

## Distinct features in circulating metabolome associate with white matter hyperintensities in females and males

Eeva Sliz<sup>1</sup>, Jean Shin<sup>1</sup>, Shahzad Ahmad<sup>2,3</sup>, Dylan M. Williams<sup>4,5</sup>, Stefan Frenzel<sup>6</sup>, Friederike Gauß<sup>7</sup>, Sarah E. Harris<sup>8</sup>, Ann-Kristin Henning<sup>7</sup>, Maria Valdes Hernandez<sup>9</sup>, Yi-Han Hu<sup>10</sup>, Beatriz Jiménez<sup>11</sup>, Muralidharan Sargurupremraj, Carole Sudre<sup>4,12,13</sup>, Ruiqi Wang<sup>14</sup>, Katharina Wittfeld<sup>6,15</sup>, Qiong Yang<sup>14</sup>, Joanna M. Wardlaw<sup>9</sup>, Henry Völzke<sup>16</sup>, Meike W. Vernooij<sup>2,17</sup>, Jonathan M Schott<sup>18</sup>, Marcus Richards<sup>4</sup>, Petroula Proitsi<sup>19</sup>, Matthias Nauck<sup>7</sup>, Matthew R. Lewis<sup>11</sup>, Lenore Launer<sup>10</sup>, Norbert Hosten<sup>20</sup>, Hans Grabe<sup>6,15</sup>, Mohsen Ghanbari<sup>2</sup>, Ian J. Deary<sup>8</sup>, Simon R. Cox<sup>8</sup>, Nishi Chaturvedi<sup>4</sup>, Josephine Barnes<sup>21</sup>, Jerome I. Rotter<sup>22</sup>, Stephanie Debette, M. Arfan Ikram<sup>2</sup>, Myriam Fornage<sup>23</sup>, Tomas Paus<sup>24,25</sup>, Sudha Seshadri<sup>26,27</sup>, and Zdenka Pausova<sup>1</sup>, for the NeuroCHARGE Working Group

<sup>1</sup> The Hospital for Sick Children, and Departments of Physiology and Nutritional Sciences, University of Toronto, Toronto, ON, Canada

<sup>2</sup> Department of Epidemiology, Erasmus Medical Centre, Rotterdam, The Netherlands

<sup>3</sup> Division of Systems Biomedicine and Pharmacology, Leiden Academic Centre for Drug Research, Leiden University, Leiden, The Netherlands

<sup>4</sup> MRC Unit for Lifelong Health and Ageing at UCL, University College London, London, UK

<sup>5</sup> Department of Medical Epidemiology and Biostatistics, Karolinska Institutet, Stockholm, Sweden

<sup>6</sup> Department of Psychiatry and Psychotherapy, University Medicine Greifswald, Greifswald, Germany

<sup>7</sup> Institute of Clinical Chemistry and Laboratory Medicine, University Medicine Greifswald, Greifswald, Germany

<sup>8</sup> Lothian Birth Cohorts group, Department of Psychology, University of Edinburgh, Edinburgh, UK

<sup>9</sup> Centre for Clinical Brain Sciences, UK Dementia Research Institute at the University of Edinburgh, Edinburgh, UK

<sup>10</sup> Laboratory of Epidemiology and Population Sciences, National Institute on Aging, Baltimore, MD, USA

<sup>11</sup> National Phenome Centre, Department of Metabolism, Digestion and Reproduction, Imperial College London, London, UK

<sup>12</sup> Centre for Medical Image Computing, Department of Computer Science, University College London

<sup>13</sup> School of Biomedical Engineering & Imaging Sciences, King's College London

<sup>14</sup> Department of Biostatistics, Boston University, Boston, MA, USA

<sup>15</sup> Germany Center for Neurodegenerative Diseases (DZNE), partner site Rostock/Greifswald, Greifswald, Germany

<sup>16</sup> Institute for Community Medicine, University Medicine Greifswald, Greifswald, Germany

<sup>17</sup> Department of Radiology and Nuclear Medicine, and Department of Neurology, Erasmus Medical Center, Rotterdam, the Netherlands

<sup>18</sup> Dementia Research Centre, UCL Queen Square Institute of Neurology, University College London, London, UK

<sup>19</sup> King's College London, Institute of Psychiatry, Psychology and Neuroscience, London, UK

<sup>20</sup> Institute of Diagnostic Radiology and Neuroradiology, University Medicine Greifswald, Greifswald, Germany

<sup>21</sup> Dementia Research Centre, UCL Queen Square Institute of Neurology, University College London, London, United Kingdom

<sup>22</sup> The Institute for Translational Genomics and Population Sciences, Department of Pediatrics, The Lundquist Institute for Biomedical Innovation at Harbor-UCLA Medical Center, Torrance, CA USA

<sup>23</sup> University of Texas Health Science Center at Houston McGovern Medical School, Houston, TX, USA

<sup>24</sup> ECOGENE-21, Chicoutimi, QC, Canada

<sup>25</sup> Departments of Psychology and Psychiatry, University of Toronto, Toronto, ON, Canada

<sup>26</sup> The Framingham Heart Study, Framingham, MA, USA

<sup>27</sup> Department of Neurology, Boston University School of Medicine, Boston, MA, USA

Corresponding author:

Zdenka Pausova, MD, FAHA

Senior Scientist, Hospital for Sick Children

Professor, Departments of Physiology and Nutritional Sciences

University of Toronto

Peter Gilgan Centre for Research and Learning

686 Bay Street, Rm. 10-9705

Toronto, ON M5G 0A4

Canada

Phone: (416) 813-7654/304340

Fax: (416) 813-5771

E-mail: zdenka.pausova@sickkids.ca

Keywords: White matter hyperintensities; Brain health; Metabolomics; Lipidomics; Hydroxyphenylpyruvate; Glucuronate; Lipid ratios

53 **ABSTRACT**

54 **Background:**

55 White matter hyperintensities (WMH) are identified on T2-weighted magnetic resonance images of the  
56 human brain as areas of enhanced brightness; WMH are a major risk factor of stroke, dementia, and  
57 death. Currently, there are no large-scale studies testing associations between WMH and circulating  
58 metabolites.

59 **Methods:**

60 We studied up to 9,290 individuals (50.7% females, average age 61 years) from 15 populations of 8  
61 community-based cohorts. WMH volume was quantified from T2-weighted or fluid-attenuated  
62 inversion-recovery images or as hypointensities on T1-weighted images. Circulating metabolomic  
63 measures were assessed with mass spectrometry and nuclear magnetic resonance spectroscopy.  
64 Associations between WMH and metabolomic measures were tested by fitting linear regression models  
65 in the pooled sample, and in sex-stratified and statin treatment-stratified subsamples. Our basic models  
66 were adjusted for age, sex, age\*sex, and technical covariates, and our fully adjusted models were  
67 additionally adjusted for statin treatment, hypertension, type 2 diabetes, smoking, body mass index, and  
68 estimated glomerular filtration rate. Population-specific results were meta-analyzed using the fixed-  
69 effect inverse variance-weighted method. Associations with false discovery rate (FDR)-adjusted  $p$ -  
70 values ( $p_{\text{FDR}} < 0.05$ ) were considered significant.

71 **Results:**

72 In the meta-analysis of results from the basic models, we identified 30 metabolomic measures  
73 associated with WMH ( $p_{\text{FDR}} < 0.05$ ), 7 of which remained significant in the fully adjusted models. The  
74 most significant association was with higher level of hydroxyphenylpyruvate in males  
75 ( $p_{\text{FDR,full,adj}} = 1.40 \times 10^{-7}$ ) and in both the pooled sample ( $p_{\text{FDR,full,adj}} = 1.66 \times 10^{-4}$ ) and statin-nontreated  
76 ( $p_{\text{FDR,full,adj}} = 1.65 \times 10^{-6}$ ) subsample. **In males, HPP explained 14% of variance in WMH.** In males and the  
77 pooled sample, WMH were also associated with lower levels of lysophosphatidylcholines and  
78 hydroxysphingomyelins, and a larger diameter of low-density lipoprotein particles, likely arising from  
79 higher triglyceride-to-total-lipids and lower cholesteryl ester-to-total-lipids ratios within these particles.  
80 In females, the only significant association was with higher level of glucuronate ( $p_{\text{FDR}} = 0.047$ ).

81 **Conclusions:**

82 **Circulating metabolomic measures, including multiple lipid measures (e.g., lysophosphatidylcholines,**  
83 **hydroxysphingomyelins, low-density lipoprotein size and composition) and non-lipid metabolites (e.g.,**  
84 **hydroxyphenylpyruvate, glucuronate) associate with WMH in a general population of middle-aged and**  
85 **older adults. Some metabolomic associations show marked sex specificities and explain sizable**  
86 **proportion of WMH variance.**

**Commented [ZP1]:** Results from an additional cohort will be incorporated later this week.

87 **CLINICAL PERSPECTIVE**

88 **What is new?**

- 89
- 90
- 91
- 92
- 93
- 94
- 95
- 96
- 97
- 98
- This is the first large-scale study to identify circulating metabolomic measures associated with white matter hyperintensities (WMH) volume, which is the most common brain-imaging marker of small vessel disease and a major risk factor for incident stroke, dementia, and all-cause mortality.
  - The metabolomic profile of WMH volume appears largely sex specific.
  - The most significant metabolite, hydroxyphenylpyruvate, explains 6% and 14% of variance in WMH volume in the pooled sample and in males, respectively, which is comparatively more than the proportions of variance explained by hypertension (1%), type 2 diabetes (1%) or smoking (0.1%).

99 **What are the clinical implications?**

- 100
- 101
- 102
- 103
- 104
- 105
- 106
- 107
- Hydroxyphenylpyruvate is a new potentially useful clinical biomarker explaining more variance in WMH volume than the established WMH-risk factors.
  - The marked sex specificities in the metabolomic associations of WMH volume add to the growing body of research suggesting that sex-specific molecular pathways underlie the associations between risk factors and cerebrovascular outcomes.
  - Some associated metabolites support vessel-related pathophysiology of WMH, while others suggest the involvement of myelin disruption and neuron injury.

108 **INTRODUCTION**

109 White matter hyperintensities (WMH) are among the most commonly encountered signal alterations on  
110 brain magnetic resonance imaging (MRI) images – they are areas of high intensity in the periventricular  
111 and deep cerebral white matter on T2-weighted (T2W) or T2 fluid-attenuated inversion recovery  
112 (FLAIR) images. The exact neuropathological mechanisms are not fully understood, but the proposed  
113 fundamental features of WMH are loss of myelin and axons, and mild gliosis (reviewed in<sup>1,2</sup>). These  
114 features might arise due to chronic ischemia and dysfunction of the blood-brain barrier (BBB) related  
115 to cerebral small vessel disease<sup>2</sup>.

116 Evidence from a recent meta-analysis of prospective, longitudinal cohort studies indicates that WMH  
117 are of major clinical significance<sup>3</sup>: the results obtained in 14,529 participants show that higher volume  
118 of WMH increases the risk of incident stroke (hazard ratio [HR] and corresponding 95% confidence  
119 interval=2.45 [1.93-3.12]), intracerebral hemorrhage (HR=3.17 [1.54-6.52]), ischemic stroke (HR=2.39  
120 [1.65-3.74]), dementia (HR=1.84 [1.40-2.43]), Alzheimer's disease (HR=1.50 [1.22-1.84]), and all-  
121 cause mortality (HR=2.00 [1.69-2.36])<sup>3</sup>. WMH are common in the general population: the prevalence  
122 is ~20% at the age of 60 years and >90% at the age of 80 years and higher<sup>4</sup>. Hypertension, type 2  
123 diabetes, and smoking are the key cardiometabolic risk factors for WMH<sup>5-7</sup>.

