

Expert Opinion on Biological Therapy



ISSN: (Print) (Online) Journal homepage: https://www.tandfonline.com/loi/iebt20

Onasemnogene abeparvovec for the treatment of spinal muscular atrophy

Hugh J. McMillan, Crystal M. Proud, Michelle A. Farrar, Ian E. Alexander, Francesco Muntoni & Laurent Servais

To cite this article: Hugh J. McMillan, Crystal M. Proud, Michelle A. Farrar, Ian E. Alexander, Francesco Muntoni & Laurent Servais (2022): Onasemnogene abeparvovec for the treatment of spinal muscular atrophy, Expert Opinion on Biological Therapy, DOI: 10.1080/14712598.2022.2066471

To link to this article: https://doi.org/10.1080/14712598.2022.2066471

<u>a</u>	© 2022 The Author(s). Published by Informa UK Limited, trading as Taylor & Francis Group.
	Published online: 02 May 2022.
	Submit your article to this journal 🗗
hil	Article views: 941
Q	View related articles 🗷
CrossMark	View Crossmark data ☑

Taylor & Francis Taylor & Francis Group

DRUG EVALUATION

OPEN ACCESS Check for updates



Onasemnogene abeparvovec for the treatment of spinal muscular atrophy

Hugh J. McMillan 6a, Crystal M. Proudb, Michelle A. Farrarcd, Ian E. Alexanderef, Francesco Muntonigh and Laurent Servais (D)

^aDepartments of Pediatrics, Neurology & Neurosurgery, Montreal Children's Hospital, McGill University Health Centre, Montreal, Canada; ^bChildren's Hospital of the King's Daughters, Norfolk, VA, United States; 'School of Clinical Medicine, UNSW Medicine and Health, UNSW Sydney, Sydney, NSW, Australia; Department of Neurology, Sydney Children's Hospital Network, Sydney, NSW, Australia; Gene Therapy Research Unit, Children's Medical Research Institute, Faculty of Medicine and Health, The University of Sydney and Sydney Children's Hospitals Network, Westmead, NSW, Australia; Discipline of Child and Adolescent Health, Sydney Medical School, Faculty of Medicine and Health, The University of Sydney, Westmead, NSW, Australia; ⁹The Dubowitz Neuromuscular Centre, University College London, Great Ormond Street Institute of Child Health London, UK; hNIHR Great Ormond Street Hospital Biomedical Research Centre, Great Ormond Street Institute of Child Health, University College London, & Great Ormond Street Hospital Trust, London, UK; Department of Pediatrics, Centre Hospitalier Universitaire de Liège & Université de Liège, Liège, Belgium; MDUK Oxford Neuromuscular Centre, University of Oxford, Oxford, UK

ABSTRACT

Introduction: Gene therapy for spinal muscular atrophy (SMA) represents a significant milestone in the treatment of neurologic diseases. SMA is a neurodegenerative disease that results in motor neuron loss because of mutations of the survival motor neuron 1 gene, which directs survival motor neuron (SMN) protein production. Onasemnogene abeparvovec, a one-time gene replacement therapy, delivers a functional transgene to restore SMN protein expression. Onasemnogene abeparvovec has demonstrated improved survival and motor milestone achievements for presymptomatic infants and patients with SMA type 1.

Areas covered: This expert review describes the current state of gene therapy for SMA, reviews the mechanism of and clinical experience with onasemnogene abeparvovec, explains future efforts to expand applications of gene therapy for SMA, and provides context for developing gene therapy for other conditions.

Expert opinion: Onasemnogene abeparvovec has demonstrated efficacy in clinical trials and, because of this, is a valuable treatment option for patients with symptomatic infantile SMA and those identified by newborn screening. Gene therapy is still in its infancy, and challenges and uncertainties associated with transgene delivery must be addressed. With ongoing development of vector technology, more specific tissue tropism, reduced 'off-target' effects, and an enhanced safety profile will continue to evolve.

ARTICLE HISTORY

Received 12 January 2022 Accepted 12 April 2022

KEYWORDS

Adeno-associated viral vector; disease-modifying treatments; gene therapy; motor milestones; newborn screening; neurodegenerative disorders; onasemnogene abeparvovec; spinal muscular atrophy; survival motor neuron; vector genomes

1. Introduction

Gene therapy has opened the door to treatments for an increasing number of rare genetic diseases. New technologies and approaches to diagnosis and care allow clinicians, patients, and caregivers to anticipate novel, lifechanging therapies for diseases that, until now, have received only supportive care. Spinal muscular atrophy (SMA) represents one such disease and its approved gene therapy, onasemnogene abeparvovec, has changed the treatment landscape of this devastating neurologic disease.

SMA is a neurodegenerative disease that results in motor neuron loss because of biallelic mutations of the survival motor neuron 1 (SMN1) gene, leading to severe muscle weakness and atrophy. SMN1 directs the production of the survival motor neuron (SMN) protein, which is essential for the development and maintenance of motor neurons [1,2]. SMA occurs in an estimated 1 in 10,000 live births and was, until recent advancements in treatment, a leading genetic cause of infant mortality [3-5]. The survival motor neuron 2 (SMN2) gene functions as a partial backup gene to SMN1, and the severity of SMA correlates inversely with the polymorphic number of SMN2 gene copies [6–8]. However, quantification of SMN2 copies, and the clinical significance of this quantification, still requires standardization [9].

Traditionally, SMA phenotypes have been described according to age at symptom onset and maximum motor milestones achieved (Table 1) [4,10,11]. The wide phenotypic spectrum of SMA ranges from profound weakness evident at birth (type 0) to relatively mild symptoms for individuals with adult onset (type 4). Type 1 is the most common and is a very severe phenotype of SMA, with symptoms usually appearing during the first few months of life. Without treatment, patients with SMA type 1 never achieve independent sitting and usually do not survive beyond 2 years of age.

Until relatively recently, most studies of SMA focused on symptom management [12-14]. The advent of an antisense oligonucleotide (nusinersen) and a small-molecule drug (risdiplam) that affect SMN2 gene splicing, as well as a gene

which permits non-commercial re-use, distribution, and reproduction in any medium, provided the original work is properly cited, and is not altered, transformed, or built upon in any way.



Article highlights

- Spinal muscular atrophy (SMA) is a neurodegenerative disease that results in motor neuron loss because of biallelic mutations of the survival motor neuron 1 (SMN1) gene, leading to inadequate concentrations of survival motor neuron (SMN) protein, which is critical for motor neuron development and maintenance.
- SMA type 1 is the most severe and most common phenotype of SMA, with symptom manifestation within the first months of life. Without treatment, patients with SMA type 1 are not predicted to survive beyond 2 years of age.
- Onasemnogene abeparvovec is a one-time, intravenously administered gene replacement therapy approved for the treatment of SMA.
- Onasemnogene abeparvovec uses an adeno-associated viral (AAV) vector that offers long-term transgene expression and has low immunogenicity compared with other viral vectors and transgenes.
- Onasemnogene abeparvovec demonstrated clinical benefit in the Phase I START and Phase III STR1VE-US and STR1VE-EU trials, which included infants with SMA type 1, most of whom were aged <6 months. Overall, patients demonstrated longer survival, achievement of motor milestones, and improved motor function after administration of onasemnogene abeparvovec. Several clinical and real-world studies and disease registries are planned or underway that will evaluate onasemnogene abeparvovec in a broader SMA patient population, as well as clinical durability and long-term safety of onasemnogene abeparvovec.
- SMA is associated with enormous medical and societal costs. The availability of gene therapy has the potential to decrease direct and indirect costs, but the considerable cost of the gene therapy and the difference between results achieved when treatment is administered before symptom onset compared with after symptom onset must be considered in the balance of costs.
- New and emerging evidence will continue to shape the way onasemnogene abeparvovec is prescribed in routine clinical practice. The lessons learned from early interventions and new treatment paradigms in SMA will influence how future screening and treatment programs are implemented for other neurologic conditions.

replacement therapy (onasemnogene abeparvovec) that delivers a functional gene to restore expression of full-length SMN protein, has changed the model of SMA treatment [15]. Multidisciplinary care in combination with disease-modifying therapies remains imperative, necessitating an approach focused on each individual patient's clinical status and current needs, to optimize motor, respiratory, and bulbar function. Although these treatments do not offer a cure for SMA, they do offer substantial gains in motor function achievements and life expectancy. With the introduction of disease-modifying treatments, SMA phenotypes are evolving, and classification is increasingly described according to functional status (e.g. non-sitter, sitter, walker) [12,13]. Age, SMN2 copy number, and baseline motor function are important determinants of outcomes [16,17].

Several consensus statements and documents to guide SMA treatment considerations are available [18-20], including recommendations specific for the use of onasemnogene abeparvovec [18,21]. These algorithms guide clinicians to provide the best possible support and improve outcomes for patients with SMA and, overall, underscore the importance of early treatment to moderate symptom development and progression. Specifically, because of the rapidly progressive nature of the disease, universal newborn screening (NBS) and early SMA treatment are critical to ameliorate irreversible motor neuron loss and maximize functional outcomes [22-28]. Prior to the availability of disease-modifying treatments for SMA, NBS for SMA was not widely accepted or implemented [29]. However, the availability of treatments that can be potentially administered before symptom onset has led to substantial gains in public and health care provider acceptance of NBS [22]. In addition, the rapid expansion of NBS for SMA will likely further impact disease classification, which will include infants treated presymptomatically stratified by SMN2 copy number.

Regional differences in access to care impact treatment of patients with SMA. Disparities among countries have been reported in genetic testing and diagnosis; support such as cough assistance and ventilation, gastric and nasogastric tubes, and scoliosis surgery; loss of ambulation; and health-related quality of life and survival. These differences are attributed to dissimilarities in health care systems (public vs. private); financial limitations or constraints; availability (or lack) of specialty care; social or cultural attitudes toward chronic, life-limiting diseases [10]; and differences in the approvals of disease-modifying treatments by regulatory bodies around the world [22,30]. Treatment algorithms and clinical guidelines continue to evolve as new evidence emerges, more knowledge is gained, and clinician and patient/caregiver acceptance for SMA treatment grows.

The availability of gene therapy for SMA provides a new paradigm both for the treatment of SMA — moving from supportive care to interventional care — and for broader treatment of neurologic disorders using gene therapy. This expert review provides an overview of the current state of gene therapy for SMA, explains future efforts to expand applications of gene therapy for this condition, and provides context for developing gene therapy for other disease states.

2. Methods

We searched PubMed to explore the breadth of literature on AAV9based gene therapy and its applications for SMA. We conducted two searches to capture literature released since publication of the pivotal Phase I START trial of onasemnogene abeparvovec (formerly AVXS-101) in 2017 through 1 November 2021. For the first search, terms included onasemnogene abeparvovec AND spinal muscular atrophy. For the second search, terms included adenoassociated virus serotype-9 OR AAV9 AND gene therapy. We also searched the reference lists of publications in the PubMed results to identify new and emerging research. Congress presentations, abstracts, and posters were also obtained by searching relevant congress web sites (including the American Academy of Neurology, the Academy of Managed Care Pharmacy, the British Paediatric Neurology Association, the European Paediatric Neurology Society, and SMA Europe). All publications were considered for this review, including those from industry, academia, and clinical practice, to ensure that an objective, independent review was conducted.

Table 1. Types of spinal muscular atrophy [4,10,11].

