

Whilst important to consider that other T cells such as Tregs, exhausted T cells and CD4⁻ exist within the CD3⁺/CD8⁻ population, given that the number of these cells were not of significance to alter the patterns found in our data or change the conclusions drawn they remained labelled as CD4⁺.

Double expression of CD38⁺HLA-DR⁺ on CD4⁺ cells was examined. Associated with increased T cell activity during active HIV infection [3] and predictors of disease progression in untreated patients [4], significantly elevated percentages of CD38⁺HLA-DR⁺ were expressed in CD4⁺ cells in those individuals with high-level rebound/non-responses. HLA-DR⁺ expression is indicative of T cell activation [13] whilst CD38 has numerous cellular functions involved with cell metabolism [14] and activity, and is expressed in up to 30% of CD4⁺ cells in healthy adults [13]. Elevated in HIV, CD4⁺CD38⁺ declines when patients demonstrate successful viral control following combination ART [15].

Co-expression of CD8⁺CD38⁺HLA-DR⁺ is reported to be indicative of both activation and proliferation of CD8⁺ T-cells, reflecting strong antiviral function by exhibiting higher effector functions including proliferation, cytotoxicity and cytokine production [16, 17] however their expression is not co-dependent [18]. During hyperacute HIV infection, up to 77% of peripheral CD8⁺ cells express both CD38⁺ and HLA-DR⁺ [19], contributing towards the setpoint for an individual. However, during chronic viral infections maintaining these levels of CD38⁺HLA-DR⁺ leads to increased expression of molecules related to exhaustion and apoptosis and both elevated percentages and absolute number of CD38⁺HLA-DR⁺ predict the progression to AIDS [20].

In individuals with high-level rebound/non-response unlike the other groups, CD8⁺CD38⁺HLA-DR⁺ continued to be strongly associated with viral load over 144 weeks and when normalised to circulating CD8⁺ cells, CD8⁺CD38⁺HLA-DR⁺ increased in number over time, which might suggest that as viral load comes under control, HLA-DR⁺/CD38⁺ is no longer co-expressed [21]. The strong correlation of CD8⁺CD38⁺HLA-DR⁺ with viral load in participants with high-level non-response/rebound by 144 weeks from initiation of second-line therapy has been associated with a poorer outcome [22] although which is the driving factor is unclear.

In those with high-level rebound/non-response more than half of CD8⁺ cells remained CD38 positive at week 144 although the percentages of CD8⁺ cells expressing CD38⁺ consistently decreased over the duration of the trial independent of viral load control.

CD38⁺ and HLA-DR⁺ expression are considered markers of activation that, either exclusively or co-expressed, correlate with viral load in non-controllers [23], although qualitative differences between HIV-1 specific CD8⁺ T cell responses, such as cytokine release, can define responses of controllers and those with progressive disease [24]. Elevated CD8⁺CD38⁺ and CD8⁺HLA-DR⁺ expression persists throughout HIV infection and has prognostic significance for progression onto AIDS [23, 25].

Over the duration of the trial, whilst the percentages of CD4⁺CD38⁺ and CD4⁺ CD38⁺HLADR⁺ T-cells decreased in all groups except high-level rebound, numbers of circulating CD4⁺CD38⁺ increased in all VL response groups when normalised to an increasing population of circulating CD4⁺ T-cells. Thus, although the percentage of activated cells decreased with functioning ART, this was offset by an overall increase in T cell number leading to a larger pool of active cells. Therefore in absolute terms, it could be argued that ART does not lead to reduced activation. Interestingly, and potentially important, CD8⁺ cells responded more quickly to fluctuations in viral replication [15], and generally demonstrated larger differences than CD4⁺ parameters when comparing the high-level rebound/non-response group and other groups. This may be an important factor for long term survival of this population.

In summary, this study demonstrates that the main drivers of immune reconstitution and activation are most likely viral, rather than being driven by specific ART combinations. Immune reconstitution is impaired in high level rebound and non-responsive individuals with viral loads above 5000 copies/ml, which is well above the WHO guidelines recommend level for switching. In contrast, provided patients remain on ART, transient blips and low-level rebound have at most small effects on reconstitution and activation. These findings suggest that the current WHO viral load threshold for switching to second-line of 1000 copies/ml should avoid the most deleterious effects of high-level rebound, given that it is mostly identified through annual viral load monitoring in many low and middle income settings.