124 We and others have shown that aberrations in the blood metabolome are associated with brain health:  
125 circulating levels of multiple lipid measures are associated with Alzheimer's disease<sup>8,9</sup> and cognitive  
126 functioning<sup>10</sup>, as well as structural properties of the brain, such as T1W signal intensity of white  
127 matter<sup>11,12</sup> and thickness of the cerebral cortex<sup>13</sup>. The metabolomic associations of WMH, however, are  
128 incompletely understood and, to date, only two small studies have been published<sup>14,15</sup>. Here we used  
129 metabolomics technologies to provide a comprehensive characterization of variations in circulating  
130 metabolomic measures as a function of WMH volume assessed in general population. We further  
131 explored whether these associations are independent of the key risk factors of WMH, including  
132 hypertension, type 2 diabetes, and smoking.

133 **METHODS**

134 **Study populations**

135 This is a large collaborative work incorporating data from 8 population-based cohort studies: Age,  
136 Gene/Environment Susceptibility-Reykjavik Study<sup>16</sup>, Framingham Heart Study<sup>17,18</sup>, Insight 46 (a sub-  
137 study of the British 1946 birth cohort)<sup>19</sup>, Lothian Birth Cohort 1936<sup>20,21</sup>, Rotterdam Study<sup>22-24</sup>, Saguenay  
138 Youth Study<sup>25</sup>, Study of Health in Pomerania<sup>26,27</sup>, and the Southall And Brent Revisited study<sup>28</sup>. Study-  
139 specific descriptions are given in the [Supplemental Methods](#). After excluding individuals with  
140 dementia, stroke, multiple sclerosis, brain surgery, or gross morphological abnormalities of the brain  
141 (e.g., cysts, brain tumors), and individuals with poor quality of MRI scans, we analyzed up to 9,290  
142 individuals of mostly European ancestries with brain MRI and blood metabolomic data. All cohort  
143 studies were approved by local ethics committees, and all participants have provided their written  
144 informed consent ([Supplemental Methods](#)).

145 **Phenotype quantifications**

146 *Brain MRI and WMH assessment*

147 The brain MRI was carried out in each cohort study separately, and the details are provided in [Table](#)  
148 [S1](#). In the present analyses, the total volume of WMH (or hypointensities on T1W images) was  
149 considered as a continuous variable.

150 *Metabolomic quantifications*

151 We used nuclear magnetic resonance (NMR)-based techniques (Nightingale Health Ltd, Helsinki,  
152 Finland; National Phenome Centre, London, UK; Bruker Biospin, Rheinstetten, Germany) and mass  
153 spectrometry (MS)-based technologies (Metabolon, Morrisville, North Carolina, USA; Biocrates Life  
154 Sciences AG, Innsbruck, Austria; Broad Institute, Cambridge, Massachusetts, USA) that are commonly  
155 employed in epidemiological research and are described elsewhere<sup>29-33</sup> (Table S1). Using these  
156 aforementioned platforms, it was possible to measure a total of 2,217 different metabolomic measures  
157 covering a broad spectrum of circulating lipids and non-lipid metabolomic measures. Of these, 1,174  
158 metabolomic measures were quantified in two or more of the study populations. **We did not include**  
159 **unknown metabolites in our study.** Metabolomic quantifications were completed using serum or plasma  
160 samples, typically extracted from blood samples drawn after overnight fasting (Table S1). Blood  
161 samples were drawn before or at the time of MRI in all cohorts except for the Rotterdam Study, in  
162 which MRI scans have been conducted at multiple time-points, and metabolomic data that were from  
163 the time-point that was closest to the MRI scans were always analyzed (Table S2).

164 **Statistical analyses**

165 *Linear regression models*

166 The cross-sectional associations between log-transformed WMH and circulating metabolomic  
167 measures were studied using linear regression. **Prior to model fitting, the log-transformed WMH and**  
168 **metabolomic measures were scaled to standard deviation (SD) units, which enables the comparison and**  
169 **meta-analysis of data in different units and varying numerical ranges that originate from the multiple**  
170 **quantification methods used (Table S1).** We fitted the regression models in five analytical samples (*i.e.*,  
171 pooled sample and sex- or statin treatment-stratified subsamples) using a simplistic covariate structure  
172 (basic models) and a more complete covariate structure (fully adjusted models) to test if the associations  
173 are independent of key risk factors. The study models were as follows:

174 *Pooled sample*

175 basic models:  $\log\text{WMH} \sim \text{metabolic measure} + \text{age} + \text{sex} + \text{age}*\text{sex} + \text{time (if applicable)} +$   
176  $\text{fasting duration (if applicable)} + \text{intracranial volume or brain size} + \text{cohort-specific covariates}$   
177 fully adjusted models: as above + statin treatment + hypertension + type 2 diabetes + BMI +  
178 eGFR + current smoking status

179 *Sex-stratified subsamples*

180 basic models:  $\log\text{WMH} \sim \text{metabolic measure} + \text{age} + \text{time (if applicable)} + \text{fasting duration (if}$   
181  $\text{applicable}) + \text{intracranial volume or brain size} + \text{cohort-specific covariates}$   
182 fully adjusted models: as above + statin treatment + hypertension + type 2 diabetes + BMI +  
183 eGFR + current smoking status

184 *Statin treatment-stratified subsamples*

185 basic models:  $\log\text{WMH} \sim \text{metabolic measure} + \text{age} + \text{sex} + \text{age}*\text{sex} + \text{time (if applicable)} +$   
186  $\text{fasting duration (if applicable)} + \text{intracranial volume or brain size} + \text{cohort-specific covariates}$   
187 fully adjusted models: as above + hypertension + type 2 diabetes + BMI + eGFR + current  
188 smoking status

189 Here, 'time' indicates the time in years between blood sampling and brain MRI, and 'fasting duration'  
190 denotes the time in hours between the last meal and blood sampling. Moreover, to investigate possible  
191 differences in the associations between the analytical subsamples, we fitted models to test for  
192 'metabolomic measure\*sex' and 'metabolomic measure\*statin use' interactions; this was done in both  
193 basic and fully adjusted models. In the Southall And Brent Revisited study where multiple major  
194 ethnicities were present, all models were fitted separately in each ethnic group.

195 *Meta-analyses and multiple testing correction*

196 The association results for metabolomic measures that were present in two or more cohorts (N=1,173)  
197 were meta-analyzed. We used inverse variance-weighted fixed-effect meta-analysis to combine the  
198 effect estimates and standard errors from each cohort. In case a cohort reported multiple association  
199 results for the same metabolic measure (*i.e.*, the metabolic measure was quantified using more than one  
200 metabolomic platform within the same cohort), the result that was obtained using a larger number of  
201 individuals was included in the meta-analysis.

202 Many metabolic measures are highly correlated and, thus, the number of independent tests is lower than  
203 the number of metabolomic measures tested. To correct for multiple testing, we estimated false  
204 discovery rate (FDR)-adjusted  $p$ -values using a method developed by Benjamini and Hochberg<sup>34</sup>. Here,  
205 all original  $p$ -values from the meta-analyses (including all analytical samples and both basic and fully  
206 adjusted models, and all metabolomic measures analyzed, including all lipid ratios) were included in  
207 the numeric vector of  $p$ -values used for estimating FDR-adjusted  $p$ -values ( $p_{FDR}$ ); all associations with  
208  $p_{FDR} < 0.05$  were considered significant. Testing for ‘metabolomic measure\*sex’ and ‘metabolomic  
209 measure\*statin treatment’ interactions were considered exploratory and no correction for multiple  
210 comparisons was applied.

211 All statistical analyses were conducted using R<sup>35</sup>. The authors declare that the summary-level results  
212 are available within the article and its online-only Data Supplement. The individual-level data analyzed  
213 in this study are available by application to the respective cohort committees.

214 **RESULTS**

215 Characteristics of the study populations are given in [Table 1](#), [Table S2](#) and [Figures S1-S2](#). In the meta-  
216 analysis, we identified 416 metabolomic measures showing nominally significant associations with  
217 WMH in at least one of the study models. Out of these, 30 (basic models) and 7 (fully adjusted models)  
218 associations remained significant after correction for multiple testing ( $p_{FDR} < 0.05$ ). An overview of the  
219 associations between circulating metabolomic measures and WMH in all basic and fully adjusted  
220 models is given in [Figure 1](#). All meta-analyzed results from the basic models and fully adjusted models  
221 are tabulated in [Tables S3](#) and [S4](#), respectively, and the results for ‘metabolomic measure’-by-sex and  
222 ‘metabolomic measure’-by-‘statin treatment’ interactions are given in [Tables S5](#) and [S6](#). Cohort-  
223 specific associations results are given in [Tables S7-S21](#). [Figure S3](#) illustrates cohort-specific results of  
224 the metabolomic measures showing FDR-significant association with WMH. [Figure S4](#) shows a  
225 comparison of the meta-analyzed results reported here versus the meta-analyzed results obtained in  
226 participants of European ancestries only. **We found the cohort-specific results to be highly similar**  
227 **across the cohorts, with very little heterogeneity observed** ([Figure S3](#), [Tables S3](#) and [S4](#)).

228 **Non-lipid measures**

229 The most robust association – in terms of both effect size and  $p$ -value – from across all studied  
230 metabolites was observed between WMH and higher circulating concentration of an amino acid  
231 derivative hydroxyphenylpyruvate (HPP; [Figure 2](#)): this association was most significant in the male  
232 subsample ( $\beta_{basic} = 0.20$ ,  $p_{FDR, basic} = 1.65 \times 10^{-6}$ ;  $\beta_{full, adj} = 0.22$ ,  $p_{FDR, full, adj} = 1.40 \times 10^{-7}$ ), but it was also  
233 significant in the pooled sample ( $\beta_{basic} = 0.13$ ,  $p_{FDR, basic} = 0.002$ ;  $\beta_{full, adj} = 0.15$ ,  $p_{FDR, full, adj} = 1.66 \times 10^{-4}$ )  
234 and in statin-nontreated subsample ( $\beta_{basic} = 0.15$ ,  $p_{FDR, basic} = 7.04 \times 10^{-5}$ ;  $\beta_{full, adj} = 0.18$ ,  
235  $p_{FDR, full, adj} = 1.65 \times 10^{-6}$ ). The HPP-by-sex interaction reached nominal significance in both the basic and  
236 fully adjusted models ( $p_{sexINT, basic} = 0.0094$ ,  $p_{sexINT, full, adj} = 0.0077$ ), suggesting that the positive effect size  
237 is larger in males than females.

238 In females, the only significant association, after correction for multiple testing, was observed between  
239 WMH and higher circulating concentration of glucuronate ( $\beta_{\text{basic}}=0.11$ ,  $p_{\text{FDR,basic}}=0.047$ ;  
240  $\beta_{\text{full,adj}}=0.11$ ,  $p_{\text{FDR,full,adj}}=0.047$ ; Figure 2); this association was significant also in statin-nontreated  
241 subsample ( $\beta_{\text{basic}}=0.10$ ,  $p_{\text{FDR,basic}}=0.025$ ;  $\beta_{\text{full,adj}}=0.10$ ,  $p_{\text{FDR,full,adj}}=0.040$ ). The glucuronate-by-sex  
242 interaction did not reach statistical significance ( $p_{\text{sexINT,basic}}=0.053$ ,  $p_{\text{sexINT,full,adj}}=0.129$ ).