SMA type	0	1	2	3	4
Approximate percentage of all SMA cases	<1%	45–60%	~20–30%	~15–20%	<1%
Typical SMN2 copies present	1 copy	2–3 copies	3 copies	3–4 copies	≥4 copies
Age at symptom onset	Prenatal	0–6 months	6–18 months	>18 months	20–30 years
Symptoms and typical features	Profound hypotonia Facial muscle weakness Inability to suck/swallow Respiratory failure at birth	Limb weakness Respiratory distress Weak cry Poor feeding Inability to sit unsupported Bell-shaped chest	May sit unsupported Poor crawling Limb weakness more profound in lower than upper limbs Some limitation of head and neck control Inability to stand or walk independently Impaired swallowing Ventilatory insufficiency Scoliosis	May stand or be able to walk independently Unable to run, jump, or climb Abnormal gait, foot deformity Able to eat independently	Ambulatory Mild, slowly progressive limb weakness
Support needed	Respiratory support beginning at birth	Respiratory and nutritional support by 5–6 months of age	Ventilatory support, feeding tube, wheelchair	Wheelchair or braces, physical and occupational therapy	Walking aids
Survival without disease-modifying treatment	Weeks	<2 years	Reduced life expectancy	Does not impact life expectancy	Does not impact life expectancy

SMA: spinal muscular atrophy; SMN2: survival motor neuron 2 gene.

3. Onasemnogene abeparvovec: overview and therapeutic mechanism

The preferred vector for a gene therapy targeting the central nervous system (CNS) is derived from adeno-associated virus (AAV), a dependent parvovirus. Recombinant AAV vectors are suitable for gene therapy because of their capacity to transduce both dividing and non-dividing cells and confer longterm transgene expression in non-dividing cells, primarily as a non-integrating episome [31-34]. In addition, AAV-based therapies have relatively low immunogenicity compared with other viral vectors, such as adenovirus. The importance of a patient's immune response to a viral vector was first demonstrated by a patient with ornithine transcarbamylase deficiency who died in 1999 after suffering complications from an adenovirus-mediated gene transfer [35]. This case highlighted the need to consider other less immunogenic viral vectors for gene therapy applications targeting genetic disease.

As the transgene is primarily maintained in an extrachromosomal episome that resides in the nucleus [36], the risk of insertional mutagenesis is less than that observed with other viral vectors, such as those derived from retroviruses in which integration is an obligate feature of the viral life cycle. However, while predominantly episomal, a small percentage of AAV vector genomes (or fragments thereof) do undergo genomic integration, and a large portion of the vector dose ends up in the liver and carries a small risk of insertional mutagenesis, which has been highlighted in several preclinical studies, but, to date, not in humans [37].

Recombinant AAVs have the same basic structure as wildtype AAVs, with an outer shell, or capsid, enclosing a singlestranded DNA genome, but the recombinant genome does not contain any AAV protein-coding sequences. Instead, the recombinant genome contains a therapeutic gene expression cassette [38,39] and retains only the flanking inverted terminal repeats, which are of viral origin and required for genome replication and packaging [39,40]. The removal of all viral coding sequences allows for a lesser risk of immunogenicity and cytotoxicity [39]. Another instrumental property of the AAV vector system is the ability to cross-package the recombinant viral genome into multiple different capsid types depending on the desired gene transfer properties conferred by the capsid. For example, the critical and unique property of the AAV9 capsid used in onasemnogene abeparvovec is the ability to cross the blood-brain barrier and transduce neurons.

One barrier to AAV administration is pre-existing anticapsid humoral immunity driven by natural exposure to wild-type AAV, particularly AAV2, which is endemic though nonpathogenic — in human populations and can lead to the production of antibodies that cross-react with the AAV9 capsid [41]. The prevalence of AAV-neutralizing antibodies is moderate at birth, decreases between 7 and 11 months of age, and progressively increases through childhood and adolescence. Overall, approximately 40-80% of adult humans have antibodies against AAV [42]. Patients who have a greater seroprevalence of anti-AAV antibodies may not be suitable candidates for AAV-based gene therapy [41-44]. Similarly, AAV vector exposure induces an immune response to the AAV capsid that currently precludes retreatment. Transgenes (or their protein products) can also stimulate immune responses, but such responses are unlikely for patients with SMA treated with onasemnogene abeparvovec because some SMN protein is produced by all patients as a result of paralogous SMN2 genes. Systemic corticosteroids are administered before and for a period of a few months after gene therapy to attenuate the immune response, providing anti-inflammatory and immunosuppressive effects [41].

The AAV serotype used in onasemnogene abeparvovec, AAV9, was isolated from the human liver, which is one of two predominant sites of natural AAV infection (the other is the spleen) [39]. AAV9 differs from other serotypes because it crosses the blood-brain barrier, which allows for intravenous delivery to achieve widespread gene expression in the CNS. Thus, AAV9 is a promising therapeutic tool for CNS and neurologic disorders [45] and has already demonstrated success in treating SMA [46–50].

Onasemnogene abeparvovec is composed of an AAV9 capsid that carries a recombinant AAV genome encoding a therapeutic cassette that is flanked by AAV2 inverted terminal repeats [51]. One of the inverted terminal repeats carries a specific deletion that allows the formation of a self-complementary genome configuration that facilitates rapid expression of the transgene [51]. Sustained, ubiquitous SMN transgene expression from the therapeutic cassette is driven by a hybrid cytomegalovirus enhancer-chicken β-actin promoter.

Following intravenous administration, onasemnogene abeparvovec is widely distributed in the CNS and peripheral tissues. The AAV9 mechanism of targeting motor neurons after systemic delivery differs from most other forms of AAV gene therapy, which are not able to efficiently cross the blood-brain barrier and target the CNS [52].

In a recent study of the biodistribution of onasemnogene abeparvovec in two symptomatic infants with SMA type 1 who died because of SMA-related complications, transduction was observed in multiple body systems [36]. Transgene copy numbers ranged from 0.04 vector genomes (vg)/cell in the thymus to 399.25 vg/cell in the liver. Distribution to skeletal muscle was also observed at 1.1-4.0 vg/cell in the diaphragm, 1.4 vg/cell in the quadriceps, 1.1-1.3 vg/cell in the psoas, and 2.5-3.3 vg/cell in the intercostal muscles. Distribution in spinal motor neurons ranged from 1.49-2.65 vg/cell. The degree of transduction in the liver was expected and consistent with studies of onasemnogene abeparvovec administration [53,54]. This AAV9 hepatotropism has important safety implications and illustrates the need for long-term follow-up. The increased delivery of vector genomes to hepatocytes could be associated with a greater theoretical risk of insertional mutagenesis and a greater potential lifetime risk of liver neoplasia.

SMN protein expression was also reported in the CNS and peripheral organs, including the brain (cortical and subcortical regions), choroid plexus, vascular structures, quadriceps, heart, liver, kidney, lungs, pancreas, spleen, thymus, stomach, large and small intestine, and inguinal lymph nodes [36]. SMN expression was greatest in the liver, with high expression also found in the heart, psoas, and diaphragm muscles.

The recommended dose of onasemnogene abeparvovec is 1.1×10^{14} vg per kilogram of body weight (vg/kg). Onasemnogene abeparvovec is administered as an intravenous infusion over 60 minutes. Treatment with systemic corticosteroids equivalent to oral prednisolone 1 mg/kg/day must be initiated 1 day before onasemnogene abeparvovec administration and continued for 30 days and then weaned over an additional 4 weeks [55]. If adverse events such as serum transaminase concentration elevations occur, increasing or continuing prednisolone dosing for a longer treatment period may be necessary [56].

4. Clinical experience with onasemnogene abeparvovec

The efficacy of onasemnogene abeparvovec in SMA type 1 has been established in clinical trials, and real-world clinical experience with onasemnogene abeparvovec is expanding rapidly, facilitated by SMA disease registries. As more patients receive onasemnogene abeparvovec, our knowledge of therapeutic effects, the overall risk of adverse events, and applications to larger patient populations is expanding.

4.1. Phase I trial of onasemnogene abeparvovec

The first clinical trial to demonstrate the benefit of onasemnogene abeparvovec, START (NCT02122952) [48], was a Phase I trial conducted in 2014 and 2015 at Nationwide Children's Hospital in Columbus, Ohio. START included 15 patients with SMA type 1, all of whom had two copies of SMN2. All patients received a single dose of onasemnogene abeparvovec. Three patients received a low dose (6.7×10¹³ vg/kg) and 12 received a high dose $(2.0\times10^{14} \text{ vg/kg})$ [equivalent to the therapeutic dose of 1.1×10^{14} vg/kg according to the potency assay]). At the time of treatment, the ages of patients in the low-dose group ranged from 5.9 to 7.2 months (mean, 6.3 months), and the ages in the high-dose group ranged from 0.9 to 7.9 months (mean, 3.4 months). Prior to onasemnogene abeparvovec treatment, three patients in the lowdose group and five patients in the high-dose group required nutritional support, and three patients in the low-dose group and two patients in the high-dose group required ventilatory support. Safety, time until death or the need for permanent ventilatory assistance, and changes in scores on the Children's Hospital of Philadelphia Infant Test of Neuromuscular Disorders (CHOP INTEND) scale of motor function were evaluated. The CHOP INTEND scale ranges from 0 to 64, with greater scores indicating better function. Historically, children with SMA type 1 are not predicted to achieve scores greater than 40 [57-59]. Endpoints were compared with a cohort of untreated patients from natural history studies [48].

Overall, treatment with onasemnogene abeparvovec resulted in longer survival, achievement of motor milestones, and improved motor function, especially for patients treated early and patients who had greater baseline function treated with the greater dose. At 20 months after administration, all 15 patients were alive and free from ventilatory support (a historic comparison cohort reported only 8% survival at the same time point). Of the 12 patients who received the high dose of onasemnogene abeparvovec, 11 sat unassisted, nine rolled over, 11 fed orally and could speak, and two walked independently. The high-dose group also achieved rapid and early increases in CHOP INTEND scores of 9.8 points from baseline at 1 month and 15.4 points at 3 months. CHOP INTEND scores declined in the historic cohort [48].

At the last follow-up 24 months after administration, the mean CHOP INTEND score in the high-dose group was 56.5, compared with 5.3 in the historic comparison cohort. All 12 infants in the high-dose START group achieved improvements in CHOP INTEND of \geq 4 points, and 11 of 12 achieved a score \geq 40.0. In addition, 11 patients sat unassisted for \geq 5 seconds, 10 for

≥10 seconds, and nine for ≥30 seconds at 24 months of follow-up. Two patients could stand and walk independently. Importantly, these two patients were treated very early and had a baseline CHOP INTEND score of 50 [60]. This is far greater than the CHOP INTEND score threshold of 40 that patients with SMA type 1 typically do not reach without treatment, which once again demonstrates the importance of early intervention for optimal outcomes.

During the 24 months of follow-up, 53 serious adverse events were reported among 10 patients. Two of the events were related to the study treatment and involved asymptomatic transaminase elevations [60].

START was the first study to demonstrate that a single dose of an AAV containing DNA coding for the SMN protein could offer clinical benefit with a favorable safety profile for patients with SMA type 1, and the long-term follow-up of the START study (LT-002; NCT04042025) supports the long-term safety and durable efficacy of onasemnogene abeparvovec [49]. The ongoing follow-up safety study includes 13 patients enrolled in START (three from the low-dose cohort and 10 from the highdose cohort). More than 6.2 years after administration, all 10 patients in the high-dose cohort were alive and free from ventilatory support at the time of follow-up. All motor milestones achieved in START have been maintained, and two patients have achieved new milestones during long-term follow-up. Eight patients experienced serious adverse events, including acute respiratory failure, pneumonia, dehydration, respiratory distress, and bronchiolitis, but none led to study discontinuation or death, and none were deemed to be related to the study drug. This study provides evidence of the clinical durability and favorable safety of onasemnogene abeparvovec, although durability is difficult to ascertain in all patients because four patients in the high-dose cohort and three patients in the low-dose cohort also received nusinersen [49].

