

Polymorphisms Predicting Phylogeny in Hepatitis B Virus (HBV)

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1 **ABSTRACT**

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Hepatitis B viruses (HBV) are compact viruses with circular genomes of ~3.2kb in length. Four genes (HBx, Core, Surface and Polymerase) generating seven products are encoded on overlapping reading frames. Ten HBV genotypes have been characterised (A-J), which may account for differences in transmission, outcomes of infection, and treatment response. However, HBV genotyping is rarely undertaken, and sequencing remains inaccessible in many settings. We used a machine learning approach based on random forest algorithms (RFA) to assess which amino acid (aa) sites in the genome are most informative for determining genotype. We downloaded 5496 genome-length HBV sequences from a public database, excluding recombinant sequences, regions with conserved indels, and genotypes I/J. Each gene was separately translated into aa, and the proteins concatenated into a single sequence (length 1614aa). Using RFA, we searched for aa sites predictive of genotype, and assessed co-variation among the sites with a Mutual Information (MI)-based method. We were able to discriminate confidently between genotypes A-H using 10 aa sites. 5/10 sites were identified in Polymerase (Pol), of which 4/5 were in the spacer domain, and a single site in reverse transcriptase. A further 4/10 sites were located in Surface protein, and a single site in HBx. There were no informative sites in Core. Properties of the aa were generally not conserved between genotypes at informative sites. Co-variation analysis identified 55 pairs of highly-linked sites. Three RFA-identified sites were represented across all pairs (two sites in spacer, and one in HBx). Residues that co-vary with these sites are concentrated in the small HBV surface gene. We also observe a cluster of sites adjacent to the Surface promoter region that co-vary with a spacer residue. Overall, we have shown that RFA analysis is a powerful tool for identifying aa sites that predict HBV lineage, with an unexpectedly high number of such sites in the spacer domain, which has conventionally been viewed as unimportant for structure or function. Our results improve ease of genotype prediction from limited regions of HBV sequence, and may have implications for understanding HBV evolution and the role of the spacer domain.

34 INTRODUCTION

35 Hepatitis B virus (HBV) is the prototype virus of the *hepadnaviridae* family, a family of small,
36 circular viruses with partially double-stranded (ds)DNA genomes of ~3.2kb in length(1). The
37 viral genome encodes seven proteins within four genes – HBx, Core, Polymerase and Surface
38 (Table 1; Figure S1) – together with associated regulatory elements(2), arranged in a series
39 of overlapping reading frames. This genome structure imposes constraints on selection, acting
40 as a stabilising selective force during replication(3, 4), and accounting for a reduced nucleotide
41 substitution rate in overlapping regions (approximately 40% lower than in the non-overlapping
42 regions)(5).

43
44 HBV DNA genomes are copied via RNA intermediates by means of an error-prone viral
45 reverse transcriptase (RT) enzyme(6), driving an evolutionary rate that is higher than would
46 be expected for a DNA virus with a high density of overlapping reading frames(1). The resulting
47 genetic diversity is the basis for the classification of HBV into ten genotypes (gt), defined by
48 $\geq 7.5\%$ nucleotide divergence(7), and designated gt-A-I, along with an unusual recombinant
49 putative gt-J (showing similarity to gt-C and gibbon *orthohepadnavirus*)(8). Genotypes are
50 further classified into subgenotypes based on $\geq 4\%$ divergence(7). There is variation in the
51 number of subgenotypes per genotype, ranging from >10 subgenotypes in gt-C(9) (reflecting
52 its status as the oldest lineage(5)), to just a single subtype in gt-E, -G and -H.

53
54 To date, HBV sequencing (and genotyping) is not recommended at baseline by clinical
55 guidelines and is not routinely undertaken to inform patient care, as there has been insufficient
56 evidence to support its role in informing surveillance or determining treatment courses(10).
57 However, as the pool of HBV sequence data expands, alongside linked clinical metadata,
58 progressive insights are emerging into associations between sequence heterogeneity
59 (including genotype, insertions, deletions and polymorphisms) and different clinical
60 phenotypes including treatment response and disease outcomes(10, 11).