#### 243 Lipid measures

244 WMH volume was associated with lower circulating concentrations of lysophosphatidylcholines  
245 (LPCs) and hydroxylated sphingomyelins (SM-OHs) (Figure 3). Among the studied LPCs, the strongest  
246 association was seen with LPC(22:6), which was significant in the pooled sample ( $\beta_{\text{basic}}=-0.078$ ,  
247  $p_{\text{FDR,basic}}=0.028$ ;  $\beta_{\text{full,adj}}=-0.082$ ,  $p_{\text{FDR,full,adj}}=0.047$ ) and in the male subsample ( $\beta_{\text{basic}}=-0.11$ ,  
248  $p_{\text{FDR,basic}}=0.025$ ;  $\beta_{\text{full,adj}}=-0.12$ ,  $p_{\text{FDR,full,adj}}=0.046$ ). Among the studied SM-OHs, the strongest  
249 association was observed with SM (OH) C22:2, and this association was significant in the male  
250 subsample only ( $\beta_{\text{basic}}=-0.11$ ,  $p_{\text{FDR,basic}}=0.017$ ;  $\beta_{\text{full,adj}}=-0.10$ ,  $p_{\text{FDR,full,adj}}=0.042$ ). Typically, the  
251 interactions between LPC measures and sex or statin treatment did not reach statistical significance  
252 (Tables S5 and S6); the exceptions were nominally significant interaction with sex for LPC(17:0) in the  
253 basic and fully adjusted models ( $p_{\text{sexINT,basic}}=0.027$ ,  $p_{\text{sexINT,fully,adj}}=0.045$ , respectively), and with statin  
254 treatment for LPC(20:4) in the fully adjusted model ( $p_{\text{statinINT,fully,adj}}=0.015$ ) (Figure 3). The SM-OH  
255 species-by-sex interactions were nominally significant in both basic and fully adjusted models for  
256 SM (OH) C14:1 ( $p_{\text{sexINT,basic}}=0.018$ ,  $p_{\text{sexINT,fully,adj}}=0.033$ ), SM (OH) C16:1 ( $p_{\text{sexINT,basic}}=0.027$ ,  
257  $p_{\text{sexINT,fully,adj}}=0.044$ ), and SM (OH) C22:2 ( $p_{\text{sexINT,basic}}=0.0061$ ,  $p_{\text{sexINT,fully,adj}}=0.013$ ). Also,  
258 SM (OH) C14:1-by-statin treatment interaction was nominally significant in the fully adjusted model  
259 ( $p_{\text{statinINT,fully,adj}}=0.047$ ).

260 A sizable proportion of the studied metabolomic measures were circulating concentrations of  
261 lipoproteins and lipoprotein lipids (approximately 24% of the meta-analyzed measures). Out of the 30  
262 metabolomic measures showing FDR-significant associations with WMH in the basic models, 12 were  
263 with measures of the lipid composition of the intermediate density and low-density lipoprotein (IDL,  
264 LDL) particles (lipid composition calculated as a ratio of a lipid concentration against total lipids  
265 concentration within individual lipoprotein subfractions). Specifically, higher WMH volume was  
266 associated with higher TG-to-total-lipids ratio and lower cholesteryl ester (CE)-to-total-lipids ratio in  
267 the pooled sample and male subsample (Figure 4). The effect sizes were attenuated in the fully adjusted  
268 models (Figure 4). Concomitantly, higher WMH volume was associated with larger LDL-particle  
269 diameter in the male subsample, and this association remained FDR-significant in the fully adjusted  
270 model ( $\beta_{\text{basic}}=0.074$ ,  $p_{\text{FDR,basic}}=0.049$ ;  $\beta_{\text{full,adj}}=0.078$ ,  $p_{\text{FDR,full,adj}}=0.047$ ). We observed a nominally  
271 significant LDL diameter-by-sex interaction ( $p_{\text{sexINT,basic}}=0.0093$ ,  $p_{\text{sexINT,full,adj}}=0.036$ ). Higher WMH  
272 volume was also associated with lower free cholesterol-to-total-lipids-ratio within medium-sized very-  
273 low-density lipoprotein (VLDL) subfraction in the statin-treated subsample (Figure 1), but no other  
274 significant association with either VLDL or high-density lipoprotein (HDL) subfraction measures was  
275 seen (Tables S3 and S4).

#### 276 DISCUSSION

277 In this study, we investigated associations between WMH volume and circulating metabolomic  
278 measures in up to 9,290 individuals. As discussed in the following text, several aspects of our  
279 metabolomic findings support a possible vessel-related pathophysiology of WMH, and some suggest  
280 the involvement of myelin disruption and neuron injury. Further, a number of the observed associations

281 showed sex specificities indicating that distinct metabolomic features accompany WMH in males and  
282 females.

283 In the present study, the most robust association was the positive association between WMH volume  
284 and HPP in males ( $p_{\text{FDR,full,adj}}=1.40 \times 10^{-7}$ ); the association was also significant in the pooled sample  
285 ( $p_{\text{FDR,full,adj}}=1.66 \times 10^{-4}$ ) and statin-nontreated individuals ( $p_{\text{FDR,full,adj}}=1.65 \times 10^{-6}$ ). HPP is a potentially  
286 toxic compound derived from the catabolism of phenylalanine and tyrosine<sup>36</sup> (Figure 2). Higher  
287 circulating levels of phenylalanine and tyrosine have been associated with higher risk of cardiovascular  
288 disease (CVD) in prior studies<sup>37,38</sup>, but, to our knowledge, HPP was not examined in those studies. We  
289 did not see strong associations with phenylalanine or tyrosine and, thus, the association between WMH  
290 and HPP likely arises downstream from phenylalanine hydroxylase (PAH) (Figure 2). The enzymatic  
291 alterations possibly contributing to higher level of HPP could be a higher activity of tyrosine  
292 aminotransferase (TAT) or lower activity of hydroxyphenylpyruvate dioxygenase (HPD). In tyrosine  
293 breakdown, TAT converts tyrosine and  $\alpha$ -ketoglutarate to HPP and glutamate<sup>39</sup> and, thus, higher TAT  
294 activity could contribute to lower  $\alpha$ -ketoglutarate and, at the same time, to higher glutamate. In males  
295 and statin-treated participants, however, we found only a nominal association between WMH and  
296 circulating  $\alpha$ -ketoglutarate that was not in the anticipated direction and no association with glutamate  
297 (Figure 2). Therefore, in the view of these results, lower HPD activity appears as the most likely  
298 mechanism driving the association between WMH and circulating HPP. **HPD requires oxygen to  
299 convert HPP to homogentisic acid (Figure 2) and, consistent with this requirement, hypoxia promotes  
300 accumulation of HPP<sup>40</sup>. The deep and periventricular white matter, which is the predilection site for  
301 WMH, is characterized by sparse vasculature consisting of long, narrow end arteries/arterioles that are  
302 vulnerable to oxygen desaturation<sup>41</sup>. Thus, higher circulating HPP in association with higher WMH  
303 may be an indicator of ischemic hypoxia promoting WMH. As indicated above, the association of HPP  
304 with WMH volumes was present in the pooled sample, in males, and in the statin-nontreated subsample,  
305 but not in females. In males, HPP explained 14% of variance in WMH, while the proportions of variance  
306 explained by age, hypertension, diabetes, and smoking were 19%, 1%, 1% and 0.1%, respectively  
307 (Table S3). Our results are consistent with previous results from prospective cohorts, demonstrating  
308 that vascular risk factors, including hypertension, explain only 0.1-2.0% of variance in the WMH  
309 volume<sup>7</sup>. Taken together, our results suggest that circulating HPP may be a strong biomarker of WMH  
310 in males, but its potential in clinical use requires further research.**

311 Glucuronate was the only metabolite demonstrating a robust association with WMH in females  
312 ( $p_{\text{FDR,full,adj}}=0.047$ ); the association was also significant in statin-nontreated participants  
313 ( $p_{\text{FDR,full,adj}}=0.040$ ). Glucuronate is derived from glucose, and, in humans, it is involved in the  
314 elimination of toxic substances by making them more water-soluble in a process called  
315 glucuronidation<sup>36</sup>. In addition, hormones can be glucuronidated to enable easier transport<sup>36</sup>. A key  
316 enzyme of glucuronidation is UDP-glucuronosyltransferase, which is highly expressed in endothelial  
317 cells of the blood-brain barrier (BBB) and associated astrocytes, where the enzyme contributes to the  
318 protection of the brain from systemic toxic substances (reviewed by Ouzzine *et al.*<sup>42</sup>). Also, endothelial  
319 cells use glucuronate to synthesize glycosaminoglycans, which are constituents of the glycocalyx layer  
320 that tightens the endothelial barrier and limits vascular permeability<sup>43</sup>. The observed WMH association  
321 with glucuronate may involve altered glucose metabolism in endothelial cells, which might compromise  
322 the molecular mechanisms enabling the protective functions of the BBB. Of note, the association  
323 between WMH and glucuronate remained significant after adjusting for type 2 diabetes in the fully  
324 adjusted model. In our study, WMH association with (mostly fasting) glucose level did not reach  
325 statistical significance, which is in line with some<sup>44</sup> but not all<sup>45</sup> previous reports.



326 In addition to the above-discussed non-lipid measures, we found that WMH were associated with  
327 several circulating lipids – most notably with LPCs and SM-OHs. These associations were almost  
328 exclusively negative and reached statistical significance predominantly in males and, in some cases,  
329 also in the pooled sample. LPCs are phospholipids generated by partial hydrolysis of  
330 phosphatidylcholines, which are the building blocks of cell membranes, such as those of blood cells  
331 and endothelial cells. In circulation, LPCs modulate inflammation and oxidative stress<sup>46,47</sup>, and they  
332 may alter the integrity of endothelial membranes, including the BBB<sup>48,49</sup>. Lower circulating levels of  
333 LPCs, such as LPC(18:2), have been associated with higher CVD risk<sup>50</sup> and adverse cognitive  
334 outcomes<sup>51</sup>. Consistent with these previous reports, we found that lower LPC(18:2) is associated with  
335 higher WMH volume, but similar to most other tested LPCs, the association reached only nominal  
336 significance in the fully adjusted model. The only LPC that remained significantly associated with  
337 WMH in the fully adjusted model was LPC(22:6) (males:  $p_{\text{FDR,full,adj}}=0.046$ ; pooled sample:  
338  $p_{\text{FDR,full,adj}}=0.047$ ). Docosahexaenoic acid (DHA, 22:6n-3) is one of the two predominant fatty acids in  
339 the human brain and a structural component of neuronal cell membranes<sup>52,53</sup>. DHA plays a crucial role  
340 in neuronal survival, neurogenesis, and synaptic function<sup>52</sup>. Evidence obtained in mice suggests that  
341 oral administration of LPC-DHA, but not free DHA, increases DHA content of the brain and improves  
342 spatial learning and memory<sup>54</sup>. Thus, the associations of circulating LPCs with WMH observed here  
343 suggest the BBB and neuron injury-related pathobiologies of WMH.