4.2. Phase III trials with onasemnogene abeparvovec

The open-label, single-arm, single-dose Phase III STR1VE trials built on the success of START by studying the safety and efficacy of the therapeutic dose of onasemnogene abeparvovec (a single intravenous infusion of 1.1×10¹⁴ vg/kg). Endpoints were again compared with patients from natural history data sets [47,50].

4.2.1. STR1VE

The first of two STR1VE trials was conducted at 12 hospitals in the United States from 2017 to 2019. STR1VE-US (NCT03306277) included 22 patients with SMA type 1 and two copies of *SMN2*. The co-primary endpoints were independent sitting for 30 seconds or longer (Bayley-III Scales of Infant and Toddler Development item #26) at 18 months of age and survival (absence of death or permanent ventilation). Secondary outcomes included ability to thrive (a composite endpoint including swallowing function, nutritional support requirements, and weight maintenance) and being free from ventilatory support at 18 months of age.

The patients' ages ranged from 0.5–5.9 months (mean, 3.7 months). The mean CHOP INTEND score at baseline was 32.0. At 14 months of age, 20 patients were alive and free from ventilatory support (compared with six patients in the natural history cohort). At 18 months of age, 13 patients achieved sitting for 30 seconds or longer (compared with zero patients in the natural history cohort). Nine patients maintained the ability to thrive and 18 were free from ventilatory support (compared with zero for either endpoint in the natural history cohort). CHOP INTEND scores increased rapidly, with mean increases from baseline reported as early as 1 month after administration. Mean increases in scores improved by 6.9 points at 1 month, 11.7 points at 3 months, and 14.6 points at 6 months [47].

All patients experienced at least one adverse event, and pyrexia was the most common. Three serious adverse events were possibly treatment-related, with two patients experiencing hepatic transaminase elevations and one patient experiencing hydrocephalus. Overall, STR1VE-US demonstrated clinically meaningful benefit for survival, motor milestone achievement, and motor function, and further supported the use of onasemnogene abeparvovec in symptomatic infants with SMA type 1 [47].

The second Phase III STR1VE trial was conducted at nine hospitals and universities in Italy, the United Kingdom, Belgium, and France from 2018 to 2020. Similar to STR1VE-US, STR1VE-EU (NCT03461289) included patients younger than 6 months of age with SMA type 1 and two copies of SMN2 [50]. Compared with STR1VE-US, STR1VE-EU comprised a broader patient population that included some patients who were receiving nutritional and/or respiratory support at baseline. Patients who required non-invasive ventilatory support for at least 6 hours per day or feeding support were excluded from STR1VE-US but were eligible for STR1VE-EU. The primary endpoint was independent sitting for at least 10 seconds at any time up to and including 18 months of age, defined by the World Health Organization (WHO) Multicentre Growth Reference Study. The secondary endpoint was ventilation-free survival at 14 months of age. Ability to thrive (defined as weight greater than the third percentile, free of nutritional support, and normal swallowing function with thin or very thin liquids at 18 months of age) was assessed as an exploratory outcome.

A total of 33 patients were included in STR1VE-EU. The patients' ages ranged from 1.8 to 6.0 months (mean, 4.1 months), and the mean CHOP INTEND score at baseline was 27.9. Nine patients reported feeding support and nine reported ventilatory support at baseline. Five patients were receiving both feeding and ventilatory support. At 14 months of age, 31 patients were alive and free from ventilatory support. At 18 months of age, 14 patients achieved independent sitting for 10 seconds. Seven patients met the ability to thrive criteria. Patients achieved rapid and sustained increases in CHOP INTEND scores, with mean increases in scores of 6.0 points from baseline at 1 month, 10.3 points at 3 months, and 13.6 points at 6 months [50].

Thirty-two patients experienced at least one adverse event, and the most common was, again, pyrexia. Six serious drug-related adverse events were reported, including two cases of pyrexia, three cases of increased hepatic transaminase concentrations, and one case of gastroenteritis. Overall, onasemnogene abeparvovec demonstrated efficacy even for patients with more severe disease than those included in studies to date, and no new safety signals were identified [50].

4.2.2. SPR1NT

SPR1NT (NCT03505099) was an open-label, single-arm, Phase III study evaluating the use of onasemnogene abeparvovec in presymptomatic infants younger than 6 weeks of age at risk of SMA type 1 with two or three copies of SMN2 [61]. In all, 29 patients were enrolled in SPR1NT. Of these, 14 patients had two copies of SMN2 and 15 patients had three copies of SMN2. Early results demonstrated that both patient groups (two and three copies of SMN2) survived and reached age-appropriate milestones. All patients in both groups were alive and free from permanent ventilation at the end of study for two- and three-copy patients [61–63]. At 18 months of age, all 14 patients in the two-copy group achieved the primary efficacy endpoint of sitting without support for at least 30 seconds, including 11 who achieved this milestone within the WHO normal developmental window [61,63]. All patients in the two-copy group achieved CHOP INTEND scores ≥58 during at least one follow-up visit [61]. All 15 patients in the three-copy group achieved the primary efficacy endpoint of standing without support, confirmed by independent video review. Fourteen of 15 did so within the WHO normal developmental window (≤514 days) [64]. Fourteen of 15 threecopy patients walked independently for at least five steps at any visit up to 24 months of age and 11 of 14 achieved this motor milestone within the WHO normal developmental window (≤534 days) [64]. No infant enrolled in SPR1NT experienced a serious adverse event that was considered treatment-related by the investigator, and no new safety signals were identified [61,62]. Overall, onasemnogene abeparvovec demonstrated clinical efficacy for children younger than 6 weeks of age with presymptomatic SMA.

4.3. SMA registries

Several registries of patients with SMA have been established to gather real-world data about the long-term safety and efficacy of SMA treatments.

4.3.1. The RESTORE registry

The RESTORE registry (NCT04174157) is a prospective, multicenter, multinational observational registry evaluating SMA history and treatment; pulmonary, nutritional, and motor milestones; health care resource utilization; work productivity; activity impairment; adverse events; health-related quality of life; caregiver burden; and survival [65–67]. Patients with genetically confirmed SMA are being enrolled during a period of 5 years, which began in September 2018. These patients will be followed for 15 years or until death [68].

Presentations at the 2022 American Academy of Neurology Annual Meeting reported interim results of RESTORE. Preliminary analyses as of 23 November 2021 (the most recent data cut) were conducted to evaluate outcomes for patients identified by NBS compared with clinical diagnosis of SMA and the use of onasemnogene

abeparvovec for patients with SMA aged at least 6 months at infusion.

To compare NBS with clinical diagnosis, patients in the RESTORE registry from the United States with ≤ 2 copies of *SMN2* were stratified according to either clinical diagnosis of SMA type 1 based on SMA symptoms or prenatal screening/ NBS. Each group was then limited to patients who had a follow-up period of at least 16 months. In all, 25 were identified as at risk for SMA type 1 based on prenatal screening or NBS and 70 were identified via clinical diagnosis. Patients identified via prenatal/NBS were diagnosed and received disease-modifying treatment significantly earlier than clinically diagnosed patients (0.8 vs. 3.7 months and 1.6 vs. 5.1 months, respectively [p < 0.0001 for both]) [66].

Patients identified via prenatal/NBS generally achieved motor milestones at earlier ages than clinically diagnosed patients. The median age for achieving independent sitting for 30 seconds was 13.7 months in the prenatal/NBS group and 21.8 months in the clinically diagnosed group. Four patients (two in each group) achieved the milestone of walking independently. The patients in the prenatal/NBS group achieved this milestone at 24.7 months, and the patients in the clinically diagnosed group achieved it at 32.2 months. Most patients in both groups achieved increases of ≥4 points on CHOP INTEND (100% in the prenatal/NBS group and 80.1% in the clinically diagnosed group). Monthly increases in CHOP INTEND scores were similar at 1.2 points in the prenatal/NBS group and 1.0 in the clinically diagnosed group. Clinically diagnosed patients received more than one SMA treatment more often than patients identified via prenatal/NBS (70.0% vs. 60.0% [p=0.3730]) [66].

To analyze the use of onasemnogene abeparvovec in a population older than those patients included in clinical trials, patients who received onasemnogene abeparvovec when they were at least 6 months of age were considered [67]. In all, 145 patients were identified from the RESTORE registry. Of these, 67 received on a semnogene abeparvovec between 6 and 12 months of age, 67 between 12 and 24 months, and 11 at age 24 months or older. Ninety-six patients were diagnosed with SMA type 1, and 11 children were presymptomatic. Forty-six patients had available CHOP INTEND scores, including 19 patients who received on a semnogene abeparvovec between 6 and 12 months of age and who achieved a score increase of ≥4 points and 14 patients who received onasemnogene abeparvovec between 12 and 24 months of age and achieved the same increased score. The adverse event profile among these patients was consistent with the overall adverse event reporting for onasemnogene abeparvovec, and no new safety signals were observed [67]. Together, these early findings from RESTORE enhance the understanding of how onasemnogene abeparvovec is used in the real world and demonstrates clinical benefit for presymptomatic children and patients older than 6 months.

4.3.2. Canadian Neuromuscular Disease Registry

The Canadian Neuromuscular Disease Registry (CNDR), established in 2011, is a longitudinal, prospective, observational study of patients with neuromuscular disease in Canada [69]. A subset of this registry focuses on SMA and will evaluate the real-world safety and effectiveness of novel therapies. As of

March 2020, 250 patients were enrolled across 37 clinics [70]. One recent abstract from the CNDR reported the patterns of switching among disease-modifying therapies for SMA. Of 217 patients in the CNDR with available data, 44 had SMA type 1. Of these, seven patients received nusinersen and then switched to onasemnogene abeparvovec [71]. Long-term outcomes and follow-up data are forthcoming.

4.3.3. SMArtCARE Registry

The SMArtCARE registry collects long-term, real-world data on the treatment of SMA in Germany, Austria, and Switzerland [72]. More than half of a subset of 76 patients were older and heavier at the time of onasemnogene abeparvovec administration (mean age, 16.8 months; mean weight, 9.1 kg) than patients in clinical trials. Fifty-eight patients had been treated with nusinersen. Of 60 patients with available data, 49 achieved improvements in CHOP INTEND scores of ≥4 points. CHOP INTEND scores increased significantly for patients younger than 24 months at the time of onasemnogene abeparvovec administration but not for those who were older than 24 months. CHOP INTEND scores also increased significantly for the 45 patients with available data who had received nusinersen. Notably, respiratory and bulbar function improved for patients with respiratory insufficiency or dysphagia despite nusinersen treatment [56].

Fifty-six patients experienced treatment-related adverse events, including eight serious adverse events. Overall, liver enzyme elevations were greater for patients who were older and heavier at the time of onasemnogene abeparvovec infusion [56]. These findings strengthen support for careful patient selection and comprehensive monitoring for potential liver dysfunction for older patients treated with onasemnogene abeparvovec.

Overall, this report adds to the evidence of a positive benefit–risk ratio of onasemnogene abeparvovec treatment for patients with SMA and extends the findings to older and heavier patients as well as to those treated with nusinersen. However, differentiating benefits related to onasemnogene abeparvovec or combination effects compared with the same potential results with nusinersen monotherapy over a longer observation period is difficult. Future research is needed to clarify these results.

