

1 **Cambrian edrioasteroid reveals new mechanism for**
2 **secondary reduction of the skeleton in echinoderms**

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22 **Abstract**

23 Echinoderms are characterized by a distinctive high-magnesium calcite endoskeleton as
24 adults, but elements of this have been drastically reduced in some groups. Herein, we
25 describe a new pentaradial echinoderm, *Yorkicystis haefneri* n. gen. n. sp., which
26 provides the oldest evidence of secondary non-mineralization of the echinoderm
27 skeleton. This material was collected from the Cambrian Kinzers Formation in York
28 (Pennsylvania, USA) and is dated as *c.* 510 Ma. Detailed morphological observations
29 demonstrate that the ambulacra (i.e. axial region) are composed of flooring and cover
30 plates, but the rest of the body (i.e. extraxial region) is preserved as a dark film and
31 lacks any evidence of skeletal plating. Moreover, X-ray fluorescence analysis reveals
32 that the axial region is elevated in iron. Based on our morphological and chemical data
33 and on taphonomic comparisons with other fossils from the Kinzers Formation, we infer
34 that the axial region was originally calcified, while the extraxial region was non-
35 mineralized. Phylogenetic analyses recover *Yorkicystis* as an edrioasteroid, indicating
36 that this partial absence of skeleton resulted from a secondary reduction. We
37 hypothesize that skeletal reduction resulted from lack of expression of the skeletogenic
38 gene regulatory network in the extraxial body wall during development. Secondary
39 reduction of the skeleton in *Yorkicystis* might have allowed for greater flexibility of the
40 body wall.

41

42 **Keywords:** Cambrian, echinoderms, skeleton, evolution, development

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44

45 **1. Introduction**

46 Animals are characterized by an enormous diversity of biomineralized skeletons, which
47 perform important functions such as protecting the internal organs or supporting the
48 body structure. Elucidating the origins of animal biomineralization is a major research
49 focus in evolutionary and geobiology; skeletons are thought to have evolved
50 independently in various clades during the latest Ediacaran or early Cambrian through
51 co-option of a conserved ancestral genetic toolkit [1,2], with possible drivers including
52 changing seawater chemistry, oxygen availability or increasing predation pressure [3,4].
53 However, the pattern and process of skeletal loss, widespread across animal phylogeny
54 [5], is much less well studied, and it remains unclear if commonalities exist in how and
55 why skeletons are reduced among species.

56 Echinoderms, with their distinctive high-magnesium calcite endoskeleton, are a
57 model system for studying the evolution and development of the skeleton [6].
58 Calcareous plates or ossicles are ubiquitous in most echinoderm bodies, but the
59 mineralized skeleton has been greatly reduced in some groups. For example, in derived
60 crinoids the ambulacral flooring plates are decalcified [7], whereas in crown-group
61 holothurians mineralization of the body-wall skeleton is mostly reduced to small
62 spicules [8]. Thus, reduction of mineralization primarily affects distinct parts of the
63 body in different classes; in crinoids, the axial region (i.e. parts of the body associated
64 with the water vascular system) is most strongly affected, whereas in holothurians there
65 is greater loss of mineralization in the extraxial region (i.e. the rest of the body).
66 Although developmental mechanisms associated with skeletal formation are well
67 understood in many echinoderms [9,10], the processes involved in skeletal reduction
68 remain enigmatic. The reasons why some echinoderm groups reduced mineralization in
69 their skeleton are also unclear, with explanations ranging from adaptation to more
70 mobile lifestyles [11] to reducing the energetic cost of skeletogenesis [12].

71 Here, we describe a new echinoderm from the Cambrian of the USA, which has
72 a unique body plan consisting of a calcified axial skeleton and a non-mineralized
73 extraxial region. This taxon demonstrates that skeletal non-mineralization in
74 echinoderms first occurred during the Cambrian, in a fundamentally different way to all
75 other species.

76

77 **2. Material and methods**

78 Two specimens (NHMUK EE 1659, 1660), preserved as external moulds in laminated
79 brown shale, were collected from the Emigsville Member of the Kinzers Formation in
80 York, Pennsylvania, USA (for more information about this formation and fossil content,
81 see [13]). Specimens were obtained from the south slope of the City View Church yard
82 by Mr. Christopher Haefner in November 2018 (with permission from the church
83 community). The coordinates of the exposure are 39.9842302 N, 76.7644671 W. At this
84 locality, the Kinzers Formation crops out in a small tectonic block that includes the
85 Emigsville and York members. Excavation yielded a rich associated fauna of trilobites,
86 echinoderms (*Lepidocystis* and *Camptostroma*) and soft-bodied organisms (e.g.
87 radiodontans). [Echinoderms from this formation always show three-dimensional](#)
88 [preservation of the skeleton](#). The Kinzers Formation extends from the middle Dyeran
89 Stage to somewhere within the Delamarian Stage of the Laurentian zonation scheme for
90 the Cambrian. The olenelloid trilobites *Wanneria walcottana* and *Olenellus rodnyi*
91 recovered from the City View Church site are consistent with this age assignment
92 (Webster pers. comm. March 26, 2019). This falls within Stage 4 of the Cambrian in the
93 global chronostratigraphic scale, approximately 510 Ma [14,15].

94 Specimens were photographed under natural light, both dry and submerged in
95 water. To investigate the elemental distribution of the fossils, NHMUK EE 1659 was

96 analysed using a Bruker Tornado M4+ micro X-ray fluorescence (XRF) scanning
97 spectrometer at University College London. Analysis with the Tornado M4+ was
98 performed under a 2 mbar vacuum. The system has an X-ray tube with a rhodium target
99 which was used with a 35 kV accelerating voltage and an 800 μ A current. The dwell
100 time was 100 μ s, the resolution was 1228 \times 888 pixels and the pixel size was 25 μ m.
101 The resulting fluorescence signal was detected using two silicon drift detector
102 spectrometers. Lastly, specimens were cast in latex, with the resulting casts whitened
103 with NH₄Cl sublimate prior to photography. Specimens are housed at the Natural
104 History Museum of London, UK (NHMUK). Comparative material of other
105 echinoderms from the Kinzers Formation is deposited in the Museum of Comparative
106 Zoology (Harvard University) and National Museum of Natural History (Smithsonian
107 Institution).