344 Similar to LPCs, the associations between WMH and SM-OHs were negative, with the most  
345 significantly associated SM-OH being SM (OH) C22:2 in males ( $p_{\text{FDR,full,adj}}=0.042$ ). SM-OHs are  
346 important components of myelin sheaths<sup>55</sup>, which are lipid-rich cell membranes wrapped around  
347 neuronal axons, protecting the axons physically and providing trophic support, as well as increasing the  
348 conduction speed of action potentials<sup>56</sup>. The negative associations between WMH and SM-OHs could  
349 point towards disruption of myelin sheaths<sup>55</sup>, which is one of the key features of WMH<sup>1</sup>. In line with  
350 our findings, previous evidence suggests that low level of serum total SM is associated with cross-  
351 sectional memory impairment<sup>57</sup>.

352 Regarding the associations with lipoprotein measures, we observed that WMH were associated with  
353 larger LDL-particle diameter ( $p_{\text{FDR,full,adj}}=0.047$  in males), likely arising from altered LDL-lipid  
354 composition, namely relatively higher TG and lower CE content. Putatively, an altered interaction  
355 between lipoproteins and enzymes modulating their lipid composition could play a role in these  
356 associations. For example, the endothelium-bound lipoprotein lipase (LPL), which catalyzes the  
357 hydrolysis of lipoprotein TGs, interacts directly with LDL-receptor related protein<sup>58</sup> that is involved in  
358 the regulation of the BBB permeability<sup>59</sup>; as such, LPL displays functions related to both blood  
359 metabolome and WMH found in areas with enhanced permeability of the BBB<sup>60</sup>. Statin treatment may  
360 have a small confounding effect on these associations, as statin-treated individuals have the highest  
361 WMH volumes together with the lowest cholesterol levels (Table 1). But considering the facts that the  
362 effect estimates of these measures were consistent across all analytical subsamples, including statin-  
363 nontreated individuals, and, that the effect estimates were only marginally attenuated in the statin use-  
364 adjusted models compared with the basic models (Figure 4), statin use does not seem to be a key factor  
365 driving the negative associations between WMH and lower CE content in the LDL subfractions.

366 In the present study, several of the FDR-significant associations in the fully adjusted models showed  
367 nominally significant metabolite-by-sex interactions suggesting that the effect sizes vary depending on  
368 sex. These associations included the amino acid-derivative HPP ( $p_{\text{sexINT,full,adj}}=0.0077$ ), and two lipid  
369 measures, namely SM (OH) C22:2 ( $p_{\text{sexINT,full,adj}}=0.013$ ) and LDL-particle size ( $p_{\text{sexINT,full,adj}}=0.036$ ): for  
370 all three, the effect sizes were nominally greater in males than in females. These sex specificities were

371 observed despite similar sample sizes of males and females, and no major sex differences in the  
372 distributions of WMH and key traits, such as BMI, cigarette smoking or clinical lipid traits (Table 1).  
373 Also, the metabolomic measures did not show major sex differences, except for the circulating levels  
374 of SM (OH) C22:2, which tended to be lower in males than in females (Figure S1), as reported  
375 previously<sup>61</sup>. Considering that hydroxysphingomyelins are important for maintenance myelination<sup>55</sup>,  
376 the lower levels of this lipid in males than in females could contribute to the nominally greater negative  
377 effect size of SM (OH) C22:2 on WMH in males than in females. Regarding HPP, the circulating levels  
378 were similar between males and females (Figure S1), and the biological mechanism of the nominally  
379 greater positive effect of HPP on WMH in males vs. females remains inconclusive. Nevertheless, it  
380 could be that circulating HPP is a marker of ischemic hypoxia that is of a similar magnitude in both  
381 males and females (as reflected by similar circulating levels of HPP in the two sexes), but the ischemic  
382 hypoxia is of a greater impact in males than in females. This is supported by previous research indicating  
383 that, in males compared with females, ischemic hypoxia in the brain induces greater oxidative stress  
384 and, thus, possibly greater tissue damage and more extensive WMH<sup>62,63</sup>. Finally, with respect to the  
385 LDL-particle size, this measure also did not show consistent sex differences across the analyzed cohorts  
386 (Figure S1), and the biological mechanisms of the nominally greater positive effect size of LDL-particle  
387 size on WMH in males than in females are not clear. Previous literature on sex differences in WMH is  
388 not extensive<sup>64-66</sup>. Sex hormones may play a role, as they show direct effects on the endothelium<sup>67</sup> and  
389 multiple brain cells, including oligodendrocytes producing myelin<sup>68</sup> and neurons<sup>69</sup>. In our study, female  
390 hormonal status was not likely a major factor in the observed sex specificities, as female participants  
391 were likely menopausal (average age >60 years). We cannot, however, exclude the possibility that the  
392 use of hormonal replacement therapy and a greater lifetime exposure to female sex hormones would  
393 modulate the associations examined in older age. Overall, our results indicating sex-specific  
394 associations between metabolomic measures and WMH add to the growing body of research reporting  
395 sex differences in the associations between risk factors and cerebrovascular outcomes<sup>70-74</sup>.

396 The strengths and limitations of this study should be considered. We analyzed data from multiple  
397 population-based cohorts that enables a large sample size, yielding the statistical power required to  
398 detect reliably the small effect sizes and to replicate/meta-analyze the results. Although the study  
399 sample included individuals from multiple geographical areas and diverse ethnic backgrounds, the vast  
400 majority of participants were of European ancestries, and, thus, replication of the findings in other  
401 ethnicities would be of high value. The fact that we analyzed metabolomic data quantified with multiple  
402 platforms could be considered as both a strength and a limitation: while covering a wide spectrum of  
403 circulating lipids and metabolites, some of the metabolic measures are underrepresented and, therefore,  
404 we may lack statistical power to detect associations with these measures. Further, due to the multiple  
405 testing correction to minimize the risk of false positive associations, some biologically relevant  
406 associations may be missed. For instance, many of the associations between WMH and lipid  
407 concentrations within IDL and LDL subfractions were negative with non-null-containing 95%  
408 confidence intervals; however, only a minority of these associations reached FDR-significance. As  
409 lipoprotein metabolism is an interconnected system, it may be more meaningful to look at the big picture  
410 instead of interpreting the associations on the level of individual metabolic measures. In most cohorts,  
411 WMH volume was quantified with automated image-analysis techniques and, thus, we cannot exclude  
412 the possibility that silent lacunar lesions (also part of small vessel disease<sup>2,75</sup>) were included in the  
413 quantified WMH volume. Manual segmentation of WMH is extremely labor-intensive and is not  
414 frequently used in large-scale cohort studies including thousands of participants<sup>6</sup>, such as the ones  
415 included here. In future studies, automatic segmentation of multi-modal MR images may allow one to  
416 compare metabolic profiles of different types of tissue abnormalities present in white matter.

417 Additionally, it would be of high value to assess the causal inferences between circulating metabolites  
418 and WMH in future studies once suitable genetic instruments are identified. Considering the results of  
419 our study, it is likely necessary to test the causality in sex-specific analyses. Finally, further insight into  
420 the pathophysiology of WMH awaits the results of LACI-2 trial, as this trial is testing two repurposed  
421 licenced drugs with effects on endothelium-dependent vasodilation and BBB integrity to prevent  
422 progression of cerebral small disease<sup>76</sup>.

423 In summary, this is the first large-scale study to determine associations between WMH and circulating  
424 metabolomic measures. While no causal associations can be inferred from the present findings, our  
425 findings indicate that alterations in circulating metabolism are closely linked with WMH in a general  
426 population of middle aged and older adults. The present findings suggest that the metabolomic profiles  
427 accompanying WMH in males and females may differ.

## Tables

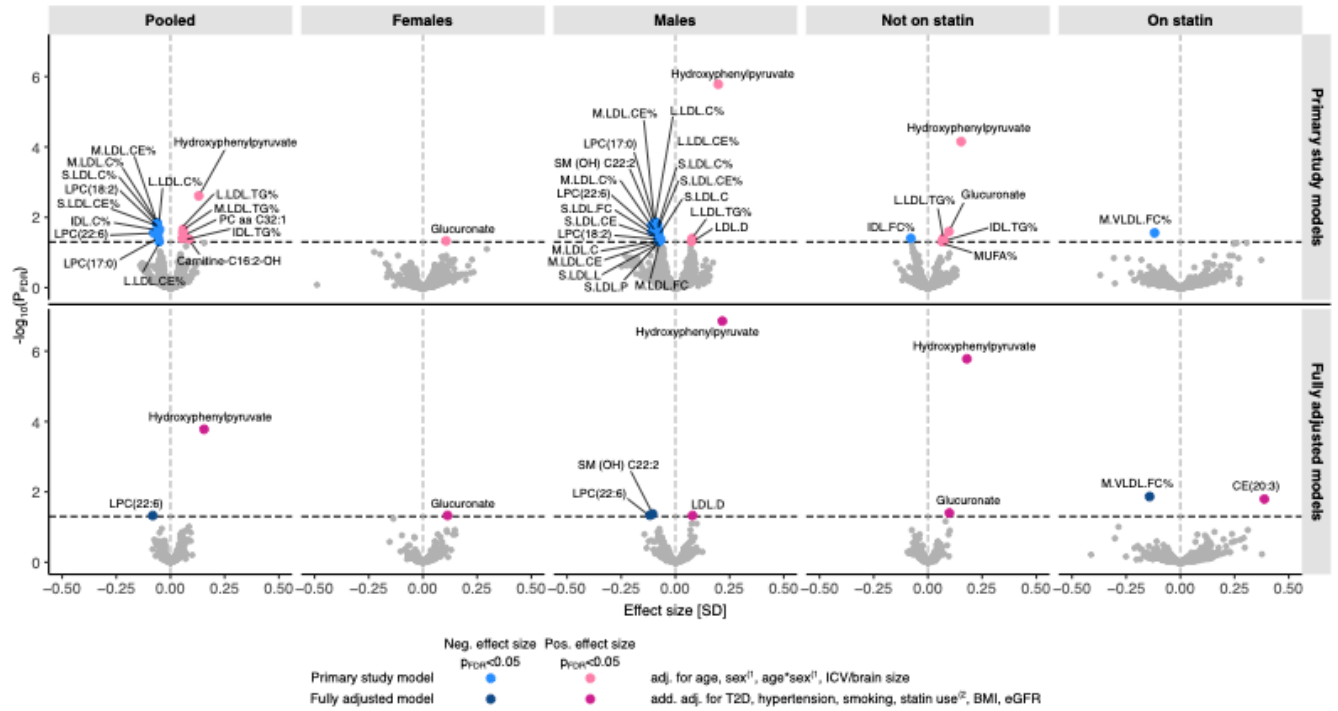
**Table 1. Sample characteristics.**

Values are mean  $\pm$  standard deviation of the pooled data of 15 populations from 8 cohort studies. The population-specific characteristics are given in [Table S2](#).