4.4. Adverse events

Onasemnogene abeparvovec has been associated with specific adverse events, which can be managed with close anticipatory surveillance. Although some adverse events may be related to total viral vector dose (and, thus, body weight), further research is needed to clarify risk factors for specific adverse effects [73,74]. Patient selection is important in preventing and/or mitigating these events. Ensuring that the patient has had no recent as well as no active infection and evaluating for any signs of pre-existing liver dysfunction is necessary to optimize safety and determine eligibility for treatment.

4.4.1. Hepatotoxicity

Hepatotoxicity has been observed following onasemnogene abeparvovec administration [53]. Hepatotoxicity is especially

concerning with high vector-dose systemic AAV regimens and deserves expanded attention. A recent study reported evidence of hepatotoxicity in 100 patients with SMA who had received onasemnogene abeparvovec [53]. The mean age of the patients at the time of onasemnogene abeparvovec administration was 2.9 months. Ninety patients had elevated serum transaminase concentrations (either alanine aminotransferase [ALT] or aspartate aminotransferase [AST]) and/or bilirubin. The elevations began as early as Week 1 and peaked approximately 1 month after administration. Thirty-four patients had at least one hepatotoxic adverse event. Sixtyone (61%) patients, including children with SMA who were presymptomatic, had elevated serum transaminase concentrations prior to onasemnogene abeparvovec administration. This reflects the early findings of the SPR1NT trial, in which 23 of 30 patients tested had elevated bilirubin and/or transaminase concentrations prior to administration of onasemnogene abeparvovec. Liver function for these patients normalized by the end of the study [53].

The early elevation in transaminases is sufficient to cause liver dysfunction that impacts the biosynthetic functions of the liver for approximately 2% of patients, evidenced by increased international normalized ratios [53]. However, the mechanism of the early increases in hepatic transaminases, while still unclear, is likely different from mechanisms involved in later increases. One report highlighted more severe hepatic dysfunction for two patients that occurred between 3 and 8 weeks after onasemnogene abeparvovec. The patients experienced increased ALT, AST, and bilirubin, which resolved with highdose corticosteroid and supportive therapy, and both patients demonstrated clinical recovery. One patient demonstrated evidence of hepatic fibrosis on liver biopsy [75]. Further research is needed to better understand the mechanism and consequences of liver function test elevations and hepatic dysfunction.

Hepatic cytolysis observed for patients treated with onasemnogene abeparvovec is different than cholestasis observed in another neuromuscular condition, X-linked myotubular myopathy (XLMTM), treated by gene therapy. Three male patients in the ASPIRO trial received 3×10¹⁴ vg/ kg of the AAV-based gene therapy and developed cholestasis and liver failure within 3 to 4 weeks after administration. Two patients experienced fatal liver dysfunction and the third was reported to have ongoing liver dysfunction. Compared with other patients who received AAV-based gene therapies, these three patients were older, heavier, and displayed evidence of pre-existing liver disease [76,77]. Increasing evidence suggests that hepatobiliary disease is a common comorbidity with XLMTM, which may present treatment challenges for children with this disorder [78], and indeed a fourth child recently developed fatal hepatic complication despite receiving a lower AAV8 viral load $(1.3\times10^{14} \text{ vg/kg})$ [79].

Unlike gene therapy with onasemnogene abeparvovec, gene therapy administered for XLMTM uses an AAV8 capsid. In addition, subclinical cholestasis has been established for patients who died following gene therapy in the XLMTM trial. The hepatotoxicity of onasemnogene abeparvovec is

evidenced by elevated AST and ALT, which demonstrates cytolysis. The hepatotoxicity in XLMTM patients was evidenced by elevated bilirubin, which demonstrates cholestasis and liver failure. In addition, the immune systems of patients with XLMTM are naïve to the missing endogenous protein, which is not the case in SMA, in which patients produce residual SMN protein. This difference could contribute to adverse events [80]. Furthermore, hepatotoxicity was not observed in murine or canine models of AAV8-based treatment in XLMTM [81,82], which highlights the importance of careful and well-conducted studies in human populations. However, these outcomes reinforce the importance of understanding the biodistribution of AAV-based gene therapy and the consequences that greater concentrations of distribution to the liver may confer.

4.4.2. Hematologic abnormalities

Thrombocytopenia and thrombotic microangiopathy (TMA) were reported after onasemnogene abeparvovec administration [73,83–85]. In clinical trials and post-marketing safety analyses, thrombocytopenia was reported as transient in most cases and often resolved without intervention [84]. In a study of eight patients who received nusinersen followed by onasemnogene abeparvovec, six experienced thrombocytopenia. The lowest blood counts were observed between Days 6 and 8 after onasemnogene abeparvovec administration. No evidence of cutaneous, mucosal, or other bleeding was observed, and blood counts normalized within a few weeks after treatment [73]. Thrombocytopenia is a feature of TMA and should be monitored following onasemnogene abeparvovec administration [83].

No cases of TMA were reported in clinical studies [84], but as of July 2020, three cases of TMA following onasemnogene abeparvovec administration had been reported. Two were reported in the RESTORE registry, and one was reported by a managed access program in Australia. Two of these cases occurred in patients who had received nusinersen. TMA developed within 1 week of onasemnogene abeparvovec infusion. All three infants recovered, one with supportive care and two with therapies such as plasmapheresis, corticosteroids, and/or blood transfusions. TMA is thought to be an immune-mediated reaction or a dose-related toxicity. Concurrent or recent infections are often related to the development of TMA, but more investigation is required to confirm risk factors for TMA [83].

4.4.3. Animal data

Dorsal root ganglia (DRG) inflammation has been observed in nonhuman primates that received a different AAV9-based gene therapy product via the intrathecal route. However, these findings were not observed in mice [84].

Clinical evaluations of human patients treated with onasemnogene abeparvovec (intrathecal or intravenous) have not demonstrated evidence of sensory symptoms consistent with DRG inflammation, such as sensory neuronopathy [84]. However, DRG damage has been reported following intrathecal administration of an AAV-based gene therapy for amyotrophic lateral sclerosis targeting defects in the *SOD1* gene in one patient [86,87]. Together, these

observations suggest that DRG findings may be a class effect of AAV therapies, though the mechanisms of damage are not defined. Electrophysiologic tests may be considered as part of onasemnogene abeparvovec monitoring, because signs and symptoms of sensory nerve abnormalities are difficult to assess for younger patients [84].

SMN overexpression associated with neuro-inflammation and innate immune response was demonstrated in a murine model, with SMN Δ 7 mice that received an AAV9-SMN viral vector. Long-term overexpression of SMN led to motor dysfunction and neurodegeneration [88]. To date, however, no evidence of SMN overexpression or clinical signs of such overexpression in humans has been presented [88,89]. The clinical significance of these animal studies is yet to be fully determined.

4.4.4. Adverse events with other gene therapies

Other AAV-based gene therapies are associated with adverse events that have not been observed with onasemnogene abeparvovec [90]. For example, complement activation has been observed following AAV-based therapies for Duchenne muscular dystrophy (DMD), a genetic disease that results in progressive muscle wasting [91,92]. This immune-mediated reaction promotes inflammation and damage to cells such as red blood cells, platelets, and endothelial cells. This cell damage can further activate the complement system and establish a feedback loop of severe hematologic injury, organ damage, and bleeding, which, if untreated, may be fatal [91]. In a Phase Ib trial of six patients, fordadistrogene movaparvovec (an AAV9-based gene therapy for DMD) was associated with atypical hemolytic uremic syndrome-like complement activation in one patient. This patient suffered acute kidney injury, hemolysis, and decreased platelet count, but symptoms resolved and renal function returned to normal within 15 days because of prompt treatment with intermittent hemodialysis and administration of a complement inhibitor [46]. This prompted changes to the study protocol, with closer monitoring and greater doses of glucocorticoids post infusion. The death of a non-ambulant young man from cardiogenic shock was reported in December 2021 and led to a hold on the clinical development of fordadistrogene movaparvovec [93].

The immunogenicity of AAV-based therapy requires continued investigation. The design and monitoring of future therapies should consider the potential for target cell destruction by T cells directed against non-self epitopes encoded by therapeutic transgenes [94]. Consequently, some DMD gene therapy clinical trials have excluded specific dystrophin mutations to minimize the number of sequence differences between the defective self-gene and the therapeutic transgene, as well as potential T-cell immunity [95]. A musclespecific promoter may minimize the risk of immune-related toxicity in non-muscle tissues [91,92].

5. Health economics

SMA is associated with substantial medical and societal costs [23]. Costs include inpatient, outpatient, and emergency care; prescription and over-the-counter medications; medical

devices and mobility aids such as wheelchairs, walkers, orthotics, and feeding products; respiratory and ventilatory assiscopayments; and home and transportation modifications. Indirect costs related to SMA include loss of productivity (by patient or caregiver); time spent caregiving; caregiver strain such as sleep problems, injury related to moving the patient in their care, anxiety, and changes to employment status; and premature death [96-98]. Quantifying the total costs of SMA is difficult, but one review reported that the average annual cost of SMA type 1 ranges from US\$75,047 to US\$196,429 [23]. Another review estimated the cost of SMA type 1 in the United States to be even greater, with annual expenses of US\$324,410 [96].

The availability of disease-modifying treatments for SMA substantially reduces direct and indirect expenses, but the costs of treatments carry their own concerns regarding benefit and cost effectiveness. Long-term benefits and future savings should be compared with out-of-pocket expenses to evaluate costs [99]. While some treatments require ongoing use, gene therapies, such as onasemnogene abeparvovec, can potentially provide a lifetime of benefit with a single treatment, although this is very difficult to ascertain today. Onasemnogene abeparvovec has a list price of US\$2.125 million, and the estimated lifetime cost for patients with SMA type 1 treated with onasemnogene abeparvovec is approximately US\$4.0 million [98,100]. However, therapies for other chronic illnesses may cost much more over patients' lifetimes. For example, bypassing agents for hemophilia A may exceed US\$15-18 million and lifetime cost of care for these patients may exceed US\$103 million [99].

Because disease-modifying treatments have been available for only a few years, considerable uncertainty persists regarding the true lifetime costs of SMA care in the era of disease-modifying treatments. Gene therapies have challenged models for conventional drug pricing and concerns for health budgets are increasing as new therapies are anticipated. The increased survival of patients who would have died and the substantial upfront costs of gene therapies may impose a strain on the pricing and economics of drug development [23]. As such, innovative approaches to costs and reimbursement models are being considered, including payover-time and pay-for-performance models, manufacturing changes [101,102], and expanded risk pools [103].

Several factors should be considered when assessing the long-term societal value compared with the price of gene therapy, including life expectancy (and, therefore, future costs unrelated to the disease), caregiver burden (including lost productivity and time spent providing care), healthrelated quality of life, and budget impact for health care payors [104,105]. Health equity and overall family well-being are also important [106,107], and one analysis proposed that hope and knowledge are substantial components of the value of gene therapies [99]. As we continue to learn more about the long-term impact that onasemnogene abeparvovec and other gene therapies will have for patients and caregivers, financial decisions will need to be made with input from health care providers, policymakers, and insurance providers, as well as patients and caregivers [103]. New models of care may further enhance health outcomes and cost effectiveness for gene therapy technologies. Specifically, NBS combined

with gene therapy will likely improve health outcomes and cost effectiveness [108].

6. Future directions and forthcoming data

Disease-modifying treatments, including onasemnogene abeparvovec, have demonstrated dramatic benefits with respect to survival and motor function for patients with SMA. However, these novel treatments have been available for only a few years and, as such, we are not yet able to describe the benefits of these therapies over patients' expected lifetimes. To date, published clinical trials of onasemnogene abeparvovec have included infants with clinically diagnosed SMA type 1 and two SMN2 gene copies, or presymptomatic children with two or three SMN2 copies. More research in a greater range of SMA patient types, reflecting real-world use scenarios and the broader indications of onasemnogene abeparvovec, is needed. Human gene therapy is still a growing field, and ongoing research and development will almost certainly define the use of this technology in future clinical practice.

