A *CADM3* VARIANT CAUSES CMT2 WITH MARKED UPPER LIMB INVOLVEMENT

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ABSTRACT

The CADM family of proteins consists of four neuronal specific adhesion molecules (CADM1, CADM2, CADM3 and CADM4) that mediate the direct contact and interaction between axons and glia. In the peripheral nerve, axon-Schwann cell interaction is essential for the structural organization of myelinated fibers and it is primarily mediated by the binding of CADM3, expressed in axons, to CADM4, expressed by myelinating Schwann cells. We have identified by whole exome sequencing three unrelated families, including one *de novo* patient, with axonal Charcot Marie Tooth disease (CMT2) sharing the same private variant in *CADM3*, Tyr172Cys. This variant is absent in 230,000 control chromosomes from gnomAD and predicted to be pathogenic. Most *CADM3* patients share a similar phenotype consisting of autosomal dominant CMT2 with marked upper limb involvement. High resolution mass spectrometry analysis detected a newly created disulfide bond in the mutant CADM3 potentially modifying the native protein conformation. Our data support a retention of the mutant protein in the endoplasmic reticulum and reduced cell surface expression *in vitro*. Stochastic Optical Reconstruction Microscopy (STORM) imaging revealed decreased co-localization of the mutant with CADM4 at intercellular contact sites. Mice carrying the corresponding human mutation (*Cadm3*^{Y170C}) showed reduced expression of the mutant protein in axons. *Cadm3*^{Y170C} mice showed normal nerve conduction and myelin morphology, but exhibit abnormal axonal organization, including abnormal distribution of Kv.1 channels and Caspr along myelinated axons. Our findings indicate the involvement of abnormal axon-glia interaction, which has been reported in CMT-like phenotype in mice, as a disease-causing mechanism in CMT patients with *CADM3* mutations.

KEYWORDS AND ABBREVIATIONS

Charcot-Marie-Tooth Disease (CMT), CADM3, cell-adhesion molecule

INTRODUCTION

In the peripheral nervous system, axon-glia interactions are essential for myelination and organization of the different axonal domains required for efficient saltatory conduction (Salzer *et al.*, 2008). Distinct sets of cell-cell adhesion molecules are known to mediate the interaction between axons and Schwann cells in different nerve domains. Those molecules are well characterized in the paranodal and juxtaparanodal regions. In the paranodal region the axonal proteins Caspr and contactin interact with neurofascin 155 (NF155) in glia, and in the juxtaparanodes Caspr2 binds to Tag1 (Salzer *et al.*, 2008). In the internodes, the axon-glia interaction is mediated by more recently characterized cell adhesion molecules, the CADM family of proteins, also known as nectin-like (Necl) and synaptic cell adhesion (SynCAM) molecules (Maurel *et al.*, 2007). CADM proteins consist of four members that are differentially expressed in Schwann cells and axons; CADM1 (Necl2), CADM2 (Necl3), and CADM3 (Necl1) are found in axons, while CADM2 and CADM4 (Necl4) are present in myelinating Schwann cells (Golan *et al.*, 2013). CADM proteins are composed of three extracellular Immunoglobulin-like (Ig) domains where cell-cell adhesion takes place, a transmembrane domain and a short cytoplasmic region (Kakunaga *et al.*, 2005). Cell-cell contact is made through different combinations of homophilic and heterophilic interactions. However, the predominant interaction is mediated between CADM3 expressed in axons and CADM4 located on Schwann cells (Maurel *et al.*, 2007) (Spiegel *et al.*, 2007). Those interactions are essential for long-range axon-guidance, myelination and maintenance of the axonal architecture domain. Besides cell-cell adhesion, CADM proteins also play a role as cell signaling molecules. They can inhibit neuregulin-mediated activation of the PI3K/AKT pathway (Chen *et al.*, 2016). Interestingly, ablation of CADM4 in mice results in myelin abnormalities and phenotype that resembles CMT (Golan *et al.*, 2013). Mice lacking *CADM4* exhibit impaired motor function, slower nerve conduction velocity and myelin abnormalities including neuropathy-like focal hypermyelination, and developed abnormal axon-glial contact and

axonal organization (Golan *et al.*, 2013). Severe myelin abnormalities, including very short internodes were also observed in transgenic mice expressing a dominant-negative mutant of CADM4 lacking its cytoplasmic domain (Cadm4dCT) (Gollan *et al.*, 2003; Elazar *et al.*, 2019). Furthermore, CADM proteins collaborate with other axon-glia CAMs to regulates oligodendrocyte myelination in the CNS (Elazar *et al.*, 2019). In contrast, mice lacking the other *Cadm* family of genes, *Cadm1*, *Cadm2* and *Cadm3*, showed no myelin or any other neurological abnormalities (Golan *et al.*, 2013), suggesting genetic redundancy in mouse. However, according to gnomAD, the four CADM genes in human have high constraint for loss-of function (LoF) mutations, displaying extremely intolerant LoF metrics (pLI>=0.9 and o/e<0.2), suggesting that humans are less tolerant to loss of CADM genes.

Here we report a pathogenic variant in *CADM3* in three unrelated families sharing the same unusual Charcot-Marie-Tooth disease with predominant upper limb involvement and atypical pyramidal features. Our genetic findings and functional studies unveil a novel pathological mechanism in CMT involving axon-glial interactions.

MATERIALS AND METHODS

CMT patients and families

All study participants were ascertained by board-certified neurologists Drs. Michael Shy and Mary Reilly. All subjects' consent was obtained according to the Declaration of Helsinki and that it has been approved by the ethical committee of the institution in which the work was performed.

Whole-exome and Sanger sequencing

Whole-exome sequencing was performed in the probands for the three CMT families. The SureSelect Human All Exon Kit (Agilent) was used for in-solution enrichment, and the HiSeq 2500 instrument (Illumina) was used to produce 100 bp paired-end sequence reads. The Burrows-Wheeler aligner and Freebayes were used for sequence alignment and variant calling. Whole-exome data were uploaded into the GENESIS 2.0 software and analyzed using strict filters for allele frequency, function and conservation. Mutations in known CMT-associated genes were absent in all families (Table S1). Sanger sequencing confirmed the missense variant in CADM3 detected by whole-exome sequencing, and it also revealed *de novo* inheritance mode of segregation in one family and autosomal dominant segregation in another family.

Paternity test

Parental consents (paternal and maternal) were obtained according to the Declaration of Helsinki and were approved by the ethical committee of the institution in which the proband was evaluated. Fragment analysis was performed to verify paternity and maternity of family 1 in order to confirm *de novo* mutation in the proband. Five different microsatellite markers were amplified by multiplex PCR using the Qiagen multiplex PCR kit: D6S1552, D6S1624, D61517, MOG-CA. PCR products were run on an ABI3730 automated sequencer (Applied Biosystems) using LIZ600 (orange) size standard. Results were analyzed with GeneMapper.

