplitudes of points on the second derivative (bottom) as (b-c-d-e)/a. The wide range of PPG-based indices is reviewed in Ref. 182, and the most pertinent indices are now discussed.

The aging index has been found to correlate with carotidfemoral PWV and chronological age (176, 319), and to be associated with the presence of atherosclerosis. The aging index was designed to increase with chronological age, with correlations of r = 0.80 and r = 0.42 with age reported in the original and a subsequent publication, respectively (176, 177). Multiple studies have found that the aging index may have utility as a measure of atherosclerosis (320, 321).

The stiffness index is calculated as height/ $\Delta T$ , providing a value in m/s to mimic PWV measurements. It has been found to correlate with carotid-femoral PWV (r = 0.65) (322), to be a genetically causal risk factor for coronary artery disease (323), and to be higher in diabetic than healthy subjects (324). The stiffness index is available in the UK Biobank Database (325), enabling extensive research into its potential utility.

The pulse rise time is the time from pulse onset to systolic peak. It has been found to be increased in subjects with peripheral arterial disease compared with healthy subjects (326), and to be increased in patients with hypertension and arteriosclerosis (327). The pulse rise time could have utility for identifying signs of peripheral arterial disease, particularly when measured at the toe.

# 4. RESEARCH DIRECTIONS

This section provides directions for future research to realize the potential of modeling and analysis of PW signals for vascular age assessment in the clinic and daily life.

#### 4.1. New Generation of Cardiovascular Models

Current state-of-the-art 1-D/0-D models typically provide PWs in steady-state, supine conditions over a period of seconds. However, to unlock the full potential of reduced-order modeling for vascular age studies, future 1-D/0-D model formulations should describe the hemodynamic effects on

arterial PWs of 1) respiration, e.g., by including intrathoracic pressure as an extra variable affecting functional vessel stiffness and ventricular preload; 2) physiological regulation by using feedback loops that dynamically adapt relevant model parameters and boundary conditions; and 3) gravity/fluid shifts by adding source terms to the governing equations, e.g., a gravity term to the conservation of linear momentum equation. These improvements will generate beat-to-beat variations under a wide range of dynamic, transient hemodynamic conditions (e.g., horizontal rest, postural changes, mental stress, exercise, sleep) over minutes and hours. Longer-term dynamic aspects of PWs from birth to old age, including sex-specific growth patterns, adiposity gain, and CV disease progression should also be formulated and coupled to the state-of-the-art 1-D/0-D governing equations. As a result, arterial PW models could be used to simulate PW signals with growth and aging for both sexes, over a time span of years.

Arterial stiffness, which varies along the arterial network and with aging, sex, and disease, has a considerable influence on simulated pulse waveforms. It is, therefore, a key physiological parameter in 1-D models. Future models should incorporate the latest knowledge on the mechanobiological homeostasis of the arteries' constituents (elastin, collagen, smooth muscle cells, proteoglycans) and their evolution from (pre-)birth to adulthood and throughout adulthood. They should therefore go beyond current approaches requiring detailed knowledge of arterial stiffness across the network, impeding model personalization. This new approach will also allow accounting for the impact of metabolic disorders, inflammation, or other factors (e.g., hormones, genetics), and processes on the mechanobiology of blood vessels and their material properties.

Current state-of-the-art 1-D/0-D models are deterministic. In the future, nondeterministic models should be created to account for biological variability and uncertainty in the input parameters of the models (e.g., due to measurement errors). Bayesian methods and Gaussian process regression can be used to quantify how uncertainty translates into variability in model-generated PWs (328, 329) and their predicted evolution during growth and aging. Measurement noise (81) and artifacts should also be considered to make simulated signals more realistic.

#### 4.2. Unleashing the Potential of In Silico Data

The data sets of in silico PWs described in Section 2.1.2 offer a novel and cost-effective approach for the development and preclinical testing of PW analysis algorithms across a wide range of CV conditions, in a relatively quick and inexpensive manner. Current in silico PWs have allowed us to 1) understand the physical mechanisms underlying observations from real populations and 2) train and test machine learning-based PW analysis algorithms; e.g., for aneurysm (81, 120) and stenosis (120) detection, arterial stiffness calculation (28, 105), and cardiac elastance assessment (330).

So far, machine learning models trained using in silico data have been tested using in silico data only. In the future, combination of in silico training of algorithms with in vivo testing in real populations could overcome the need to acquire large data sets in vivo. This will require data sets of in silico PWs created using a new generation of CV models (see Section 4.1) that can replicate trajectories of CV growth, remodeling, and aging in children, adolescents, and adults, for both sexes and in a wide variety of physiological and pathological conditions, including early vascular aging (4).

#### 4.3. Al-Based Algorithms and Digital Twins

We envision PW analysis algorithms based on AI for vascular age assessment from PW signals acquired under varying physiological conditions in daily life, from infants to adults. AI-based algorithms could be constructed using in silico data (see Section 4.2) to assess arterial stiffness from basic clinical data (age, sex, body height/weight) and PW signals acquired by noninvasive wearable devices (e.g., the PPG signal). We also envision digital twins of the CV system capable of predicting an individual's CV aging trajectory. This could lead to an early assessment of vascular age for patient stratification. Data assimilation and AI techniques will allow the new generation of CV models (see Section 4.1) to be used as digital twins for personalized diagnosis, prognosis, and therapy. Current state-of-the-art models require detailed anatomical and physiological data sets to estimate model parameters, yet currently rely on only a few anatomical data sets of "representative" adult males. This unworkable and biased approach should be abandoned. Instead, generation of digital twins will require data assimilation algorithms and morphing/scaling methods to generate an individual's changing arterial network throughout life, matching body size for males/females. Existing imaging and deep phenotyping data [e.g., UK Biobank (325)] could be used for this purpose. UK Biobank also contains PW data together with epidemiological, demographic, and genomics data, which enables investigation of the genome-wide associations of PW signals, their prognostic value for incident CV disease (331), and their use in Mendelian randomization studies (332).

#### 4.4. Clinical Perspectives

Ultimately, AI-based algorithms and digital twins should be tested using longitudinal studies in large populations, such as UK Biobank (331), Framingham (333), or Asklepios (334). These methods could be calibrated to the individual's clinical and measured data at study onset. Follow-up data could then be used to assess how well the effective evolution of the subject's CV system and the aging process of their arteries matches model predictions and, hence, facilitates the early identification of at-risk citizens. This should be compared with current practice where CV disease screening mostly requires direct contact with patients. Application of these digital solutions in children and adolescents offers the prospect to detect adverse CV trajectories with accelerated stiffening of arteries, elevated blood pressures, and concomitant cardiac problems (ventricular hypertrophy, heart failure). Preventive measures could then be targeted at high-risk individuals to protect the CV system from prolonged insults and accumulating damage that manifest as an increased CV risk in later life. A combination of population level- and high-risk-targeted prevention would represent a cost-efficient solution with a high societal impact.



#### 5. CONCLUSIONS

This review has shown that modeling and analysis of arterial PWs play a key role in vascular age studies, in the clinic and in daily life. The following main conclusions arise from our review:

- 1) Blood pressure, blood flow velocity, blood flow rate, arterial distension, and PPG PW signals contain a wealth of information suitable for vascular age assessment and identification of individuals at elevated CV risk.
- 2) PW signals can be measured by a variety of invasive and noninvasive devices, including wearable technologies. Their characteristics and morphologies, which differ between different signal types, measurement sites, and ages need to be considered when measuring, preprocessing, and analyzing PW signals.
- Further research is needed to identify the most accurate PW analysis method considering the characteristics and morphology of the available input PW signal/s. Modeling can facilitate this research by providing reference data sets of in silico PWs to benchmark PW analysis methods and thereby identify reliable methods that are ready for implementation in real subjects.
- 4) Physics-based, reduced-order 1-D and 0-D models can simulate PWs in large arteries, often in steady state, in supine conditions, and over one cardiac cycle, with a reasonable computational cost and with accuracies comparable with those obtained by 3-D models. However, to unlock the full potential of reduced-order modeling for vascular age studies, models should simulate PW signals under a wide range of dynamic hemodynamic conditions, accounting for uncertainty in the input parameters, biological variability, and long-term mechanobiological processes related to growth, aging, sex, and disease, from birth to old age.
- 5) We envision AI-based algorithms and digital twins capable of predicting an individual's CV aging trajectory through model-based, automated interpretation of PW signals, from frequent recordings by noninvasive, wearable technologies, throughout life. The development of these tools will require a combination of in silico and in vivo PW data, to overcome the need to acquire large data sets in vivo.

#### SUPPLEMENTAL DATA

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#### DISCLOSURES

No conflicts of interest, financial or otherwise, are declared by the authors.

#### AUTHOR CONTRIBUTIONS

J.A. and B.E.W. conceived and designed research; J.A., P.H.C., B.P., and B.E.W. prepared figures; J.A., P.H.C., V.B., B.P., B.H., A.D.H., P.S., and B.E.W., drafted manuscript; J.A., P.H.C., V.B., B.P., B.H., R.M.B., M.P.M., S.V., S.P., A.W.K., A.G., C.C.M., J.M., A.D.H., P.S., and B.E.W., edited and revised manuscript; J.A., P.H.C., V.B., B.P., B.H., R.M.B., M.P.M., S.V., S.P., A.W.K., A.G., C.C.M., J.M., A.D.H., P.S., and B.E.W. approved final version of manuscript.

#### REFERENCES

- Olsen MH, Angell SY, Asma S, Boutouyrie P, Burger D, Chirinos JA, Damasceno A, Delles C, Gimenez-Roqueplo A-P, Hering D, López-Jaramillo P, Martinez F, Perkovic V, Rietzschel ER, Schillaci G. Schutte AE. Scuteri A. Sharman JE. Wachtell K. Wang JG. A call to action and a lifecourse strategy to address the global burden of raised blood pressure on current and future generations: the Lancet Commission on hypertension. Lancet 388: 2665–2712, 2016. doi:10.1016/S0140-6736(16)31134-5.
- 2. Climie RE, Bruno RM, Hametner B, Mayer CC, Terentes-Printzios D. Vascular age is not only atherosclerosis, it is also arteriosclerosis. J Am Coll Cardiol 76: 229-230, 2020. doi:10.1016/j.jacc.2020.03.081.
- Climie RE, Alastruey J, Mayer CC, Schwarz A, Laucyte-Cibulskiene A Voicehovska J Bianchini E Bruno R-M Charlton PH Grillo A Guala A, Hallab M, Hametner B, Jankowski P, Königstein K, Lebedeva A, Mozos I, Pucci G, Puzantian H, Terentes-Printzios D, Yetik-Anacak G, Park C, Nilsson PM, Weber T. Vascular ageingmoving from bench towards bedside. Eur J Prev Cardiol. 2023 May 08 [Epub ahead of print]. doi:10.1093/eurjpc/zwad028.
- Nilsson PM. Early vascular aging (EVA): consequences and prevention. Vasc Health Risk Manag 4: 547-552, 2008. doi:10.2147/vhrm.
- Broomé M, Maksuti E, Bjällmark A, Frenckner B, Janerot-Sjöberg B. Closed-loop real-time simulation model of hemodynamics and oxygen transport in the cardiovascular system. Biomed Eng Online 12: 69, 2013. doi:10.1186/1475-925X-12-69.
- Euler L. Principia pro motu sanguinis per arterias determinando. Opera Posthuma Mathematica Et Physica Anno 1844 Detecta, edited by Fuss PH, Petropoli NF, Apund Eggers et Socios, vol. 2, 1775, p. 814-823.
- Parker KH. A brief history of arterial wave mechanics. Med Biol Eng Comput 47: 111-118, 2009. doi:10.1007/s11517-009-0440-5.
- Westerhof N. Analog Studies of Human Systemic Arterial Hemodynamics (PhD thesis). Philadelphia, Pennsylvania: University of Pennsylvania, 1968.
- Hughes TJR. A Study of the One-Dimensional Theory of Arterial Pulse Propagation (PhD thesis Report 74-13). University of California Berkeley, Structural Engineering Laboratory, 1974.
- 10. Hughes TJR, Lubliner J. On the one-dimensional theory of blood flow in the larger vessels. Math Biosci 18: 161-170, 1973. doi:10.1016/ 0025-5564(73)90027-8.
- van de Vosse FN, Stergiopulos N. Pulse wave propagation in the arterial tree. Annu Rev Fluid Mech 43: 467-499, 2011. doi:10.1146/ annurev-fluid-122109-160730.
- Barnard ACL, Hunt WA, Timlake WP, Varley E. A theory of fluid flow in compliant tubes. Biophys J 6: 717-724, 1966. doi:10.1016/ S0006-3495(66)86690-0.
- Caro CG, Pedley TJ, Schroter RC, Seed WA. The Mechanics of the Circulation. Oxford: Oxford University Press, 1978.
- Robertson AM, Zakaria H. One-dimensional non-Newtonian models for arterial flows. American Physical Society, 57th Annual Meeting of the Division of Fluid Dynamics, Seattle, Washington, November 21-23, 2004.

