

The Many Roles of Cholesterol in Sepsis: A Review

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MS conceived the idea for the article. DH performed the initial literature search and drafted the first version of the manuscript. DH, AK, AP, MB and MS revised subsequent drafts of the manuscript. All authors have approved the final version of the manuscript.

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The authors are performing laboratory studies investigating the potential of cholesterol as a therapy for sepsis and have no current commercial conflicts.

Abstract

Rationale: The biological functions of cholesterol are diverse, ranging from cell membrane integrity and signalling, immunity, to the synthesis of steroid and sex hormones, Vitamin D, bile acids and oxysterols. Multiple studies have demonstrated hypocholesterolemia in sepsis, the degree of which is an excellent prognosticator of poor outcomes. However, the clinical significance of hypocholesterolemia has been largely unrecognized.

Objectives/Methods: We undertook a detailed review of the biological roles of cholesterol, the impact of sepsis, its reliability as a prognosticator in sepsis, and the potential utility of cholesterol as a treatment.

Measurements and Main Results: Sepsis affects cholesterol synthesis, transport and metabolism. This likely impacts upon its biological functions including immunity, hormone and vitamin production, and cell membrane receptor sensitivity. Early preclinical studies show promise for cholesterol as a pleiotropic therapeutic agent.

Conclusions: Hypocholesterolemia is a frequent condition in sepsis and an important early prognosticator. Low plasma levels are associated with wider changes in cholesterol metabolism and its functional roles, and these appear to play a significant role in sepsis pathophysiology. The therapeutic impact of cholesterol elevation warrants further investigation.

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Sepsis, cholesterol, hypocholesterolemia, lipid metabolism

Introduction

Sepsis, the dysregulated host response to infection resulting in organ dysfunction [1], is a major worldwide cause of mortality [2] and morbidity. Current management focuses on adequate fluid resuscitation, organ support, and treating the infection with antibiotics and source control. To date, no available treatments that directly target underlying pathophysiological mechanisms have been clearly demonstrated to improve outcomes.

Cholesterol, a sterol lipid, plays an integral role in multiple body functions including maintenance of cellular membrane processes, immunity, signalling, pathway regulation, and as a precursor for the synthesis of steroid hormones, Vitamin D, bile acids and oxysterols. Sepsis-induced hypocholesterolemia was first recognized a century ago [3]; multiple studies demonstrate a worse prognosis associated with the magnitude of decline. However, mechanisms by which plasma levels fall, the impact on organ functionality, the relationship of plasma cholesterol to intracellular concentrations, and the potential role of cholesterol as a therapeutic all require elucidation.

There is increasing interest in the therapeutic possibilities of lipoproteins and modulation of cholesterol transport in sepsis, particularly in immune-inflammatory modulation and pathogen scavenging. There has, however, been little focus on cholesterol itself rather than its carriers. In this article, we provide an overview of the biology of cholesterol, its possible roles in sepsis pathophysiology, and its potential utility as a specific adjunctive treatment.

Cholesterol synthesis, structure, metabolism and functional roles

Cholesterol consists of four linked aromatic hydrophobic rings, a small hydrophilic hydroxyl group, and a hydrophobic chain. Due to its high hydrophobicity, cholesterol is only present within cells predominantly as a component of lipid membranes or bound to lipid-binding proteins [4] (Figure 1). Animals obtain cholesterol through diet and, primarily, by endogenous synthesis. Cholesterol synthesis is a multistep (~30 reaction) process that is highly energy-consuming; synthesis of one cholesterol molecule requires 18 acetyl-CoA, 36 ATP, 16 NADPH and 11 oxygen molecules. Endogenous cholesterol synthesis is tightly regulated by negative feedback (Figure 2). Hydroxymethylglutaryl-coenzyme A reductase (HMG-CoA reductase), the target of statin therapy, is the rate-limiting enzyme within the pathway and the predominant mechanism by which cells adapt to changes in cholesterol bioavailability.

To enable transport in plasma, cholesterol must be bound to lipoproteins or albumin. Lipoproteins are categorized into chylomicrons, chylomicron remnants, very-low (VLDL), low (LDL) and high-density (HDL) lipoprotein by density, size and the type of particle-forming and other associated proteins. Cholesterol bound to LDL is transported from liver to peripheral tissues whereas HDL carries cholesterol to the liver and steroidogenic tissues - "*reverse cholesterol transport*" [4]. Mammalian cells lack an enzyme system to catabolize and recycle cholesterol and its derivatives. The liver clears cholesterol from the circulation via LDL and HDL receptors [5]. It is then metabolized or excreted either unmodified or as bile acids, a large proportion of which is reabsorbed.

Cholesterol and its metabolites provide multiple biological functions (Figure 3):

(i) Cholesterol is an integral part of cell membranes, and plays a crucial role in modulating membrane thickness, permeability, fluidity and functionality [6, 7]. Within the

membrane, cholesterol distributes non-homogeneously, accumulating within lipid rafts. These small, highly dynamic, sterol- and sphingolipid-enriched membrane micro-domains attract many transmembrane proteins such as ion channels, transporters and receptors, including G-protein coupled receptors (GPCRs) [7]. Alterations in membrane cholesterol affect the membrane's physical properties and influence the presence and activity of transmembrane proteins such as the sodium-potassium-ATPase and β -adrenergic receptors [7].

(ii) Both cholesterol and its lipoprotein carriers have immunomodulatory properties including binding of endotoxin and other toxins [8, 9]. This scavenging mechanism may play an important role in neutralizing toxins as part of the innate immune system response, preventing activation of Toll-like receptors (TLRs) by pathogen-associated molecular patterns (PAMPs). Of note, key receptors regulating the immune response such as Toll-like receptors and T- and B-cell receptors are localized within lipid rafts [10].

(iii) Cholesterol is the only steroidogenic substrate used to synthesize adrenocortical (glucocorticoids, aldosterone) and sex hormones (e.g. estrogen, progesterone, testosterone) and vitamin D through multi-step processes [11]. During a triggered stress response, approximately 80% of circulating cortisol may be derived from plasma cholesterol [12].

The impact of Vitamin D on multiple diseases, including musculoskeletal disorders, insulin resistance and metabolic syndrome, and on cardiovascular and immunological dysfunction has been studied extensively [13].

(iv) Conversion of cholesterol to bile acids involves 17 distinct enzymatic steps within hepatocytes and is the principal route of cholesterol metabolism. Bile acids undergo

enterohepatic recirculation, allowing recycling with *de novo* hepatocyte synthesis compensating for physiological intestinal losses. Bile acids aid metabolite excretion by the liver, absorption of lipids, hydrophobic nutrients and fat-soluble vitamins, and prevent bacterial overgrowth within the small bowel and biliary tree. They also regulate multiple functions within various liver cell types, e.g. cell differentiation and regeneration [14].

(v) Oxysterols represent a large family of oxidized derivatives of cholesterol with multiple biological actions, including immunomodulation [15]. Cholesterol can be oxidized either enzymatically or non-enzymatically by reactive oxygen species. Oxysterols can exert their functions through GPCRs, nuclear receptors and other molecular pathways, regulating many processes from cytokine production to virus entry into cells [16, 17]. Oxysterols modulate neutrophil, B- and T-cell functionality, enhance innate immunity and regulate production of the anti-inflammatory cytokine, IL-10 [17, 18].

