

Geoff Burnstock, purinergic signalling, and chemosensory control of breathing

Alexander V. Gourine and K. Michael Spyer

Centre for Cardiovascular and Metabolic Neuroscience, Department of Neuroscience, Physiology & Pharmacology, University College London, Gower Street, London WC1E 6BT, UK

Abstract

This article is the authors' contribution to the tribute issue in honour of Geoffrey Burnstock, the founder of this journal and the field of purinergic signalling. We give a brief account of the results of experimental studies which at the beginning received valuable input from Geoff, who both directly and indirectly influenced our research undertaken over the last two decades. Research into the mechanisms controlling breathing identified ATP as the common mediator of the central and peripheral chemosensory transduction. Studies of the sources and mechanisms of chemosensory ATP release in the CNS suggested that this signalling pathway is universally engaged in conditions of increased metabolic demand by brain glial cells - astrocytes. Astrocytes appear to function as versatile CNS metabolic sensors that detect changes in brain tissue pH, CO₂, oxygen, and cerebral perfusion pressure. Experimental studies on various aspects of astrocyte biology generated data indicating that the function of these omnipresent glial cells and communication between astrocytes and neurons are governed by purinergic signalling, - first discovered by Geoff Burnstock in the 70's and researched through his entire scientific career.

Introduction

It is a real privilege to have this opportunity to present a review of some of our research in a volume dedicated to the memory of Geoff Burnstock. He was a colleague, collaborator, and friend who both directly and indirectly influenced our research undertaken over the last two decades.

One of us (KMS) first met Geoff in 1974 at an international meeting in Tokyo which led to several informal meetings over the years in London. We were together in New York in 1978 attending a meeting organised by Chandler Brooks and spent a great deal of extramural time sampling the dining delights of the city. Geoff then tried to recruit KMS to his Department of Anatomy and Developmental Biology at the University College London (UCL). The Department had an extraordinary mixture of truly excellent scientists and had a strong emphasis on Neuroscience. Geoff offered a wonderful suite of laboratories, but for many reasons it was not the right time for KMS to move from Birmingham and the offer was declined at that time. However, two years later KMS moved to London to head the Department of Physiology at the Royal Free Hospital School of Medicine.

It took until the mid-90's for a collaboration to develop between the Burnstock and Spyer laboratories, coinciding with KMS becoming Head of UCL's Department of Physiology. This collaboration centred on the role of ATP and adenosine in the CNS control of circulation. Geoff was forceful in persuading us that the effects of adenosine are "not that important" and that we should focus on ATP-mediated signalling. In a series of pilot studies with Theresa Thomas it slowly emerged that signalling via P2 receptors might indeed play an important role in modulation of the activity of brainstem circuits that control breathing.

Our collaboration intensified in 1997 when Geoff retired from the headship of the Anatomy Department and moved to the Royal Free Hospital School of Medicine to occupy laboratory space adjacent to the Spyer's laboratories. This led to the creation of the Autonomic Neuroscience Institute, with Burnstock and Spyer as co-directors, aimed at developing a unique centre for research on the autonomic nervous system. This venture was very successful in regard of allowing Geoff to continue to be fully research active and motivate the line enquiry described in detail below. The Autonomic Neuroscience

Institute was closed on Geoff's return to the University of Melbourne in 2017 and succeeded by UCL Centre for Cardiovascular and Metabolic Neuroscience.

In 1975 Geoff together with Marcello Costa published a book "Adrenergic Neurons: Their Organization, Function and Development in the Peripheral Nervous System". AVG was familiar with Geoff's name from an early age, as the Russian edition of the book edited by AVG's father Valery Gourine and published in 1979 was always in a prominent place in the home library. AVG joined the Department of Physiology in 2000 to work in collaboration with KMS on the mechanisms of chemosensory control of breathing and had the first opportunity to meet Geoff in person. The subsequent sections of this article give a general summary of the outcomes of our research on the role of ATP in the chemosensory control of breathing which in its early years received valuable input from Geoff, which is evident from several joint publications.

Early work on the role of adenosine in the brainstem mechanisms of cardiovascular control

Early studies on purinergic signalling in the brainstem mechanisms of cardiovascular control focused on the role played by adenosine (Spyer & Thomas, 2000; Spyer *et al.*, 1997). In experiments using experimental animals (rats) it was shown that the cardiovascular changes that accompany the defence reaction are associated with the release of adenosine in the nucleus of the solitary tract and the rostral ventrolateral medulla (RVLM). Blockade of adenosine receptors was found to modify the pressor response, indicating that adaptive cardiovascular changes during fight-or-flight reactions are modulated by adenosine. The pharmacological data suggested that this adenosine is likely to be produced extracellularly following the breakdown of the released ATP (St Lambert *et al.*, 1997). This hypothesis was subsequently tested and supported by experimental data (Dale *et al.*, 2002) obtained in collaboration with Nick Dale (University of Warwick) who developed enzymatic microelectrode biosensors for real-time detection of adenosine release in the brain (Llaudet *et al.*, 2003).