6.1. Onasemnogene abeparvovec in expanded patient populations

Clinical trials of onasemnogene abeparvovec have included only young infants who weighed 8.5 kg or less, but future and ongoing studies will assess intravenous onasemnogene abeparvovec for 'heavier' patients (≥8.5 kg to ≤21 kg [Phase IIIb SMART trial; NCT04851873]) [109]. An intrathecal formulation was assessed for patients aged 6 to 50 months (Phase I STRONG; NCT03381729) [110,111] and will be assessed for patients with SMA type 2 and two to four copies of *SMN2* (Phase III STEER trial; NCT05089656) [112] (Table 2).

6.2. Use of multiple disease-modifying treatments

Clarification regarding treatment with multiple disease-modifying treatments for SMA and switching between disease-modifying treatments is needed [113,114]. Currently no evidence of additive benefit for combination or sequential therapy has been reported, but this is an active area of investigation [115,116].

RESPOND (NCT04488133) is a single-arm Phase IV study that will evaluate the safety and efficacy of nusinersen for patients aged 2 to 36 months who have already received onasemnogene abeparvovec. RESPOND is expected to enroll 60 patients who, in the opinion of the investigator, may achieve additional benefit from nusinersen after first-line treatment with onasemnogene abeparvovec. This study has an expected completion date of September 2024 [117].

6.3. Multidisciplinary care

Ongoing multidisciplinary care is critical for patients with SMA. Disease-modifying treatments greatly improve the prognosis for the disease, but these treatments are not curative, and care is still required over the patient's lifetime. Multidisciplinary care is tailored to the clinical and



functional status of the patient to address ongoing pulmonary, orthopedic, nutritional, and neuromuscular complications resulting from the underlying disease, and the care required will likely change during the patient's lifetime [12,13,118].

New treatments suggest an evolving paradigm for supporting patients with SMA. With the evolution of phenotypes, an approach focused on clinical status and current needs is necessary to optimize outcomes. A patient's baseline motor function and age at initiation of therapy are important parameters to consider when communicating with families regarding informed decision-making and establishing expectations for potential treatment options.

6.4. Vector technology

Vector technology may also evolve and change gene therapy for SMA. Specifically, the development and use of capsids with reduced hepatotropism, and changes to vector design, particularly for promoter-enhancer specificity, could improve the theoretical safety profile [119]. In addition, vectors that are more efficient than the AAV9 capsid at crossing the bloodbrain barrier could improve systemic administration of gene therapies for CNS diseases [120-122].

Recent work in barcoding capsid variants (i.e. linking a specific nucleotide sequence to each capsid for visualization and improved efficiency of transduction [123]) resulted in improved muscle targeting with AAV-based gene therapy products. An AAV9 mutant was directed to target skeletal muscle, including the heart and diaphragm, and demonstrated superior efficiency and specificity compared with non-barcoded vectors [124]. A similar study of muscle-targeting AAV9 capsids allowed for administration of doses that were up to 250 times less than those used in preclinical and clinical trials [125]. However, the peptide used for improved efficiency of transduction would potentially impose immunologic concerns, and barcoding technologies do not necessarily lead to avoidance of AAV pre-existing immunity. In addition, the importance of the difference between mouse models used for barcoding work and humans is yet to be determined. Results obtained with specific AAV capsids in mice likely do not reliably predict performance in equivalent human tissues and cell types, but the development of predictive preclinical models of the blood-brain barrier and human CNS remained challenging. Still, this work will likely help optimize AAV vectors for gene therapy and may have applications in gene therapies that target the CNS.

7. Conclusions

The therapeutic landscape for SMA has changed dramatically over the past few years. Although most cases were essentially untreatable and fatal within the first years of life, diseasemodifying treatments, including onasemnogene abeparvovec, are improving survival and permitting many patients to thrive. Prognoses for most patients with SMA are greatly improved, and the importance of ongoing multidisciplinary care remains undiminished. While onasemnogene abeparvovec for SMA represents a significant milestone in human gene therapy, this field is still in its infancy, and challenges and uncertainties,

such as patient and disease selection, need to be clarified. The safety and efficacy of gene therapy may be affected by many factors, including patient age, weight, and disease severity, as well as delivery mechanisms and targets of the gene therapy vector.

8. Expert opinion

8.1. Prioritizing early diagnosis and intervention

Onasemnogene abeparvovec is the only one-time diseasemodifying treatment for SMA. Outcomes in presymptomatic children highlight the need for universal NBS, and NBS needs to be linked with best practice. Today, only approximately 3% of the world population is screened at birth for SMA [22]. NBS has demonstrated that approximately 30% of infants with two copies of SMN2 have symptoms around the time of diagnosis and exhibit early, rapid decline in motor nerve function [126]. Facilitating timely access to therapy is critical to optimize outcomes. A multidisciplinary team and strong partnerships among stakeholders are key to implementation of NBS and subsequent health care [127].

Onasemnogene abeparvovec represents an attractive option for parents of presymptomatic children identified by NBS. The possibility of further treatment in case of transgene loss of expression should be considered for inclusion in therapeutic options and health economic models. Further investigation is necessary to determine if these approaches are safe and beneficial.

8.2. Overcoming barriers to gene therapy administration

The implementation and integration of gene therapy into health practice requires establishment of appropriate infrastructure, timely delivery, and multidisciplinary medical management. Delays in administration of onasemnogene abeparvovec, such as confirmation of diagnosis, testing of AAV9 serology, navigation of coverage issues, and drug shipment, must be minimized.

Barriers to administration of onasemnogene abeparvovec that deserve attention include facility-specific challenges such as discomfort with or being ill-equipped to facilitate administration of gene therapy, reimbursement decisions, and patient/family willingness to accept gene therapy. Specifically, the rare occurrence of a serious adverse effect is likely more difficult for families to accept in presymptomatic children. More research is needed to identify potential predispositions to adverse reactions and options for minimizing or mitigating these risks. In addition, a better understanding of individual serious adverse reactions, as the mechanisms of adverse events related to gene therapy are not fully understood, is needed.

8.3. Enhancing AAV9 technology

AAV9 therapy is an evolving technology. Manufacturing bottlenecks for the research and development of new gene therapies also present a barrier to innovation. In the future, we anticipate that gene therapies will become safer and more efficient and

Table 2. Future and ongoing clinical trials of onasemnogene abeparvoyec.

	SMART (NCT04851873) [106]	STRONG (NCT03381729) [107,108]	STEER (NCT05089656) [109]
Purpose	To assess safety and tolerability over 12 months after administration of intravenous onasemnogene abeparvovec	To evaluate safety and tolerability of an intrathecal formulation of onasemnogene abeparvovec	To evaluate efficacy, safety, and tolerability of an intrathecal formulation of onasemnogene abeparvovec
Population	Patients who weigh ≥8.5 kg to ≤21 kg	Patients aged 6–50 months with three copies of SMN2	Patients with SMA type 2 and two to four copies of SMN2 who are aged 2–18 years
Estimated enrollment	24 patients	32 patients enrolled as of December 2019 interim analysis	125 patients
Design	Phase IIIb, open-label, single-arm multicenter study Motor-milestone achievement and function will be used to assess efficacy	Phase I, open-label, dose-comparison study The trial assessed three doses of onasemnogene abeparvovec administered via one-time intraspinal injection (Dose A: 6.0×10^{13} ; Dose B: 1.2×10^{14} ; Dose C: 2.4×10^{14} vg).	Phase III, multicenter, randomized, sham-controlled, double-blind study
Preliminary results/ notable findings to date		The US Food and Drug Administration halted the trial in October 2019 for safety concerns prompted by an animal study of intrathecal onasemnogene abeparvovec. The hold was lifted in August 2021. As of an interim analysis in December 2019, patients achieved clinically meaningful motor milestones in all dose groups and age groups. No deaths were reported, and no new safety signals were identified.	
Expected completion	August 2023	No longer recruiting	October 2024

SMA: spinal muscular atrophy; SMN2: survival motor neuron 2 gene.

use less expensive manufacturing technology through advances in AAV manufacturing, as well as advances in other gene transfer and editing technologies. Specifically, future AAV technologies will likely exhibit decreased hepatotropism, which will enhance efficiency and potentially reduce adverse events, ultimately improving future outcomes and successes.

In addition, AAV technology will potentially expand to deliver CRISPR/Cas9 nucleases [126,127]. This approach has shown promise in preclinical trials by demonstrating greater concentrations of AAV integration with high target specificity [128–130] and no evidence of genome-wide genotoxicity [128]. Importantly, a report of CRISPR/Cpf1-mediated correction of patient-specific induced pluripotent stem cells from a patient with SMA demonstrated successful expression of SMN. This provides support for future investigations into therapeutic approaches using CRISPR technology for patients with SMA [131132]

To date, CRISPR/Cas systems have been investigated in animal and cell models of human diseases, including cancer; cardiovascular, pulmonary, and metabolic diseases; hemophilia; and monogenic diseases such as muscular dystrophy [133–136]. CRISPR/Cas systems have also been combined with induced pluripotent stem cells to investigate cell replacement therapy and precision medicine for human diseases. Further, CRISPR has been applied to diagnostic testing, demonstrating high sensitivity and specificity in a fast and inexpensive system [134].

Currently, CRISPR technology is being investigated in several early stage clinical trials. Studies are investigating its use in the treatment of viral diseases, including human papillomavirus-related cervical neoplasia, refractory viral keratitis, human immunodeficiency virus, and coronavirus disease; solid tumors, including esophageal cancer, T- and B-cell malignancies, gastrointestinal malignancies, renal carcinoma, and tumors of the central nervous system; blood disorders, including leukemia and lymphoma, multiple myeloma, sickle cell disease, and β -thalassemia; and rare genetic disorders, including Kabuki Syndrome [137]. CRISPR technology has the opportunity to revolutionize treatment for countless patients and though its use in humans is limited, early results are promising.

8.4. Implementing personalized care for SMA

The discovery of biomarkers to identify individual capacity for response will improve the use of onasemnogene abeparvovec and allow targeted treatment to more specific populations and assessment of meaningful prognostic endpoints. Several genetic, epigenetic, proteomic, electrophysiologic, and imaging biomarkers have been considered for SMA, but their reproducibility and applicability should be confirmed [138]. Once identified and confirmed, these biomarkers will allow for a personalized approach to SMA treatment. Similarly, identifying factors that predispose patients to TMA or severe hepatotoxicity could help to appropriately select patients for gene therapy treatment rather than treatment by another disease-modifying approach.



Drug name (generic)	Onasemnogene abeparvovec
Phase (for indication under discussion)	Approved in the United States in 2019
Indication (specific to discussion)	Onasemnogene abeparvovec is an adeno- associated virus vector-based gene therapy indicated for the treatment of pediatric patients less than 2 years of age with spinal muscular atrophy (SMA) with biallelic mutations in the survival motor neuron 1 gene.
Pharmacology description/ mechanism of action	Onasemnogene abeparvovec is a recombinant AAV9-based gene therapy designed to deliver a copy of the gene encoding the human SMN protein. SMA is caused by a biallelic mutation in the SMN1 gene, which results in insufficient SMN protein expression.
Chemical structure	Not available
Pivotal trial(s)	START, STR1VE-US, STR1VE-EU

8.5. Clarifying the role of combination or sequential treatments

The goal of providing treatment to patients with SMA has shifted to optimizing their function and health-related quality of life to ensure that they 'thrive' instead of only 'survive.' This has provoked discussion about a potential role for combination or sequential treatments, including combination SMNdirected therapies or the addition of non-SMN directed therapies such as an anti-myostatin therapy [138]. Clinical trials will be important to answer this question because no evidence that combination or sequential therapy offers meaningful clinical benefit for patients with SMA currently exists. We predict that the future of SMA treatment may involve using complementary mechanisms of action or different targets along the motor unit [36].