108 To establish the relationships between *Yorkicystis* and other early echinoderms,
109 parsimony and Bayesian phylogenetic analyses were performed. We chose to focus
110 exclusively on pentaradial forms because all recent phylogenetic analyses
111 [16,17,18,19,20,21] have recovered these as derived in echinoderm phylogeny. We
112 selected all the major pentaradial morphotypes present in the Cambrian and early
113 Ordovician for which fossil material is sufficiently well known, including a spiral form
114 (*Helicocystis*), early edrioasteroids (*Kailidiscus* and *Stromatocystites*), isorophid
115 edrioasteroids (*Isorophus* and *Argodiscus*), plesiomorphic blastozoans (*Lepidocystis*,
116 *Gogia*, and *Vyscystis*), an early glyptocystitid (*Ridersia*) and the earliest crinoids
117 (*Titanocrinus* and *Apektocrinus*). *Helicoplacus* was chosen as the outgroup because it is
118 a triradial form that is widely regarded as the sister group to pentaradial forms [17,22].
119 Morphological information was obtained from direct observations of fossil specimens
120 and the published literature. Most of the taxa were coded as in previous phylogenetic

121 analyses [23,24,25]. The final character matrix consisted of 13 taxa and 28 characters
122 (supplementary table 1). Parsimony analyses were run using the branch and bound
123 algorithm in the program Paup* v. 4.0a [26]. Bayesian analyses were run using
124 MrBayes v. 3.2 [27] using the Mkv model [28]. Rate variation was modeled using a
125 gamma distribution with a prior of exponential (1.0). Branch lengths were
126 unconstrained with a compound [Dirichlet](#) prior. Two differing values of the symmetric
127 [Dirichlet](#) hyperprior were used to account for differing transition rate [asymmetries](#). The
128 joint posterior distribution of model parameters, branch lengths, and tree topologies was
129 estimated using Markov chain Monte Carlo (MCMC). Additional details of
130 phylogenetic analyses and MCMC convergence are provided in the supplementary
131 information.

132

133 **3. Results**

134 **(a) Morphology of *Yorkicystis gen. nov.***

135 The body of *Yorkicystis haefneri* gen. et sp. nov. is divided into two main regions that
136 are preserved in different ways and correspond to the extraxial and axial body walls
137 (*sensu* [29, 30]). The extraxial region of *Y. haefneri* gen. et sp. nov. consists of a
138 globular body (figures 1*a,b*, 2*a,b*; supplementary material, figure S3), measuring 23 mm
139 wide and 18 mm long in the most complete specimen (NHMUK EE 1659). It is
140 preserved as a dark film (figure 2*a*), with no evidence of skeletal plating (figure 1*a,b*;
141 supplementary material, figure S3). No attachment structure or body openings (e.g.
142 periproct, gonopore, hydropore) are preserved.

143 The axial region is composed of five large, probably straight, recumbent
144 ambulacra that converge in the peristomial region and presumably originate at the
145 centrally located mouth (figures 1*a,b*, 2*a,b*). The ambulacra are long (maximum length

146 of 20 mm in the holotype and 24 mm in the paratype), wide in plan view, and slightly
147 taper in width distally (figure 1*a–e*). Each ambulacrum is composed of two series of
148 plates, which we interpret as flooring and cover plates, similar to those in other
149 edrioasteroid-grade echinoderms [31]. Flooring plates are biserially arranged, triangular
150 in shape, and very thick (figure 1*f–i*). Their internal surfaces are smooth, with notches at
151 the lateral margins that we interpret as podial pores (*sensu* [32]) (figure 1*f,i*). Externally,
152 these plates have prominent rims converging at a central point that extend to the corners
153 of each plate (figure 1*c,d,e,g*). These rims are elevated compared to the main body of
154 the plates. Cover plates are tessellate and organized into multiple series of tiny
155 polygonal plates, which decrease in size towards the perradial line (figure 1*c–e*). The
156 first series of cover plates articulate with the apices of the flooring plates. Ambulacra
157 are preserved in reddish-orange (figure 2*a*). The peristome is covered by tiny plates with
158 no obvious organization preserved (figure 1*a,b*).

159

160 **(b) *Elemental analysis***

161 XRF mapping showed that NHMUK EE 1659 is enriched in P, Fe, Ca, Zn and S and
162 depleted in Si, Al, K and Mg compared to the surrounding matrix (figure 2*b*;
163 supplementary material, figure S2). Within the fossil, the axial region is especially
164 elevated in Fe (figure 2*b*; supplementary material, figure S2), with local enrichment of
165 Zn and S in some parts (supplementary material, figure S2); P and Ca are elevated in
166 both the axial and extraxial regions (figure 2*b*; supplementary material, figure S2).

167

168 **(c) *Phylogenetic position***

169 In both our Bayesian and parsimony analyses, *Yorkicystis* was recovered as an
170 edrioasteroid, in a clade with *Argodiscus*, *Isorophus*, and *Kailidiscus* (figure 3). This

171 clade of edrioasteroids was part of a larger clade consisting of all pentaradial forms
172 excluding *Helicocystis*. In both analyses, *Helicocystis* was retrieved as the earliest-
173 diverging pentaradial echinoderm.

174

175 **4. Discussion**

176 Variation in the morphology and preservation of the axial and extraxial parts of the
177 body in *Yorkicystis* demonstrates that these regions had different compositions in life.
178 While the specimens preserve no trace of original calcite, the presence of ambulacra
179 with characteristic flooring and cover plates (figures 1*c–i*, 4), preserved as 3-
180 dimensional moulds elevated in Fe (figure 2*b*; supplementary material, figure S2), and
181 the relict ‘ghost’ stereom microstructure in the external part of flooring plates are clear
182 indicators of a typical echinoderm skeleton. In contrast, the extraxial region is generally
183 preserved as a dark film (figure 2*a*). This mode of preservation is common among
184 Burgess Shale-type deposits like the Kinzers Formation [33, 34], and soft tissues are
185 often preserved in this way [33, 35]. This, together with the apparent absence of skeletal
186 plating, as revealed by direct study of the fossils and latex casting (figures 1*a,b*, 2*a,b*;
187 supplementary material, figure S3), strongly suggests the extraxial region of *Yorkicystis*
188 was originally non-mineralized. Although the fossils appear preserved flattened and
189 with curved ambulacra, taphonomic observations indicate that their original shape was
190 globular and probably with straight ambulacra (supplementary material, figure S1).