- Zamir M. The Physics of Pulsatile Flow. Biological and Medical Physics Series. New York: Springer-Verlag, 2000.
- Armentano R, Barra J, Levenson J, Simon A, Pichel RH. Arterial wall mechanics in conscious dogs: assessment of viscous, inertial, and elastic moduli to characterize aortic wall behavior. Circ Res 76: 468-478, 1995. doi:10.1161/01.res.76.3.468.
- 17. Devault K, Gremaud P, Novak V, Olufsen M, Vernières G, Zhao P. Blood flow in the circle of Willis: modeling and calibration. Multiscale Model Simul 7: 888-909, 2008. doi:10.1137/07070231X.
- Bessems D, Giannopapa CG, Rutten MCM, van de Vosse FN. Experimental validation of a time-domain based wave propagation model of blood flow in viscoelastic vessels. J Biomech 41: 284-291, 2008. doi:10.1016/j.jbiomech.2007.09.014.
- Valdez-Jasso D, Haider MA, Banks HT, Santana DB, Zócalo Germán YZ, Armentano RL, Olufsen MS. Analysis of viscoelastic wall properties in ovine arteries. IEEE Trans Biomed Eng 56: 210-219, 2009. doi:10.1109/TBME.2008.2003093.
- 20. Armentano R, Megnien JL, Simon A, Bellenfant F, Barra J, Levenson J. Effects of hypertension on viscoelasticity of carotid and femoral arteries in humans. Hypertension 26: 48-54, 1995. doi:10.1161/01.hyp.26.1.48.
- 21. Reymond P, Merenda F, Perren F, Rüfenacht D, Stergiopulos N. Validation of a one-dimensional model of the systemic arterial tree. Am J Physiol Heart Circ Physiol 297: H208-H222, 2009. doi:10.1152/ ajpheart.00037.2009.
- 22. Valdez-Jasso D, Bia D, Zócalo Y, Armentano RL, Haider MA, Olufsen MS. Linear and nonlinear viscoelastic modeling of aorta and carotid pressure-area dynamics under in vivo and ex vivo conditions. Ann Biomed Eng 39: 1438-1456, 2011. doi:10.1007/s10439-010-0236-7.
- 23. Formaggia L, Lamponi D, Quarteroni A. One-dimensional models for blood flow in arteries. J Eng Math 47: 251-276, 2003. doi:10.1023/B:ENGI.0000007980.01347.29.
- 24. Alastruey J, Parker KH, Sherwin SJ. Arterial pulse wave haemodynamics. In: 11th International Conference on Pressure Surges, edited by Anderson S. Lisbon: Virtual PiE Led t/a BHR Group, 2012, p. 401-442.
- 25. Mynard JP, Valen-Sendstad K. A unified method for estimating pressure losses at vascular junctions. Int J Numer Method Biomed Eng 31: e02717, 2015. doi:10.1002/cnm.2717.
- 26. Blanco PJ, Watanabe SM, Passos MARF, Lemos PA, Feijóo RA. An anatomically detailed arterial network model for one-dimensional computational hemodynamics. IEEE Trans Biomed Eng 62: 736-753, 2015. doi:10.1109/TBME.2014.2364522.
- 27. Qureshi MU, Vaughan GA, Sainsbury C, Johnson M, Peskin Olufsen CM, Hill NA. Numerical simulation of blood flow and pressure drop in the pulmonary arterial and venous circulation. Biomech Model Mechanobiol 13: 1137-1154, 2014. doi:10.1007/s10237-014-
- 28. Charlton P, Mariscal-Harana J, Vennin S, Li Y, Chowienczyk P, Alastruey J. Modeling arterial pulse waves in healthy aging: a database for in silico evaluation of hemodynamics and pulse wave indexes. Am J Physiol Heart Circ Physiol 317: H1062-H1085, 2019. doi:10.1152/ajpheart.00218.2019.
- 29. Stergiopulos N, Young DF, Rogge TR. Computer simulation of arterial flow with applications to arterial and aortic stenoses. J Biomech 25: 1477-1488, 1992. doi:10.1016/0021-9290(92)90060-e.
- 30. Formaggia L, Lamponi D, Tuveri M, Veneziani A. Numerical modeling of 1D arterial networks coupled with a lumped parameters description of the heart. Comput Methods Biomech Biomed Eng 9: 273-288, 2006. doi:10.1080/10255840600857767.
- Müller LO, Toro EF. A global multiscale mathematical model for the human circulation with emphasis on the venous system. Int J Numer Method Biomed Eng 30: 681-725, 2014. doi:10.1002/cnm.2622.
- 32. Mynard JP, Nithiarasu P. A 1D arterial blood flow model incorporating ventricular pressure, aortic valve and regional coronary flow using the locally conservative Galerkin (LCG) method. Commun Numer Method Eng 24: 367-417, 2008. doi:10.1002/cnm.1117.
- 33. Olufsen MS. Structured tree outflow condition for blood flow in larger systemic arteries. Am J Physiol Heart Circ Physiol 276: H257-H268, 1999. doi:10.1152/ajpheart.1999.276.1.H257.
- 34. Azer K, Peskin CS. A one-dimensional model of blood flow in arteries with friction and convection based on the Womersley

- velocity profile. Cardiovasc Eng 7: 51-73, 2007. doi:10.1007/s10558-
- 35. Huo Y, Kassab GS. A hybrid one-dimensional/Womersley model of pulsatile blood flow in the entire coronary arterial tree. Am J Physiol Heart Circ Physiol 292: H2623-H2633, 2007. doi:10.1152/ajpheart. 00987.2006.
- 36. Olufsen MS, Hill NA, Vaughan GDA, Sainsbury C, Johnson M. Rarefaction and blood pressure in systemic and pulmonary arteries. J Fluid Mech 705: 280–305, 2012. doi:10.1017/jfm.2012.220.
- Quarteroni A, Ragni S, Veneziani A. Coupling between lumped and distributed models for blood flow problems. Comput Visual Sci 4: 111-124, 2001. doi:10.1007/s007910100063.
- 38. Liang F, Takagi S, Himeno R, Liu H. Multi-scale modeling of the human cardiovascular system with applications to aortic valvular and arterial stenoses. Med Biol Eng Comput 47: 743-755, 2009. doi:10.1007/s11517-009-0449-9.
- 39. Mynard JP, Smolich JJ. One-dimensional haemodynamic modeling and wave dynamics in the entire adult circulation. Ann Biomed Eng 43: 1443-1460, 2015 [Erratum in Ann Biomed Eng 44: 1324, 2016]. doi:10.1007/s10439-015-1313-8.
- 40. Blanco P, Feijóo R. A dimensionally-heterogeneous closed-loop model for the cardiovascular system and its applications. Med Eng Phys 35: 652-667, 2013. doi:10.1016/j.medengphy.2012.07.011.
- Gallo C, Ridolfi L, Scarsoglio S. Cardiovascular deconditioning during long-term spaceflight through multiscale modeling. NPJ Microgravity 6: 27, 2020. doi:10.1038/s41526-020-00117-5.
- Streeter VL, Keitzer WF, Bohr DF. Pulsatile pressure and flow distensible vessels. Circ Res 13: 3-20, 1963. doi:10.1161/01.res.13.1.3.
- Schaaf BW, Abbrecht PH. Digital computer simulation of human systemic arterial pulse wave transmission: a nonlinear model. J Biomech 5: 345-364, 1972. doi:10.1016/0021-9290(72)90064-4.
- 44. Wemple RR, Mockros LF. Pressure and flow in the systemic arterial system. J Biomech 5: 629-641, 1972. doi:10.1016/0021-9290(72) 90035-8.
- Raines JK, Jaffrin MY, Shapiro AH. A computer simulation of the human arterial system. Proceedings of the Summer Computer Simulation Conf. (SCSC), Boston, 1971.
- 46. Avolio AP. Multi-branched model of the human arterial system. Med Biol Ena Comput 18: 709-718, 1980, doi:10.1007/BF02441895.
- Stettler JC, Niederer P, Anliker M. Theoretical analysis of arterial hemodynamics including the influence of bifurcations. I. Mathematical model and prediction of normal pulse patterns. Ann Biomed Eng 9: 145-164, 1981. doi:10.1007/BF02363533.
- Stettler JC, Niederer P, Anliker M, Casty M. Theoretical analysis of arterial hemodynamics including the influence of bifurcations. II. Critical evaluation of theoretical model and comparison with noninvasive measurements of flow patterns in normal and pathological cases. Ann Biomed Eng 9: 165-175, 1981. doi:10.1007/BF02363534.
- 49. Kufahl RH, Clark ME. A circle of Willis simulation using distensible vessels and pulsatile flow. J Biomech Eng 107: 112-122, 1985. doi:10.1115/1.3138531.
- Hillen B, Hoogstraten HW, Post L. A mathematical model of the flow in the circle of Willis. J Biomech 19: 187-194, 1986. doi:10.1016/ 0021-9290(86)90151-x.
- 51. Papapanayotou CJ, Cherruault Y, De La Rochefoucauld B. A mathematical model of the circle of Willis in the presence of an arteriovenous anomaly. Computer Math Appl 20: 199-206, 1990. doi:10.1016/ 0898-1221(90)90327-G.
- 52. Fitchett DH. LV-arterial coupling: interactive model to predict effect of wave reflections on LV energetics. Am J Physiol Heart Circ Physiol 261: H1026-H1033, 1991. doi:10.1152/ajpheart.1991.261.4.H1026.
- 53. Cassot F, Zagzoule M, Marc-Vergnes JP. Hemodynamic role of the circle of Willis in stenoses of internal carotid arteries. An analytical solution of a linear model. J Biomech 33: 395-405, 2000. doi:10.1016/s0021-9290(99)00193-1.
- 54. Wan J, Steele B, Spicer SA, Strohband S, Feijóo GR, Hughes TJR, Taylor CA. A one-dimensional finite element method for simulationbased medical planning for cardiovascular disease. Comput Methods Biomech Biomed Eng 5: 195-206, 2002. doi:10.1080/ 10255840290010670.
- 55. Sherwin SJ, Franke VE, Peiró J, Parker KH. One-dimensional modelling of a vascular network in spacetime variables. J Eng Maths 47: 217-250, 2003. doi:10.1023/B:ENGI.0000007979.32871.e2.

- 56. Wang JJ, Parker KH. Wave propagation in a model of the arterial circulation. J Biomech 37: 457-470, 2004. doi:10.1016/j.jbiomech. 2003 09 007
- 57. Bessems D, Rutten M, van de Vosse F. A wave propagation model of blood flow in large vessels using an approximate velocity profile function. J Fluid Mech 580: 145-168, 2007. doi:10.1017/ S0022112007005344.
- 58. Acosta S, Puelz C, Rivière B, Penny DJ, Brady KM, Rusin CG. Cardiovascular mechanics in the early stages of pulmonary hypertension: a computational study. Biomech Model Mechanobiol 16: 2093-2112, 2017. doi:10.1007/s10237-017-0940-4.
- 59. Carson J, Lewis M, Rassi D, van Loon R. A data-driven model to study utero-ovarian blood flow physiology during pregnancy. Biomech Model Mechanobiol 18: 1155-1176, 2019. doi:10.1007/ s10237-019-01135-3.
- 60. Westerhof BE, van Gemert MJC, van den Wijngaard JP. Pressure and flow relations in the systemic arterial tree throughout development from newborn to adult. Front Pediatr 8: 251, 2020. doi:10.3389/ fped.2020.00251.
- Charlton PH, Kyriaco PA, Mant J, Marozas V, Chowienczyk P, Alastruey J. Wearable photoplethysmography for cardiovascular monitoring. Proc IEEE Inst Electr Electron Eng 110: 355-381, 2022. doi:10.1109/JPROC.2022.3149785.
- 62. Alastruey J, Nagel SR, Nier B, Hunt AAE, Weinberg PD, Peiró J. Modelling pulse wave propagation in the rabbit systemic circulation to assess the effects of altered nitric oxide synthesis. J Biomech 42: 2116-2123, 2009. doi:10.1016/j.jbiomech.2009.05.028.
- 63. Charlton P, Celka P, Farukh B, Chowienczyk P, Alastruey J. Assessing mental stress from the photoplethysmogram: a numerical study. Physiol Meas 39: 054001, 2018. doi:10.1088/1361-6579/
- 64. Reymond P, Bohraus Y, Perren F, Lazeyras F, Stergiopulos N. Validation of a patient-specific one-dimensional model of the systemic arterial tree. Am J Physiol Heart Circ Physiol 301: H1173-H1182, 2011. doi:10.1152/ajpheart.00821.2010.
- 65. Guala A, Camporeale C, Tosello F, Canuto C, Ridolfi L. Modelling and subject-specific validation of the heart-arterial tree system. Ann Biomed Eng 43: 222-237, 2015. doi:10.1007/s10439-014-1163-9.
- 66. Tosello F. Guala A. Leone D. Camporeale C. Bruno G. Ridolfi L. Veglio F, Milan A. Central pressure appraisal: clinical validation of a subjectspecific mathematical model. PLoS One 11: e0151523, 2016 [Erratum in PLoS One 11: e0157117, 2016]. doi:10.1371/journal.pone.0151523.
- 67. Guala A, Camporeale C, Ridolfi L. Compensatory effect between aortic stiffening and remodelling during ageing. PLoS One 10: e0139211, 2015. doi:10.1371/journal.pone.0139211.
- 68. Guala A, Scalseggi M, Ridolfi L. Coronary fluid mechanics in an ageing cardiovascular system. Meccanica 52: 503-514, 2017. doi:10.1007/ s11012-015-0283-0.
- 69. Coccarelli A, Hasan HM, Carson J, Parthimos D, Nithiarasu P. Influence of ageing on human body blood flow and heat transfer: a detailed computational modelling study. Int J Numer Method Biomed Eng 34: e3120, 2018. doi:10.1002/cnm.3120.
- 70. Maksuti E, Westerhof N, Westerhof BE, Broom'e M, Stergiopulos N. Contribution of the arterial system and the heart to blood pressure during normal aging—a simulation study. PLoS One 11: e0157493, 2016. doi:10.1371/journal.pone.0157493.
- 71. Yashin Al, Akushevich IV, Arbeev KG, Akushevich L, Ukraintseva SV, Kulminski A. Insights on aging and exceptional longevity from longitudinal data: novel findings from the Framingham Heart Study. Age (Dordr) 28: 363-374, 2006. doi:10.1007/s11357-006-9023-7.
- Echocardiographic Normal Ranges Meta-Analysis of the Left Heart Collaboration. Ethnic-specific normative reference values for echocardiographic LA and LV size, LV mass, and systolic function: the EchoNoRMAL Study. JACC Cardiovasc Imaging 8: 656-665, 2015. doi:10.1016/j.jcmg.2015.02.014.
- 73. Le TT, Tan RS, De Deyn M, Goh EPC, Han Y, Leong BR, Cook SA, Chin CW-L. Cardiovascular magnetic resonance reference ranges for the heart and aorta in Chinese at 3T. J Cardiovasc Maan Reson 18: 21, 2016. doi:10.1186/s12968-016-0236-3.
- 74. Gerstenblith G, Frederiksen J, Yin FC, Fortuin NJ, Lakatta EG, Weisfeldt ML. Echocardiographic assessment of a normal adult aging population. Circulation 56: 273-278, 1977. doi:10.1161/01. cir.56.2.273.

