Regulation of mitophagy by the NSL complex underlies genetic risk for Parkinson's disease at 16q11.2 and MAPT H1 loci

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Running Title: New mitophagy Parkinson's risk genes

ABSTRACT

Parkinson's disease is a common incurable neurodegenerative disease. The identification of genetic variants via genome-wide association studies has considerably advanced our understanding of the Parkinson's disease genetic risk. Understanding the functional significance of the risk loci is now a critical step towards translating these genetic advances into an enhanced biological understanding of the disease. Impaired mitophagy is a key causative pathway in familial Parkinson's disease, but its relevance to idiopathic Parkinson's disease is unclear. We used a mitophagy screening assay to evaluate the functional significance of risk genes identified through genome-wide association studies. We identified two new regulators of PINK1-dependent mitophagy initiation, KAT8 and KANSL1, previously shown to modulate lysine acetylation. These findings suggest PINK1-mitophagy is a contributing factor to idiopathic Parkinson's disease. *KANSL1* is located on chromosome 17q21 where the risk associated gene has long been considered to be *MAPT*. While our data does not exclude a possible association between the *MAPT* gene and Parkinson's disease, it provides strong evidence that *KANSL1* plays a crucial role in the disease. Finally, these results enrich our understanding of physiological events regulating mitophagy and establish a novel pathway for drug targeting in neurodegeneration.

Keywords: GWAS; KANSL1; KAT8; mitophagy; Parkinson's disease

Abbreviations: ARA-C = Cytosine β-D-arabinofuranoside; ASE = Allele-Specific Expression; BRD4 = Bromodomain-containing protein 4; Cas = CRISPR associated protein; Coloc = Colocalization; CRISPR = Clustered Regularly Interspaced Short Palindromic Repeats; CRISPRi = CRISPR inteference; dCas9 = enzymatically dead Cas9; DMEM = Dulbecco's Modified Eagle Medium; eQTL = Expression Quantitative Trait Loci; FBS = Foetal Bovine Serum; FDR = False Discovery Rate; GTEx = Genotype-Tissue Expression; GWAS = Genome-Wide Association Study; HBSS = Hanks' Balanced Salt Solution; HCFC1 = Host Cell Factor C1; HCS = High Content Screen; hiPSC = Human Induced Pluripotent Stem Cell; IB = Immunoblot; IF = Immunofluorescence; IMEx = International Molecular Exchange; KANSL1 = KAT8 Regulatory NSL Complex Subunit 1; KANSL2 = KAT8 Regulatory NSL Complex Subunit 2; KANSL3 = KAT8 Regulatory NSL Complex Subunit 3; KAT = Lysine Acetyltransferase; KAT8 = Lysine Acetyltransferase 8; KD = Knockdown; LD = Linkage Disequilibrium; LoF = Loss of Function; MCRS1 = Microspherule Protein 1; Ψ_m = Mitochondrial Membrane Potential; MSL = Male-Specific Lethal; NA = Numerical Aperture; NEAAs = Non-Essential Amino Acids; No TD = No Transduction; NSL = Non-Specific Lethal; O/A = Oligomycin/Antimycin; OGT = O-linked N-

acetylglucosaminyltransferase; ORFs = Open Reading Frames; PBS = Phosphate Buffered Saline; PD = Parkinson's Disease; PFA = Para-Formaldehyde; PHF20 = PHD Finger Protein 20; PINK1 = PTEN-induced kinase 1; POE = Parkin Overexpressing; pParkin(Ser65) = Phosphorylated Parkin (Serine65); PPI = Protein-Protein Interaction; pRab8A(Ser111) = Phosphorylated Rab8a (Serine111); pUb(Ser65) = Phosphorylated Ubiquitin (Serine 65); ROCKi = Rho Kinase Inhibitor; SCR = Scrambled; SD = Standard Deviation; sgRNA = Single Guide RNA; siRNA = Small Interfering RNA; SNPs = Single Nucleotide Polymorphisms; TET-ON = Tetracycline-ON; TMRM = Tetramethylrhodamine, Methyl Ester, Perchlorate; TWAS = Transcriptome-Wide Association Analysis; WDR5 = WD Repeat Domain 5; WPPINA = Weighted Protein-Protein Interaction Network Analysis

INTRODUCTION

Parkinson's disease (PD) is the most common movement disorder of old age and afflicts more than 125,000 in the UK [1]. Temporary symptomatic relief remains the cornerstone of current treatments, with no disease-modifying therapies yet available [2]. Until recently, the genetic basis for PD was limited to family-based linkage studies, favouring the identification of rare Mendelian genes of high penetrance and effect. However, genome-wide association studies (GWAS) have identified large numbers of common genetic variants linked to increased risk of developing the disease [3,4]. While these genetic discoveries have led to a rapid improvement in our understanding of the genetic architecture of PD [5], they have resulted in two major challenges for the research community. First, conclusively identifying the causal gene(s) for a given risk locus, and secondly dissecting their contribution to disease pathogenesis. Addressing these challenges is critical for moving beyond genetic insights to developing new disease-modifying strategies for PD.

Previous functional analyses of *PINK1* and *PRKN*, two genes associated with autosomal recessive PD, have highlighted the selective degradation of damaged mitochondria (mitophagy) as a key contributor to disease pathogenesis. In mammalian cells, the mitochondrial kinase PINK1 selectively accumulates at the surface of damaged mitochondria, where it phosphorylates ubiquitin, leading to the recruitment and phosphorylation of the E3 ubiquitin ligase Parkin. The recruitment of autophagy receptors leads to the engulfment of damaged mitochondria in autophagosomes, and ultimately fusion with lysosomes [6–11]. It has subsequently become clear that other PD-associated Mendelian genes, such as *FBXO7*, *DJ-1* and *VPS35* [12], are implicated in the regulation of PINK1-mediated mitochondrial quality control. Based upon these data, we hypothesised that PD-GWAS candidate genes may also be involved in this process, providing a mechanistic link between these genes and the aetiology of idiopathic PD. In order to test that hypothesis, we used functional genomics to prioritise candidate genes at the PD GWAS loci, and we developed a phenotypic high content screening assay as a tool to identify genes that regulate PINK1-dependent mitophagy initiation, and as such, are likely to be genes that influence the risk of developing PD.

In this study, we show that PD GWAS risk gene candidates KAT8 and KANSL1, that are both part of the non-specific lethal (NSL) complex, are new and important regulators of PINK1-mediated mitochondrial quality control. KAT8 is a histone acetyltransferase belonging to the MYST family that represents the major catalytic constituent of two distinct protein complexes: the male-specific lethal (MSL) and NSL complex [13,14]. Alongside KAT8,

the NSL complex consists of 8 additional proteins HCFC1, KANSL1, KANSL2, KANSL3, MCRS1, OGT, PHF20, WDR5 [13]. KAT8 is the catalytically active acetytransferase of the NSL complex, responsible for the deposition of acetylation modifications on lysine 16 of histone H4 (H4K16ac) facilitating chromatin decompaction making it more permissible for transcriptional machinery and target gene expression[15], in addition to protranscriptional H4K5ac and H4K8ac [16]. The NSL complex regulates the expression of genes involved in a multitude of crucial biological processes including proliferation, metabolism, transcription, DNA replication and autophagy [15–23]. In addition to its canonical role in the nucleus, components of the NSL complex have also been suggested to partially localise to the mitochondria (KAT8, KANSL1, KANSL2, KANSL3 and MCRS1) where they regulate mtDNA transcription and mitochondrial oxidative metabolism [24].

These findings suggest that mitophagy contributes to idiopathic PD and provides a proof of principle for functional screening approaches to identify causative genes in GWAS loci. Finally, these results suggest lysine acetylation as a potential new avenue for mitophagy modulation and therapeutic intervention.

MATERIALS AND METHODS

The following methods are available on protocols.io: Cell-Based in vitro Assays, dx.doi.org/10.17504/protocols.io.5jyl89648v2w/v1; Bioinformatic Prioritisation and Hit Validation, dx.doi.org/10.17504/protocols.io.3byl4br2zvo5/v1; Drosophila stocks, husbandry, locomotor and lifespan assays, and immunohistochemistry and sample preparation, dx.doi.org/10.17504/protocols.io.eq2lyn1dqvx9/v1.

Reagents

Oligomycin (mitochondrial complex V inhibitor) was purchased from Cayman Chemicals (11341) and from Sigma-Aldrich (O4876), and antimycin A (mitochondrial complex III inhibitor) were purchased from Sigma-Aldrich (A8674). All siRNAs were purchased as pre-designed siGENOME SMARTpools from Dharmacon: non-targeting (D-001206-13), PINK1 (M-004030-02), PLK1 (L-003290-00), KIF-11 (L-003317-00), KAT8 (M-014800-00), KANSL1 (M-031748-00), KANSL2 (M-020816-01), KANSL3 (M-016928-01), HCFC1 (M-019953-01), MCRS1 (M-018557-00), OGT (M-019111-00), PHF20 (M-015234-02), WDR5

(M-013383-01). The following antibodies were used for immunocytochemistry: mouse anti TOM20 (Santa Cruz, sc-17764, RRID:AB_628381, 1:1000), rabbit anti phospho-ubiquitin (Ser65) (Cell Signaling, 37642, 1:1000), rabbit anti phospho-Parkin (Ser65) (Abcam/Michael J. Fox Foundation, MJF17, 1:250), rabbit anti FLAG (Sigma-Aldrich, F7425, RRID:AB_439687, 1:500), AlexaFluor 488 goat anti rabbit (Invitrogen, A11008, RRID:AB_143165, 1:2000), AlexaFluor 568 goat anti mouse (Invitrogen, A11004, RRID:AB_2534072, 1:2000), AlexaFluor 488 donkey anti-rabbit (Invitrogen, A21206, RRID:AB_2535792, 1:2000), AlexaFluor 647 donkey anti-mouse (Invitrogen, A31571, RRID:AB_162542, 1:2000). The following antibodies were used for immunoblotting: mouse anti TIM23 (BD Biosciences, 611223, RRID:AB_398755, 1:1000), rabbit anti TOM20 (Santa Cruz, sc-11415, RRID:AB_2207533, 1:1000), rabbit anti phospho-ubiquitin (Ser65) (Merck Millipore, ABS1513-I, RRID:AB_2858191, 1:1000; and Cell Signaling, 37642, 1:1000), mouse anti GAPDH (Abcam, ab110305, RRID:AB_10861081, 1:10000), rabbit anti KAT8 (Abcam, ab200660, RRID:AB_2891127, 1:1000), rabbit anti total-Tau (DAKO, A0024, RRID:AB_10013724, 1:10000), mouse anti V5 tag (Invitrogen, R960-25, RRID:AB_2556564, 1:1000), rabbit anti KANSL1 (Sigma-Aldrich, HPA006874, RRID:AB_1852393, 1:500), rabbit anti pParkin(Ser65) (Abcam/Michael J. Fox Foundation, MJF17, 1:1000), mouse anti FLAG M2 (Sigma-Aldrich, F3165, RRID:AB_259529, 1:1000), sheep anti pRab8A(Ser111) (MRC Protein Phosphorylation and Ubiquitylation Unit, University of Dundee, 1ug/ml preblocked with 10ug/ml non-phosphorylated peptide [25]), rabbit anti total Rab8 (Cell Signaling, 6975, RRID:AB_10827742, 1:1000), IRDye 680LT donkey anti mouse (LI-COR Biosciences, 925-68022, RRID:AB_2814906, 1:20000), IRDye 800CW donkey anti rabbit (LI-COR Biosciences, 925-32213, RRID:AB_2715510, 1:20000), IRDye 800CW Donkey anti-Goat (LI-COR Biosciences, 926-32214, RRID:AB_621846, 1:20000), IRDye® 680RD Goat anti-Rabbit (LI-COR Biosciences, 926-68071, RRID:AB_10956166, 1:20000). The generation of rabbit monoclonal anti PINK1 antibody has been described elsewhere [26], and is available on reasonable request to the corresponding author.

Selection of genes for High Content Screening

Candidates for High Content Screening were selected based on i) Weighted Protein-Protein Interaction Network Analysis (WPPINA); ii) complex prioritization; and, iii) coloc analysis.

WPPINA analysis is reported in [27] where the 2014 PD GWAS [28] was analysed; candidate genes where selected among those prioritised and with a linkage disequilibrium (LD) $r2 \geq 0.8$. The same pipeline has then been additionally applied to the 2017 PD GWAS [3] to update the list of candidate genes. Briefly, a protein-protein interaction network has been created based on the Mendelian genes for PD (seeds) using data from databases within the IMEx consortium. The network has been topologically analysed to extract the core network (i.e. the most interconnected part of the network). The core network contains the proteins/genes that can connect >60% of the initial seeds and are therefore considered relevant for sustaining communal processes and pathways, shared by the seeds. These processes have been evaluated by Gene Ontology Biological Processes enrichment analysis. The top single nucleotide polymorphisms (SNPs) of the 2017 PD GWAS have been used to extract open reading frames (ORFs) in cis-haplotypes defined by LD $r2 \geq 0.8$ (analysis performed in October 2017). These ORFs have been matched to the core network to identify overlapping proteins/genes in relevant/shared pathways. Results of complex prioritization (neurocentric prioritization strategy) were gathered from [3] where this strategy was applied to the 2017 PD GWAS. The coloc analysis was performed as reported in [29], posterior probabilities for the hypothesis that both traits, the regulation of expression of a given gene and the risk for PD share a causal variant (PPH4), were calculated for each gene, and genes with $PPH4 \geq 0.75$ were considered to have strong evidence for colocalization. Summary statistics were obtained from the most recent PD GWAS [4] and were used for regional association plotting using LocusZoom [30]. With the exception of *PM20D1* all genes are expressed in SHSY5Y cells according to publically available expression data deposited in the Human Protein Atlas (proteinatlas.org) [31] and EBI Expression Atlas (http://www.ebi.ac.uk/gxa) [32].