First studies of the ATP effects in the brainstem

Subsequent work undertaken in close collaboration with Burnstock's laboratory aimed at understanding the role of ATP in modulation of the neuronal activity in the RVLM. First it

was found that ATP acting via P2X and P2Y receptors has a strong excitatory effect on the majority of RVLM neurons, including cells with monosynaptic projections to the spinal cord – pre-sympathetic (or sympathoexcitatory) neurons (Ralevic *et al.*, 1999). Strong cardiovascular and respiratory responses were recorded following microinjections of ATP or P2X receptor agonists into the RVLM (Thomas *et al.*, 2001). It was also observed that in anaesthetized rats microinjections of the broad spectrum P2 receptor blocker suramin or P2X receptor agonist $\alpha\beta$ -metATP (to desensitize the receptors) into the RVLM region reduced the respiratory responses to CO₂ (Thomas *et al.*, 1999). CO₂-induced increases in the activity of the medullary inspiratory neurons were found to be blocked by P2 receptor antagonists (Thomas & Spyer, 2000).

Studies of the role of P2X receptors in the chemosensory control of breathing

Reduction of the respiratory response to CO₂ in conditions of P2 receptor blockade localised to the ventral regions of the brainstem led to a hypothesis that central respiratory chemosensitivity to CO₂ is mediated via a proxy of pH changes detected by P2X₂ receptors, known to be highly sensitive to changes in pH within the physiological range (King *et al.*, 1997). In the absence of selective P2X₂ receptor ligands, this hypothesis was tested in P2X₂ and P2X₂/P2X₃ receptor knockout mice, which became available for this project from a collaboration between Geoff and the research group led by Anthony Ford in Roche Pharmaceutical (Palo Alto). In conscious mice, the respiratory responses to the increases in the level of inspired CO₂ were found to be unaffected by genetic deletion of P2X₂, P2X₃ or both receptor subunits (Rong *et al.*, 2003). Moreover, analysis of the P2 receptor expression demonstrated that only a small proportion of inspiratory neurons identified within the ventral respiratory column express P2X₂ (Gourine *et al.*, 2003), - a result supported by a recent report showing low level of P2X₂ expression in the ventral regions of the brainstem (Kim *et al.*, 2020).

Demonstration of the pivotal role of ATP as a key mediator of chemosensory signalling in the carotid body

While the respiratory sensitivity to CO₂ was unaffected in P2X₂ knockout mice, the responses to hypoxia were found to be dramatically reduced in conditions of P2X₂ receptor deficiency (Rong *et al.*, 2003), pointing to a critical role of ATP-mediated signalling in the carotid body function. In an *ex vivo* preparation of the carotid body,

Weifang Rong observed that the increases in the carotid sinus nerve discharge evoked by hypoxia are dramatically reduced by pharmacological P2X receptor blockade, or in conditions of P2X₂ receptor deficiency (Rong *et al.*, 2003). Further reductions of the carotid body response to hypoxia were observed in P2X₂/P2X₃ double knockout mice, suggesting that the ventilatory responses in low oxygen conditions are mediated by ATP acting at heteromeric P2X_{2/3} receptors expressed by the afferent terminals of the carotid sinus nerve (Gourine, 2005). The resulting publication (Rong *et al.*, 2003) was the last in a series of articles published from the collaboration between Burnstock and Spyer laboratories and has led to ATP being generally regarded as the key signalling molecule of chemosensory transduction in the carotid body. Moreover, the carotid body's P2X receptors are now recognized as a potential target for the treatment of circulatory system disease (Pijacka *et al.*, 2016).