Characteristic	Combined	Females	Males	Not on statin	On statin
<b>Number of individuals</b>	9,290	4,711	4,579	7,565	1,633
<b>Males (%)</b>	49.3	0	100	47.2	58.7
<b>Smokers (%)</b>	13.6	14.0	13.3	14.5	12.4
<b>On statin (%)</b>	17.7	14.4	21.0	0	100
<b>Hypertension (%)</b>	47.1	44.9	49.3	41.3	74.4
<b>Type 2 diabetes (%)</b>	7.0	6.1	7.9	4.0	20.7
<b>Age (years)</b>	61.0 $\pm$ 7.3	60.7 $\pm$ 7.4	61.4 $\pm$ 7.2	59.6 $\pm$ 7.5	67.9 $\pm$ 5.6
<b>BMI (kg/m<sup>2</sup>)</b>	27.3 $\pm$ 4.4	27.1 $\pm$ 4.7	27.5 $\pm$ 4.0	27.1 $\pm$ 4.4	28.2 $\pm$ 4.1
<b>logWMH</b>	5.35 $\pm$ 0.95	5.17 $\pm$ 0.93	5.35 $\pm$ 0.95	5.05 $\pm$ 0.91	6.78 $\pm$ 1.07
<b>Total-TG (mmol/L)*</b>	1.38 $\pm$ 0.65	1.37 $\pm$ 0.62	1.39 $\pm$ 0.65	1.36 $\pm$ 0.63	1.45 $\pm$ 0.69
<b>Total-C (mmol/L)*</b>	4.95 $\pm$ 1.00	5.10 $\pm$ 0.97	4.81 $\pm$ 0.97	5.18 $\pm$ 0.93	4.17 $\pm$ 0.89
<b>LDL-C (mmol/L)**</b>	2.39 $\pm$ 0.67	2.49 $\pm$ 0.67	2.31 $\pm$ 0.66	2.57 $\pm$ 0.64	1.74 $\pm$ 0.54
<b>HDL-C (mmol/L)**</b>	1.51 $\pm$ 0.36	1.58 $\pm$ 0.34	1.43 $\pm$ 0.31	1.54 $\pm$ 0.63	1.39 $\pm$ 0.33

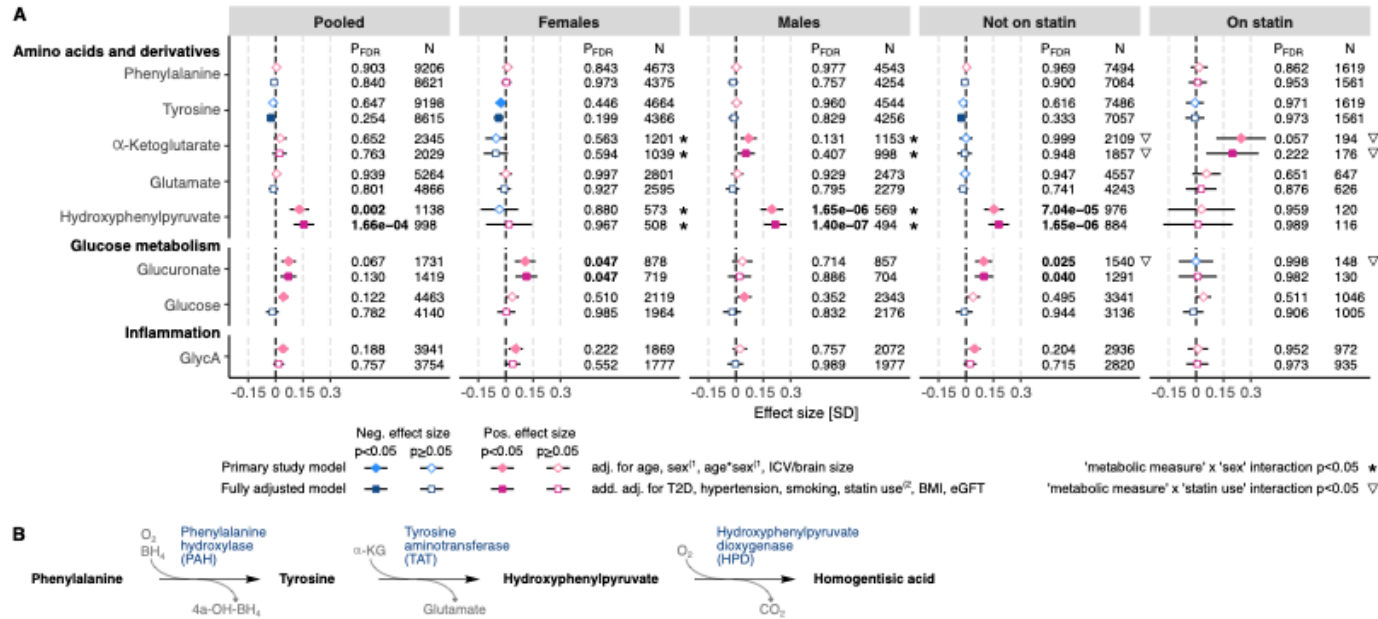
\* Pooled data of 9 populations for which total-TG and total-C were available.

\*\* Pooled data of 10 populations for which LDL-C and HDL-C were available.



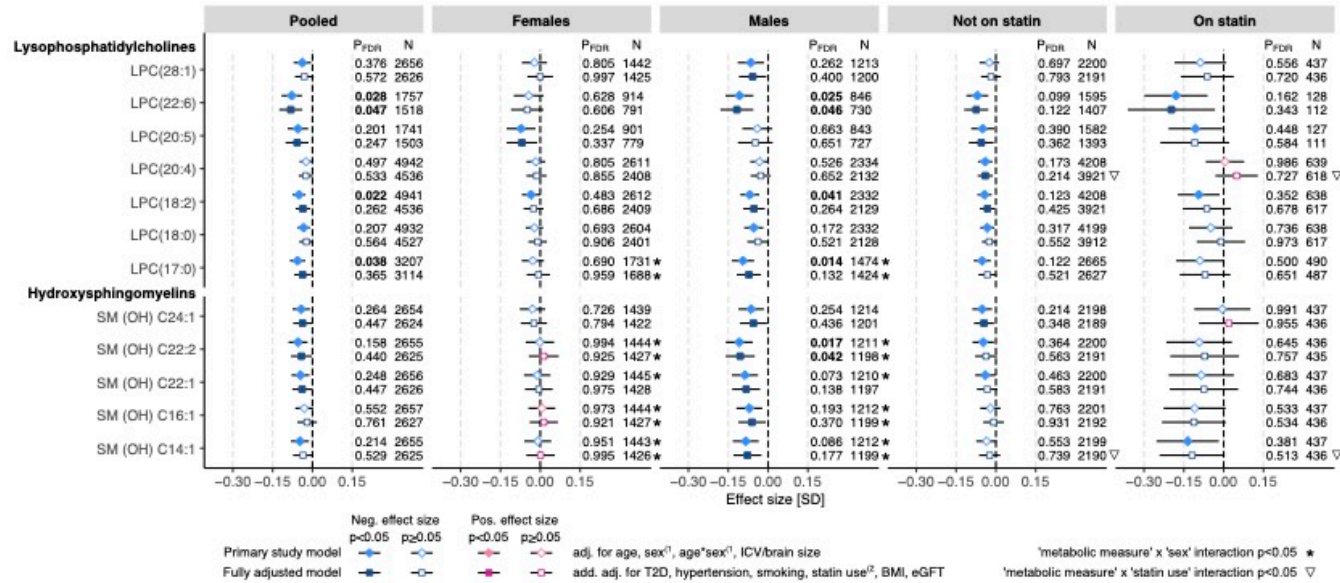
**Figure 1. Metabolic associations of WMH.**

The plot shows the effect sizes (x-axis) and statistical significance (y-axis) of the metabolic associations of WMH as obtained in the meta-analysis of up to 9,290 individuals from 15 populations of mostly European ancestry. Metabolic associations of log-transformed WMH were determined by fitting linear regression models separately in the pooled sample, and in sex or statin treatment-stratified subsamples (columns). Population-specific effect sizes and standard errors were combined with inverse variance-weighted fixed effect meta-analysis. The metabolomic measures showing a significant ( $P_{FDR} < 0.05$ ) association with WMH are labeled. *Primary study models (top row)*: The associations were adjusted for age and intracranial volume or brain size, and also for sex and age-by-sex interaction in the pooled sample and in statin treatment-stratified subsamples<sup>(1)</sup>. *Fully adjusted models (bottom row)*: The associations were additionally adjusted for type 2 diabetes (T2D), hypertension, current smoking status, body mass index (BMI), estimated glomerular filtration rate (eGFR), and hypertension, and for statin treatment in the pooled sample and in sex-stratified subsamples<sup>(2)</sup>. Where relevant, all models were also adjusted for fasting duration, time between blood sampling and brain MRI, and cohort study-specific covariates.



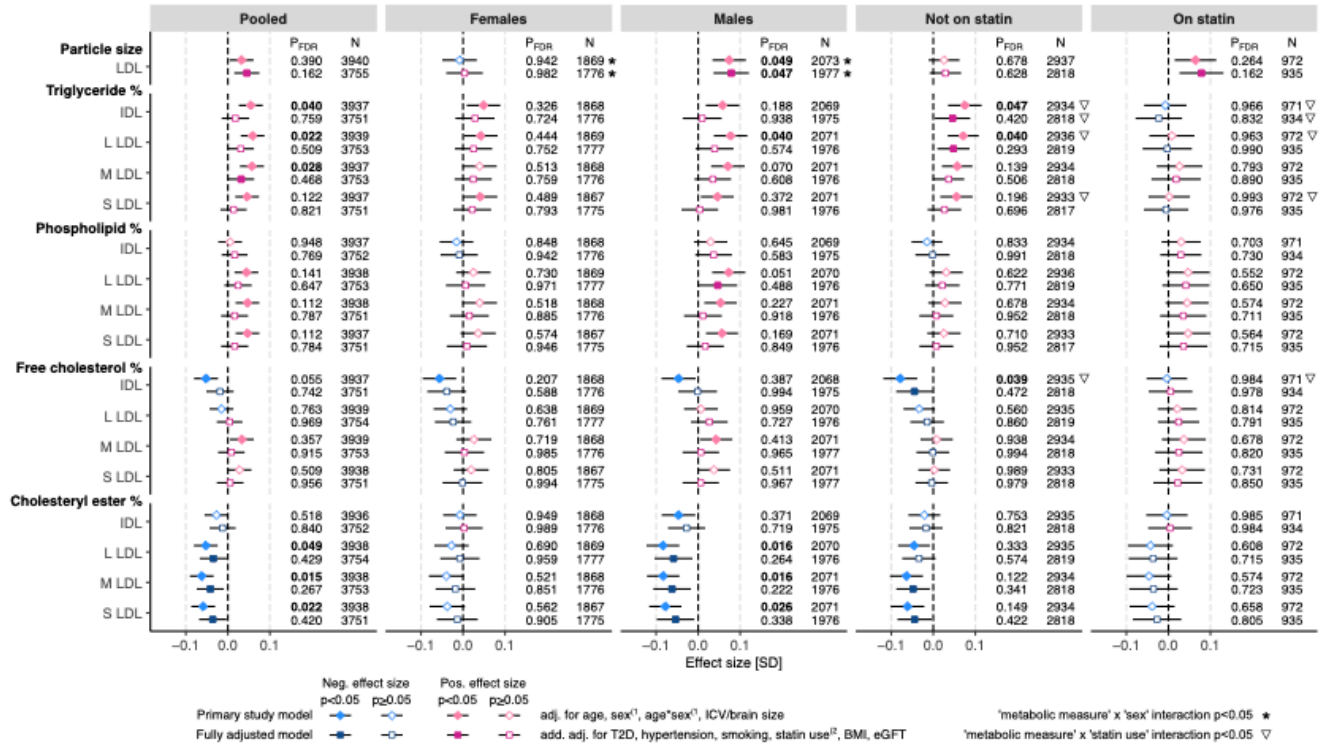
**Figure 2. WMH associations with circulating levels of selected non-lipid metabolites.**

A) The associations between WMH and circulating metabolomic measures were determined using linear regression. The primary study models (diamonds) were adjusted for age and intracranial volume or brain size, and, in the pooled sample and in statin treatment-stratified subsamples also for sex and age-by-sex interaction<sup>1</sup>. The fully adjusted models (squares) were additionally adjusted for type 2 diabetes (T2D), hypertension, current smoking status, body mass index (BMI), and estimated glomerular filtration rate (eGFR), and, in the pooled sample and in sex-stratified subsamples, also for statin treatment<sup>2</sup>. Where relevant, all models were adjusted for fasting duration, time between blood sampling and brain MRI, and possible cohort study-specific covariates. Blue color indicates negative effect size and red color indicates positive effect size. Error bars indicate 95% confidence intervals. P-values below the threshold for multiple testing correction ( $P_{FDR}<0.05$ ) are indicated with bold font. B) Catabolic pathway of phenylalanine and tyrosine into hydroxyphenylpyruvate and homogentisic acid by phenylalanine hydroxylase (PAH), tyrosine aminotransferase (TAT) and hydroxyphenylpyruvate dioxygenase (HPD).



