



Spatial distribution of metabolites in the retina and its relevance to studies of metabolic retinal disorders

Roberto Bonelli^{1,2} · Sasha M. Woods³ · Sarah Lockwood⁴ · Paul N. Bishop^{5,6} · Kamron N. Khan⁷ · Melanie Bahlo^{1,2} · Brendan R. E. Ansell^{1,2} · Marcus Fruttiger³

Received: 29 June 2022 / Accepted: 21 December 2022
© The Author(s) 2023

Abstract

Introduction The primate retina has evolved regional specialisations for specific visual functions. The macula is specialised towards high acuity vision and is an area that contains an increased density of cone photoreceptors and signal processing neurons. Different regions in the retina display unique susceptibility to pathology, with many retinal diseases primarily affecting the macula.

Objectives To better understand the properties of different retinal areas we studied the differential distribution of metabolites across the retina.

Methods We conducted an untargeted metabolomics analysis on full-thickness punches from three different regions (macula, temporal peri-macula and periphery) of healthy primate retina.

Results Nearly half of all metabolites identified showed differential abundance in at least one comparison between the three regions. Furthermore, mapping metabolomics results from macula-specific eye diseases onto our region-specific metabolite distributions revealed differential abundance defining systemic metabolic dysregulations that were region specific.

Conclusions The unique metabolic phenotype of different retinal regions is likely due to the differential distribution of different cell types in these regions reflecting the specific metabolic requirements of each cell type. Our results may help to better understand the pathobiology of retinal diseases with region specificity.

Melanie Bahlo, Brendan R. E. Ansell, and Marcus Fruttiger jointly supervised the preparation and publication of this manuscript.

✉ Marcus Fruttiger
m.fruttiger@ucl.ac.uk

- ¹ Population Health & Immunity Division, The Walter & Eliza Hall Institute of Medical Research, 1G Royal Parade, Parkville, VIC 3052, Australia
- ² Department of Medical Biology, The University of Melbourne, Parkville, VIC 3010, Australia
- ³ UCL Institute of Ophthalmology, University College London, 11-43 Bath St, London EC1V 9EL, UK
- ⁴ UC Davis, CA National Primate Research Centre, Davis, CA 95616, USA
- ⁵ Division of Evolution, Infection and Genomics, School of Biological Sciences, Faculty of Biology, Medicine, and Health, University of Manchester, Manchester M13 9PT, UK
- ⁶ Manchester Royal Eye Hospital, Manchester University NHS Foundation Trust, Manchester Academic Health Science Centre, Manchester M13 9WL, UK
- ⁷ The Leeds Teaching Hospitals NHS Trust, St. James's Hospital, Leeds LS9 7TF, UK

1 Introduction

Systemic metabolic dysregulation can cause pathology in the retina with diabetic retinopathy being a prime example of this. It has been recognised that diabetes long-term metabolic dysregulations can lead to complications in the retina and vision loss with hyperglycaemia believed to be one of the main disease drivers. More recently, several studies have aimed to investigate potential links between other retinal disorders and systemic metabolic changes. For example, metabolomic studies performed on serum from age-related macular degeneration (AMD) have identified associations between dysregulations of lipids as well as amino acids with AMD disease status or severity (Acar et al., 2020; Brown et al., 2018; Laíns et al., 2017, 2019). Similarly, metabolomic profiling of Macular telangiectasia type 2 (MacTel) patients identified serum levels of serine, and sphingolipids as an important MacTel risk factor (Bonelli et al., 2021; Bonelli et al., 2020; Bonelli et al., 2021; Gantner et al., 2019; Scerri et al., 2017; Tyynismaa, 2019). However, it is not clear whether the systemic manifestations (i.e. changes in

the serum) are causally related to retinal pathology in AMD or MacTel, or whether they are just indicators of underlying disturbances affecting both retina and peripheral blood.

Complexity is also introduced when retinal diseases do not affect the tissue uniformly due to its spatial structural variation. High acuity vision depends on a region called the macula, which has a peak density of cone photoreceptors as well as retinal ganglion cells and is thicker than the peripheral retina, which is rich in rod photoreceptors. This differential distribution of cells across the retina is reflected by different transcriptional profiles in different retinal regions (Hu et al., 2019; Ratnapriya et al., 2019; Voigt et al., 2019, 2020; Whitmore et al., 2014; Yan et al., 2020; Yi et al., 2021). However, spatially differential gene expression data presented, so far, limited success explaining why, for example, the macula is particularly affected by diseases such as AMD or MacTel. Furthermore, it is not clear how changes in metabolism, mentioned above, may have a differential impact on retinal diseases. It is also not clear how differential cell distribution impacts metabolite levels in different areas of the retina.

Understanding the spatial distribution of metabolites in the retina, and how that relates to different retinal cell types may therefore be useful to understand how systemic risk factors might affect specific cells in the retina and in particular the macula. This study presents an untargeted metabolomics analysis performed on primates measuring the main metabolic profiles of the different regions of the retina. We identify metabolites and metabolic pathways that differentiate the macula from more peripheral regions and investigate the relationship between these findings and the distribution of different cell types in the retina. Lastly, using our results, we investigate how systemic metabolic risk factors found in both AMD and MacTel relate to the specific metabolic characteristics of the macula.

2 Results

2.1 Differential metabolite abundance across the retina

To map the distribution of metabolites across the primate retina, samples from three different retinal areas were collected from eleven primate (*Macaca fascicularis*) postmortem eyes and analysed by untargeted metabolomics mass spectrometry analysis (Metabolon Inc.). The raw data (peak areas) underwent log transformation, quality and low abundance filtering, normalisation and missing value imputation. One sample was discarded from further analyses as it was confirmed to be incorrectly collected when checking positive controls (Materials and Methods). The final dataset consisted of 32 retinal samples from six primates. Three

females and three males were included in this study with an average age of 2.1 years. For each sample, 371 metabolite abundances were retained. Linear modelling correcting for several covariates was performed as described in Materials and Methods to detect metabolites with differential abundance between retinal areas. A schematic overview of the study methods and analyses is presented in Fig. 1. We found a total of 197 (53%) metabolites whose abundance was significantly different in at least one statistical contrast (Table S1). The number and intersection of differentially abundant metabolites between retinal areas are presented in Figure S1. The largest number of differences was found between the macula and periphery (190), the two regions with the largest spatial distance in the retina. We did however find more differentially abundant metabolites between the macula and temporal areas (97), than between the temporal and peripheral areas (53).

Comparing the abundance of individual metabolites in the macula versus periphery (Figure S2-3, Table S1), the most significantly enriched metabolites were carotene diol 1 and 3, corresponding to the macular pigments lutein and zeaxanthin. These two diet-derived xenobiotics are known to be enriched in the macula of primates and were used in this study to validate the correct dissection of the macular samples (Materials and Methods). Of note, ergothioneine and beta-guanidinopropanoate, also diet-derived, were the third and fourth most significantly enriched metabolites, as well as xenobiotics, in the macula. In contrast, the positive control euthanasia drug, pentobarbital, was equally distributed across the retina (Table S1). The most enriched endogenous metabolites in the macula were *N*-acetyl-3-methylhistidine, *N*-acetyl-aspartyl-glutamate (NAAG) and 2-hydroxyglutarate. The NAA precursors *N*-acetylaspartate (NAA) and aspartate were also significantly enriched metabolites (rank 15 and 25) (Table S1). The most depleted non-lipid metabolites in the macula were putrescine and taurine (rank 2 and 4, Table S1).