Generation of knock-in mice

Knockin mice containing the human CADM3 mutation Y172C were generated using the CRISPR/Cas9 system. SgRNA (crRNA and tracrRNA) and repair DNA oligonucleotides were ordered from IDT. All reagents were prepared according to the manufacturer's instructions and were microinjected or electroporated with a cas9 purified protein, into fertilized oocytes of C57BL/6J mice. SgRNA sequence CCACAGAAACCCATAATCAC, repair DNA sequence TGAACTGTGACTTCATGGTGCCTCTCCTTCCTATCCTGGCCATTCCCTATGCATGGCA GGGATCCCTCAGAAGCCTATTATTACTGGTTGTAAGTCATCATTGCGGGAAAAGGAG ACAGCCACTCTAAATTGTCAGTCTTCTGGGAGCAA. The repair sequence included silent mutations in the area corresponding to the guide sequence including a restriction enzyme site. Mosaic mice were screened by primers that were specific for the edited allele (forward: CTCCACCCCACCTACACTAC; reverse: GTAATAATAGGCTTCTGAGG), and with primers flanking the edited area of which the products were further used for RFLP (forward: CTCCACCCCACCTACACTAC; reverse: TGTCTGCCATTCATTTCCACT, product was cut with BamHI). Positive mosaics confirmed by Sanger sequencing were crossed on WT C57BL/6J mice. All positive F1 mice were sequenced as well (as mosaic mice can contain more than one form of edited alleles). A single sequence-verified male and female F1 mice were crossed to establish a working colony, in parallel a single sequence-verified mail was crossed on WT females to establish a family crossed continuously on a WT background. All mice were genotyped at 3 weeks after birth. All experiments in this study were approved by the Institutional Animal Care and Use Committees (IACUC) at the Weizmann Institute.

For RT PCR analysis, total RNA was isolated from freshly dissected brains using TRI-reagent (Sigma-Aldrich), cDNAs were obtained with SuperScript II reverse transcriptase (Invitrogen) using oligo-dT. Specific PCR primer sets were designed according to mRNA sequences (NCBI). The following primer pairs were used for mRNA-based RT-PCR analysis: Cadm3 WT allele CCTATTTAATCCCGGCGACTGC and

TAACCAGTGATTATGGGTTTCTG, mutant allele CCTATTTAATCCCGGCGACTGC and GTAATAATAGGCTTCTGAGGG; actin GTGGTGGTGAAGCTGTAGCCACGCT and GAGCACCCTGTGCTGCTCACCGAGG.

Constructs generation

Plasmids encoding the open-reading frame (ORF) of CADM3 transcript1 (NM_021189.4) fused with either an HA tag or GFP tagged at the C-terminus end were obtained from Genecopoeia. Plasmid encoding the open-reading frame of CADM4 transcript 1 (NM_145296.1) fused with a myc-tag at the C-terminus was also obtained from Genecopoeia. Site-direct-mutagenesis was performed to generate plasmids with the patients' mutation corresponding to Tyr172Cys. Point mutation was introduced by PCR using the Q5 Site-Direct Mutagenesis kit (NEB) with the following primers: 5'-ATCACTGGTTGTAAATCTTCATTACG-3' and 5'-GATGGGCTTCTGTGGAATTC-3'. Addition of N-terminus HA tag and the A519G mutation to rat Cadm3 Spiegel et al., 2007) (corresponding to rat and human Tyr170Cys and Tyr172Cys, respectively), were generated using the Site Directed Mutagenesis (SDM) TPCR[1]. Primers (5-GCGCCCGGCGGGGCCAATTACCCATACGATGTTCCAGATTACGCTCTTTCCCAGGAC GGCTACTG-3; 5-TTCCCGCAACGATGACTTACAACCAGTGATT ATGGGTTTC-3) and (5-GCAGGCGAGGAGCAGGAGC-3; 5-GG GAGCAAACCTGCAGCC-3) were used. The addition of C-terminus Myc tag was done by PCR using (5-ACGACTCACTATAGG GAGACC-3) and (5-CTAGCGGATCCT ACAGATCCTCTTCTGAGATGAGTTTTTGTTCGATGAAATATTCCTTCTTGTCATC-3) primers. The ORF of all constructs was confirmed by DNA sequencing.

Endoplasmic reticulum (ER) stress analysis

RT4 Schwann cells stably expressing a dual luciferase reporter assay of XbpI splicing (Bai et al., 2018) were used to monitor activation of the unfolded protein (UPR) response. CADM3 constructs

were transfected with lipofectamine 3000 (Thermofisher) into RT4 Schwann cells. Firefly luciferase and nano-luciferase activity was assayed as described by Bai Y et al(Bai et al., 2018) ER localization was analyzed by confocal microscopy in cells co-stained with an HA antibody and the ER marker calnexin.

Co-immunoprecipitation assay and Western Blot

HEK293T cells, 48-72 hours after transfection, were lysed in solubilization buffer (50mM HEPES pH 7, 150mM NaCl, 5mM EGTA, 10% glycerol, 1% Triton X100 and 1% Proteinase Inhibitors mix (Calbiochem 539134)). Lysates were centrifuged for 20 min at 4°C and the supernatants were kept at -20°C. Sample buffer X5 (0.5M Tris HCl pH 6.8, 50% glycerol, 5% SDS and 3.7% DTT) was added and the samples were boiled for 5 min and were loaded for separation on a 10%-12% polyacrylamide gel. The gel was transferred to a Polyvinylidene difluoride (PVDF) membrane for blotting. The blots were blocked in TBS-T containing 3% BSA for 1 hour in room temperature and incubated for 1 hour at room temperature and then for 1 hour or for overnight at 4°C with different primary antibodies diluted in the blocking buffer. Following 3 washes in TBS-T, blots were incubated with horseradish-peroxidase-coupled secondary antibodies for 30 min at room temperature and were washed again with TBS-T.

Three months old mice SNs were dissected and immediately frozen in dry ice. Homogenization was performed in RIPA buffer (25 mM Tris-HCl pH 7.4, 1% NP40, 0.5% sodium deoxycholate, 0.1% SDS,150 mM NaCl) supplemented with 1% protease inhibitors cocktail (Calbiochem 539134). Homogenates were incubated on ice for 40 min, centrifuged at 15,000 rpm for 30 min and supernatant was collected and kept at −80°C.

For the preparation of mice brain membranes, three months old mice brains were homogenized in membrane homogenization buffer (20mM HEPES pH 7.5, 0.32M sucrose, 1mM EGTA, 1.5mM MgSO4) supplemented with 1% protease inhibitors cocktail (Calbiochem 539134). The homogenates were centrifuged at 1000g for 10 min at 4°C for removal of nuclei and heavy cell

debris. Supernatants were collected and centrifuged at 20,000g for 1 hour at 4°C. The pellets were resuspended in membrane extraction solubilization buffer (2% NP40, 2mM MgCl2, 2mM PMSF) supplemented with 1% protease inhibitors cocktail (Calbiochem 539134) and kept at -80°C. Protein concentration was determined using a BCA kit (Pierce). Sample buffer X5 (0.5M Tris-HCl pH 6.8, 50% glycerol, 5% SDS, 3.7% DTT and a bit of bromophenol blue) was added and the samples were boiled for 5 min. Equal amounts of protein were loaded for separation on a 10% SDS-PAGE gel and transferred to PVDF membrane (semi-dry transfer using Trans-Blot Turbo system (25V, 10 min), Bio-Rad). Membranes were then incubated with blocking solution (3% BSA in TBS-T) for 30 min at room temperature and then reacted for 1 hour at room temperature or for overnight at 4°C with the appropriate primary antibodies diluted in TBS-T. Membranes were then washed in TBS-T (3 × 5 min) and incubated with horseradish-peroxidase-coupled secondary antibody for 45 min at room temperature. Membranes were washed again and reacted with ECL (Thermo Scientific). Membranes were visualized using the ChemiDoc MP imaging system (Bio-Rad).