Cell Culture and siRNA transfection

Parkin over-expressing (POE) SHSY5Y cells are a kind gift from H. Ardley [33] and the mt-Keima POE SHSY5Y cells were a kind gift of C. Luft [34]. PINK1-HA overexpressing SHSY5Y cells were a kind gift from E. Deas [35]. WT SHSY5Y (RRID:CVCL_0019) and H4 (RRID:CVCL_1239) cells were sourced from American Type Culture Collection (ATCC, RRID:SCR_00167). Lenti-X 293T human embryonic kidney (HEK) cells were sourced from Takara Bio (632180, RRID: CVCL_4401). Cells were cultured in Dulbecco's Modified Eagle Medium (DMEM, Gibco, 11995-065) supplemented with 10% heat-inactivated foetal bovine

serum (FBS, Gibco) in a humidified chamber at 37 °C with 5% CO_2. For siRNA transfection, cells were transfected using DharmaFECT1 transfection reagent (Dharmacon, T-2001-03) according to the manufacturer's instructions (for concentrations of siRNA, see sections below). Whole genome-sequencing shows SHSY5Ys are of the H1/H1 haplotype (data not shown).

KANSL1 iNeuron Culture and Differentiation

Isogenic human induced pluripotent stem cell (hiPSC) lines with/without a heterozygous loss of function (LoF) frameshift mutation in Exon2 of the KANSL1 gene (c.531insT), which have also being stable transduced with transgenes permitting doxycycline-inducible overexpression of murine Ngn2 were a kind gift from the lab of N. Nadif Kasri and have been published elsewhere [22]. Whole genome-sequencing shows the parental line is of the H1/H2 haplotype, with Sanger sequencing of KANSL1 cDNA revealing the LoF frameshift mutation in the KANSL1$^{+/-}$ line is on the H2 haplotype (i.e. H1/-) (data not shown). hiPSCs were cultured on Geltrex (Thermofisher) coated culture dishes in mTeSR1 (StemCell Technologies) and maintained in a humidified 37 °C incubator, 5% CO2.

Isogenic KANSL1$^{+/+}$ and KANSL1$^{+/-}$ hiPSCs were differentiated into excitatory cortical neurons by doxycycline induced overexpression of murine Ngn2 by adapting recently published protocols [22,36]. On d0 hiPSCs were first dissociated into a single cell suspension using accutase (Sigma) before plating in induction medium consisting of DMEM/F12 supplemented with 1x Glutamax, 1x non-essential amino acids (NEAAs), 1x N2-supplement (all Thermofisher), 4 µg/ml doxycyxline (Sigma). Induction media was additionally supplemented with 10 µM Y-27632 Rho Kinase inhibitor (ROCKi, Peprotech) during initial seeding. 7.5x10^5 cells were seeded onto geltrex coated 6-well plates. 24 h later (d1) and 48 h later (d2) a full medium change was performed with freshly prepared induction media without Y-27632 ROCKi. On d3 a full medium change was performed with freshly prepared N2-B27 media consisting of a 1:1 mixture of DMEM/F12:Neurobasal supplemented with 0.5x N2-supplement, 0.5x B27 supplement, 0.5x NEAAs, 0.5x Glutamax, 45 µM 2-Mercaptoethanol (all Thermofisher), 2.7 µg/ml insulin (Sigma). N2-B27 media was additionally supplemented with 2 µM Cytosine β-D-arabinofuranoside (ARA-C) (Sigma) on d3. A half media change with N2-B27 lacking ARA-C was performed every 3-4 days

thereafter. A half media change with N2-B27 was performed on d16 with cells collected for experimental assays 24 h later on d17.

CRISPRi-i3N iNeuron Culture and Differentiation

A hiPSC line stably transduced with transgenes permitting doxycycline-inducible overexpression of murine Ngn2 at the AAVS1 safe-harbor locus, and stably transduced with constitutively expressed enzymatically dead Cas9 (dCas9)-KRAB transcriptional repressor fusion protein at the CLYBL promoter safe-harbor locus (CRISPRi-i3N hiPSCs) was a kind gift from the labs of M. E. Ward and M. Kampmann, and have been published elsewhere [37,38]. hiPSCs were cultured as outlined above for KANSL1 hiPSCs.

CRISPRi-i3N hiPSCs were first transduced with lentiviral particles encoding a mCherry-reporter and sgRNA sequences targeting the promoter regions of *KANSL1*, *KAT8* or *PINK1*, or a non-targeting sgRNA control (see Supplementary Table 1 for sgRNA sequences, and below for lentiviral production). Transduced cells were then differentiated into excitatory cortical neurons by doxycycline induced overexpression of Ngn2 by adapting recently published protocols [37,38].

On d-1 CRISPRi-i3N hiPSCs were first dissociated into a single cell suspension using accutase and reverse transduced with sgRNA lentiviral supernatant in mTeSR1 supplemented with 5ug/ml polybrene and 10 µM Y-27632 ROCKi. 4.5x10^5 cells were seeded onto Geltrex-coated 6-well plates. 24 h later (d0) media was changed to induction media (composition outlined above for KANSL1 iNeurons). 24 h later (d1) and 48 h later (d2) a full medium change was performed with freshly prepared induction media. On d3, differentiating iNeurons were dissociated into a single cell suspension using accutase (Sigma) and seeded in N2-B27 media (composition outlined above for KANSL1 iNeurons) into Geltrex-coated 96-well CellCarrier Ultra plates for immunofluorescence (IF) (3x10^4 cells per well) and 12-well plates (5x10^5 cells per well) for biochemistry purposes. A half media change with N2-B27 was performed the following day (d4) and every 3-4 days thereafter. A half media change with N2-B27 was performed on d16 with cells collected for experimental assays 24 h later on d17.

Lentiviral Particle Generation

70-90% confluent Lenti-X 293T HEK cells cultured in DMEM 10% FBS media were transfected with pMD2.G and pCMVR8.74 alongside appropriate delivery plasmids: pLV[Exp]-U6>sgRNA-hPGK>mApple (Vectorbuilder) plasmids (for sgRNA), empty pLVX-EF1α-IRES-Puro (Clontech, Takara Bio, 631988), V5-KANSL1 pLVX-EF1α-IRES-Puro, or V5-KAT8 pLV[Exp]-EF1α-IRES-Puro at a 1:1:2 molar mass ratio using Lipofectamine 3000 (Invitrogen). The next day a full media change was performed with mTeSR1 (for sgRNA lentivirus) or DMEM 10% FBS media and cells cultured for 24 h. The lentivirus containing mTeSR1 / DMEM 10% FBS was collected and diluted 1:2 with fresh mTeSR1 or 10% FBS before filtering through 0.44 µm PES filters. pMD2.G (Addgene plasmid #12259, RRID:Addgene_12259) and pCMVR8.74 (Addgene plasmid #22036, RRID:Addgene_22036) were gifts from Didier Trono. KANSL1 cDNA (ENST00000432791.7) with N-terminal V5 tag was cloned into the pLVX-EF1α-IRES-Puro plasmid using SpeI and NotI restriction sites (doi:10.5281/zenodo.6903553). See Supplementary Table 1 for sgRNA plasmids and V5-KAT8 pLV[Exp]-EF1α-IRES-Puro plasmids (Vectorbuilder).

ASEs

Sites of allele-specific expression (ASE) were identified as described by Guelfi and colleagues [39] by mapping RNA-seq data to personalised genomes, an approach specifically chosen because it aims to minimise the impact of mapping biases. RNA-seq data generated from 49 putamen and 35 substantia nigra tissue samples from the UK Brain Expression Consortium was used for this analysis[39] and can be accessed through European Genome-phenome Archive numbers EGAS00001002113 and EGAS00001003065. All samples were obtained from neuropathologically normal individuals of European descent and sites with greater than 15 reads in a sample were tested for ASE. Only sites present in non-overlapping genes were considered and data from both the tissues were considered together to increase power. Sites with minimum false discovery rate (FDR) < 5% across samples were marked as ASE sites. Plots were generated using Gviz3 (https://bioconductor.org/packages/Gviz/), with gene and transcript details obtained from Ensembl v92 (http://www.ensembl.org/, RRID:SCR_002344).

High Content siRNA Screen

Cell plating and siRNA transfection

siRNA was dispensed into Geltrex-coated 96-well CellCarrier Ultra plates (Perkin Elmer) at a final concentration of 30 nM using the Echo 555 acoustic liquid handler (Labcyte). For each well, 25 µl of DMEM containing 4.8 µl/ml of DharmaFECT1 transfection reagent was added and incubated for 30 min before POE SHSY5Y cells were seeded using the CyBio SELMA (Analytik Jena) at 15,000 cells per well, 100 µl per well in DMEM + 10% FBS. Cells were incubated for 72 h before treatment with 10 µM oligomycin/10 µM antimycin for 3 h to induce mitophagy. Positive hits from the screen were validated further, however without determining functional siRNA KD of all gene targets in the screens, none of the negative hits can be formally excluded as regulators of PINK1-dependent mitophagy initiation.

IF and Image Capture and Analysis

Cells were fixed with 4% PFA (Sigma-Aldrich, F8775), then blocked and permeabilised with 10% FBS, 0.25% Triton X-100 in PBS for 1 h, before immunostaining with pUb(Ser65) and TOM20 primary antibodies (in 10% FBS/PBS) for 2 h at room temperature. After 3x PBS washes, AlexaFluor 568 anti-mouse and 488 anti-rabbit secondary antibodies and Hoechst 33342 (Thermo Scientific, 62249) were added (in 10% FBS/PBS, 1:2000 dilution for all) and incubated for 1 h at room temperature. Following a final 3x PBS washes, plates were imaged using the Opera Phenix (Perkin Elmer). 5x fields of view and 4x 1 µm Z-planes were acquired per well, using the 40X water objective, NA1.1. Images were analysed in an automated way using the Columbus 2.8 analysis system (Perkin Elmer, https://www.perkinelmer.com/en-ca/product/image-data-storage-and-analysis-system-columbus) to measure the integrated intensity of pUb(Ser65) within the whole cell. First of all, the image was loaded as a maximum projection, before being segmented to identify the nuclei using the Hoechst 33342 channel (method B). The cytoplasm was then identified using the "Find Cytoplasm" building block (method B) on the sum of the Hoechst and Alexa 568 channels. pUb(Ser65) was identified as spots (method B) on the Alexa 488 channel, before measuring their integrated intensity.

Screen quality control, data processing and candidate selection

Screen plates were quality controlled based on the efficacy of the PINK1 siRNA control and O/A treatment window (minimum 3-fold). Data were checked for edge effects using Dotmatics Vortex visualization software. Raw data was quality controlled using robust Z prime > 0.5. Data were processed using Python (http://www.python.org/, RRID:SCR_008394) for Z score calculation before visualization in Dotmatics Vortex. Candidates were considered a hit where Z score was ≥ 2 or ≤ -2, and where replication of efficacy was seen both within and across plates.

siRNA libraries

The siRNA libraries were purchased from Dharmacon as an ON-TARGETplus SMARTpool Cherry-pick siRNA library, 0.25 nmol in a 384-well plate. siRNAs were resuspended in RNase-free water for a final concentration of 20 µM. SCR, PINK1 and PLK1 or KIF11 controls were added to the 384-well plate at a concentration of 20 µM before dispensing into barcoded assay-ready plates.

Mitochondrial enrichment and Western blotting

POE SHSY5Y and H4 cells were transfected with 100 nM siRNA and incubated for 72 h. KANSL1 iNeurons were cultured as detailed above. Whole cell lysates were used from POE SHSY5Y cells, H4 cells, and KANSL1 iNeurons. For some experiments, POE SHSY5Y lysates were first fractionated into cytoplasmic and mitochondria-enriched preparations to facilitate detection of mitochondrial localised proteins of interest. Samples were run on SDS-PAGE before immunoblot (IB) with the Odyssey® CLx Imager (LI-COR Biosciences). Mitochondrial enrichment and Western blotting protocols were described previously [26].

siRNA KD Rescue

POE SHSY5Ys were transfected with 25 nM siRNA (d0) and incubated for 48 h. siRNA KD cells were then transduced with lentivirus in the presence of 10 µg/ml polybrene (d2), full media change performed the following day (d3) and collected 4 days post siRNA transfection (and 2 days post lentivirus transduction) (d4).

pRab8A(Ser111) Measurements

SHSY5Y cells stably overexpressing PINK1-HA were transfected with 100 nM siRNA and incubated for 72 h. 200 ug of protein (whole cell lysate) were immunoprecipitated with Protein A Dynabeads™ (Invitrogen) prebound with 0.5 ug of rabbit anti total-Rab8 antibody (Cell Signaling, 6975) at 4 °C overnight. Samples were eluted from the beads by heating at 95 °C in 2x LDS supplemented with 50mM DTT for 5min.

Immunofluorescence

SHSY5Y cells were reverse transfected with 50 nM siRNA in 96-well CellCarrier Ultra plates according to the manufacturer's instructions and incubated for 72 h. CRISPRi-i3N iNeurons were cultured as described above. Cells were then treated, fixed and stained as per the screening protocol detailed above (for treatment concentrations and times, see corresponding figures). For visualisation purposes, brightness and contrast settings were selected on the SCR (siRNA KD SHSY5Y) or no transduction (No TD) (CRISPRi-i3N iNeurons) controls and applied to all other images. Images are presented as maximum projections of the channels for one field of view.

