

Type of article: Research Article (Short Communication)

Title of the article: Translation through collaboration: practice applied in BAMOS project in *in vivo* testing of innovative osteochondral scaffolds

Running title: *In vivo* tests of osteochondral scaffolds in BAMOS

Author names: Ricardo Donate¹, Maryam Tamaddon², Viviana Ribeiro^{3,4}, Mario Monzón^{1,*}, J. Miguel Oliveira^{3,4,*}, Chaozong Liu^{2,*}

Author affiliation:

¹ Departamento de Ingeniería Mecánica, Grupo de Investigación en Fabricación Integrada y Avanzada, Universidad de Las Palmas de Gran Canaria, Campus Universitario de Tafira s/n, 35017 Las Palmas, Spain;

² Division of Surgery & Interventional Science, University College London, Royal National Orthopaedic Hospital, Stanmore HA7 4LP, UK;

³ 3B's Research Group, I3B's-Research Institute on Biomaterials, Biodegradables and Biomimetics, University of Minho, Parque de Ciência e Tecnologia, Zona Industrial da Gandra, AvePark, Barco, Guimarães, 4805-017, Portugal;

⁴ ICVS/3B's-PT Government Associated Laboratory, Braga, Guimarães, Portugal.

***Corresponding author(s):** mario.monzon@ulpgc.es; miguel.oliveira@i3bs.uminho.pt; chaozong.liu@ucl.ac.uk

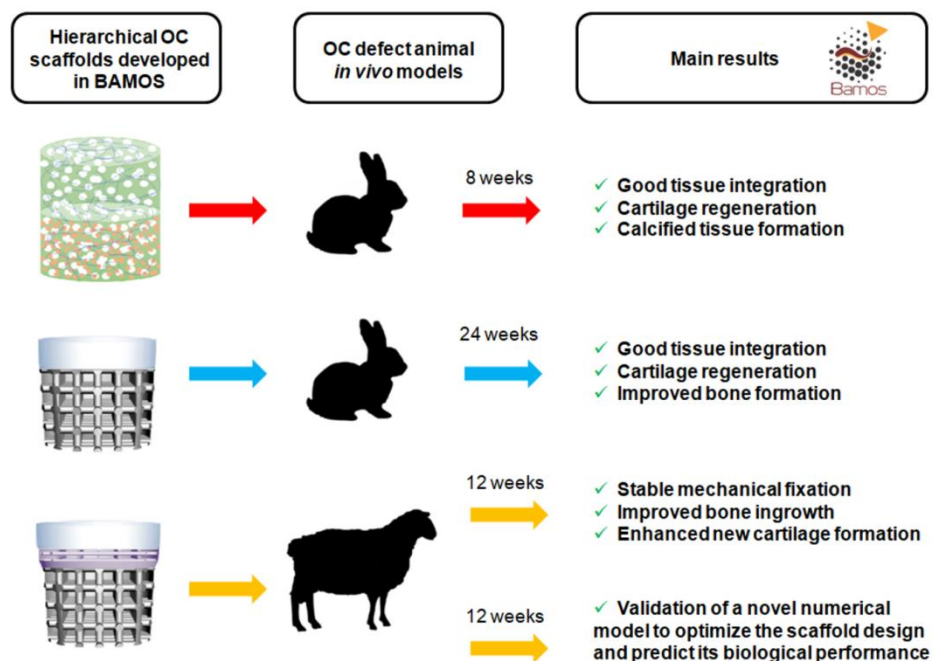
Key Words: Bone; Cartilage; In vivo evaluation; Regenerative Medicine; Tissue Engineering

Abstract:

Osteoarthritis is the most common chronic degenerative joint disease, recognized by the World Health Organization as a public health problem that affects millions of people worldwide. BAMOS project, funded under the frame of the Horizon 2020 RISE program, aims to delay or avoid the use of joint replacements by developing novel cost-effective osteochondral scaffold technology for early intervention of osteoarthritis. This project brings together five internationally leading research organisations (Universidad de Las Palmas de Gran Canaria, Spain; University of Minho, Portugal;

University College London, UK; Xi'an Jiaotong University and Zhejiang University, China) and two healthcare providers (Royal National Orthopaedic Hospital, UK; and Saúde Atlântica - Gestão Hospitalar, S.A., Portugal) to work on the development, manufacturing and marketing of osteochondral scaffolds for the repair of large cartilage damages in osteoarthritis patients. The multidisciplinary consortium of BAMOS collaborates through research and innovation staff exchanges. The project covers all the stages of the development before the clinical trials: design of scaffolds, biomaterials development, processability under additive manufacturing, *in vitro* test, and *in vivo* test. This paper reports the translational practice adopted in the project in *in vivo* assessment of the osteochondral scaffolds developed.