Limitations

In labelling our cells we have classified those CD3⁺ CD8⁻ as CD4⁺ cells and acknowledge that this population of CD8⁻ may be CD4⁻/CD8⁻. We maintain that as a percentage of cells analysed over time, our labelled CD8⁻ follow the same independently analysed patterns of total CD4⁺ count reported previously [2] and have therefore maintained the labelling of CD8⁻ as CD4⁺. In the analysis of the subsets, the significantly elevated percentages of CD38⁺ HLADR⁺ in CD8⁻ cells in patients with high viral load in this study are likely to be CD8⁻CD4⁺ and not CD8⁻CD4⁻ as this population decreases with high viral load [26].

Acknowledgements

We thank all the participants and staff from all the centres participating in the EARNEST trial. Members of the EARNEST Trial Team are:

Participating Sites

Uganda

JCRC Kampala (African trial co-ordinating centre; 231): E Agweng, P Awio, G Bakeinyaga, C Isabirye, U Kabuga, S Kasuswa, M Katuramu, C Kityo, F Kiweewa, H Kyomugisha, E Lutalo, P Mugenyi, D Mulima, H Musana, G Musitwa, V Musiime, M Ndigendawan, H Namata, J Nkalubo, P Ocitti Labejja, P Okello, P Olal, G Pimundu, P Segonga, F Ssali, Z Tamale, D Tumukunde, W Namala, R Byaruhanga, J Kayiwa, J Tukamushaba, S Abunyang,

D Eram, O Denis, R Lwalanda, L Mugarura, J Namusanje, I Nankya, E Ndashimye, E Nabulime, D Mulima, O Senfuma.

IDI, Kampala (216): G Bihabwa, E Buluma, P Easterbrook, A Elbireer, A Kambugu, D Kanya, M Katwere, R Kiggundu, C Komujuni, E Laker, E Lubwama, I Mambule, J Matovu, A Nakajubi, J Nakku, R Nalumenya, L Namuyimbwa, F Semitala, B Wandera, J Wanyama

JCRC, Mbarara (97): H Mugerwa, A Lugemwa, E Ninsiima, T Ssenkindu, S Mwebe, L Atwine, H William, C Katemba, S Abunyang, M Acaku, P Ssebutinde, H Kitizo, J Kukundakwe, M Naluguza, K Ssegawa, Namayanja, F Nsibuka, P Tuhirirwe, M Fortunate

JCRC Fort Portal (66): J Acen, J Achidri, A Amone, M. Chamai, J Ditai, M Kemigisa, M Kiconco, C Matama, D Mbanza, F Nambaziira, M Owor Odoi, A Rweyora, G. Tumwebaze

San Raphael of St Francis Hospital, Nsambya (48): H Kalanzi, J Katabaazi, A Kiyingi, M Mbidde, M. Mugenyi, R Mwebaze, P Okong, I Senoga

JCRC Mbale (47): M Abwola, D Baliruno, J Bwomezi, A Kasede, M Mudoola, R Namisi, F Ssenono, S Tuhirwe

JCRC Gulu (43): G Abongomera, G Amone, J Abach, I Aciro, B Arach, P Kidega, J Omongin, E Ocung, W Odong, A Phillipam

JCRC Kabale (33): H Alima, B Ahimbisibwe, E Atuhaire, F Atukunda, G Bekusike, A Bulegyeya, D. Kahatano, S Kamukama, J Kyoshabire, A Nassali, A Mbonye, T M Naturinda, Ndukukire, A Nshabohurira, H. Ntawiha, A Rogers, M Tibyasa;

JCRC Kakira (31): S. Kiirya, D. Atwongyeire, A. Nankya, C. Draleku, D. Nakiboneka, D. Odoch, L. Lakidi, R. Ruganda, R. Abiriga, M. Mulindwa, F. Balmoi, S. Kafuma, E. Moriku

Zimbabwe

University of Zimbabwe Clinical Research Centre, Harare (265): J Hakim, A Reid, E Chidziva, G Musoro, C Warambwa, G Tinago, S Mutsai, M Phiri, S Mudzingwa, T Bafana, V Masore, C Moyo, R Nhema, S Chitongo