Membrane proteins extraction

Transfected HEK293T cells were lysed with homogenization buffer (20mM HEPES pH 7.5, 0.32M sucrose, 1mM EGTA, 1.5mM MgSO4, 1% proteinase inhibitors mix (Calbiochem 539134) using homogenizer. The lysates were centrifuged at 3000g for 10 min at 4°C for removing nuclei and cell debris, supernatants were collected and centrifuged again for 60 min in 20000g. The pellets were resuspended in solubilization buffer (2% IGEPAL, 2mM MgCl2, 1% proteinase inhibitors mix), incubated for 30 min with alternately vortex and again were centrifuged at 20000g for 30 min. samples were kept in -20°C and later analyzed by immunoblot, as described above, using rabbit anti HA tag antibody.

Cell surface biotinylation

Surface biotinylation assay was done on transfected HEK293T cells at 48-72 hours after transfection. Cells were washed three times with ice-cold PBS+Ca+Mg pH 8 and biotinylated with 1 mg/ml Sulfo-NHS-biotin (Thermo Scientific, 21217) as previously described (Gollan et al., 2003). Proteins were immunoprecipitated by anti Myc tag antibody and anti-mouse IgG agarose beads (Sigma, A6531) rotated overnight at 4°C. The beads were washed once with ice-cold solubilization buffer and twice with ice-cold TNTG and eluted with the SDS-PAGE sample buffer and 5 min boiling. Samples were then then subjected to Western blot analysis as described above. Membranes were blotted with horseradish-peroxidase-coupled streptavidin for biotinylated protein detection.

Binding assay

For Fc-fusion protein binding experiments, transfected COS7 cells were washed with fresh DMEM and incubated for 45 min at room temperature with conditioned media containing 0.5–1 μg/ml of the different Fc-fusion Cadm proteins, preincubated with Cy3 anti human Fc antibody. The cells were washed twice with PBS to remove unbound proteins and were analyzed by immunofluorescence as previously described.

Electron microscopy

WT and homozygous mice were sacrificed, and their sciatic nerves exposed and fixed by continuous dripping of fresh Karnovsky fixative for 20 min (fixative containing 4% paraformaldehyde, 0.1 M sodium cacodylate, and 2.5% glutaraldehyde, pH 7.4, in PBS). Sciatic nerves were then carefully dissected, placed in fresh fixative, and left to rotate over-night at room temperature. Samples were then transferred to 4°C until processed by an EM professional as previously described.(Feinberg et al., 2010) Semithin sections (1.5 μm) were stained with toluidine blue and analyzed under a light microscope (Nikon Eclipse E800).

Electrophysiology

Sciatic nerve conduction velocity measurements were performed by Prof. Mike Shy (University of Iowa, IA USA) on adult WT (n=5) and homozygous (CADM3Y172C/Y172C) mice (n=3). Mice were anesthetized with a mix of Ketamine/Xylazine (87.5mg/kg Ketamine,12.5mg/kg Xylazine). Anesthetized mice were placed on a heating pad (constant 37°C) to maintain constant body temperature throughout the electrophysiological study (Neuromax, XLTEK). Sciatic nerve motor nerve conduction velocity was recorded stimulating a distal (ankle) and a proximal (sciatic notch) site along the nerve with platinum monopolar needle electrodes. One pair of stimulating electrodes was positioned percutaneously at the sciatic notch. A second pair was inserted adjacent to the tibial nerve at the ankle. Compound muscle action potentials (CMAP) were recorded from the intrinsic foot muscle using needle electrodes. CMAP amplitudes were measured from baseline to the peak of negative deflection.

Disulfide-bond analysis by mass spectrometry

HEK293T cells were transfected with either Y172C-CADM3-HA or Y132C-CADM3-HA. Protein lysates were extracted and immunoprecipitated with magnetic beads containing HA antibodies. IP eluate was submitted for analysis to MtoZ Biolabs. Briefly, protein samples were digested with trypsin and analyzed on Liquid Chromatography with tandem mass spectrometry (LC-MS-MS). Raw data was analyzed by BiopharmFinder.

Transfection and Immunocytochemistry

L Cells stabled transfected with the CADM3 constructs either wild-type, WT-CADM3-GFP, or mutant, Y172C-CADM3-GFP, were plated in glass-bottom 4-well chambers (Ibidi). Cells were co-transfected with CADM4-Myc using lipofectamine 3000 (ThermoFisher). 24 hours after transfection cells were fixed with 4% paraformaldehyde and permeabilized with cold methanol. Cells were incubated with primary antibodies, diluted in 2% BSA for 2hrs. Cells were subsequently

washed three times with PBS and incubated with the secondary antibodies. COS7 and HEK293T cells were grown in DMEM medium (Gibco) which contained 10% FBS (Gibco) and 1% Penicillin/Streptomycin (Gibco). HEK293T transfection was preformed using Calcium Phosphate method and COS7 transfection was done using Lipofectamine 2000 reagent (Invitrogen). Transfected COS7 cells seeded on 13mm glass cover slips (Thermo Scientific) were fixed in 4% paraformaldehyde (PFA) for 10 min and were blocked for 1 hour in blocking solution (5% normal goat serum without 0.1% Triton X-100 in PBS) at room temperature. Fixed cells were then washed in PBS and incubated for 1 hour at room temperature or for overnight at 4°C with the relevant mixture of primary antibodies diluted in blocking solution. Cover slips were then washed in PBS and incubated with secondary antibodies diluted in blocking solution for 45 min at room temperature. Cover slips were washed again in PBS and then mounted with fluoromount-G (SouthernBiotech).

Nerve immunofluorescence

Sciatic nerves were dissected and immersed in 4% PFA for 20 min at room temperature. Nerves were transferred to 20% sucrose (in PBS) and stored at 4°C till teasing. Nerves were desheathed and teased using fine forceps on SuperFrost Plus slides (Menzel-Glaser; Thermo Scientific), air dried overnight, and then kept frozen at -20°C till use. When staining for Kv1.2 or NrCAM, samples were postfixed for 2–5 min using cold methanol (-20°C) and washed with PBS. Samples were incubated for 1 hour in blocking solution (5% normal goat serum and 0.1% Triton X-100 in PBS) at room temperature. If no postfixation was performed, blocking solution contained 0.5% Triton X-100. Samples were incubated overnight at 4°C with the relevant mixture of primary antibodies diluted in blocking solution (5% normal goat serum and 0.1% Triton X-100, in PBS). Nerve samples were then washed in PBS and incubated with secondary antibodies for 45 min at room temperature (same blocking solution as for the primary antibodies). Samples were washed again in PBS, mounted as above, and observed using an Axioskop2 microscope equipped with an ApoTom

imaging system (Carl Zeiss) fitted with a Hamamatsu ORCA-ER CCD camera. Images were acquired and processed using the Zen 2012 software (Carl Zeiss), Photoshop CC2019 software (Adobe) and Power point 2019 software (Microsoft).

Antibodies

Primary antibodies that were used: monoclonal rabbit anti HA (Cell Signaling, C29F4), mouse anti Myc (9E10), anti-GFP (Cell signaling), rabbit anti extracellular domain Cadm3 (190), rabbit anti extracellular domain of Cadm3 (Spiegel et al., 2007; polyclonal antibodies 876 and 878), mouse anti GAPDH (Millipore, MAB374), rabbit anti Kv1.2 (Rhodes et al., 1995), rabbit anti Caspr (Peles et al., 1997; polyclonal antibodies 60-62), rat anti NrCAM (Horresh et al., 2010; RP12B3). Fluorophore-coupled secondary antibodies were purchased from Jackson Laboratories. Alexa Fluor 555 from ThermoFisher, and Janelia Fluor 646 from Novus Biologicals. For the binding experiments Fc fusion Cadm4 protein was used (Spiegel et al., 2007).