RT-qPCR

Total RNA was extracted from cells using the Monarch Total RNA Miniprep Kit (New England Bioscience) with inclusion of the optional on-column DNAse treatment and quantified using a NanoDrop One Spectrophotometer (Thermofisher Scientific). RNA was then reverse transcribed in a 10 µl reaction with 2.5 U/µl SuperScript IV reverse transcriptase with 2.5 µM random hexamers, 0.5 mM dNTPs, 5 mM DTT and 2 U/µl RNAseOUT (all Invitrogen). Equal amounts of RNA were reverse transcribed for all samples of a single experiment, with 500ng of RNA in a 10 µl reverse transcription reaction being the most common. The cDNA product was then diluted such that 500ng of reverse transcribed RNA would be in a 600 ul final volume (i.e. 0.83 ng/µl). 4 ul (i.e. 3.33 ng) of the diluted cDNA was then subjected to quantitative real-time PCR (qPCR) using 1x Fast SYBR™ Green Master Mix (Applied Biosystems) and 500 nM gene specific primer pairs (Supplementary Table 2) on a QuantStudio™ 7 Flex Real-Time PCR System (Applied Biosystems). At least

2x technical replicates were performed for each sample and gene target combination, and a RT^- control for all samples and gene target combinations was performed alongside in most assays, with rare exceptions due to test well number limitations. Relative mRNA expression levels were calculated using the $2^{-\Delta\Delta Ct}$ method and *RPL18A* (SHSY5Y and H4 cells) or *UBC* (iNeurons) as the house-keeping gene.

Mitophagy measurement using the mt-Keima reporter

Stable mt-Keima expressing POE SHSY5Y cells were reverse transfected with 50 nM siRNA in 96-well CellCarrier Ultra plates according to the manufacturer's instructions and incubated for 72 h. For the assay, the cell medium was replaced with phenol-free DMEM + 10% FBS containing Hoechst 33342 (1:10000) and either DMSO or 1 µM oligomycin/1 µM antimycin to induce mitophagy. Cells were immediately imaged on the Opera Phenix (PerkinElmer) at 37 °C with 5% CO_2, acquiring 15x single plane fields of view, using the 63X water objective, NA1.15. The following excitation wavelengths and emission filters were used: cytoplasmic Keima: 488 nm, 650–760 nm; lysosomal Keima: 561 nm, 570–630 nm; Hoechst 33342: 375 nm, 435–480 nm. Images were analysed in an automated way using the Columbus 2.8 analysis system (Perkin Elmer) to measure the mitophagy index. Cells were identified using the nuclear signal of the Hoechst 33342 channel, before segmenting and measuring the area of the cytoplasmic and lysosomal mt-Keima. The mitophagy index was calculated as the ratio between the total area of lysosomal mitochondria and the total area of mt-Keima (sum of the cytoplasmic and lysosomal mtKeima areas) per well.

***Drosophila* stocks and husbandry**

Flies were raised under standard conditions in a humidified, temperature-controlled incubator with a 12h:12h light:dark cycle at 25°C, on food consisting of agar, cornmeal, molasses, propionic acid and yeast. The following strains were obtained from the Bloomington *Drosophila* Stock Center (RRID:SCR_006457): *mof* RNAi lines, P{TRiP.JF01701} (RRID:BDSC_31401); and P{TRiP.HMS00537} (RRID:BDSC_58281); *nsl1* RNAi lines, P{TRiP.HMJ22458} (RRID:BDSC_58328); the pan-neuronal *nSyb-GAL4* driver (RRID:BDSC_51941); and dopaminergic neuron driver (TH-GAL4; RRID:BDSC_8848); and control (*lacZ*) RNAi P{GD936}v51446) (RRID:FlyBase_FBst0469426) from the Vienna

Drosophila Resource Center (RRID:SCR_013805). All experiments were conducted using male flies.

Locomotor and lifespan assays

The startle induced negative geotaxis (climbing) assay was performed using a counter-current apparatus. Briefly, 20-23 males were placed into the first chamber, tapped to the bottom, and given 10 s to climb a 10 cm distance. This procedure was repeated five times (five chambers), and the number of flies that has remained into each chamber counted. The weighted performance of several group of flies for each genotype was normalized to the maximum possible score and expressed as *Climbing index* [40].

For lifespan experiments, flies were grown under identical conditions at low-density. Progeny were collected under very light anaesthesia and kept in tubes of approximately 20 males each, around 50-100 in total. Flies were transferred every 2-3 days to fresh medium and the number of dead flies recorded. Percent survival was calculated at the end of the experiment after correcting for any accidental loss.

Immunohistochemistry and sample preparation

Drosophila brains were dissected from aged flies and immunostained as described previously [41]. Adult brains were dissected in PBS and fixed in 4% formaldehyde for 30 min on ice, permeabilized in 0.3% Triton X-100 for 3 times 20 min, and blocked with 0.3% Triton X-100 plus 4% goat serum in PBS for 4 h at RT. Tissues were incubated with anti-tyrosine hydroxylase (Immunostar Inc. #22491, RRID:AB_572268), diluted in 0.3% Triton X-100 plus 4% goat serum in PBS for 72 h at 4°C, then rinsed 3 times 20 min with 0.3% Triton X-100 in PBS, and incubated with the appropriate fluorescent secondary antibodies overnight at 4°C. The tissues were washed 2 times in PBS and mounted on slides using Prolong Diamond Antifade mounting medium (Thermo Fisher Scientific). Brains were imaged with a Zeiss LMS 880 confocal. Tyrosine hydroxylase-positive neurons were counted under blinded conditions.

Statistical Analysis

Intensity measurements from imaging experiments were normalised for each experiment (see figure legends and graphs). N numbers are shown in figure legends and refer to the number of

independent, replicate experiments. Within each experiment, the mean values of every condition were calculated from a minimum of 3 technical replicates. Integrated density measurements from Western blot experiments were normalised to control wells (see figure legends and graphs). Wherever possible, normalisation to conditions for statistical comparisons were avoided in order to maintain experimental error associated. GraphPad Prism 9 (La Jolla, California, USA) was used for statistical analyses and graph production. Data were subjected to either one-way or two-way ANOVA with Dunnett's post-hoc analysis for multiple comparisons, unless otherwise stated. All error bars indicate mean ± standard deviation (SD) from replicate experiments.

Data Availability

Exome and genome variant data is available from Genome Aggregation Database (gnomAD): https://gnomad.broadinstitute.org/about Version 2.1.1 and subsequent releases are available for download: https://gnomad.broadinstitute.org/downloads, Google Cloud Public Datasets (https://cloud.google.com/public-datasets), the Registry of Open Data on AWS (https://registry.opendata.aws/), and Azure Open Datasets (https://azure.microsoft.com/en-us/services/open-datasets/).

Data sets from the IMEx consortium [42] (https://www.imexconsortium.org/) used for WPPINA analysis data are reported by Ferrari et al. [27]: and available through doi:10.1186/s12864-018-4804-9. 2014 PD GWAS data [28] analysed is available through doi:10.1038/ng.3043. 2017 PD GWAS [3] data is available through doi:10.1038/ng.3955. PD GWAS summary statistics [4] are available through doi:10.1016/S1474-4422(19)30320-5.

Genes expressed in SHSY5Y cells were accessed from expression data deposited in the Human Protein Atlas (proteinatlas.org) and EBI Expression Atlas (https://www.ebi.ac.uk/gxa/home).

Tabulated data for figures of the manuscript are available through doi:10.5281/zenodo.6952972. The datasets generated, analysed, and reported in this manuscript are available from the corresponding author on reasonable request.

Software

LocusZoom 30 (http://locuszoom.org/, RRID:SCR_021374) was used for the regional association plotting of summary statistics from PD GWAS [4]. Dotmatics Vortex v5.1 software was used to check for edge effects and for visualisation (https://www.dotmatics.com/capabilities/vortex). GraphPad Prism 9 (La Jolla, California, USA) was used for statistical analyses and graph production: GraphPad Prism (RRID:SCR_002798) (https://www.graphpad.com/).

RESULTS

Bioinformatic prioritisation of PD GWAS candidates

Genomic analyses of PD have identified over 80 loci associated with an increased lifetime risk for disease [3]. In contrast to Mendelian PD genes, however, the assignment of a causative gene to a risk locus is often challenging. In order to identify new risk genes for PD, we undertook a triage of PD GWAS candidate genes using a combination of methods: i) Colocalization (Coloc) and transcriptome-wide association analysis (TWAS) [43] using expression quantitative trait loci (eQTLs) information derived from Braineac [44], GTEx and CommonMind resources [29,45] to link PD risk variants with specific genes, ii) weighted protein-protein interaction (PPI) network analysis (WPPINA)[27] based on Mendelian genes associated with PD, and iii) the prioritised gene set as described in PD-GWAS [3,28]. This resulted in the nomination of 31 open reading frames (ORFs) as putatively causal for associations at PD risk loci. 55% of these genes were prioritised through multiple techniques, with three out of 31 genes (*KAT8, CTSB* and *NCKIPSD*) identified through all three prioritization methods (Extended Data Fig. 1A). The 31 genes, together with 7 PD Mendelian genes and lysosomal storage disorder genes, previously shown to be enriched for rare, likely damaging variants in PD [46], were then taken forward for functional analysis (Table 1).

pUb(Ser65) based screen of prioritised genes identifies KAT8 as novel regulator of PINK1-dependent mitophagy intitiation.

Based upon extensive data implicating impaired mitophagy in the aetiology of familial PD, we hypothesized that PD-GWAS candidate genes, involved in the most common, idiopathic

form of the disease, may play a role in this process. In order to test whether the 38 prioritised genes have a role in PINK1-mitophagy, we developed and optimized a high content screening (HCS) assay for phosphorylation of ubiquitin at serine 65 (pUb(Ser65)), a PINK1-dependent mitophagy initiation marker [47], following mitochondrial depolarization (Fig. 1A). The 38 prioritised genes were individually knocked down (KD) using siRNA in Parkin over-expressing (POE)-SHSY5Y human neuroblastoma cells. Increased mitochondrial clearance following mitochondrial depolarization induced by treatment with 10 μM of oligomycin/antimycin A (O/A) was validated as an endpoint for mitophagy (Extended Data Fig. 1B). Over 97% of the pUb(Ser65) signal colocalised with the TOM20 mitochondrial marker in O/A treated cells (Extended Data Fig. 1C, D). KD efficiency was validated using both a pool of *PINK1* siRNA, which decreased O/A induced pUb(Ser65) and subsequent TOM20 degradation (Extended Data Fig. 1E-G) without decreasing cell viability (Extended Data Fig. 2A-B), and a pool of Polo-like kinase 1 (PLK-1) siRNA that decreased cell viability by apoptosis (Extended Data Fig. 2A-B). The siRNA pools for the 38 candidate genes, together with controls, were screened in duplicate on each plate, across three replicate plates per run. Hits were identified as those wells where O/A-induced pUb(Ser65) was decreased or increased at greater than two standard deviations from the mean of the scramble (SCR) negative control siRNA.

KAT8 was selected based on reproducible downregulation of O/A-induced PINK1-dependent pUb(Ser65) across all three replicates (Fig. 1B and Extended Data Fig. 1H), without affecting cell viability (Extended Data Fig. 2C). Notably, *KAT8* was selected as a candidate gene on the basis of all three prioritization criteria – namely, proximity of the lead SNP to an ORF (Fig 1D), colocalization of a brain-derived eQTL signal with a PD GWAS association signal (Extended Data Fig. 3) and evidence of PPI with a known PD gene (Table 1). Furthermore, we find that colocalization and TWAS [48] analyses at this locus are consistent with the KD models in the HCS assay (Supplementary Tables 3 and 4) [29]. Both methods predict that the risk allele operates by reducing *KAT8* expression in PD cases versus controls.

The effect of KAT8 KD on pUb(Ser65) was further validated in POE SHSY5Y cells treated with 1 μM O/A, using both immunoblotting (IB) and immunofluorescence (IF) (Fig. 1D-G and Extended Data Fig. 4). In order to assess whether other lysine acetyltransferases (KATs) could regulate PINK1-dependent mitophagy, the pUb(Ser65) screen was repeated in POE SHSY5Y cells silenced for 22 other KATs (Supplementary Table 5) [49,50]. Only KAT8

KD led to a decreased pUb(Ser65) signal, emphasising the specificity of the KAT8 KD effect on pUb(Ser65) (Fig. 1H).

KANSL1, another NSL member and PD GWAS candidate, regulates PINK1-dependent mitophagy and dopaminergic neuron viability

These functional data complement and support the omic prioritization of *KAT8* as a causative gene candidate for the chromosome 16q11.2 PD associated locus (Fig. 1C). To gain further insight into a possible role for KAT8 in the aetiology of PD, we explored the known functional interactions of this protein.

KAT8 has previously been shown to partially localise to mitochondria as part of the NSL complex together with KANSL1, KANSL2, KANSL3, and MCRS1 [24]. To test whether other components of the NSL complex also modulate mitophagy, the pUb(Ser65) screen was repeated in POE SHSY5Y cells silenced for each of the nine NSL components. Notably, reduction of KANSL1, KANSL2, KANSL3, MCRS1 and KAT8 expression led to decreased pUb(Ser65) after 1.5 or 3 h O/A treatment (Fig. 2A and Extended Data Fig. 5), suggesting that the NSL complex modulates PINK1-dependent mitophagy initiation. Interestingly, *KANSL1* is another PD GWAS candidate gene [3]. The effect of KANSL1 KD on pUb(Ser65) was further validated in POE SHSY5Y cells treated with 1 µM O/A, using both IF and IB (Fig. 2B-F).