- 75. Redfield MM, Jacobsen SJ, Borlaug BA, Rodeheffer RJ, Kass DA. Age- and gender-related ventricular-vascular stiffening: a community-based study. Circulation 112: 2254-2262, 2005. doi:10.1161/ CIRCULATIONAHA.105.541078.
- 76. Hickson SS, Butlin M, Graves M, Taviani V, Avolio AP, McEniery CM, Wilkinson IB. The relationship of age with regional aortic stiffness and diameter. JACC Cardiovasc Imaging 3: 1247-1255, 2010. doi:10.1016/j.jcmg.2010.09.016.
- 77. Hansen F, Mangell P, Sonesson B, Länne T. Diameter and compliance in the human common carotid artery-variations with age and sex. Ultrasound Med Biol 21: 1-9, 1995. doi:10.1016/0301-5629(94) 00090-5
- 78. The Reference Values for Arterial Stiffness' Collaboration. Determinants of pulse wave velocity in healthy people and in the presence of cardiovascular risk factors: 'establishing normal and reference values'. Eur Heart J 31: 2338-2350, 2010. doi:10.1093/ eurheartj/ehq165.
- McVeigh GE, Bratteli CW, Morgan DJ, Alinder CM, Glasser SP, Finkelstein SM, Cohn D. JN. Agerelated abnormalities in arterial compliance identified by pressure pulse contour analysis: aging and arterial compliance. Hypertension 33: 1392-1398, 1999. doi:10.1161/ 01.hvp.33.6.1392.
- 80. Guala A, Leone D, Milan A, Ridolfi L. In silico analysis of the anti-hypertensive drugs impact on myocardial oxygen balance. Biomech Model Mechanobiol 16: 1035-1047, 2017. doi:10.1007/s10237-017-
- Wang T, Jin W, Liang F, Alastruey J. Machine learning-based pulse wave analysis for early detection of abdominal aortic aneurysms using in silico pulse waves. Symmetry 13: 804, 2021. doi:10.3390/ sym13050804.
- Swillens A, Lanoye L, De Backer J, Stergiopulos N, Verdonck PR, Vermassen F, Segers P. Effect of an abdominal aortic aneurysm on wave reflection in the aorta. IEEE Trans Biomed Eng 55: 1602-1611, 2008. doi:10.1109/TBME.2007.913994.
- van den Wijngaard JPHM, Siebes M, Westerhof BE. Comparison of arterial waves derived by classical wave separation and wave intensity analysis in a model of aortic coarctation. Med Biol Eng Comput 47: 211-220, 2009. doi:10.1007/s11517-008-0387-y.
- 84. Strocchi M, Contarino C, Zhang Q, Bonmassari R, Toro EF. A global mathematical model for the simulation of stenoses and bypass placement in the human arterial system. Appl Math Comput 300: 21-39, 2017. doi:10.1016/j.amc.2016.11.028.
- Willemet M, Chowienczyk P, Alastruey J. A database of virtual healthy subjects to assess the accuracy of foot-to-foot pulse wave velocities for estimation of aortic stiffness. Am J Physiol Heart Circ Physiol 309: H663-H675, 2015. doi:10.1152/ajpheart.00175.2015.
- 86. Charlton PH. Pulse Wave Database (PWDB): a database of arterial pulse waves representative of healthy adults, 2020. https:// peterhcharlton.github.jo/pwdb/pwdb.html.
- Reymond P, Crosetto P, Deparis S, Quarteroni A, Stergiopulos N. Physiological simulation of blood flow in the aorta: comparison of hemodynamic indices as predicted by 3-D FSI, 3-D rigid wall and 1-D models. Med Eng Phys 35: 784-791, 2013. doi:10.1016/j.medengphy.
- 88. Grinberg L, Cheever E, Anor T, Madsen JR, Karniadakis GE. Modeling blood flow circulation in intracranial arterial networks: a comparative 3D/1D simulation study. Ann Biomed Eng 39: 297-309, 2011. doi:10.1007/s10439-010-0132-1.
- 89. Xiao N, Alastruey J, Figueroa CA. A systematic comparison between 1-D and 3-D hemodynamics in compliant arterial models. Int J Numer Method Biomed Eng 30: 204-231, 2014. doi:10.1002/ cnm.2598.
- 90. Boileau E, Nithiarasu P, Blanco PJ, MüLler LO, Fossan FE, Hellevik LR, Donders WP, Huberts W, Willemet M, Alastruey J. A benchmark study of numerical schemes for one-dimensional arterial blood flow modelling. Int J Numer Method Biomed Eng 31: e02732, 2015. doi:10.1002/cnm.2732.
- Alastruey J, Xiao N, Fok H, Schaeffter T, Figueroa CA. On the impact of modelling assumptions in multi-scale, subject-specific models of aortic haemodynamics. J R Soc Interface 13: 20160073, 2016. doi:10.1098/rsif.2016.0073.
- 92. Jin W, Alastruey J. Arterial pulse wave propagation across stenoses and aneurysms: assessment of 1-D simulations against 3-D

- simulations and in vitro measurements. J R Soc Interface 18: 20200881, 2021. doi:10.1098/rsif.2020.0881.
- 93. Alastruey J, Khir AW, Matthys KS, Segers P, Sherwin SJ, Verdonck P, Parker KH, Peiró J. Pulse wave propagation in a model human arterial network: assessment of 1-D visco-elastic simulations against in vitro measurements. J Biomech 44: 2250-2258, 2011. doi:10.1016/j. jbiomech.2011.05.041.
- 94. Saito M, Ikenaga Y, Matsukawa M, Watanabe Y, Asada T, Lagr'Ee P-Y. One-dimensional model for propagation of a pressure wave in a model of the human arterial network: comparison of theoretical and experimental results. J Biomech Eng 133: 121005, 2011. doi:10.1115/1.4005472.
- 95. Huberts W, Van Canneyt K, Segers P, Eloot S, Tordoir JHM, Verdonck P, van de Vosse FN, Bosboom EMH. Experimental validation of a pulse wave propagation model for predicting hemodynamics after vascular access surgery. J Biomech 45: 1684-1691, 2012. doi:10.1016/j.jbiomech.2012.03.028.
- 96. Olufsen MS, Peskin CS, Kim WY, Pedersen EM, Nadim A, Larsen J. Numerical simulation and experimental validation of blood flow in arteries with structured-tree outflow conditions. Ann Biomed Eng 28: 1281-1299, 2000. doi:10.1114/1.1326031.
- 97. Willemet M, Lacroix V, Marchandise E. Validation of a 1D patientspecific model of the arterial hemodynamics in bypassed lowerlimbs: simulations against in vivo measurements. Med Eng Phys 35: 1573-1583, 2013. doi:10.1016/j.medengphy.2013.04.012.
- 98. Steele BN, Wan J, Ku JP, Hughes TJR, Taylor CA. In vivo validation of a one-dimensional finite-element method for predicting blood flow in cardiovascular bypass grafts. IEEE Trans Biomed Eng 50: 649-656, 2003. doi:10.1109/TBME.2003.812201.
- 99. Mynard JP, Penny DJ, Smolich JJ. Scalability and in vivo validation of a multiscale numerical model of the left coronary circulation. Am J Physiol Heart Circ Physiol 306: H517-H528, 2014. doi:10.1152/ ajpheart.00603.2013.
- 100. Pagoulatou S, Stergiopulos N. Evolution of aortic pressure during normal ageing: a model-based study. PLoS One 12: e0182173, 2017. doi:10.1371/journal.pone.0182173.
- Kraskov A, Stogbauer H, Grassberger P. Estimating mutual information. Phys Rev E Stat Nonlin Soft Matter Phys 69: 066138, 2004 [Erratum in Phys Rev E Stat Nonlin Soft Matter Phys 83: 019903, 2011]. doi:10.1103/PhysRevE.69.066138.
- 102. Stetson PF. Independent component analysis of pulse oximetry signals. Annual International Conference of the IEEE Engineering in Medicine and Biology Society IEEE Engineering in Medicine and Biology Society Annual Conference 2004. San Francisco, CA, 2006, p. 231-234. doi:10.1109/IEMBS.2004.1403134.
- 103. Yan J, Zhou C, Xia C, Shen Y, Wang Y, Li F, Yan H, Guo R. Mutual information based recognition of pulse signal in TCM. 4th International Conference on Bioinformatics and Biomedical Engineering. Chengdu, China, 2010, p. 1-4.
- Mariscal-Harana J, Charlton PH, Vennin S, Aramburu J, Florkow MC, van Engelen A, Schneider T, de Bliek H, Ruijsink B, Valverde I, Beerbaum P, Grotenhuis H, Charakida M, Chowienczyk P, Sherwin SJ, Alastruey J. Estimating central blood pressure from aortic flow: development and assessment of algorithms. Am J Physiol Heart Circ Physiol 320: H494–H510, 2021. doi:10.1152/ajpheart.00241.2020.
- 105. Willemet M, Vennin S, Alastruey J. Computational assessment of hemodynamics-based diagnostic tools using a database of virtual subjects: application to three case studies. J Biomech 49: 3908-3914, 2016. doi:10.1016/j.jbiomech.2016.11.001.
- Liang FY, Takagi S, Himeno R, Liu H. Biomechanical characterization of ventricular-arterial coupling during aging: a multi-scale model study. J Biomech 42: 692-704, 2009. doi:10.1016/j.jbiomech.2009.01.010.
- 107. Mynard J, Penny DJ, Smolich JJ. Wave intensity amplification and attenuation in non-linear flow: implications for the calculation of local reflection coefficients. J Biomech 41: 3314-3321, 2008. doi:10.1016/j. jbiomech.2008.10.002.
- 108. Heusinkveld MHG, Delhaas T, Lumens J, Huberts W, Spronck B, Hughes AD, Reesink KD. Augmentation index is not a proxy for wave reflection magnitude: mechanistic analysis using a computational model. J Appl Physiol (1985) 127: 491-500, 2019. doi:10.1152/ japplphysiol.00769.2018.
- 109. Reymond P, Westerhof N, Stergiopulos N. Systolic hypertension mechanisms: effect of global and local proximal aorta stiffening on