Stochastic Optical Reconstruction Microscopy (STORM) Imaging

Imaging experiments were carried out with a Nikon eclipse Ti2 microscope equipped with Nikon Instruments (NSTORM). For two color dSTORM imaging, Janelia 646, and Janelia 549 secondary antibodies were used with MEA STORM imaging buffer and were imaged continuously with 10000 frames collected per filter range at a frequency of 20 ms. The images were acquired using a 100x, 1.49 NA objective, and imaged onto a Hamamatsu C11440 ORCA-flash 4.0 camera. Storm analysis was carried out with Nikon Elements Analysis 5.02.01 for identification of molecules. Molecule list files were then exported from Nikon elements to be further analyzed using software Clus-Doc software in MATLAB R2018b. Cluster analysis, specifically DBSCAN function, was carried out after manually selecting region of interest.

Data Availability

The data presented in this study are available from the corresponding author upon request.

RESULTS

Identification of *CADM3* as a disease-causing gene

Whole-exome sequencing was performed in three unrelated index patients diagnosed with unusual neuropathy with marked upper limb involvement (Table 1). Pathogenic variants in known CMT genes were absent in the three families. Sequencing data was processed and analyzed by bioinformatics tools including BWA aligner, FreeBayers, GATK and GENESIS (Gonzalez *et al.*, 2015). The GENESIS platform currently harbors 9,569 whole exomes/genomes from individuals with different phenotypes, including 1,257 CMT affected individuals. *CADM3* was identified by utilizing a strategy of searching for very rare ('private') alleles shared across these CMT families. The required filters were non-synonymous variants, never observed in gnomAD ($MAF_{gnomad} = 0$, highly conserved (GERP>3)(Cooper *et al.*, 2005) and predicted to cause functional damage (CADD>25) (Rentzsch *et al.*, 2019). This approach yielded only one heterozygous variant: c.515A>G (NM_021189.4), p.Tyr172Cys (chr1:159163243, hg19), which was confirmed by Sanger sequencing (Figure 1A). This *CADM3* variant co-segregated with disease in all three families. The sporadic patient in family 1 carried Tyr172Cys as a *de novo* variant; confirmed by paternity testing (Figure 1A and Supplemental Figure 2), which suggests an independent mutation event rather than a founder effect. Family 2 is sporadic, and DNA of parents are not available. Family 3 shows dominant segregation of mutation in both affected individuals (mother and son) (Figure 1A). This variant is highly conserved across species, predicted to be pathogenic by seven different prediction tools including DANN, DEOGEN2, EIGEN, FATHMM-MKL, M-CAP, MutationAssessor and MutationTaster. Variant is predicted to be "likely pathogenic" (greater than 90% certainty) based on ACMG (Richards *et al.*, 2015) standards (PS2, PM2 and PP3). Further, the Tyr172Cys allele is absent from our internal controls (8,312 non-CMT WES/WGS

chromosomes on the GENESIS platform). A Fisher's exact test statistic formally supports a significance value of $p<0.01$ (Table S2).

Clinical description of individuals with CADM3 variants

Family 1

Patient II:1 was a 30-year-old woman presenting with a pronounced neuropathy affecting upper and lower extremities. She was born 3 months prematurely weighing only 2 pounds nine ounces and required intubation and care in the intensive care unit. She had a patent ductus arteriosus that required surgery. She did not walk independently until 18 months of age, required bracing to walk between the ages of 2 and 3 years and ankle foot orthotics (AFOs) to ambulate between the age of 5 and 12. She had multiple surgeries in both ankles after the age of 12 throughout her teenage years including tendon transfers and eventually bone fusions on the right ankle. She also had tendon transfers on the right forearm. As a child she had pronounced difficulties with balance. She required adaptations on her bicycle at eight years of age because she was unable to use hand or foot brakes. At age 30 she utilizes AFOs ambulate and has virtually no use of her left hand for fine motor movements. On neurological examination she has mild scoliosis, pronounced atrophy in both arms below her elbows and in both legs below her knees. Her left wrist strength was weak to extension and flexion (4-/5) while her right wrist extension was 4- but 5/5 for flexion. Left grip strength was 3/5 and 4+/5 on the right. Finger extensors were 2/5 on the left and 4-/5 on the right. Her intrinsic hand muscles, abductor pollicis brevis, first dorsal interosseous and abductor digiti minimi were all 0/5 on the left. Her abductor pollicis brevis was 3/5 on the right and the first dorsal interosseous and abductor digiti minimi were 1/5. In the lower extremities her anterior tibialis was 4/5 on the left and 4+/5 on the right. Her ankle extenders were 3+/5 bilaterally and inversion was full strength. She had no movement of either great toe. Touch sensation was reduced at both great toes but normal in her ankles and fingers. Vibratory sensation was absent at her toes, but normal at her ankles and fingers. Position sense was reduced in her fingers and left great toe. Deep tendon reflexes were

absent at her ankles but present in her knees and upper extremities. There is no family history of these symptoms.

Family 2

Patient II:1 was a seven years old male who was first noted to have weakness at three years of age. Early developmental milestones were normal, including rolling by 5 months, sitting by 6 months, walking by 12 months, normal fine motor and language skills. He gradually developed complete foot drop and cannot walk without AFO. He denies sensory symptoms. On neurological examination he has facial weakness, nasal speech, no voluntary movement in the lower extremities below his knees and severe foot deformity. He has wrist and finger contractures with moderate weakness in his first dorsal interosseous muscle of the hand. while he had weakness in wrist extension and flexion his contractures prevented grading strength for these muscles Vibratory sense was decreased in fingers bilaterally, position sense and cutaneous sensation were normal in all four extremities. Deep tendon reflexes were absent. Nerve conduction studies were mildly reduced in the ulnar and peroneal nerves with pronounced reduction of the CMAP amplitudes and were slow in the median nerve with a markedly reduced CMAP amplitude (Tables 2 and 3). Sensory SNAP amplitudes were unobtainable. He also had nerve and muscle biopsy. A sural nerve biopsy showed a reduced number of large myelinated fibers and very few groups of small regenerating fibers (Figure 5). Of interest, we overserved a number of abnormally myelinated fibers with thick folded myelin and focal myelin swelling or hypermyelination. The muscle biopsy reported a neurogenic atrophy.

Family 3

Case II:1 had normal birth and milestones. Around the age of 3 he was noted to have flat feet and between 3 and 4 years became slow at running and began to turn in his feet. He was prescribed AFOs at the age of 5 and was no longer able to run since the age of 8. He had multiple operations

in his feet including bilateral arthrodesis at the age of 11. He also complained of impaired dexterity of his hands from the first decade of life. There was no mention of sensory symptoms in the lower limbs, but he complained of sensory symptoms in the ulnar distribution bilaterally, which improved after ulnar nerve decompression, although muscle weakness did not. Examination at the age of 14 years showed distal atrophy and severe weakness of intrinsic hand muscles, ankle dorsiflexion and eversion, but preserved strength of ankle plantar flexion and of proximal muscle groups (Table 1). Reflexes were brisk throughout. Tone was normal and plantars were down-going. Pinprick sensation was normal in the lower limbs but reduced in the ulnar nerve distribution bilaterally, being worse on the left where is also involved other nerve territories up to the distal arm. Vibration sensation was also reduced in the lower limbs up to the knee.