8.6. Establishing the durability of clinical benefit of gene therapy

The duration of gene expression is still largely unknown, and it is likely that evaluation of onasemnogene abeparvovec administration in presymptomatic children will help describe median gene expression duration. The stability of episomes in postmitotic cells (e.g., motor neurons) is unclear. While no evidence of phenomena that could affect long-term expression (e.g., promoter/enhancer methylation) exists, longer follow-up studies are needed.

Acknowledgments

Medical writing assistance and editorial support was provided by Jennifer Gibson, PharmD, from Kay Square Scientific, Newtown Square, PA. This support was funded by Novartis Gene Therapies, Inc.

Declaration of interest

HJ McMillan has received honoraria for scientific advisory boards from Novartis Gene Therapies, Inc., and has received research support from Roche. CM Proud is a site principal investigator for Biogen and Novartis Gene Therapies, Inc., clinical trials, and has received honoraria for advisory board participation from Biogen, Novartis, and Roche, and speaker's fees from Biogen and Novartis. MA Farrar has received honoraria for scientific

advisory boards from Novartis Gene Therapies, Inc., Biogen, and Roche, and has received research grants from Biogen. IE Alexander is an inventor on patent applications filed by Children's Medical Research Institute related to AAV capsid sequences and in vivo function of novel AAV variants, is cofounder of Exigen Biotherapeutics, and has consulted on technologies and has stock and/or equities in companies and technology broadly addressed in this paper. F Muntoni has received honoraria for scientific advisory boards from Novartis Gene Therapies, Inc., Biogen, Novartis, PTC, Roche, and Sarepta and reports grants and personal fees from Novartis Gene Therapies, Inc., Biogen, and Roche. L Servais has received personal compensation as an advisory committee board member/consultant from Novartis Gene Therapies, Inc., Biogen, Biophytis, Cytokinetics, Dynacure, Roche, Santhera, and Sarepta Therapeutics; and has received research support from Novartis Gene Therapies, Biogen, Dynacure, and Roche. The authors have no other relevant affiliations or financial involvement with any organization or entity with a financial interest in or financial conflict with the subject matter or materials discussed in the manuscript apart from those disclosed

Reviewer disclosures

A reviewer on this manuscript has disclosed that they attended the clinical trials entitled 'Safety and Efficacy of Intravenous OAV101 (AVXS-101) in Pediatric Patients With Spinal Muscular Atrophy (SMA) (SMART)(NCT04851873)' and 'Efficacy and Safety of Intrathecal OAV101 (AVXS-101) in Pediatric Patients With Type 2 Spinal Muscular Atrophy (SMA) (STEER) (NCT05089656).' Peer reviewers on this manuscript have no other relevant financial relationships or otherwise to disclose.

Funding

This paper was funded by Novartis Gene Therapies, Inc.

ORCID

Hugh J. McMillan http://orcid.org/0000-0001-8927-2018 Laurent Servais http://orcid.org/0000-0001-9270-4061

References

Papers of special note have been highlighted as either of interest (•) or of considerable interest (••) to readers.

- 1. Lefebvre S, Burglen L, Reboullet S, et al. Identification and characterization of a spinal muscular atrophy-determining gene. Cell. 1995;80(1):155-165.
- 2. Wirth B. Spinal muscular atrophy: in the challenge lies a solution. Trends Neurosci. 2021;44(4):306-322.
- 3. Verhaart IEC, Robertson A, Wilson IJ, et al. Prevalence, incidence and carrier frequency of 5q-linked spinal muscular atrophy a literature review. Orphanet J Rare Dis. 2017;12(1):124.
- 4. Lally C, Jones C, Farwell W, et al. Indirect estimation of the prevalence of spinal muscular atrophy type I, II, and III in the United States. Orphanet J Rare Dis. 2017;12(1):175.
- 5. Strunk A, Abbes A, Stuitje AR, et al. Validation of a fast, robust, inexpensive, two-tiered neonatal screening test algorithm on dried blood spots for spinal muscular atrophy. Int J Neonatal Screen. 2019;5(2):21.
- 6. Burghes AHM, Beattie CE. Spinal muscular atrophy: why do low levels of survival motor neuron protein make motor neurons sick? Nat Rev Neurosci. 2009;10(8):597-609.
- 7. Calucho M, Bernal S, Alías L, et al. Correlation between SMA type and SMN2 copy number revisited: an analysis of 625 unrelated Spanish patients and a compilation of 2834 reported cases. Neuromuscul Disord. 2018;28(3):208-215.



- Tiziano FD, Bertini E, Messina S, et al. The Hammersmith functional score correlates with the SMN2 copy number: a multicentric study. Neuromuscul Disord. 2007;17(5):400–403.
- Schorling DC, Becker J, Pechmann A, et al. Discrepancy in redetermination of SMN2 copy numbers in children with SMA. Neurology. 2019;93(6):267–269.
- Bladen CL, Thompson R, Jackson JM, et al. Mapping the differences in care for 5,000 spinal muscular atrophy patients, a survey of 24 national registries in North America, Australasia and Europe. J Neurol. 2014;261:152–163.
- Butchbach MER. Copy number variations in the survival motor neuron genes: implications for spinal muscular atrophy and other neurodegenerative diseases. Front Mol Biosc. 2016; 3:7.
- 12. Finkel RS, Mercuri E, Meyer OH, et al. Diagnosis and management of spinal muscular atrophy: part 2: pulmonary and acute care; medications, supplements and immunizations; other organ systems; and ethics. Neuromuscul Disord. 2018;28(3): 197–207.
- Reviews the importance of managing spinal muscular atrophy as a disease that affects multiple body systems.
- 13. Mercuri E, Finkel RS, Muntoni F, et al. Diagnosis and management of spinal muscular atrophy: part 1: recommendations for diagnosis, rehabilitation, orthopedic and nutritional care. Neuromuscul Disord. 2018;28:103–115.
- Reviews the importance of multidisciplinary management in the treatment of spinal muscular atrophy.
- Wang CH, Finkel RS, Bertinin ES, et al. Consensus statement for standard of care in spinal muscular atrophy. J Child Neurol. 2007;22 (8):1027–1049.
- 15. Ramdas S, Servais L. New treatments in spinal muscular atrophy: an overview of currently available data. Expert Opin Pharmacother. 2020;21(3):307–315.
- 16. De Vivo DC, Bertini E, Swoboda KJ, et al. Nusinersen initiated in infants during the presymptomatic stage of spinal muscular atrophy: interim efficacy and safety results from the phase 2 NURTURE study. Neuromuscul Disord. 2019;29(11):842–856.
- Dangouloff T, Servais L. Clinical evidence supporting early treatment of patients with spinal muscular atrophy: current perspectives. Ther Clin Risk Manag. 2019;15:1153–1161.
- Kirschner J, Butoianu N, Goemans N, et al. European ad-hoc consensus statement on gene replacement therapy for spinal muscular atrophy. Eur J Paediatr Neurol. 2020;28:38–43.
- Slayter J, Hodgkinson V, Lounsberry J, et al. A Canadian adult spinal muscular atrophy outcome measures toolkit: results of a national consensus using a modified delphi method. J Neuromuscul Dis. 2021;8(4):579–588.
- Glascock J, Sampson J, Connolly AM, et al. Revised recommendations for the treatment of infants diagnosed with spinal muscular atrophy via newborn screening who have 4 copies of SMN2.
 J Neuromuscul Dis. 2020;7(2):97–100.
- Kichula EA, Proud CM, Farrar MA, et al. Expert recommendations and clinical considerations in the use of onasemnogene abeparvovec gene therapy for spinal muscular atrophy. Muscle Nerve. 2021;64(4):413–427.
- Dangouloff T, Vrscaj E, Servais L, et al. Newborn screening programs for spinal muscular atrophy worldwide: where we stand and where to go. Neuromuscul Disord. 2021;31(6):574–582.
- 23. Dangouloff T, Botty C, Beaudart C, et al. Systematic literature review of the economic burden of spinal muscular atrophy and economic evaluations of treatments. Orphanet J Rare Dis. 2021;16(1):47.
- 24. Hale K, Ojodu J, Singh S. Landscape of spinal muscular atrophy newborn screening in the United States: 2018–2021. Int J Neonatal Screen. 2021;7(3):33.
- 25. Kariyawasam DST, D'Silva AM, Vetsch J, et al. "We needed this": perspectives of parents and healthcare professionals involved in a pilot newborn screening program for spinal muscular atrophy. EClinicalMedicine. 2021;33:100742.
- 26. Lopez-Chacon M, Buehner AN, Rao VK. Spinal muscular atrophy diagnosed by newborn screening. Pediatr Neurol Briefs. 2019;33:5.
- Lowes LP, Alfano LN, Arnold WD, et al. Impact of age and motor function in a phase 1/2A study of infants with SMA type 1 receiving

- single-dose gene replacement therapy. Pediatr Neurol. 2019;98:39–45.
- 28. Dangouloff T, Burghes A, Tizzano EF, et al. 244th ENMC international workshop: newborn screening in spinal muscular atrophy May 10–12, 2019, Hoofdorp, The Netherlands. Neuromuscul Disord. 2020;30(1):93–103.
- 29. Boardman FK, Sadler C, Young PJ. Newborn genetic screening for spinal muscular atrophy in the UK: the views of the general population. Mol Genet Genomic Med. 2018;6(1):99–108.
- Novartis Gene Therapies. Novartis Gene Therapies recommits to Global Managed Access Program for 2021. Novartis. 2021 Jan 15. Available at: www.novartis.com/news/novartis-gene-therapies-recommits-global-managed-access-program-2021 [Last accessed 2021 Jan 15]
- 31. Colella P, Ronzitti G, Mingozzie F. Emerging issues in AAV-mediated *in vivo* gene therapy. Mol Ther Methods Clin Dev. 2018;8:87–104.
- 32. Dayton RD, Wang DB, Klein RL. The advent of AAV9 expands applications for brain and spinal cord gene delivery. Expert Opin Biol Ther. 2012;12(6):757–766.
- 33. Murlidharan G, Sakamoto K, Rao L, et al. CNS-restricted transduction and CRISPR/Cas9-mediated gene deletion with an engineered AAV vector. Mol Ther Nucleic Acids. 2016;5:e338.
- 34. Rashnonejad A, Chermahini GA, Li S, et al. Large-scale production of adeno-associated viral vector serotype-9 carrying the human survival motor neuron gene. Mol Biotechnol. 2016;58(1):30–36.
- 35. Marshall E. Gene therapy death prompts review of adenovirus vector. Science. 1999;286(5448):2244–2245.
- 36. Thomsen G, Burghes AHM, Hsieh C, et al. Biodistribution of onasemnogene abeparvovec DNA, mRNA and SMN protein in human tissue. Nat Med. 2021;27(10):1701–1711.
- 37. Dalwadi DA, Calabria A, Tiyaboonchai A, et al. AAV integration in human hepatocytes. Mol Ther. 2021;29(10):2898–2909.
- 38. Domenger C, Grimm D. Next-generation AAV vectors—do not judge a virus (only) by its cover. Hum Mol Genet. 2019;28(R1):R3–R14.
- 39. Wang D, Tai PWL, Gao G. Adeno-associated virus vector as a platform for gene therapy delivery. Nat Rev Drug Discov. 2019;18(5):358–378
- 40. Ronzitti G, Gross D-A, Mingozzi F. Human immune responses to adeno-associated virus (AAV) vectors. Front Immunol. 2020;11:670.
- 41. Chu WS, Ng J. Immunomodulation in administration of rAAV: preclinical and clinical adjuvant pharmacotherapies. Front Immunol. 2021:12:658038.
- 42. Calcedo R, Morizono H, Wang L, et al. Adeno-associated virus antibody profiles in newborns, children, and adolescents. Clin Vaccine Immunol. 2011;18(9):1586–1588.
- 43. Day JW, Finkel RS, Mercuri E, et al. Adeno-associated virus serotype 9 antibodies in patients screened for treatment with onasemnogene abeparvovec. Mol Ther Methods Clin Dev. 2021;21:76–82.
- 44. Gorovits B, Azadeh M, Buchlis G, et al. Evaluation of the humoral response to adeno-associated virus-based gene therapy modalities using total antibody assays. AAPS J. 2021;23(6):108.
- 45. Saraiva J, Nobre RJ, Almeida LP. Gene therapy for the CNS using AAVs: the impact of systemic delivery by AAV. J Control Release. 2016;241:94–109.
- Mendell JR, Al-Zaidy SA, Rodino-Klapac LR, et al. Current clinical applications of *in vivo* gene therapy with AAVs. Mol Ther. 2021;29 (2):464–488
- 47. Day JW, Finkel RS, Chiriboga CA, et al. Onasemnogene abeparvovec gene therapy for symptomatic infantile-onset spinal muscular atrophy in patients with two copies of SMN2 (STR1VE): an open-label, single-arm, multicentre, phase 3 trial. Lancet Neurol. 2021;20 (4):284–293.
- One of two Phase III trials that studied the safety and efficacy of the therapeutic dose of onasemnogene abeparvovec for patients younger than 6 months old with spinal muscular atrophy type 1.
- Mendell JR, Al-Zaidy S, Shell R, et al. Single-dose gene-replacement therapy for spinal muscular atrophy. N Engl J Med. 2017;377: 1713–1722.