**Figure 3. WMH associations with circulating levels of lysophosphatidylcholines (LPC) and hydroxylated sphingomyelins (SM (OH)).**

The associations between WMH and circulating metabolomic measures were determined using linear regression. The primary study models (diamonds) were adjusted for age and intracranial volume or brain size, and, in the pooled sample and in statin treatment-stratified subsamples also for sex and age-by-sex interaction<sup>†</sup>. The fully adjusted models (squares) were additionally adjusted for type 2 diabetes (T2D), hypertension, current smoking status, body mass index (BMI), and estimated glomerular filtration rate (eGFR), and, in the pooled sample and in sex-stratified subsamples, also for statin treatment<sup>‡</sup>. Where relevant, all models were adjusted for fasting duration, time between blood sampling and brain MRI, and possible cohort study-specific covariates. Blue color indicates negative effect size and red color indicates positive effect size. Error bars indicate 95% confidence intervals. P-values below the threshold for multiple testing correction ( $p_{FDR} < 0.05$ ) are indicated with bold font.



**Figure 4. WMH associations with LDL particle size and lipid composition measures in IDL and LDL subfractions.**

The associations between WMH and circulating metabolic measures were determined using linear regression. The primary study models (diamonds) were adjusted for age and intracranial volume or brain size, and, in the pooled sample and in statin treatment-stratified subsamples also for sex and age-by-sex interaction<sup>(1)</sup>. The fully adjusted models (squares) were additionally adjusted for type 2 diabetes (T2D), hypertension, current smoking status, body mass index (BMI), and estimated glomerular filtration rate (eGFR), and, in the pooled sample and in sex-stratified subsamples, also for statin treatment<sup>(2)</sup>. Where relevant, all models were adjusted for fasting duration, time between blood sampling and brain MRI, and possible cohort study-specific covariates. Blue color indicates negative effect size and red color indicates positive effect size. Error bars indicate 95% confidence intervals. P-values below the threshold for multiple testing correction ( $p_{FDR} < 0.05$ ) are indicated with bold font.



## Disclosures

None.

## References

1. Moroni F, Ammirati E, Hainsworth AH, Camici PG. Association of White Matter Hyperintensities and Cardiovascular Disease: The Importance of Microcirculatory Disease. *Circ Cardiovasc Imaging*. 2020;1–13.
2. Wardlaw JM, Smith C, Dichgans M. Small vessel disease: mechanisms and clinical implications. *Lancet Neurol*. 2019;18:684–696.
3. Debette S, Schilling S, Duperron MG, Larsson SC, Markus HS. Clinical Significance of Magnetic Resonance Imaging Markers of Vascular Brain Injury: A Systematic Review and Meta-analysis. *JAMA Neurol*. 2019;76:81–94.
4. Debette S, Markus HS. The clinical importance of white matter hyperintensities on brain magnetic resonance imaging: Systematic review and meta-analysis. *BMJ*. 2010;341:288.
5. Power MC, Deal JA, Sharrett AR, Jack CR, Knopman D, Mosley TH, Gottesman RF. Smoking and white matter hyperintensity progression. *Neurology*. 2015;84:841 LP – 848.
6. Sargurupremraj M, Suzuki H, Jian X, Sarnowski C, Evans TE, Bis JC, Eiriksdottir G, Sakaue S, Terzikhan N, Habes M, Zhao W, Armstrong NJ, Hofer E, Yanek LR, Hagenaars SP, Kumar RB, van den Akker EB, McWhirter RE, Trompet S, Mishra A, Saba Y, Satizabal CL, Beaudet G, Petit L, Tsuchida A, Zago L, Schilling S, Sigurdsson S, Gottesman RF, Lewis CE, Aggarwal NT, Lopez OL, Smith JA, Valdés Hernández MC, van der Grond J, Wright MJ, Knol MJ, Dörr M, Thomson RJ, Bordes C, Le Grand Q, Duperron MG, Smith A V., Knopman DS, Schreiner PJ, Evans DA, Rotter JI, Beiser AS, Maniega SM, Beekman M, Trollor J, Stott DJ, Vernooij MW, Wittfeld K, Niessen WJ, Soumaré A, Boerwinkle E, Sidney S, Turner ST, Davies G, Thalamuthu A, Völker U, van Buchem MA, Bryan RN, Dupuis J, Bastin ME, Ames D, Teumer A, Amouyel P, Kwok JB, Bülow R, Deary IJ, Schofield PR, Brodaty H, Jiang J, Tabara Y, Setoh K, Miyamoto S, Yoshida K, Nagata M, Kamatani Y, Matsuda F, Psaty BM, Bennett DA, De Jager PL, Mosley TH, Sachdev PS, Schmidt R, Warren HR, Evangelou E, Trégouët DA, Amouyel P, de Andrade M, Basu S, Berr C, Brody JA, Chasman DI, Dartigues JF, et al. Cerebral small vessel disease genomics and its implications across the lifespan. *Nat Commun*. 2020;11.
7. Wardlaw JM, Allerhand M, Doubal FN, Hernandez MV, Morris Z, Gow AJ, Bastin M, Starr JM, Dennis MS, Deary IJ. Vascular risk factors, large-artery atheroma, and brain white matter hyperintensities. *Neurology*. 2014;82:1331–1338.
8. Bernath MM, Bhattacharyya S, Nho K, Barupal DK, Fiehn O, Baillie R, Risacher SL, Arnold M, Jacobson T, Trojanowski JQ, Shaw LM, Weiner MW, Doraiswamy PM, Kaddurah-Daouk R, Saykin AJ. Serum triglycerides in Alzheimer disease. *Neurology*. 2020;10.1212/WNL.0000000000009436.
9. Varma VR, Oommen AM, Varma S, Casanova R, An Y, Andrews RM, O'Brien R, Pletnikova O, Troncoso JC, Toledo J, Baillie R, Arnold M, Kastenmueller G, Nho K, Doraiswamy PM, Saykin AJ, Kaddurah-Daouk R, Legido-Quigley C, Thambisetty M. Brain and blood metabolite signatures of pathology and progression in Alzheimer disease: A targeted metabolomics study. *PLoS Med*. 2018;15:1–31.
10. van der Lee SJ, Teunissen CE, Pool R, Shipley MJ, Teumer A, Chouraki V, Melo van Lent D, Tynkkynen J, Fischer K, Hernesniemi J, Haller T, Singh-Manoux A, Verhoeven A, Willemsen G, de Leeuw FA, Wagner H, van Dongen J, Hertel J, Budde K, Willems van Dijk K, Weinhold L, Ikram MA, Pietzner M, Perola M, Wagner M, Friedrich N, Slagboom PE, Scheltens P, Yang Q, Gertzen RE, Egert S, Li S, Hankemeier T, van Beijsterveldt CEM, Vasani RS, Maier W, Peeters CFW, Jörgen Grabe H, Ramirez A, Seshadri S, Metspalu A, Kivimäki M, Salomaa V,

- Demirkan A, Boomsma DI, van der Flier WM, Amin N, van Duijn CM. Circulating metabolites and general cognitive ability and dementia: Evidence from 11 cohort studies. *Alzheimer's Dement.* 2018;14:707–722.
11. Syme C, Pelletier S, Shin J, Abrahamowicz M, Leonard G, Perron M, Richer L, Veillette S, Gaudet D, Pike B, Strug LJ, Wang Y, Xu H, Taylor G, Bennett S, Paus T, Pausova Z. Visceral fat-related systemic inflammation and the adolescent brain: a mediating role of circulating glycerophosphocholines. *Int J Obes.* 2019;43:1223–1230.
  12. Sliz E, Shin J, Syme C, Patel Y, Parker N, Richer L, Gaudet D, Bennett S, Paus T, Pausova Z. A variant near DHCR24 associates with microstructural properties of white matter and peripheral lipid metabolism in adolescents. *Mol Psychiatry.* 2020;10.1038/s41380-019-0640–9.
  13. Sliz E, Shin J, Syme C, Black S, Seshadri S, Paus T, Pausova Z. Thickness of the cerebral cortex shows positive association with blood levels of triacylglycerols carrying 18-carbon fatty acids. *Commun Biol.* 2020;3:1–9.
  14. de Leeuw FA, Karamujić-Čomić H, Tijms BM, Peeters CFW, Kester MI, Scheltens P, Ahmad S, Vojinovic D, Adams HHH, Hankemeier T, Bos D, van der Lugt A, Vernooij MW, Ikram MA, Amin N, Barkhof F, Teunissen CE, van Duijn CM, van der Flier WM. Circulating metabolites are associated with brain atrophy and white matter hyperintensities. *Alzheimer's Dement.* 2020;205–214.
  15. Li D, Misialek JR, Jack CR, Mielke MM, Knopman D, Gottesman R, Mosley T, Alonso A. Plasma metabolites associated with brain MRI measures of neurodegeneration in older adults in the atherosclerosis risk in communities– neurocognitive study (ARIC-NCS). *Int J Mol Sci.* 2019;20:1–12.
  16. Harris TB, Launer LJ, Eiriksdottir G, Kjartansson O, Jonsson P V., Sigurdsson G, Thorgeirsson G, Aspelund T, Garcia ME, Cotch MF, Hoffman HJ, Gudnason V. Age, gene/environment susceptibility-reykjavik study: Multidisciplinary applied phenomics. *Am J Epidemiol.* 2007;165:1076–1087.
  17. DeCarli C, Massaro J, Harvey D, Hald J, Tullberg M, Au R, Beiser A, D'Agostino R, Wolf PA. Measures of brain morphology and infarction in the framingham heart study: Establishing what is normal. *Neurobiol Aging.* 2005;26:491–510.
  18. Tsao CW, Vasan RS. Cohort Profile: The Framingham Heart Study (FHS): Overview of milestones in cardiovascular epidemiology. *Int J Epidemiol.* 2015;44:1800–1813.
  19. Lane CA, Parker TD, Cash DM, Macpherson K, Donnachie E, Murray-Smith H, Barnes A, Barker S, Beasley DG, Bras J, Brown D, Burgos N, Byford M, Jorge Cardoso M, Carvalho A, Collins J, De Vita E, Dickson JC, Epie N, Espak M, Henley SMD, Hoskote C, Hutel M, Klimova J, Malone IB, Markiewicz P, Melbourne A, Modat M, Schrag A, Shah S, Sharma N, Sudre CH, Thomas DL, Wong A, Zhang H, Hardy J, Zetterberg H, Ourselin S, Crutch SJ, Kuh D, Richards M, Fox NC, Schott JM. Study protocol: Insight 46 - a neuroscience sub-study of the MRC National Survey of Health and Development. *BMC Neurol.* 2017;17:1–25.
  20. Taylor AM, Pattie A, Deary IJ. Cohort Profile Update : The Lothian Birth Cohorts of 1921 and 1936. *Int J Epidemiol.* 2018;47:1042-1042r.
  21. Wardlaw JM, Bastin ME, Valde MC, Royle NA, Morris Z, Clayden JD, Sandeman EM, Eadie E, Murray C, Starr JM, Deary IJ. Brain aging , cognition in youth and old age and vascular disease in the Lothian Birth Cohort 1936: rationale , design and methodology of the imaging protocol. 2011;6:547–559.
  22. Hofman A, Breteler MMB, Van Duijn CM, Krestin GP, Pols HA, Stricker BHC, Tiemeier H, Uitterlinden AG, Vingerling JR, Witteman JCM. The Rotterdam Study: Objectives and design update. *Eur J Epidemiol.* 2007;22:819–829.
  23. Ikram MA, Brusselle GGO, Murad SD, van Duijn CM, Franco OH, Goedegebure A, Klaver CCW, Nijsten TEC, Peeters RP, Stricker BH, Tiemeier H, Uitterlinden AG, Vernooij MW,