Studies of the chemosensory ATP release in the brainstem

The data discussed above did not support the hypothesis that pH sensitivity of P2X receptors expressed by the neurons of the medullary respiratory network underlie central respiratory sensitivity to CO₂. This led to the development of a modified hypothesis that the central respiratory chemosensitivity is mediated not by pH-sensitivity of P2X receptors expressed by the respiratory neurons, but by the CO₂/acidification-induced release and actions of ATP. We suggested that chemosensory stimuli induce the release of ATP in the brain and this ATP activates neurons of the medullary respiratory networks to trigger adaptive changes in breathing (Gourine, 2005). This hypothesis was tested in collaboration with Nick Dale and Enrique Llaudet (University of Warwick) who at that time were working on the development of enzymatic amperometric biosensors for real time detection of ATP release (Llaudet *et al.*, 2005). The very first experiments with the new biosensor showed that systemic chemosensory stimuli, such as increases in inspired CO₂ (hypercapnia) or decreases in inspired oxygen (hypoxia), trigger ATP release from the chemosensitive regions of the ventral medulla oblongata (Gourine *et al.*, 2005b; Gourine *et al.*, 2005a; Huckstepp *et al.*, 2016). Blockade of ATP receptors was found to reduce the respiratory responses to both hypercapnia and hypoxia (Gourine *et al.*, 2005b; Gourine *et al.*, 2005a)), suggesting that ATP mediates (at least in part) the effects of chemosensory stimuli on the activity of the brainstem respiratory network (but these actions of ATP are mediated by P2 receptors other than P2X₂ or P2X₃, considering the phenotype of P2X₂/P2X₃ receptor knockout mice, as discussed above).

Identifying the source(s) and mechanisms of ATP release in response to CO₂

Electrically non-excitabile cells communicate via the release of ATP, therefore, we next tested the hypothesis that in response to chemosensory stimuli (CO₂ or hypoxia) ATP is released by glial cells. Earlier investigators had noted dense glial layer covering the ventral surface of the brainstem at the locations corresponding to the classical chemosensory areas (Loeschcke, 1982) and the sites of chemosensory ATP release (Gourine *et al.*, 2005a) (for a detailed histological analysis of brainstem astrocytes see (Sheikhabaei *et al.*, 2018a)). Subsequent studies conducted in London by AVG in collaboration with Sergey Kasparov (University of Bristol) and in Warwick by Nick Dale, revealed distinct and parallel mechanisms underlying the sensitivity of brainstem astrocytes to changes in pH and CO₂, leading to the release of ATP as a common mediator of chemosensory signalling (Huckstepp *et al.*, 2010b;Gourine *et al.*, 2010) (Figure 1a). It was found that in the brainstem astrocytes, acidification activates Na⁺/HCO₃⁻ cotransport, leading to increases in [Na⁺]_i, activation of the Na⁺/Ca²⁺ exchanger to operate in a reverse mode, Ca²⁺ entry and exocytosis of ATP-containing vesicular compartments (Kasymov *et al.*, 2013;Turovsky *et al.*, 2016). CO₂ is sensed directly by connexin 26 hemichannels which increase their open probability (allowing egress of ATP) proportionally to the concentration of CO₂ which forms carbamate bridges between subunits (Huckstepp *et al.*, 2010a;Meigh *et al.*, 2013;Dospinescu *et al.*, 2019).

Follow up studies using *in vivo* animal models provided further evidence that ATP released by brainstem astrocytes contributes to the development of the ventilatory responses to CO₂ and hypoxia (Huckstepp *et al.*, 2010b;Gourine *et al.*, 2010;Angelova *et al.*, 2015;Sheikhabaei *et al.*, 2018b;van de Wiel *et al.*, 2020). An important role played by astrocytes and purinergic signalling in mediating the effects of chemosensory stimuli on the activities of the brainstem cardiovascular and respiratory control networks is supported by the results of experimental studies conducted by other research groups (Lorier *et al.*, 2007;Lorier *et al.*, 2008;Huxtable *et al.*, 2009;Huxtable *et al.*, 2010;Wenker *et al.*, 2010;Zwicker *et al.*, 2011;Wenker *et al.*, 2012;Funk, 2013;Sobrinho *et al.*, 2014;Barna *et al.*, 2016;Rajani *et al.*, 2016;Cinelli *et al.*, 2017;Sobrinho *et al.*, 2017;Hawkins *et al.*, 2017;Rajani *et al.*, 2018;Reklow *et al.*, 2019;Patterson *et al.*, 2021). The notion that ATP release by astrocytes contributes to the hypoxic ventilatory response centrally was met with some scepticism and the readers are invited to evaluate

the arguments in favour and against this hypothesis presented in a series of opinion articles (Gourine and Funk, 2017; Funk and Gourine, 2018a, 2018b; Teppema, 2018).