191 Our phylogenetic analyses recover *Yorkicystis* as an edrioasteroid (figure 3),
192 revealing that the reduction of the skeleton was a derived trait, and not representative of
193 the plesiomorphic condition among pentaradial echinoderms. *Yorkicystis* is thus
194 interpreted as the oldest echinoderm with a secondarily non-mineralized body wall
195 (figure 5). While a small number of Ediacaran and lower Cambrian fossils have

196 previously been considered as uncalcified echinoderms (e.g. [18,36,37,38]), their
197 echinoderm affinities are highly dubious and debated [20,22,39,40]. However,
198 comparable patterns of incomplete calcification have been documented in other
199 echinoderm groups based on younger fossil and living forms. For example, in most
200 crinoids, flooring plates are weakly or not calcified, while other elements of the axial
201 skeleton are mineralized [7, 41]; in some taxa, components of the extraxial region, such
202 as the anal sac, are also uncalcified [12]. Some edrioasteroids like isorophids and
203 *Walcottidiscus* also reduce part of the aboral region to attach on substrates [42]. More
204 radical reduction of skeletal mineralization is present among holothurians, in which the
205 skeleton typically consists of small spicules embedded in the body wall and a ring of
206 ossicles surrounding the pharynx [8,43], or is lost entirely [11, 44]; skeletal reduction
207 thus affects both the axial and extraxial regions. Consequently, *Yorkicystis* is unique
208 among echinoderms in having a clear differentiation between calcified axial and
209 uncalcified extraxial regions.

210 Developmental mechanisms underpinning the formation of the skeleton in
211 *Yorkicystis* may shed light on skeletal development in echinoderms more broadly.
212 Transcriptomic comparisons of crinoids, echinoids, asteroids, and ophiuroids [10], as
213 well as numerous experimental studies in larval and adult skeletons of extant
214 echinoderm classes, have identified a conserved biomineralization ‘toolkit’ of genes and
215 proteins that underlies the process of skeletogenesis. This includes transcription factors
216 such as *Alx1* and *Ets1*, signalling pathways like FGF and VEGF, and numerous
217 downstream differentiation genes such as c-lectin protein domain-bearing genes,
218 metalloproteases, and cell-surface molecules [9,10,45,46]. The existence of this
219 conserved set of genes across echinoderms points to a distinct genetic regulatory
220 module directing the development of the echinoderm skeleton. During development,

221 aspects of this regulatory network are deployed in distinct spatial contexts based on
222 ectodermal signalling cues, resulting in the growth of skeleton in particular parts of the
223 body [45–48]. Inhibiting these signalling pathways results in downregulation of
224 skeletogenic genes, and obstruction of biomineralization in both embryos and adult
225 echinoderms [46–48]. Given the conserved nature of the main components of the
226 echinoderm skeletal biomineralization toolkit, we expect that the echinoderm skeletal
227 gene regulatory network has been expressed during skeletal growth in both axial and
228 extraxial skeletons of all fossil echinoderms. We hypothesize that, where this
229 skeletogenic module is not deployed, skeletal loss or reduction will result. In this
230 context, *Yorkicystis*, with its uncalcified extraxial body, indicates that components of
231 the skeletogenic gene regulatory network were expressed only in the axial region.

232 The morphology of *Yorkicystis* could also indicate that the activation of the
233 skeletogenic gene regulatory network in axial and extraxial tissues is independent and
234 that the expression of this gene network in these tissues may be controlled by different
235 upstream developmental genetic mechanisms, such as differential deployment of
236 signalling pathways. This might imply different molecular mechanisms underlying the
237 development of axial and extraxial tissue [49], although the existence of other
238 echinoderms with varying degrees of calcification across axial and extraxial regions
239 (see above) could point to greater variability in developmental processes between taxa.
240 Recent advances in CRISPR genome editing have facilitated functional perturbations of
241 gene expression in adult echinoderms in distinct morphological regions of the adult
242 body plan [50]. Based on the morphology of *Yorkicystis*, with its calcified axial and
243 uncalcified extraxial skeletons, we hypothesize that that functional knockout of
244 transcription factors or signalling genes toward the top of the skeletal gene regulatory
245 network hierarchy, such as *Alx1* or *VegfR*, in the extraxial regions of growing adult

246 echinoderms will result in a distinct loss of skeleton in those regions. Developments in
247 functional perturbations, paired with recent advances in localizing skeletal gene
248 expression in post-metamorphic echinoderms [51], make testing this hypothesis
249 possible.

250 Early echinoderms displayed great plasticity in terms of their body-plan
251 construction, with bilateral, asymmetrical, triradial and pentaradial forms described
252 from the Cambrian [22,52]. Recent phylogenetic analyses [18,20,17,19] place bilateral
253 and asymmetrical forms as stem-group echinoderms, indicating that some of the
254 synapomorphies of crown-group echinoderms, such as pentaradial symmetry and
255 ambulacra with flooring and cover plates, are not plesiomorphic for the phylum [16,22].
256 Our phylogenetic analyses recover *Yorkicystis* as a derived pentaradial form, most
257 closely related to Cambrian edrioasteroids such as *Kailidiscus*, with the absence of
258 skeleton in the extraxial part of the body due to secondary loss. This represents the
259 oldest example of skeletal reduction yet documented in echinoderms. Moreover, the
260 secondary loss of skeleton in the extraxial region alone differs from the situation in all
261 other echinoderms with reduced skeletons (e.g. crinoids and holothurians), and strongly
262 implies a distinct mechanism for reducing the skeleton. The absence of skeleton in the
263 extraxial region suggests that *Yorkicystis* preferentially directed energy towards
264 skeletogenesis in the axial region. This would have conserved energy for other
265 metabolic requirements, while still ensuring the external soft parts of the water vascular
266 system (i.e. the tube feet, figure 4k) were supported and protected. The absence of
267 plating in the extraxial part would also explain why the ambulacra in *Yorkicystis* depart
268 morphologically from other edrioasteroids. Loss of the extraxial skeleton might have
269 enabled greater flexibility of the body wall, allowing the animal to vary its body shape
270 in response to changing currents.

271

272 **5. Systematic palaeontology**

273 Phylum: Echinodermata

274 Class: Edrioasteroidea

275 Family: Yorkicystitidae nov.

276 Genus: *Yorkicystis* gen. nov.

277 Type species: *Yorkicystis haefneri* sp. nov.

278

279 *Etymology.* Genus name refers to the city of York, Pennsylvania, where specimens

280 were collected.

