

1 **Patients with XLP Type 1 have variable numbers of NKT cells**

2 Elizabeth Ralph, MSc

3 Great Ormond Street Hospital for Children NHS Foundation Trust

4 Josie Evans, PhD

5 Great Ormond Street Hospital for Children NHS Foundation Trust

6 Claire Booth, MBBS, PhD

7 Great Ormond Street Hospital for Children NHS Foundation Trust

8 Kimberly Gilmour, PhD

9 Great Ormond Street Hospital for Children NHS Foundation Trust

10

11 Corresponding author:

12 Elizabeth Ralph, Immunology Laboratory, Camelia Botnar Laboratories, Great

13 Ormond Street Hospital for Children NHS Foundation Trust, London, WC1N 3JH.

14 +44 20 7829 8835 elizabeth.ralph@gosh.nhs.uk

15

16

17 **Patients with XLP Type 1 have variable numbers of NKT cells**

18

19 **Abstract**

20 X-linked lymphoproliferative disease (XLP1) is a rare primary immunodeficiency that
21 usually presents in early childhood. Patients with XLP1 have been reported to have
22 absent NKT cells, and it has been suggested that this can be diagnostic for the
23 disorder. Whilst NKT frequency in adults is variable, little is known about their
24 frequency in children. Therefore, we established a paediatric reference range for
25 these cells. In contrast to previous reports, in our cohort of XLP1 patients NKT cell
26 numbers were found to be variable, and we would advise against using the finding of
27 NKT cells to exclude a diagnosis of XLP1.

28

29 **Background**

30 NKT cells are a sublineage of T cells with unique properties, that include expression
31 of an invariant T cell receptor ($V\alpha_{24}V\beta_{11}$ in humans) which binds glycosphingolipids
32 presented by the MHC class I-like molecule, CD1d, and they have the ability to
33 rapidly produce many cytokines after stimulation¹. NKT cells develop in the thymus
34 and are positively selected by CD1d expressing bone marrow cells, rather than by
35 cortical epithelial cells like conventional T cells. NKT cells are most prevalent in the
36 thymus, spleen, liver and bone marrow and are much less abundant in lymph nodes.
37 NKT cells make up only approximately 0.1% of human peripheral blood T cells.
38 There is a high degree of variability in the frequency of NKT cells in the peripheral
39 blood of normal individuals. In a study of 70 normal controls, NKT cell numbers
40 varied from 10 – 30,000 per mL². Although SLAM-associated protein (SAP) is not
41 required for the development of the majority of lymphocytes, it plays a crucial role in

42 the development of NKT cells. *SAP*^{-/-} mice have greatly reduced numbers of NKT
43 cells in secondary lymphoid organs^{3,4}.

44

45 X-linked lymphoproliferative disease (XLP1) is a rare primary immunodeficiency
46 disorder in which the gene (*SH2D1A*) which encodes the protein SAP is defective.
47 XLP1 patients have been reported to have greatly reduced numbers of NKT cells in
48 peripheral blood^{4,5}. In one study⁴ XLP1 patients (n = 17) had 0 – 58 NKT cells/mL
49 peripheral blood which was greatly reduced compared to the numbers in normal
50 individuals: 10 controls with 120 – 1596 NKT cells/mL. These studies used small
51 numbers of control individuals (8 age matched controls (1 to 27 years old)⁵, 10
52 healthy individuals (unspecified ages)⁴). As the number of NKT cells in peripheral
53 blood is known to be extremely variable in adults² and the average age of onset of
54 XLP1 is 2.5 years⁶, reference ranges for peripheral blood NKT cells in children need
55 to be established using larger numbers of controls as a preliminary step to
56 determining whether NKT cell enumeration may aid in the diagnosis of XLP1.

57

58 **Aim**

59 The aim of this study was to develop a paediatric reference range for NKT cells in
60 peripheral blood and compare NKT cell numbers in children and in adults. We then
61 aimed to enumerate NKT cells in known XLP1 patients and to determine the
62 usefulness of NKT cell enumeration in the diagnosis of XLP1.

63

64 **Methods and materials**

65 The method for NKT cell enumeration was based on a published method².
66 Peripheral blood was stained with directly conjugated fluorescent monoclonal

67 antibodies to CD3 and V α 24/V β 11 subunits of the NKT cell invariant TCR. Following
68 lysis of erythrocytes, cells were washed and re-suspended in fixative and analysed
69 by flow cytometry. Absolute cell counts were calculated based on the proportion of
70 beads acquired compared to cells acquired, **and also by using the lymphocyte**
71 **count obtained from the full blood count.** Samples from 28 pediatric controls, 15
72 adult controls, and 5 XLP1 patients were analysed.