Understanding the role of purinergic signalling in the neuronal-activity dependent control of cerebral blood flow and local brain tissue pH

There is significant evidence that neuronal activation leads to the release of purines (Pankratov *et al.*, 1998; Pascual *et al.*, 2005; Pankratov *et al.*, 2006; Wall & Dale, 2013; Sims & Dale, 2014; Badimon *et al.*, 2020; Peng *et al.*, 2020). What is the functional significance of ATP and adenosine release that parallels the increases in the neuronal activity? Using biosensor recordings in anaesthetised rats, we demonstrated the release of ATP from the central terminals of visceral afferents (Gourine *et al.*, 2008) and in the forepaw region of the cerebral cortex in response to activation of somatosensory pathways (Wells *et al.*, 2015). We also found that ATP-mediated signalling plays an important role in the mechanisms of neurovascular coupling in the cerebral cortex (Wells *et al.*, 2015). In a parallel study involving our laboratory, ATP was shown to mediate neuronal activity-dependent cerebrovascular responses at the capillary level (Mishra *et al.*, 2016). More recent data suggest that ATP triggers bicarbonate secretion by astrocytes and this release of bicarbonate helps to maintain local brain extracellular pH homeostasis in conditions of enhanced acid loads associated with increases in neuronal activity (Theparambil *et al.*, 2020) (Figure 1b).

Studies of the role of purinergic signalling in the control of circulation and breathing in pathological conditions

Astroglial dysfunction had been shown to contribute to neuropathology and disordered breathing pattern in Rett's syndrome, - an autism spectrum disorder caused by loss of function of the transcription factor MeCP2 (Lioy *et al.*, 2011). Mouse models of the disease showed that MeCP2 deficiency impairs the ability of brainstem astrocytes to detect changes in CO₂/H⁺ (Turovsky *et al.*, 2015), while MeCP2 deletion specifically from astrocytes markedly reduces the ventilatory sensitivity to CO₂ (Garg *et al.*, 2015). These data provided further evidence supporting the hypothesis of an important role played by astrocytes and purinergic signalling in chemosensory control of breathing.

There is also evidence that upregulated chemosensory glial responses leading to the release and high 'ambient' concentrations of purines may play an important role in the mechanisms underlying sympathetic activation which accompanies and contributes to the progression of the circulatory system diseases, such as hypertension and heart failure. Experiments in animal models showed that blockade of ATP-mediated signalling in the RVLM slows the remodelling process in heart failure induced by myocardial infarction (Marina *et al.*, 2013), and reduces systemic arterial blood pressure in hypertension (Marina *et al.*, 2015).

Studies of the mechanosensory properties of astrocytes

In the late 90's Geoff proposed a concept of purinergic mechanosensory transduction, involving ATP as one of the signalling molecules released by cells in response to mechanical stress (Burnstock, 1999). Studies in mice with genetic deletion of P2X₂/P2X₃ receptors provided strong experimental evidence that this mechanism operates in the peripheral organs, such as the bladder (Cockayne *et al.*, 2000; Vlaskovska *et al.*, 2001). Recent results suggest that Geoff's concept also applies to our understanding of mechanosensory signalling in the brain. Experimental evidence was obtained suggesting that astrocytes function as intracranial baroreceptors that detect decreases in brain perfusion and trigger compensatory increases in arterial blood pressure and heart rate to preserve cerebral blood flow and oxygen delivery (Marina *et al.*, 2020). TRPV4-dependent opening of connexin 43 hemichannels leading to the release of ATP appears to be the key central event underlying mechanosensory signalling in astrocytes (Turovsky *et al.*, 2020).

Conclusion

This article gives a brief account of the research which at the beginning was motivated by the results of exploratory studies undertaken in collaboration with Geoff's laboratory. When in the year 2000 we started to explore in detail the potential role of ATP in the mechanisms underlying chemosensory control of breathing, all our research methods were tuned to study neurons and the first series of the experiments focused on studies of the neuronal mechanisms and neuronal responses. The first breakthrough came in late 2002 with the development of purine biosensors by Nick Dale and our joint first recordings of chemosensory release of ATP in the living brain. Release of ATP pointed to

the potential involvement of astrocytes, which at that time were generally considered to merely provide neurons with structural and metabolic support, while the research methods to study astrocytes *in vivo* were at early stages of development. The next significant advance came ~2008 as a result of collaboration with Sergey Kasparov and Anja Teschemacher (University of Bristol) who pioneered the use of viral vectors to target brainstem astrocytes; first to express genetically-encoded Ca^{2+} sensors and then light-sensitive proteins, allowing monitoring and control of astroglial activity *in vivo*. All the subsequent studies on various aspects of astrocyte biology generated data, indicating that the function of these omnipresent glial cells and communication between astrocytes and neurons are governed by purinergic signalling, - first discovered by Geoff Burnstock in the 70's and researched through his entire scientific career. Our collaboration with Geoff was hugely valuable to us. It allowed us to take initial steps that would otherwise have been difficult in a timely manner. He did not always understand the issues that were bedevilling us, but he knew what the end point might be and was happy to trust our judgement on the way to proceed. We believe he would take pride in what our research has achieved and particularly in the body of evidence suggesting that purinergic mechanisms may have a true potential in providing novel therapeutic targets for the treatments of some common respiratory and cardiovascular diseases.