- .. The pivotal Phase I trial of onasemnogene abenaryovec that established its clinical benefit and safety for patients with SMA
- 49. Mendell JR, Al-Zaidy SA, Lehman KJ, et al. Five-year extension results of the phase 1 START trial of onasemnogene abeparvovec in spinal muscular atrophy. JAMA Neurol. 2021;78(7):834-841.
- 50. Mercuri E, Muntoni F, Baranello F, et al. Onasemnogene abeparvovec gene therapy for symptomatic infantile-onset spinal muscular atrophy type 1 (STR1VE-EU): an open-label, single-arm, multicentre, phase 3 trial. Lancet Neurol. 2021;20:832-841
- .. One of two Phase III trials that studied the safety and efficacy of the therapeutic dose of onasemnogene abeparvovec in patients younger than 6 months old with spinal muscular atrophy type 1.
- 51. McCarty DM, Monahan PE, Samulski RJ. Self-complementary recombinant adeno-associated virus (scAAV) vectors promote efficient transduction independently of DNA synthesis. Gene Ther. 2001;8 (16):1248-1254.
- 52. Kotulska K, Fattal-Valevski A, Haberlova J. Recombinant adeno-associated virus serotype 9 gene therapy in spinal muscular atrophy. Front Neurol. 2021;12:726468.
- 53. Chand D, Mohr F, McMillan H, et al. Hepatotoxicity following administration of onasemnogene abeparvovec (AVXS-101) for the treatment of spinal muscular atrophy. J Hepatol. 2021;74(3):540-566.
- 54. Zincarelli C, Soltys S, Rengo G, et al. Analysis of AAV serotypes 1-9 mediated gene expression and tropism in mice after systemic injection. Mol Ther. 2008;16(6):1073-1080.
- 55. ZOLGENSMA. Prescribing Information. Novartis; October 2021. Available at: www.novartis.us/sites/www.novartis.us/files/zolgen sma.pdf [Last accessed 2021 Nov 8]
- 56. Weiß C, Ziegler A, Becker LL, et al. Gene replacement therapy with onasemnogene abeparvovec in children with spinal muscular atrophy aged 24 months or younger and bodyweight up to 15 kg: an observational cohort study. Lancet Child Adolesc Health. 2022;6(1): 17-27.
- 57. Mercuri E, Lucibello S, Perulli M, et al. Longitudinal natural history of type I spinal muscular atrophy: a critical review. Orphanet J Rare Dis. 2020:15(1):84.
- 58. Finkel RS, McDermott MP, Kaufmann P, et al. Observational study of spinal muscular atrophy type I and implications for clinical trials. Neurology. 2014;83(9):810-817.
- A natural history study of patients with spinal muscular atrophy type 1.
- 59. Kolb SJ, Coffey CS, Yankey JW, et al., Natural history of infantile-onset spinal muscular atrophy. Ann Neurol. 2017;82(6):883-891.
- · A natural history study of patients with spinal muscular atrophy type 1.
- 60. Al-Zaidy SA, Kolb SJ, Lowes L, et al. AVXS-101 (onasemnogene abeparvovec) for SMA1: comparative study with a prospective natural history cohort. J Neuromuscul Dis. 2019;6(3):307-317.
- 61. Strauss K, Farrar M, Muntoni F, et al. OPR-201 Onasemnogene abeparvovec for presymptomatic infants with spinal muscular atrophy and 2 copies of SMN2: a phase III study. Eur J Neurol. 2021;28 (Suppl 1):S950-S951.
- 62. Strauss KA, Muntoni F, Farrar MA, et al. Onasemnogene abeparvovec gene therapy in presymptomatic spinal muscular atrophy: final results of SPR1NT study for children with three copies of SMN2. Neurology.2022;In press:Vol. 98.
- 63. SPR1NT: an open-label, single-arm clinical trial of presymptomatic patients with SMA. Novartis Gene Therapies, Inc., 2021. Available at: www.zolgensma-hcp.com/clinical-experiences/spr1nt-trial-efficacy. [Last accessed 2021 Nov 5]
- 64. De Onis M. WHO child growth standards based on length/height, weight and age. Acta Paediatr. 2006;450:76-85.
- 65. Finkel RS, Day JW, De Vivo DC, et al. RESTORE: a prospective multinational registry of patients with genetically confirmed spinal muscular atrophy - rationale and study design. J Neuromuscul Dis. 2020;7(2):145-152.
- 66. Servais L, De Vivo DC, Kirschner J, et al. Outcomes in US spinal muscular atrophy patients identified by newborn screening or

- RESTORE registry. clinical diagnosis: findings from the Neurology.2022;In press:Vol. 98.
- 67. Servais L, De Vivo DC, Kirschner J, et al. Effectiveness and safety of onasemnogene abeparvovec in older patients with spinal muscular atrophy: real-world outcomes from the RESTORE registry. Neurology.2022;In press:Vol. 98.
- 68. Registry of patients with a diagnosis of spinal muscular atrophy (SMA). ClinicalTrials.gov identifier: NCT04174157. 2019 Nov 22. Available at: clinicaltrials.gov/ct2/show/NCT04174157?term= NCT04174157&draw=2&rank=1 [Last accessed 2021 Nov 5]
- 69. Wei Y, McCormick A, MacKenzie A, et al. The Canadian Neuromuscular Disease Registry: connecting patients to national and international research opportunities. Paediatr Child Health. 2018:23(1):20-26.
- 70. Hodgkinson VL, Oskoui M, Lounsberry J, et al. A national spinal muscular atrophy registry for real-world evidence. Can J Neurol Sci. 2020;47(6):810-815.
- 71. Hodgkinson-Brechenmacher V, Oskoui M, Brais B, et al. SMA -OUTCOME MEASURES AND REGISTRIES, EP.262. The Canadian Neuromuscular Disease Registry: a national spinal muscular atrophy (SMA) registry for real-world evidence. Neuromuscul Disord. 2021;31(Supplement 1):S129.
- 72. Pechmann A, Konig K, Bernert G, et al. SMArtCARE A platform to collect real-life outcome data of patients with spinal muscular atrophy. Orphanet J Rare Dis. 2019;14(1):18.
- 73. Friese J, Geitmann S, Holzwarth D, et al. Safety monitoring of gene therapy for spinal muscular atrophy with onasemnogene abeparvovec -a single centre experience. J Neuromuscul Dis. 2021;8 (2):209-216.
- 74. Waldrop MA, Karingada C, Storey MA, et al. Gene therapy for spinal muscular atrophy: safety and early outcomes. Pediatrics. 2020;146 (3):e20200729.
- 75. Feldman AG, Parsons JA, Dutmer CM, et al. Subacute liver failure following gene replacement therapy for spinal muscular atrophy type 1. J Pediatr. 2020;225:252-258.
- 76. Shieh PB, Bonnemann CG, Muller-Felber W, et al. Re: "Moving forward after two deaths in a gene therapy trial of myotubular myopathy" by Wilson and Flotte. Hum Gene Ther. 2020;31(15-16):787.
- 77. Wilson JM, Flotte TR. Moving forward after two deaths in a gene therapy trial of myotubular myopathy. Hum Gene Ther. 2020;31(13-14):695-696.
- 78. D'Amico A, Longo A, Fattori F, et al. Hepatobiliary disease in XLMTM: a common comorbidity with potential impact on treatment strategies. Orphanet J Rare Dis. 2021;16(1):425.
- 79. Philippidis A. Fourth boy dies in clinical trial of astellas' AT132. Hum Gene Ther. 2021;32(19-20):1008.
- 80. Perez BA, Shutterly A, Chan YK, et al. Management of neuroinflammatory responses to AAV-mediated gene therapies for neurodegenerative diseases. Brain Sci. 2020;10(2):119.
- 81. Childers MK, Joubert R, Poulard K, et al. Gene therapy prolongs survival and restores function in murine and canine models of myotubular myopathy. Sci Transl Med. 2014;6(220):220ra10.
- 82. Morales L, Gambhir Y, Bennett J, et al. Broader implications of progressive liver dysfunction and lethal sepsis in two boys following systemic high-dose AAV. Mol Ther. 2020;28(8):1753-1755.
- 83. Chand DH, Zaidman C, Arya K, et al. Thrombotic microangiopathy following onasemnogene abeparvovec for spinal muscular atrophy: a case series. J Pediatr. 2021;231:265-268.
- 84. Day JW, Mendell JR, Mercuri E, et al. Clinical trial and postmarketing safety of onasemnogene abeparvovec therapy. Drug Saf. 2021;44:1109-1119.
- · A description of risks associated with onasemnogene abeparvovec therapy and their potential management.
- 85. D'Silva AM, Holland S, Kariyawasam D, et al. Onasemnogene abeparvovec in spinal muscular atrophy: an Australian experience of safety and efficacy. Ann Clin Transl Neurol. 2022;9(3):339-350.
- 86. Bonnemann CG. AAV related immunological safety and toxicity: preliminary clinical observations in the GAN and MTM1 trials. Presented at: Virtual Workshop on Systemic Immunogenicity