- pulse pressure. Ann Biomed Eng 40: 742-749, 2012. doi:10.1007/ s10439-011-0443-x.
- Segers P, Rietzschel ER, De Buyzere ML, Stergiopulos N, Westerhof N, Van Bortel LM, Gillebert T, Verdonck PR. Three- and four-element Windkessel models: assessment of their fitting performance in a large cohort of healthy middle-aged individuals. Proc Inst Mech Eng H 222: 417-428, 2008. doi:10.1243/09544119JEIM287.
- Stergiopulos N, Segers P, Westerhof N. Use of pulse pressure method for estimating total arterial compliance in vivo. Am J Physiol Heart Circ Physiol 276: H424-H428, 1999. doi:10.1152/ajpheart.1999.276.2.H424.
- Obeid H, Soulat G, Mousseaux E, Laurent S, Stergiopulos N, Boutouyrie P, Segers P. Numerical assessment and comparison of pulse wave velocity methods aiming at measuring aortic stiffness. Physiol Meas 38: 1953–1967, 2017. doi:10.1088/1361-6579/aa905a.
- Gaddum NR, Alastruey J, Beerbaum P, Chowienczyk P, Schaeffter T. A technical assessment of pulse wave velocity algorithms applied to non-invasive arterial waveforms. Ann Biomed Eng 41: 2617-2629, 2013. doi:10.1007/s10439-013-0854-v.
- Vardoulis O, Papaioannou TG, Stergiopulos N. Validation of a novel and existing algorithms for the estimation of pulse transit time: advancing the accuracy in pulse wave velocity measurement. Am J Physiol Heart Circ Physiol 304: H1558-H1567, 2013. doi:10.1152/ ajpheart.00963.2012.
- Papaioannou TG, Vardoulis O, Stergiopulos N. The "systolic volume balance" method for the noninvasive estimation of cardiac output based on pressure wave analysis. Am J Physiol Heart Circ Physiol 302: H2064-H2073, 2012. doi:10.1152/ajpheart.00052.2012.
- Bikia V, Pagoulatou S, Trachet B, Soulis D, Protogerou AD, Papaioannou TG, Stergiopulos N. Noninvasive cardiac output and central systolic pressure from cuff-pressure and pulse wave velocity. IEEE J Biomed Health Inform 24: 1968-1981, 2020. doi:10.1109/ JBHI.2019.2956604.
- Stergiopulos N, Westerhof BE, Westerhof N. Physical basis of pressure transfer from periphery to aorta: a model-based study. Am J Physiol Heart Circ Physiol 274: H1386-H1392, 1998. doi:10.1152/ ajpheart.1998.274.4.H1386.
- Vennin S, Mayer A, Li Y, Fok H, Clapp B, Alastruey J, Chowienczyk P. Non-invasive calculation of the aortic blood pressure waveform from the flow velocity waveform: a proof of concept. Am J Physiol Heart Circ Physiol 309: 969-976, 2015. doi:10.1152/ajpheart.00152.2015.
- Guala A, Camporeale C, Ridolfi L, Mesin L. Non-invasive aortic systolic pressure and pulse wave velocity estimation in a primary care setting: an in silico study. Med Eng Phys 42: 91-98, 2017. doi:10.1016/j.medengphy.2017.02.007.
- 120. Jones G, Parr J, Nithiarasu P, Pant S. Machine learning for detection of stenoses and aneurysms: application in a physiologically realistic virtual patient database. Biomech Model Mechanobiol 20: 2097-2146, 2021. doi:10.1007/s10237-021-01497-7.
- Pagoulatou SZ, Stergiopulos N. Estimating left ventricular elastance from aortic flow waveform, ventricular ejection fraction, and brachial pressure: an in silico study. Ann Biomed Eng 46: 1722-1735, 2018. doi:10.1007/s10439-018-2072-0.
- 122. Kowalski R, Beare R, Willemet M, Alastruey J, Smolich JJ, Cheung MHH, Mynard JP. Robust and practical non-invasive estimation of local arterial wave speed and mean blood velocity waveforms. Physiol Meas 38: 2081–2099, 2017. doi:10.1088/1361-6579/aa8de3.
- Campos Arias D, Stergiopulos N, Rodríguez Moliner T, Segers P. Mapping the site-specific accuracy of loop-based local pulse wave velocity estimation and reflection magnitude: a 1D arterial network model analysis. Physiol Meas 40: 075002, 2019. doi:10.1088/1361-
- Trachet B, Reymond P, Kips J, Swillens A, De Buyzere M, Suys B, Stergiopulos N, Segers P. Numerical validation of a new method to assess aortic pulse wave velocity from a single recording of a brachial artery waveform with an occluding cuff. Ann Biomed Eng 38: 876-888, 2010. doi:10.1007/s10439-010-9945-1.
- 125. Wang J-J, O'Brien AB, Shrive NG, Parker KH, Tyberg JV. Time-domain representation of ventricular-arterial coupling as a Windkessel and wave system. Am J Physiol Heart Circ Physiol 284: H1358-H1368, 2003. doi:10.1152/ajpheart.00175.2002.
- 126. Mynard JP, Penny DJ, Davidson MR, Smolich J. The reservoirwave paradigm introduces error into arterial wave analysis: a computer modelling and in-vivo study. J Hypertens 20: 734–743, 2012. doi:10.1097/HJH.0b013e32834f9793.

- 127. Ghigo AR, Abou Taam S, Wang X, Lagrée PY, Fullana JM. A onedimensional arterial network model for bypass graft assessment. Med Eng Phys 43: 39-47, 2017. doi:10.1016/j.medengphy.2017.02.002.
- 128. Caroli A, Manini S, Antiga L, Passera K, Ene-lordache B, Rota S, Remuzzi G, Bode A, Leermakers J, van de Vosse FN, Vanholder R, Malovrh M, Tordoir J, Remuzzi A; ARCH project Consortium. Validation of a patient-specific hemodynamic computational model for surgical planning of vascular access in hemodialysis patients. Kidney Int 84: 1237-1245, 2013. doi:10.1038/ki.2013.188.
- 129. Vardoulis O, Coppens E, Martin B, Reymond P, Tozzi P, Stergiopulos N. Impact of aortic grafts on arterial pressure: a computational fluid dynamics study. Eur J Vasc Endovasc Surg 42: 704-710, 2011. doi:10.1016/j.ejvs.2011.08.006.
- 130. Westerhof N, Lankhaar J-W, Westerhof BE. The arterial Windkessel. Med Biol Eng Comput 47: 131-141, 2009. doi:10.1007/s11517-008-0359-2
- Stergiopulos N, Westerhof BE, Westerhof N. Total arterial inertance as the fourth element of the Windkessel model. Am J Physiol Heart Circ Physiol 276: H81-H88, 1999. doi:10.1152/ajpheart.1999.276.1.H81.
- 132. Noordergraaf A, Verdouw PD, Boom HBK. The use of an analog computer in a circulation model. Prog Cardiovasc Dis 5: 419-439, 1963. doi:10.1016/s0033-0620(63)80009-2.
- 133. Westerhof N, Bosman F, de Vries CJ, Noordergraaf A. Analog studies of the human systemic arterial tree. J Biomech 2: 121-143, 1969. doi:10.1016/0021-9290(69)90024-4.
- 134. Alastruey J, Parker KH, Peiró J, Sherwin SJ. Analysing the pattern of pulse waves in arterial networks: a time-domain study. J Eng Math 64: 331-351, 2009. doi:10.1007/s10665-009-9275-1.
- 135. Frank O. Die Grundform des arteriellen Pulses. Erste Abhandlung Mathematische Analyse Z Biol. Z Biol 37: 483-526, 1899.
- Westerhof N, Elzinga G, Sipkema P. An artificial arterial system for pumping hearts. J Appl Physiol 31: 776-781, 1971. doi:10.1152/ iappl.1971.31.5.776.
- 137. Westerhof N, O'Rourke MF. Haemodynamic basis for the development of left ventricular failure in systolic hypertension and for its logical therapy. J Hypertens 13: 943-952, 1995. doi:10.1097/00004872-199509000-00002.
- 138. Beneken JEW. A Mathematical Approach to Cardio-Vascular Function: The Uncontrolled Human System (PhD thesis). The Netherlands: Medisch-Fysisch Instituut, T.N.O., University of Utrecht,
- 139. Senzaki H, Chen C-H, Kass DA. Single-beat estimation of end-systolic pressure-volume relation in humans. Circulation 94: 2497-2506, 1996. doi:10.1161/01.cir.94.10.2497.
- 140. Burkhoff D, Mirsky L, Suga H. Assessment of systolic and diastolic ventricular properties via pressure-volume analysis: a guide for clinical, translational, and basic researchers. Am J Physiol Heart Circ Physiol 289: H501-H512, 2005. doi:10.1152/ajpheart.00138.2005.
- Westerhof N, Stergiopulos N, Noble MIM. Snapshots of Hemodynamics: An Aid for Clinical Research and Graduate Education (3rd ed.). New York: Springer, 2019.
- 142. Segers P, Stergiopulos N, Westerhof N. Relation of effective arterial elastance to arterial system properties. Am J Physiol Heart Circ Physiol 282: H1041-H1046, 2002. doi:10.1152/ajpheart.00764.2001.
- 143. Burkhoff D, Sagawa K. Ventricular efficiency predicted by an analytical model. Am J Physiol Regul Integr Comp Physiol 250: R1021-R1027, 1986. doi:10.1152/ajpregu.1986.250.6.R1021.
- 144. Stergiopulos N, Meister JJ, Westerhof N. Determinants of stroke volume and systolic and diastolic aortic pressure. Am J Physiol Heart Circ Physiol 270: H2050-H2059, 1996. doi:10.1152/ajpheart. 1996.270.6.H2050.
- 145. Segers P, Stergiopulos N, Westerhof N. Quantification of the contribution of cardiac and arterial remodeling to hypertension. Hypertension 36: 760-765, 2000. doi:10.1161/01.hyp.36.5.760.
- 146. Vennin S, Li Y, Willemet M, Fok H, Gu H, Charlton P, Alastruey J, Chowienczyk P. Identifying hemodynamic determinants of pulse pressure: a combined numerical and physiological approach. Hypertension 70: 1176-1182, 2017. doi:10.1161/HYPERTENSIONAHA.117.09706.
- 147. Wesseling KH, Jansen JR, Settels JJ, Schreuder JJ. Computation of aortic flow from pressure in humans using a nonlinear, three-element model. J Appl Physiol (1985) 74: 2566-2573, 1993. doi:10.1152/ jappl.1993.74.5.2566.
- 148. Charlton P, Smith J, Camporota L, Beale R, Alastruey J. Optimising the Windkessel model for cardiac output monitoring during changes

- in vascular tone. 2014 36th Annual International Conference of the IEEE Engineering in Medicine and Biology Society. Chicago, IL, 2014. p. 3759-3762.
- 149. Murgo JP, Westerhof N, Giolma JP, Altobelli SA. Aortic input impedance in normal man: relationship to pressure wave forms. Circulation 62: 105-116, 1980. doi:10.1161/01.cir.62.1.105.
- 150. Mitchell GF. Pulse pressure, arterial compliance and cardiovascular morbidity and mortality. Curr Opin Nephrol Hypertens 8: 335-342, 1999. doi:10.1097/00041552-199905000-00010.
- Ursino M. Interaction between carotid baroregulation and the pulsating heart: a mathematical model. Am J Physiol Heart Circ Physiol 275: H1733-H1747, 1998. doi:10.1152/ajpheart.1998.275.5.H1733.
- 152. Segers P, Stergiopulos N, Westerhof N, Wouters P, Kolh P, Verdonck P. Systemic and pulmonary hemodynamics assessed with a lumped-parameter heart-arterial interaction model. J Eng Math 47: 185-199, 2003. doi:10.1023/B:ENGI.0000007975.27377.9c.
- 153. Danielsen M, Ottesen JT. A cardiovascular model. Applied Mathematical Models in Human Physiology, edited by Ottesen JT, Olufsen MS, Larsen JK. Philadelphia: SIAM monographs on mathematical human physiology, 2004, chapt. 6.
- Goodwin JA, van Meurs WL, Sá Couto CD, Beneken JEW, Graves SA. A model for educational simulation of infant cardiovascular physiology. Anesth Analg 99: 1655-1664, 2004. doi:10.1213/01. ANE.0000134797.52793.AF.
- 155. Arts T, Delhaas T, Bovendeerd P, Verbeek X, Prinzen FW. Adaptation to mechanical load determines shape and properties of heart and circulation: the CircAdapt model. Am J Physiol Heart Circ Physiol 288: H1943–H1954, 2005. doi:10.1152/ajpheart.00444.2004.
- 156. Mulder MP, Broomé M, Donker DW, Westerhof BE. Distinct morphologies of arterial waveforms reveal preload-, contractility-, and afterload-deficient hemodynamic instability: an in silico simulation study. Physiol Rep 10: e15242, 2022. doi:10.14814/phy2.15242.
- 157. Suga H. Sagawa K. Shoukas AA. Load independence of the instantaneous pressure-volume ratio of the canine left ventricle and effects of epinephrine and heart rate on the ratio. Circ Res 32: 314-322, 1973. doi:10.1161/01.res.32.3.314.
- Mitchell GF, Parise H, Benjamin EJ, Larson MG, Keyes MJ, Vita JA, Vasan RS, Levy D. Changes in arterial stiffness and wave reflection with advancing age in healthy men and women: the Framingham Heart Study. Hypertension 43: 1239-1245, 2004. doi:10.1161/01. HYP.0000128420.01881.aa.
- Campos-Arias D, De Buyzere ML, Chirinos JA, Rietzschel ER, Segers P. Longitudinal changes of input impedance, pulse wave velocity, and wave reflection in a middle-aged population: the Asklepios study. Hypertension 77: 1154-1165, 2021. doi:10.1161/ HYPERTENSIONAHA.120.16149.
- 160. Chiesa ST, Masi S, Shipley MJ, Ellins EA, Fraser AG, Hughes AD, Patel RS, Khir AW, Halcox JP, Singh-Manoux A, Kivimaki M, Celermajer DS, Deanfield JE. Carotid artery wave intensity in midto late-life predicts cognitive decline: the Whitehall II study. Eur Heart J 40: 2300-2309, 2019. doi:10.1093/eurheartj/ehz189.
- Li Y, Hickson SS, McEniery CM, Wilkinson IB, Khir AW. Stiffening and ventricular-arterial interaction in the ascending aorta using MRI: ageing effects in healthy humans. J Hypertens 37: 347-355, 2019. doi:10.1097/HJH.0000000000001886.
- Borlotti A, Khir AW, Rietzschel ER, De Buyzere ML, Vermeersch S, Segers P. Noninvasive determination of local pulse wave velocity and wave intensity: changes with age and gender in the carotid and femoral arteries of healthy human. J Appl Physiol (1985) 113: 727-735, 2012. doi:10.1152/japplphysiol.00164.2012.
- Bhuva AN, D'Silva A, Torlasco C, Nadarajan N, Jones S, Boubertakh R, Van Zalen J, Scully P, Knott K, Benedetti G, Augusto JB, Bastiaenen R, Lloyd G, Sharma S, Moon JC, Parker KH, Manisty CH, Hughes AD. Non-invasive assessment of ventriculo-arterial coupling using aortic wave intensity analysis combining central blood pressure and phase-contrast cardiovascular magnetic resonance. Eur Heart J Cardiovasc Imaging 21: 805-813, 2020. doi:10.1093/ehjci/jez227.
- McEniery CM, Yasmin , Hall IR, Qasem A, Wilkinson IB, Cockcroft JR; ACCT Investigators. Normal vascular aging: differential effects on wave reflection and aortic pulse wave velocity: the Anglo-Cardiff Collaborative Trial (ACCT). J Am Coll Cardiol 46: 1753-1760, 2005. doi:10.1016/j.jacc.2005.07.037.