73

74 **Results**

75 NKT cells were identified by flow cytometry as CD3+ T lymphocytes expressing the
76 invariant TCR, V α 24V β 11 (**Figure 1**, R1 and R2 and R4). NKT cell counts were very
77 variable and showed a skewed distribution, tending towards lower NKT cell numbers.
78 (**Figure S1, and Table S2**).

79

80 **The range for paediatric samples was 24-8766 NKT cells/mL, with a median of**
81 **658 NKT cells/mL The adult range was determined as 27-7572 NKT cells/mL**
82 **with a median value of 1142 NKT cells/mL** There was no significant difference
83 between paediatric NKT cell counts and adult NKT cell counts ($p=0.168$ using 2-
84 tailed Mann-Whitney U test)

85

86 Peripheral blood NKT cells were enumerated in 5 known XLP1 patients. NKT cell
87 counts ranged from 12 to 115 cells/mL Two patients (Patient 1 and Patient 2) had an
88 NKT cell count within the paediatric reference range (**Figure 2**). Patients with very
89 low NKT cell numbers had several different mutations, while the two patients with an
90 NKT cell count within the paediatric reference range had a deletion of exon 2 and 4
91 in SH2D1A (**Table S1**).

92

93 Discussion

94 **Peripheral blood NKT cell numbers are variable in children and adults, and**
95 **showed a skewed distribution.** There was no significant difference between
96 paediatric and adult NKT cell counts in this study. NKT cell numbers in 2 known
97 XLP1 patients (Patient 1 and Patient 2) were low but within the normal range, whilst
98 NKT cell numbers were low and below the reference range in 3 patients. This is in
99 contrast to previous reports^{4,5} which found that NKT cells were absent in XLP1
100 patients. Patient 1 and 2 are noted to be siblings and share the same mutation; it is
101 unclear if and how this exon deletion affects NKT cell development. **Further work to**
102 **establish NKT numbers and any correlation of genotype and phenotype in a**
103 **larger cohort of XLP1 patients through a multi-centre study would be**
104 **beneficial, although the rarity of the condition and the fact that most of the**
105 **patient have been treated with HSCT makes this difficult.**

106

107 In summary, as NKT cell numbers are variable and are not completely absent in
108 XLP1 patients, 'normal' numbers of NKT cells should not exclude a diagnosis of
109 XLP1. We recommend that measurement of SAP protein by flow cytometry, followed
110 by confirmatory genetic analysis should be undertaken where a diagnosis of XLP1 is
111 suspected.

112

113 Acknowledgments

114 Gene sequencing was done at the Molecular Genetics Laboratory at Great Ormond
115 Street Hospital

116

117 *Declaration of funding*

118 This report is independent research by the National Institute for Health Research Biomedical
119 Research Centre Funding Scheme. The views expressed in this publication are those of the
120 author(s) and not necessarily those of the NHS, the National Institute for Health Research or
121 the Department of Health. This report is independent research supported by the National
122 Institute for Health Research Great Ormond Street Hospital Biomedical Research Centre.
123 The views expressed in this publication are those of the author(s) and not necessarily those
124 of the NHS, the National Institute for Health Research or the Department of Health.

125

126 *Authorship*

127 ER, JH and KG designed the research study, ER and JH performed the research
128 and analysed the data, CB contributed patient data, ER wrote the manuscript, all
129 authors critically revised the manuscript and approved the final version.

130

131 **References**

132 **1** Kronenberg M. Toward an understanding of NKT cell biology: progress and
133 paradoxes. *Annu Rev Immunol.* 2005; 23:877-900

134 **2** van der Vliet HJ, Molling JW, von Blomberg BM, Kölgen W, Stam AG, de Gruijl TD
135 *et al.* Circulating Va24+Vb11+ NKT cell numbers and dendritic cell CD1d expression
136 in hepatitis C virus infected patients. *Clin Immunol.* 2005 Feb;114(2):183-9.

137 **3.** Chung B, Aoukaty A, Dutz J, Terhorst C, Tan R. Signaling lymphocytic activation
138 molecule-associated protein controls NKT cell functions. *J Immunol.* 2005 Mar
139 15;174(6):3153-7.