Cholesterol levels fall during sepsis, in line with severity and outcome

Reductions in total plasma cholesterol, high-density (HDL-C) and low-density lipoprotein cholesterol (LDL-C) are well recognized in sepsis [19-28]. Levels are decreased at the time of diagnosis [21] and often decline further during the disease course [25]. Serum HDL-C levels reach a nadir around day 3 post-admission, whereas LDL-C is lowest at the time of diagnosis [21]. Variable recovery in serum levels occurs over subsequent days [25]. The kinetics of VLDL-C in sepsis are poorly characterized in human sepsis.

Multiple studies report a greater mortality risk in patients with lower levels of total, HDL- and LDL-cholesterol [23-28]. Of note, a recent genetic study suggested that low LDL levels in sepsis may be associative rather than causal of an increased mortality risk [27]

while low HDL cholesterol may be a causal factor [29]. Increased LDL clearance may contribute to a lower sepsis mortality via enhanced pathogen lipid clearance [27].

Survivors show a slow return to almost normal values over the disease course. The magnitude of fall is associated with a higher incidence of multi-organ dysfunction, an increased duration of ICU stay and more nosocomial infection [23, 26]. Elevated serum markers of inflammation correlate negatively with cholesterol levels [20, 24, 28].

Infusion of recombinant TNF-alpha or IL-6 into cancer patients also produced large falls in plasma cholesterol in inverse correlation to markers of inflammation [30, 31]. Animal experiments can replicate these findings and can be used as a therapeutic test bed. However, this is model-dependent as some rodent models injected with endotoxin or TNF-alpha actually demonstrate hypercholesterolemia [32]. However, we and others have found large falls in total and HDL cholesterol levels in rats given a more realistic peritonitis insult [33-35]. Hypocholesterolemia has also been demonstrated in septic models using primates, sheep and dogs [36-38].

Why does serum cholesterol fall in sepsis?

Biological mechanisms leading to hypocholesterolemia in sepsis remain incompletely understood. Apart from decreased intake and impaired intestinal absorption of fat in critical illness [39], decreased synthesis, impaired cholesterol transport, increased metabolism and depletion through toxin scavenging may be implicated.

Data on the impact of sepsis on cholesterol synthesis are limited and conflicting. Old studies in rodent models reported increased hepatic cholesterogenesis [32, 40] and concurrent hypercholesterolemia [32]. Vasconcelos et al however noted a decrease in HMG-

CoA reductase activity compared to healthy, fed rats [40]. Our currently unpublished data reveal decreased expression of transcriptional regulators (SREBP-1, SREBP-2, INSIG) and enzymes (HMG-CoA reductase) within the hepatic cholesterol synthesis pathway in our rat peritonitis model.

Pro-inflammatory cytokines may contribute to hypocholesterolemia by reducing hepatic synthesis of apolipoproteins that bind cholesterol to form lipoproteins [41]. Falls in plasma LDL-C are commonly but variably reported whereas low HDL-C is a consistent finding. Those changes suggest reverse cholesterol transport, i.e. transfer of cholesterol from peripheral tissues to the liver, may be more affected [19]. Figure 4 illustrates different cholesterol metabolic and transfer pathways affected by sepsis. Transporters (e.g. the ATP-binding cassette (ABC) transporter superfamily which transforms lipid-poor apolipoprotein A1 (apoA-1) particles into mature HDL particles) and enzymes such as lecithin-cholesterol acyltransferase (LCAT), which converts free cholesterol to more hydrophobic cholesterol esters enabling incorporation into HDL, are affected by sepsis [22, 41]. The binding capacity of HDL is also affected by alterations in its structure and protein composition, and by accumulation of oxidized lipids [42].

Cholesteryl ester transfer protein (CETP) mediates triglyceride and cholesteryl ester transfer between triglyceride-rich lipoproteins and HDL particles, with lower plasma CETP levels increasing the proportion of HDL cholesterol. However, total circulating cholesterol levels are unaffected [43]. Literature on the relevance of changes in plasma CETP levels in sepsis and relationship to outcomes is conflicting [29, 44-46].

Similarly, conflicting patient data are seen with regard to alterations in plasma proprotein convertase subtilisin kexin 9 (PCSK9) levels, an enzyme that degrades hepatic LDL and adipocyte VLDL receptors, resulting in hypercholesterolemia [47-49].

Cholesterol metabolism can be increased in sepsis by enzymatic and non-enzymatic oxidation. Cholesterol-25-hydroxylase is strongly induced by inflammation and its product, 25-hydroxycholesterol [50]. The acute-phase protein phospholipase A2 (PLA2) rises during inflammation and promotes increased metabolism of cholesterol esters and apolipoproteins, thereby reducing serum cholesterol [51]. PLA2 activity is enhanced by another acute-phase reactant, serum amyloid A (SAA), which also affects cholesterol transport [52]. Sepsis however decreases bile flow [53]. Impaired biotransformation and hepatobiliary transport of bile acids occur within hours of induction of polymicrobial sepsis [54]. As a consequence, bile acids can be elevated in the blood compartment.

Impact of sepsis on the biological roles of cholesterol

As described earlier, cholesterol and its various metabolites exert many complex biological functions, many of which are disrupted during sepsis. The specific contribution of cholesterol deficiency to these abnormalities requires further elucidation, but there is sufficient direct and circumstantial evidence to suggest cholesterol deficiency may play an important role.

- *Cell membrane function*

The cholesterol composition within lipid rafts modifies intrinsic function and downstream signaling, such as the adrenergic receptor pathway. Cholesterol depletion in human neutrophil cell membranes induced a more pro-inflammatory phenotype including

priming, enhanced activation, increased adhesion and oxidant production [55, 56]. Raft-dependent signaling of multiple cell types may be altered due to changes in membrane cholesterol levels affecting, for example, GPCR density and activity [6, 7]. This may be of particular relevance in septic shock where myocardial and vascular hyporeactivity to exogenous catecholamines is a defining characteristic, with the magnitude of hyporesponsiveness associated with increased mortality [57].

- *Immunomodulatory and anti-bacterial properties of cholesterol*

Notwithstanding the scavenging and immunosuppressive roles of HDL and other lipoproteins, a low cholesterol may itself negatively impact on innate and adaptive immune cells [58]. Intracellular cholesterol plays a pivotal role in TLR signaling in macrophages [59]. The cholesterol concentration within membrane lipid rafts significantly impacts on raft levels of TLR-4 and -9 [59]. Depletion of the ABC-A1 transporter in knockout macrophages, impacting on intracellular cholesterol transport, was associated with enlarged, cholesterol-containing lipid rafts that were rich in TLR-4 and hyperresponsive to LPS [59]. In lymphocytes, enrichment of cholesterol in lipid rafts was associated with increased formation of an immune synapse between signalling complexes and T-cell receptors. Low serum and low membrane cholesterol concentrations also influence natural killer cell (NK cell) function [60].

- *Steroid, sex hormone and vitamin D deficiency*

Adrenal insufficiency is a recognized complication in patients with sepsis and septic shock and associated with increased mortality [61]. Even though plasma cortisol levels are frequently raised, there is decreased responsiveness to ACTH stimulation, particularly in

eventual non-survivors [62], suggesting the possibility of diminished reserves. As mentioned earlier, some 80% of circulating cortisol during stress is derived from plasma cholesterol [12]. The contribution of hypocholesterolemia in sepsis is uncertain as the downstream cortisol production pathway may also be compromised, e.g. expression of steroidogenic acute regulatory protein (StAR), the rate-limiting step in steroidogenesis which orchestrates transport of cholesterol from outer to inner mitochondrial membranes [63]. Pharmacological suppression of HDL-C does however disrupt adrenal steroidogenesis [64]. Nonetheless, human data are conflicting [65-67].