61
62 Machine learning approaches are frequently applied to omics-based data, including
63 transcriptomics and proteomics(12). We set out to apply a machine learning approach based
64 on a random forest algorithm informed by full-length HBV sequences. Our aim was to identify
65 genome regions through which genotype can be predicted and to cast light on the selection
66 pressures that determine HBV genetic population structure.

67
68 **METHODS**

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70 Random forest algorithms (RFA) are a type of decision tree-based analysis, providing a
71 relatively hypothesis-free approach to interrogating complex data sets. The method has been
72 applied widely, including in host tropism studies in influenza(13), to identify molecules
73 inhibiting flaviviruses(14), to analyse mutational fitness effects in picornaviruses(15), and in
74 the identification of genes related to immunogenicity and pathogenicity in *Streptococcus*
75 *pneumoniae* infection(16, 17).

76
77 Briefly, this study included nucleotide alignments (n=5496) of HBV genotypes A-H(9) (Table
78 S1). Recombinant sequences were excluded from the analysis, as were genotypes I and J
79 which are recombinant in origin. Each of the overlapping HBV genes was separately translated
80 into amino acid (aa) sequences, which were then concatenated into a single sequence for
81 each genome (total length 1614 aa, Figure S1B). Residues were numbered and reported using
82 X02763 (gt-A) as a reference sequence, as is convention in the field(9). The RFA pipeline, as
83 detailed in the supplementary methods and Figures S3 and S4, was then applied to the
84 concatenated HBV sequences, using the known genotype of each sequence as the
85 classification variable and aa sites as predictive variables, in search of a parsimonious number
86 of sites that maximised prediction of sample genotype (feature selection).

87
88 To address the impact of site co-variation on feature selection, we quantified amino acid co-
89 variation among all pairs of sites in the HBV genome using a Mutual Information (MI) approach
90 as previously applied to *Plasmodium falciparum* sequence data(18). A full description of the
91 methods can be found in the supplementary material.

92

93 **RESULTS**

94 **HBV genotypes can be distinguished through 10 amino acid sites**

95 The machine learning approach discriminated confidently between HBV gt-A-H using just 10
96 amino acid sites (Figure 1). Half of these sites (5/10) were identified in Pol, with four in the
97 spacer region of Pol, and a single site in the reverse transcriptase (RT) domain. A further 4/10
98 sites were located in the Surface protein, particularly in pre-S1 (2/4), and a single site was
99 identified in HBx. The majority of the sites (9/10) were in overlapping regions (the single site
100 in RT being the exception), with the pre-S1/spacer overlap accounting for 6/10 sites. None of
101 the 10 sites identified were in Core protein.

102

103 We classified the amino acid sites based on chemical properties (Figure 1). Properties were
104 generally not conserved across the genotypes at informative sites, with the exception of HBx-
105 40 which was almost always hydrophobic apart from in gt-F/H. This observation suggests that
106 there may be consistent selection pressure to maintain different chemical properties between

107 genotypes, and that the sites are not located in key regions required for host interactions, as
108 this would typically require functional conservation.

109

110 **General location of the top genotype-informative sites**

111 We considered the top 50 most informative sites to determine whether this changed the
112 distribution throughout the genome compared to the top 10 sites. The distribution of these
113 sites within the genome remained comparable. In particular, amino acid 40 in HBx remained
114 the only informative site in HBx, 28/50 (56%) were located in Polymerase, with 16/28 of these
115 sites located in the spacer domain. The Surface protein contained 21/50 sites. The majority of
116 sites were identified in overlapping regions of the genome, with a low number of sites in the
117 TP, RT and RNAse H domains of the Polymerase polyprotein (Table S2, Figure S2).

118

119 **Sites defining genotypes**

120 Although 10 sites were sufficient to discriminate confidently between all genotypes considered,
121 the majority of sites identified were conserved within each genotype, albeit with a few sites
122 presenting variation at the subgenotype level. For example, gt-B sequences could be identified
123 by a single site, 40A in HBx, with all other genotypes having 40P/S (Figures 1, S5). The 988H
124 residue (283H in RT) was also key for identifying gt-A (Figures 1, 2). Other sites were
125 polymorphic within a particular genotype, but genotype-specificity could be distinguished by
126 the *absence* of particular residues (e.g. non-V/G at site 1435 (221 in surface) indicates gt-C)
127 (Figures 1, S6). The close evolutionary history of gt-F and H could be seen by homology at
128 many of the top-10 sites (Figure 1), with site 637 (87 in spacer domain) demonstrating the
129 clearest discrimination between gt-F (637N/Y) and gt-H (637D) (Figure S5). Sites 40 and 570
130 also showed differences in the distribution of amino acids between gt-F and gt-H.