Malawi

Department of Medicine, University of Malawi College of Medicine and the Malawi-Liverpool-Wellcome Trust Clinical Research Programme, University of Malawi College of Medicine (92): Robert Heyderman, Lucky Kabanga, Symon Kaunda, Aubrey Kudzala, Linly Lifa, Jane Mallewa, Mike Moore, Chrissie Mtali, George Musowa, Grace Mwimaniwa, Rosemary Sikwese, Joep van Oosterhout, Milton Ziwoya

Mzuzu Central Hospital, Mzuzu (19): H Chimbaka, B Chitete, S Kamanga, T Kayinga, E Makwakwa, R Mbiya, M Mlenga, T Mphande, C Mtika, G Mushani, O Ndhlovu, M Ngonga, I Nkhana, R Nyirenda

Kenya

Moi Teaching and Referral Hospital (52): P Cheruiyot, C Kwobah, W Lokitala Ekiru, M Mokaya, A Mudogo, A Nzioka, A Siika, M Tanui, S Wachira, K Wools-Kaloustian

Zambia

University Teaching Hospital (37): P Alipalli, E Chikatula, J Kipaila, I Kunda, S Lakhi, J Malama, W Mufwambi, L Mulenga, P Mwaba, E Mwamba, A Mweemba, M Namfukwe

The Aids Support Organisation (TASO), Uganda: E Kerukadho, B Ngwatu, J Birungi

MRC Clinical Trials Unit: N Paton, J Boles, A Burke, L Castle, S Ghuman, L Kendall, A Hoppe, S Tebbs, M Thomason, J Thompson, S Walker, J Whittle, H Wilkes, N Young

Monitors: C Kapuya, F Kyomuhendo, D Kyakundi, N Mkandawire, S Mulambo, S Senyonjo

Clinical Expert Review Committee: B Angus, A Arenas-Pinto, A Palfreeman, F Post, D Ishola

European Collaborators:

J Arribas (Hospital La Paz, Madrid, Spain), R Colebunders (Institute of Tropical Medicine, Antwerp, Belgium), M Floridia (ISS, Italy), M Giuliano (ISS, Italy), P Mallon (University College Dublin, Ireland), P Walsh (University College Dublin, Ireland), M De Rosa (CINECA, Italy), E Rinaldi (CINECA, Italy)

Trial Steering Committee: I Weller (Chair), C Gilks, J Hakim, A Kangewende, S Lakhi, E Luyirika, F Miiro, P Mwamba, P Mugenyi, S Ojoo, N Paton, S Phiri, J van Oosterhout, A Siika, S Walker, A Wapakabulo,

Data Monitoring Committee: T Peto (Chair), N French, J Matenga

Pharmaceutical companies: G Cloherty, J van Wyk, M Norton, S Lehrman, P Lamba, K Malik, J Rooney, W Snowden, J Villacian

Contributing Authors

Francesca I F ARRIGONI: design and performance

Moira SPYER: conception, design, and performance

Patricia HUNTER: design, and performance

Dagmar ALBER: design, and performance

Cissy KITYO: conception, design, and performance

James HAKIM: conception, design, and performance

Allen MATUBU: conception, design, and performance

Patrick OLAL: conception, design, and performance

Nicholas I PATON: conception, design, and performance

Sarah WALKER: conception, design, and performance

Nigel KLEIN : conception, design, and performance

References

1. Paton NI, Kityo C, Hoppe A, Reid A, Kambugu A, Lugemwa A, et al. **Assessment of second-line antiretroviral regimens for HIV therapy in Africa.** *The New England journal of medicine* 2014; 371(3):234-247.
2. Paton NI, Bagenda L, van Oosterhout JJ, Bertagnolio S, Easterbrook PJ, Walker AS, et al. **Nucleoside reverse-transcriptase inhibitor cross-resistance and outcomes from second-line antiretroviral therapy in the public health approach: an observational analysis within the randomised, open-label, EARNEST trial.** *The Lancet HIV* 2017; 4(8):e341-e348.
3. Meditz AL, Haas MK, Folkvord JM, Melander K, Young R, McCarter M, et al. **HLA-DR+ CD38+ CD4+ T lymphocytes have elevated CCR5 expression and produce the majority of R5-tropic HIV-1 RNA in vivo.** *J Virol* 2011; 85(19):10189-10200.
4. Deeks SG, Kitchen CM, Liu L, Guo H, Gascon R, Narváez AB, et al. **Immune activation set point during early HIV infection predicts subsequent CD4+ T-cell changes independent of viral load.** *Blood* 2004; 104(4):942-947.
5. Prendergast AJ, Arrigoni F, D A, Hunter P, Walker AS, Klein N. **The Impact of temporary and persistent viraemia on inflammatory biomarkers and CD4 cell subpopulations in HIV- infected children in sub-Saharan Africa.** 2019.