Falls in sex hormone [68] and vitamin D levels [69] are also well recognized in sepsis and carry prognostic and potential therapeutic implications. Pharmacological activation of the estrogen receptor-beta improved survival in pneumonia and peritonitis models of sepsis [70]. Administration of high-dose vitamin D to critically ill patients with severe vitamin D deficiency have produced conflicting outcomes [71, 72]. An association has been described between low cholesterol and low testosterone in male septic shock patients [73], however causation remains unclear. Low LDL-C levels have also been linked to low testosterone levels in chronically ill patients [68].

- *Bile acids*

Impaired biotransformation and hepatobiliary transport of bile acids occur within hours following induction of polymicrobial sepsis [54]. In septic patients, bile acids are significantly elevated and predictive of poor outcomes [74]. This appears to relate to diminished or even obstructed bile flow from liver rather than increased synthesis. To what extent changes in cholesterol levels in different body compartments during sepsis alter the complex mechanisms of bile acid metabolism remains to be elucidated.

Cholesterol supplementation and lipoprotein therapies

The idea of a lipid treatment for infection is not new, whether this be cholesterol, HDL or analogues, oxysterols or phospholipid emulsions. Indeed, Bayer took out a patent for cholesterol therapy for blackwater fever (malaria) in 1910. The possible impact of cholesterol therapy on a wide range of infectious diseases was suggested soon after [75].

Published studies remain relatively scanty and are often based on model systems. What benefit derives from the lipoprotein itself or from elevation of cholesterol levels is unclear.

Cholesterol nanoparticles elevated intracellular levels and prevented the cytotoxic effect of the pneumococcal antigen, pneumolysin on hepatocytes [76]. Administration of 25-hydroxycholesterol decreased viral load and improved outcomes in a porcine viral pneumonitis model [77]. In terms of carriers of cholesterol, intravenous application of reconstituted HDL or HDL mimetics (based on apolipoprotein A-1) reduced organ damage, improved hemodynamics and survival in a variety of septic or endotoxemic rodent models [35, 78-83]. Inhibition of CETP with anacetrapib preserved high-density lipoprotein cholesterol levels and improved survival in septic mice [46]. Pharmacological inhibition of PCSK9 has however delivered variable results. Whereas improved survival was noted in a murine polymicrobial peritonitis model [47], no protection was afforded in a murine endotoxin model [84].

Human studies are limited. Reconstituted HDL decreased proinflammatory cytokine release in human volunteer endotoxemia [85]. A multicenter study enrolling nearly 1400 patients with presumed Gram negative sepsis [86] reported that a 10% phospholipid-

lipoprotein emulsion that contained no cholesterol, given with the aim of neutralizing endotoxin, failed to deliver any benefit. A two-centre Phase I/II clinical protocol has been recently published [87] in which an anti-inflammatory lipid emulsion containing fish oil is being administered intravenously to septic patients with the objective of raising plasma cholesterol levels. The impact of cholesterol infusions on lipoprotein levels (HDL-C, LDL-C, VLDL-C) remains unknown. More experimental *in vitro* and *in vivo* studies are needed to address mechanisms, feasibility, dose finding and possible adverse events.

Statin therapy for sepsis – is there a paradox?

How can the above arguments related to cholesterol therapy be reconciled with the putative benefits of statins in critical illness, agents which are conventionally used to treat hypercholesterolemia? Statins inhibit HMG-CoA reductase, the rate-limiting enzyme in the mevalonate pathway which commences with acetyl CoA. This pathway later splits into branches that synthesize cholesterol, heme A, ubiquinone, dolichol and other isoprenoids. Statins also affect other pathways either directly related or not to mevalonate, such as endothelial NO synthase activation [88]. Thus, other than lowering cholesterol, statins have multiple other immunomodulatory, anti-inflammatory and metabolic effects such as activation of PPARs, increased production of endothelial NO, reduced synthesis of endothelin-1 and thromboxane A₂, and NADPH oxidase inactivation [88-90]. These may be both beneficial or harmful, e.g. statin-induced myopathy has been linked to reductions in ubiquinone and thus mitochondrial functionality or alterations in sarcolemma and/or membrane binding proteins [91]. The impact of statins on mortality in cardiovascular disease specifically related to cholesterol lowering is questioned [92].

With respect to sepsis, epidemiological studies reported an association with improved survival from sepsis in patients on pre-existing statin treatment, however, this likely relates to population lifestyle differences [93-95]. Two randomized controlled, multicenter trials found no benefit from *de novo* statin therapy in sepsis [96, 97]. Notably, plasma cholesterol levels were markedly subnormal in both atorvastatin and control groups (2.4 vs 2.6 mmol/l, respectively) [96]. The HARP-2 trial of patients with ARDS, of whom 40% had sepsis, showed no outcome effect from simvastatin [98]. Of note, a post-hoc analysis suggested patients with a hyperinflammatory phenotype could benefit [99], indicating non-cholesterol lowering effects may be more pertinent. Based on current evidence, we cannot recommend continuation or addition of statins in sepsis; prospective randomised studies are needed to clarify their potential utility in specific patient subsets.

Conclusions

Low cholesterol levels are a well-recognized manifestation of sepsis and septic shock. The magnitude of hypocholesterolemia relates to disease severity and outcome and is an early prognostic marker. Several pathophysiologic mechanisms can participate in the development of hypocholesterolemia in sepsis and its impact on multiple downstream biochemical pathways. Further studies are needed to extend our knowledge about the importance and interactions of these mechanisms and the role of cholesterol \pm lipoproteins as therapeutics.

References:

1. Singer M, Deutschman CS, Seymour CW, Shankar-Hari M, Annane D, Bauer M, Bellomo R, Bernard GR, Chiche JD, Coopersmith CM, Hotchkiss RS, Levy MM, Marshall JC, Martin GS, Opal SM, Rubenfeld GD, van der Poll T, Vincent JL, Angus DC. The Third International Consensus Definitions for Sepsis and Septic Shock (Sepsis-3). *JAMA* 2016; 315:801-810.
2. Rudd KE, Johnson SC, Agesa KM, Shackelford KA, Tsoi D, Kievlan DR, Colombara DV, Ikuta KS, Kissoon N, Finfer S, Fleischmann-Struzek C, Machado FR, Reinhart KK, Rowan K, Seymour CW, Watson RS, West TE, Marinho F, Hay SI, Lozano R, Lopez AD, Angus DC, Murray CJL, Naghavi M. Global, regional, and national sepsis incidence and mortality, 1990-2017: analysis for the global burden of disease study. *Lancet* 2020; 395:200-211.
3. Macadam W, Shiskin C. The Cholesterol Content of the Blood in Relation to Genito-urinary Sepsis. *Proc R Soc Med* 1924; 17:53-55.
4. Ikonen E. Cellular cholesterol trafficking and compartmentalization. *Nat Rev Mol Cell Biol* 2008; 9:125-138.
5. Shen WJ, Asthana S, Kraemer FB, Azhar S. Scavenger receptor B type 1: expression, molecular regulation, and cholesterol transport function. *J Lipid Res* 2018;59:1114-1131.
6. Jafurulla M, Chattopadhyay A. Membrane lipids in the function of serotonin and adrenergic receptors. *Curr Med Chem* 2013; 20:47-55.