To better understand which aspects of the metabolism differ between the retinal periphery and the macula, metabolites were grouped according to metabolic pathways and biochemical families and tested for directional enrichment (Fig. 2, Figure S2, Table S1). The pathway with the most significant enrichment in the macula was the alanine/aspartate pathway, followed by the tricarboxylic acid (TCA) cycle. Conversely, the macula was relatively depleted of phosphatidylethanolamine, ceramide, sphingomyelin, diacylglycerol, polyamine and benzoate (in order of statistical significance). Overall, there was a similar number of metabolites enriched (99) versus metabolites depleted in the macula (98), but these differences were contributed to by metabolites from different classes. Overall, lipids were depleted in the macula and non-lipid metabolites were enriched, as displayed in Fig. 2, Figure S3). Furthermore, four of the six significantly

Study Schematic

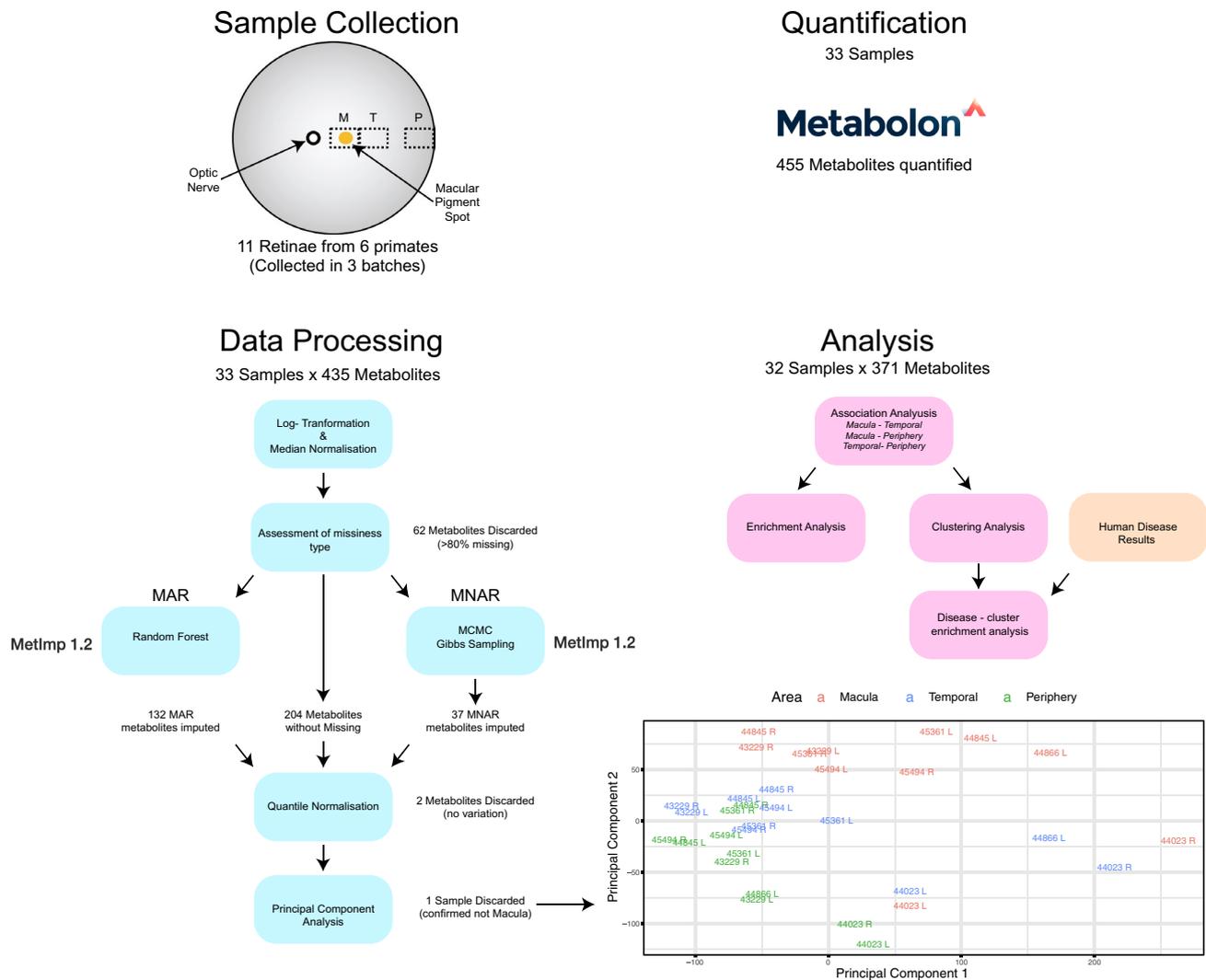


Fig. 1 Study schematic and principal components analysis plot

depleted metabolic groups were lipids (note, the bottom four rows in Fig. 2 are primarily depleted).

Plotting the distribution of metabolic pathways and biochemical families in the three sampled regions (macula, temporal and peripheral) further illustrates the relative lack of ceramide, phosphatidylethanolamine, sphingomyelin and diacylglycerol in the macula compared to the periphery (Supplementary Figure 2). Of note, phosphatidylcholine and plasmenylethanolamine were much more evenly distributed, and carnitine-related fatty acid metabolism and lysophosphatidylcholine were the only lipid groups enriched in the macula.

In addition, we tested for metabolic pathways enriched with differentially abundant metabolites represented in both

directions ('mixed directionality enrichment'). Pathways with significant enrichment/depletion in the macula were the leucine, methionine, glycine/serine, urea cycle, glutathione, purine, phosphatidylcholine and the plasmenylethanolamine metabolite classes (Fig. 2, Table S1). The individual differentially-abundant metabolites that constitute these mixed directionality results (pathways labelled with a star in Fig. 2) are displayed in Fig. 3. In contrast to the other lipid groups, the number of different phosphatidylcholines (dark green in Fig. 3) was similar in the enriched and depleted categories. However, the unsaturated forms were typically enriched, and the saturated ones tended to be depleted in the macula. Also of note were the purine family metabolites, wherein phosphorylated purines (ADP, AMP, GDP and IMP) were

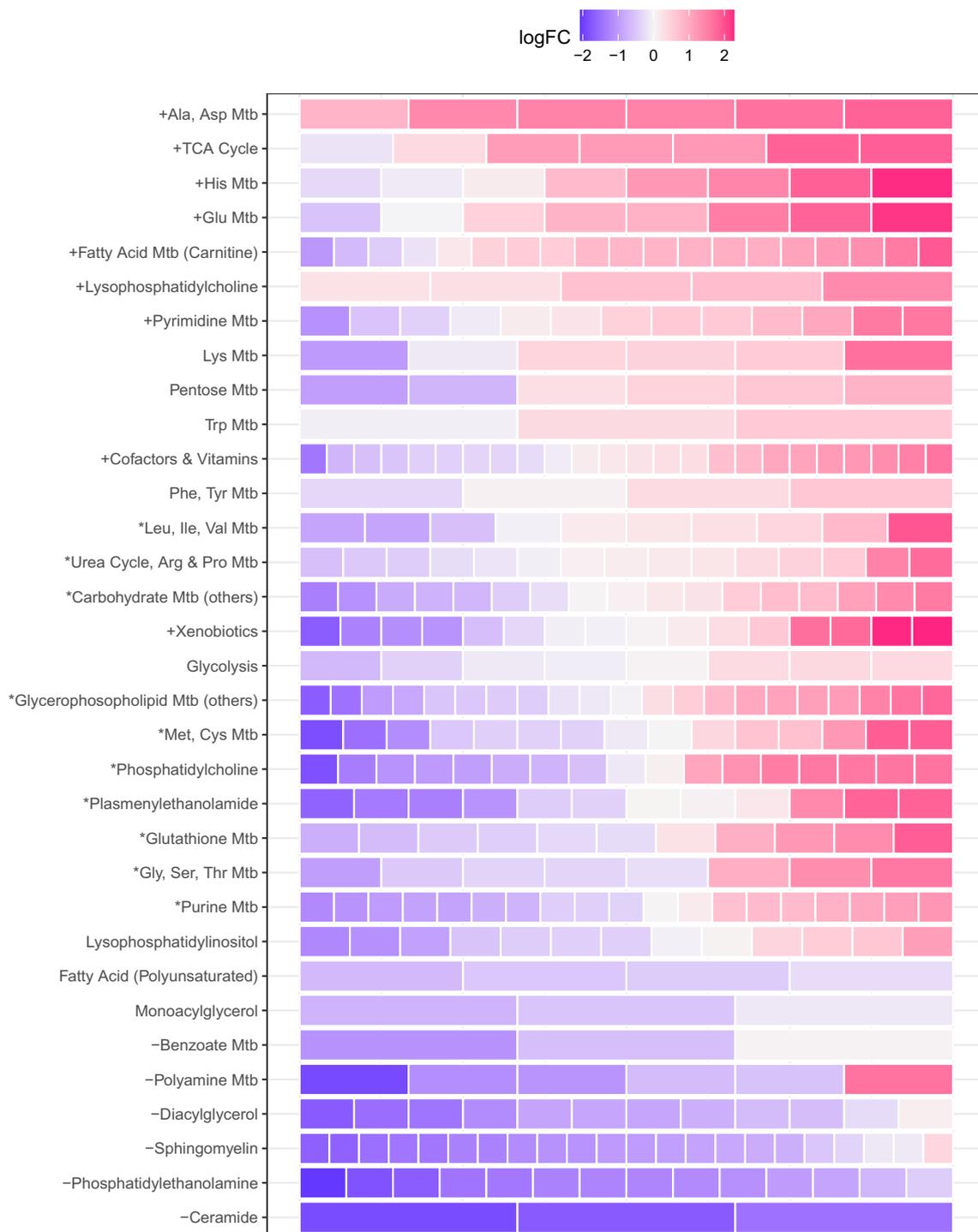


Fig. 2 Differential abundance of metabolites across retinal space. Relative abundance of N metabolites (filled rectangles, or 'bricks') grouped by biochemical family/pathway (y axis) and ordered by greater family-wise abundance in the macula (top) to the periphery (bottom). The number of bricks indicates the number of metabolites in the pathway available for this study. The colour of each brick represents the log-fold change of that metabolite between macula and periphery. Positive log-fold change (reds) indicates that the