281

282 *Diagnosis.* Extraxial body uncalcified. Axial body composed of five large,

283 recumbent ambulacra incorporated into the thecal wall. Ambulacra consist of

284 biserially arranged, large triangular adradial flooring plates with external rims;

285 flooring plates internally smooth, with podial pores along lateral margins. Cover

286 plates organized into multiple series.

287

288 *Discussion.* The new family Yorkicystitidae is here created to accommodate the new

289 genus and species *Yorkicystis haefneri*, which is differentiated from all other

290 edrioasteroids based on its unique body construction (uncalcified extraxial region

291 and ambulacral construction). The ambulacra, consisting of multiple series of cover

292 plates and biserial flooring plates, differ from those of derived isorophid

293 edrioasteroids, which have uniserial flooring plates [23,53], but are more similar to

294 stromatocystitids (i.e. *Cambraster* and *Stromatocystites*) and edrioasterids in which

295 flooring plates are biserially arranged [32,42,54,55]. The flooring plate system in

296 *Yorkicystis* probably corresponds to and is thus homologous with the adradial series
297 of *Walcottidiscus* [42]. In *Cambraster*, *Stromatocystites*, and edrioasterids the
298 flooring plates are externally exposed and articulate aborally with the tessellate
299 interambulacral membrane; this is not the case in *Yorkicystis* because of the absence
300 of calcified elements in the extraxial region. But the floor plates do not have an
301 externally exposed shelf that would be expected if they were the abradial set. Instead
302 they are strongly rimmed in *Yorkicystis*.

303 The cover plate system in *Yorkicystis* is similar to the multitiered systems in
304 some other Cambrian echinoderms, including the edrioasteroid *Kailidiscus* and the class
305 *Cincta*, where larger platelets abradially articulate with tiers of smaller platelets towards
306 the midline. This type of cover plate system is unknown among taxa with biserial
307 abradial floor plates, except for the edrioasterid *Pseudedriophus guensburgi*, which is
308 otherwise dissimilar [56].

309

310 *Yorkicystis haefneri* sp. nov.

311

312 *Etymology.* Species name dedicated to Mr. Christopher Haefner, who discovered the
313 two known specimens and made them available for research.

314

315 *Diagnosis.* As for genus.

316

317 *Material.* Holotype: NHMUK EE 1659, includes part and counterpart. Paratype:
318 NHMUK EE 1660, is a complete specimen with well preserved ambulacra.

319

320 *Locality and horizon.* York, Pennsylvania, USA; Stage 4, Series 2, Cambrian.

321

322 *Description.* See above and supplementary information.

323

324 **Data accessibility.** Data from elemental analysis are available from the Dryad Digital
325 Repository: XXXXXXXXXXXX [REF].

326

327 **Authors' contributions.** S.Z. conceived the study, cast and photographed specimens,
328 conceived the phylogenetic analysis, analysed and interpreted data, and drafted the
329 manuscript. I.A.R. conceived the elemental analysis, helped draft the manuscript,
330 analysed and interpreted data, and critically revised the manuscript. C.D.S. conceived
331 the phylogenetic analysis, analysed and interpreted data and critically revised the
332 manuscript. J.R.T. ran the phylogenetic analyses, analysed and interpreted data and
333 critically revised the manuscript. A.P.G. conceived and carried out the elemental
334 analysis, analysed and interpreted data, and critically revised the manuscript. All authors
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337

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357

358 **References**

- 359 1. Ewin DH. 2020 The origin of animal body plans: a view from fossil evidence
360 and the gene regulatory genome. *Development* **147**, dev182899.
361 (doi:10.1242/dev.182899)
- 362 2. Murdock DJE. 2020 The “biomineralization toolkit” and the origin of animal
363 skeletons. *Biol. Rev.* **95**, 1372–1392. (doi:10.1111/brv.12614)
- 364 3. Wood R, Ivantsov AY, Zhuravlev AY. 2017 First macrobiota biomineralization
365 was environmentally triggered. *Proc. R. Soc. B* **284**, 20170059.
366 (doi:10.1098/rspb.2017.0059)
- 367 4. Wood R. 2018 Exploring the drivers of early biomineralization. *Emerg. Top.*
368 *Life Sci.* **2**, 201–212. (doi:10.1042/ETLS20170164)
- 369 5. Nielsen C. 2011 Animal Evolution: Interrelationships of the Living Phyla.
370 Oxford University Press, 416 pp.

- 371 6. Cary GA, Hinman VF. 2017 Echinoderm development and evolution in the post-
372 genomic era. *Dev. Biol.* **427**, 203–211. (doi:10.1016/j.ydbio.2017.02.003)
- 373 7. Guensburg TE, Sprinkle J. 2009 Solving the mystery of crinoid ancestry: new
374 fossil evidence of arm origin and development. *J. Paleontol.* **83**, 350–364.
375 (doi:10.1666/08-090.1)
- 376 8. Smith AB, Reich M. 2013 Tracing the evolution of the holothurian body plan
377 through stem group fossils. *Biol. J. Linn. Soc.* **109**, 670–681.
378 (doi:10.1111/bij.12073)
- 379 9. Gao F, Thompson JR, Petsios E, Erkenbrack E, Moats RA, Bottjer DJ, Davidson
380 EH, 2015 Juvenile skeletogenesis in anciently diverged sea urchin clades. *Dev.*
381 *Biol.* **400**, 148–158. (doi:10.1016/j.ydbio.2015.01.017)
- 382 10. Dylus DV, Czarkwiani A, Blowes LM, Elphick MR, Oliveri P. 2018
383 Developmental transcriptomics of the brittle star *Amphiura filiformis* reveals
384 gene regulatory network rewiring in echinoderm larval skeleton evolution.
385 *Genome Biol.* **19**, 26. (doi:10.1186/s13059-018-1402-8).
- 386 11. Smirnov AV. 2016. Parallelisms in the evolution of sea cucumbers
387 (Echinodermata: Holothuroidea). *Paleontol. J.* **50**, 1610–1625.
- 388 12. Kammer TW, Ausich WI. 2007 Soft-tissue preservation of the hind gut in a new
389 genus of cladid crinoid from the Mississippian (Visean, Asbian) at St Andrews,
390 Scotland. *Palaeontology* **50**, 951–959. (doi:10.1111/j.1475-4983.2007.00687.x)
- 391 13. Thomas RDK, Runnegar B, Matt K. 2020 *Pelagiella exigua*, an early Cambrian
392 stem gastropod with chaetae: lophotrochozoan heritage and conchiferan novelty.
393 *Palaeontology* **63**, 601–627. (doi:10.1111/j.1475-4983.2007.00687.x)