- 165. Gardner AW, Parker DE. Association between arterial compliance and age in participants 9 to 77 years old. Angiology 61: 37-41, 2010. doi:10.1177/0003319709339588.
- 166. Waddell TK, Dart AM, Gatzka CD, Cameron JD, Kingwell BA. Women exhibit a greater age-related increase in proximal aortic stiffness than men. J Hypertens 19: 2205-2212, 2001. doi:10.1097/ 00004872-200112000-00014.
- 167. Mazzaro L, Almasi SJ, Shandas R, Seals DR, Gates PE. Aortic input impedance increases with age in healthy men and women. Hypertension 45: 1101-1106, 2005. doi:10.1161/01.HYP.0000164579.
- 168. Segers P, Rietzschel ER, De Buyzere ML, Vermeersch SJ, De Bacquer D, Van Bortel LM, De Backer G, Gillebert TC, Verdonck PR; Asklepios investigators. Noninvasive (input) impedance, pulse wave velocity, and wave reflection in healthy middle-aged men and women. Hypertension 49: 1248-1255, 2007. doi:10.1161/HYPERTENSIONAHA.106.085480.
- Segers P, Mahieu D, Kips J, Rietzschel E, De Buyzere M, De Bacquer D, Bekaert S, De Backer G, Gillebert T, Verdonck P, Van Bortel L; Asklepios investigators. Amplification of the pressure pulse in the upper limb in healthy, middle-aged men and women. Hypertension 54: 414-420, 2009. doi:10.1161/HYPERTENSIONAHA.109.133009.
- 170. Ishida A, Miyagi M, Kinjo K, Ohya Y. Age- and sex-related effects on ankle-brachial index in a screened cohort of Japanese: the Okinawa Peripheral Arterial Disease Study (OPADS). Eur J Prev Cardiol 21: 712-718, 2014. doi:10.1177/2047487312462822.
- Li Y, Wang JG, Dolan E, Gao PJ, Guo HF, Nawrot T, Stanton AV, Zhu DL, O'Brien E, Staessen JA. Ambulatory arterial stiffness index derived from 24-hour ambulatory blood pressure monitoring. Hypertension 47: 359-364, 2006. doi:10.1161/01.HYP.0000200695.34024.4c.
- 172. Choi SY, Oh BH, Bae Park J, Choi DJ, Rhee MY, Park S. Age-associated increase in arterial stiffness measured according to the cardioankle vascular index without blood pressure changes in healthy adults. J Atheroscler Thromb 20: 911–923, 2013. doi:10.5551/jat.18267.
- 173. Hashimoto J, Westerhof BE, Ito S. Carotid flow augmentation, arterial aging, and cerebral white matter hyperintensities. Arterioscler Thromb Vasc Biol 38: 2843-2853, 2018. doi:10.1161/ATVBAHA.118.311873.
- 174. Hashimoto J, Ito S. Pulse pressure amplification, arterial stiffness, and peripheral wave reflection determine pulsatile flow waveform of the femoral artery. Hypertension 56: 926-933, 2010. doi:10.1161/ HYPERTENSIONAHA.110.159368.
- 175. Hashimoto J, Ito S. Aortic stiffness determines diastolic blood flow reversal in the descending thoracic aorta: potential implication for retrograde embolic stroke in hypertension. Hypertension 62: 542-549, 2013. doi:10.1161/HYPERTENSIONAHA.113.01318.
- 176. Takazawa K, Tanaka N, Fujita M, Matsuoka O, Saiki T, Aikawa M, Tamura S, Ibukiyama C. Assessment of vasoactive agents and vascular aging by the second derivative of photoplethysmogram waveform. Hypertension 32: 365-370, 1998. doi:10.1161/01.hyp.32.2.365.
- Hashimoto J, Watabe D, Kimura A, Takahashi H, Ohkubo T, Totsune K, Imai Y. Determinants of the second derivative of the finger photoplethysmogram and brachial-ankle pulse-wave velocity: the Ohasama study. Am J Hypertens 18: 477-485, 2005. doi:10.1016/j. amjhyper.2004.11.009.
- 178. Choi J, Park MG. Variations in the second derivative of a photoplethysmogram with age in healthy Korean adults. Int J Environ Res Public Health 20: 236, 2022. doi:10.3390/ijerph20010236.
- 179. Yamakoshi KI, Shimazu H, Togawa T. Indirect measurement of instantaneous arterial blood pressure in the human finger by the vascular unloading technique. IEEE Trans Biomed Eng 27: 150-155, 1980. doi:10.1109/TBME.1980.326616.
- 180. Ding XR, Zhao N, Yang GZ, Pettigrew RI, Lo B, Miao F, Li Y, Liu J, Zhang YT. Continuous blood pressure measurement from invasive to unobtrusive: celebration of 200th birth anniversary of Carl Ludwig. IEEE J Biomed Health Inform 20: 1455-1465, 2016. doi:10.1109/ JBHI.2016.2620995.
- Bianchini E, Lønnebakken MT, Wohlfahrt P, Piskin S, Terentes-Printzios D, Alastruey J, Guala A. Magnetic resonance imaging and computed tomography for the non-invasive assessment of arterial ageing—a review by the VascAgeNet COST Action. J Am Heart Assoc 12: e027414, 2023 May 15 [Epub ahead of print]. doi:10.1161/ JAHA.122.027414.
- 182. Charlton PH, Paliakaitė B, Pilt K, Bachler M, Zanelli S, Kulin D, Allen J, Hallab M, Bianchini E, Mayer CC, Terentes-Printzios D, Dittrich V, Hametner B, Veerasingam D, Žikić D, Marozas V. Assessing

- hemodynamics from the photoplethysmogram to gain insights into vascular age: a review from VascAgeNet. Am J Physiol Heart Circ Physiol 322: H493-H522, 2022. doi:10.1152/ajpheart.00392.2021.
- 183. Gardner RM. Direct blood pressure measurement—dynamic response requirements. Anesthesiology 54: 227–236, 1981. doi:10.1097/ 00000542-198103000-00010.
- 184. Truijen J, Westerhof BE, Kim YS, Stok WJ, de Mol BA, Preckel B, Hollmann MW, van Lieshout JJ. The effect of haemodynamic and peripheral vascular variability on cardiac output monitoring: thermodilution and non-invasive pulse contour cardiac output during cardiothoracic surgery. Anaesthesia 73: 1489-1499, 2018. doi:10.1111/ anae.14380.
- Zaitsev M, Maclaren J, Herbst M. Motion artifacts in MRI: a complex problem with many partial solutions. J Magn Reson Imaging 42: 887-901, 2015. doi:10.1002/jmri.24850.
- 186. Gill RW. Measurement of blood flow by ultrasound: accuracy and sources of error. Ultrasound Med Biol 11: 625-641, 1985. doi:10.1016/ 0301-5629(85)90035-3.
- 187. Michard F. Changes in arterial pressure during mechanical ventilation. Anesthesiology 103: 419-428, 2005. doi:10.1097/00000542-200508000-00026.
- 188. **Wang D**, **Zhang D**, **Lu G**. A robust signal preprocessing framework for wrist pulse analysis. Biomed Signal Process Control 23: 62-75, 2016. doi:10.1016/j.bspc.2015.08.002.
- Li J-Q, Li R, Chen Z-Z, Deng G-Q, Wang H, Mavromoustakis CX, Song H, Ming Z. Design of a continuous blood pressure measurement system based on pulse wave and ECG signals. IEEE J Transl Eng Health Med 6: 1900114, 2018. doi:10.1109/JTEHM.2017.2788885.
- 190. **Nyquist H.** Certain topics in telegraph transmission theory. *Trans Am* Inst Electr Eng 47: 617–644, 1928. doi:10.1109/T-AIEE.1928.5055024.
- Hughes AD, Park C, Davies J, Francis D, McG Thom SA, Mayet J, Parker KH. Limitations of augmentation index in the assessment of wave reflection in normotensive healthy individuals. PLoS One 8: e59371, 2013. doi:10.1371/journal.pone.0059371.
- 192. Charlton PH. PulseAnalyse: a signal processing tool for cardiovascular pulse waves, 2020.
- 193. Vest AN, Da Poian G, Li Q, Liu C, Nemati S, Shah AJ, Clifford GD. An open source benchmarked toolbox for cardiovascular waveform and interval analysis. Physiol Meas 39: 105004, 2018. doi:10.1088/1361-6579/aae021. https://zenodo.org/record/4106324#.ZF5KGy8w2fU.
- 194. Charlton PH, Kotzen K, Mejía-Mejía E, Aston PJ, Budidha K, Mant J, Pettit C, Behar JA, Kyriacou PA. Detecting beats in the photoplethysmogram: benchmarking open-source algorithms. Physiol Meas 43: 085007, 2022. doi:10.1088/1361-6579/ac826d.
- Gambarotta N, Aletti F, Baselli Ferrario GM. A review of methods for the signal quality assessment to improve reliability of heart rate and blood pressures derived parameters. Med Biol Eng Comput 54: 1025-1035, 2016. doi:10.1007/s11517-016-1453-5.
- Mejia-Mejia E, Allen J, Budidha K, El-Hajj C, Kyriacou PA, Charlton P. Photoplethysmography signal processing and synthesis. In: Photoplethysmography (1st ed.), edited by Kyriacou PA, Allen J. Elsevier, 2021, p. 69-146.
- Charlton PH, Marozas V. Wearable photoplethysmography devices. In: Photoplethysmography (1st ed.), edited by Kyriacou PA, Allen J. Elsevier, 2021, p. 401-439.
- Jedrzejewski D, McFarlane E, Lacy PS, Williams B. Pulse wave calibration and implications for blood pressure measurement: systematic review and meta-analysis. Hypertension 78: 360-371, 2021. doi:10.1161/HYPERTENSIONAHA.120.16817.
- 199. Karamanoglu M, O'Rourke MF, Avolio AP, Kelly RP. An analysis of the relationship between central aortic and peripheral upper limb pressure waves in man. Eur Heart J 14: 160-167, 1993. doi:10.1093/ eurheartj/14.2.160.
- 200. Joly L, Perret-Guillaume C, Kearney-Schwartz A, Salvi P, Mandry D, Marie PY, Karcher G, Rossignol P, Zannad F, Benetos A. Pulse wave velocity assessment by external noninvasive devices and phase-contrast magnetic resonance imaging in the obese. Hypertension 54: 421-426, 2009. doi:10.1161/HYPERTENSIONAHA.109.133645.
- 201. Savitzky A, Golay MJE. Smoothing and differentiation of data by simplified least squares procedures. Anal Chem 36: 1627-1639, 1964. doi:10.1021/ac60214a047.
- 202. Xu L, Zhou S, Wang L, Yao Y, Hao L, Qi L, Yao Y, Han H, Mukkamala R, Greenwald SE. Improving the accuracy and robustness of carotid-femoral pulse wave velocity measurement using a