metabolite is more abundant in the macula while negative values (blues) indicate that the metabolite is more abundant in the periphery. + denotes family is significantly enriched in the macula (mainly red); - denotes family is significantly depleted in the macula (mainly blue); * denotes family is more differentially abundant in both directions than would be expected by chance ('mixed directionality'/signal mixture effect)

enriched in the macula, and their non-phosphorylated counterparts (adenosine, guanosine and inosine) were depleted.

Abundance differences of each metabolite in the macula and peripheral regions are displayed in Figures S2, Figure S4, and Figure S5. To explore the distribution characteristics of metabolites across all three sampled retinal areas, we grouped metabolites according to differential abundance patterns that progressed laterally from the macula to the temporal region to the periphery. We defined ten major pattern clusters (Table 1, Figure S6, Figure S7), the largest of which contained metabolites that progressively increased or decreased according to proximity to the retinal centre. A total of 173 metabolites showed the greatest abundance differences at the extremes of the macula > temporal > peripheral axis, with only seven showing significant differences in the temporal retina relative to the other regions.

Similarly, we clustered metabolic families by first deriving principal components (PCs) for each family and then testing for differences in their magnitude between retinal regions. As expected, when these differences in PC abundance were grouped, most pathways/families recapitulated spatial distribution patterns observed using their constituent metabolite abundances (Table 1, Table S1).

Lastly, we tested for the effects of sex and age on the metabolic composition of the retina. Interestingly, we found female primates to exhibit increased levels of phosphatidyl-ethanolamine and plasmenyl-ethanolamide metabolic pathways, and decreased levels of lysophosphatidylcholine lysine-related pathways compared to males (Table S2, Figure S8). Older age tended to positively correlate with sphingomyelins and negatively impact levels of the leucine, isoleucine and valine-related metabolites. Testing for differential effect for age and sex on metabolic abundances across the areas did not reveal any major signal (Table S2).

2.2 The regional retinal context of disease-related metabolites

To place results from our analysis in healthy primate retina into the context of pathology, we mapped our data against serum-based investigations of two retinal disorders that exclusively affect the macula and have known associations with systemic metabolic changes, Macular Telangiectasia type 2 (MacTel) (Bonelli et al., 2021; Bonelli et al., 2020; Bonelli et al., 2021; Gantner et al., 2019; Scerri et al., 2017; Tynismaa, 2019) and Age-related Macular Degeneration (AMD) (Acar et al., 2020; Brown et al., 2018; Láíns et al., 2017, 2019). To this end, we leveraged differential serum metabolite abundance results from our previously published MacTel study (Bonelli et al., 2020), to compare serum metabolites in patients with MacTel compared to controls. In addition to MacTel, we also recruited a cohort of 205 individuals with AMD and 146 healthy controls. Patients

were divided into sub-phenotypes of choroidal neovascularization (CNV), geographic atrophy (GA) and 'mixed'. Abundances of 763 serum metabolites were compared between each AMD patient subgroup and the controls. Although similar size studies of AMD have already been published (Acar et al., 2020; Brown et al., 2018; Láíns et al., 2017, 2019), our data and its analysis ensured that the data generation, pre-processing, cleaning and statistical analysis were consistent across datasets. In all statistical contrasts, only four metabolites were identified as being significantly differentially abundant after correcting for multiple testing: tryptophan betaine, heptanoate, 1-linoleolglycerol (18:2) and 1-pentadecanoylglycerol (15:0). All four were depleted in the serum of AMD-CNV patients compared to controls. A further 73 metabolites were differentially abundant in this patient subgroup at the nominal threshold of $p < 0.05$ (Supplementary Results and Table S3). We found 36 and 37 nominally significant metabolites when comparing AMD-GA and AMD-Mixed to controls. None of these remained significant after accounting for multiple testing.

To investigate the spatial dimension of patient serum results in the retina, we separated the log fold-change results from the human serum-based studies into three groups according to their specific abundance in the primate macula or periphery, or annotated them as having a non-significant difference (Fig. 4A–B). Several of the metabolites that were changed in patient serum were found to be differentially distributed in healthy primates' retinas. For instance, metabolites of the glycine/serine/threonine metabolism as well as alanine/asparagine metabolism were systemically depleted in the serum of MacTel subjects and were defined by our analyses to be more abundant in the macula compared with periphery or not differentially abundant (Fig. 4A). The opposite trend was evident for phosphatidylethanolamines, which exhibited lower abundance in the macula than periphery but were enriched in the serum of MacTel patients. Sphingomyelins were significantly more abundant in the macula and depleted in MacTel patient serum. This analysis revealed however some interesting diversification patterns across the methionine and cysteine metabolism whose metabolites depleted in MacTel were mostly macula enriched rather than the non-differentiated ones. In AMD, purine and ceramide metabolism was reduced in patient serum and healthy primate retina (Fig. 4B).

3 Discussion

In this study, we measured metabolic abundances in three different regions of the primate retina: the central macular, temporal, and peripheral regions. Using a large and untargeted panel of 371 metabolites for 32 retinal samples, our