Deconvolution of the individual siRNA within the respective pools (of 4x individual siRNA against the target gene) (Extended Data Fig. 6) and rescue of the siRNA KD phenotype with overexpression of V5-tagged KANSL1/KAT8 (Extended Data Fig. 7) are both supportive of on-target effect associated with KANSL1/KAT8 KD. The effect of the KAT8 and KANSL1 KD on pUb(Ser65) was confirmed in WT SHSY5Y cells and the astroglioma H4 cell line, both of which are expressing endogenous levels of Parkin (Extended Data Fig. 8). In order to further assess the effect of KAT8 and KANSL1 KD on PINK1-dependent mitophagy initiation, we measured pUb(Ser65) levels over time (Fig. 3A-B), as well as PINK1 recruitment (Fig. 1D and F, Fig. 2D and F), Parkin recruitment (Fig. 3C-D), PINK1-dependent phosphorylation of Parkin at Ser65 (pParkin(Ser65)) (Fig. 3E-F and Extended Data Fig. 9) [9], and PINK1-dependent (but indirect) phosphorylation of Rab8A at Ser111 (pRab8A(Ser111)) [25] (Extended Data Fig. 10). The reduction in pUb(Ser65) levels

is not associated with reduced availability of Parkin (Extended Data Fig. 9) or ubiquitin (Extended Data Fig. 11).

Given the canonical function of the NSL complex as a pro-transcriptional epigenetic remodelling complex [15,16], a strong candidate mechanism accounting for the reduced PINK1 protein accumulation and PINK1-dependant mitophagy initiation could be reduced *PINK1* gene expression. In fact, RT-qPCR assessments of *PINK1* mRNA levels in WT and POE SHSY5Ys, reveal that KANSL1 and to a lesser degree KAT8 KD, both reduce *PINK1* gene expression (Fig. 4). These data are mirrored by both *PINK1* gene expression and PINK1 protein accumulation in H4 cells (Extended Data Fig. 8C,D,F-H). Finally, the reduction in *PINK1* mRNA following KANSL1/KAT8 KD in POE SHSY5Ys is also rescued with KANSL1/KAT8 overexpression (Extended Data Fig. 7C,D,G,H).

KD of both KAT8 and KANSL1 reduced subsequent mitochondrial clearance in live POE-SHSY5Y cells, as measured by the mitophagy reporter mt-Keima [51] (Fig. 5).

In order to assess the role of KAT8/KANSL1 in neuronal function and survival *in vivo*, we used *Drosophila* as a model system. Notably, the NSL complex was originally discovered in *Drosophila* through the homologs of *KAT8* and *KANSL1* (*mof* and *nsl1*, respectively), but null mutations for these genes are associated with developmental lethality owing to profound transcriptional remodelling during development [52]. Therefore, we utilised inducible transgenic RNAi strains to target the KD of *mof* and *nsl1* specifically in neuronal tissues. Using behavioural assays as a sensitive readout of neuronal function we found that pan-neuronal KD of *mof* or *nsl1* caused progressive loss of motor (climbing) ability (Extended Data Fig. 12A, B), and also significantly shortened lifespan (Extended Data Fig. 12C, D). Interestingly, loss of *nsl1* had a notably stronger effect than loss of *mof*. Consistent with this, KD of *nsl1* but not *mof*, in either all neurons or only in dopaminergic (DA) neurons, caused the loss of DA neurons (Extended Data Fig. 12E, F).

KANSL1 is a likely PD GWAS candidate at the 17q21 locus

KANSL1 is located within the extensively studied inversion polymorphism on chromosome 17q21 (Extended Data Fig. 13A, B), which also contains *MAPT* - a gene frequently postulated to drive PD risk at this locus [53]. While the majority of individuals inherit this region in the direct orientation, up to 25% of individuals of European descent have a ~1mb sequence in the opposite orientation [54,55], inducing a larger ~1.3–1.6 Mb region of linkage

disequilibrium (LD). Since this inversion polymorphism precludes recombination over a region of ~1.3–1.6 Mb, haplotype-specific polymorphisms have arisen resulting in the generation of two major haplotype clades, termed H1 (common haplotype) and H2 (inversion carriers), with H1 previously strongly linked to neurodegenerative disease including PD [56–58]. Due to high LD, the genetics of this region have been hard to dissect, and robust eQTL analyses have been challenging due to the issue of polymorphisms within probe sequences in microarray-based analyses or mapping biases in RNA-seq-based analyses. Several variants (rs34579536, rs35833914 and rs34043286) are in high LD with the H1/H2 haplotype and are located within *KANSL1* (Fig. 6A,B), raising the possibility that they could directly impact on KANSL1 protein function. In particular, one of the variants is associated with a serine (H1 haplotype) or proline (H2 haplotype) amino acid change in KANSL1 protein sequence (NM_001193465:c.T2152C:p.S718P), and would therefore be predicted to alter the gross secondary structure of the KANSL1 protein (Fig. 6B). Furthermore, we explored the possibility that PD risk might be mediated at this locus through an effect on *KANSL1* expression. Using RNA sequencing data generated from 84 brain samples (substantia nigra n=35; putamen n=49), for which we had access to whole exome sequencing and SNP genotyping data thus enabling mapping to personalised genomes [39], we performed allele-specific expression analysis. More specifically, we quantified the variation in expression between the H1 and H2 haplotypes (Supplementary Table 8) amongst heterozygotes. While we identified allele-specific expression sites within *MAPT* (Extended Data Fig. 14 and Supplementary Table 9), we also identified 4 sites of allele-specific expression in *KANSL1* (Fig. 6A), suggesting that the high PD risk H1 allele is associated with lower *KANSL1* expression, consistent with our functional assessment. Interestingly, sequence analysis of the human *KANSL1* haplotype revealed that the high risk H1 haplotype is the more recent "mutant" specific to *Homo sapiens*, and that other primates and mammals share the rarer non-risk ancestral H2 haplotype (Fig. 6B). To assess the specificity of the KANSL1 KD effect on PINK1-dependent mitophagy initiation, 32 open reading frames in linkage disequilibrium on the H1 haplotype at the 17q21 locus (Extended data Fig. 13A, B and Supplementary Table 10) were knocked down individually and their effect on pUb(Ser65) was assessed. While the effect of KANSL1 KD on pUb(Ser65) was confirmed, neither the KD of MAPT (see also Extended Data Fig. 15), nor the KD of each of the other 30 genes on this locus, led to a decrease in the pUb(Ser65) signal (Fig. 6C). These data confirm the selectivity of our mitophagy screening assay and suggest that *KANSL1* is likely to be a key PD risk gene at the 17q21 locus.

Impairments in PINK1-dependent mitophagy initiaition are also observed in human iNeuron models of KANSL1 and KAT8 deficiency

Finally, we sought to validate the mitophagy initiation impairments associated with reductions in KANSL1 and KAT8, in more disease-relevant human induced pluripotent stem cell (hiPSC) derived neuron systems.

To this end, further experiments were performed using isogenic hiPSC lines with/without a heterozygous loss of function (LoF) frameshift mutation (c.531insT) in KANSL1 introduced through CRISPR/Cas technology [22]. These lines have also been stably transduced with transgenes conferring overexpression of murine neurogenin-2 (Ngn2) under a tetracycline-ON (TET-ON) system, permitting differentiation into human cortical neurons following doxycycline treatment (iNeurons). After 17 days in vitro (DIV), KANSL1 control ($KANSL1^{+/+}$) and heterozygous LoF ($KANSL1^{+/-}$) iNeurons with ~50% reduction in *KANSL1* gene expression (Extended Data Fig. 16A) were subjected to assessments of mitophagy initiation. Treatment of $KANSL1^{+/+}$ iNeurons with 1 µM O/A resulted in pUb(Ser65) deposition, which continued to increase across a prolonged 12 h treatment window (Fig. 7A, B). Whilst $KANSL1^{+/-}$ iNeurons showed detectable pUb(Ser65) deposition, the levels were lower than that of the isogenic $KANSL1^{+/+}$ iNeurons, with this difference being significant at 9 h and 12 h of O/A treatment (Fig. 7B).

Similar experiments were also performed using the CRISPRi-i3N iNeuron system which in addition to the presence of a TET-ON system for dox-induced Ngn2 mediated neuronal differentiation [37], also express an enzymatically dead Cas9 (dCas9)-KRAB transcriptional repressor fusion protein that permits target gene KD through introduction of sgRNA molecules for specific gene promoters [38]. Similar to observations in the $KANSL1^{+/-}$ iNeurons, and KANSL1/KAT8/PINK1 siRNA KD cell line models, CRISPRi KD of all three gene targets (Extended Data Fig. 16B-D) significantly reduced mitochondrial pUb(Ser65) following 9 h O/A treatment as assessed through IF (Fig. 7C, D).

These data support an important functional link between KANSL1, KAT8 and PINK1-dependent mitophagy initiation. While our data support reductions in *PINK1* gene expression as the most likely underlying mechanism for mitophagy impairment in KAT8 and KANSL1 deficient cell lines, further experiments will be required to determine whether KAT8/KANSL1- dependent *PINK1* gene expression regulation also explains mitophagy

impairments in other cellular and *in vivo* models, or/and whether other mechanisms could be involved.

DISCUSSION

Since the first PD GWAS study was performed in 2006 [59], GWAS have identified over 80 independent loci for PD [4]. However, translating GWAS findings into a new molecular understanding of PD-associated pathways and new therapeutic targets has remained a major challenge for the scientific community. In order to screen for PD GWAS candidate genes that play a role in PINK1-mitophagy, and thus are likely to be genuine risk genes for PD, we have set up and optimised a HCS for pUb(Ser65), a marker of PINK1-dependent mitophagy initiation, a key pathway in PD pathogenesis. This approach allowed the successful identification of two new genes associated with increased PD risk, that play a role in mitophagy. Interestingly, these two genes were previously shown to be part of the same complex, the NSL complex.

This study demonstrates the substantial potential of functional screens to exploit genetic data by providing orthogonal information that can confidently identify new risk genes. This is particularly important in genomic regions with uniformly high linkage disequilibrium, such as the 17q21 inversion region which includes 32 ORFs of which many are highly expressed in brain and where existing fine-mapping and functional genomic analyses have been inconclusive. Interestingly, while *MAPT* has long been considered the risk associated gene at this locus, this has recently been questioned by Dong and colleagues, who also raised the significance of KANSL1 in driving PD risk at the locus [60]. In line with our data, eQTL analysis by other groups has shown the PD risk H1 haplotype is associated with reduced *KANSL1* mRNA expression [61,62], and non-synonymous H1 vs H2 haplotype KANSL1 amino acid changes including K104T, N225D, S718P and I1084 we describe in this manuscript have been confirmed to be in high linkage disequilibrium through Sanger sequencing by another group [63]. Similar to our data, eQTL analysis by others has revealed that lower levels of *KAT8* gene expression are also linked to an increased PD risk [3].

Furthermore, functional screening can simultaneously provide mechanistic insights as exemplified in this case by the novel insights we provide into the molecular events regulating mitochondrial quality control and which support a role for mitophagy as a contributing factor to sporadic PD. KANSL1 is part of the NSL complex and functions as a scaffolding protein by binding other subunits, including KAT8 [14]. Through the deposition of pro-transcriptional histone acetylation marks, the NSL complex underscores an important regulator of target gene expression

[16]. In this study we show that PINK1-dependent phosphorylation of ubiquitin is reduced in the context of KANSL1 and KAT8 LoF and that this appears to be largely caused by a reduction in *PINK1* gene expression, at least in the cell line models. We hypothesise that as a consequence of reduced *PINK1* gene and PINK1 protein expression, mitochondrial accumulation of activated PINK1 is reduced, leading to reduced pUb(Ser65) deposition, Parkin activation, and subsequent mitochondrial clearance.

It was previously shown that depletion of KAT8/KANSL1 causes significant downregulation of mitochondrial DNA transcription and translation, and ultimately impaired mitochondrial respiration [24]. Future studies will need to determine whether KAT8/KANSL1-dependent modulation of mitochondrial DNA or nuclear DNA encoded mitochondrial genes could regulate PINK1 mitochondrial accumulation, activation and subsequent mitophagy. It has been further proposed that the KAT8/KANSL1 complex has targets in the mitochondria other than the mitochondrial DNA [24]. It will be interesting to determine whether the KAT8/KANSL1 complex could acetylate ubiquitin, which has previously been shown to be acetylated on six out of its seven lysines (K6, K11, K27, K33, K48, K63) [64]. It will also be interesting to understand further whether the regulation of mitophagy related genes such as *PINK1*, is associated with a direct effect of the NSL complex at target gene promoters, or as a consequence of other upstream biochemical cascades, as previously described in the context of KANSL1 LoF [22].

KANSL1 haploinsufficiency caused by heterozygous pathogenic genetic variants in KANSL1 is associated with the neurodevelopmental disorder Koolen-de Vries syndrome (KdVS; OMIM #610443) [65]. Pathogenic variants in KAT8 have also been associated with developmental disorders inclusive of a strong neurological phenotype [66], underpinning the functional importance of the NSL complex in neurodevelopment. Using iNeurons differentiated from KdVS-derived patient hiPSCs and genome-edited KANSL1 heterozygous hiPSCs, we have recently shown that KANSL1 deficiency leads to impairments in autophagic flux and lysosomal function which appear to be largely caused by elevated cellular reactive oxygen species [22], and impairments in the transcriptional regulation of autophagy related genes have also been described elsewhere in the context of both KAT8 and KANSL1 LoF [20,23]. Together with our current work highlighting *KANSL1* and *KAT8* as PD risk genes, these studies suggest that impairments in the autophagic process could be a contributing pathomechanism for idiopathic disease. Alongside our own data implicating impairments in PINK1-dependent mitophagy initiation, synergistic dysregulation in autophagosome and lysosomal-dependent steps, downstream in the mitophagy process, highlight clearance of damaged mitochondria as a particular vulnerability in the context of KAT8/KANSL1

LoF. While more severe KAT8/KANSL1 haploinsufficiency leads to impaired neurodevelopment, more subtle changes in KAT8/KANSL1 and associated mitochondrial deficits, associated with impaired bulk autophagy might lead to accumulation of cellular damage leading to selective vulnerability of dopaminergic neurons later in life.