Acknowledgements: Results of the authors' experimental studies described in this review article were obtained with generous support of Biotechnology and Biological Sciences Research Council, British Heart Foundation and The Wellcome Trust. AVG is a Wellcome Trust Senior Research Fellow (Ref. 200893).

Disclosures: No conflicts of interest, financial or otherwise, are declared by the authors.

Author contributions: AVG and KMS drafted, edited, and approved the final version of manuscript.

Author notes: Address for correspondence: A. V. Gourine, Centre for Cardiovascular and Metabolic Neuroscience, Department of Neuroscience, Physiology & Pharmacology, University College London, London WC1E 6BT, United Kingdom. a.gourine@ucl.ac.uk

Figure legend

Figure 1 | Purinergic signalling mediates communication between astrocytes and neurons in conditions of increased metabolic demand. **a**, Astrocytes function as versatile metabolic sensors of the brain milieu, exquisitely sensitive to changes in brain tissue pH, partial pressures of oxygen and carbon dioxide, as well as cerebral perfusion pressure. In the brainstem, astrocytes are adjacent to, and intermingled with, the networks of neurons that generate and modulate the central respiratory and sympathetic drives. Brainstem astrocytes respond to the potential metabolic threats and via the release of ATP stimulate breathing and increase sympathetic activity (Marina *et al.*, 2018). **b**, In the cerebral cortex, astrocytes contribute to the protection of the brain milieu from acidification locally. At least one third of all astrocytes release bicarbonate to buffer extracellular H⁺ loads associated with increases in neuronal activity. The underlying signalling mechanism involves neuronal activity-dependent release of ATP triggering bicarbonate secretion by astrocytes via activation of P2Y₁ receptors (Theparambil *et al.*, 2020).

References

- Angelova PR, Kasymov V, Christie I, Sheikhabaehi S, Turovsky E, Marina N, Korsak A, Zwicker J, Teschemacher AG, Ackland GL, Funk GD, Kasparov S, Abramov AY, & Gourine AV (2015). Functional oxygen sensitivity of astrocytes. *J Neurosci* **35**, 10460-10473.
- Badimon A, Strasburger HJ, Ayata P, Chen X, Nair A, Ikegami A, Hwang P, Chan AT, Graves SM, Uweru JO, Ledderose C, Kutlu MG, Wheeler MA, Kahan A, Ishikawa M, Wang YC, Loh YE, Jiang JX, Surmeier DJ, Robson SC, Junger WG, Sebra R, Calipari ES, Kenny PJ, Eyo UB, Colonna M, Quintana FJ, Wake H, Gradinaru V, & Schaefer A (2020). Negative feedback control of neuronal activity by microglia. *Nature* **586**, 417-423.
- Barna BF, Takakura AC, Mulkey DK, & Moreira TS (2016). Purinergic receptor blockade in the retrotrapezoid nucleus attenuates the respiratory chemoreflexes in awake rats. *Acta Physiol (Oxf)* **217**, 80-93.
- Burnstock G (1999). Release of vasoactive substances from endothelial cells by shear stress and purinergic mechanosensory transduction. *J Anat* **194**, 335-342.
- Cinelli E, Iovino L, & Mutolo D (2017). ATP and astrocytes play a prominent role in the control of the respiratory pattern generator in the lamprey. *J Physiol* **595**, 7063-7079.
- Cockayne DA, Hamilton SG, Zhu QM, Dunn PM, Zhong Y, Novakovic S, Malmberg AB, Cain G, Berson A, Kassotakis L, Hedley L, Lachnit WG, Burnstock G, McMahon SB, & Ford AP (2000). Urinary bladder hyporeflexia and reduced pain-related behaviour in P2X₃-deficient mice. *Nature* **407**, 1011-1015.
- Dale N, Gourine AV, Llaudet E, Bulmer D, Thomas T, & Spyer KM (2002). Rapid adenosine release in the nucleus tractus solitarius during defence response in rats: real-time measurement *in vivo*. *Journal of Physiology* **544**, 149-160.
- Dospinescu VM, Nijjar S, Spanos F, Cook J, de WE, Biscotti MA, Gerdol M, & Dale N (2019). Structural determinants of CO₂-sensitivity in the beta connexin family suggested by evolutionary analysis. *Commun Biol* **2**, 331.
- Funk GD (2013). Neuromodulation: purinergic signaling in respiratory control. *Compr Physiol* **3**, 331-363.
- Funk GD, Gourine AV (2018a). CrossTalk proposal: a central hypoxia sensor contributes to the excitatory hypoxic ventilatory response. *J Physiol* **596**, 2935-2938
- Funk GD, Gourine AV (2018b). Rebuttal from Gregory D. Funk and Alexander V. Gourine. *J Physiol* **596**, 2943-2944.
- Garg SK, Liroy DT, Knopp SJ, & Bissonnette JM (2015). Conditional depletion of methyl-CpG-binding protein 2 in astrocytes depresses the hypercapnic ventilatory response in mice. *J Appl Physiol (1985)* **119**, 670-676.
- Gourine AV (2005). On the peripheral and central chemoreception and control of breathing: an emerging role of ATP. *J Physiol* **568**, 715-724.
- Gourine AV, Atkinson L, Deuchars J, & Spyer KM (2003). Purinergic signalling in the medullary mechanisms of respiratory control in the rat: respiratory neurones express the P2X₂ receptor subunit. *J Physiol* **552**, 197-211.
- Gourine AV, Dale N, Korsak A, Llaudet E, Tian F, Huckstepp R, & Spyer KM (2008). Release of ATP and glutamate in the nucleus tractus solitarius mediate pulmonary stretch receptor (Breuer-Hering) reflex pathway. *J Physiol* **586**, 3963-3978.
- Gourine AV, Funk GD (2017). On the existence of a central respiratory oxygen sensor. *J Appl Physiol (1985)*. **123**, 1344-1349.