His mother, I:2 now aged 49, had no definite symptoms until the fourth decade of life, when she started complaining of cramping in her right hand and progressive sensory loss in the ulnar nerve territory. She underwent two right ulnar nerve decompression, which improved the sensory symptoms, although weakness in her right hand continued to progress. In the last decade she noticed similar symptoms in her left hand. When asked, she admits she always thought she had thin hands compared to the rest of her body and as a teenager she used to get occasional cramping in her legs. Her current examination showed distal atrophy and moderate weakness in her hands and reduced sensation distally in her upper limbs, worse on the right and in the ulnar nerve territory (Table 1). Lower limb muscle bulk, strength and sensation were normal, but she did had difficulty walking on her heels. Reflexes were pathologically brisk throughout. Muscle tone was normal and Babinski reflex was absent.

In conclusion, II-1 and I-1 had a phenotype consistent with Silver Syndrome, eg an upper limb predominant motor predominant sensory-motor neuropathy with brisk reflexes throughout. There was a prominent involvement of ulnar nerves, with partial improvement of sensory symptoms after ulnar nerve decompression. Nerve conduction studies were consistent in both cases with an axonal sensory-motor neuropathy with intermediate reduction of conduction velocities

(Tables 2 and 3).

Tyr172Cys mutation interferes with the native disulfide bond conformation

The mutation is located in the extracellular domain adjacent to the second Ig (Ig2) loop, essential for cell-cell adhesion (Figure 1B). The Ig-like loops are formed by three pairs of cysteines connected by disulfide bonds at positions 84-144, 186-243, 288-333 (Figure 1C). All six cysteines are conserved across the four different CADMs showing by multiple sequence alignment (Supplemental Figure 3). Interestingly, those are the only cysteines present in the entire CADM3 sequence, with the exception of the cysteines located at the membrane signal peptide that is supposed to be cleaved (Zhou *et al.*, 2005). Because the mutation creates a new cysteine, we hypothesized that this new cysteine forms a disulfide bond interfering with native disulfide bonds conformation. To test this hypothesis, HEK293T cells were transfected with a plasmid encoding the HA-tagged human or the Y172C mutant Cadm3 in order to pull-down the proteins using an HA antibody. The purified CADM3 proteins were then digested with trypsin and subjected to Liquid Chromatography with tandem mass spectrometry (LC-MS-MS). Analysis of the MS data identified a non-native disulfide bond between the mutant site 172C and 186C (Figure 1C and Supplemental Figure 1). In addition, the native disulfide bond between 186C and 243C was also detected, suggesting either disulfide bond shuffling or co-occurrence of different folded molecules.

CADM3 mutation activates the Unfolded Protein Response

In order to investigate whether the disulfide bond rearrangement could cause activation of the unfolded protein response (UPR) due to protein misfolding, we used a dual luciferase reporter assay of XBP1 splicing using stably transfected RT4 Schwann cells (Bai *et al.*, 2018). These cells encode XBP1, a key modulator of the UPR, fused to luciferase reporters (firefly and nano-luciferase) sequences, which are separated by an intronic sequence. XBP1 is fused out of frame with the

luciferases, and both are pulled into frame only when XBP-1 gets spliced. During the process of UPR, IRE1α gets activated and it splices XBP1, leading to the coding sequence of XBP1 to come into frame with the reporters, resulting in expression of the luciferase. Dual reporter RT4 cells were transiently transfected with cDNAs expressing both wild-type (WT-CADM3-HA) and mutant (Y172C-CADM3-HA) CADM3. Results showed that both firefly luciferase and nano luciferase levels in the mutant were significantly different than those obtained from transfections with the wild-type CADM3 ($p<0.001$, two-tailed t-test) (Figure 2A). This result suggests that a large fraction of the protein is retained in the endoplasmic reticulum (ER) triggering the UPR. To analyze the subcellular localization of CADM3, cells were transfected with both WT and mutant CADM3 plasmids and immunofluorescence labeled with the ER marker, GRP78-BiP. Confocal images show that both WT and mutant CADM3 cytoplasmic staining co-localizes in the ER (Figure 2B). When cells are co-transfected with both CADM3 and CADM4 (CADM4-myc) plasmids, a fraction of the CADM3 protein (both WT and mutant) is targeted to the membrane at cell-cell contact sites, where it co-localized with CADM4, although a large fraction remains retained in the cytoplasm (Figure 2C). In contrast, CADM4 appears more enriched at the membrane with minimum retention at the cytoplasm.

CADM3 mutant reaches the cell surface and bind CADM4

We have previously shown in rodents that glial CADM4 preferentially bind axonal CADM3 and CADM2 (Spiegel *et al.*, 2007; Elazar *et al.*, 2019). To examine whether the human mutation in *CADM3* affects its ability to interact with CADM4, we mutated the corresponding residue in the rat *Cadm3* cDNA (Y170C). The difference in the position of the mutation between rodents (Y170; XP_006250385.1) and humans (Y172; NP_067012.1) is due to the presence of additional two amino acids in position 5 of the human cDNA. Wild type and mutant CADM3 proteins carrying extracellular HA and intracellular Myc tags, were expressed in HEK293T transfected cells and as expected, were enriched in the cell membrane fraction (Figure 3A). However, in contrast to the

HA-tag antibody which recognized both the wild type and the mutated CADM3, an antibody directed to a short sequence in the extracellular region of CADM3 which contains the mutated tyrosine (QKPIITGYKSSLREKETATL (Spiegel *et al.*, 2007); showed preferential recognition of the wild type protein (Figure 3B). Nevertheless, immunolabeling of non-permeabilized COS7 cells expressing the doubly-tagged Cadm3 and Cadm3-Y170C, as well as a cell surface biotinylation experiment (Figure 3E), demonstrated that both the wild type and the mutant proteins reached the cell surface.

The Tyr172Cys CADM3 variant shows reduced co-localization with CADM4 at cellular contact sites in the plasma membrane

To investigate whether the mutation affects CADM3 cell localization, we performed immunofluorescence co-localization experiments. We used L-cells, which lack cell-cell adhesion molecules, stably expressing the human encoding CADM3 plasmids, WT-CADM3-GFP or Y172C-CADM3-GFP and co-transfected with CADM4-Myc. Cells imaged by conventional confocal microscopy show CADM4 co-localized with both WT and mutant CADM3 (Figure 2C). However, the spatial resolution of confocal microscopy is limited to 200 nm, which is well above the scale of the plasma membrane, therefore it cannot distinguish between adjacent molecules. To investigate a more precise localization of CADM3 in cells, we performed stochastic optical reconstruction microscopy (STORM) imaging which has a spatial resolution of 20nm (Rust *et al.*, 2006) (Figure 4E). While conventional confocal microscopy shows perfect co-localization between CADM3 and CADM4, STORM shows that CADM3 and CADM4 are located in close opposition at their respective plasma membranes (Figure 4A and 4B) with overlapping points predominantly localized between cell-cell contact sites (Figure 4B). In contrast, the mutant Y172C-CADM3 appears more disorganized with significantly decreased co-localization with CADM4 at contact sites (Figure 4C, 4D and 4F), and in addition, it showed predominant localization in the cytoplasm (Figure 4D). We cannot conclude based on that experiment whether the loss of co-localization is

due to a decreased ability of the mutant CADM3 to reach the membrane or due to impaired molecular interaction between CADM3 and CADM4. However, the decreased physical interaction between CADM3 and CADM4 could potentially compromise the axon-glia interaction. ~~These results further support impaired interaction between the axolemma and myelin, possibly due reduced interaction between CADM3 and CADM4.~~