7. Allen JA, Halverson-Tamboli RA, Rasenick MM. Lipid raft microdomains and neurotransmitter signalling. *Nat Rev Neurosci* 2007; 8:128-140.
8. Morin EE, Guo L, Schwendeman A, Li XA. HDL in sepsis - risk factor and therapeutic approach. *Front Pharmacol* 2015; 6:244.
9. Guo L, Ai J, Zheng Z, Howatt DA, Daugherty A, Huang B, Li XA. High density lipoprotein protects against polymicrobe-induced sepsis in mice. *J Biol Chem* 2013; 288:17947-17953.
10. Varshney P, Yadav V, Saini N. Lipid rafts in immune signalling: current progress and future perspective. *Immunology* 2016; 149:13-24.
11. Payne AH, Hales DB. Overview of steroidogenic enzymes in the pathway from cholesterol to active steroid hormones. *Endocr Rev* 2004; 25:947-970.
12. Borkowski AJ, Levin S, Delcroix C, Mahler A, Verhas V. Blood cholesterol and hydrocortisone production in man: quantitative aspects of the utilization of circulating cholesterol by the adrenals at rest and under adrenocorticotropin stimulation. *J Clin Invest* 1967; 46:797-811.
13. Bikle DD. Vitamin D metabolism, mechanism of action, and clinical applications. *Chem Biol* 2014; 21:319-329.
14. Marin JJ, Macias RI, Briz O, Banales JM, Monte MJ. Bile Acids in Physiology, Pathology and Pharmacology. *Curr Drug Metab* 2015; 17:4-29.
15. Mutemberezi V, Guillemot-Legris O, Muccioli GG. Oxysterols: From cholesterol metabolites to key mediators. *Prog Lipid Res* 2016; 64:152-169.

16. Spann NJ, Glass CK. Sterols and oxysterols in immune cell function. *Nat Immunol* 2013; 14:893-900.
17. Abrams ME, Johnson KA, Perelman SS, Zhang LS, Endapally S, Mar KB, Thompson BM, McDonald JG, Schoggins JW, Radhakrishnan A, Alto NM. Oxysterols provide innate immunity to bacterial infection by mobilizing cell surface accessible cholesterol. *Nat Microbiol* 2020; 5:929-942.
18. Perucha E, Melchiotti R, Bibby JA, Wu W, Frederiksen KS, Roberts CA, Hall Z, LeFric G, Robertson KA, Lavender P, Gerwien JG, Taams LS, Griffin JL, de Rinaldis E, van Baarsen LGM, Kemper C, Ghazal P, Cope AP. The cholesterol biosynthesis pathway regulates IL-10 expression in human Th1 cells. *Nat Commun* 2019; 10:498.
19. Golucci APBS, Marson FAL, Ribeiro AF, Nogueira RJN. Lipid profile associated with the systemic inflammatory response syndrome and sepsis in critically ill patients. *Nutrition* 2018; 55-56:7-14.
20. Levels JH, Lemaire LC, van den Ende AE, van Deventer SJ, van Lanschot JJ. Lipid composition and lipopolysaccharide binding capacity of lipoproteins in plasma and lymph of patients with systemic inflammatory response syndrome and multiple organ failure. *Crit Care Med* 2003; 31:1647-1653.
21. van Leeuwen HJ, Heezius EC, Dallinga GM, van Strijp JA, Verhoef J, van Kessel KP. Lipoprotein metabolism in patients with severe sepsis. *Crit Care Med* 2003; 31:1359-1366.

22. Levels JH, Pajkrt D, Schultz M, Hoek FJ, van Tol A, Meijers JC, van Deventer SJ. Alterations in lipoprotein homeostasis during human experimental endotoxemia and clinical sepsis. *Biochim Biophys Acta* 2007; 1771:1429-1438.
23. Cirstea M, Walley KR, Russell JA, Brunham LR, Genga KR, Boyd JH. Decreased high-density lipoprotein cholesterol level is an early prognostic marker for organ dysfunction and death in patients with suspected sepsis. *J Crit Care* 2017; 38:289-294.
24. Lekkou A, Mouzaki A, Siagris D, Ravani I, Gogos C. Serum lipid profile, cytokine production, and clinical outcome in patients with severe sepsis. *J Crit Care* 2014; 29:723-727.
25. Lee SH, Park MS, Park BH, Jung WJ, Lee IS, Kim SY, Kim EY, Jung JY, Kang YA, Kim YS, Kim SK, Chang J, Chung KS. Prognostic implications of serum lipid metabolism over time during sepsis. *Biomed Res Int* 2015; 2015:789298.
26. Chien JY, Jerng JS, Yu CJ, Yang PC. Low serum level of high-density lipoprotein cholesterol is a poor prognostic factor for severe sepsis. *Crit Care Med* 2005; 33:1688-1693.
27. Walley KR, Boyd JH, Kong HJ, Russell JA. Low low-density lipoprotein levels are associated with, but do not causally contribute to, increased mortality in sepsis. *Crit Care Med* 2019;47:463-466.
28. Vavrova L, Rychlikova J, Mrackova M, Novakova O, Zak A, Novak F. Increased inflammatory markers with altered antioxidant status persist after clinical recovery

- from severe sepsis: a correlation with low HDL cholesterol and albumin. *Clin Exp Med* 2016; 16:557-569.
29. Trinder M, Genga KR, Kong HJ, Blauw LL, Lo C, Li X, Cirstea M, Wang Y, Rensen PCN, Russell JA, Walley KR, Boyd JH, Brunham LR. Cholesteryl Ester Transfer Protein Influences High-Density Lipoprotein Levels and Survival in Sepsis. *Am J Respir Crit Care Med* 2019; 199:854-862.
30. Spriggs DR, Sherman ML, Michie H, Arthur KA, Imamura K, Wilmore D, Frei E, Kufe DW. Recombinant human tumor necrosis factor administered as a 24-hour intravenous infusion. A phase I and pharmacologic study. *J Natl Cancer Inst* 1988; 80:1039-1044.
31. van Gameren MM, Willemse PH, Mulder NH, Limburg PC, Groen HJ, Vellenga E, de Vries EG. Effects of recombinant human interleukin-6 in cancer patients: a phase I-II study. *Blood* 1994; 84:1434-1441.
32. Memon RA, Grunfeld C, Moser AH, Feingold KR. Tumor necrosis factor mediates the effects of endotoxin on cholesterol and triglyceride metabolism in mice. *Endocrinology* 1993; 132:2246-2253.
33. Hill NE, Saeed S, Phadke R, Ellis MJ, Chambers D, Wilson DR, Castells J, Morel J, Freysennet DG, Brett SJ, Murphy KG, Singer M. Detailed characterization of a long-term rodent model of critical illness and recovery. *Crit Care Med* 2015; 43:e84-96.
34. Morel J, Hargreaves I, Brealey D, Neergheen V, Backman JT, Lindig S, Bläss M, Bauer M, McAuley DF, Singer M. Simvastatin pre-treatment improves survival and mitochondrial function in a 3-day fluid-resuscitated rat model of sepsis. *Clin Sci* 2017; 131:747-758.