140 **4.** Nichols KE, Hom J, Gong SY, Ganguly A, Ma CS, Cannons JL *et al.* Regulation of
141 NKT cell development by SAP, the protein defective in XLP. *Nat Med.* 2005
142 Mar;11(3):340-5.

- 143 5. Pasquier B, Yin L, Fondanèche MC, Relouzat F, Bloch-Queyrat C, Lambert N *et*
 144 *al.* Defective NKT cell development in mice and humans lacking the adapter SAP,
 145 the X-linked lymphoproliferative syndrome gene product. *J Exp Med.* 2005 Mar
 146 7;201(5):695-701.
- 147 6. Purtilo DT, Cassel CK, Yang JP, Harper R. X-linked recessive progressive
 148 combined variable immunodeficiency (Duncan's disease). *Lancet.* 1975 Apr
 149 26;1(7913):935-40.

150

151

152 **Figures:**

153

154

155

156

157

158

159

160

161

162

163

164

165

166

167

168

169

170

171

172

173

174

175

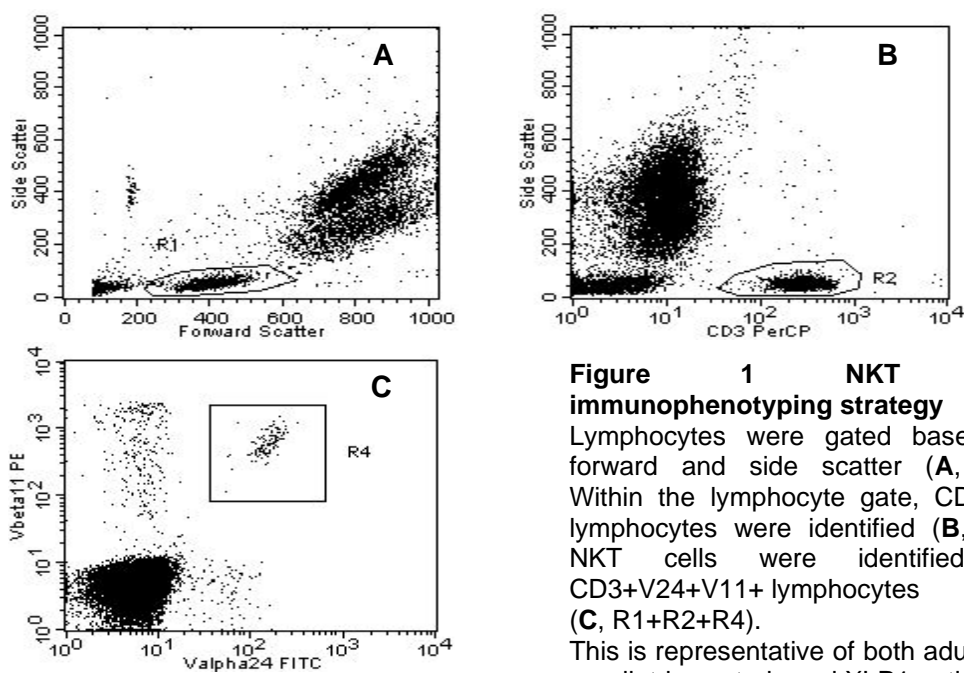
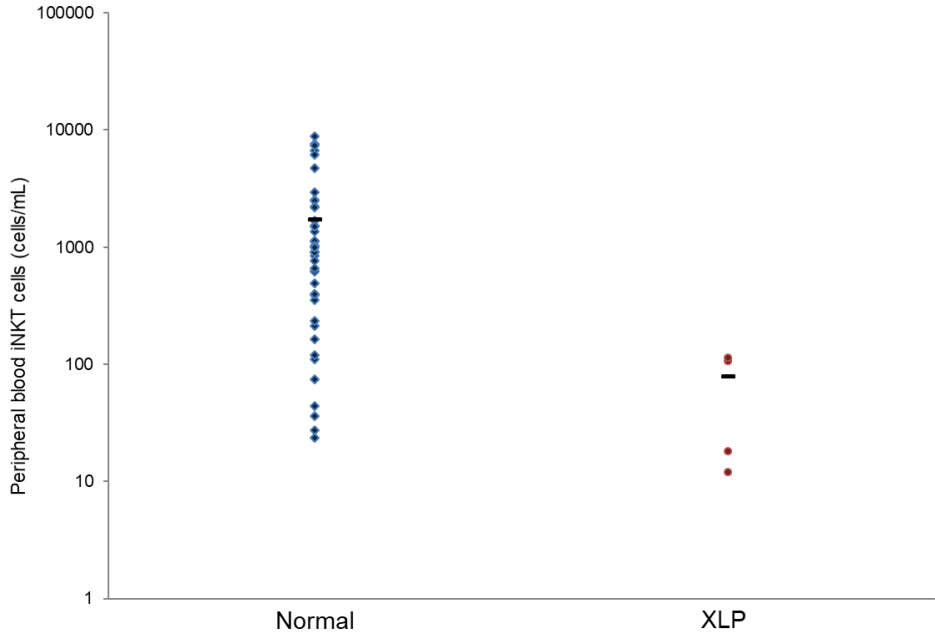