- Considerations of AAV-Mediated Gene Therapy, NIH, NCATS; 2020. Available at: videocast.nih.gov/watch=38547 [Last accessed 2021 Nov 81
- 87. Mueller C, Berry JD, McKenna-Yasek DM, et al. SOD1 Suppression with adeno-associated virus and MicroRNA in familial ALS. N Engl J Med. 2020;383(2):151-158.
- 88. Van Alstyne M, Tattoli I, Delestree N, et al. Gain of toxic function by long-term AAV9-mediated SMN overexpression in the sensorimotor circuit. Nat Neurosci. 2021;24(7):930-940.
- 89. Crawford TO, Sumner CJ. Assuring long-term safety of highly effective gene-modulating therapeutics for rare diseases. J Clin Invest. 2021:131(15):e152817.
- 90. Huang L. Wan J. Wu Y. et al. Challenges in adeno-associated virus-based treatment of central nervous system diseases through systemic injection. Life Sci. 2021;270:119142.
- 91. Duan D. Systemic AAV micro-dystrophin gene therapy for Duchenne muscular dystrophy. Mol Ther. 2018;26(10):2337-2356.
- 92. Ramos J, Chamberlain JS. Gene therapy for duchenne muscular dystrophy. Expert Opin Orphan Drugs. 2015;3(11):1255-1266.
- 93. Philippidis A. After patient death, FDA places hold on Pfizer Duchenne muscular dystrophy gene therapy trial. Hum Gene Ther. 2022;33(3-4):111-115.
- 94. Mendell JR, Campbell K, Rodino-Klapac L, et al. Dystrophin Immunity in Duchenne's Muscular Dystrophy. N Engl J Med. 2010;363(15):1429-1437.
- 95. A study to evaluate the safety and tolerability of PF-06939926 gene therapy in Duchenne Muscular Dystrophy. ClinicalTrials.gov identifier: NCT03362502. 2021 Oct 15. Available at: clinicaltrials.gov/ct2/ show/NCT03362502. [Last accessed 2021 Dec 13]
- 96. Landfeldt E, Pechmann A, McMillan HJ, et al. Costs of illness of spinal muscular atrophy: a systematic review. Appl Health Econ Health Policy. 2021;19(4):501-520.
- 97. McMillan HJ, Gerber B, Cowling T, et al. Burden of spinal muscular atrophy (SMA) on patients and caregivers in Canada. J Neuromuscul Dis. 2021;8(4):553-568.
- 98. Belter L, Cruz R, Kulas S, et al. Economic burden of spinal muscular atrophy: an analysis of claims data. J Mark Access Health Policy. 2020;8(1):1843277.
- 99. Garrison LP, Jiao B, Dabbous O. Gene therapy may not be as expensive as people think: challenges in assessing the value of single and short-term therapies. J Manag Care Spec Pharm. 2021;27(5):674-681
- 100. Dean R, Jensen I, Cyr P, et al. An updated cost-utility model for onasemnogene abeparvovec (Zolgensma®) in spinal muscular atrophy type 1 patients and comparison with evaluation by the institute for clinical and effectiveness review (ICER). J Mark Access Health Policy. 2021;9:1889841.
- 101. Harris E. Potential solutions to current pricing models for cell and gene therapies. Horsham, PA: Life Science Leader, 2019 Oct 1. Available at: www.lifescienceleader.com/doc/potentialsolutions-to-current-pricing-models-for-cell-and-gene-therapies -0001 [Last accessed 2021 Nov 5]
- 102. Seymore B. Payment models for pricey transformative pharmaceuticals. PharmacyTimes.com. Updated 2020 Feb 4. Available at: https://www. pharmacytimes.com/view/payment-models-for-pricey-transformativepharmaceuticals. [Last accessed 2021 Nov 5]
- 103. Wong CH, Li D, Wang N, et al. Estimating the financial impact of gene therapy, medRxiv, Available at: www.medrxiv.org/content/10. 1101/2020.10.27.20220871v1.full [Last accessed 2021 Nov 8]
- 104. Aballéa S, Thokagevistk K, Velikanova R, et al. Health economic evaluation of gene replacement therapies: methodological issues and recommendations. J Mark Access Health Policy. 2020;8: 1822666.
- 105. Broekhoff TF, Sweegers CCG, Krijkamp EM, et al. Early cost-effectiveness of onasemnogene abeparvovec-xioi (Zolgensma) and Nusinersen (Spinraza) treatment for spinal muscular atrophy I in the Netherlands with relapse scenarios. Value Health. 2021:24:759-769.
- 106. Jena AB, Lakdawalla DN. Value frameworks for rare diseases: should they be different? Washington, DC: Health Affairs Blog, 2017 Apr

- 12. Available at: www.healthaffairs.org/do/10.1377/hblog20170412. 059563/full/ [Last accessed 2021 Apr 12].
- 107. Ollendorf DA, Chapman RH, Pearson SD. Evaluating and valuing drugs for rare conditions: no easy answers. Value Health. 2018;21:547–552.
- 108. Shih STF, Farrar M, Wiley V, et al. Newborn screening for spinal muscular atrophy with disease-modifying therapies: a cost-effectiveness analysis. J Neurol Neurosurg Psychiatry. 2021;92(12):1296-1304.
- 109. Safety and efficacy of intravenous OAV101 (AVXS-101) in pediatric patients with spinal muscular atrophy (SMA) (SMART). clinicalTrials. gov identifier: NCT04851873, 2021 Apr 21, Available at: clinicaltrials. gov/ct2/show/NCT04851873 [Last accessed 2021 Nov 5]
- 110. Study of intrathecal administration of onasemnogene abeparvovec-xioi for spinal muscular atrophy (STRONG). ClinicalTrials.gov identifier: NCT03381729. 2021 June 9. Available at: https://clinicaltrials.gov/ct2/show/NCT03381729?term= NCT03381729 [Last accessed 2021 Nov 18]
- 111. Finkel RS, Day JW, Darras BT, et al. One-time administration of AVXS-101 IT for spinal muscular atrophy: phase I/II study (STRONG). Presented online at the 16th International Child Neurology Congress/49th International Child Neurology Society 2020 Virtual Annual Meeting; 2020 Oct 12-20.
- 112. Efficacy and safety of intrathecal OAV101 (AVXS-101) in pediatric patients with type 2 spinal muscular atrophy (SMA) (STEER). ClinicalTrials.gov identifier: NCT05089656, 2021 Oct 22, Available at: clinicaltrials.gov/ct2/ show/NCT05089656. [Last accessed 2021 Nov 5]
- 113. Harada Y, Rao VK, Arya K, et al. Combination molecular therapies for type 1 spinal muscular atrophy. Muscle Nerve. 2020;62(4):550-554.
- 114. Kray KM, McGovern VL, Chugh D, et al. Dual SMN inducing therapies can rescue survival and motor unit function in symptomatic Δ7SMA mice. Neurobiol Dis. 2021;159:105488.
- 115. Pagliarini V, Guerra M, Di Rosa V, et al. Combined treatment with the histone deacetylase inhibitor LBH589 and a splice-switch antisense oligonucleotide enhances SMN2 splicing and SMN expression in spinal muscular atrophy cells. J Neurochem. 2020;153:264-275.
- 116. Poletti A, Fischbeck KH. Combinatorial treatment for spinal muscular atrophy: an editorial for 'Combined treatment with the histone deacetylase inhibitor LBH589 and a splice-switch antisense oligonucleotide enhances SMN2 splicing and SMN expression in spinal muscular atrophy cells' on page 264. J Neurochem. 2020;153(2):146-149.
- 117. A study of nusinersen among participants with spinal muscular atrophy who received onasemnogene abeparvovec (RESPOND). ClinicalTrials. gov identifier: NCT04488133. 2021 Sept 29. Available at: www.clinical trials.gov/ct2/show/NCT04488133 [Last accessed 2021 Oct 20]
- 118. Masson R, Brusa C, Scoto M, et al. Brain, cognition, and language development in spinal muscular atrophy type 1: a scoping review. Dev Med Child Neurol. 2021;63(5):527-536.
- 119. Nance ME, Shi R, Hakim CH, et al. AAV9 edits muscle stem cells in normal and dystrophic adult mice. Mol Ther. 2019;27 (9):1568-1585.
- 120. Fischell JM, Fishman PS. A multifaceted approach to optimizing AAV delivery to the brain for the treatment of neurodegenerative diseases. Front Neurosci. 2021;15:747726.
- 121. Liu D, Zhu M, Zhang Y, et al. Crossing the blood-brain barrier with AAV vectors. Metab Brain Dis. 2021;36(1):45-52.
- 122. Nonnenmacher M, Wang W, Child MA, et al. Rapid evolution of blood-brain-barrier-penetrating AAV capsids by RNA-driven biopanning. Mol Ther. 2021;20:366-378.
- 123. Marsic D, Mendez-Gomez HR, Zolotukhin S. High-accuracy biodistribution analysis of adeno-associated virus variants by double barcode sequencing. Mol Ther Methods Clin Dev. 2015;2:15041.
- 124. Weinmann J, Weis S, Sippel J, et al. Identification of a myotropic AAV by massively parallel in vivo evaluation of barcoded capsid variants. Nature. 2020;11:5432.
- 125. Tabebordbar M, Lagerborg KA, Stanton A, et al. Directed evolution of a family of AAV capsid variants enabling potent muscle-directed gene delivery across species. Cell. 2021;184(19):4919-38.e22.
- 126. Kariyawasam DST, Russell JS, Wiley V, et al. The implementation of newborn screening for spinal muscular atrophy: the Australian experience. Genet Med. 2020;22(3):557-565.



- 127. D'Silva AM, Kariyawasam DST, Best S, et al. Integrating newborn screening for spinal muscular atrophy into health care systems: an Australian pilot programme. Dev Med Child Neurol. 2022;64 (5):625-632.
- 128. Hanlon KS, Kleinstiver BP, Garcia SP, et al. High levels of AAV vector integration into CRISPR-induced DNA breaks. Nat Commun. 2019;10:4439.
- 129. Yu W, Wu Z. Use of AAV vectors for CRISPR-mediated in vivo genome editing in the retina. Methods Mol Biol. 2019;1950:123-139.
- 130. Li JJ, Lin X, Tang C, et al. Disruption of splicing-regulatory elements using CRISPR/Cas9 to rescue spinal muscular atrophy in human iPSCs and mice. Natl Sci Rev. 2020;7:92-101.
- 131. Zhou M, Hu Z, Qiu L, et al. Seamless genetic conversion of SMN2 to SMN1 via CRISPR/Cpf1 and single-stranded oligodeoxynucleotides in spinal muscular atrophy patient-specific induced pluripotent stem cells. Hum Gene Ther. 2018;29:1252-1263.
- 132. Waddington SN, Privolizzi R, Karda R, et al. A broad overview and review of CRISPR-Cas technology and stem cells. Curr Stem Cell Rep. 2016;2:9-20.

- 133. Xu Y, Li Z. CRISPR-Cas systems: overview, innovations and applications in human disease research and gene therapy. Comput Struct Biotechnol J. 2020;18:2401-2415.
- 134. Khalaf K, Janowicz K, Dyszkiewicz-Konwinska M, et al. CRISPR/Cas9 in cancer immunotherapy: animal models and human clinical trials. Genes (Basel). 2020;11:921.
- 135. Hirakawa MP, Krishnakumar R, Timlin JA, et al. Gene editing and CRISPR in the clinic: current and future perspectives. Biosci Rep. 2020;40(4):BSR20200127.
- 136. U.S. National Library of Medicine. ClinicalTrials.gov. Available at: https://clinicaltrials.gov/ct2/results?cond=&term=crispr&cn try=&state=&city=&dist= [Last accessed 2022 Mar 10]
- 137. Kariyawasam DST, D'Silva A, Lin C, et al. Biomarkers and the development of a personalized medicine approach in spinal muscular atrophy. Front Neurol. 2019;10:898
- 138. Servais L, Baranello G, Scoto M, et al. Therapeutic interventions for spinal muscular atrophy: preclinical and early clinical development opportunities. Expert Opin Investig Drugs. 2021;30 (5):519-527.