Our data highlight the utility of a cellular function high-content siRNA KD screen for prioritisation of GWAS candidates, however it is important to be aware of limitations to the use of such a strategy. While we have confirmed successful KD of KAT8 and KANSL1, without evaluating KD of all genes screened (which can be challenging with high-throughput screens), the potential pitfall for false-negatives remains. siRNA KD strategies are limited to delineating the functional effect of reductions in the expression of a target gene, whereas increased expression might be more disease and functionally relevant in some cases.

Important genetic discoveries in PD, in particular, the identification of the *PINK1* [67] and *PRKN* genes [68], opened the field of selective mitophagy [7]. However, there is still a clear need for a better molecular understanding of mitochondrial quality control. Here we provide new insights into the mechanism by identifying two new molecular players, KAT8 and KANSL1. These new regulators of mitophagy provide the first direct evidence for a role of the PINK1-mitophagy pathway in idiopathic PD and the convergence between familial and idiopathic pathways in disease. Taken together, these findings open a novel avenue for the therapeutic modulation of mitophagy in PD, with potential implications across drug discovery in frontotemporal dementia and Alzheimer's disease, where mitophagy also plays an important role in disease pathogenesis [69].

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COMPETING INTERESTS

The authors declare that they have no conflict of interest

SUPPLEMENTARY MATERIAL

Supplementary material is available at *Brain* online.

REFERENCES

1. Hardy J, Lewis P, Revesz T, Lees A, Paisan-Ruiz C. The genetics of Parkinson's syndromes: a critical review. *Curr Opin Genet Dev*. 2009;19(3):254-265. doi:https://doi.org/10.1016/j.gde.2009.03.008

2. Connolly BS, Lang AE. Pharmacological Treatment of Parkinson Disease: A Review. *JAMA*. 2014;311(16):1670-1683. doi:10.1001/jama.2014.3654

3. Chang D, Nalls MA, Hallgrímsdóttir IB, et al. A meta-analysis of genome-wide association studies identifies 17 new Parkinson's disease risk loci. *Nat Genet*. 2017;49(10):1511-1516. doi:10.1038/ng.3955

4. Nalls MA, Blauwendraat C, Vallerga CL, et al. Identification of novel risk loci, causal insights, and heritable risk for Parkinson's disease: a meta-analysis of genome-wide association studies. *Lancet Neurol*. 2019;18(12):1091-1102. doi:10.1016/S1474-4422(19)30320-5

5. Nalls MA, Plagnol V, Hernandez DG, et al. Imputation of sequence variants for identification of genetic risks for Parkinson's disease: a meta-analysis of genome-wide association studies. *Lancet*. 2011;377(9766):641-649. doi:10.1016/S0140-6736(10)62345-8

6. Narendra D, Tanaka A, Suen D-F, Youle RJ. Parkin is recruited selectively to impaired mitochondria and promotes their autophagy. *J Cell Biol*. 2008;183(5):795-803. doi:10.1083/jcb.200809125

7. McWilliams TG, Muqit MM. PINK1 and Parkin: emerging themes in mitochondrial homeostasis. *Curr Opin Cell Biol*. 2017;45:83-91. doi:10.1016/j.ceb.2017.03.013

8. Narendra DP, Jin SM, Tanaka A, et al. PINK1 Is Selectively Stabilized on Impaired Mitochondria to Activate Parkin. Green DR, ed. *PLoS Biol*. 2010;8(1):e1000298. doi:10.1371/journal.pbio.1000298

9. Kazlauskaite A, Kondapalli C, Gourlay R, et al. Parkin is activated by PINK1-dependent phosphorylation of ubiquitin at Ser65. *Biochem J*. 2014;460(1):127-141. doi:10.1042/BJ20140334

10. Shiba-Fukushima K, Arano T, Matsumoto G, et al. Phosphorylation of Mitochondrial Polyubiquitin by PINK1 Promotes Parkin Mitochondrial Tethering. *PLoS Genet*. 2014;10(12). doi:10.1371/journal.pgen.1004861

11. Lazarou M, Sliter DA, Kane LA, et al. The ubiquitin kinase PINK1 recruits autophagy receptors to induce mitophagy. *Nature*. 2015;524(7565):309-314. doi:10.1038/nature14893

12. Plotegher N, Duchen MR. Crosstalk between lysosomes and mitochondria in Parkinson's disease. *Front Cell Dev Biol*. 2017;5(DEC):2011-2018. doi:10.3389/fcell.2017.00110

13. Cai Y, Jin J, Swanson SK, et al. Subunit Composition and Substrate Specificity of a MOF-containing Histone Acetyltransferase Distinct from the Male-specific Lethal (MSL) Complex. *J Biol Chem*. 2010;285(7):4268-4272. doi:10.1074/JBC.C109.087981

14. Dias J, Van Nguyen N, Georgiev P, et al. Structural analysis of the KANSL1/WDR5/KANSL2 complex reveals that WDR5 is required for efficient assembly and chromatin targeting of the NSL complex. *Genes Dev*. 2014;28(9). doi:10.1101/GAD.240200.114

15. Sheikh BN, Guhathakurta S, Akhtar A. The non-specific lethal (NSL) complex at the crossroads of transcriptional control and cellular homeostasis . *EMBO Rep*. 2019;20(7). doi:10.15252/embr.201847630

16. Radzisheuskaya A, Shliaha P V., Grinev V V., et al. Complex-dependent histone acetyltransferase activity of KAT8 determines its role in transcription and cellular homeostasis. *Mol Cell*. 2021;81(8):1749-1765.e8. doi:10.1016/J.MOLCEL.2021.02.012

17. Feller C, Prestel M, Hartmann H, Straub T, Söding J, Becker PB. The MOF-containing NSL complex associates globally with housekeeping genes, but activates only a defined subset. *Nucleic Acids Res*. 2012;40(4):1509-1522. doi:10.1093/NAR/GKR869

18. Ravens S, Fournier M, Ye T, et al. Mof-associated complexes have overlapping and unique roles in regulating pluripotency in embryonic stem cells and during differentiation. *Elife*. 2014;2014(3). doi:10.7554/ELIFE.02104.001

19. Gaub A, Sheikh BN, Basilicata MF, et al. Evolutionary conserved NSL complex/BRD4 axis controls transcription activation via histone acetylation. *Nat Commun 2020 111*. 2020;11(1):1-17. doi:10.1038/s41467-020-16103-0

20. Füllgrabe J, Lynch-Day MA, Heldring N, et al. The histone H4 lysine 16

acetyltransferase hMOF regulates the outcome of autophagy. *Nature*. 2013;500(7463):468-471.

21. Hale CM, Cheng Q, Ortuno D, et al. Identification of modulators of autophagic flux in an image-based high content siRNA screen. *Autophagy*. 2016;12(4):713-726. doi:10.1080/15548627.2016.1147669/SUPPL_FILE/KAUP_A_1147669_SM6249.ZIP

22. Linda K, Lewerissa EI, Verboven AHA, et al. Imbalanced autophagy causes synaptic deficits in a human model for neurodevelopmental disorders. *Autophagy*. Published online 2021. doi:10.1080/15548627.2021.1936777/SUPPL_FILE/KAUP_A_1936777_SM2241.ZIP

23. Li T, Lu D, Yao C, et al. Kansl1 haploinsufficiency impairs autophagosome-lysosome fusion and links autophagic dysfunction with Koolen-de Vries syndrome in mice. *Nat Commun 2022 131*. 2022;13(1):1-16. doi:10.1038/s41467-022-28613-0

24. Chatterjee A, Seyfferth J, Lucci J, et al. MOF Acetyl Transferase Regulates Transcription and Respiration in Mitochondria. *Cell*. 2016;167(3):722-738.e23. doi:10.1016/j.cell.2016.09.052

25. Lai Y-C, Kondapalli C, Lehneck R, et al. Phosphoproteomic screening identifies Rab GTPases as novel downstream targets of PINK1. *EMBO J*. 2015;34(22):2840-2861. doi:10.15252/EMBJ.201591593

26. Soutar MPM, Kempthorne L, Miyakawa S, et al. AKT signalling selectively regulates PINK1 mitophagy in SHSY5Y cells and human iPSC-derived neurons. *Sci Rep*. 2018;8(1):8855. doi:10.1038/s41598-018-26949-6

27. Ferrari R, Kia DA, Tomkins JE, et al. Stratification of candidate genes for Parkinson's disease using weighted protein-protein interaction network analysis. *BMC Genomics*. 2018;19(1):452. doi:10.1186/s12864-018-4804-9

28. Nalls MA, Pankratz N, Lill CM, et al. Large-scale meta-analysis of genome-wide association data identifies six new risk loci for Parkinson's disease. *Nat Genet*. 2014;46(9):989-993. doi:10.1038/ng.3043

29. Kia DA, Zhang D, Guelfi S, et al. Integration of eQTL and Parkinson's disease GWAS data implicates 11 disease genes. *bioRxiv*. Published online 2019:627216. doi:10.1101/627216

30. Pruim RJ, Welch RP, Sanna S, et al. LocusZoom: regional visualization of genome-

wide association scan results. *Bioinformatics*. 2010;26(18):2336-2337. doi:10.1093/bioinformatics/btq419

31. Marti-Solano M, Crilly SE, Malinverni D, et al. Combinatorial expression of GPCR isoforms affects signalling and drug responses. *Nat 2020 5877835*. 2020;587(7835):650-656. doi:10.1038/s41586-020-2888-2

32. Papatheodorou I, Moreno P, Manning J, et al. Expression Atlas update: from tissues to single cells. *Nucleic Acids Res*. 2020;48(D1):D77-D83. doi:10.1093/NAR/GKZ947

33. Ardley HC, Scott GB, Rose SA, Tan NGS, Markham AF, Robinson PA. Inhibition of proteasomal activity causes inclusion formation in neuronal and non-neuronal cells overexpressing Parkin. *Mol Biol Cell*. 2003;14(11):4541-4556. doi:10.1091/mbc.E03-02-0078

34. Soutar MPM, Kempthorne L, Annuario E, et al. FBS/BSA media concentration determines CCCP's ability to depolarize mitochondria and activate PINK1-PRKN mitophagy. *Autophagy*. 2019;15(11):2002-2011. doi:10.1080/15548627.2019.1603549

35. Deas E, Plun-Favreau H, Gandhi S, et al. PINK1 cleavage at position A103 by the mitochondrial protease PARL. *Hum Mol Genet*. 2011;20(5):867-879. doi:10.1093/HMG/DDQ526

36. Frega M, van Gestel SHC, Linda K, et al. Rapid Neuronal Differentiation of Induced Pluripotent Stem Cells for Measuring Network Activity on Micro-electrode Arrays. *J Vis Exp*. 2017;(119):e54900. doi:10.3791/54900

37. Fernandopulle MS, Prestil R, Grunseich C, Wang C, Gan L, Ward ME. Transcription Factor-Mediated Differentiation of Human iPSCs into Neurons. *Curr Protoc cell Biol*. 2018;79(1). doi:10.1002/CPCB.51

38. Tian R, Gachechiladze MA, Ludwig CH, et al. CRISPR interference-based platform for multimodal genetic screens in human iPSC-derived neurons. *Neuron*. 2019;104(2):239. doi:10.1016/J.NEURON.2019.07.014

39. Guelfi S, D'Sa K, Botía JA, et al. Regulatory sites for splicing in human basal ganglia are enriched for disease-relevant information. *Nat Commun 2020 111*. 2020;11(1):1-16. doi:10.1038/s41467-020-14483-x

40. Greene JC, Whitworth AJ, Kuo I, Andrews LA, Feany MB, Pallanck LJ. Mitochondrial pathology and apoptotic muscle degeneration in Drosophila

parkin mutants. *Proc Natl Acad Sci*. 2003;100(7):4078 LP - 4083. doi:10.1073/pnas.0737556100

41. Whitworth AJ, Theodore DA, Greene JC, Beneš H, Wes PD, Pallanck LJ. Increased glutathione S-transferase activity rescues dopaminergic neuron loss in a Drosophila model of Parkinson's disease. *Proc Natl Acad Sci*. 2005;102(22):8024 LP - 8029. doi:10.1073/pnas.0501078102

42. Orchard S, Ammari M, Aranda B, et al. The MIntAct project—IntAct as a common curation platform for 11 molecular interaction databases. *Nucleic Acids Res*. 2014;42(D1):D358-D363. doi:10.1093/NAR/GKT1115

43. Giambartolomei C, Vukcevic D, Schadt EE, et al. Bayesian Test for Colocalisation between Pairs of Genetic Association Studies Using Summary Statistics. Williams SM, ed. *PLoS Genet*. 2014;10(5):e1004383. doi:10.1371/journal.pgen.1004383

44. Ramasamy A, Trabzuni D, Guelfi S, et al. Genetic variability in the regulation of gene expression in ten regions of the human brain. *Nat Neurosci*. 2014;17(10):1418-1428. doi:10.1038/nn.3801

45. Lonsdale J, Thomas J, Salvatore M, et al. The Genotype-Tissue Expression (GTEx) project. *Nat Genet*. 2013;45(6):580-585. doi:10.1038/ng.2653

46. Robak LA, Jansen IE, Van Rooij J, et al. Excessive burden of lysosomal storage disorder gene variants in Parkinson's disease. *Brain*. 2017;140(12):3191-3203. doi:10.1093/brain/awx285

47. Hou X, Fiesel FC, Truban D, et al. Age- and disease-dependent increase of the mitophagy marker phospho-ubiquitin in normal aging and Lewy body disease. *Autophagy*. 2018;14(8):1404-1418. doi:10.1080/15548627.2018.1461294