35. Moreira RS, Irigoyen M, Sanches TR, Volpini RA, Camara NO, Malheiros DM, Shimizu MH, Seguro AC, Andrade L. Apolipoprotein A-I mimetic peptide 4F attenuates kidney injury, heart injury, and endothelial dysfunction in sepsis. *Am J Physiol Regul Integr Comp Physiol* 2014; 307:R514-524.
36. Ettinger WH, Miller LD, Albers JJ, Smith TK, Parks JS. Lipopolysaccharide and tumor necrosis factor cause a fall in plasma concentration of lecithin: cholesterol acyltransferase in cynomolgus monkeys. *J Lipid Res* 1990; 31:1099-1107.
37. El-Deeb WM, Tharwat M. Lipoproteins profile, acute phase proteins, proinflammatory cytokines and oxidative stress biomarkers in sheep with pneumonic pasteurellosis. *Comp Clin Pathol* 2015; 24: 581–588.
38. Hardy JP, Streeter EM, DeCook RR. Retrospective evaluation of plasma cholesterol concentration in septic dogs and its association with morbidity and mortality: 51 cases (2005-2015). *J Vet Emerg Crit Care* 2018; 28:149-156.
39. Ali Abdelhamid Y, Cousins CE, Sim JA, Bellon MS, Nguyen NQ, Horowitz M, Chapman MJ, Deane AM. Effect of Critical Illness on Triglyceride Absorption. *JPEN J Parenter Enteral Nutr* 2015; 39:966-972.
40. de Vasconcelos PR, Kettlewell MG, Gibbons GF, Williamson DH. Increased rates of hepatic cholesterologenesis and fatty acid synthesis in septic rats in vivo: evidence for the possible involvement of insulin. *Clin Sci* 1989;76:205-211.
41. de la Llera Moya M, McGillicuddy FC, Hinkle CC, Byrne M, Joshi MR, Nguyen V, Tabita-Martinez J, Wolfe ML, Badellino K, Pruscino L, Mehta NN, Asztalos BF, Reilly MP.

- Inflammation modulates human HDL composition and function in vivo. *Atherosclerosis* 2012; 222:390-394.
42. Tall AR, Yvan-Charvet L. Cholesterol, inflammation and innate immunity. *Nat Rev Immunol* 2015; 15:104-116.
43. Brousseau ME, Schaefer EJ, Wolfe ML, Bloedon LT, Digenio AG, Clark RW, Mancuso JP, Rader DJ. Effects of an inhibitor of cholesteryl ester transfer protein on HDL cholesterol. *N Engl J Med* 2004;350:1505-1515.
44. Dusuel A, Deckert V, Pais de Barros JP, van Dongen K, Choubley H, Charron É, Le Guern N, Labbé J, Mandard S, Grober J, Lagrost L, Gautier T. Human cholesteryl ester transfer protein lacks lipopolysaccharide transfer activity, but worsens inflammation and sepsis outcomes in mice. *J Lipid Res* 2020;62:100011. (in press)
45. Grion CM, Cardoso LT, Perazolo TF, Garcia AS, Barbosa DS, Morimoto HK, Matsuo T, Carrilho AJ. Lipoproteins and CETP levels as risk factors for severe sepsis in hospitalized patients. *Eur J Clin Invest* 2010; 40:330-338.
46. Trinder M, Wang Y, Madsen CM, Ponomarev T, Bohunek L, Daisely BA, Julia Kong H, Blauw LL, Nordestgaard BG, Tybjaerg-Hansen A, Wurfel MM, Russell JA, Walley KR, Rensen PCN, Boyd JH, Brunham LR. Inhibition of cholesteryl ester transfer protein preserves high-density lipoprotein cholesterol and improves survival in sepsis. *Circulation* 2021; 143:921-934.

47. Walley KR, Thain KR, Russell JA, Reilly MP, Meyer NJ, Ferguson JF, Christie JD, Nakada TA, Fjell CD, Thair SA, Cirstea MS, Boyd JH. PCSK9 is a critical regulator of the innate immune response and septic shock outcome. *Sci Transl Med* 2014;6:258ra143.
48. Feng Q, Wei WQ, Chaugai S, Carranza Leon BG, Kawai V, Carranza Leon DA, Jiang L, Zhong X, Liu G, Ihegword A, Shaffer CM, Linton MF, Chung CP, Stein CM. A Genetic Approach to the Association Between PCSK9 and Sepsis. *JAMA Netw Open* 2019; 2:e1911130.
49. Vecchié A, Bonaventura A, Meessen J, Novelli D, Minetti S, Elia E, Ferrara D, Ansaldo AM, Scaravilli V, Villa S, Ferla L, Caironi P, Latini R, Carbone F, Montecucco F; ALBIOS Biomarkers Study Investigators. PCSK9 is associated with mortality in patients with septic shock: data from the ALBIOS study. *J Intern Med* 2021; 289:179-192.
50. Diczfalusy U, Olofsson KE, Carlsson AM, Gong M, Golenbock DT, Rooyackers O, Fläring U, Björkbacka H. Marked upregulation of cholesterol 25-hydroxylase expression by lipopolysaccharide. *J Lipid Res* 2009; 50:2258-2264.
51. Tietge UJ, Maugeais C, Cain W, Grass D, Glick JM, de Beer FC, Rader DJ. Overexpression of secretory phospholipase A(2) causes rapid catabolism and altered tissue uptake of high density lipoprotein cholesteryl ester and apolipoprotein A-I. *J Biol Chem* 2000; 275:10077-10084.
52. Kisilevsky R, Subrahmanyam L. Serum amyloid A changes high density lipoprotein's cellular affinity. A clue to serum amyloid A's principal function. *Lab Invest* 1992; 66:778-785.

53. Bhogal HK, Sanyal AJ. The molecular pathogenesis of cholestasis in sepsis. *Front Biosci* 2013; 5:87-96.
54. Recknagel P, Gonnert FA, Westermann M, Lambeck S, Lupp A, Rudiger A, Dyson A, Carré JE, Kortgen A, Krafft C, Popp J, Sponholz C, Fuhrmann V, Hilger I, Claus RA, Riedemann NC, Wetzker R, Singer M, Trauner M, Bauer M. Liver dysfunction and phosphatidylinositol-3-kinase signalling in early sepsis: experimental studies in rodent models of peritonitis. *PLoS Med* 2012; 9(11) :e1001338.
55. Solomkin JS, Robinson CT, Cave CM, Ehmer B, Lentsch AB. Alterations in membrane cholesterol cause mobilization of lipid rafts from specific granules and prime human neutrophils for enhanced adherence-dependent oxidant production. *Shock* 2007; 28:334-338.
56. White MM, Geraghty P, Hayes E, Cox S, Leitch W, Alfawaz B, Lavelle GM, McElvaney OJ, Flannery R, Keenan J, Meleady P, Henry M, Clynes M, Gunaratnam C, McElvaney NG, Reeves EP. Neutrophil Membrane Cholesterol Content is a Key Factor in Cystic Fibrosis Lung Disease. *EBioMedicine* 2017; 23:173-184.
57. Dünser MW, Ruokonen E, Pettilä V, Ulmer H, Torgersen C, Schmittinger CA, Jakob S, Takala J. Association of arterial blood pressure and vasopressor load with septic shock mortality: a post hoc analysis of a multicenter trial. *Crit Care* 2009; 13:R181.
58. Catapano AL, Pirillo A, Bonacina F, Norata GD. HDL in innate and adaptive immunity. *Cardiovasc Res* 2014; 103:372-383.