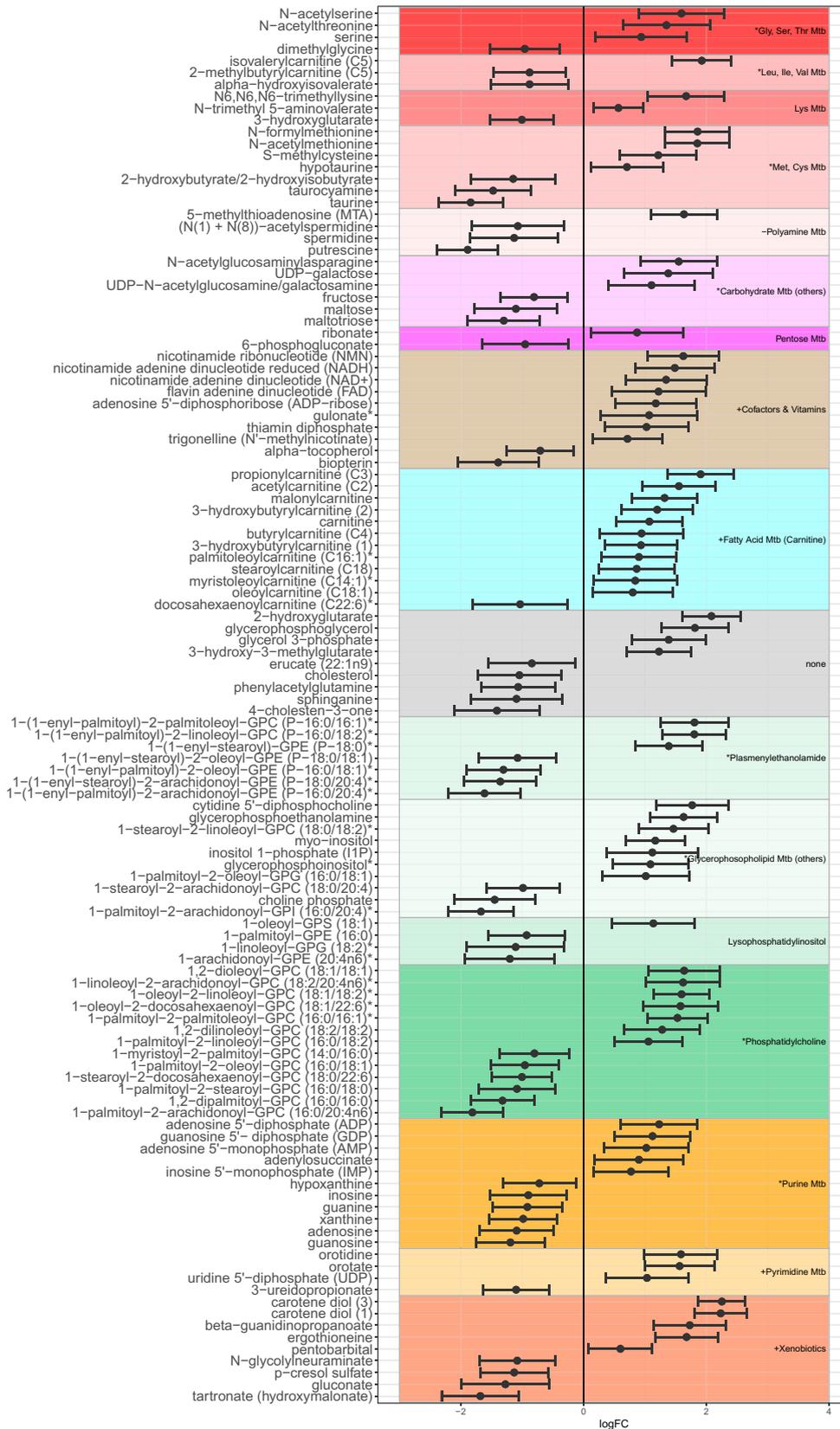


Fig. 3 Log-fold changes and 95% confidence interval of metabolite families/pathways with mixed abundances between the macula and periphery. Only metabolites which are individually significantly differentially abundant are displayed. Positive log-fold changes values in this figure indicate that the metabolite abundance was higher in the macula compared to the periphery and vice versa. Metabolites are grouped and coloured by their respective biochemical families/biological pathways. +denotes family is significantly enriched in the macula (mainly red); —denotes family is significantly depleted in the macula (mainly blue); * denotes family is more differentially abundant in both directions than would be expected by chance (signal mixture effect). Families with significant abundance entirely in the macula or periphery are omitted for clarity (provided in Figure S5–7)

study is the first of its kind to elucidate the metabolic profile of retinal regional areas by identifying multiple differences. We found a total of almost 200 metabolites whose abundance differed between the three areas, highlighting a clear metabolic separation between the macular region of the retina compared to the outer, temporal or peripheral, areas.

The most enriched metabolites in the macula were identified in our study as carotene diol 1 and 3. These are carotenoids containing an oxygenated carotene backbone and are known as xanthophylls. It is well established that lutein and zeaxanthin are two diet-derived xanthophylls highly enriched in the macula and responsible for the characteristic primate yellow pigment spot (Landrum & Bone, 2001). Since zeaxanthin is relatively more concentrated in the macula compared to lutein (Bone et al., 1988), the distribution pattern observed in our study (Figure S9) indicates that carotene 3 corresponds to zeaxanthin and carotene diol 1 to lutein. Furthermore, the finding also validates that the tissue samples were dissected from the correct retinal region.

The function of these yellow macular pigments has not been firmly established yet, although their natural antioxidative properties are proposed to protect the retina from light-induced oxidative damage. In this context, it is interesting that we also identified ergothioneine, a diet-derived, colourless xenobiotic with antioxidative properties (Halliwell et al., 2018), as the third most enriched xenobiotic in the primate macula—lending further credence to the hypothesis of macula-specific anti-oxidative mechanisms. This concept is further supported by the macular enrichment of glutathione (Table S1), which is an endogenously generated antioxidative compound.

The fourth most enriched xenobiotic in the macula, beta-guanidinopropanoate, is known to decrease intracellular creatine and phosphocreatine levels and, in skeletal muscle, to increase fatigue tolerance (Oudman et al., 2013). Considering the important role of phosphocreatine in photoreceptor energy metabolism (Linton et al., 2010; Wallimann et al., 2011), it is plausible that the specific accumulation of beta-guanidinopropanoate contributes to the fine-tuning of energy usage in the macula.

One of the most striking observations in our study was the reduced abundance of many lipid groups in the macula compared to the periphery. Since many of the lipids detected in our study are membrane constituents, it is likely that this differential distribution is linked to the anatomy of the retina. More specifically, photoreceptors contain more lipids than other cells in the retina because their outer segments consist of densely stacked disc-shaped membranes. The thickness of the outer segment layer is more or less uniform across the entire retina, whilst the inner retina (composed of four major neuronal subtypes) approximately doubles in thickness in the macula compared to the periphery. As all samples in this study (full-thickness retinal punches) were normalised, this roughly halves the relative contribution of outer segments (from rods and cones) in the macular sample. Furthermore, the molar ratio of phosphatidylethanolamine to phosphatidylcholine in disk membranes is much higher (1:1) than in normal plasma membranes (1:6) (Boesze-Battaglia & Schimmel, 1997), which explains the relatively low abundance of phosphatidylethanolamine in the macula, compared to phosphatidylcholine. By extension, it is plausible that the pronounced relative reduction of sphingomyelin and ceramide abundance in the macula is also based on enrichment of these lipids in photoreceptor outer segments, although this has yet to be validated.

The second-most depleted non-lipid compound in the macula was taurine, which is known to be highly enriched in the retina, more so than any other bodily tissue (Ripps & Shen, 2012). Photoreceptors accumulate taurine and critically depend on it for long-term survival (Rascher et al., 2004; Ripps & Shen, 2012; Schmidt et al., 1976). Thus, as in the case of phosphatidylethanolamine described above, the relative depletion of taurine in the macula can be attributed to its known enrichment in photoreceptors. Consequently, we may speculate that putrescine (the most depleted macula non-lipid metabolite) and the polyamine family metabolites (the most depleted macula non-lipid metabolic group) are overrepresented in photoreceptors. However, supporting evidence for this (Ientile et al., 1986) is scarce. The spatial distribution of cones and rods might also contribute to the differential abundance of the phosphorylated versus non-phosphorylated purines such as GDP/GTP and guanosine as these metabolites are important components of the visual signalling cascade and might reflect activity differences between cones versus rods in the light-adapted retina.

The most enriched metabolite in the macula was NAAG, which is the third-most-prevalent neurotransmitter in the mammalian nervous system (Neale et al., 2000) after glutamate and taurine. Although the physiological function of NAAG in the retina is not well understood, its synthesis in and release from retinal ganglion cells in mammals and birds is well established by immunohistochemistry (Anderson et al., 1987; Williamson et al., 1991) and by the enrichment

Table 1 Abundance pattern cluster descriptions

Cluster name	Description	Significant pathways
Steep enrichment	Metabolites whose abundance very quickly increases when moving from the periphery to the macula of the retina. Differences are consistent in direction and significant between all three areas	Alanine and Aspartate Metabolism; Histidine Metabolism; Methionine, Cysteine, SAM and Taurine Metabolism
Shallow enrichment	Metabolites whose abundance slowly increases when moving from the periphery to the macula of the retina. A significant difference is only perceived when comparing macula to periphery	Glutamate Metabolism; Xenobiotics; Carbohydrate Metabolism (other); Glycine, Serine and Threonine Metabolism
Macula enriched	Metabolites whose abundance is particularly elevated only in the macula	TCA Cycle; Fatty Acid Mtb (Carnitine); Lysophospholipid; Lysine Metabolism
Periphery depleted	Metabolites whose abundance is particularly depleted only in the periphery	none
Steep depletion	Metabolites whose abundance very quickly decreases when moving from the periphery to the macula of the retina	Phosphatidylethanolamine (PE); Ceramides; Polyamine Metabolism
Shallow depletion	Metabolites whose abundance slowly decreases when moving from the periphery to the macula of the retina. A significant difference is only perceived when comparing macula to periphery	none
Periphery enriched	Metabolites whose abundance is particularly elevated only in the periphery	Sphingomyelin; Benzoate Metabolism
Macula depleted	Metabolites whose abundance is particularly depleted only in the macula	Diacylglycerol; Phospholipid Metabolism
Temporal enriched	Metabolites whose abundance is particularly elevated only in the temporal region	Cofactors & Vitamins; Monoacylglycerol; Lysophospholipid
Temporal depleted	Metabolites whose abundance is particularly depleted only in the temporal region	none

This table represents the ten identified pattern clusters that divide biochemical families according to their different abundances across retinal areas. A description for each cluster is provided. Lastly, metabolic families that were significant for each cluster (Table S1) are presented in the last column. More detailed metabolic abundances per area divided by pattern clusters are provided in Figure S6

of the NAAG producing enzymes (N-acetylaspartate synthetase and N-acetylaspartylglutamate synthase) visible in published single-cell transcription profiles (Swamy et al., 2021). This suggests that the increased abundance of NAAG is due to the much higher numbers of retinal ganglion cells in the macula versus the periphery. These results show that future studies that allow single-cell metabolomics are needed. These will be able to identify cell type-specific metabolite repertoires and will alleviate the problem of unknown cell type proportion contributions to these results (Seydel, 2021).