131

132 **Sites defining subgenotypes**

133 A number of the top-10 informative sites were also highly discriminatory for some HBV
134 subgenotypes, including p40-P/S (gt-F), p599-A/T (gt-B), p637-D/G (gt-A) and p659-A/S (gt-
135 D) (Figures 2, S5 and S6). Differences in the amino acids selected at some of the sites in gt-
136 D, in particular p40-S and p1253-R (Figures S5-6), also support the designation of gt-D5 as a
137 unique subtype(9). These sequences cluster distantly from other gt-D sequences on a long
138 branch and show strong geographical clustering, with all sequences isolated from India and
139 Bangladesh.

140

141 **Co-varying sites with top 10 informative sites**

142 When using Random Forests, high co-variation between two predictor variables can result in
143 their importance for classification being shared and thus penalized (relative to other predictor

144 variables). Within our pipeline, this could have resulted in the exclusion of pairs of sites that
145 present high co-variation, or the exclusion of single sites that had high co-variation with the
146 top 10 selected sites. To address these possibilities, we first used Mutual Information theory
147 to quantify co-variation between all pairs of amino acid sites across the genome (see
148 Supplementary Text for full details), and found that the vast majority of site pairs present low
149 co-variation (Figure S7).

150
151 Among the 55 site pairs with the highest co-variation (Table S4), all included at least one site
152 from the top 10 most informative sites that discriminate genotype, thus ruling out the possibility
153 that pairs of sites that discriminate genotype and that present high co-variation were excluded
154 in our Random Forest approach. Of the top 10 sites, only three featured in 55 site pairs with
155 the highest co-variation (HBx site 40, and spacer sites p599 and p637) presenting varying
156 degrees of co-variation with a range of sites across the genome (Table S4, Figure S8). These
157 other sites, although not in the top 10 list of sites that discriminate genotype, could still be of
158 biological interest. For example, we find that HBx 40 is highly co-variable with aa227 in Pol
159 (p596) and a series of amino acids at the start of small-HBs (p1403, p1404, p1406, p1411;
160 corresponding to aa15-23), which also co-varied with site p599 but to a lesser degree. The
161 highest co-variations were found between p599 and both HBx aa39 (p39) and pre-S1 aa97
162 (p1311), which were also found to intermediately co-vary with HBx 40. Reasons for the strong
163 association between this site in HBx, spacer and the start of small-HBs are unclear. The
164 majority of past work on HBx interactions has focused on interactions with host proteins rather
165 than considering influences on other viral proteins(19, 20).

166
167 In comparison, site p637 (aa268 in Pol) had the lowest degrees of co-variation with other sites,
168 but nonetheless presented a varied list of connections, showing associations with two sites in
169 core (aa100 and 123; p254 and 277 respectively) and a cluster of closely-located sites in Pol
170 (p632, p633, p636). This cluster of sites in Pol overlaps a regulatory region in the nucleotide
171 code adjacent to a 'CCAAT' box, known to be the S-promoter region(21).

172

173 **DISCUSSION**

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175 **HBV genotypes can be defined by 10 key sites**

176 Our analysis demonstrates that HBV genetic population structure can be determined from as
177 few as 10 amino acid sites across the viral proteome (Figure 1, Table S3). Four of the top-10
178 sites were identified by previous studies as informative for HBV genotyping (Table S3).
179 Analysing the aa sequences individually by protein has enabled us to determine which
180 residues are key, avoiding difficulties in interpretation that could otherwise arise as a result of

181 the overlapping genome structure. Our analysis further suggests that Core is uninformative
182 for distinguishing HBV genotypes (Table S2). This is in keeping with a high conservation rate
183 of >75% of amino acid sites(22), as expected for a highly structural capsid protein which also
184 plays diverse roles in the viral replication cycle. HBx was also found to be a relatively
185 uninformative region of the HBV genome, with a single site identified in the top-50 most
186 informative sites (Table S2).