59. Fessler MB, Parks JS. Intracellular lipid flux and membrane microdomains as organizing principles in inflammatory cell signaling. *J Immunol* 2011; 187:1529-1535.
60. Hillyard DZ, Nutt CD, Thomson J, McDonald KJ, Wan RK, Cameron AJ, Mark PB, Jardine AG. Statins inhibit NK cell cytotoxicity by membrane raft depletion rather than inhibition of isoprenylation. *Atherosclerosis* 2007; 191:319-325.
61. Marik PE, Pastores SM, Annane D, Meduri GU, Sprung CL, Arlt W, Keh D, Briegel J, Beishuizen A, Dimopoulou I, Tsagarakis S, Singer M, Chrousos GP, Zaloga G, Bokhari F, Vogeser M; American College of Critical Care Medicine. Recommendations for the diagnosis and management of corticosteroid insufficiency in critically ill adult patients: consensus statements from an international task force by the American College of Critical Care Medicine. *Crit Care Med* 2008; 36:1937-1949.
62. Annane D. The Role of ACTH and Corticosteroids for Sepsis and Septic Shock: An Update. *Front Endocrinol* 2016;7:70.
63. Kanczkowski W, Tymoszek P, Chavakis T, Janitzky V, Weirich T, Zacharowski K, Ehrhart-Bornstein M, Bornstein SR. Upregulation of TLR2 and TLR4 in the human adrenocortical cells differentially modulates adrenal steroidogenesis. *Mol Cell Endocrinol* 2011; 336:41-46.
64. Haeno S, Maeda N, Yamaguchi K, Sato M, Uto A, Yokota H. Adrenal steroidogenesis disruption caused by HDL/cholesterol suppression in diethylstilbestrol-treated adult male rat. *Endocrine* 2016; 52:148-156.

65. Festti J, Grion CM, Festti L, Mazzuco TL, Lima-Valassi HP, Brito VN, Barbosa DS, Carrilho AJ. Adrenocorticotrophic hormone but not high-density lipoprotein cholesterol or salivary cortisol was a predictor of adrenal insufficiency in patients with septic shock. *Shock* 2014; 42:16-21.
66. Molenaar N, Bijkerk RM, Beishuizen A, Hempen CM, de Jong MF, Vermes I, van der Sluijs Veer G, Girbes AR, Groeneveld AB. Steroidogenesis in the adrenal dysfunction of critical illness: impact of etomidate. *Crit Care* 2012; 16:R121.
67. Etogo-Asse FE, Vincent RP, Hughes SA, Auzinger G, Le Roux CW, Wendon J, Bernal W. High density lipoprotein in patients with liver failure; relation to sepsis, adrenal function and outcome of illness. *Liver Int* 2012; 32:128-136.
68. Arem R, Ghusn H, Ellerhorst J, Comstock JP. Effect of decreased plasma low-density lipoprotein levels on adrenal and testicular function in man. *Clin Biochem* 1997; 30:419-424.
69. de Haan K, Groeneveld AB, de Geus HR, Egal M, Struijs A. Vitamin D deficiency as a risk factor for infection, sepsis and mortality in the critically ill: systematic review and meta-analysis. *Crit Care* 2014; 18:660.
70. Christaki E, Opal SM, Keith JC Jr, Kessinian N, Palardy JE, Parejo NA, Lavallie E, Racie L, Mounts W, Malamas MS, Mewshaw RE, Harris HA, Vlasuk GP. Estrogen receptor beta agonism increases survival in experimentally induced sepsis and ameliorates the genomic sepsis signature: a pharmacogenomic study. *J Infect Dis* 2010; 201:1250-1257.

71. Amrein K, Schnedl C, Holl A, Riedl R, Christopher KB, Pachler C, Urbanic Purkart T, Waltensdorfer A, Münch A, Warnkross H, Stojakovic T, Bisping E, Toller W, Smolle KH, Berghold A, Pieber TR, Dobnig H. Effect of high-dose vitamin D3 on hospital length of stay in critically ill patients with vitamin D deficiency: the VITdAL-ICU randomized clinical trial. *JAMA* 2014; 312:1520-1530.
72. National Heart, Lung, and Blood Institute PETAL Clinical Trials Network, Ginde AA, Brower RG, Caterino JM, Finck L, Banner-Goodspeed VM, Grissom CK, Hayden D, Hough CL, Hyzy RC, Khan A, Levitt JE, Park PK, Ringwood N, Rivers EP, Self WH, Shapiro NI, Thompson BT, Yealy DM, Talmor D. Early High-Dose Vitamin D3 for Critically Ill, Vitamin D-Deficient Patients. *N Engl J Med* 2019; 381:2529-2540.
73. Christeff N, Benassayag C, Carli-Vielle C, Carli A, Nunez EA. Elevated oestrogen and reduced testosterone levels in the serum of male septic shock patients. *J Steroid Biochem* 1988; 29:435-440.
74. Horvatits T, Drolz A, Rutter K, Roedl K, Langouche L, Van den Berghe G, Fauler G, Meyer B, Hülsmann M, Heinz G, Trauner M, Fuhrmann V. Circulating bile acids predict outcome in critically ill patients. *Ann Intensive Care* 2017; 7:48.
75. Kipp HA. Variation in the cholesterol content of the serum in pneumonia. *J Biol Chem* 1920; 44:215-237.
76. Press AT, Traeger A, Pietsch C, Mosig A, Wagner M, Clemens MG, Jbeily N, Koch N, Gottschaldt M, Bézière N, Ermolayev V, Ntziachristos V, Popp J, Kessels MM, Qualmann B, Schubert US, Bauer M. Cell type-specific delivery of short interfering RNAs by dye-functionalised theranostic nanoparticles. *Nat Commun* 2014; 5:5565.

77. Song Z, Bai J, Nauwynck H, Lin L, Liu X, Yu J, Jiang P. 25-Hydroxycholesterol provides antiviral protection against highly pathogenic porcine reproductive and respiratory syndrome virus in swine. *Vet Microbiol* 2019; 231:63-70.
78. McDonald MC, Dhady P, Cockerill GW, Cuzzocrea S, Mota-Filipe H, Hinds CJ, Miller NE, Thiemermann C. Reconstituted high-density lipoprotein attenuates organ injury and adhesion molecule expression in a rodent model of endotoxic shock. *Shock* 2003; 20:551-557.
79. Zhang Z, Datta G, Zhang Y, Miller AP, Mochon P, Chen YF, Chatham J, Anantharamaiah GM, White CR. Apolipoprotein A-I mimetic peptide treatment inhibits inflammatory responses and improves survival in septic rats. *Am J Physiol Heart Circ Physiol* 2009; 297: H866-873.
80. Datta G, Gupta H, Zhang Z, Mayakonda P, Anantharamaiah GM, White CR. HDL mimetic peptide administration improves left ventricular filling and cardiac output in lipopolysaccharide-treated rats. *J Clin Exp Cardiol* 2011; 2:1000172.
81. Kwon WY, Suh GJ, Kim KS, Kwak YH, Kim K. 4F, apolipoprotein AI mimetic peptide, attenuates acute lung injury and improves survival in endotoxemic rats. *J Trauma Acute Care Surg* 2012; 72:1576-1583.
82. Zhang X, Wang L, Chen B. Recombinant HDL (Milano) protects endotoxin-challenged rats from multiple organ injury and dysfunction. *Biol Chem* 2015; 396:53-60.
83. Tanaka S, Genève C, Zappella N, Yong-Sang J, Planesse C, Louedec L, Viranaïcken W, Bringart M, Montravers P, Denamur E, Duranteau J, Couret D, Meilhac O. Reconstituted

high-density lipoprotein therapy improves survival in mouse models of sepsis.