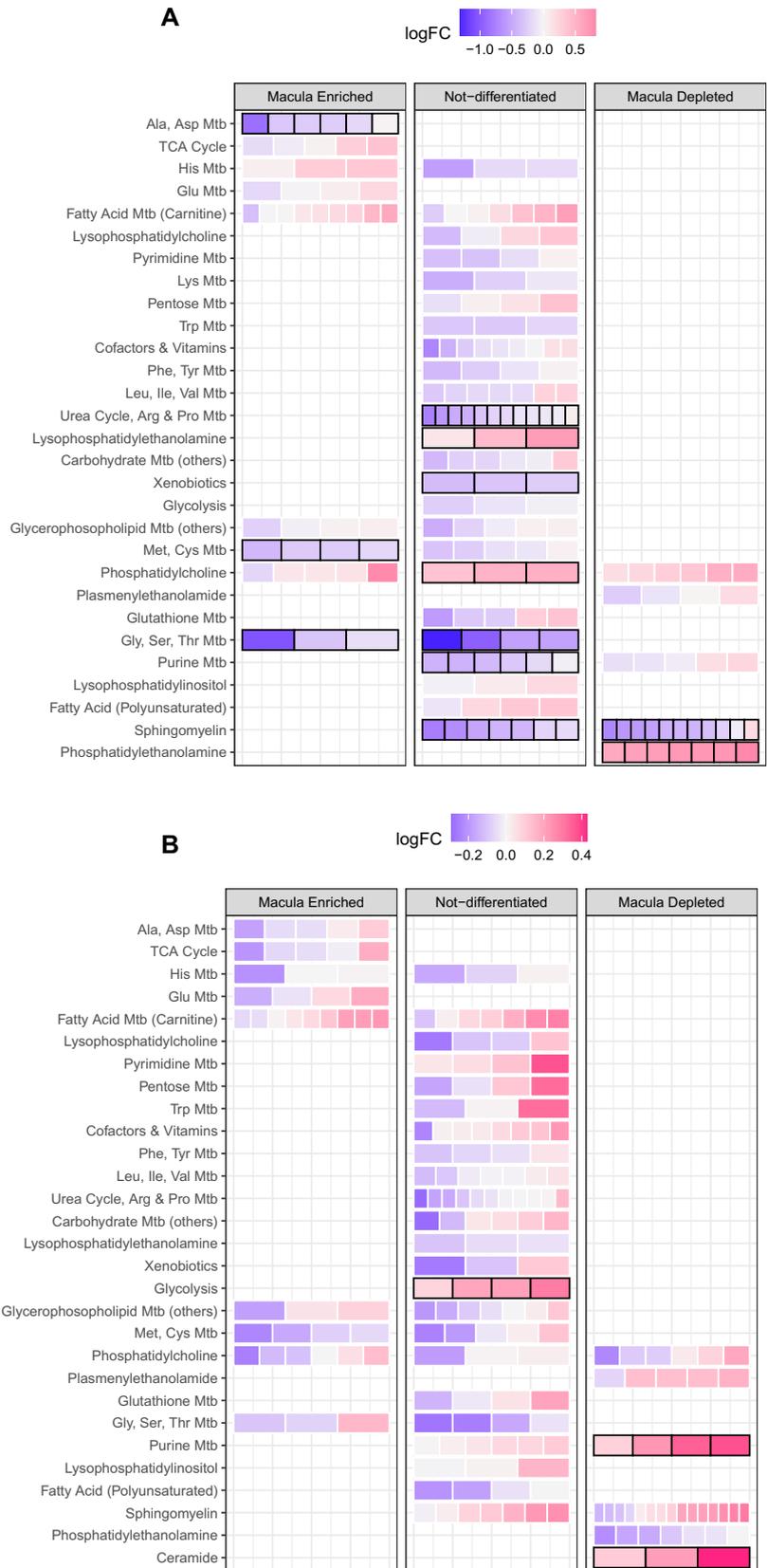
Considering the important roles specific metabolites play in cytoarchitecture and cellular function it is arguably not surprising that we found in this study over half of all identified metabolites were differentially distributed in at least one comparison. This appears to apply in particular to metabolites associated with retinal ganglion cells, rods and cones, which display the most extreme spatial distribution gradients across the retina. In contrast, we would expect metabolites that are linked to cell types with a more even spread, such as Müller cells or bipolar cells, to show less pronounced abundance differences in our three sampling locations. An illustrative example of this may be gamma-aminobutyrate

(GABA) and glutamine which are both evenly distributed (Table S1) and which are part of dominant metabolic pathways in Müller cells (Bringmann et al., 2013).

In this study, we were unable to analyse the retinal pigment epithelium, which acts as a major metabolic support hub for the retina (Hass et al., 2022; Hurley, 2021). Future studies on the metabolic composition of the RPE may provide further insight regarding the differential spatial distribution of metabolites in the RPE and their potential relationship to retinal health.

To explore the relevance of regional metabolic phenotypes in the retina in the context of disease, we explored the potential overlap between our primate retina dataset and serum metabolomics data from patients with retinal eye diseases. We chose to focus on MacTel and AMD because both of these diseases are associated with systemic metabolic changes and both manifest themselves region-specifically in the macula. The retina and the serum studies were all based on the same metabolomics platform (Metabolon) and the bioinformatic pre-processing of all datasets was uniform, enabling meaningful comparisons whereas other studies often have this as a confounder. This approach revealed that the metabolic pathways that were changed in the serum of

Fig. 4 Area Region specific metabolic pathways enrichment results for two retinal diseases: **A** MacTel and **B** AMD. Each row represents a biological pathway that contains metabolites either enriched, not differentiated or depleted in the macula of the primates compared to the periphery. Metabolites are presented as “bricks” in each row. The colour of each metabolite represents the log-fold changes of that metabolite between MacTel cases and controls. High log-fold change indicates that the metabolite is more abundant in the cases compared to controls while negative values indicate a depletion of such metabolite. Sub-pathways significantly enriched (FDR < 0.05 for MacTel and nominal p < 0.05 for AMD) present a thick black stroke around them. There were no metabolites in the AMD study that achieved FDR < 0.05



MacTel patients and that also displayed a region-specific distribution in the healthy retina were in the alanine/asparagine metabolism, parts of glycine/serine metabolism, the phosphatidylethanolamine metabolism and the sphingomyelin metabolism. In AMD, we found that the purine and ceramide metabolic pathways showed retinal region-specificity (in the healthy retina) as well as changes in patient serum.

Although the overlaps between healthy primate retina and human patient serum do not establish causality or imply a functional role in pathobiological mechanisms, they are likely to aid a better understanding of why diseases such as MacTel and AMD primarily affect the macula.

4 Materials and methods

4.1 Sample collection and processing

4.1.1 Macaque retina collection and processing

Metabolic abundances were measured in the retinas of 6 healthy adult primates (*Macaca fascicularis*). Postmortem retinal tissue was collected at the California National Primate Research Center (Animal Welfare Assurance ID: D16-00272 (A3433-01), Public Health Service Policy on Humane Care and Use of Laboratory Animals Policy). For all animals but one, both eyes were included in the study for a total of 11 retinas. Fresh neural retinal tissue in the light-adapted state was dissected 10–20 min after euthanasia from three different eccentricities from the fovea. The yellow macular pigment spot (macula lutea) was clearly visible in the dissected retinas and used to locate the fovea, from where a piece of retinal tissue was cut out (centred on the fovea). The diameter of the sample was equivalent to the distance between the fovea and the temporal edge of the optic disc. A second, adjacent sample of the same size was taken temporally to the foveal sample. A third sample (same size) was taken further temporally in the peripheral retina, resulting in a total of 33 samples. The primates presented an average age of 2.1 years ($sd = 0.94$) and were divided into 3 females and 3 males. Samples were collected and immediately flash frozen on 3 separate dates and then submitted to Metabolon Inc. (Durham, USA) for mass spectrometry analysis. Briefly, this involved analysis of five fractions per sample: two for analysis by two separate reverse phase (RP)/UPLC-MS/MS methods with positive ion mode electrospray ionization (ESI); one for analysis by RP/UPLC-MS/MS with negative ion mode ESI; and one for analysis by HILIC/UPLC-MS/MS with negative ion mode ESI. Raw data was extracted, peak-identified and QC processed using Metabolon's hardware and software. Compounds were identified by comparison to library entries of purified standards and peaks were

quantified using the area-under-the-curve technique, providing relative abundances of 435 metabolites.