- 394 14. Sundberg FA, Geyer G, Kruse PD, McCollum LB, Pegel TV, Żylińska A,
395 Zhuravlev AY. 2016 International correlation of the Cambrian Series 2-3, Stages
396 4-5 boundary interval. *Australas. Palaeontol. Mem.* **49**, 83–124.
- 397 15. Geyer G. 2019 A comprehensive Cambrian correlation chart. *Episodes* **42**, 321–
398 332. (doi:10.18814/epiiugs/2019/019026)
- 399 16. Zamora S, Rahman IA, Smith AB. 2012 Plated Cambrian Bilaterians Reveal the
400 Earliest Stages of Echinoderm Evolution. *PLoS One* **7**, e38296.
401 (doi:10.1371/journal.pone.0038296)
- 402 17. Smith AB, Zamora S. 2013 Cambrian spiral-plated echinoderms from
403 Gondwana reveal the earliest pentaradial body plan. *Proc. R. Soc. B* **280**
404 20131197. (doi:10.1098/rspb.2013.1197)
- 405 18. Topper TP, Guo J, Clausen S, Skovsted CB, Zhang Z. 2019 A stem group
406 echinoderm from the basal Cambrian of China and the origins of Ambulacraria.
407 *Nat. Commun.* **10**, 1366. (doi:10.1038/s41467-019-09059-3)
- 408 19. Nanglu K, Caron JB, Cameron CB. 2020 Cambrian tentaculate worms and the
409 origin of the hemichordate body plan. *Curr. Biol.* **30**, 4238–4244.e1.
410 (doi:10.1016/j.cub.2020.07.078)
- 411 20. Zamora S, Wright DF, Mooi R, Lefebvre B, Guensburg TE, Gorzelak P, David
412 B, Sumrall CD, Cole SR, Hunter AW, Sprinkle J, Thompson JR, Ewin TAM,
413 Fatka O, Nardin E, Reich M, Nohejlová M, Rahman IA. 2020 Re-evaluating the
414 phylogenetic position of the enigmatic early Cambrian deuterostome
415 *Yanjiahella*. *Nat. Commun.* **11**, 1286. (doi:10.1038/s41467-019-09059-3)
- 416 21. Hunter AW, Ortega-Hernández J. 2021 A new somasteroid from the Fezouata
417 Lagerstätte in Morocco and the Early Ordovician origin of Asterozoa. *Biol. Lett.*
418 **17**, 20200809. (doi:10.1098/rsbl.2020.0809)

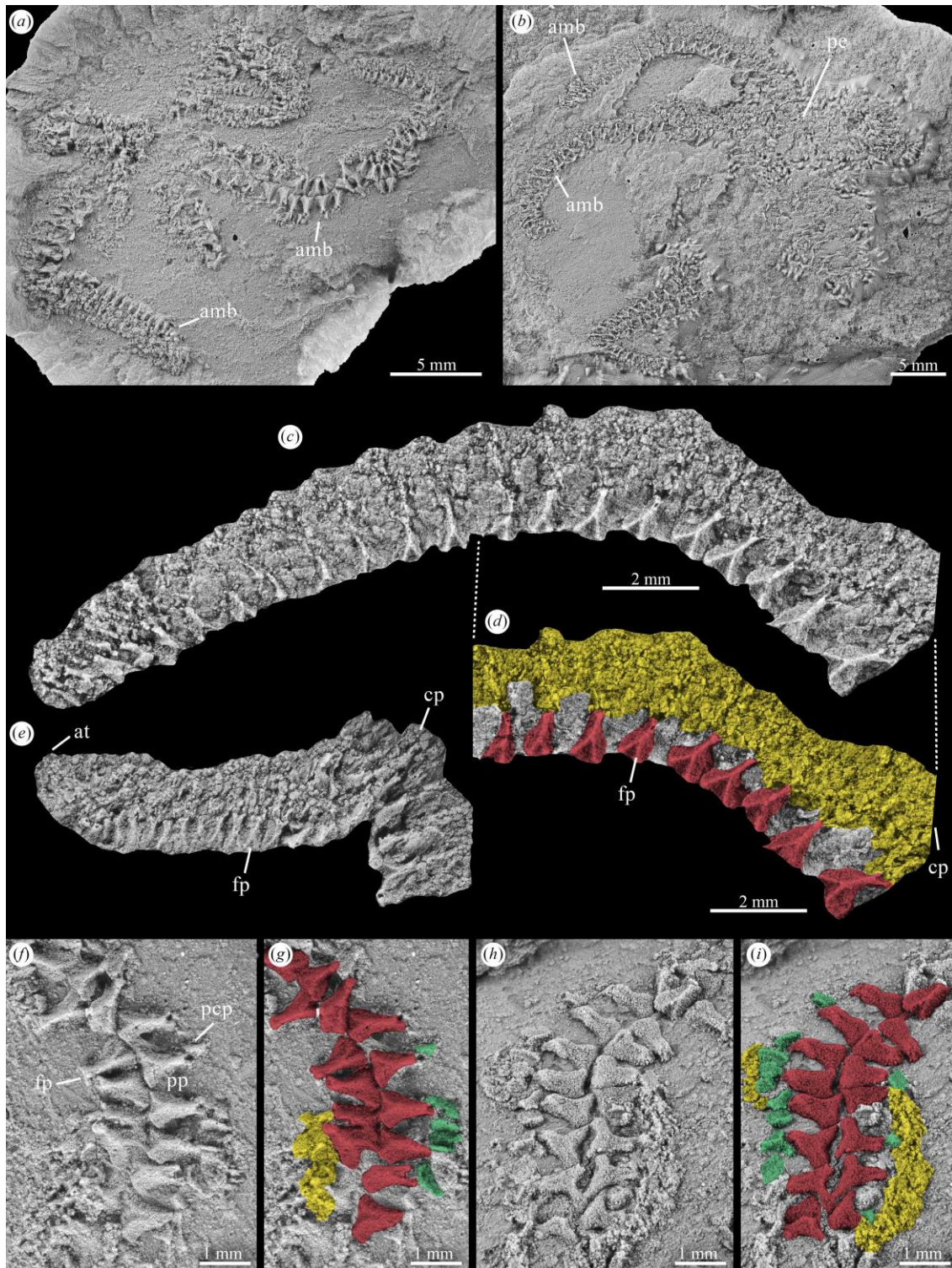
- 419 22. Zamora S, Rahman IA. 2014 Deciphering the early evolution of echinoderms
420 with Cambrian fossils. *Palaeontology* **57**, 1105–1119. (doi:10.1111/pala.12138)
- 421 23. Sumrall CD, Zamora S. 2011 Ordovician edrioasteroids from Morocco: Faunal
422 exchanges across the Rheic Ocean. *J. Syst. Palaeontol.* **9**, 425–454.
423 (doi:10.1080/14772019.2010.499137)
- 424 24. Zamora S, Smith AB. 2012 Cambrian stalked echinoderms show unexpected
425 plasticity of arm construction. *Proc. R. Soc. B* **279**, 293–298.
426 (doi:10.1098/rspb.2011.0777)
- 427 25. Nohejlová M, Nardin E, Fatka O, Kašička L, Szabad M. 2019 Morphology,
428 palaeoecology and phylogenetic interpretation of the Cambrian echinoderm
429 *Vyscystis* (Barrandian area, Czech Republic). *J. Syst. Palaeontol.* **17**, 1619–
430 1634. (doi:10.1080/14772019.2018.1541485)
- 431 26. Swofford DL. 2002 PAUP*. *Phylogenetic analysis using parsimony (*and other*
432 *methods)*. Version 4. Sunderland, MA: Sinauer Associates.
- 433 27. Ronquist F, Teslenko M, van der Mark P, Ayres DL, Darling A, Höhna S,
434 Larget B, Liu L, Suchard MA, Huelsenbeck, JP. 2012 MrBayes 3.2: efficient
435 Bayesian phylogenetic inference and model choice across a large model space.
436 *Syst. Biol.* **61**, 539–542. (doi:10.1093/sysbio/sys029)
- 437 28. Lewis PO. 2001 A likelihood approach to estimating phylogeny from discrete
438 morphological character data. *Syst. Biol.* **50**, 913–925
439 (doi:10.1080/106351501753462876)
- 440 29. Mooi R, David B, Marchand D. 1994 Echinoderm skeletal homologies: classical
441 morphology meets modern phylogenetics. In *Echinoderms through time* (eds B
442 David, A Guille, JP Féral, M Roux), pp. 87–95. Rotterdam, The Netherlands:
443 Balkema.