48. Gusev A, Ko A, Shi H, et al. Integrative approaches for large-scale transcriptome-wide association studies. *Nat Genet*. 2016;48(3):245-252. doi:10.1038/ng.3506

49. Simon RP, Robaa D, Alhalabi Z, Sippl W, Jung M. KATching-Up on Small Molecule Modulators of Lysine Acetyltransferases. *J Med Chem*. 2016;59(4):1249-1270. doi:10.1021/acs.jmedchem.5b01502

50. Sheikh BN, Akhtar A. The many lives of KATs — detectors, integrators and modulators of the cellular environment. *Nat Rev Genet*. 2019;20(January). doi:10.1038/s41576-018-0072-4

51. Katayama H, Kogure T, Mizushima N, Yoshimori T, Miyawaki A. A sensitive and quantitative technique for detecting autophagic events based on lysosomal delivery. *Chem Biol*. 2011;18(8):1042-1052. doi:10.1016/j.chembiol.2011.05.013

52. Raja SJ, Charapitsa I, Conrad T, et al. The Nonspecific Lethal Complex Is a Transcriptional Regulator in Drosophila. *Mol Cell*. 2010;38(6):827-841. doi:10.1016/j.molcel.2010.05.021

53. Wray S, Lewis PA. A tangled web - tau and sporadic Parkinson's disease. *Front Psychiatry*. 2010;1(DEC):1-7. doi:10.3389/fpsyt.2010.00150

54. Stefansson H, Helgason A, Thorleifsson G, et al. A common inversion under selection in Europeans. *Nat Genet*. 2005;37(2):129-137. doi:10.1038/ng1508

55. Zody MC, Jiang Z, Fung HC, et al. Evolutionary toggling of the MAPT 17q21.31 inversion region. *Nat Genet*. 2008;40(9):1076-1083. doi:10.1038/ng.193

56. Pittman AM, Myers AJ, Abou-Sleiman P, et al. Linkage disequilibrium fine mapping and haplotype association analysis of the tau gene in progressive supranuclear palsy and corticobasal degeneration. *J Med Genet*. 2005;42(11):837-846. doi:10.1136/jmg.2005.031377

57. Hutton M, Lendon CL, Rizzu P, et al. Association of missense and 5′-splice-site mutations in tau with the inherited dementia FTDP-17. *Nature*. 1998;393(6686):702-705. doi:10.1038/31508

58. Healy DG, Abou-Sleiman PM, Lees AJ, et al. Tau gene and Parkinson's disease: a case–control study and meta-analysis. *J Neurol Neurosurg Psychiatry*. 2004;75(7):962-965. doi:10.1136/JNNP.2003.026203

59. Fung H-C, Scholz S, Matarin M, et al. Genome-wide genotyping in Parkinson's disease and neurologically normal controls: first stage analysis and public release of data. *Lancet Neurol*. 2006;5(11):911-916. doi:10.1016/S1474-4422(06)70578-6

60. Dong X, Liao Z, Gritsch D, et al. Enhancers active in dopamine neurons are a primary link between genetic variation and neuropsychiatric disease. *Nat Neurosci*. 2018;21(10):1482-1492. doi:10.1038/s41593-018-0223-0

61. Ferrari R, Wang Y, Vandrovcova J, et al. Genetic architecture of sporadic frontotemporal dementia and overlap with Alzheimer's and Parkinson's diseases. *J Neurol Neurosurg Psychiatry*. 2017;88(2):152-164. doi:10.1136/JNNP-2016-314411

62. Koks S, Pfaff AL, Bubb VJ, Quinn JP. Transcript Variants of Genes Involved in Neurodegeneration Are Differentially Regulated by the APOE and MAPT Haplotypes. *Genes 2021, Vol 12, Page 423*. 2021;12(3):423. doi:10.3390/GENES12030423

63. Soto-Beasley AI, Walton RL, Valentino RR, et al. Screening non-MAPT genes of the Chr17q21 H1 haplotype in Parkinson's disease. *Parkinsonism Relat Disord*. 2020;78:138-144. doi:10.1016/J.PARKRELDIS.2020.07.022

64. Swatek KN, Komander D. Ubiquitin modifications. *Cell Res*. 2016;26(4):399-422. doi:10.1038/cr.2016.39

65. Koolen DA, Pfundt R, Linda K, et al. The Koolen-de Vries syndrome: a phenotypic comparison of patients with a 17q21.31 microdeletion versus a KANSL1 sequence variant. *Eur J Hum Genet 2016 245*. 2015;24(5):652-659. doi:10.1038/ejhg.2015.178

66. Li L, Ghorbani M, Weisz-Hubshman M, et al. Lysine acetyltransferase 8 is involved in cerebral development and syndromic intellectual disability. *J Clin Invest*. 2020;130(3):1431-1445. doi:10.1172/JCI131145

67. Valente EM, Abou-Sleiman PM, Caputo V, et al. Hereditary early-onset Parkinson's disease caused by mutations in PINK1. *Science (80-)*. 2004;304(5674):1158-1160. doi:10.1126/science.1096284

68. Kitada T, Asakawa S, Hattori N, et al. Mutations in the parkin gene cause autosomal recessive juvenile parkinsonism. *Nature*. 1998;392(6676):605-608. doi:10.1038/33416

69. Chu CT. Mechanisms of selective autophagy and mitophagy: Implications for neurodegenerative diseases. *Neurobiol Dis*. 2019;122(July 2018):23-34. doi:10.1016/j.nbd.2018.07.015

70. Lek M, Karczewski KJ, Minikel E V, et al. Analysis of protein-coding genetic variation in 60,706 humans. *Nature*. 2016;536(7616):285-291. doi:10.1038/nature19057

FIGURE LEGENDS

Figure 1 High content mitophagy screen of PD risk genes identifies KAT8 as a modulator of pUb(Ser65) levels. A. Workflow of the high content screen for O/A-induced pUb(Ser65) levels. **B.** pUb(Ser65) Z-scores of one representative mitophagy screen plate. **C.** Overview of the PD GWAS genetic signal at the *KAT8* locus. **D.** Representative IB of mitochondrial fractions from SCR, PINK1 and KAT8 KD POE SHSY5Y treated with 1 μM O/A for 1.5 or 3 h. **E.** Quantification of pUb(Ser65) in E (n=5, one-way ANOVA with Dunnett's correction). **F.** Quantification of PINK1 in E (n=4, one-way ANOVA with Dunnett's correction). **G.** Quantification of KAT8 in E (n=5, one-way ANOVA with Dunnett's correction). **H.** pUb(Ser65) Z-scores of one representative KAT screen plate. See Supplementary Table 5 for the complete list of the genes screened. Data are shown as mean ± SD.

Figure 2 Knockdown of KANSL1 affects pUb(Ser65) levels. A. Quantification of pUb(Ser65) following treatment of SCR, PINK1 or NSL components siRNA KD POE SHSY5Y cells with 1 μM O/A for 1.5 h. Data are shown as mean ± SD; n=6, one-way ANOVA with Dunnett's correction. **B.** Representative images of pUb(Ser65) (green) following treatment of SCR, PINK1 and KANSL1 KD POE SHSY5Y cells with 1 μM O/A for 3 h. Insets show the nuclei (blue) for the same fields. Scale bar = 20 μm. **C.** Quantification of pUb(Ser65) in B (n=3, two-way ANOVA with Dunnett's correction). **D**. Representative IB of mitochondrial fractions from SCR, PINK1 and KANSL1 KD POE SHSY5Y treated with 1 μM O/A for 1.5 or 3 h. **E.** Quantification of pUb(Ser65) in D (n=5, one-way ANOVA with Dunnett's correction). **F.** Quantification of PINK1 in D (n=4, one-way ANOVA with Dunnett's correction). Data are shown as mean ± SD.

Figure 3 KAT8 and KANSL1 knockdown decreases PINK1-dependent mitophagy initiation. A. Representative images of pUb(Ser65) (green) following treatment of SCR, PINK1, KAT8 and KANSL1 KD POE SHSY5Y cells with 1 μM O/A for 0-7 h. Insets show the nuclei (blue) for the same fields. Scale bar = 20 μm. **B.** Quantification of pUb(Ser65) in A (n=6, two-way ANOVA with Dunnett's correction). For details on the statistical test, see Supplementary Table 6. **C.** Representative images of FLAG-Parkin (green) with Hoechst nuclei counterstain (blue) following treatment of SCR, PINK1 and KAT8 siRNA KD POE

SHSY5Y with 1 µM O/A for 3 h. Scale bar = 20 µm. **D.** Quantification of FLAG-Parkin recruitment to the mitochondria as a ratio of FLAG intensity in the mitochondria and in the cytosol in C (n=5, two-way ANOVA with Dunnett's correction). **E.** Representative images of pParkin (green) with Hoechst nuclei counterstain (blue) following treatment of SCR, PINK1 and KAT8 siRNA KD POE SHSY5Y with 1 µM O/A for 3 h. Scale bar = 20 µm. **F.** Quantification of pParkin levels in E (n=5, two-way ANOVA with Dunnett's correction). Data are shown as mean ± SD.

Figure 4 RT-qPCR Assessments of *KANSL1, KAT8, PINK1* and *PRKN* gene expression in POE and WT SHSY5Ys. A-C. RT-qPCR quantification of *KANSL1* mRNA (A), *KAT8* mRNA (B) and *PINK1* mRNA (C) in POE SHSY5Ys (n=6, one-way ANOVA with Dunnett's correction). **D-G.** RT-qPCR quantification of *KANSL1* mRNA (D), *KAT8* mRNA (E), *PINK1* mRNA (F) and *PRKN* mRNA (G) in WT SHSY5Ys (n=5, one-way ANOVA with Dunnett's correction). Data are shown as mean ± SD.

Figure 5 KANSL1 and KAT8 knockdown decrease mitochondrial clearance. A. Representative images of mt-Keima following treatment of SCR, PINK1 and KAT8 siRNA KD POE SHSY5Y with 1 µM O/A for 0-8 h. The first and third rows show the neutral Keima-green signal (green) counterstained with Hoechst (blue) after 0 h and 6 h respectively of DMSO vs O/A. The second and fourth rows show the acidic lysosomal Keima-red signal (red) counterstained with Hoechst (blue) after 0 h and 6 h respectively of DMSO vs O/A. Scale bar = 25 µm. **B.** Quantification of the mitophagy index, calculated as the ratio of the area of lysosomal mt-Keima signal and total mt-Keima signal in A (n=3, one-way ANOVA with Dunnett's correction). For details on the statistical test, see Supplementary Table 7. Data are shown as mean ± SD.

Figure 6 *KANSL1* presents ASE sites in LD with the H1/H2 SNP, and pUb(Ser65) levels are altered by siRNA KD of KANSL1 but not other genes present at the 17q21 locus. A. ASEs derived from putamen and substantia nigra in high linkage disequilibrium with the H1/H2 tagging SNP, rs12185268 and their position along the *KANSL1* gene. The missense variants track displays the variants annotated as missense by gnomAD v2.1.1[70]. The valid track displays the heterozygous sites (orange = missense) with an average read depth greater

than 15 reads across all samples, which were examined for ASE. The topmost track displays the FDR-corrected minimum -log10 p-value across samples for the sites that show an ASE in at least one sample. **B.** Conservation of the KANSL1 protein across species. The four coding variants (NM_001193465) in the *KANSL1* gene are in high LD (r2 >0.8) with the H1/H2 haplotypes. **C.** pUb(Ser65) Z-scores of one representative 17q21 locus screen plate. See Supplementary Table 10 for the complete list of the genes screened.

Figure 7 pUb(Ser65) levels are reduced in isogenic iNeurons with heterozygous KANSL1$^{+/-}$ loss of function and CRISPRi-i3N iNeurons with KANSL1 and KAT8 sgRNA KD A. Representative IB of isogenic d17 iNeurons with/without heterozygous LoF frameshift mutation in KANSL1 treated with 1 µM O/A over a 12 h extended time-course. **B.** Quantification of pUb(Ser65) in A (n=4 inductions, two-way ANOVA with Dunnett's correction). **C.** Representative images of pUb(Ser65) (yellow) with Hoechst nuclei counterstain (blue) following treatment of non-transduced (No TD), non-targeting, KANSL1, KAT8 and PINK1 sgRNA KD d17 CRISPRi-i3N iNeurons with 1 µM O/A vs DMSO for 9 h. Inserts show staining for TOM20 (cyan) and mCherry transduction reporter (magenta) with Hoechst nuclei counterstain (blue) for the same field of view. Scale bar = 100 µm. **D.** Quantification of pUb(Ser65) intensity in TOM20 defined mitochondrial area in D (n=3 inductions, two-way ANOVA with Dunnett's correction). Data are shown as mean ± SD.

Table 1 Overview of bioinformatic evidence for genes prioritised and taken forward for downstream functional analysis

Gene	ColB	ColG	PPI	GWAS	MPD	MLS
ATP13A2					X	X
CCNT2				X		
CD38	X	X		X		
CTSB	X	X	X			
DDRGK1				X		
DGKQ				X		
DJ1					X	
DNAJC13					X	
FBXO7					X	
GALC	X			X		X
GBA			X		X	X
GPNMB	X	X		X		
HSD3B7		X				
IDUA						X
INPP5F			X			
KAT8		X	X	X		
KLHL7		X		X		
LRRK2			X	X	X	
LSM7	X	X		X		
MAPT			X	X		
NCKIPSD	X	X	X	X		
NEK1	X					
NSF			X			
NUCKS1		X		X		
NUPL2	X	X		X		
PDLIM2		X		X		
PM20D1	X					
PRKN					X	
RAB7L1	X	X	X			
SH3GL2			X			
SLC41A1		X		X		
SNCA					X	
SPPL2B	X					
STK39				X		
VAMP4	X	X				
VPS35					X	
WDR6	X	X				
ZNF646				X		

ColB = coloc analysis using Braineac; ColG = coloc analysis using GTEx; GWAS = genes prioritised in PD-GWAS[3]; MLS = Mendelian genes associated with lysosomal storage disorders; MPD = Mendelian genes associated with PD; WPPINA = weighted protein interaction network.