187

188 **Informative sites are concentrated in the spacer domain**

189 The spacer domain, which spans aa 184-348, is an intrinsically disordered protein and poorly
190 conserved region of Pol, unique to hepadnaviridae. Previous literature has shown that the
191 spacer domain can tolerate significant deletions and insertions without a significant impact on
192 polymerase function(23, 24).

193

194 The unexpected clustering of sites that predict genotype in the spacer domain indicates that
195 whilst the domain retains a considerable amount of plasticity, this is highly lineage-specific
196 rather than stochastic. Other studies have also found that spacer mutations are relevant in
197 distinguishing between simian hepatitis B viruses(25), as well as human HBV genotypes(9,
198 26, 27) and subgenotypes(28–30). Importantly the four top-10 sites we identified in spacer
199 map to regions previously identified as useful for lineage distinction(24). This suggests
200 selection pressures may be acting to conserve genotype-specific sequence within spacer.
201 Furthermore, it substantiates the hypothesis that spacer plays a central role in the co-evolution
202 of the overlapping P and S genes, potentially related to selection pressure from antiviral drugs,
203 vaccines and the host immune response(24). In addition to encoding regions within proteins,
204 the promoter region for the RNA transcript encoding medium- and small-HBs is present in the
205 pre-S1/spacer overlap region. Mutation of the spacer region is therefore likely to interfere with
206 the generation of M-/S-HBs transcripts, the biological significance of which is unclear(31).
207 Current models are poorly equipped to study this, suggesting our understanding of the role of
208 spacer may be limited by the tools used to analyse its function.

209

210 **Limitations of the methodology**

211 Recombinant sequences, combining two or more different HBV genotypes, were intentionally
212 excluded from this analysis. As such, the short list of 10 sites that can discriminate genotypes
213 is not expected to be able to adequately classify recombinant samples(32). Since our
214 algorithm used the amino acid sequences to compare isolates, synonymous mutations are not
215 considered in the analysis. As several regions of the HBV genome contain promoter regions,
216 such as the well-described basal core promoter of pre-core, synonymous changes in the DNA

217 sequence would have important functional effects. Conservation of the nucleotide sequence
218 in these regions would therefore also be key and may be lineage-specific.

219

220 **CONCLUSIONS**

221 We present the observation that HBV can be reliably genotyped using information from as few
222 as ten sites, and for the distinction of some genotypes by a single site. This is of potential
223 practical importance if a genotype identification is desired but limited sequence data are
224 available. Our finding that discriminatory sites are concentrated in spacer underlines the role
225 and evolutionary importance of the spacer domain in the viral polymerase. With emerging
226 importance of genotypes in HBV disease outcomes, quick approaches to genotyping from
227 short fragments of sequence data may be of increasing practical utility, particularly in low-
228 resource settings. Furthermore, describing the impact of selection pressure at different sites
229 in the genome can provide insights into viral evolution, and potentially contribute to
230 mechanistic insights regarding viral persistence and pathogenesis.

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Table 1: Summary of HBV genes and proteins, and their roles and functions

Gene	Protein(s)	Roles and function
X	X	<ul style="list-style-type: none"> • Small regulatory protein (154aa) • Role in subversion of host restriction factors • Transactivating properties that are implicated in oncogenesis(33).
Core (C)	Pre-core (e-antigen); Core	<ul style="list-style-type: none"> • Post-translational processing to derive capsid protein and e-antigen • Roles in intracellular trafficking and stabilisation of covalently closed circular (ccc)DNA • Soluble e-antigen is secreted into blood, can cross the placenta; acts as an immune tolerogen
Polymerase (Pol)	Polymerase	<ul style="list-style-type: none"> • Four distinct domains: terminal protein (TP); spacer; reverse transcriptase (RT); RNase H. • Takes up approximately two-thirds of the genome(34). • RT and RNase H domains show homology to HIV proteins, and some HIV nucleos(t)ide inhibitors can be used for HBV treatment(34). • TP and spacer domains are unique, and no known homologues have been identified to date(34).
Surface (S)	Short (S) Medium (Pre-S2 + S) Long (Pre-S1 + Pre-S2 + S) surface proteins	<ul style="list-style-type: none"> • External envelope • Receptor binding domains • Surface epitopes neutralised by vaccine-mediated or naturally arising antibodies • Produced in excess, with a potential role as a tolerogen/immunological decoy. • Gene is completely overlapped by polymerase, representing the longest known gene overlap of any animal virus(35).