Knock-in mouse model carrying the mutant allele

To further study the cellular and molecular mechanisms that underlie the pathogenicity of the human CADM3 mutation *in vivo*, we have generated knock-in mice carrying the corresponding mutation (*Cadm3*^{Y170C}). *Cadm3*^{Y170C} mice were born at the expected Mendelian frequency, were viable, and did not display any overt neurological abnormality. To examine the expression of the mutant allele we performed a reverse transcription (RT) PCR on RNA isolated from brains of WT, heterozygous and homozygous *Cadm3*^{Y170C} mice. This analysis showed that the mutant mRNA is expressed in both heterozygous and homozygous mice (Figure 6A). Western blot analysis using a CADM3 antibody that recognizes both the WT and the mutant protein revealed the presence of Cadm3-Y170C mutant in membrane protein extract prepared from three-months-old homozygous mice brains (Figure 6B). The expression of the mutant Cadm3-Y170C protein in these mice brains was similar to that of Cadm3 in WT mice. In contrast, Western blot analysis using the same antibody done on sciatic nerve extract isolated from three-months-old homozygous *Cadm3*^{Y170C} mice revealed that the expression of the mutant protein in the nerve was lower than the expression of CADM3 protein in nerves from WT mice of the same age (Figure. 6C-D). However, and in line with the lack of clear neurological phenotype, *Cadm3*^{Y170C} mice exhibit nerve conduction velocity similar to their WT control littermates (Figure 6E).

***Cadm3*^{Y170C} mice display abnormal axonal organization**

To analyze *Cadm3*^{Y172C} mice, we focused on potential molecular and structural abnormalities in

their myelin unit. Toluidine blue stained sections, as well as electron microscopy analysis of cross sections of sciatic nerves isolated from WT and homozygous *Cadm3*^{Y172C} mice showed no major difference in peripheral myelin except few individual fibers with thickened myelin, reminiscent of tomaculae seen in the patient sural nerve biopsy (Figure 6F). We also examined whether the mutation affect the molecular organization of the nerve, which is critical to its function. Immunofluorescence analysis of teased sciatic nerve preparations from homozygous *Cadm3*^{Y172C} mice using antibodies to Caspr (i.e., paranodal junction marker), and Kv1.2 (i.e., juxtaparanodal marker) and NrCAM (i.e., nodal marker), revealed abnormal accumulation of Kv1.2 channels around the juxtaparanodes, as well as in patches along the internodes and some aberrant appearance of Caspr at the edges of the paranodes (Figure 7A-C). The abnormal localization of Kv1.2 channels was mostly seen along larger caliber axons. We also compared the localization of Kv1.2 channels of Cadm3^{Y172C} mice to mice lacking either *Cadm3* (*Cadm3*$^{-/-}$) or *Cadm4* (*Cadm4*$^{-/-}$) (Gollan *et al.*, 2003). We noticed that the aberrant distribution of Kv1.2 channels in *Cadm3*^{Y170C} was milder than sciatic nerves isolated from *Cadm4*$^{-/-}$ mice, but surprisingly, completely absent in *Cadm3*$^{-/-}$ nerves (Figure 7D).

DISCUSSION

The present study identifies *CADM3* as a novel CMT disease gene. *CADM3* encodes for a neuronal specific cell-adhesion molecule localized to the internodal domain of myelinated fibers. It plays a major role in axon-glia interaction and domain organization maintenance in the internodes. Several studies have indicated that loss of axon-glia interactions lead to neurologic abnormalities in mice due to axonal and myelination dysfunction. Mice lacking the paranodal junctional component Caspr exhibit tremor, ataxia and motor paresis (Bhat *et al.*, 2001; Gollan *et al.*, 2003; Elazar *et al.*, 2019). Mice lacking *Cadm4*, the *Cadm3* interaction partner located in Schwann cells, results in a phenotype that resembles CMT, such as impaired motor function and reduced nerve conduction velocity (Golan *et al.*, 2013). In humans, biallelic mutations in *CNTNAP1* which encodes a

paranodal cell-adhesion molecule, the contactin-associated protein (Caspr), causes severe phenotypes including congenital hypomyelinating neuropathy-3 (Hengel *et al.*, 2017) and lethal congenital contracture syndrome (Lakhani *et al.*, 2017). Finally, biallelic mutations in neurofascin (*NFASC*) were also recently identify to cause neurodevelopmental impairment and peripheral demyelination (Efthymiou *et al*, 2019). Thus, the paranodal region is of recognized importance to the pathogenicity of CMT2; yet, we demonstrate this first direct genetic evidence for a cell adhesion molecule causing axonal CMT in patients. The clinical features we observed in the *CADM3* patients are in keeping with early onset autosomal dominant CMT2 with marked upper limb involvement and, in one family, brisk reflexes suggesting upper motor neuron involvement. There was phenotypic heterogeneity among our patients ranging from severe early onset (Family1-II:1) to a milder adult onset (Family3-I:2). Nerve conduction studies suggested a predominantly axonal neuropathy but with motor slowing in some patients. Unusually, all patients analyzed carry the same missense variant, Tyr172Cys, located in the extracellular domain adjacent to the second Ig loop. Mass spectrometry analysis studies revealed the formation of a new disulfide bond created by the mutated cysteine residue, indicating structural protein rearrangement. CADM proteins mediate specific cell-cell contact through homophilic and heterophilic interactions within the family of proteins. Trans interactions in this family occur through the first Ig-like domain, whereas the second Ig-like domain is necessary for cis interactions. Ig-like domains are stabilized by disulfide bonds, which maintain their structure. In the mutant CADM3, tyrosine 172 was changed to a cysteine residue. A missense variation that forms a new cysteine may perturb an existing disulfide bond and create an alternative bond with a nearby cysteine. The disulfide bond in the second Ig-like domain of CADM3 occurs between cysteines 186 and 243. The mutation is located in the beginning of the second Ig-like domain, thus the new cysteine 172 may interfere with the second disulfide bond, what could likely be disturbing the cis and not trans interactions of the protein.

Overexpression of the mutated *CADM3* in RT4 Schwann cells resulted in activation of the

unfolded protein response, suggesting protein misfolding as a potential pathological mechanism. We also explored the spatial resolution of cell-cell contact site at the nanoscale level with STORM microscopy to more precisely evaluate CADM3 interaction with CADM4. STORM images revealed significantly decreased co-localization between CADM3 and CADM4 at cell-cell contact sites, indicating axon-glia interface disturbance as a potential pathological mechanism of the disease. Interestingly, a sural nerve biopsy from family 1 revealed the presence of tomaculous swellings, further supporting abnormal axon-glia contacts. Similar abnormal structures were observed in *Cadm4* knock-out mice resulting in a CMT-like phenotype (Gollan *et al.*, 2013).