- simplified tube-load model. Sci Rep 12: 5147, 2022. doi:10.1038/
- 203. Vennin S, Li Y, Mariscal-Harana J, Charlton PH, Fok H, Gu H, Chowienczyk P, Alastruey J. Novel pressure wave separation analysis for cardiovascular function assessment highlights major role of aortic root. IEEE Trans Biomed Eng 69: 1707-1716, 2022. doi:10.1109/ TBMF 2021 3127799
- 204. Parker KH, Jones CJH. Forward and backward running waves in the arteries: analysis using the method of characteristics. J Biomech Eng 112: 322-326, 1990. doi:10.1115/1.2891191.
- 205. Chirinos JA, Kips JG, Jacobs DR, Brumback L, Duprez DA, Kronmal R, Bluemke DA, Townsend RR, Vermeersch S, Segers P. Arterial wave reflections and incident cardiovascular events and heart failure: MESA (Multiethnic Study of Atherosclerosis). J Am Coll Cardiol 60: 2170-2177, 2012. doi:10.1016/j.jacc.2012.07.054.
- 206. Weber T, Wassertheurer S, Rammer M, Haiden A, Hametner B, Eber B. Wave reflections, assessed with a novel method for pulse wave separation, are associated with end-organ damage and clinical outcomes. Hypertension 60: 534-541, 2012. doi:10.1161/HYPERTENSIONAHA.
- 207. Westerhof BE, van den Wijngaard JP, Murgo JP, Westerhof N. Location of a reflection site is elusive: consequences for the calculation of aortic pulse wave velocity. Hypertension 52: 478-483, 2008. doi:10.1161/HYPERTENSIONAHA.108.116525.
- 208. Willemet M, Alastruey J. Arterial pressure and flow wave analysis using time-domain 1-D hemodynamics. Ann Biomed Eng 43: 190-206, 2015. doi:10.1007/s10439-014-1087-4.
- 209. Westerhof N, Segers P, Westerhof BE. Wave separation, wave intensity, the reservoir-wave concept, and the instantaneous wave-free ratio: presumptions and principles. Hypertension 66: 93-98, 2015 [Erratum in Hypertension 66: e21, 2015]. doi:10.1161/HYPERTENSIONAHA.115.
- 210. Mynard JP, Smolich JJ. The case against the reservoir-wave approach. Int J Cardiol 176: 1009-1012, 2014. doi:10.1016/j.ijcard. 2014.07.070.
- Parker KH. An introduction to wave intensity analysis. Med Biol Eng Comput 47: 175-188, 2009. doi:10.1007/s11517-009-0439-y.
- 212. Jones CJH, Sugawara M. "Wavefronts" in the aorta—implications for the mechanisms of left ventricular ejection and aortic valve closure. Cardiovasc Res 27: 1902-1905, 1993. doi:10.1093/cvr/27.11.1902.
- 213. Khir AW, Henein MY, Koh T, Das SK, Parker KH, Gibson DG. Arterial waves in humans during peripheral vascular surgery. Clin Sci 101: 749-757, 2001, doi:10.1042/CS20010046.
- 214. Ohte N, Narita H, Sugawara M, Niki K, Okada T, Harada A, Hayano J, Kimura G. Clinical usefulness of carotid arterial wave intensity in assessing left ventricular systolic and early diastolic performance. Heart Vessels 18: 107-111, 2003. doi:10.1007/s00380-003-0700-5.
- 215. Zambanini A, Cunningham SL, Parker KH, Khir AW, McG. Thom SA, Hughes AD. Wave-energy patterns in carotid, brachial, and radial arteries: a noninvasive approach using wave-intensity analysis. Am J Physiol Heart Circ Physiol 289: H270-H276, 2005. doi:10.1152/ aipheart.00636.2003.
- 216. Avgeropoulou C, Illmann A, Schumm-Draeger P-M, Kallikazaros J, von Bibra H. Assessment of arterioventricular coupling by tissue Doppler and wave intensity in type 2 diabetes. Diabetes Vasc Dis 6: 271-278, 2006. doi:10.1177/14746514060060060401.
- 217. Sugawara M, Niki K, Ohte N, Okada T, Harada A. Clinical usefulness of wave intensity analysis. Med Biol Eng Comput 47: 197-206, 2009. doi:10.1007/s11517-008-0388-x.
- 218. Kolyva C, Pantalos GM, Giridharan GA, Pepper JR, Khir AW. Discerning aortic waves during intra-aortic balloon pumping and their relation to benefits of counterpulsation in humans. J Appl Physiol (1985) 107: 1497-1503, 2009. doi:10.1152/japplphysiol.00413.2009.
- 219. Liu R, Kwok YL, Li Y, Lao TT, Zhang X. The effects of graduated compression stockings on cutaneous surface pressure along the path of main superficial veins of lower limbs. Wounds 18: 150-157, 2006.
- 220. Sun YH, Anderson TA, Parker KH, Tyberg JV. Effects of left ventricular contractility and coronary vascular resistance on coronary dynamics. Am J Physiol Heart Circ Physiol 286: H1590-H1595, 2004. doi:10.1152/aipheart.01100.2001.
- 221. Davies JE, Whinnett ZI, Francis DP, Manisty CH, Aguado-Sierra J, Willson K, Foale RA, Malik IS, Hughes AD, Parker KH, Mayet J. Evidence of a dominant backward-propagating "suction" wave

- responsible for diastolic coronary filling in humans, attenuated in left ventricular hypertrophy. Circulation, 113: 1768-1778, 2006. doi:10.1161/ CIRCULATIONAHA.105.603050.
- 222. Mynard JP, Penny DJ, Smolich JJ. Major influence of a 'smoke and mirrors' effect caused by wave reflection on early diastolic coronary arterial wave intensity. J Physiol 596: 993-1017, 2018. doi:10.1113/
- 223. Penny DJ, Mynard JP, Smolich JJ. Aortic wave intensity analysis of ventricular-vascular interaction during incremental dobutamine infusion in adult sheep. Am J Physiol Heart Circ Physiol 294: H481-H489, 2008. doi:10.1152/ajpheart.00962.2006.
- 224. Rivolo S, Patterson T, Asrress KN, Marber M, Redwood S, Smith NP, Lee J. Accurate and standardized coronary wave intensity analysis. IEEE Trans Biomed Eng 64: 1187-1196, 2017. doi:10.1109/TBME.
- 225. Francis N, Selwanos PP, Yacoub MH, Parker KH. The use of maximum entropy to enhance wave intensity analysis: an application to coronary arteries in hypertrophic obstructive cardiomyopathy. Front Cardiovasc Med 8: 701267, 2021. doi:10.3389/fcvm.2021.701267.
- 226. Mynard JP, Davidson MR, Penny DJ, Smolich JJ. Robustness of the P-U and InD-U loop wave speed estimation methods: effects of the diastolic pressure decay and vessel wall non-linearities. 33rd Annual International Conference of the IEEE EMBS, Boston, MA, 2011. https://researchr.org/publication/embc-2011.
- Alastruey J, Hunt AAE, Weinberg PW. Novel wave intensity analysis of arterial pulse wave propagation accounting for peripheral reflections. Int J Numer Method Biomed Eng 30: 249-279, 2014. doi:10.1002/cnm.2602.
- 228. Mynard JP, Smolich JJ. Wave potential and the one-dimensional Windkessel as a wave-based paradigm of diastolic arterial hemodynamics. Am J Physiol Heart Circ Physiol 307: H307-H318, 2014 [Erratum in Am J Physiol Heart Circ Physiol 308: H151, 2015]. doi:10.1152/aipheart.00293.2014.
- 229. Moens Al. Over de voortplantingssnelheid von den pols (On the Speed of Propagation of the Pulse). Technical report. Leiden, 1877,
- 230. Korteweg DJ. Uber die Fortpflanzungsgeschwindigkeit des Schalles in elastischen Röhern. Ann Phys Chem 241: 525-542, 1878. doi:10.1002/ andp.18782411206.
- 231. Khir AW, O'Brien A, Gibbs JS, Parker KH. Determination of wave speed and wave separation in the arteries. J Biomech 34: 1145-1155, 2001. doi:10.1016/s0021-9290(01)00076-8.
- 232. Feng J, Khir AW. Determination of wave speed and wave separation in the arteries using diameter and velocity. J Biomech 43: 455-462, 2010. doi:10.1016/j.jbiomech.2009.09.046.
- 233. Rabben SI, Baerum S, Sørhus V, Torp H. Ultrasound-based vessel wall tracking: an auto-correlation technique with RF center frequency estimation. Ultrasound Med Biol 28: 507-517, 2002. doi:10.1016/s0301-5629(02)00487-8.
- 234. Alastruey J. Numerical assessment of time-domain methods for the estimation of local arterial pulse wave speed. J Biomech 44: 885-891, 2011. doi:10.1016/j.jbiomech.2010.12.002.
- 235. Davies JE, Whinnett ZI, Francis DP, Willson K, Foale RA, Malik IS, Hughes AD, Parker KH, Mayet J. Use of simultaneous pressure and velocity measurements to estimate arterial wave speed at a single site in humans. Am J Physiol Heart Circ Physiol 290: H878-H885, 2006. doi:10.1152/ajpheart.00751.2005.
- 236. Weber T, Wassertheurer S, Hametner B, Parragh S, Eber B. Noninvasive methods to assess pulse wave velocity: comparison with the invasive gold standard and relationship with organ damage. J Hypertens 33: 1023-1031, 2015. doi:10.1097/HJH.000000000000518.
- Redheuil A, Yu W-C, Wu CO, Mousseaux E, de Cesare A, Yan R, Kachenoura N, Bluemke D, Lima JAC. Reduced ascending aortic strain and distensibility: earliest manifestations of vascular aging in humans. Hypertension 55: 319-326, 2010. doi:10.1161/HYPERTENSIONAHA. 109.141275.
- 238. Laurent S, Cockcroft J, Van Bortel L, Boutouyrie P, Giannattasio C, Hayoz D, Pannier B, Vlachopoulos C, Wilkinson I, Struijker-Boudier H; European Network for Non-invasive Investigation of Large Arteries. Expert consensus document on arterial stiffness: methodological issues and clinical applications. Eur Heart J 27: 2588-2605, 2006. doi:10.1093/eurheartj/ehl254.
- 239. Lu Y, Pechlaner R, Cai J, Yuan H, Huang Z, Yang G, Wang J, Chen Z, Kiechl S, Xu Q. Trajectories of age-related arterial stiffness in

- Chinese men and women. J Am Coll Cardiol 75: 870-880, 2020. doi:10.1016/j.jacc.2019.12.039.
- 240. Bachler M, Mayer C, Hametner B, Wassertheurer S. Increasing stability of real-time pulse wave velocity estimation by combining established and new approaches. 8th EUROSIM Congress on Modelling and Simulation, Cardiff, UK, 2013, p. 47-51.
- 241. Kimoto E, Shoji T, Shinohara K, Inaba M, Okuno Y, Miki T, Koyama H, Emoto M, Nishizawa Y. Preferential stiffening of central over peripheral arteries in type 2 diabetes. Diabetes 52: 448-452, 2003. doi:10.2337/diabetes.52.2.448.
- 242. Bruno RM, Nilsson PM, Engström G, Wadström BN, Empana JP, Boutouyrie P, Laurent S. Early and supernormal vascular aging: clinical characteristics and association with incident cardiovascular events. Hypertension 76: 1616-1624, 2020. doi:10.1161/HYPERTENSIONAHA.
- 243. Weber T, Mayer CC. "Man is as old as his arteries" taken literally: in search of the best metric. Hypertension 76: 1425-1427, 2020. doi:10.1161/HYPERTENSIONAHA.120.16128.
- 244. Nilsson Wadström B, Fatehali AH, Engström G, Nilsson PM. A vascular aging index as independent predictor of cardiovascular events and total mortality in an elderly urban population. Angiology 70: 929-937, 2019. doi:10.1177/0003319719857270.
- 245. Vlachopoulos C, Aznaouridis K, Stefanadis C. Prediction of cardiovascular events and all-cause mortality with arterial stiffness: a systematic review and meta-analysis. J Am Coll Cardiol 55: 1318-1327, 2010. doi:10.1016/j.jacc.2009.10.061.
- 246. Ben-Shlomo Y, Spears M, Boustred C, May M, Anderson SG, Benjamin EJ, Boutouyrie P, Cameron J, Chen CH, Cruickshank JK, Hwang SJ, Lakatta EG, Laurent S, Maldonado J, Mitchell GF, Najjar SS, Newman AB, Ohishi M, Pannier B, Pereira T, Vasan RS, Shokawa T, Sutton-Tyrell K, Verbeke F, Wang KL, Webb DJ, Willum Hansen T, Zoungas S, McEniery CM, Cockcroft JR, Wilkinson IB. Aortic pulse wave velocity improves cardiovascular event prediction: an individual participant meta-analysis of prospective observational data from 17,635 subjects. J Am Coll Cardiol 63: 636-646, 2014. doi:10.1016/j.jacc.2013.09.063.
- 247. Swillens A, Taelman L, Degroote J, Vierendeels J, Segers P. Comparison of non-invasive methods for measurement of local pulse wave velocity using FSI-simulations and In vivo data. Ann Biomed Eng 41: 1567-1578, 2013. doi:10.1007/s10439-012-0688-z.
- 248. Chemla D, Hébert JL, Coirault C, Zamani K, Suard I, Colin P, Lecarpentier Y. Total arterial compliance estimated by stroke volumeto-aortic pulse pressure ratio in humans. Am J Physiol Heart Circ Physiol 274: H500-H505, 1998. doi:10.1152/ajpheart.1998.274.2.H500.
- 249. Segers P, Verdonck P, Deryck Y, Brimioulle S, Naeije R, Carlier S, Stergiopulos N. Pulse pressure method and the area method for the estimation of total arterial compliance in dogs: sensitivity to wave reflection intensity. Ann Biomed Eng 27: 480-485, 1999. doi:10.1114/ 1.192
- 250. Stergiopulos N, Meister J-J, Westerhof N. Evaluation of methods for estimation of total arterial compliance. Am J Physiol Heart Circ Physiol 268: H1540-H1548, 1995. doi:10.1152/ajpheart.1995.268.4. H1540.
- 251. Liu Z, Brin KP, Yin FC. Estimation of total arterial compliance: an improved method and evaluation of current methods. Am J Physiol Heart Circ Physiol 251: H588-H600, 1986. doi:10.1152/ajpheart.1986.
- 252. Stergiopulos N, Meister JJ, Westerhof N. Simple and accurate way for estimating total and segmental arterial compliance: the pulse pressure method. Ann Biomed Eng 22: 392-397, 1994. doi:10.1007/
- 253. Goldwyn RM, Wat TB. Arterial pressure pulse contour analysis via a mathematical model for the clinical quantification of human vascular properties. IEEE Trans Biomed Eng BME-14: 11-17, 1967. doi:10.1109/ TBME.1967.4502455.
- 254. Fogliardi R, Di Donfrancesco M, Burattini R. Comparison of linear and nonlinear formulations of the three-element Windkessel model. Am J Physiol Heart Circ Physiol 271: H2661-H2668, 1996. doi:10.1152/ajpheart.1996.271.6.H2661.
- 255. Quick CM, Berger DS, Noordergraaf A. Apparent arterial compliance. Am J Physiol Heart Circ Physiol 274: H1393-H1403, 1998. doi:10.1152/ajpheart.1998.274.4.H1393.
- 256. Pollack GH, Reddy RV, Noordergraaf A. Input impedance, wave travel, and reflections in the human pulmonary arterial tree: studies