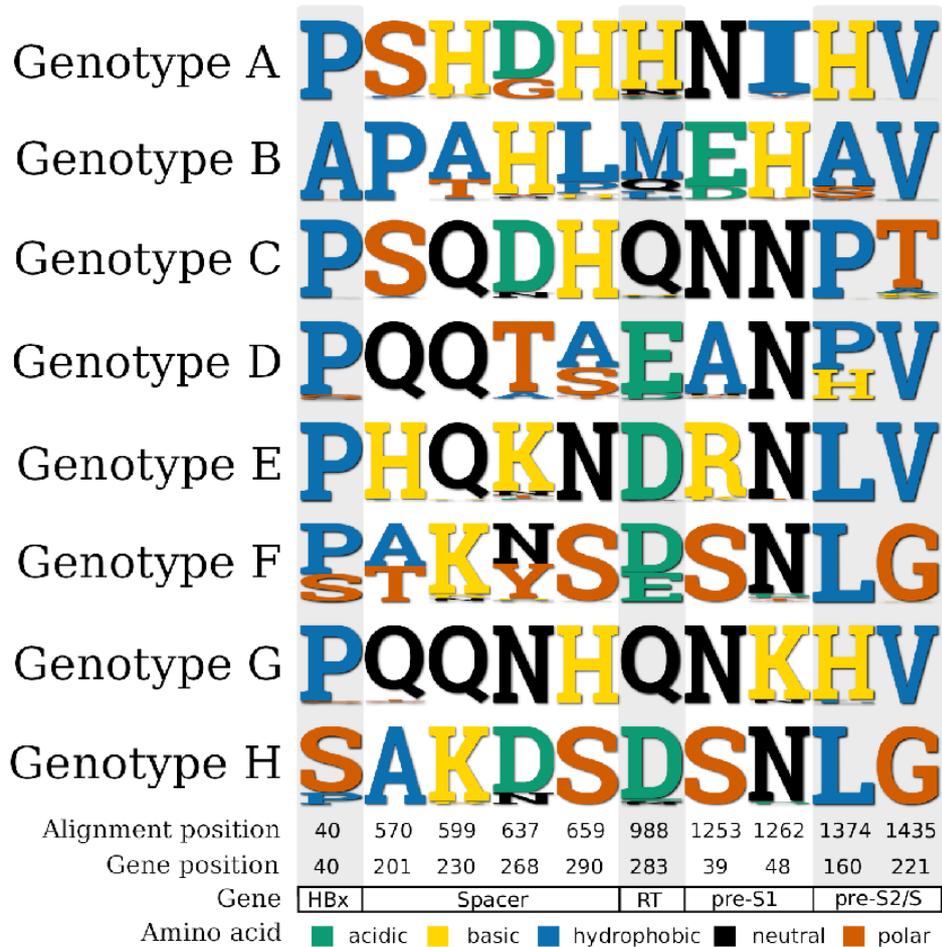


Figure 1: Top 10 amino acid sites discriminating between HBV genotypes, and the residues found at the sites in each genotype. Residues have been coloured according to their properties (key at bottom of the figure). Position of the sites is given in the concatenated amino construct used for analysis (Figure S1) and the equivalent locations in each gene are given at the foot of the figure. In Pol, residues in spacer are given assuming the first amino acid is at the start of the terminal protein, and sites within the reverse transcriptase (RT) domain are counted separately, as is convention in the field (Table S3).