6. Song CB, Zhang LL, Wu X, Fu YJ, Jiang YJ, Shang H, et al. **CD4(+)CD38(+) central memory T cells contribute to HIV persistence in HIV-infected individuals on long-term ART.** *J Transl Med* 2020; 18(1):95.
7. Organisation WH. **Updated recommendations on first-line and second-line antiretroviral regimens and post-exposure prophylaxis and recommendations on early infant diagnosis of HIV.** In; 2018.
8. Hakim JG, Thompson J, Kityo C, Hoppe A, Kambugu A, van Oosterhout JJ, et al. **Lopinavir plus nucleoside reverse-transcriptase inhibitors, lopinavir plus raltegravir, or lopinavir monotherapy for second-line treatment of HIV (EARNEST): 144-week follow-up results from a randomised controlled trial.** *The Lancet Infectious diseases* 2018; 18(1):47-57.
9. de Martino M, Galli L, Chiarelli F, Rossi ME, Vierucci A. **Do nucleoside analogues directly influence T-lymphocyte subset counts? The pediatric model.** *JAIDS Journal of Acquired Immune Deficiency Syndromes* 1998; 18(4):391-392.
10. Serrano-Villar S, Sainz T, Lee SA, Hunt PW, Sinclair E, Shacklett BL, et al. **HIV-infected individuals with low CD4/CD8 ratio despite effective antiretroviral therapy exhibit altered T cell subsets, heightened CD8+ T cell activation, and increased risk of non-AIDS morbidity and mortality.** *PLoS Pathog* 2014; 10(5):e1004078.
11. McBride JA, Striker R. **Imbalance in the game of T cells: What can the CD4/CD8 T-cell ratio tell us about HIV and health?** *PLoS Pathog* 2017; 13(11):7.
12. Leung V, Gillis J, Raboud J, Cooper C, Hogg RS, Loutfy MR, et al. **Predictors of CD4:CD8 ratio normalization and its effect on health outcomes in the era of combination antiretroviral therapy.** *PloS one* 2013; 8(10):e77665.
13. Ramzaoui S, Jouen-Beades F, Gilbert D, Borsa-Lebas F, Michel Y, Humbert G, et al. **During HIV infection, CD4+ CD38+ T-cells are the predominant circulating CD4+ subset whose HLA-DR positivity increases with disease progression and whose V beta repertoire is similar to that of CD4+ CD38- T-cells.** *Clin Immunol Immunopathol* 1995; 77(1):33-41.
14. Chatterjee S, Chakraborty P, Mehrotra S. **CD38-NAD+-Sirt1 axis in T cell immunotherapy.** *Aging (Albany NY)* 2019; 11(20):8743.
15. Benito JM, Lopez M, Lozano S, Ballesteros C, Martinez P, Gonzalez-Lahoz J, et al. **Differential upregulation of CD38 on different T-cell subsets may influence the ability to reconstitute CD4+ T cells under successful highly active antiretroviral therapy.** *Journal of acquired immune deficiency syndromes (1999)* 2005; 38(4):373-381.