4.1.2 Human blood collection and processing

To assess the utility of our results when exploring systemic metabolic profiles of retinal disorders, we collected metabolic abundances from plasma of 351 participants. Of these 205 were AMD patients, while the rest were age-matched controls without AMD. AMD patients were divided into three sub-disease categories according to their retinal diagnosis, patients with choroidal neovascularization (CNV), geographic atrophy (GA) and patients with CNV and GA (mixed). This data consisted of 127 CNV, 45 GA and 33 Mixed AMD patients. The metabolic measurements were processed in 12 batches. The participants presented an average age of 78.2 years ($sd = 7.31$) with 54% females and 46% males. For each sample, we received abundances for 1403 metabolites, 431 of which were discarded a priori given that they were not defined as specific metabolites by Metabolon Inc.. Metabolic missingness rate was 14% ($sd = 27%$) among all samples with a similar missingness rate between healthy individuals (13.8%) and all the AMD cases (CNV 13.7%, GA 13.5%, Mixed 14.4%). Missingness varied across metabolites (Table S3).

4.2 Data pre-processing and imputation

Given their strongly skewed distribution, metabolic abundances were log-transformed to achieve distribution symmetry. Metabolic abundances were normalised by dividing the global median log-abundance of area/batch combination. To validate that samples were dissected from the correct locations in the retina we monitored the abundance of the metabolites carotene diol 1 and 3. Although Metabolon has not yet directly assigned the two carotenoids to specific structures, carotene diol 1 and 3 almost certainly represent the macular pigments lutein and zeaxanthin, respectively. The distribution of these two metabolites across the three sample locations matches well with the known enrichment of lutein and zeaxanthin in the macula. Furthermore, zeaxanthin is known to have a "sharper peak" than lutein in the fovea, when comparing macula versus periphery (Figure S9).

The metabolic missingness rate was 26% ($sd = 34%$) among all samples with an increasing missingness rate between the areas (24% macula; 25.7% temporal, 28.4% periphery). Missingness in metabolomics study is usually assumed to arise from two different mechanisms, either Missingness At Random (MAR, the missingness of the metabolite does not depend on the value of the metabolite itself) or Missingness Not At Random (MNAR, the missingness of the metabolite depends on the value of the metabolite itself). Imputation of metabolic abundance

depends on such the assumption of mechanism, however, no common consensus has yet been reached on what type should be assumed (Bingol, 2018; Do et al., 2018; Wei et al., 2018a, b). Given the aforementioned result it is plausible to assume that some metabolites in this study have retinal region specific abundance and additionally present as MNAR. Which metabolites were responsible for this effect was, however, unclear. To this end, we performed imputation of metabolic abundances in a flexible manner by using two different imputation approaches depending on the type of missingness that was most likely for each metabolite. Firstly, we discarded from further analyses 62 metabolites (Table S4) with missingness rates of $\geq 80\%$. Secondly, for each metabolite i with at least one missing value, we identified two auxiliary metabolites (J_i and Z_i) which had the two highest correlations with metabolite i (calculated over at least three observable abundances). The average correlation between metabolites and the first auxiliary variable J was $r = 0.92$ ($sd = 0.06$) while a correlation of $r = 0.89$ ($sd = 0.07$) was observed for the auxiliary variable Z . Thirdly, auxiliary variables were then used for each metabolite to determine whether the missingness for that metabolite was MAR or MNAR. To this end, we performed a T-test comparing the observed values of both auxiliary variables when the metabolite of interest was missing or non-missing. We could not perform such a test for 41 metabolites as these presented with either one or two missing values. If metabolic values of either auxiliary variable were identified as being significantly lower ($p < 0.05$) when metabolite i presented missing values then the missingness of metabolite i was considered MNAR. If neither of the two auxiliary variables were significantly lower or metabolite i had no auxiliary variables available it was considered to present as MAR. This resulted in 132 metabolites flagged as MAR and 37 as MNAR missingness. The metabolomics dataset was then imputed using the MetImp software v1.2 (<https://metabolomics.cc.hawaii.edu/software/MetImp/>) (Wei et al., 2018a, b; Wei et al., 2018a, b) using a MAR random forest approach which uses all available metabolic observations to impute missing values. Metabolites with MNAR missingness were imputed using the same tool with a Gibbs sampling approach. Both approaches have been used by several previous studies (Bingol, 2018; Do et al., 2018; Wei et al., 2018a, b).

We then proceeded to quantile normalise each sample to reduce the effect of potential confounding due to batch effect using the *NormaliseBetweenArrays* function of the Limma package v 3.44.3. We assessed the gain of biological separation between samples by comparing the first two metabolic principal components in the non-normalised data versus the normalised data (Figure S10 A-B). Investigation of preparation batches in the normalised data revealed an

effect of batch preparation on metabolic abundance which was orthogonal to the biological differences (Figure S10 C).

Two metabolites were discarded due to the lack of any variability. The macula sample of one animal was discarded from further analyses since it was identified as incorrectly labelled based on clustering in the PC plots and examination of its carotene diol 1 and 3 levels. The final dataset comprised 32 samples and 371 metabolites. Metabolites were divided into 34 biological pathway groups. The list of metabolites and respective pathway membership is available in Table S5.

The data processing of the human serum metabolic data for the AMD study is presented in Supplementary Methods and are similar to those described above as well as a previously reported MacTel metabolomics study (Bonelli et al., 2020).

5 Statistical analysis

To test for differential abundance between different areas of the retina we adopted the same strategy as in our previous MacTel metabolite study (Bonelli et al., 2020). This strategy involved the usage of the Limma software for gene expression analysis which exploits multivariate linear regression combined with the empirical Bayes approach (Ritchie et al., 2006, 2015; Smyth, 2005). Metabolites were average-centred and standard deviation scaled. Primate age and gender, as well as preparation batch, were included as covariates in the model. Intra-primate correlation was taken into account by considering samples as biological replicates of the same primate using the function *duplicateCorrelation* in Limma (Smyth et al., 2005). For each metabolite, we tested three contrasts: *Macular vs Temporal*, *Temporal vs Periphery* and *Macula vs Periphery*. For this study, we used a false discovery rate cut-off of 5%. All metabolites with a Benjamini–Hochberg corrected p-value less than the FDRcut off of 0.05 were considered significant.

To test for pathway differential abundance between retinal areas we tested for enrichment of differential abundance in the pathways by using the *fry* function from the R/Limma package. The reason behind using these two different approaches has been described elsewhere (Bonelli et al., 2020). Additionally, we prepared global pathway abundance by calculating the first principal component on all metabolites in each pathway. The specific methodology to perform this has been described elsewhere (Bonelli et al., 2020).

Metabolites and metabolic pathways were then divided into clusters reflecting the patterns of significance and effect direction between contrasts (*Macular vs Temporal*, *Temporal vs Periphery*, *Macula vs Periphery*). A table of cluster names with contrast combinations is presented in Table S6.

To assess the utility of our primate metabolomic study results for the interpretation of human retinal disease we extracted the results from our previous MacTel study (Bonelli et al., 2020) as well as the new study presented in this manuscript for AMD. Each metabolic family was divided into three subgroups dividing metabolites that were either, “enriched”, “not-differentiated” or “depleted” in the macula compared to the rest of the retina. With the new definition, we then tested for enrichment in each newly defined pathway using the R/limma *fr* module (Giner & Smyth, 2016). Pathways with enrichment FDR < 0.05 were considered significant in MacTel. Given the very low level of significance in the AMD study, a less stringent threshold of a nominal p-value < 0.05 was used instead for the enrichment analysis of this disorder.