- Gourine AV, Kasymov V, Marina N, Tang F, Figueiredo MF, Lane S, Teschemacher AG, Spyer KM, Deisseroth K, & Kasparov S (2010). Astrocytes control breathing through pH-dependent release of ATP. *Science* **329**, 571-575.
- Gourine AV, Llaudet E, Dale N, & Spyer KM (2005a). ATP is a mediator of chemosensory transduction in the central nervous system. *Nature* **436**, 108-111.
- Gourine AV, Llaudet E, Dale N, & Spyer KM (2005b). Release of ATP in the ventral medulla during hypoxia in rats: role in hypoxic ventilatory response. *J Neurosci* **25**, 1211-1218.
- Hawkins VE, Takakura AC, Trinh A, Malheiros-Lima MR, Cleary CM, Wenker IC, Dubreuil T, Rodriguez EM, Nelson MT, Moreira TS, & Mulkey DK (2017). Purinergic regulation of vascular tone in the retrotrapezoid nucleus is specialized to support the drive to breathe. *Elife* **6**.
- Huckstepp RT, Eason R, Sachdev A, & Dale N (2010a). CO₂-dependent opening of connexin 26 and related beta connexins. *J Physiol* **588**, 3921-3931.
- Huckstepp RT, Id BR, Eason R, Spyer KM, Dicke N, Willecke K, Marina N, Gourine AV, & Dale N (2010b). Connexin hemichannel-mediated CO₂-dependent release of ATP in the medulla oblongata contributes to central respiratory chemosensitivity. *J Physiol* **588**, 3901-3920.
- Huckstepp RT, Llaudet E, & Gourine AV (2016). CO₂-induced ATP-dependent release of acetylcholine on the ventral surface of the medulla oblongata. *PLoS One* **11**, e0167861.
- Huxtable AG, Zwicker JD, Alvares TS, Ruangkittisakul A, Fang X, Hahn LB, Posse de CE, Baker GB, Ballanyi K, & Funk GD (2010). Glia contribute to the purinergic modulation of inspiratory rhythm-generating networks. *J Neurosci* **30**, 3947-3958.
- Huxtable AG, Zwicker JD, Poon BY, Pagliardini S, Vrouwe SQ, Greer JJ, & Funk GD (2009). Tripartite purinergic modulation of central respiratory networks during perinatal development: the influence of ATP, ectonucleotidases, and ATP metabolites. *J Neurosci* **29**, 14713-14725.
- Kasymov V, Larina O, Castaldo C, Marina N, Patrushev M, Kasparov S, & Gourine AV (2013). Differential sensitivity of brainstem versus cortical astrocytes to changes in pH reveals functional regional specialization of astroglia. *J Neurosci* **33**, 435-441.
- Kim SH, Bahia PK, Patil M, Sutton S, Sowell I, Hadley SH, Kollarik M, & Taylor-Clark TE (2020). Development of a Mouse Reporter Strain for the Purinergic P2X₂ Receptor. *eNeuro* **7**.
- King BF, Wildman SS, Ziganshina LE, Pintor J, & Burnstock G (1997). Effects of extracellular pH on agonism and antagonism at a recombinant P2X₂ receptor. *Br J Pharmacol* **121**, 1445-1453.
- Lioy DT, Garg SK, Monaghan CE, Raber J, Foust KD, Kaspar BK, Hirrlinger PG, Kirchhoff F, Bissonnette JM, Ballas N, & Mandel G (2011). A role for glia in the progression of Rett's syndrome. *Nature* **475**, 497-U90.
- Llaudet E, Botting NP, Crayston JA, & Dale N (2003). A three-enzyme microelectrode sensor for detecting purine release from central nervous system. *Biosensors and Bioelectronics* **18**, 43-52.
- Llaudet E, Hatz S, Droniou M, & Dale N (2005). Microelectrode biosensor for real-time measurement of ATP in biological tissue. *Analytical Chemistry* **77**, 3267-3273.
- Loeschcke HH (1982). Central chemosensitivity and the reaction theory. *J Physiol* **332**, 1-24.
- Lorier AR, Huxtable AG, Robinson DM, Lipski J, Housley GD, & Funk GD (2007). P2Y₁ receptor modulation of the pre-Botzinger complex inspiratory rhythm generating network in vitro. *J Neurosci* **27**, 993-1005.
- Lorier AR, Lipski J, Housley GD, Greer JJ, & Funk GD (2008). ATP sensitivity of preBotzinger complex neurones in neonatal rat in vitro: mechanism underlying a P2 receptor-mediated increase in inspiratory frequency. *J Physiol* **586**, 1429-1446.
- Marina N, Ang R, Machhada A, Kasymov V, Karagiannis A, Hosford PS, Mosienko V, Teschemacher AG, Vihko P, Paton JF, Kasparov S, & Gourine AV (2015). Brainstem hypoxia contributes to the development of hypertension in the spontaneously hypertensive rat. *Hypertension* **65**, 775-783.