16. Miller JD, van der Most RG, Akondy RS, Glidewell JT, Albott S, Masopust D, et al. **Human effector and memory CD8+ T cell responses to smallpox and yellow fever vaccines.** *Immunity* 2008; 28(5):710-722.
17. Lindgren T, Ahlm C, Mohamed N, Evander M, Ljunggren HG, Bjorkstrom NK. **Longitudinal analysis of the human T cell response during acute hantavirus infection.** *J Virol* 2011; 85(19):10252-10260.
18. Psomas C, Younas M, Reynes C, Cezar R, Portalès P, Tuailon E, et al. **One of the immune activation profiles observed in HIV-1-infected adults with suppressed viremia is linked to metabolic syndrome: the ACTIVIH study.** *EBioMedicine* 2016; 8:265-276.
19. Ndhlovu ZM, Kanya P, Mewalal N, Klooverpris HN, Nkosi T, Pretorius K, et al. **Magnitude and Kinetics of CD8+ T Cell Activation during Hyperacute HIV Infection Impact Viral Set Point.** *Immunity* 2015; 43(3):591-604.
20. Kestens L, Vanham G, Gigase P, Young G, Hannel I, Vanlangendonck F, et al. **Expression of activation antigens, HLA-DR and CD38, on CD8 lymphocytes during HIV-1 infection.** *AIDS* 1992; 6(8):793-797.
21. Han J, Mu W, Zhao H, Hao Y, Song C, Zhou H, et al. **HIV-1 low-level viremia affects T cell activation rather than T cell development in school-age children, adolescents, and young adults during antiretroviral therapy.** *International Journal of Infectious Diseases* 2020; 91:210-217.
22. Collaboration ATC. **Causes of death in HIV-1—infected patients treated with antiretroviral therapy, 1996–2006: collaborative analysis of 13 HIV cohort studies.** *Clinical Infectious Diseases* 2010; 50(10):1387-1396.
23. Doisne JM, Urrutia A, Lacabartz-Porret C, Goujard C, Meyer L, Chaix ML, et al. **CD8+ T cells specific for EBV, cytomegalovirus, and influenza virus are activated during primary HIV infection.** *J Immunol* 2004; 173(4):2410-2418.
24. Hersperger AR, Migueles SA, Betts MR, Connors M. **Qualitative features of the HIV-specific CD8+ T cell response associated with immunologic control.** *Current Opinion in HIV and AIDS* 2011; 6(3):169.
25. Liu Z, Cumberland WG, Hultin LE, Prince HE, Detels R, Giorgi JV. **Elevated CD38 antigen expression on CD8+ T cells is a stronger marker for the risk of chronic HIV disease progression to AIDS and death in the Multicenter AIDS Cohort Study than CD4+ cell count, soluble immune activation markers, or combinations of HLA-DR and CD38 expression.** *J Acquir Immune Defic Syndr Hum Retrovirol* 1997; 16(2):83-92.

- 26, Liang, Qi, Yanmei Jiao, Tong Zhang, Rui Wang, Wei Li, Hongwei Zhang, Xiaojie Huang, Zhong Tang, and Hao Wu. **Double Negative (DN)[CD3 (+) CD4 (-) CD8 (-)] T cells correlate with disease progression during HIV infection.** *Immunological investigations* 42, no. 5 (2013): 431-437.

Figure 1. Number and proportion of activated CD4 and CD8 cells over time on second-line ART. Note: Showing mean plus 95% confidence interval for each randomised group at each timepoint.