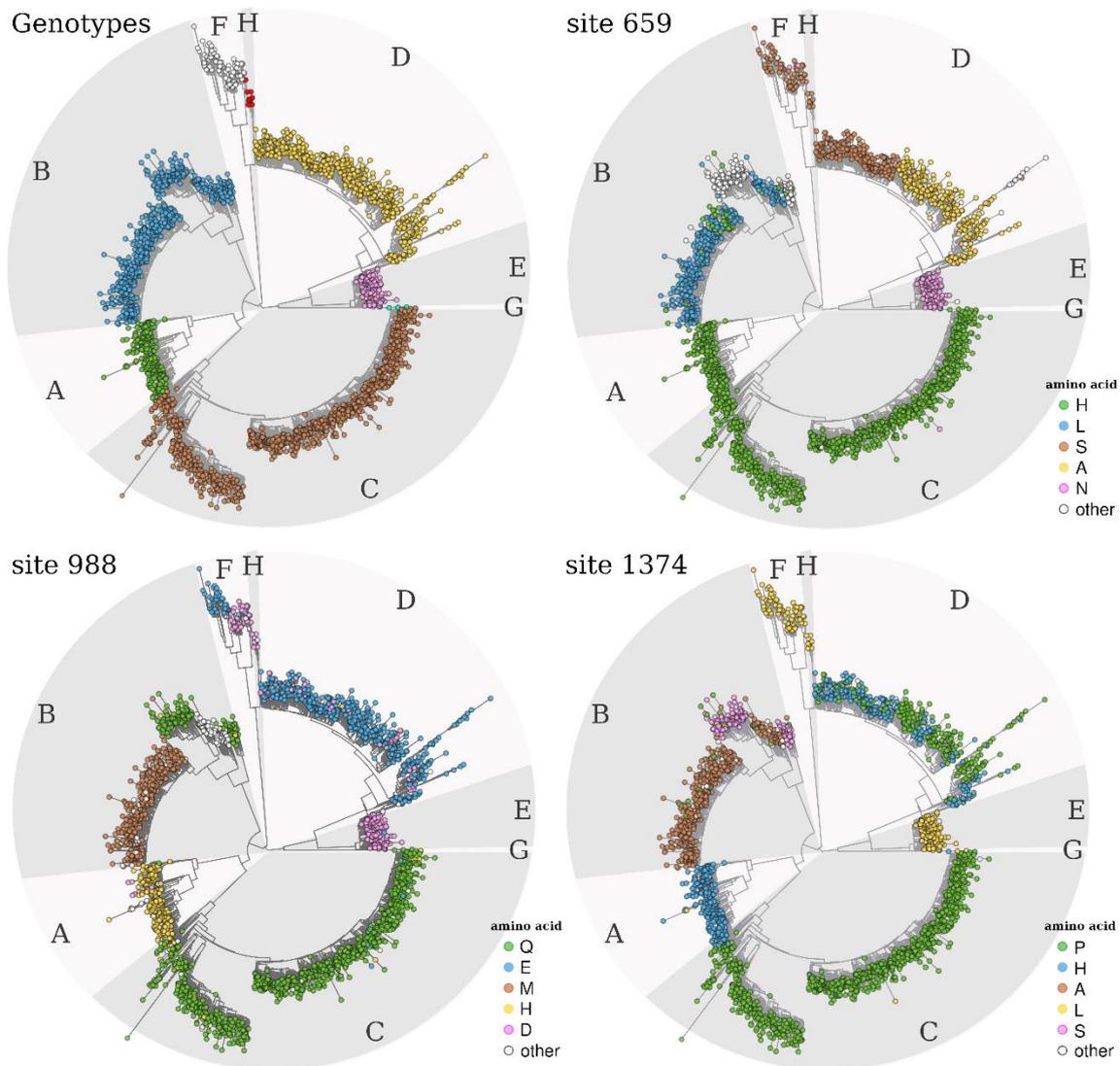


Figure 2: Phylogenetic trees showing overall genotype lineage, and distribution of three exemplar amino acid sites that predict lineage. Maximum likelihood phylogenetic trees were available to download as a part of the online resource from which we obtained nucleotide sequences(9). The top left tree highlights the different HBV genotypes (A to H) with capital letters and with shaded, alternating, lighter and darker grey areas. For the trees of sites 659, 988 and 1374, nodes are coloured on the basis of the amino acid residue at each site (inner legends) using an inhouse R script based on the R package ‘Analyses of Phylogenetics and Evolution’ (ape v5.4 (36)). On each tree, only the top five most frequent amino acids are presented, with the rest under the category “other”. Phylogenies for the other 7 top-10 sites are shown in Supplementary Figures S5 and S6.

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