- Hofman A. The Rotterdam Study: 2018 update on objectives, design and main results. *Eur J Epidemiol.* 2017;32:807–850.
24. Ikram MA, Brusselle G, Ghanbari M, Goedegebure A, Ikram MK, Kavousi M, Kieboom BCT, Klaver CCW, de Kneegt RJ, Luik AI, Nijsten TEC, Peeters RP, van Rooij FJA, Stricker BH, Uitterlinden AG, Vernooij MW, Voortman T. Objectives, design and main findings until 2020 from the Rotterdam Study. Springer Netherlands; 2020.
  25. Pausova Z, Paus T, Abrahamowicz M, Bernard M, Gaudet D, Leonard G, Peron M, Pike GB, Richer L, Séguin JR, Veillette S. Cohort Profile: The Saguenay Youth Study (SYS). *Int J Epidemiol.* 2017;46:e19.
  26. John U, Hensel E, Lüdemann J, Piek M, Sauer S, Adam C, Born G, Alte D, Greiser E, Haertel U, Hense HW, Haerting J, Willich S, Kessler C. Study of Health in Pomerania (SHIP): A health examination survey in an east German region: Objectives and design. *Soz Präventivmed.* 2001;46:186–194.
  27. Völzke H, Alte D, Schmidt CO, Radke D, Lorbeer R, Friedrich N, Aumann N, Lau K, Piontek M, Born G, Havemann C, Ittermann T, Schipf S, Haring R, Baumeister SE, Wallaschofski H, Nauck M, Frick S, Arnold A, Jünger M, Mayerle J, Kraft M, Lerch MM, Dörr M, Reffellmann T, Empen K, Felix SB, Obst A, Koch B, Gläser S, Ewert R, Fietze I, Penzel T, Dören M, Rathmann W, Haerting J, Hannemann M, Röpcke J, Schminke U, Jürgens C, Tost F, Rettig R, Kors JA, Ungerer S, Hegenscheid K, Kühn JP, Kühn J, Hosten N, Puls R, Henke J, Gloger O, Teumer A, Homuth G, Völker U, Schwahn C, Holtfreter B, Polzer I, Kohlmann T, Grabe HJ, Roskopf D, Kroemer HK, Kocher T, Biffar R, John U, Hoffmann W. Cohort profile: The study of health in Pomerania. *Int J Epidemiol.* 2011;40:294–307.
  28. Jones S, Tillin T, Park C, Williams S, Rapala A, Al Saikhan L, Eastwood S V, Richards M, Hughes AD, Chaturvedi N. Cohort profile update: Southall and Brent revisited (SABRE) study: a UK population-based comparison of cardiovascular disease and diabetes in people of European, South Asian and African Caribbean heritage. *Int J Epidemiol.* 2020;1441–1442.
  29. Würtz P, Kangas AJ, Soinen P, Lawlor DA, Davey Smith G, Ala-Korpela M. Quantitative Serum Nuclear Magnetic Resonance Metabolomics in Large-Scale Epidemiology: A Primer on -Omic Technologies. *Am J Epidemiol.* 2017;186:1084–1096.
  30. Dona AC, Jiménez B, Schafer H, Humpfer E, Spraul M, Lewis MR, Pearce JTM, Holmes E, Lindon JC, Nicholson JK. Precision high-throughput proton NMR spectroscopy of human urine, serum, and plasma for large-scale metabolic phenotyping. *Anal Chem.* 2014;86:9887–9894.
  31. Römisch-Margl W, Prehn C, Bogumil R, Röhring C, Suhre K, Adamski J. Procedure for tissue sample preparation and metabolite extraction for high-throughput targeted metabolomics. *Metabolomics.* 2012;8:133–142.
  32. Jiménez B, Holmes E, Heude C, Tolson RF, Harvey N, Lodge SL, Chetwynd AJ, Cannet C, Fang F, Pearce JTM, Lewis MR, Viant MR, Lindon JC, Spraul M, Schäfer H, Nicholson JK. Quantitative Lipoprotein Subclass and Low Molecular Weight Metabolite Analysis in Human Serum and Plasma by 1H NMR Spectroscopy in a Multilaboratory Trial. *Anal Chem.* 2018;90:11962–11971.
  33. Harris SE, Ritchie SJ, Correia GDS, Jiménez B, Fawns-Ritchie C, Pattie A, Corley J, Maniega SM, Hernández MV, Starr JM, Hill D, Wren P, Bastin ME, Lewis MR, Wardlaw JM, Deary IJ. Plasma lipid and lipoprotein biomarkers in LBC1936: Do they predict general cognitive ability and brain structure? *bioRxiv.* 2020;1–30.
  34. Benjamini Y, Hochberg Y. Controlling the False Discovery Rate: A Practical and Powerful Approach to Multiple Testing. *J R Stat Soc Ser B.* 1995;57:289–300.
  35. R Core Team. R: A language and environment for statistical computing. 2021;URL <https://www.R-project.org/>.
  36. Wishart DS, Feunang YD, Marcu A, Guo AC, Liang K, Vázquez-Fresno R, Sajed T, Johnson

- D, Li C, Karu N, Sayeeda Z, Lo E, Assempour N, Berjanskii M, Singhal S, Arndt D, Liang Y, Badran H, Grant J, Serra-Cayuela A, Liu Y, Mandal R, Neveu V, Pon A, Knox C, Wilson M, Manach C, Scalbert A. HMDB 4.0: The human metabolome database for 2018. *Nucleic Acids Res.* 2018;46:D608–D617.
37. Würtz P, Havulinna AS, Soininen P, Tynkkynen T, Prieto-Merino D, Tillin T, Ghorbani A, Artati A, Wang Q, Tiainen M, Kangas AJ, Kettunen J, Kaikkonen J, Mikkilä V, Jula A, Kähönen M, Lehtimäki T, Lawlor DA, Gaunt TR, Hughes AD, Sattar N, Illig T, Adamski J, Wang TJ, Perola M, Ripatti S, Vasani RS, Raitakari OT, Gerszten RE, Casas JP, Chaturvedi N, Ala-Korpela M, Salomaa V. Metabolite profiling and cardiovascular event risk: A prospective study of 3 population-based cohorts. *Circulation.* 2015;131:774–785.
  38. Würtz P, Raiko JR, Magnussen CG, Soininen P, Kangas AJ, Tynkkynen T, Thomson R, Laatikainen R, Savolainen MJ, Laurikka J, Kuukasjärvi P, Tarkka M, Karhunen PJ, Jula A, Viikari JS, Kähönen M, Lehtimäki T, Juonala M, Ala-Korpela M, Raitakari OT. High-throughput quantification of circulating metabolites improves prediction of subclinical atherosclerosis. *Eur Heart J.* 2012;33:2307–2316.
  39. Jassal B, Matthews L, Viteri G, Gong C, Lorente P, Fabregat A, Sidiropoulos K, Cook J, Gillespie M, Haw R, Loney F, May B, Milacic M, Rothfels K, Sevilla C, Shamovsky V, Shorsler S, Varusai T, Weiser J, Wu G, Stein L, Hermjakob H, D'Eustachio P. The reactome pathway knowledgebase. *Nucleic Acids Res.* 2020;48:D498–D503.
  40. Jones DP, Mason HS. Metabolic hypoxia: accumulation of tyrosine metabolites in hepatocytes at low pO<sub>2</sub>. *Biochem Biophys Res Commun.* 1978;80:477–83.
  41. Sosa SM, Smith KJ. Understanding a role for hypoxia in lesion formation and location in the deep and periventricular white matter in small vessel disease and multiple sclerosis. *Clin Sci.* 2017;131:2503–2524.
  42. Ouzzine M, Gulberti S, Ramalanjaona N, Magdalou J, Fournel-Gigleux S. The UDP-glucuronosyltransferases of the blood-brain barrier: Their role in drug metabolism and detoxication. *Front Cell Neurosci.* 2014;8:1–12.
  43. Eelen G, de Zeeuw P, Treps L, Harjes U, Wong BW, Carmeliet P. Endothelial cell metabolism. *Physiol Rev.* 2018;98:3–58.
  44. Jeerakathil T, Wolf PA, Beiser A, Massaro J, Seshadri S, D'Agostino RB, DeCarli C. Stroke risk profile predicts white matter hyperintensity volume: The Framingham study. *Stroke.* 2004;35:1857–1861.
  45. Scharf EL, Graff-Radford J, Przybelski SA, Lesnick TG, Mielke MM, Knopman DS, Preboske GM, Schwarz CG, Senjem ML, Gunter JL, Machulda M, Kantarci K, Petersen RC, Jack CR, Vemuri P. Cardiometabolic Health and Longitudinal Progression of White Matter Hyperintensity: The Mayo Clinic Study of Aging. *Stroke.* 2019;50:3037–3044.
  46. Marathe GK, Pandit C, Lakshmikanth CL, Chaithra VH, Jacob SP, D'Souza CJM. To hydrolyze or not to hydrolyze: the dilemma of platelet-activating factor acetylhydrolase. *J Lipid Res.* 2014;55:1847–1854.
  47. Yan JJ, Jung JS, Lee JEJ, Lee JEJ, Huh SO, Kim HS, Jung KC, Cho JY, Nam JS, Suh HW, Kim YH, Song DK. Therapeutic effects of lysophosphatidylcholine in experimental sepsis. *Nat Med.* 2004;10:161–167.
  48. Sevastou I, Kaffe E, Mouratis MA, Aidinis V. Lysoglycerophospholipids in chronic inflammatory disorders: The PLA<sub>2</sub>/LPC and ATX/LPA axes. *Biochim Biophys Acta - Mol Cell Biol Lipids.* 2013;1831:42–60.
  49. Ousman SS, David S. Rapid Recruitment and Activation of Macrophages In the Adult Mouse Spinal Cord. *Glia.* 2000;104:92–104.
  50. Ganna A, Salihovic S, Sundström J, Broeckling CD, Hedman ÅK, Magnusson PKE, Pedersen