Statistical analysis methodology used to analyse the serum metabolites of human samples comparing AMD to healthy controls is presented in Supplementary Methods.

Supplementary Information The online version contains supplementary material available at <https://doi.org/10.1007/s11306-022-01969-6>.

Acknowledgements We thank Prof Tony Moore and Prof John Yates for their help with samples and data collection from AMD patients. We thank Dr Valentina Cipriani for assisting with matching the serum donor demographics. Finally, we thank patients and their families for their involvement in this study.

Author contributions Conceptualization: MF, Methodology: SMW RB MF, Software: RB, Formal analysis: RB BREA MF, Investigation: RB, BREA MB MF, Data collection: SMW PNB KNK, Resources: RB SMW SL MF, Data Curation: SMW SL, Writing – Original, Draft: RB BREA MF, Writing—Review & Editing: RB BREA MB MF, Visualization: RB, Supervision: MB BREA MF, Project administration: BREA MF, Funding acquisition: MB MF.

Funding The authors would like to thank the Lowy Medical Research Institute for funding to conduct this study. BREA was supported by Australian National Health and Medical Research Council (NHMRC) Fellowship (GNT1157776) and MB by an NHMRC Investigator Grant (GNT1195236). This work was also supported by the Australian State of Victoria’s Government’s Operational Infrastructure Support Program, the NHMRC Independent Research Institute Infrastructure Support Scheme (IRIIS) and the NIHR Moorfields Biomedical Research Centre.

Data availability Primate retinal metabolomic data and human plasma metabolomic data are available on request.

Declarations

Conflict of interest All authors certify that they have no affiliations with or involvement in any organization or entity with any financial interest or non-financial interest in the subject matter or materials discussed in this manuscript.

Open Access This article is licensed under a Creative Commons Attribution 4.0 International License, which permits use, sharing, adaptation, distribution and reproduction in any medium or format, as long as you give appropriate credit to the original author(s) and the source, provide a link to the Creative Commons licence, and indicate if changes

were made. The images or other third party material in this article are included in the article's Creative Commons licence, unless indicated otherwise in a credit line to the material. If material is not included in the article's Creative Commons licence and your intended use is not permitted by statutory regulation or exceeds the permitted use, you will need to obtain permission directly from the copyright holder. To view a copy of this licence, visit <http://creativecommons.org/licenses/by/4.0/>.

References

- Acar, İE., Lores-Motta, L., Colijn, J. M., Meester-Smoor, M. A., Verzijden, T., Cougnard-Gregoire, A., Ajana, S., Merle, B. M. J., de Breuk, A., Heesterbeek, T. J., van den Akker, E., Daha, M. R., Claes, B., Pauleikhoff, D., Hense, H.-W., van Duijn, C. M., Fauser, S., Hoyng, C. B., Delcourt, C., EYE-RISK Consortium. (2020). Integrating metabolomics, genomics, and disease pathways in age-related macular degeneration: The EYE-RISK consortium. *Ophthalmology*, 127(12), 1693–1709.
- Anderson, K. J., Borja, M. A., Cotman, C. W., Moffett, J. R., Namboodiri, M. A., & Neale, J. H. (1987). N-acetylaspartylglutamate identified in the rat retinal ganglion cells and their projections in the brain. *Brain Research*, 411(1), 172–177.
- Bingol, K. (2018). Recent advances in targeted and untargeted metabolomics by NMR and MS/NMR methods. *High-Throughput*, 7(2), 9. <https://doi.org/10.3390/ht7020009>
- Boesze-Battaglia, K., & Schimmel, R. (1997). Cell membrane lipid composition and distribution: Implications for cell function and lessons learned from photoreceptors and platelets. *The Journal of Experimental Biology*, 200(Pt 23), 2927–2936.
- Bone, R. A., Landrum, J. T., Fernandez, L., & Tarsis, S. L. (1988). Analysis of the macular pigment by HPLC: Retinal distribution and age study. *Investigative Ophthalmology & Visual Science*, 29(6), 843–849.
- Bonelli, R., Woods, S. M., Ansell, B. R. E., Heeren, T. F. C., Egan, C. A., Khan, K. N., Guymer, R., Trombley, J., Friedlander, M., Bahlo, M., & Fruttiger, M. (2020). Systemic lipid dysregulation is a risk factor for macular neurodegenerative disease. *Scientific Reports*, 10(1), 1–13.
- Bonelli, R., Ansell, B. R. E., Lotta, L., Scerri, T., Clemons, T. E., Leung, I., Peto, T., Bird, A. C., Sallo, F. B., Langenberg, C., Bahlo, M., The MacTel Consortium. (2021). Genetic disruption of serine biosynthesis is a key driver of macular telangiectasia type 2 aetiology and progression. *Genome Medicine*, 13(1), 39.
- Bonelli, R., Jackson, V. E., Prasad, A., Munro, J. E., Farashi, S., Heeren, T. F. C., Pontikos, N., Schepke, L., Friedlander, M., MacTel Consortium, Egan, C. A., Allikmets, R., Ansell, B. R. E., & Bahlo, M. (2021). Identification of genetic factors influencing metabolic dysregulation and retinal support for MacTel, a retinal disorder. *Communications Biology*, 4(1), 274.
- Bringmann, A., Grosche, A., Pannicke, T., & Reichenbach, A. (2013). GABA and glutamate uptake and metabolism in retinal glial (Müller) cells. *Frontiers in Endocrinology*, 4, 48.
- Brown, C. N., Green, B. D., Thompson, R. B., den Hollander, A. I., Lengyel, I., EYE-RISK consortium. (2018). Metabolomics and age-related macular degeneration. *Metabolites*, 9(1), 4. <https://doi.org/10.3390/metabo9010004>
- Do, K. T., Wahl, S., Raffler, J., Molnos, S., Laimighofer, M., Adamski, J., Suhre, K., Strauch, K., Peters, A., Gieger, C., Langenberg, C., Stewart, I. D., Theis, F. J., Grallert, H., Kastenmüller, G., & Krumsiek, J. (2018). Characterization of missing values in untargeted MS-based metabolomics data and evaluation of missing