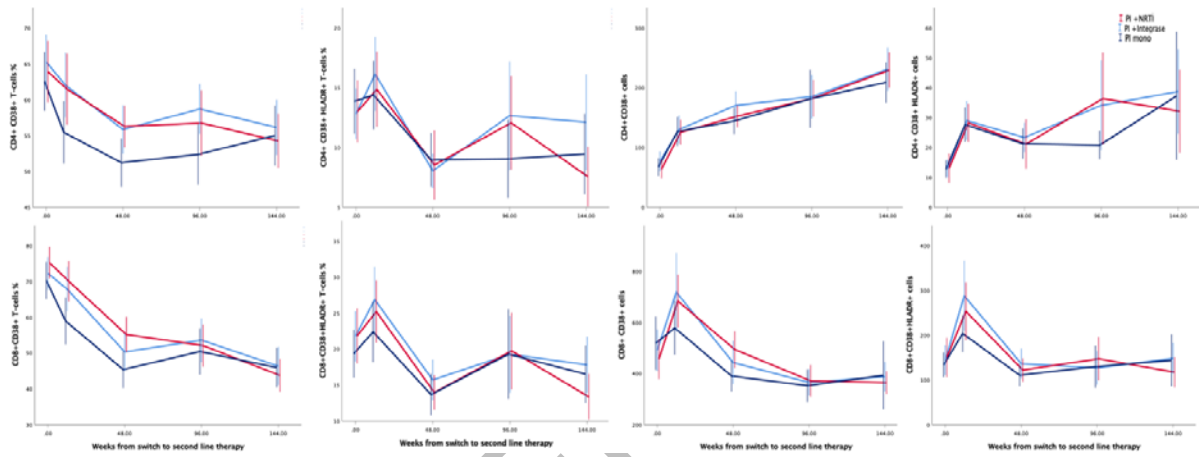


Figure 2. Viral load responses over time on second line therapy

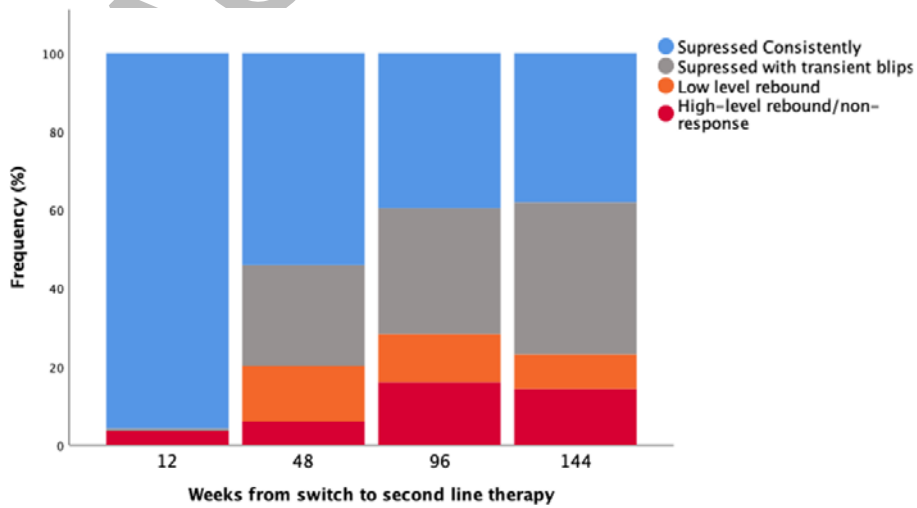


Figure 3: Changes in CD4 and activated CD4 sub-populations over time, expressed both as percentages and normalised to circulating CD4 levels. Note: showing mean plus 95% confidence interval for each group at each timepoint.

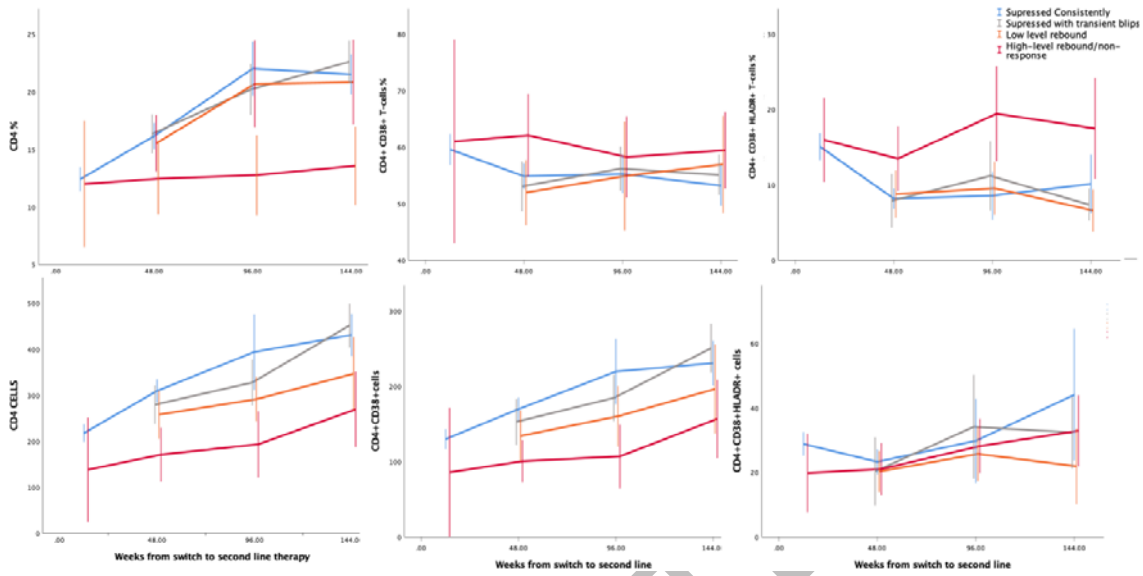


Figure 4: CD4/CD38/HLADR expression and viral load, by VL response at baseline (A), 12 weeks (B), 48 weeks (C), 96 weeks (D) and 144 weeks (E) for viral loads >40 copies/ml. Note: Definition of suppressed consistently at 12 weeks includes VL response