- using an electrical analog. IEEE Trans Biomed Eng 15: 151-164, 1968. doi:10.1109/tbme.1968.4502559.
- 257. Nichols WW, Pepine CJ. Left ventricular afterload and aortic input impedance: implications of pulsatile blood flow. Prog Cardiovasc Dis 24: 293-306, 1982. doi:10.1016/0033-0620(82)90007-x.
- 258. Patel DJ, Defreitas FM, Fry DL. Hydraulic input impedance to aorta and pulmonary artery in dogs. J Appl Physiol 18: 134-140, 1963. doi:10.1152/jappl.1963.18.1.134.
- 259. O'Rourke MF, Taylor MG. Input impedance on the systemic circulation. Circ Res 20: 365-380, 1967. doi:10.1161/01.res.20.4.365.
- 260. Nichols WW, O'Rourke MF. McDonald's Blood Flow in Arteries (5th ed.). New York: Oxford University Press, 2005.
- 261. Li JK. Time domain resolution of forward and reflected waves in the aorta. IEEE Trans Biomed Eng 33: 783-785, 1986. doi:10.1109/ TBME.1986.325903.
- 262. Zong W, Heldt T, Moody GB, Mark RG. An open-source algorithm to detect onset of arterial blood pressure pulses. Comput Cardiol 2003: 259-262, 2003. doi:10.1109/CIC.2003.1291140.
- Obeid H, Khettab H, Marais L, Hallab M, Laurent S, Boutouyrie P. Evaluation of arterial stiffness by finger-toe pulse wave velocity: optimization of signal processing and clinical validation. J Hypertens 35: 1618-1625, 2017. doi:10.1097/HJH.000000000001371.
- 264. Paliakaite B, Charlton PH, Rapalis A, Pluščiauskaite V, Piartli P, Kaniusas E, Marozas V. Blood pressure estimation based on photoplethysmography: finger versus wrist. Comput Cardiol 2021: 1-4, 2021. doi:10.23919/CinC53138.2021.9662716.
- 265. Esper SA, Pinsky MR. Arterial waveform analysis. Best Pract Res Clin Anaesthesiol 28: 363–380, 2014. doi:10.1016/j.bpa.2014.08.002.
- 266. Schwid HA, Taylor LA, Smith NT. Computer model analysis of the radial artery pressure waveform. J Clin Monit 3: 220-228, 1987. doi:10.1007/BF03337375.
- 267. Oppenheim MI, Sittig DF. An innovative dicrotic notch detection algorithm which combines rule-based logic with digital signal processing techniques. Comput Biomed Res 28: 154-170, 1995. doi:10.1006/cbmr.1995.1011.
- 268. Hoeksel SA, Jansen JR, Blom JA, Schreuder JJ. Detection of dicrotic notch in arterial pressure signals. J Clin Monit 13: 309-316, 1997. doi:10.1023/a:1007414906294.
- 269. Dawber TR. Thomas HE. McNamara PM. Characteristics of the dicrotic notch of the arterial pulse wave in coronary heart disease. Angiology 24: 244-255, 1973. doi:10.1177/000331977302400407.
- 270. Mathieu A, Charlton PH, Alastruey J. Using smart wearables to monitor cardiac ejection. Proceedings (MDPI) 14: 48, 2018. doi:10.3390/ ecsa-5-05744.
- 271. Nichols WW. Clinical measurement of arterial stiffness obtained from noninvasive pressure waveforms. Am J Hypertens 18: 3S-10S, 2005. doi:10.1016/j.amjhyper.2004.10.009.
- 272. Kelly R, Hayward C, Avolio A, O'Rourke M. Noninvasive determination of age-related changes in the human arterial pulse. Circulation 80: 1652-1659, 1989. doi:10.1161/01.cir.80.6.1652.
- 273. Segers P, Rietzschel ER, De Buyzere ML, De Bacquer D, Van Bortel LM, de Backer G, Gillebert TC, Verdonck PR. Assessment of pressure wave reflection: getting the timing right! Physiol Meas 28: 1045-1056, 2007. doi:10.1088/0967-3334/28/9/006.
- 274. Millasseau SC, Patel SJ, Redwood SR, Ritter JM, Chowienczyk PJ. Pressure wave reflection assessed from the peripheral pulse: is a transfer function necessary? Hypertension 41: 1016-1020, 2003. doi:10.1161/01.HYP.0000057574.64076.A5.
- 275. Millasseau SC, Ritter JM, Takazawa K, Chowienczyk PJ. Contour analysis of the photoplethysmographic pulse measured at the finger. J Hypertens 24: 1449-1456, 2006. doi:10.1097/01.hjh.0000239277. 05068.87.
- 276. Elgendi M, Norton I, Brearley M, Abbott D, Schuurmans D. Detection of a and b waves in the acceleration photoplethysmogram. Biomed Eng Online 13: 139, 2014. doi:10.1186/1475-925X-13-
- 277. Elgendi M. Detection of c, d, and e waves in the acceleration photoplethysmogram. Comput Methods Programs Biomed 117: 125-136, 2014. doi:10.1016/j.cmpb.2014.08.001.
- 278. Franklin SS, Gustin W 4th, Wong ND, Larson MG, Weber MA, Kannel WB, Levy D. Hemodynamic patterns of age-related changes in blood pressure. The Framingham Heart Study. Circulation 96: 308-315, 1997. doi:10.1161/01.cir.96.1.308.

- 279. Roman MJ, Devereux RB, Kizer JR, Okin PM, Lee ET, Wang W, Umans JG, Calhoun D, Howard BV. High central pulse pressure is independently associated with adverse cardiovascular outcome the strong heart study. J Am Coll Cardiol 54: 1730-1734, 2009. doi:10.1016/ j.jacc.2009.05.070.
- 280. Chirinos JA, Segers P. Noninvasive evaluation of left ventricular afterload: part 2: arterial pressure-flow and pressure-volume relations in humans. Hypertension 56: 563-570, 2010. doi:10.1161/ HYPERTENSIONAHA.110.157339.
- 281. Kaess BM, Rong J, Larson MG, Hamburg NM, Vita JA, Levy D, Benjamin EJ. Vasan RS. Mitchell GF. Aortic stiffness, blood pressure progression, and incident hypertension. JAMA 308: 875-881, 2012. doi:10.1001/2012.jama.10503.
- 282. O'Rourke MF, Safar ME. Relationship between aortic stiffening and microvascular disease in brain and kidney: cause and logic of therapy. Hypertension 46: 200-204, 2005. doi:10.1161/01.HYP. 0000168052.00426.65.
- 283. Mitchell GF. Aortic stiffness, pressure and flow pulsatility, and target organ damage. J Appl Physiol (1985) 125: 1871-1880, 2018. doi:10.1152/ japplphysiol.00108.2018.
- 284. Avolio AP, Van Bortel LM, Boutouyrie P, Cockcroft JR, McEniery CM, Protogerou AD, Roman MJ, Safar ME, Segers P, Smulyan H. Role of pulse pressure amplification in arterial hypertension: experts' opinion and review of the data. Hypertension 54: 375-383, 2009 [Erratum in Hypertension 58: e30, 2011]. doi:10.1161/ HYPERTENSIONAHA.109.134379.
- 285. Benetos A, Gautier S, Labat C, Salvi P, Valbusa F, Marino F, Toulza O, Agnoletti D, Zamboni M, Dubail D, Manckoundia P, Rolland Y, Hanon O, Perret-Guillaume C, Lacolley P, Safar ME, Guillemin F. Mortality and cardiovascular events are best predicted by low central/peripheral pulse pressure amplification but not by high blood pressure levels in elderly nursing home subjects: the PARTAGE (Predictive Values of Blood Pressure and Arterial Stiffness in Institutionalized Very Aged Population) study. J Am Coll Cardiol 60: 1503-1511, 2012. doi:10.1016/j.jacc.2012.04.055.
- 286. McEniery CM, Yasmin , McDonnell B, Munnery M, Wallace SM, Rowe CV, Cockcroft JR. Wilkinson IB; Anglo-Cardiff Collaborative Trial Investigators. Central pressure: variability and impact of cardiovascular risk factors: the Anglo-Cardiff Collaborative Trial II. Hypertension 51: 1476-1482, 2008. doi:10.1161/HYPERTENSIONAHA.107.105445.
- 287. Wilkinson IB, Mohammad NH, Tyrrell S, Hall IR, Webb DJ, Paul VE, Levy T, Cockcroft JR. Heart rate dependency of pulse pressure amplification and arterial stiffness. Am J Hypertens 15: 24-30, 2002. doi:10.1016/s0895-7061(01)02252-x.
- 288. Herbert A, Cruickshank JK, Laurent S, Boutouyrie P; Reference Values for Arterial Measurements Collaboration. Establishing reference values for central blood pressure and its amplification in a general healthy population and according to cardiovascular risk factors. Eur Heart J 35: 3122-3133, 2014. doi:10.1093/eurheartj/ehu293.
- 289. Nijdam M, Plantinga EY, Hulsen HT, Bos WJ, Grobbee DE, van der Schouw YT, Bots ML. Pulse pressure amplification and risk of cardiovascular disease. Am J Hypertens 21: 388-392, 2008. doi:10.1038/
- 290. Breet Y, Huisman HW, Kruger R, van Rooyen JM, Gafane-Matemane LF, Ware LJ, Schutte AE. Pulse pressure amplification and its relationship with age in young, apparently healthy black and white adults: the African-PREDICT study. Int J Cardiol 15: 387-391, 2017. doi:10.1016/j.ijcard.2017.08.070.
- 291. Jeroncic A, Gunjaca G, Mrsic DB, Mudnic I, Brizic I, Polasek O, Boban M. Normative equations for central augmentation index: assessment of inter-population applicability and how it could be improved. Sci Rep 6: 27016, 2016. doi:10.1038/srep27016.
- 292. Yasmin, Brown MJ. Similarities and differences between augmentation index and pulse wave velocity in the assessment of arterial stiffness. QJM 92: 595-600, 1999. doi:10.1093/gjmed/92.10.595.
- 293. Sakurai M, Yamakado T, Kurachi H, Kato T, Kuroda K, Ishisu R, Okamoto S, Isaka N, Nakano T, Ito M. The relationship between aortic augmentation index and pulse wave velocity: an invasive study. J Hypertens 25: 391-397, 2007. doi:10.1097/HJH.0b013e3280115b7c.
- 294. Fantin F, Mattocks A, Bulpitt CJ, Banya W, Rajkumar C. Is augmentation index a good measure of vascular stiffness in the elderly? Age Ageing 36: 43-48, 2007. doi:10.1093/ageing/afl115.