- NL, Larsson A, Siegbahn A, Zilmer M, Prenni J, Ärnlöv J, Lind L, Fall T, Ingelsson E. Large-scale Metabolomic Profiling Identifies Novel Biomarkers for Incident Coronary Heart Disease. *PLoS Genet.* 2014;10.
51. Mapstone M, Cheema AK, Fiandaca MS, Zhong X, Mhyre TR, Macarthur LH, Hall WJ, Fisher SG, Peterson DR, Haley JM, Nazar MD, Rich SA, Berlau DJ, Peltz CB, Tan MT, Kawas CH, Federoff HJ. Plasma phospholipids identify antecedent memory impairment in older adults. *Nat Med.* 2014;20:415–418.
  52. Bazinet RP, Layé S. Polyunsaturated fatty acids and their metabolites in brain function and disease. *Nat Rev Neurosci.* 2014;15:771–785.
  53. Chen CT, Kitson AP, Hopperton KE, Domenichiello AF, Trépanier MO, Lin LE, Ermini L, Post M, Thies F, Bazinet RP. Plasma non-esterified docosahexaenoic acid is the major pool supplying the brain. *Sci Rep.* 2015;5:1–12.
  54. Sugasini D, Thomas R, Yalagala PCR, Tai LM, Subbaiah P V. Dietary docosahexaenoic acid (DHA) as lysophosphatidylcholine, but not as free acid, enriches brain DHA and improves memory in adult mice. *Sci Rep.* 2017;7:1–11.
  55. Zöller I, Meixner M, Hartmann D, Büssow H, Meyer R, Gieselmann V, Eckhardt M. Absence of 2-hydroxylated sphingolipids is compatible with normal neural development but causes late-onset axon and myelin sheath degeneration. *J Neurosci.* 2008;28:9741–9754.
  56. Stadelmann C, Timmler S, Barrantes-Freer A, Simons M. Myelin in the central nervous system: Structure, function, and pathology. *Physiol Rev.* 2019;99:1381–1431.
  57. Mielke MM, Bandaru VVR, Haughey NJ, Rabins P V., Lyketsos CG, Carlson MC. Serum sphingomyelins and ceramides are early predictors of memory impairment. *Neurobiol Aging.* 2010;31:17–24.
  58. Beisiegel U, Heeren J. Lipoprotein lipase (EC 3.1.1.34) targeting of lipoproteins to receptors. *Proc Nutr Soc.* 1997;56:731–737.
  59. Lillis AP, Mikhailenko I, Strickland DK. Beyond endocytosis: LRP function in cell migration, proliferation and vascular permeability. *J Thromb Haemost.* 2005;3:1884–1893.
  60. Wardlaw JM, Makin SJ, Valdés Hernández MC, Armitage PA, Heye AK, Chappell FM, Muñoz-Maniega S, Sakka E, Shuler K, Dennis MS, Thrippleton MJ. Blood-brain barrier failure as a core mechanism in cerebral small vessel disease and dementia: evidence from a cohort study. *Alzheimer's Dement.* 2017;13:634–643.
  61. Muilwijk M, Callender N, Goorden S, Vaz FM, van Valkengoed IGM. Sex differences in the association of sphingolipids with age in Dutch and South-Asian Surinamese living in Amsterdam, the Netherlands. *Biol Sex Differ.* 2021;12:1–14.
  62. Netto CA, Sanches E, Odorcyk FK, Duran-Carabali LE, Weis SN. Sex-dependent consequences of neonatal brain hypoxia-ischemia in the rat. *J Neurosci Res.* 2017;95:409–421.
  63. Demarest TG, Schuh RA, Waddell J, McKenna MC, Fiskum G. Sex-dependent mitochondrial respiratory impairment and oxidative stress in a rat model of neonatal hypoxic-ischemic encephalopathy. *J Neurochem.* 2016;137:714–729.
  64. Van Den Heuvel DMJ, Admiraal-Behloul F, Ten Dam VH, Olofsen H, Bollen ELEM, Murray HM, Blauw GJ, Westendorp RGJ, De Craen AJM, Van Buchem MA. Different progression rates for deep white matter hyperintensities in elderly men and women. *Neurology.* 2004;63:1699–1701.
  65. Ammirati E, Moroni F, Magnoni M, Rocca MA, Anzalone N, Cacciaguerra L, Di Terlizzi S, Villa C, Sizzano F, Palini A, Scotti I, Besana F, Spagnolo P, Rimoldi OE, Chiesa R, Falini A, Filippi M, Camici PG. Progression of brain white matter hyperintensities in asymptomatic patients with carotid atherosclerotic plaques and no indication for revascularization. *Atherosclerosis.* 2019;287:171–178.

66. Jiménez-Sánchez Lorena, Hamilton Olivia K, Backhouse Ellen V, Clancy Una, Steward Catriona R, Wardlaw Joanna M. Sex differences in cerebral small vessel disease: a systematic review and meta-analysis. *medRxiv*. 2021;1–21.
67. Stanhewicz AE, Wenner MM, Stachenfeld NS. Sex differences in endothelial function important to vascular health and overall cardiovascular disease risk across the lifespan. *Am J Physiol - Hear Circ Physiol*. 2018;315:H1569–H1588.
68. Cerghet M, Skoff RP, Swamydas M, Bessert D. Sexual dimorphism in the white matter of rodents. *J Neurol Sci*. 2009;286:76–80.
69. Pesaresi M, Soon-Shiong R, French L, D.R. Kaplan, Miller FD, Paus T. Axon diameter and axonal transport: In vivo and in vitro effects of androgens. *Neuroimage*. 2015;115:191–201.
70. Howard VJ, Madsen TE, Kleindorfer DO, Judd SE, Rhodes JD, Soliman EZ, Kissela BM, Safford MM, Moy CS, McClure LA, Howard G, Cushman M. Sex and Race Differences in the Association of Incident Ischemic Stroke with Risk Factors. *JAMA Neurol*. 2019;76:179–186.
71. Peters SAE, Huxley RR, Woodward M. Diabetes as a risk factor for stroke in women compared with men: A systematic review and meta-analysis of 64 cohorts, including 775 385 individuals and 12 539 strokes. *Lancet*. 2014;383:1973–1980.
72. Madsen TE, Howard G, Kleindorfer DO, Furie KL, Oparil S, Manson JE, Liu S, Howard VJ. Sex Differences in Hypertension and Stroke Risk in the REGARDS Study: A Longitudinal Cohort Study. *Hypertension*. 2019;74:749–755.
73. Madsen TE, Khoury JC, Leppert M, Alwell K, Moomaw CJ, Sucharew H, Woo D, Ferioli S, Martini S, Adeoye O, Khatri P, Flaherty M, De Los Rios La Rosa F, MacKey J, Mistry E, Demel SL, Coleman E, Jasne A, Slavin SJ, Walsh K, Star M, Broderick JP, Kissela BM, Kleindorfer DO. Temporal Trends in Stroke Incidence over Time by Sex and Age in the GCNKSS. *Stroke*. 2020;1070–1076.
74. Peters SAE, Carcel C, Millett ERC, Woodward M. Sex differences in the association between major risk factors and the risk of stroke in the UK Biobank cohort study. *Neurology*. 2020;95:e2715–e2726.
75. Wardlaw JM, Smith EE, Biessels GJ, Cordonnier C, Fazekas F, Frayne R, Lindley RI, O'Brien JT, Barkhof F, Benavente OR, Black SE, Brayne C, Breteler M, Chabriat H, DeCarli C, de Leeuw FE, Doubal F, Duering M, Fox NC, Greenberg S, Hachinski V, Kilimann I, Mok V, Oostenbrugge R van, Pantoni L, Speck O, Stephan BCM, Teipel S, Viswanathan A, Werring D, Chen C, Smith C, van Buchem M, Norrving B, Gorelick PB, Dichgans M. Neuroimaging standards for research into small vessel disease and its contribution to ageing and neurodegeneration. *Lancet Neurol*. 2013;12:822–838.
76. Wardlaw J, Bath PMW, Doubal F, Heye A, Sprigg N, Woodhouse LJ, Blair G, Appleton J, Cvoro V, England T, Hassan A, John Werring D, Montgomery A. Protocol: The Lacunar Intervention Trial 2 (LACI-2). A trial of two repurposed licenced drugs to prevent progression of cerebral small vessel disease. *Eur Stroke J*. 2020;2.

## **Acknowledgements**

SYS: The authors thank the following individuals for their contributions in data acquisition and storage in the SYS: Manon Bernard (database architect, The Hospital for Sick Children) and H el ene Simard (C EGEP de Jonqui ere). The authors thank all participants who took part in the Saguenay Youth Study.

Support in part by the National Center for Advancing Translational Sciences, CTSI grant UL1TR001881, and the National Institute of Diabetes and Digestive and Kidney Disease Diabetes Research Center (DRC) grant DK063491 to the Southern California Diabetes Endocrinology Research Center.

Insight 46: The authors thank the study members who helped in the design of the study and the LHA and Insight 46 study teams for their contributions in data acquisition, analysis and storage.

We thank the LBC1936 participants and team members who contributed to these studies.

The Rotterdam Study is funded by Erasmus Medical Center and Erasmus University, Rotterdam, Netherlands Organization for the Health Research and Development (ZonMw), the Research Institute for Diseases in the Elderly (RIDE), the Ministry of Education, Culture and Science, the Ministry for Health, Welfare and Sports, the European Commission (DG XII), and the Municipality of Rotterdam. All study participants, general practitioners, specialists, researchers, institutions and funders of all other studies from this thesis are appreciatively acknowledged. Metabolomics measurements of Nightingales platform were funded by Biobanking and Biomolecular Resources Research Infrastructure (BBMRI)–NL(184.021.007).

## **Funding**

SYS: The Saguenay Youth Study has been funded by the Canadian Institutes of Health Research, Heart and Stroke Foundation of Canada, Canadian Foundation for Innovation, and National Institutes for Health.

Insight 46 is principally funded by grants from Alzheimer’s Research UK, the Medical Research Council Dementias Platform UK and the Wolfson Foundation, and the British Heart Foundation. The National Survey of Health and Development is funded by the Medical Research Council). JS is supported by the UCL/ UCLH NIHR Biomedical Research Centre.

The LBC1936 is supported by Age UK (Disconnected Mind project, which supports S.E.H.), the Medical Research Council (MRC) (G0701120, G1001245, MR/M013111/1, MR/R024065/1, which supports S.R.C.), and the University of Edinburgh. MRI brain imaging was supported by MRC grants G0701120, G1001245, MR/M013111/1 and MR/R024065/1. Metabolomics were supported by the Dementia Platform UK, MRC grant MR/L023784/2, the MRC & National Institute for Health Research [grant number MC\_PC\_12025] and infrastructure support was provided by the National Institute for Health Research (NIHR) Imperial Biomedical Research Centre (BRC).

Support in part by the National Center for Advancing Translational Sciences, CTSI grant UL1TR001881, and the National Institute of Diabetes and Digestive and Kidney Disease Diabetes Research Center (DRC) grant DK063491 to the Southern California Diabetes Endocrinology Research Center.

### **Author contributions**

SS and ZP conceptualized the study. ES, JS, and ZP contributed to the analysis plan. ES, JS, SA, DMW, SF, FG, SEH, AKH, YHH, RW, and KW analyzed the data and generated results. ES and ZP interpreted the results and wrote the original manuscript. QY, HV, JMS, MR, MN, LL, MAI, NH, HG, IJD, SRC, JMW, MRL, BJ, CS, MWV, PP, MG, JB, MVH, NC, JIR, MF, TP, SS, and ZP obtained funding, provided additional study resources, and contributed to metabolic and/or MRI data. All authors contributed to revising the content and approved the final version.