- Marina N, Christie IN, Korsak A, Doronin M, Brazhe A, Hosford PS, Wells JA, Sheikhabaei S, Humoud I, Paton JFR, Lythgoe MF, Semyanov A, Kasparov S, & Gourine AV (2020). Astrocytes monitor cerebral perfusion and control systemic circulation to maintain brain blood flow. *Nat Commun* **11**, 131.
- Marina N, Tang F, Figueiredo M, Mastitskaya S, Kasimov V, Mohamed-Ali V, Roloff E, Teschemacher AG, Gourine AV, & Kasparov S (2013). Purinergic signalling in the rostral ventrolateral medulla controls sympathetic drive and contributes to the progression of heart failure following myocardial infarction in rats. *Basic Res Cardiol* **108**, 317.
- Marina N, Turovsky E, Christie IN, Hosford PS, Hadjihambi A, Korsak A, Ang R, Mastitskaya S, Sheikhabaei S, Theparambil SM, & Gourine AV (2018). Brain metabolic sensing and metabolic signaling at the level of an astrocyte. *Glia* **66**, 1185-1199.
- Meigh L, Greenhalgh SA, Rodgers TL, Cann MJ, Roper DI, & Dale N (2013). CO₂ directly modulates connexin 26 by formation of carbamate bridges between subunits. *Elife* **2**, e01213.
- Mishra A, Reynolds JP, Chen Y, Gourine AV, Rusakov DA, & Attwell D (2016). Astrocytes mediate neurovascular signaling to capillary pericytes but not to arterioles. *Nat Neurosci* **19**, 1619-1627.
- Pankratov Y, Castro E, Miras-Portugal MT, & Krishtal O (1998). A purinergic component of the excitatory postsynaptic current mediated by P2X receptors in the CA1 neurons of the rat hippocampus. *Eur J Neurosci* **10**, 3898-3902.
- Pankratov Y, Lalo U, Verkhratsky A, & North RA (2006). Vesicular release of ATP at central synapses. *Pflugers Arch* **452**, 589-597.
- Pascual O, Casper KB, Kubera C, Zhang J, Revilla-Sanchez R, Sul JY, Takano H, Moss SJ, McCarthy K, & Haydon PG (2005). Astrocytic purinergic signaling coordinates synaptic networks. *Science* **310**, 113-116.
- Patterson KC, Kahanovitch U, Goncalves CM, Hablitz JJ, Staruschenko A, Mulkey DK, & Olsen ML (2021). Kir 5.1-dependent CO₂/H⁺-sensitive currents contribute to astrocyte heterogeneity across brain regions. *Glia* **69**, 310-325.
- Peng W, Wu Z, Song K, Zhang S, Li Y, & Xu M (2020). Regulation of sleep homeostasis mediator adenosine by basal forebrain glutamatergic neurons. *Science* **369**.
- Pijacka W, Moraes DJ, Ratcliffe LE, Nightingale AK, Hart EC, da Silva MP, Machado BH, McBryde FD, Abdala AP, Ford AP, & Paton JF (2016). Purinergic receptors in the carotid body as a new drug target for controlling hypertension. *Nat Med* **22**, 1151-1159.
- Rajani V, Zhang Y, Jalubula V, Rancic V, Sheikhabaei S, Zwicker JD, Pagliardini S, Dickson CT, Ballanyi K, Kasparov S, Gourine AV, & Funk GD (2018). Release of ATP by pre-Botzinger complex astrocytes contributes to the hypoxic ventilatory response via a Ca²⁺-dependent P2Y₁ receptor mechanism. *J Physiol* **596**, 3245-3269.
- Rajani V, Zhang Y, Revill AL, & Funk GD (2016). The role of P2Y₁ receptor signaling in central respiratory control. *Respir Physiol Neurobiol* **226**, 3-10.
- Ralevic V, Thomas T, Burnstock G, & Spyer KM (1999). Characterization of P2 receptors modulating neural activity in rat rostral ventrolateral medulla. *Neuroscience* **94**, 867-878.
- Reklow RJ, Alvares TS, Zhang Y, Miranda Tapia AP, Biancardi V, Katzell AK, Frangos SM, Hansen MA, Toohey AW, Cass CE, Young JD, Pagliardini S, Boison D, & Funk GD (2019). The Purinome and the preBotzinger Complex - A Menage of Unexplored Mechanisms That May Modulate/Shape the Hypoxic Ventilatory Response. *Front Cell Neurosci* **13**, 365.
- Rong W, Gourine AV, Cockayne DA, Xiang Z, Ford AP, Spyer KM, & Burnstock G (2003). Pivotal role of nucleotide P2X₂ receptor subunit of the ATP-gated ion channel mediating ventilatory responses to hypoxia. *Journal of Neuroscience* **23**, 11315-11321.
- Sheikhabaei S, Morris B, Collina J, Anjum S, Znati S, Gamarra J, Zhang R, Gourine AV, & Smith JC (2018a). Morphometric analysis of astrocytes in brainstem respiratory regions. *J Comp Neurol* **526**, 2032-2047.