Figure 1 NKT cell immunophenotyping strategy

Lymphocytes were gated based on forward and side scatter (A, R1). Within the lymphocyte gate, CD3⁺ T lymphocytes were identified (B, R2). NKT cells were identified as CD3⁺V24⁺V11⁺ lymphocytes (C, R1+R2+R4).

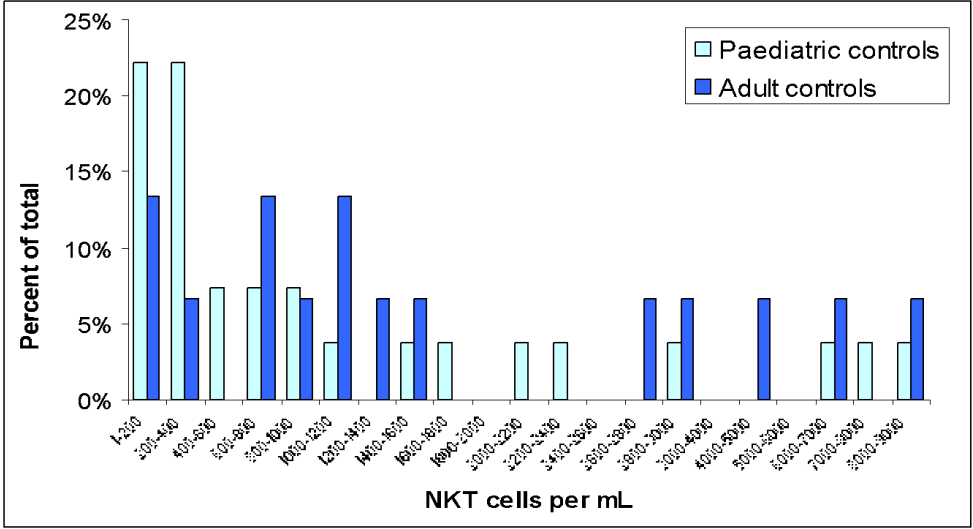
This is representative of both adult and paediatric controls and XLP1 patients



176
177
178
179
180
181
182
183
184
185

Figure 2
Paediatric peripheral blood NKT cells in normal controls and known XLP1 patients

NKT cells were enumerated in 28 normal controls (diamonds, line represents 50th percentile) and 5 known XLP1 patients (circles, line represents 50th percentile).



186
187
188
189
190
191
192
193
194
195
196

Figure S1
Distribution of NKT cell counts in adults and children

NKT cells were enumerated in duplicate in 28 children (light blue bars) and 15 adults (dark blue bars).

Patient	Mutation in <i>SH2D1A</i>	NKT cell count (/ml)
1	Deletion Exon 2 and 4	107
2	Deletion Exon 2 and 4	115
3	C245DUP	18
4	Deletion Exon 2	12
5	C163c>t	18

197
198
199
200

Table S1
Comparison of mutation in *SH2D1A* with absolute NKT cell count

Paediatric control	NKT cell count (/ml) (calculated using lymphocyte count)	Paediatric control	NKT cell count (/ml) (calculated using lymphocyte count)
1	374	15	1869
2	633	16	196
3	2478	17	32
4	383	18	2021
5	265	19	1670
6	861	20	2091
7	925	21	Lymphocyte count not available
8	290	22	115
9	637	23	49
10	113	24	1157
11	332	25	23
12	165	26	6028
13	178	27	8363
14	610		

201
202
203
204

Table S2
Paediatric peripheral blood NKT cells in normal controls, calculated using lymphocyte count.
NKT cells were enumerated in 28 normal controls