Our knock-in mouse model carrying the human Tyr172Cys mutation showed weaker expression of the mutant protein in sciatic nerve compared to *Cadm3* wild type mice. This result may indicate that the targeting of mutant Cadm3 to the axonal membrane or its stabilization at the axonal membrane is impaired. This result is in accordance with the observed reduction in its colocalization with CADM4 as observed by super resolution microscopy in L-cells. Our *in-vivo* analysis of teased sciatic nerve fibers from homozygous mice revealed abnormal accumulation of Kv1.2 channels around the juxtaparanodes and in patches along the internodes, as well as mild aberrant appearance of Caspr at the paranodes. We have previously showed that that absence of *Cadm4* in mice Schwann cells leads to abnormal organization of the underlying axonal membrane, whereas mice lacking *Cadm3* have normal PNS myelin (Gollan *et al.*, 2003). It was suggested that the difference between these two mutants reflects compensatory mechanism by other axonal CADM proteins. The presence of axonal organization abnormalities in Cadm3^{Y172C} mice may suggests that this mutation interferes not only with the interaction between CADM3 and CADM4, but also with the interactions between CADM4 and other axonal CADMs, or even other axoglial CAMs as recently was demonstrated in the CNS for CADM4, MAG and Caspr (Djannatian *et al.*, 2019; Elazar *et al.*, 2019). Humans, in contrast to the knock-out mice, which can survive without CADM3, appear to be intolerant to loss-of-function mutations in CADM3, since it is highly constraint for LoF mutations in gnomAD (pLI>0.9). In gnomAD the expected number of LoF

variants for CADM3 is 21.9, and the observed number is 1 variant for the canonical transcript, however that only allele is flagged as “low call.” Likewise, we didn’t observe any LoF mutation in our GENESIS database, which contains 9,569 exomes/genome. This observation suggests that the human *CADM3* is less redundant than the mouse gene and consequently a compensatory mechanism by other CADMs might be less effective.

The clinical and molecular findings of three independent families with axonal CMT, marked upper limb involvement, and brisk reflexes, harboring the same pathogenic variant in *CADM3*, unveils a new pathological mechanism involving axon-glial interaction abnormalities in patients with CMT.

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COMPETING INTERESTS

The authors report no competing interests.

SUPPLEMENTAL INFORMATION

Supplemental Figure 1: Tandem Mass Spectrometry analysis. MS/MS spectra showing detected disulfide-linked peptides.

Supplemental Figure 2: RFLP analysis of the proband and her parents (family 1) confirm the paternity of the father. The following microsatellites markers were used: D6S1552, D6S1624 and MOG-CA. The fragment sizes labeled in red and green correspond to father's and mother's respectively.

Supplemental Figure 3: Multiple sequence alignment between CADM1, CADM2, CADM3 and CADM4 with ClustalW. Sequence highlighted in blue indicates membrane signal peptide. The transmembrane region is underlined in orange. Cysteines highlighted in yellow form disulfide bonds of Ig loops. Patients mutation shown in red (Y172C).

FIGURE LEGENDS

Figure 1: *CADM3* mutation in CMT2 families. (A) Pedigree and Sanger sequencing traces of CMT2 families sharing the same heterozygous *CADM3* variant Tyr172Cys. **(B)** Diagram shows *CADM3* mutation at exon 5 (C.515A>G) and its corresponding position in the protein (Y172C) located in the extracellular domain between Ig1 and Ig2 loops. **(C)** Diagram shows native disulfide bonds represented by connected arrows (green) and new disulfide bond C172-C186 (red) generated by the mutation Y172C, which was detected by liquid chromatography with tandem mass spectrometry (LC-MS-MS).

Figure 2: Overexpression of *CADM3* mutant activates the unfolded protein response. (A) Dual reporter luciferase RT4 cells were transiently transfected with plasmids encoding the human wild-type (WT) and mutant (Y172C) *CADM3*. Increased levels of firefly and nano luciferase were observed in the mutant *CADM3*. **(B)** Immunofluorescence shows co-localization of both WT and mutant *CADM3* with the endoplasmic reticulum marker GPR78-BiP. **(C)** Cells co-transfected with CADM3 and CADM4 show CADM3 co-localization with CADM4 at cell-cell contact sites.

Figure 3: CADM3 mutant reaches the cell surface of transfected cells and interacts with CADM4. (**A**) WB analysis of total cell lysate (Total) and membrane preparation showing the expression of HA-tagged (tag located extracellularly) and Myc-tagged (tag located intracellularly) of rat *Cadm3* (WT) and *Cadm3* mutant (Y170C) in transfected HEK293 cells, using an antibody to HA. Cells transfected with GFP were used as control. Molecular size markers (kilodalton) are shown on the right. (**B**) Western blot analysis of membrane preparations from HEK293 cells expressing the same constructs as in panel A, using a polyclonal antibody directed to the extracellular domain of CADM3 or an antibody to HA-tag as indicated. Note that the CADM3 antibody recognize the wild type but not the mutant Cadm3 protein. Molecular size markers (kilodalton) are shown on the right. (**C**) Both wild type and mutant proteins are present at the cell

surface. Immunolabeling of HEK293 cells transfected with the double-tagged versions of wild type or mutant Cadm3 was done using either live (- Tx100) or permeabilized (+ Tx100) cells. The single channels and merged images are shown as indicated. (**D**) Surface biotinylation of HEK293 cells transfected with wild type (WT), mutant (Y170C), or GFP, followed by immunoprecipitation (IP) with an antibody to Myc and immunoblot using streptavidin-HRP (Strep-HRP) or an anti-HA antibody as indicated. Molecular size markers (kilodalton) are shown on the right. (**E**) The mutant Cadm3 interacts with the extracellular domain of Cadm4. COS7 cells expressing Myc-tagged wild type (WT) or mutant (Y170C) Cadm3 were incubated with medium containing a soluble extracellular domain of Cadm4 fused to Fc region of human IgG. Expression of Cadm3 and binding of Cadm4-Fc was monitored using antibodies to human Fc (Fc-Binding) and Myc tag, as indicated. Dapi-labeling (blue) of cell nuclei is shown in the merged images. Scale bars: 20μm.

Figure 4: Super-resolution microscopy shows CADM3 co-localization with CADM4 at plasma membrane contact sites. (A-B) Storm imaging reveals close proximity between WT-CADM3 (red) and CADM4 (green) at cell-cell contact sites in L-cells, which lack cell-cell adhesion molecules. A representative image shows two distinct color lines formed at cell contact sites with some co-localization overlapping between red and green. **(B)** Zoomed-in views from regions demarcated by purple boxes. **(C-D)** Mutant Y172C-CADM3 shows less specific localization to the membrane and diminished co-localization with CADM4. **(D)** Zoomed-in views from regions demarcated by purple boxes. **(E)** Illustration of different results obtained with conventional confocal microscopy and STORM imaging. Adjacent CADM molecules cannot be distinguished by conventional microscopy which shows co-localization (yellow), while STORM is able distinct the molecules (red and green). **(F)** Quantification of co-localization between CADM4 and CADM3. Twenty images from each group were obtained to calculate the results (t-test, $p<0.005$).

Figure 5: Nerve biopsy from CADM3 patient. (A) A cross section of sural nerve from patient of family 2 show a reduced number of large myelinated fibers and very few groups of small regenerating fibers. Of interest, we overserved a number of abnormally myelinated fibers with thick folded myelin. A teased fiber from the same biopsy **(B)** shows such a focal myelin swelling or hypermyelination, a histological feature of demyelinating peripheral neuropathies.