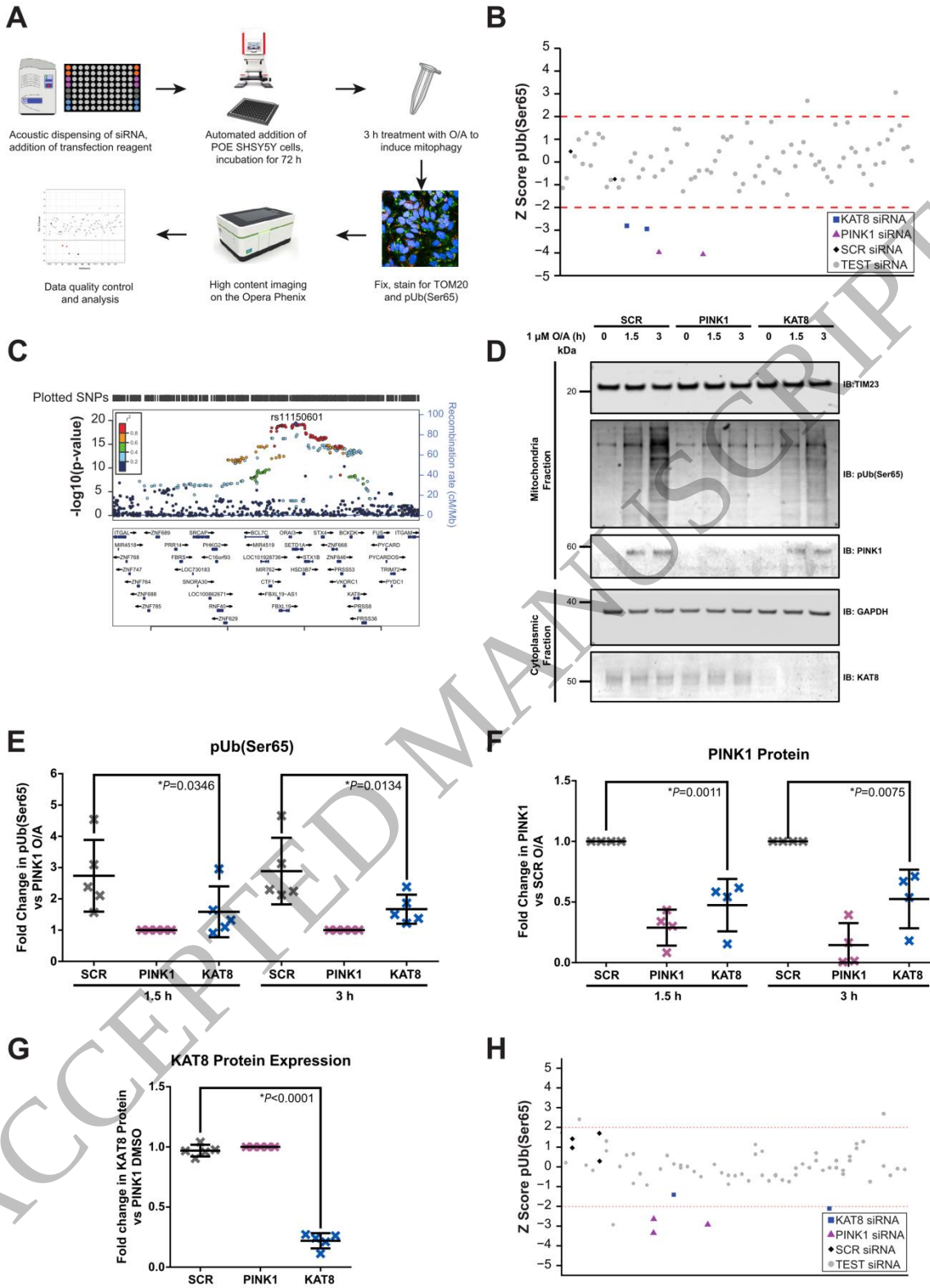

Figure 1
421x559 mm (.59 x DPI)

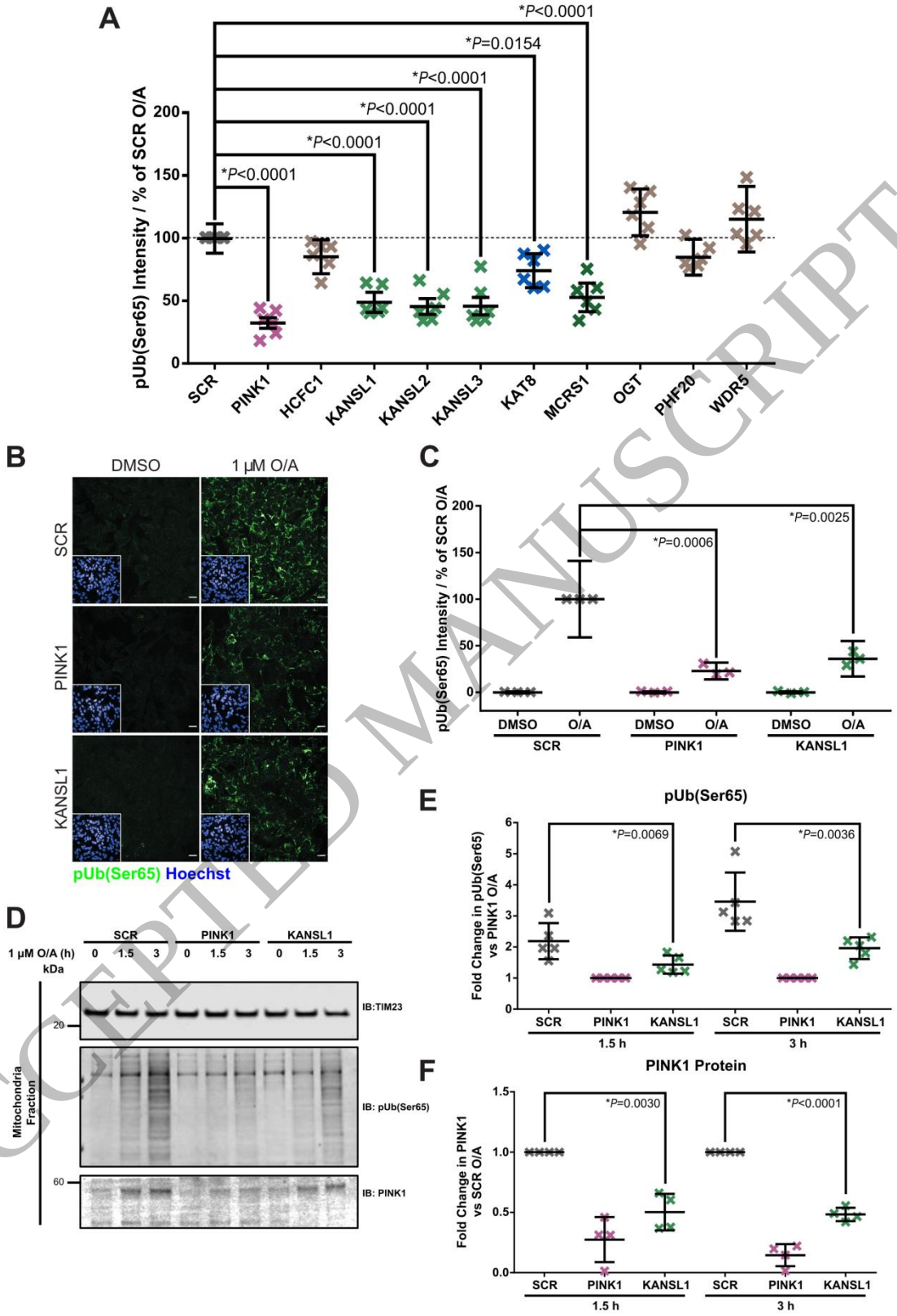

Figure 2
395x559 mm (.59 x DPI)

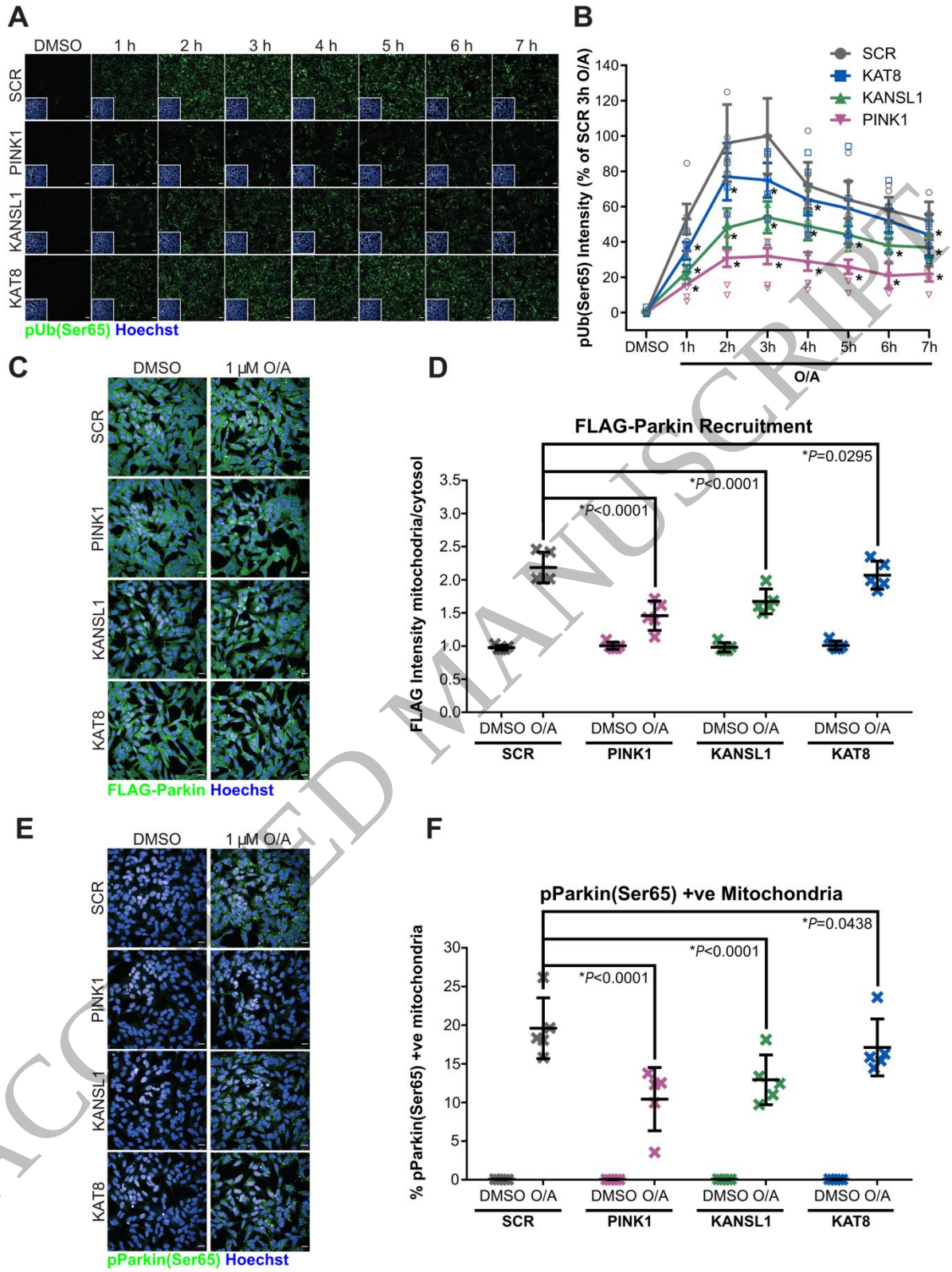

Figure 3
411x559 mm (.59 x DPI)