- 444 30. David B, Mooi R. 1998 Major events in the evolution of echinoderms viewed by
445 the light of embryology. In *Echinoderms: San Francisco* (eds R Mooi, M.
446 Telford), pp. 21–28. Rotterdam, The Netherlands: Balkema.
- 447 31. Bell BM. 1976 A study of North American Edrioasteroidea. *N. Y. State Mus.*
448 *Mem.* **21**, 446 pp.
- 449 32. Smith AB. 1985 Cambrian eleutherozoan echinoderms and the early
450 diversification of edrioasteroids. *Palaeontology* **28**, 715–756.
- 451 33. Gaines RR, Briggs DEG, Yuanlong Z. 2008 Cambrian Burgess Shale-type
452 deposits share a common mode of fossilization. *Geology* **36**, 755–758.
453 (doi:10.1130/G24961A.1)
- 454 34. Butterfield NJ. 1995 Secular distribution of Burgess-Shale-type preservation.
455 *Lethaia* **28**, 1–13. (doi:10.1111/j.1502-3931.1995.tb01587.x)
- 456 35. Walcott CD. 1911 Middle Cambrian holothurians and medusa. *Smithson. Misc.*
457 *Collect.* **57**, 41–68.
- 458 36. Gehling JG. 1987 Earliest known echinoderm — a new Ediacaran fossil from
459 the Pound Subgroup of South Australia. *Alcheringa* **11**, 337–345.
- 460 37. Shu D-G, Conway Morris S, Han J, Zhang Z-F, Liu J-N. 2004 Ancestral
461 echinoderms from the Chengjiang deposits of China. *Nature* **430**, 422–428.
462 (doi:10.1038/nature02648)
- 463 38. Swalla BJ, Smith AB. 2008 Deciphering deuterostome phylogeny: molecular,
464 morphological and palaeontological perspectives. *Phil. Trans. R. Soc. B* **363**,
465 1557–1568. (doi:10.1098/rstb.2007.2246)
- 466 39. Clausen S, Hou X-G, Bergström J, Franzén C. 2010 The absence of
467 echinoderms from the Lower Cambrian Chengjiang fauna of China:

- 468 palaeoecological and palaeogeographical implications. *Palaeogeogr.*
469 *Palaeoclimatol. Palaeoecol.* **294**, 133–141. (doi:10.1016/j.palaeo.2010.01.001)
- 470 40. Topper TP, Guo J, Clausen S, Skovsted CB, Zhang Z. 2020 Reply to ‘Re-
471 evaluating the phylogenetic position of the enigmatic early Cambrian
472 deuterostome *Yanjiahella*’. *Nat. Commun.* **11**, 1287. (doi:10.1038/s41467-020-
473 14922-9)
- 474 41. Guensburg TE, Sprinkle J, Mooi R, Lefebvre B, David B, Roux M, Derstler K.
475 2020 *Athenacrinus* n. gen. and other early echinoderm taxa inform crinoid origin
476 and arm evolution. *J. Paleontol.* **94**, 311–333. (doi:10.1017/jpa.2019.87)
- 477 42. Zhao Y, Sumrall CD, Parsley RL, Peng J. 2010 *Kailidiscus*, a new
478 plesiomorphic edrioasteroid from the basal Middle Cambrian Kaili Biota of
479 Guizhou Province, China. *J. Paleontol.*, **84**, 668–680.
480 (doi:10.1017/S0022336000058388)
- 481 43. Rahman IA, Thompson JR, Briggs DEG, Siveter DJ, Siveter DJ, Sutton MD.
482 2019 A new ophiocistoid with soft-tissue preservation from the Silurian
483 Herefordshire Lagerstätte, and the evolution of the holothurian body plan. *Proc.*
484 *R. Soc. B* **286**, 20182792. (doi:10.1098/rspb.2018.2792)
- 485 44. Martins L, Souto C. 2020 Taxonomy of the Brazilian Apodida (Holothuroidea),
486 with the description of two new genera. *Marine Biology Research* **16**, 219–255.
487 (doi: 10.1080/17451000.2020.1761027)
- 488 45. Gao F, Davidson EH. 2008 Transfer of a large gene regulatory apparatus to a
489 new developmental address in echinoid evolution. *Proc. Natl. Acad. Sci.*
490 *U.S.A.* **105**, 6091–6096. (doi:10.1073/pnas.0801201105)