Figure 6: Evaluation of mouse *Cadm3*^{Y170C} mutation. (A) RT-PCR analysis using WT and mutant-specific primers showing the presence of the wild type and mutant Cadm3 variants in the mice. **(B)** WB analysis showing the expression of *Cadm3* in wild type and the knock-in homozygous mice brains. **(C)** WB showing reduced expression of the mutant protein in sciatic nerves. **(D)** Quantification of panel C. **(E)** Mutant (KI) and wild type mice show similar nerve conduction velocity. **(F)** *Cadm3*-Y170C mutant sciatic nerve exhibits normal myelin. Toluidine blue stained cross sections of sciatic nerves isolated from one half-month-old mouse of the indicated genotype (left panel). Electron microscopy analysis of sciatic nerves showing normal myelin morphology in wild type (WT) and mutant (Cadm3-Y170C) nerves. Scale bars: (A) 10um, (B) 2um, (C) 0.4um.

Figure 7: *Cadm3*^{Y170C} mutant nerves exhibit abnormal axonal organization. Tease sciatic nerves were immunolabeled using antibodies to Kv1.2, NrCAM and Caspr as indicated. **(A-B)**. Abnormal (arrowheads) distribution of Kv1.2 potassium channels. The location of the internodes (IND) is marked in **(B)** and the nodes are marked by asterisks. **(C)** Mild abnormalities in Caspr (line marked by an arrowhead in lower left panel), as well as abnormal distribution of Caspr along the internodes (Arrowheads in lower right panel). **(D)** Kv1.2 abnormalities are present in the knock-in (left) but not in *Cadm3* null mice (middle) and are similar (although less severe) than *Cadm4* null mice (right). The location of the nodes is marked by asterisks. Scale bars: 20μm (A, B, C left, D), 10μm (C right).

TABLES

Table 1. Clinical features of CADM3 patients

Table 2. Motor nerve studies

Table 3. Sensory nerve studies

Table S1. CMT-associated genes analyzed by whole exome sequencing

Table S2. Fisher's exact test statistic shows a value of $p<0.01$.

	Tyr172Cys alleles	**Wild-type chromosomes**	**Total**
CMT chromosomes (GENESIS)	3	2,508	2,514
Controls (GENESIS)	0	16,624	16,624
Total	3	19,132	19,138

REFERENCES

Bai Y, Wu X, Brennan KM, Wang DS, D'Antonio M, Moran J, *et al.* Myelin protein zero mutations and the unfolded protein response in Charcot Marie Tooth disease type 1B. Ann Clin Transl Neurol 2018; 5(4): 445-55.

Bhat MA, Rios JC, Lu Y, Garcia-Fresco GP, Ching W, St Martin M, *et al.* Axon-glia interactions and the domain organization of myelinated axons requires neurexin IV/Caspr/Paranodin. Neuron 2001; 30(2): 369-83.

Chen MS, Kim H, Jagot-Lacoussiere L, Maurel P. Cadm3 (Necl-1) interferes with the activation of the PI3 kinase/Akt signaling cascade and inhibits Schwann cell myelination in vitro. Glia 2016; 64(12): 2247-62.

Cooper GM, Stone EA, Asimenos G, Program NCS, Green ED, Batzoglou S, *et al.* Distribution and intensity of constraint in mammalian genomic sequence. Genome Res 2005; 15(7): 901-13.

Djannatian M, Timmler S, Arends M, Luckner M, Weil MT, Alexopoulos I, *et al.* Two adhesive systems cooperatively regulate axon ensheathment and myelin growth in the CNS. Nat Commun 2019; 10(1): 4794.

Efthymiou S, Salpietro V, Malintan N, Poncelet M, Kriouile Y, Fortuna S, De *et al*. Biallelic mutations in neurofascin cause neurodevelopmental impairment and peripheral demyelination. Brain. 2019 Oct 1;142(10):2948-2964.

Elazar N, Vainshtein A, Rechav K, Tsoory M, Eshed-Eisenbach Y, Peles E. Coordinated internodal and paranodal adhesion controls accurate myelination by oligodendrocytes. J Cell Biol 2019; 218(9): 2887-95.

Golan N, Kartvelishvily E, Spiegel I, Salomon D, Sabanay H, Rechav K, *et al.* Genetic deletion of Cadm4 results in myelin abnormalities resembling Charcot-Marie-Tooth neuropathy. J Neurosci 2013; 33(27): 10950-61.

Gollan L, Salomon D, Salzer JL, Peles E. Caspr regulates the processing of contactin and inhibits its binding to neurofascin. J Cell Biol 2003; 163(6): 1213-8.

Gonzalez M, Falk MJ, Gai X, Postrel R, Schule R, Zuchner S. Innovative genomic collaboration using the GENESIS (GEM.app) platform. Human mutation 2015; 36(10): 950-6.

Hengel H, Magee A, Mahanjah M, Vallat JM, Ouvrier R, Abu-Rashid M, *et al.* CNTNAP1 mutations cause CNS hypomyelination and neuropathy with or without arthrogryposis. Neurol Genet 2017; 3(2): e144.

Kakunaga S, Ikeda W, Itoh S, Deguchi-Tawarada M, Ohtsuka T, Mizoguchi A, *et al.* Nectin-like molecule-1/TSLL1/SynCAM3: a neural tissue-specific immunoglobulin-like

cell-cell adhesion molecule localizing at non-junctional contact sites of presynaptic nerve terminals, axons and glia cell processes. J Cell Sci 2005; 118(Pt 6): 1267-77.

Lakhani S, Doan R, Almureikhi M, Partlow JN, Al Saffar M, Elsaid MF, *et al.* Identification of a novel CNTNAP1 mutation causing arthrogryposis multiplex congenita with cerebral and cerebellar atrophy. Eur J Med Genet 2017; 60(5): 245-9.

Maurel P, Einheber S, Galinska J, Thaker P, Lam I, Rubin MB, *et al.* Nectin-like proteins mediate axon Schwann cell interactions along the internode and are essential for myelination. J Cell Biol 2007; 178(5): 861-74.

Murphy SM, Herrmann DN, McDermott MP, Scherer SS, Shy ME, Reilly MM, *et al*. Reliability of the CMT neuropathy score (second version) in Charcot-Marie-Tooth disease. J Peripher Nerv Syst. 2011 Sep;16(3):191-8.

Rentzsch P, Witten D, Cooper GM, Shendure J, Kircher M. CADD: predicting the deleteriousness of variants throughout the human genome. Nucleic Acids Res 2019; 47(D1): D886-D94.

Richards S, Aziz N, Bale S, Bick D, Das S, Gastier-Foster J, *et al.* Standards and guidelines for the interpretation of sequence variants: a joint consensus recommendation of the American College of Medical Genetics and Genomics and the Association for Molecular Pathology. Genet Med 2015; 17(5): 405-24.

Rust MJ, Bates M, Zhuang X. Sub-diffraction-limit imaging by stochastic optical reconstruction microscopy (STORM). Nat Methods 2006; 3(10): 793-5.

Salzer JL, Brophy PJ, Peles E. Molecular domains of myelinated axons in the peripheral nervous system. Glia 2008; 56(14): 1532-40.

Spiegel I, Adamsky K, Eshed Y, Milo R, Sabanay H, Sarig-Nadir O, *et al.* A central role for Necl4 (SynCAM4) in Schwann cell-axon interaction and myelination. Nat Neurosci 2007; 10(7): 861-9.

Zhou Y, Du G, Hu X, Yu S, Liu Y, Xu Y, *et al.* Nectin-like molecule 1 is a protein 4.1N associated protein and recruits protein 4.1N from cytoplasm to the plasma membrane. Biochim Biophys Acta 2005; 1669(2): 142-54.