Anesthesiology 2020; 132:825-838.

84. Berger JM, Loza Valdes A, Gromada J, Anderson N, Horton JD. Inhibition of PCSK9 does not improve lipopolysaccharide-induced mortality in mice. *J Lipid Res* 2017;58:1661-1669.
85. Pajkrt D, Doran JE, Koster F, Lerch PG, Arnet B, van der Poll T, ten Cate JW, van Deventer SJ. Antiinflammatory effects of reconstituted high-density lipoprotein during human endotoxemia. *J Exp Med* 1996; 184:1601-1608.
86. Dellinger RP, Tomayko JF, Angus DC, Opal S, Cupo MA, McDermott S, Ducher A, Calandra T, Cohen J; Lipid Infusion and Patient Outcomes in Sepsis (LIPOS) Investigators. Efficacy and safety of a phospholipid emulsion (GR270773) in Gram-negative severe sepsis: results of a phase II multicenter, randomized, placebo-controlled, dose-finding clinical trial. *Crit Care Med* 2009; 37:2929-2938.
87. Guirgis FW, Black LP, Rosenthal MD, Henson M, Ferreira J, Leeuwenburgh C, Kalynych C, Moldawer LL, Miller T, Jones L, Crandall M, Reddy ST, Wu SS, Moore FA. LIPid Intensive Drug therapy for Sepsis Pilot (LIPIDS-P): Phase I/II clinical trial protocol of lipid emulsion therapy for stabilising cholesterol levels in sepsis and septic shock. *BMJ Open* 2019; 9:e029348.
88. Oesterle A, Laufs U, Liao JK. Pleiotropic effects of statins on the cardiovascular system. *Circ Res* 2017;120:229-243.
89. Zeiser R. Immune modulatory effects of statins. *Immunology* 2018;154:69-75.

90. Durant R, Klouche K, Delbosc S, Morena M, Amigues L, Beraud JJ, Canaud B, Cristol JP. Superoxide anion overproduction in sepsis: effects of vitamin e and simvastatin. *Shock* 2004;22:34-39.
91. Camerino GM, Tarantino N, Canfora I, De Bellis M, Musumeci O, Pierno S. Statin-Induced Myopathy: Translational Studies from Preclinical to Clinical Evidence. *Int J Mol Sci* 2021;22:2070.
92. DuBroff R, de Lorgeril M. Cholesterol confusion and statin controversy. *World J Cardiol* 2015;7:404-409.
93. Hackam DG, Mamdani M, Li P, Redelmeier DA. Statins and sepsis in patients with cardiovascular disease: a population-based cohort analysis. *Lancet* 2006;367:413-418.
94. Kruger P, Fitzsimmons K, Cook D, Jones M, Nimmo G. Statin therapy is associated with fewer deaths in patients with bacteraemia. *Intensive Care Med* 2006;32:75-79.
95. Liappis AP, Kan VL, Rochester CG, Simon GL. The effect of statins on mortality in patients with bacteremia. *Clin Infect Dis* 2001;33:1352-1357.
96. Kruger P, Bailey M, Bellomo R, Cooper DJ, Harward M, Higgins A, Howe B, Jones D, Joyce C, Kostner K, McNeil J, Nichol A, Roberts MS, Syres G, Venkatesh B; ANZ-STATInS Investigators—ANZICS Clinical Trials Group. A multicenter randomized trial of atorvastatin therapy in intensive care patients with severe sepsis. *Am J Respir Crit Care Med* 2013;187:743-750.
97. National Heart, Lung, and Blood Institute ARDS Clinical Trials Network, Truwit JD, Bernard GR, Steingrub J, Matthay MA, Liu KD, Albertson TE, Brower RG, Shanholtz C,

Rock P, Douglas IS, deBoisblanc BP, Hough CL, Hite RD, Thompson BT. Rosuvastatin for sepsis-associated acute respiratory distress syndrome. *N Engl J Med* 2014;370:2191-2200.

98. McAuley DF, Laffey JG, O'Kane CM, Perkins GD, Mullan B, Trinder TJ, Johnston P, Hopkins PA, Johnston AJ, McDowell C, McNally C; HARP-2 Investigators; Irish Critical Care Trials Group. Simvastatin in the acute respiratory distress syndrome. *N Engl J Med* 2014;371:1695-1703.

99. Calfee CS, Delucchi KL, Sinha P, Matthay MA, Hackett J, Shankar-Hari M, McDowell C, Laffey JG, O'Kane CM, McAuley DF; Irish Critical Care Trials Group. Acute respiratory distress syndrome subphenotypes and differential response to simvastatin: secondary analysis of a randomised controlled trial. *Lancet Respir Med* 2018;6:691-698.

Figure legends

Fig. 1 Cholesterol structure and location within cell membranes

Fig. 2 Cholesterol synthesis and metabolism pathways and impact of sepsis

ABC1; ATP-binding cassette transporter-1, AMP; adenosine monophosphate, AMPK; AMP-activated protein kinase, ApoA1; apolipoprotein A1, HDL; high-density lipoprotein, HMGCoA reductase; Hydroxymethylglutaryl-coenzyme A reductase, INSIG; insulin-induced gene-1 protein, LDL; low-density lipoprotein, SIRT-1; sirtuin-1, SREBP; sterol regulatory element-binding protein, SCAP; SREBP cleavage-activating protein, VLDL; very low-density lipoprotein.

* plasma levels may be normal or raised for adrenocorticoid hormones and bile acids but this may relate to decreased metabolism/excretion rather than increased production.

Cortisol levels frequently fail to augment with exogenous ACTH stimulation

Fig. 3 Functional roles of cholesterol

Fig. 4 Impact of sepsis on cholesterol transport

VLDL; very low-density lipoprotein, LDL; low-density lipoprotein, HDL; high-density lipoprotein; LDL-R; low-density lipoprotein receptor; ABC, ATP-binding cassette transporter; SR-BI, scavenger receptor B type 1; LCAT; lecithin-cholesterol acyltransferase; CETP; cholesteryl ester transfer protein; PCSK9; proprotein convertase subtilisin kexin 9.