Graphical Abstract: The main results of the *in vivo* evaluations carried out in BAMOS project, funded under Horizon 2020 RISE program, are summarized. Animal models of osteochondral defect have been used to assess the biological performance of the different multimaterial and multilayered scaffolds developed.



Introduction

Osteoarthritis (OA) is mainly characterized by articular cartilage progressive loss, osteophyte formation, synovial membrane inflammation and thickening of the subchondral bone, which leads to the generation of osteochondral defects with limited self-healing capacity. Osteochondral (OC) scaffold is a tissue engineering approach aiming to repair a joint defect and restore its function to delay or eliminate the need for a joint replacement. Although they have been established and are promising for the treatment of small osteochondral defects, no products to date have demonstrated the appropriate biomechanical properties required to promote successful long-lasting regeneration of large osteochondral defects. BAMOS project addresses this challenge in OA treatment [1] by the following main objectives: a) Define clinical specifications for OC scaffolds, b) Develop new OC scaffolds biomaterials, c) Develop innovative Additive Manufacturing (AM) techniques to produced patient-tailored OC scaffolds, d) Assess the OC scaffold in both *in vitro* and *in vivo*, and e) Train early-stage researchers in the context of collaborative research.

Methods

After a complete physicochemical and *in vitro* characterization, the different hierarchical OC scaffolds developed in the context of BAMOS project were evaluated using *in vivo* models. Thus, for example, a rabbit knee critical size OC defect model was used for assessing *in vivo* OC regeneration when implanting a horseradish peroxidase cross-linked silk fibroin-based (HRP-SF) scaffold [2]. These 3D structures were prepared by combining two distinct layers: an HRP-SF cartilage-like layer; and a subchondral bone-like layer composed of HRP-SF and beta-tricalcium phosphate (β -TCP) particles doped with zinc and strontium (HRP-SF/ZnSr- β -TCP) [3].

Also using a rabbit model, bilayered scaffolds were implanted into osteochondral defects created at the distal femoral trochlea and tested for 24 weeks [4]. These scaffolds were composed of a titanium (Ti) matrix that served as bone layer, and a collagen/PLGA layer intended for cartilage regeneration.

Following a different approach, multi-material trilayered OC scaffolds were also developed in BAMOS by combining AM and other conventional technologies [5]:

- Casting and freeze-drying methods were used to obtain a collagen/PLGA composite layer to act as cartilage-like layer.
- Material extrusion of polymers from a heated nozzle (MEX-TRB/P), commonly referred as fused deposition modelling (FDM), was used to manufacture a PLA-based two-part junction layer that served as calcified cartilage of the hierarchical scaffold.
- Powder bed fusion of metal (PBF-LB/M) was used to produce a porous Ti matrix intended for bone regeneration.

The developed OC trilayered scaffold were evaluated *in vivo* using a sheep stifle condyle model for 12 weeks.

Results

After 8 weeks of implantation, the hierarchical (HRP-SF)-based scaffold showed good tissue integration (with no signs of inflammatory reactions), cartilage tissue regeneration and calcified tissue formation [3]. Formation of collagen type II and glycosaminoglycans (GAGs) in the HRP-SF articular cartilage-like layer was confirmed by histological analyses, while *de novo* bone ingrowth and blood vessels infiltration were observed in the HRP-SF/ZnSr- β -TCP bone-like layer [2].

In the case of Ti/collagen-PLGA bilayered scaffolds, we concluded that the mechanical support provided by the Ti layer promoted subchondral bone formation and new tissue integration, which led to better cartilage regeneration.

The 12-week *in vivo* evaluation of the proposed OC trilayered scaffold, carried out in a sheep stifle condyle model, showed a stable mechanical fixation of the 3D structure on the implantation site with no adverse effects observed on the surrounding tissues [6]. Improved bone ingrowth into the Ti matrix, as well as enhanced formation of hyaline-like cartilage tissue, were reported after histological examinations. The up-regulation of the cartilage-related markers aggrecan and collagen type-II confirmed the capacity of the proposed 3D structures to regenerate cartilage tissue.

Discussion

Apart from their good biological response *in vivo*, the OC bilayered (HRP-SF)-based scaffolds have also shown to possess adequate structural integrity, memory-shape properties, and excellent mechanical and *in vitro* biological properties, even preventing bacterial biofilm formation [3], which in sum confirms the potential of these structures to be used in OC Tissue Engineering applications.