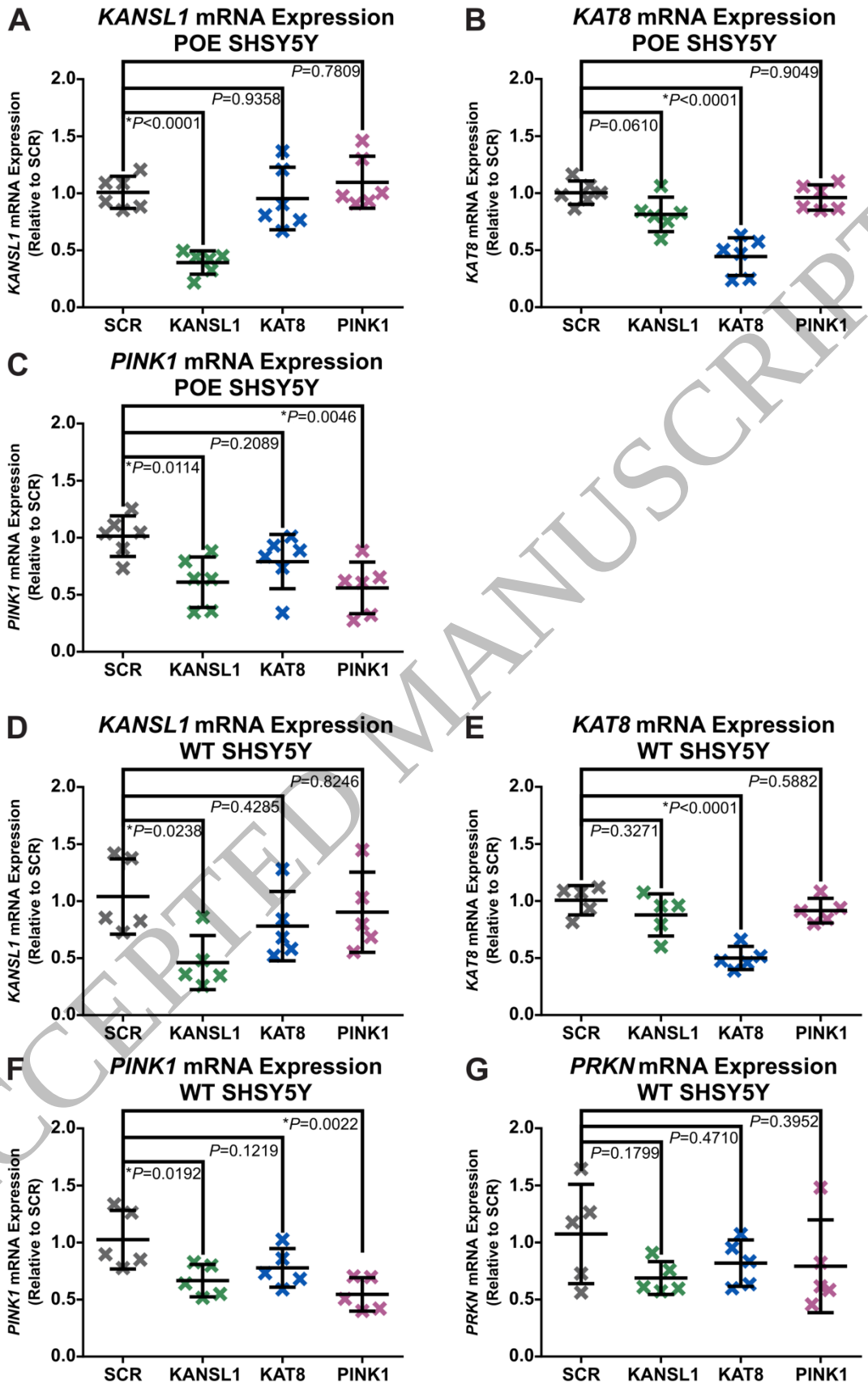

Figure 4
353x559 mm (.59 x DPI)

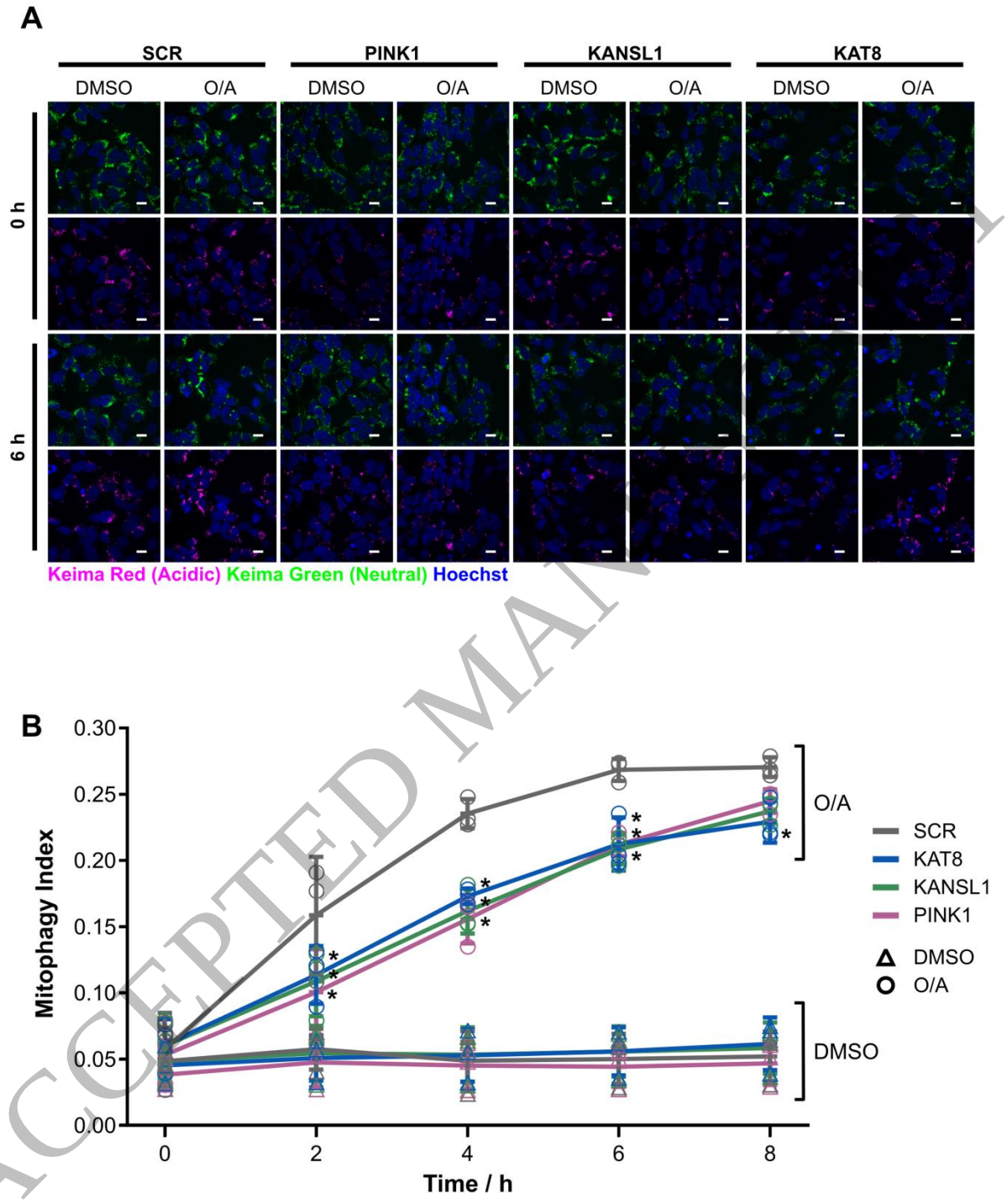

Figure 5
460x559 mm (.59 x DPI)

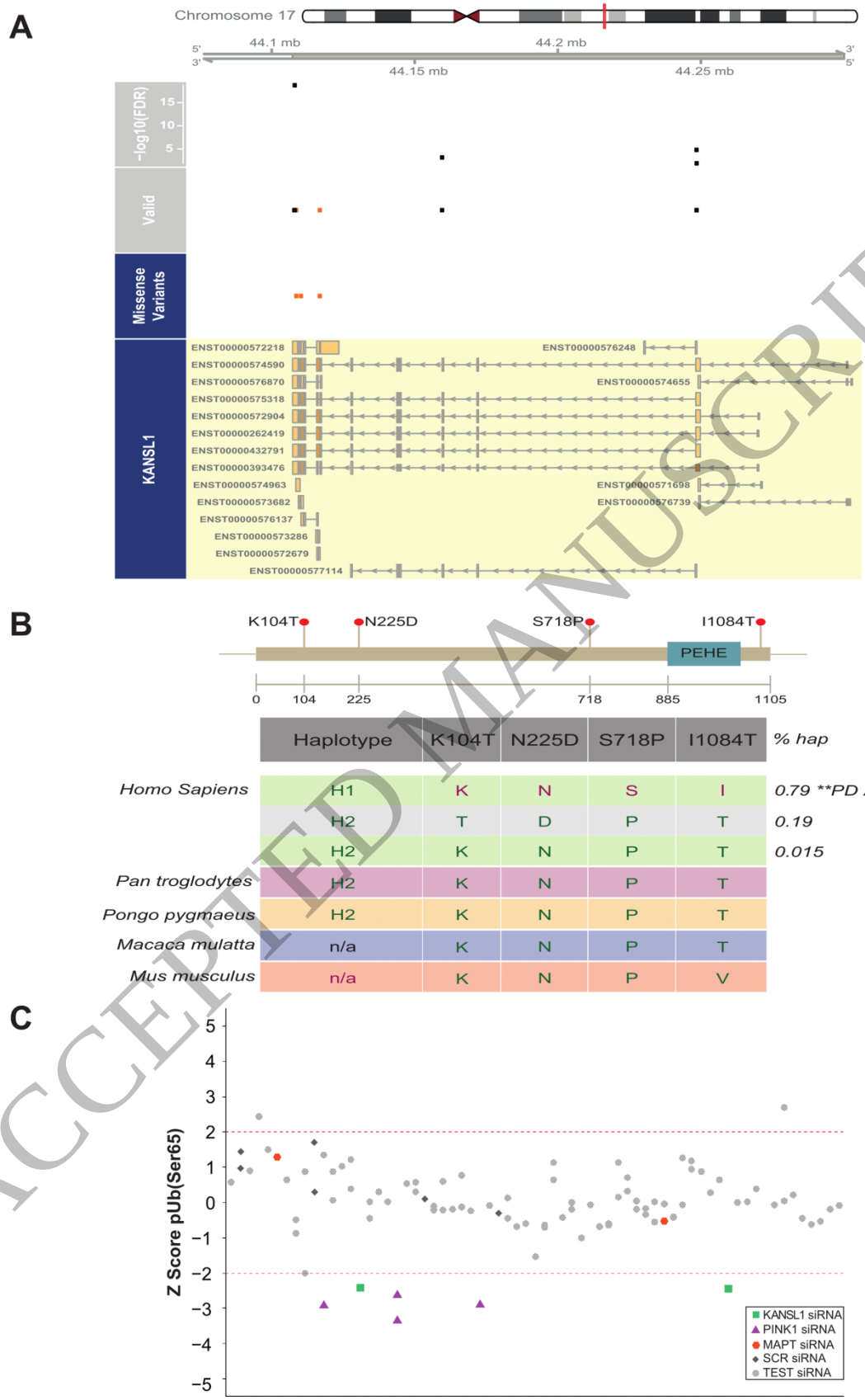

Figure 6
380x559 mm (.59 x DPI)

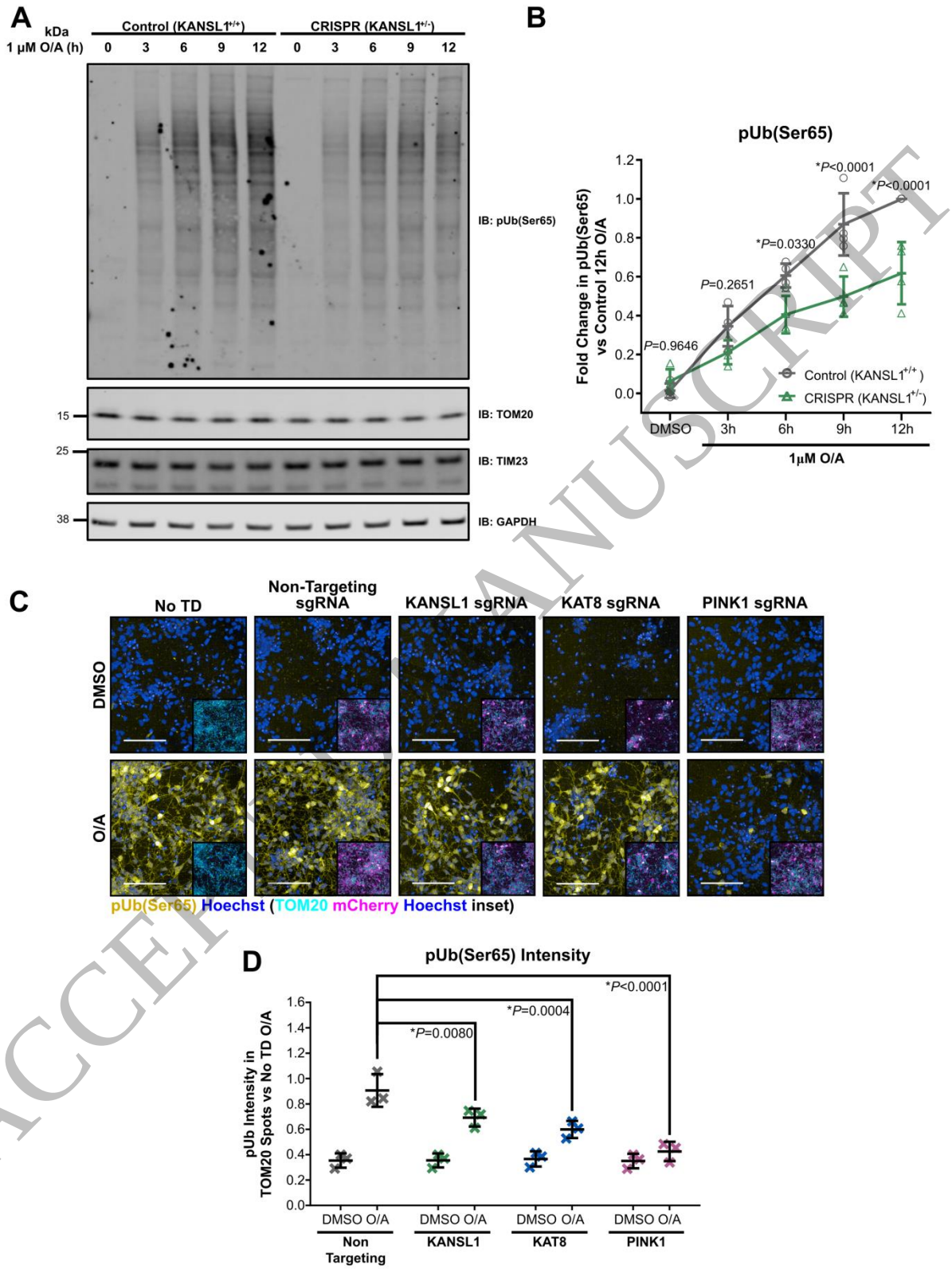

Figure 7
414x559 mm (.59 x DPI)