- 295. London GM, Blacher J, Pannier B, GúErin AP, Marchais SJ, Safar ME. Arterial wave reflections and survival in end-stage renal failure. Hypertension 38: 434-438, 2001. doi:10.1161/01.hyp.38.3.434.
- 296. Nürnberger J, Keflioglu-Scheiber A, Opazo Saez AM, Wenzel RR, Philipp T, Schäfers RF. Augmentation index is associated with cardiovascular risk. J Hypertens 20: 2407–2414, 2002. doi:10.1097/ 00004872-200212000-00020.
- 297. Weber T, Auer J, O'rourke MF, Kvas E, Lassnig E, Lamm G, Stark N, Rammer M, Eber B. Increased arterial wave reflections predict severe cardiovascular events in patients undergoing percutaneous coronary interventions. Eur Heart J 26: 2657-2663, 2005. doi:10.1093/ eurheartj/ehi504.
- 298. Chirinos JA, Zambrano JP, Chakko S, Veerani A, Schob A, Willens HJ, Perez G, Mendez AJ. Aortic pressure augmentation predicts adverse cardiovascular events in patients with established coronary artery disease. Hypertension 45: 980-985, 2005. doi:10.1161/01. HYP.0000165025.16381.44.
- 299. Torjesen AA, Wang N, Larson MG, Hamburg NM, Vita JA, Levy D, Benjamin EJ, Vasan RS, Mitchell GF. Forward and backward wave morphology and central pressure augmentation in men and women in the Framingham Heart Study. Hypertension 64: 259-265, 2014. doi:10.1161/HYPERTENSIONAHA.114.03371.
- 300. Fok H, Guilcher A, Li Y, Brett S, Shah A, Clapp B, Chowienczyk P. Augmentation pressure is influenced by ventricular contractility/ relaxation dynamics: novel mechanism of reduction of pulse pressure by nitrates. Hypertension 63: 1050-1055, 2014. doi:10.1161/ HYPERTENSIONAHA.113.02955.
- 301. van de Velde L, Eeftinck Schattenkerk DW, Venema PAHT, Best HJ, van den Bogaard B, Stok WJ, Westerhof BE, van den Born BJH. Myocardial preload alters central pressure augmentation through changes in the forward wave. J Hypertens 36: 544-551, 2018. doi:10.1097/HJH.0000000000001583.
- Li Y, Jiang B, Keehn L, Gu H, Boguslavskyi A, Cecelja M, Vennin S, Spector T, Alastruey J, Chowienczyk P. Hemodynamic mechanism of the age-related increase in pulse pressure in women-insights from Twins United Kingdom. Hypertension 73: 1018-1024, 2019. doi:10.1161/HYPERTENSIONAHA.118.12402.
- 303. Pagoulatou S, Adamopoulos D, Rovas G, Bikia V, Stergiopulos N. The effect of left ventricular contractility on arterial hemodynamics: a model-based investigation. PLoS One 16: e0255561, 2021. doi:10.1371/ journal.pone.0255561.
- 304. Wilkinson IB, MacCallum H, Flint L, Cockcroft JR, Newby DE, Webb DJ. The influence of heart rate on augmentation index and central arterial pressure in humans. J Physiol 525: 263-270, 2000. doi:10.1111/j.1469-7793.2000.t01-1-00263.x.
- 305. Baksi AJ, Treibel TA, Davies JE, Hadjiloizou N, Foale RA, Parker KH, Francis DP, Mayet J, Hughes AD. A meta-analysis of the mechanism of blood pressure change with aging. J Am Coll Cardiol 54: 2087-2092, 2009. doi:10.1016/j.jacc.2009.06.049.
- 306. Westerhof BE, Westerhof N. Uniform tube models with single reflection site do not explain aortic wave travel and pressure wave shape. Physiol Meas 39: 124006, 2018. doi:10.1088/1361-6579/aaf3dd.
- 307. Westerhof BE, van Loon L, Westerhof N. Reflections revisited, reinterpretation required. Artery Res 26: 117-120, 2020. doi:10.2991/ artres.k.200423.001.
- 308. Khir AW, Hughes AD, Parker KH. The inflection point is not a reliable method for the determination of the augmentation index (Abstract), JAm Coll Cardiol 47: 315A, 2006.
- 309. Westerhof BE, Westerhof N. Magnitude and return time of the reflected wave: the effects of large artery stiffness and aortic geometry. J Hypertens 30: 932–939, 2012. doi:10.1097/HJH.0b013e3283524932.
- 310. Westerhof N. Westerhof BE. Wave transmission and reflection of waves "The myth is in their use". Artery Res 6: 1-6, 2012. doi:10.1016/ j.artres.2012.01.004.
- 311. Aboyans V, Criqui MH, Abraham P, Allison MA, Creager MA. Measurement and interpretation of the ankle-brachial index: a scientific statement from the American Heart Association. Circulation 11: 2890-2909, 2012. doi:10.1161/CIR.0b013e318276fbcb.
- 312. Zhang S, Mao HJ, Sun B, Wang NN, Zhang B, Zeng M, Xu L, Yu XB, Liu J, Xing CY. The relationship between AASI and arterial atherosclerosis in ESRD patients. Ren Fail 37: 22-28, 2015. doi:10.3109/ 0886022X.2014.963787.
- 313. Westerhof N, Lankhaar JW, Westerhof BE. Ambulatory arterial stiffness index is not a stiffness parameter but a ventriculo-arterial

- coupling factor. Hypertension 49: e7, 2007. doi:10.1161/01.HYP. 0000254947.07458.90.
- 314. Kips JG, Vermeersch SJ, Reymond P, Boutouyrie P, Stergiopulos N, Laurent S, Van Bortel LM, Segers P. Ambulatory arterial stiffness index does not accurately assess arterial stiffness. J Hypertens 30: 574-580, 2012. doi:10.1097/HJH.0b013e32834fca18.
- 315. Shirai K, Hiruta N, Song M, Kurosu T, Suzuki J, Tomaru T, Miyashita Y, Saiki A, Takahashi M, Suzuki K, Takata M. Cardioankle vascular index (CAVI) as a novel indicator of arterial stiffness: theory, evidence and perspectives. J Atheroscler Thromb 18: 924-938, 2011. doi:10.5551/jat.7716.
- 316. Hayashi K, Handa H, Nagasawa S, Okumura A, Moritake K. Stiffness and elastic behavior of human intracraneal and extracraneal arteries. J Biomech 13: 175-184, 1980. doi:10.1016/0021-9290(80)90191-8.
- 317. Spronck B, Avolio AP, Tan I, Butlin M, Reesink KD, Delhaas T. Arterial stiffness index β and cardio-ankle vascular index inherently depend on blood pressure but can be readily corrected. J Hypertens 35: 98-104, 2017. doi:10.1097/HJH.000000000001132.
- 318. Hirata K, Yaginuma T, O'Rourke MF, Kawakami M. Age-related changes in carotid artery flow and pressure pulses: possible implications for cerebral microvascular disease. Stroke 37: 2552-2556, 2006. doi:10.1161/01.STR.0000242289.20381.f4.
- von Wowern E, Ostling G, Nilsson PM, Olofsson P. Digital photoplethysmography for assessment of arterial stiffness: repeatability and comparison with applanation tonometry. PLoS One 10: e0135659, 2015. doi:10.1371/journal.pone.0135659.
- 320. Bortolotto LA, Blacher J, Kondo T, Takazawa K, Safar ME. Assessment of vascular aging and atherosclerosis in hypertensive subjects: second derivative of photoplethysmogram versus pulse wave velocity. Am J Hypertens 13: 165-171, 2000. doi:10.1016/s0895-7061(99)00192-2.
- 321. Peltokangas M, Telembeci AA, Verho J, Mattila VM, Romsi P, Vehkaoja A, Lekkala J, Oksala N. Parameters extracted from arterial pulse waves as markers of atherosclerotic changes: performance and repeatability. IEEE J Biomed Health Inform 22: 750-757, 2018. doi:10.1109/JBHI.2017.2679904.
- 322. Millasseau SC, Kelly RP, Ritter JM, Chowienczyk PJ. Determination of age-related increases in large artery stiffness by digital pulse contour analysis. Clin Sci (Lond) 103: 371-377, 2002. doi:10.1042/ cs1030371.
- 323. Zekavat SM, Aragam K, Emdin C, Khera AV, Klarin D, Zhao H, Natarajan P. Genetic association of finger photoplethysmographyderived arterial stiffness index with blood pressure and coronary artery disease. Arterioscler Thromb Vasc Biol 39: 1253-1261, 2019. doi:10.1161/ATVBAHA.119.312626.
- 324. Wei HC, Xiao MX, Chen HY, Li YQ, Wu HT, Sun CK. Instantaneous frequency from Hilbert-Huang transformation of digital volume pulse as indicator of diabetes and arterial stiffness in upper-middle-aged subjects. Sci Rep 8: 15771, 2018. doi:10.1038/s41598-018-34091-6.
- 325. Sudlow C, Gallacher J, Allen N, Beral V, Burton P, Danesh J, Downey P, Elliott P, Green J, Landray M, Liu B, Matthews P, Ong G, Pell J, Silman A, Young A, Sprosen T, Peakman T, Collins R. UK Biobank: an open access resource for identifying the causes of a wide range of complex diseases of middle and old age. PLoS Med 31: e1001779, 2015. doi:10.1371/journal.pmed.1001779.

- 326. Allen J, Overbeck K, Nath AF, Murray A, Stansby G. A prospective comparison of bilateral photoplethysmography versus the anklebrachial pressure index for detecting and quantifying lower limb peripheral arterial disease. J Vasc Surg 47: 794-802, 2008. doi:10.1016/ j.jvs.2007.11.057.
- 327. Dillon JB, Hertzman AB. The form of the volume pulse in the finger pad in health, arteriosclerosis, and hypertension. Am Heart J 21: 172-190, 1941. doi:10.1016/S0002-8703(41)90966-3.
- 328. Eck VG, Donders WP, Sturdy J, Feinberg J, Delhaas T, Hellevik L, Huberts RW. A guide to uncertainty quantification and sensitivity analysis for cardiovascular applications. Int J Numer Meth Biomed Eng 32: e02755, 2016. doi:10.1002/cnm.2755.
- 329. Melis A, Clayton RH, Marzo A. Bayesian sensitivity analysis of a 1D vascular model with Gaussian process emulators. Int J Numer Method Biomed Eng 33: e2882, 2017. doi:10.1002/cnm.2882.
- 330. Bikia V, Lazaroska M, Scherrer Ma D, Zhao M, Rovas G, Pagoulatou S, Stergiopulos N. Estimation of left ventricular end-systolic elastance from brachial pressure waveform via deep learning. Front Bioeng Biotechnol 9: 754003, 2021. doi:10.3389/fbioe.2021.
- 331. Cunningham JW, Di Achille P, Morrill VN, Weng L-C, Choi SH, Khurshid S, Nauffal V, Pirruccello JP, Solomon SD, Batra P, Ho JE, Philippakis AA, Ellinor PT, Lubitz SA. Machine learning to understand genetic and clinical factors associated with the pulse waveform dicrotic notch. Circ: Genom Precis Med 16: e003676, 2023. doi:10.1161/CIRCGEN.121.003676.
- Cecelja M, Keehn L, Ye L, Spector TD, Hughes AD, Chowienczyk P. Genetic aetiology of blood pressure relates to aortic stiffness with bi-directional causality: evidence from heritability, blood pressure polymorphisms, and Mendelian randomization. Eur Heart J 41: 3314-3322, 2020. doi:10.1093/eurheartj/ehaa238.
- 333. Mitchell GF, Hwang SJ, Vasan RS, Larson MG, Pencina MJ, Hamburg NM, Vita JA, Levy D, Benjamin EJ. Arterial stiffness and cardiovascular events: the Framingham Heart study. Circulation 121: 505-511, 2010. doi:10.1161/CIRCULATIONAHA.109.886655.
- Rietzschel ER, De Buyzere ML, Bekaert S, Segers P, De Bacquer D, Cooman L, Van Damme P, Cassiman P, Langlois M, van Oostveldt P, Verdonck P, De Backer G, Gillebert TC, Asklepios Investigators. Rationale, design, methods and baseline characteristics of the Asklepios Study. Eur J Cardiovasc Prev Rehabil 14: 179–191, 2007. doi:10.1097/HJR.0b013e328012c380.
- Jin W, Chowienczyk P, Alastruey J. An in silico simulation of flowmediated dilation reveals that blood pressure and other factors may influence the response independent of endothelial function. Am J Physiol Heart Circ Physiol 318: H1337-H1345, 2020. doi:10.1152/ ajpheart.00703.2019.
- 336. Bianchini E, Guala A, Golemati S, Alastruey J, Climie RE, Dalakleidi K, Francesconi M, Fuchs D, Hartman Y, Malik AEF, Makūnaitė M, Nikita KS, Park C, Pugh CJA, Šatrauskienė A, Terentes-Printizios D, Teynor A, Thijssen D, Schmidt-Trucksäss A, Zupkauskienė J, Boutouyrie P, Bruno RM, Reesink KD. The Ultrasound Window Into Vascular Ageing: A Technology Review by the VascAgeNet COST Action. J Ultrasound Med 2023 May 6 [Epub ahead of print]. doi:10.1002/jum.16243.