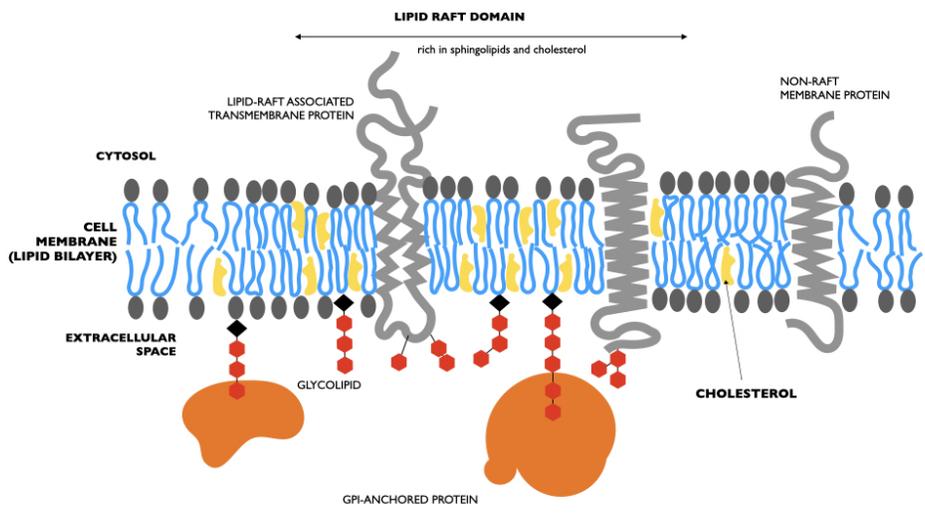


Fig. 1: Cholesterol structure and location within cell membranes

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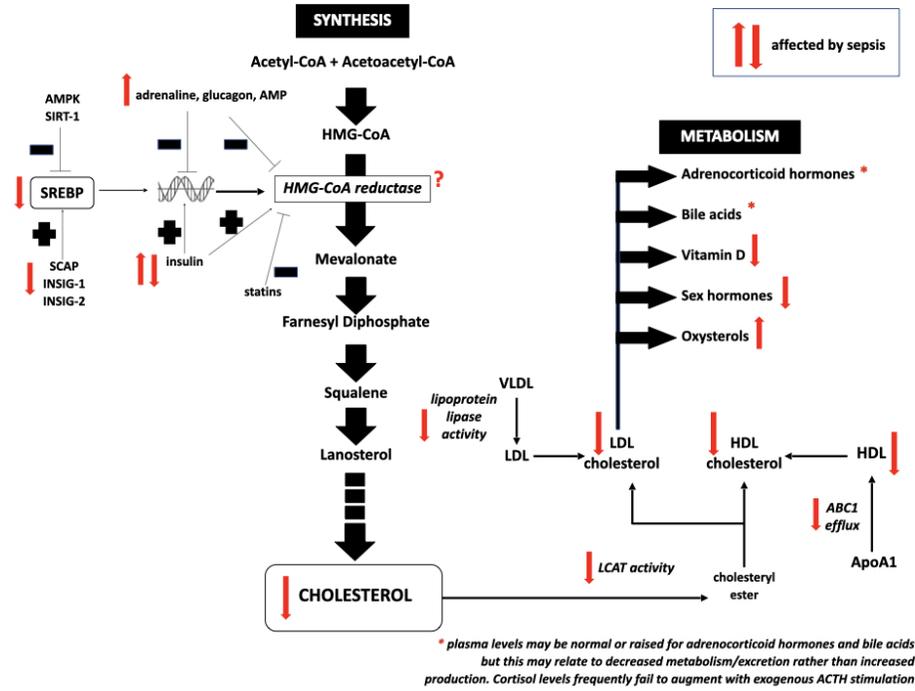


Fig 2: Cholesterol synthesis and metabolism pathways and impact of sepsis

361x270mm (72 x 72 DPI)

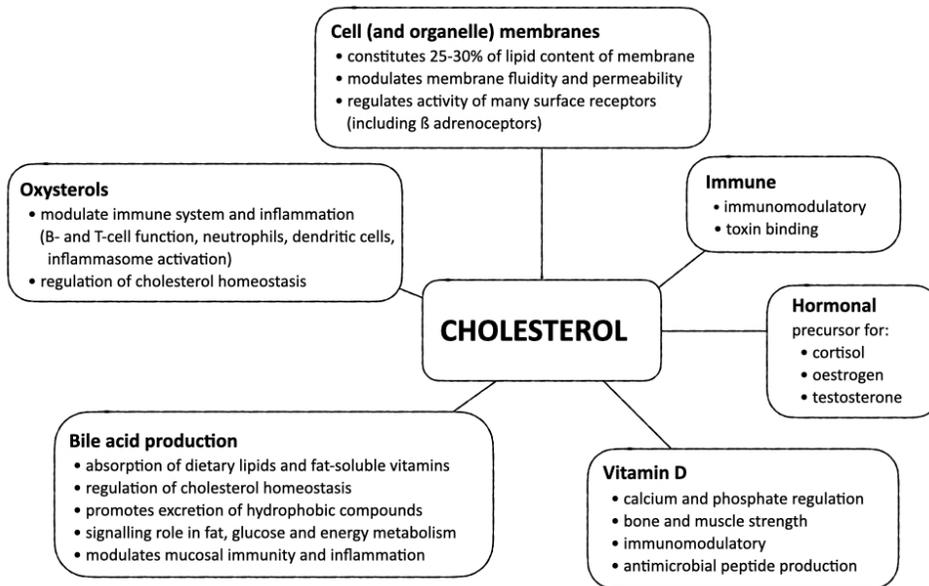


Fig 3: Functional roles of cholesterol

361x270mm (72 x 72 DPI)

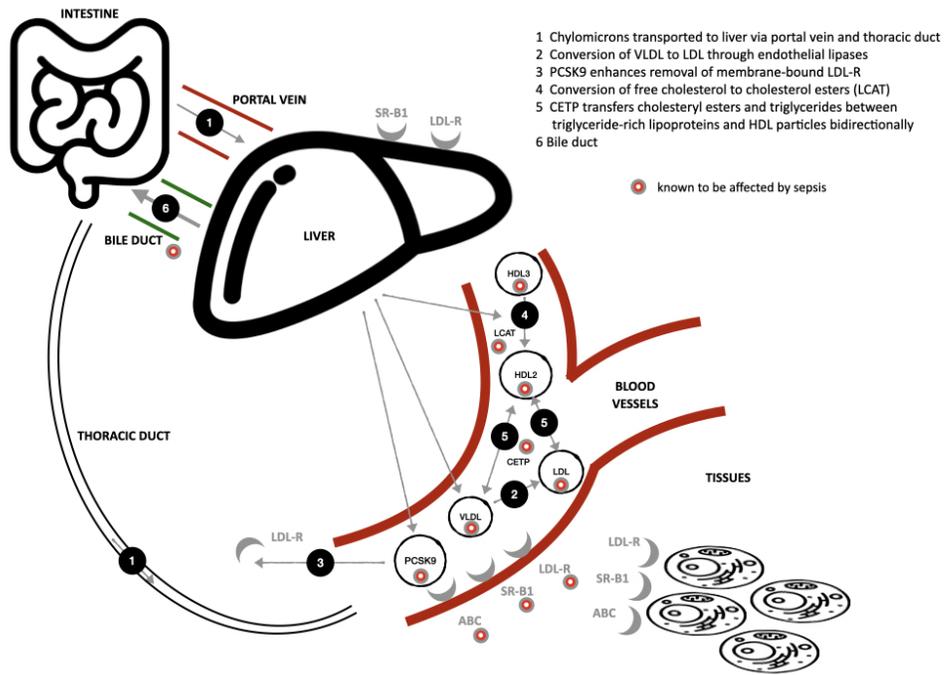


Fig 4: Impact of sepsis on cholesterol transport

361x270mm (72 x 72 DPI)