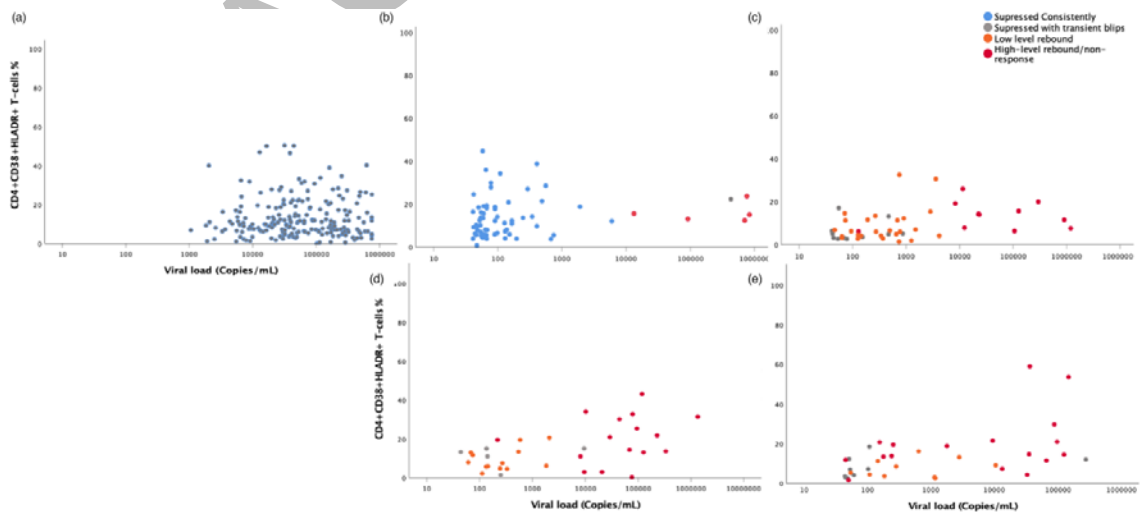
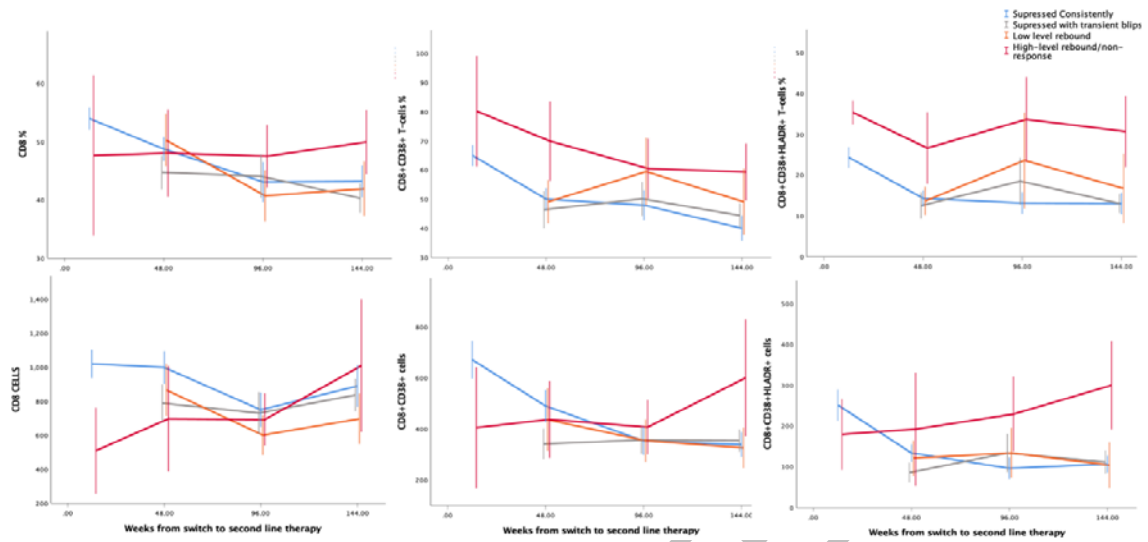


Figure 5: Changes in CD8 and activated CD8 sub-populations over time, expressed both as percentages and normalised to circulating CD8 levels. Note: Showing mean plus 95% confidence interval for each group at each timepoint.