On the other hand, the results obtained for the OC trilayered scaffolds also showed the potential of these cell-free scaffolds to be applied in the treatment of large OC defects. Interestingly, the incorporation of bone marrow concentrate (BMC) to the trilayered scaffolds has led to a non significant improvement on bone regeneration when treating OC defects [7]. In this case, an ovine stifle condyle model was used during a 6-month test. Despite obtaining no significantly higher quantity of newly formed bone when using the Ti-PLA-PLGA/collagen scaffold, the results suggested that enhanced bone homogeneity and biomechanical durability were obtained when implanting the trilayered scaffolds (seeded with BMC), thus producing a higher quality of new subchondral bone tissue. Similarly, not statistically significant differences in terms of OC regeneration were obtained between collagen/hydroxyapatite scaffolds with or without BMC when tested *in vivo* using an ovine femoral condyle model for 6 months [8].

Noteworthy, an ovine condyle model was also used to validate a novel numerical model developed in the context of BAMOS project, which is intended to optimize the scaffold design and material properties but also to predict its final biological performance [9]. The simulated cell distribution in the scaffold matched well with the *in vivo* regenerated bone tissue distribution. Therefore, this model could help reduce the number of preliminary time- and cost-consuming *in vivo* and *in vitro* tests needed to optimize the scaffold design.

Author contributions: Mario Monzón: Resources, Writing—Original Draft, Project administration, Funding acquisition. Ricardo Donate: Writing—Original Draft, Visualization. Maryam Tamaddon: Writing—Review and Editing. Viviana Ribeiro: Writing—Review and Editing. Chaozong Liu: Resources, Writing—Review and Editing. J. Miguel Oliveira: Resources, Writing—Review and Editing. All authors approved the final version of this manuscript.

Financial support: The authors would like to thank H2020-MSCA-RISE program, as this work is part of the developments carried out in BAMOS project, funded from the European Union's Horizon 2020 research and innovation programme under grant agreement No. 734156.

Acknowledgement: Not applicable.

Conflicts of interests statement: The authors have no competing interests to declare.

References

1. Monzón M. Biomaterials and additive manufacturing: osteochondral scaffold innovation applied to osteoarthritis (BAMOS project). *J Zhejiang Univ A* 2018;19:329–30. <https://doi.org/10.1631/JZUS.A18NW001>.
2. Ribeiro VP, Pina S, Canadas RF, da Silva Morais A, Vilela C, Vieira S, et al. In Vivo Performance of Hierarchical HRP-Crosslinked Silk Fibroin/ β -TCP Scaffolds for Osteochondral Tissue Regeneration. *Regen Med Front* 2019;1:e190007. <https://doi.org/10.20900/rmf20190007>
3. Ribeiro VP, Pina S, Costa JB, Cengiz IF, García-Fernández L, Fernández-Gutiérrez MDM, et al. Enzymatically Cross-Linked Silk Fibroin-Based Hierarchical Scaffolds for Osteochondral Regeneration. *ACS Appl Mater Interfaces* 2019;11:3781–99. <https://doi.org/10.1021/acsami.8b21259>.
4. Yang T, Tamaddon M, Jiang L, Wang J, Liu Z, Liu Z, et al. Bilayered scaffold with 3D printed stiff subchondral bony compartment to provide constant mechanical support for long-term cartilage regeneration. *J Orthop Transl* 2021;30:112–21. <https://doi.org/10.1016/j.jot.2021.09.001>.
5. Blunn G, Liu C, Tamaddon M. Improved bone and cartilage regeneration in a rapid-manufactured

functionally-graded osteochondral. Hangzhou International Conference on Biomaterials, Bio-Design and Manufacturing (BDMC2018). Aug. 26 - 28, 2018; Hangzhou (China).

https://2020.aconf.org/conf_156776/abstract/77.html

6. Tamaddon M, Blunn G, Tan R, Yang P, Sun X, Chen SM, Luo J, Liu Z, Xu W, Lu X, Donate R, Monzón M, Liu C. In vivo evaluation of additively manufactured multi-layered scaffold for the repair of large osteochondral defects. *Bio-Design Manuf* 2022. <https://doi.org/10.1007/s42242-021-00177-w>
7. Flaherty T, Tamaddon M, Liu C. Micro-computed tomography analysis of subchondral bone regeneration using osteochondral scaffolds in an ovine condyle model. *Appl Sci* 2021;11:1–14. <https://doi.org/10.3390/app11030891>.
8. Tamaddon M, Blunn G, Xu W, Alemán Domínguez ME, Monzón M, Donaldson J, et al. Sheep condyle model evaluation of bone marrow cell concentrate combined with a scaffold for repair of large osteochondral defects. *Bone Joint Res* 2021;10:677–89. <https://doi.org/10.1302/2046-3758.1010.BJR-2020-0504.R1>.
9. Liu Z, Tamaddon M, Chen SM, Wang H, San Cheong V, Gang F, et al. Determination of an Initial Stage of the Bone Tissue Ingrowth Into Titanium Matrix by Cell Adhesion Model. *Front Bioeng Biotechnol* 2021;9:1–13. <https://doi.org/10.3389/fbioe.2021.7360>