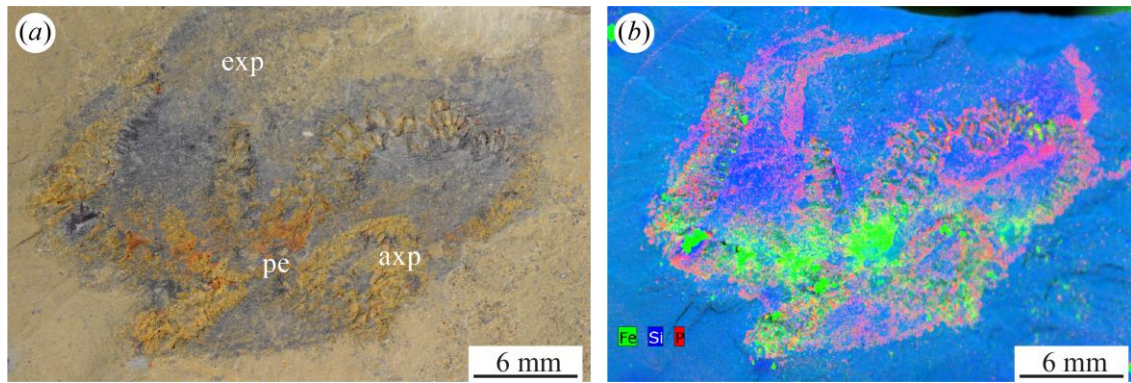
- 491 46. Czarkwiani A, Dylus DV, Carballo L, Oliveri P. 2019 FGF signalling plays
492 similar roles in development and regeneration of the skeleton in the brittle star
493 *Amphiura filiformis*. *Development* **148**, dev180760. (doi:10.1242/dev.180760)
- 494 47. Duloquin L, Lhomond G, Gache C. 2007 Localized VEGF signaling from
495 ectoderm to mesenchyme cells controls morphogenesis of the sea urchin embryo
496 skeleton. *Development* **134**, 2293–2302. (doi:10.1242/dev.005108)
- 497 48. Morgulis M, Gildor T, Roopin M, Sher N, Malik A, Lalar M, Dines M, de-
498 Leon SBT, Khalaily L, de-Leon SBT. 2019 Possible cooption of a VEGF-driven
499 tubulogenesis program for biomineralization in echinoderms. *Proc. Natl. Acad.*
500 *Sci. U.S.A.* **116**, 12353–12362. (doi:10.1073/pnas.1902126116)
- 501 49. Mooi R, David B, Wray GA. 2005 Arrays in rays: terminal addition in
502 echinoderms and its correlation with gene expression. *Evol. Dev.* **7**, 542–555.
503 (doi:10.1111/j.1525-142X.2005.05058.x)
- 504 50. Wessel GM, Kiyomoto M, Shen T-L, Yajima, M. 2020 Genetic manipulation of
505 the pigment pathway in a sea urchin reveals distinct lineage commitment prior to
506 metamorphosis in the bilateral to radial body plan transition. *Sci. Rep.* **10**, 1973.
507 (doi:10.1038/s41598-020-58584-5)
- 508 51. Thompson JR, Paganos P, Benvenuto G, Arnone MI, Oliveri P. 2021 Post-
509 metamorphic skeletal growth in the sea urchin *Paracentrotus lividus* and
510 implications for body plan evolution. *EvoDevo* **12**, 3. (doi:10.1186/s13227-021-
511 00174-1)
- 512 52. Smith AB, Zamora S, Álvaro JJ. 2013. The oldest echinoderm faunas from
513 Gondwana show that echinoderm body plan diversification was rapid. *Nat.*
514 *Commun.* **4**, 1385. (doi:10.1038/ncomms2391)

- 515 53. Kammer TW, Sumrall CD, Ausich WI, Deline B, Zamora S. 2013 Oral region
516 homologies in Paleozoic crinoids and other plesiomorphic pentaradial
517 echinoderms. *PLoS One* **8**, e77989. (doi:10.1371/journal.pone.0077989)
- 518 54. Zamora S, Sumrall CD, Vizcaíno D. 2013 Morphology and ontogeny of the
519 Cambrian edrioasteroid echinoderm *Cambraster cannati* from western
520 Gondwana. *Acta Palaeontol. Pol.* **58**, 545–559. (doi:10.4202/app.2011.0152)
- 521 55. Zamora S, Lefebvre B, Hosgör I, Franzen C, Nardin E, Fatka O, Álvaro JJ. 2015
522 The Cambrian edrioasteroid *Stromatocystites* (Echinodermata): systematics,
523 palaeogeography and palaeoecology. *Geobios* **48**, 417–426.
524 (doi:10.1016/j.geobios.2015.07.004)
- 525 56. Sprinkle J, Sumrall CD. 2015 New edrioasterine and astrocystitid
526 (Echinodermata: Edrioasteroidea) from the Ninemile Shale (Lower Ordovician),
527 central Nevada. *J. Paleontol.* **89**, 346–352. (doi:10.1017/jpa.2014.29)
- 528
529
- 530 **Figure captions**



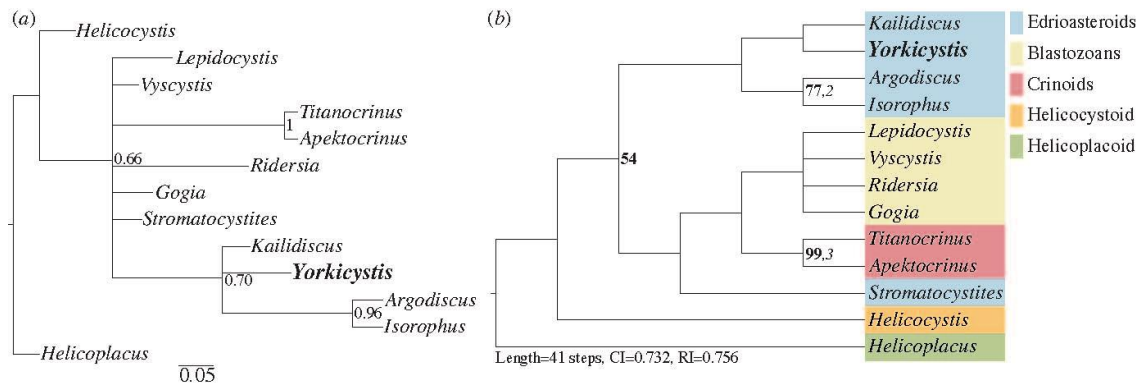
531
 532 **Figure 1.** *Yorkicystis haefneri* from the early Cambrian Kinzers Formation (York,
 533 Pennsylvania). (a) NHMUK EE 1659, holotype complete specimen. (b) NHMUK EE
 534 1660, paratype complete specimen. (c,d) NHMUK EE 1660, ambulatory region in lateral
 535 view. (e) NHMUK EE 1659, ambulatory region in lateral view. (f–i). NHMUK EE 1659,
 536 ambulatory region in plan view, part (f,g) and counterpart (h,i). Colours: red, flooring