- data handling strategies. *Metabolomics: Official Journal of the Metabolomic Society*, 14(10), 128.
- Gantner, M. L., Eade, K., Wallace, M., Handzlik, M. K., Fallon, R., Trombley, J., Bonelli, R., Giles, S., Harkins-Perry, S., Heeren, T. F. C., Sauer, L., Ideguchi, Y., Baldini, M., Scheppke, L., Dorrell, M. I., Kitano, M., Hart, B. J., Cai, C., Nagasaki, T., ... Friedlander, M. (2019). Serine and lipid metabolism in macular disease and peripheral neuropathy. *The New England Journal of Medicine*. <https://doi.org/10.1056/NEJMoa1815111>
- Giner, G., & Smyth, G. K. (2016). FRY: A fast approximation to ROAST gene set test with mean aggregated set statistics. *F1000Research*. <https://doi.org/10.7490/f1000research.1113351.1>
- Halliwell, B., Cheah, I. K., & Tang, R. M. Y. (2018). Ergothioneine—a diet-derived antioxidant with therapeutic potential. *FEBS Letters*, 592(20), 3357–3366.
- Hass, D. T., Bisbach, C. M., Robbins, B. M., Sadilek, M., Sweet, I. R., & Hurley, J. B. (2022). Succinate metabolism in the retinal pigment epithelium uncouples respiration from ATP synthesis. *Cell Reports*, 39(10), 110917.
- Hu, Y., Wang, X., Hu, B., Mao, Y., Chen, Y., Yan, L., Yong, J., Dong, J., Wei, Y., Wang, W., Wen, L., Qiao, J., & Tang, F. (2019). Dissecting the transcriptome landscape of the human fetal neural retina and retinal pigment epithelium by single-cell RNA-seq analysis. *PLoS Biology*, 17(7), e3000365.
- Hurley, J. B. (2021). Retina metabolism and metabolism in the pigmented epithelium: A busy intersection. *Annual Review of Vision Science*, 7, 665–692.
- Ientile, R., Russo, P., & Macaione, S. (1986). Polyamine localization and biosynthesis in chemically fractionated rat retina. *Journal of Neurochemistry*, 47(5), 1356–1360.
- Laíns, I., Chung, W., Kelly, R. S., Gil, J., Marques, M., Barreto, P., Murta, J. N., Kim, I. K., Vavvas, D. G., Miller, J. B., Silva, R., Lasky-Su, J., Liang, L., Miller, J. W., & Husain, D. (2019). Human plasma metabolomics in age-related macular degeneration: Meta-analysis of two cohorts. *Metabolites*, 9(7), 127. <https://doi.org/10.3390/metabo9070127>
- Laíns, I., Duarte, D., Barros, A. S., Martins, A. S., Gil, J., Miller, J. B., Marques, M., Mesquita, T., Kim, I. K., da Cachulo, M. L., Vavvas, D., Carreira, I. M., Murta, J. N., Silva, R., Miller, J. W., Husain, D., & Gil, A. M. (2017). Human plasma metabolomics in age-related macular degeneration (AMD) using nuclear magnetic resonance spectroscopy. *PLoS One*, 12(5), e0177749.
- Landrum, J. T., & Bone, R. A. (2001). Lutein, zeaxanthin, and the macular pigment. *Archives of Biochemistry and Biophysics*, 385(1), 28–40.
- Linton, J. D., Holzhausen, L. C., Babai, N., Song, H., Miyagishima, K. J., Stearns, G. W., Lindsay, K., Wei, J., Chertov, A. O., Peters, T. A., Caffè, R., Pluk, H., Seeliger, M. W., Tanimoto, N., Fong, K., Bolton, L., Kuok, D. L. T., Sweet, I. R., Bartoletti, T. M., ... Hurley, J. B. (2010). Flow of energy in the outer retina in darkness and in light. *Proceedings of the National Academy of Sciences of the United States of America*, 107(19), 8599–8604.
- Neale, J. H., Bzdega, T., & Wroblewska, B. (2000). N-Acetylaspartylglutamate: The most abundant peptide neurotransmitter in the mammalian central nervous system. *Journal of Neurochemistry*, 75(2), 443–452.
- Oudman, I., Clark, J. F., & Brewster, L. M. (2013). The effect of the creatine analogue beta-guanidinopropionic acid on energy metabolism: A systematic review. *PLoS One*, 8(1), e52879.
- Rascher, K., Servos, G., Berthold, G., Hartwig, H.-G., Warskulat, U., Heller-Stilb, B., & Häussinger, D. (2004). Light deprivation slows but does not prevent the loss of photoreceptors in taurine transporter knockout mice. *Vision Research*, 44(17), 2091–2100.
- Ratnapriya, R., Sosina, O. A., Starostik, M. R., Kwicklis, M., Kaphahn, R. J., Fritsche, L. G., Walton, A., Arvanitis, M., Gieser, L., Pietraszkiewicz, A., Montezuma, S. R., Chew, E. Y., Battle, A., Abecasis, G. R., Ferrington, D. A., Chatterjee, N., & Swaroop, A. (2019). Retinal transcriptome and eQTL analyses identify genes associated with age-related macular degeneration. *Nature Genetics*. <https://doi.org/10.1038/s41588-019-0351-9>
- Ripps, H., & Shen, W. (2012). Review: Taurine: A “very essential” amino acid. *Molecular Vision*, 18, 2673–2686.
- Ritchie, M. E., Diyagama, D., Neilson, J., van Laar, R., Dobrovic, A., Holloway, A., & Smyth, G. K. (2006). Empirical array quality weights in the analysis of microarray data. *BMC Bioinformatics*, 7, 261.
- Ritchie, M. E., Phipson, B., Wu, D., Hu, Y., Law, C. W., Shi, W., & Smyth, G. K. (2015). limma powers differential expression analyses for RNA-sequencing and microarray studies. *Nucleic Acids Research*, 43(7), e47.
- Scerri, T. S., Quagliari, A., Cai, C., Zernant, J., Matsunami, N., Baird, L., Scheppke, L., Bonelli, R., Yannuzzi, L. A., Friedlander, M., Egan, C. A., Fruttiger, M., Leppert, M., Allikmets, R., & Bahlo, M. (2017). Genome-wide analyses identify common variants associated with macular telangiectasia type 2. *Nature Genetics*. <https://doi.org/10.1038/ng.3799>
- Schmidt, S. Y., Berson, E. L., & Hayes, K. C. (1976). Retinal degeneration in cats fed casein. *I. Taurine Deficiency. Investigative Ophthalmology*, 15(1), 47–52.
- Seydel, C. (2021). Single-cell metabolomics hits its stride. *Nature Methods*, 18(12), 1452–1456.
- Smyth, G. K. (2005). limma: Linear Models for Microarray Data. In R. Gentleman, V. J. Carey, W. Huber, R. A. Irizarry, & S. Dudoit (Eds.), *Bioinformatics and Computational Biology Solutions Using R and Bioconductor* (pp. 397–420). Springer.
- Smyth, G. K., Michaud, J., & Scott, H. S. (2005). Use of within-array replicate spots for assessing differential expression in microarray experiments. *Bioinformatics*, 21(9), 2067–2075.
- Swamy, V. S., Fufa, T. D., Hufnagel, R. B., & McGaughey, D. M. (2021). Building the mega single-cell transcriptome ocular meta-atlas. *GigaScience*, 10(10), giab061. <https://doi.org/10.1093/gigascience/giab061>
- Tyynismaa, H. (2019). A metabolic vulnerability of vision. *The New England Journal of Medicine*. <https://doi.org/10.1056/NEJMe1910322>
- Voigt, A. P., Binkley, E., Flamme-Wiese, M. J., Zeng, S., DeLuca, A. P., Scheetz, T. E., Tucker, B. A., Mullins, R. F., & Stone, E. M. (2020). Single-cell RNA sequencing in human retinal degeneration reveals distinct glial cell populations. *Cells*, 9(2), 438. <https://doi.org/10.3390/cells9020438>
- Voigt, A. P., Mulfaul, K., Mullin, N. K., Flamme-Wiese, M. J., Giacalone, J. C., Stone, E. M., Tucker, B. A., Scheetz, T. E., & Mullins, R. F. (2019). Single-cell transcriptomics of the human retinal pigment epithelium and choroid in health and macular degeneration. *Proceedings of the National Academy of Sciences of the United States of America*, 116(48), 24100–24107.
- Wallimann, T., Tokarska-Schlattner, M., & Schlattner, U. (2011). The creatine kinase system and pleiotropic effects of creatine. *Amino Acids*, 40(5), 1271–1296.
- Wei, R., Wang, J., Jia, E., Chen, T., Ni, Y., & Jia, W. (2018a). GSimp: A Gibbs sampler based left-censored missing value imputation approach for metabolomics studies. *PLoS Computational Biology*, 14(1), e1005973.
- Wei, R., Wang, J., Su, M., Jia, E., Chen, S., Chen, T., & Ni, Y. (2018b). Missing value imputation approach for mass spectrometry-based metabolomics data. *Scientific Reports*, 8(1), 663.
- Whitmore, S. S., Wagner, A. H., DeLuca, A. P., Drack, A. V., Stone, E. M., Tucker, B. A., Zeng, S., Braun, T. A., Mullins, R. F., & Scheetz, T. E. (2014). Transcriptomic analysis across nasal, temporal, and macular regions of human neural retina and RPE/choroid by RNA-Seq. *Experimental Eye Research*, 129, 93–106.

- Williamson, L. C., Eagles, D. A., Brady, M. J., Moffett, J. R., Nambodiri, M. A. A., & Neale, J. H. (1991). Localization and synaptic release of N-acetylaspartylglutamate in the chick retina and optic tectum. *The European Journal of Neuroscience*, 3(5), 441–451.
- Yan, W., Peng, Y.-R., van Zyl, T., Regev, A., Shekhar, K., Juric, D., & Sanes, J. R. (2020). Cell atlas of the human fovea and peripheral retina. *In bioRxiv*. <https://doi.org/10.1101/2020.02.11.943779>
- Yi, W., Lu, Y., Zhong, S., Zhang, M., Sun, L., Dong, H., Wang, M., Wei, M., Xie, H., Qu, H., Peng, R., Hong, J., Yao, Z., Tong, Y., Wang, W., Ma, Q., Liu, Z., Ma, Y., Li, S., & Xue, T. (2021). A single-cell transcriptome atlas of the aging human and macaque retina. *National Science Review*, 8(4), nwaal79. <https://doi.org/10.1093/nsr/nwaa179>

Publisher's Note Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.