ACCEPTED

Table 1: Cell numbers and subpopulations over 144 weeks on second-line therapy.

	Weeks from switch to second-line therapy			p	Weeks from switch to second-line therapy		p	p	p
	0	12	48	Week 48 v 0	96	144	week 144 v 0	week 144 v 12	week 144 v 48
Numbers with results	208	140	183		106	147		ANCOVA adjusted for BL	
CD4 cells/mm ³	112 (98 to 125) N=193	213 (194 to 232)	286 (267 to 305) N=182	p<0.0005	328 (288 to 367)	408 (378 to 438)	p<0.0005	p<0.0005	p<0.0005
CD4 %	10 (9 to 11) N=93	12 (11 to 13)	16 (15 to 17) N=82	p<0.0005	20 (18 to 21)	21 (20 to 22)	p<0.0005	p<0.0005	p<0.0005
CD4+ CD38+ T-cells %	64 (62 to 66)	60 (57 to 62)	55 (53 to 56)	p<0.0005	56 (54 to 58)	55 (53 to 57)	p<0.0005	p=0.001	p=0.85
CD4+CD38+H LADR+ T-cells %	13 (12 to 15)	15 (13 to 17)	9 (7 to 10)	p<0.0005	11 (9 to 14)	10 (8 to 12)	p=0.003	p<0.0005	p=0.39
CD4+CD38+ cells	69 (60 to 78)	127 (114 to 140)	156 (144 to 168)	p<0.0005	183 (161 to 205)	224 (205 to 243)	p<0.0005	p<0.0005	p<0.0005

	N=1 93		N=1 82		20 5)				
CD4+CD38+H LADR+ cells	13 (11 to 15)	28 (25 to 32)	22 (18 to 25)	p<0.00 05	30 (23 to 38)	36 (27 to 45)	p<0.00 05	p=0.05 2	p<0.00 05
	N=1 93		N=1 82						
	Weeks from switch to second- line therapy			p	Weeks from switch to second- line therapy		p	p	p
CD8 Cells	690 (627 to 753)	999 (917 to 1081)	908 (842 to 974)	p<0.00 05	71 6 (65 to 77)	867 (787 to 947)	p<0.00 05	p<0.00 05	p=0.36 1
	N=1 93		N=1 82						
CD8 %	58 (57 to 60)	54 (52 to 56)	48 (46 to 49)	p<0.00 05	44 (42 to 46)	43 (41 to 45)	p<0.00 05	p<0.00 05	p<0.00 05
	N=1 93		N=1 82			N=1 46			
CD8+CD38+ T-cells %	73 (70 to 76)	66 (62 to 69)	50 (47 to 53)	p<0.00 05	52 (49 to 55)	45 (42 to 48)	p<0.00 05	p<0.00 05	p=0.02
CD8+CD38+H LADR+ T-cells %	21 (19 to 23)	25 (22 to 27)	14 (13 to 16)	p<0.00 05	19 (16 to 23)	16 (14 to 18)	p<0.00 05	p<0.00 05	p=0.66
CD8+CD38+ cells	488 (438 to 537)	661 (590 to 732)	441 (399 to 483)	p=0.09 4	36 2 (33 to 40)	378 (336 to 420)	p<0.00 05	p<0.00 05	p=0.04

	N=1 93		N=1 82		39 5)				
CD8+CD38+H LADR+ cells	143 (122 to 163) N=1 93	249 (212 to 285)	123 (107 to 139) N=1 82	p=0.06 7	13 4 (10 9 to 15 9)	134 (112 to 157)	p=0.10 4	p<0.00 05	p=0.53
CD4:CD8 ratio	0.2 (0.2 to 0.2) N=1 93	0.3 (0.2 to 0.3)	0.4 (0.3 to 0.4) N=1 82	p<0.00 05	0.5 (0. 4 to 0.5)	0.5 (0.5 to 0.6)	p<0.00 05	p<0.00 05	p<0.00 05

Note: showing mean and 95% confidence interval for each randomised group at each timepoint. 208 patients were immunophenotyped at baseline; 193 patients had CD4 and CD8 data on the same sample as immunophenotyping at baseline (15 had CD4 and CD8 on screening sample only)

ACCEPTED