537 plates; green, [primary](#) of cover plates; yellow, cover plates. Abbreviations: amb,
538 ambulacrum; at, ambulacral tip; cp, cover plate; fp, flooring plate; pcp, primary
539 cover plate; pe, peristome; pp, podial pore. All images are photographs of latex casts
540 whitened with NH₄Cl sublimate.



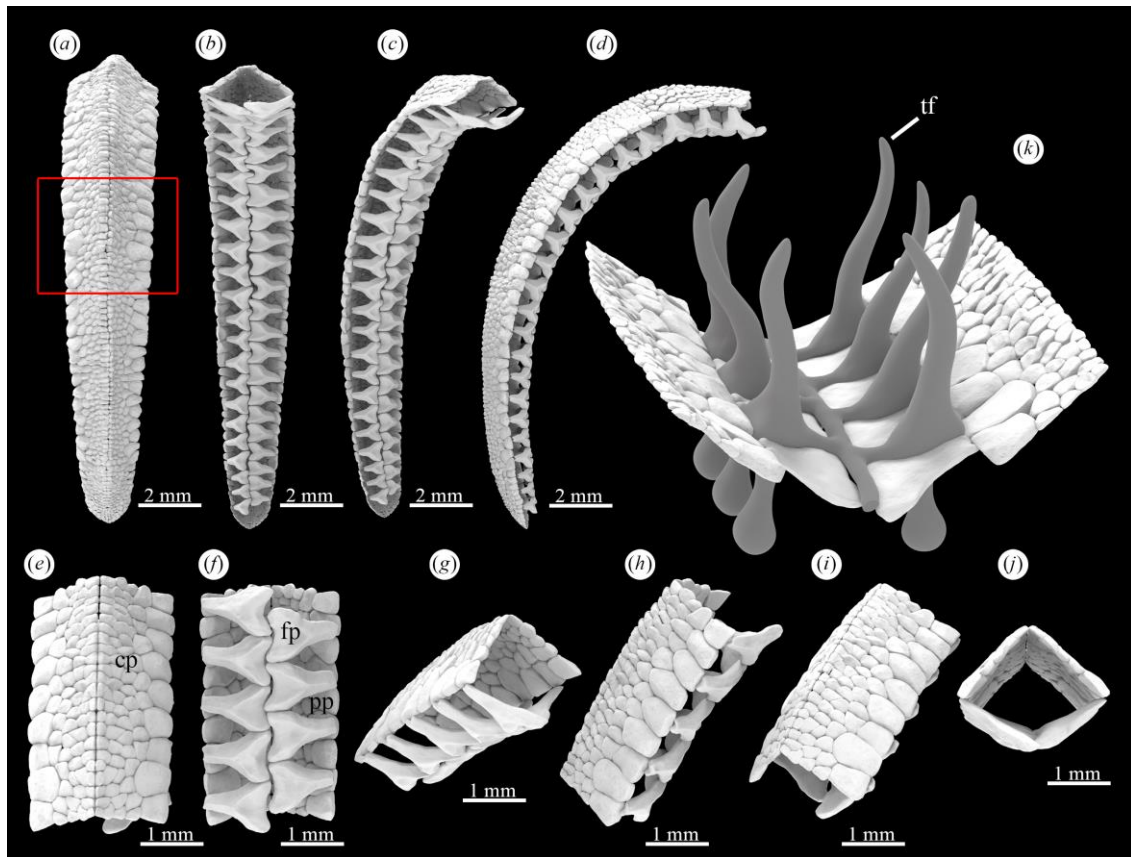
541

542 **Figure 2.** *Yorkicystis haefneri* from the early Cambrian Kinzers Formation (York,
543 Pennsylvania). (NHMUK EE 1659). (a) Photograph of specimen submerged in
544 water. (b) False-colour element map generated using X-ray fluorescence analysis
545 showing Fe (green), Si (blue) and P (red); intensity of colour corresponds to element
546 intensity. Abbreviations: axp, axial part; exp, extraxial part; pe, peristome.



547

548 **Figure 3.** Phylogenetic position of *Yorkicystis*. (a) 50% majority-rule consensus tree
 549 resulting from Bayesian analyses. Numbers next to nodes represent the proportion of
 550 trees in the post-burn in posterior sample that contained that node. Analysis performed
 551 with a symmetric Dirichlet hyperprior set to ∞ , corresponding to symmetric character
 552 transition rates. Branch lengths represent the number of expected changes per character.
 553 (b) Strict consensus of the four most parsimonious trees resulting from parsimony
 554 analyses. Bootstrap support (BS; bold) and decay indices (DI; italics) are shown for
 555 each node with BS > 50 and DI > 0. Both trees are rooted on *Helicoplacus*.



556

557 **Figure 4.** Digital reconstruction of the ambulacral construction of *Yorkicystis*
 558 *haefneri*. (a–d) Single ambulacrum in different views. (e–j) Detail of part of the
 559 ambulacrum in different views. (k) Inferred soft tissues of the water vascular system
 560 housed within the ambulacrum during life. Abbreviations: cp, cover plate; fp,
 561 flooring plate; pp, podial pore; tf, tube foot.



562
563 **Figure 5.** Life reconstruction of *Yorkicystis haefneri*.

564

565

566 **Electronic supplementary material**

567 **Supplementary Information.** The supplementary information of this paper includes
568 taphonomic observations, additional results from X-ray fluorescence analysis, a full
569 description of the fossils and further details on the phylogenetic analyses.

570 Supplementary table 1 includes character states for all taxa included in the phylogenetic
571 analyses.