Sheikhabaei S, Turovsky EA, Hosford PS, Hadjihambi A, Theparambil SM, Liu B, Marina N, Teschemacher AG, Kasparov S, Smith JC, & Gourine AV (2018b). Astrocytes modulate brainstem respiratory rhythm-generating circuits and determine exercise capacity. *Nat Commun* **9**, 370.

Sims RE & Dale N (2014). Activity-dependent adenosine release may be linked to activation of Na⁺/K⁺ ATPase: an *in vitro* rat study. *PLoS One* **9**, e87481.

Sobrinho CR, Goncalves CM, Takakura AC, Mulkey DK, & Moreira TS (2017). Fluorocitrate-mediated depolarization of astrocytes in the retrotrapezoid nucleus stimulates breathing. *J Neurophysiol* **118**, 1690-1697.

Sobrinho CR, Wenker IC, Poss EM, Takakura AC, Moreira TS, & Mulkey DK (2014). Purinergic signalling contributes to chemoreception in the retrotrapezoid nucleus but not the nucleus of the solitary tract or medullary raphe. *J Physiol* **592**, 1309-1323.

Spyer KM, Lambert JH, & Thomas T (1997). Central nervous system control of cardiovascular function: neural mechanisms and novel modulators. *Clin Exp Pharmacol Physiol* **24**, 743-747.

Spyer KM & Thomas T (2000). A role for adenosine in modulating cardio-respiratory responses: a mini-review. *Brain Res Bull* **53**, 121-124.

St Lambert JH, Thomas T, Burnstock G, & Spyer KM (1997). A source of adenosine involved in cardiovascular responses to defense area stimulation. *Am J Physiol* **272**, R195-R200.

Teppema LJ (2018). CrossTalk opposing view: the hypoxic ventilatory response does not include a central, excitatory hypoxia sensing component. *J Physiol*. **596**, 2939-2941.

Theparambil SM, Hosford PS, Ruminot I, Kopach O, Reynolds JR, Sandoval PY, Rusakov DA, Barros LF, & Gourine AV (2020). Astrocytes regulate brain extracellular pH via a neuronal activity-dependent bicarbonate shuttle. *Nat Commun* **11**, 5073.

Thomas T, Ralevic V, Bardini M, Burnstock G, & Spyer KM (2001). Evidence for the involvement of purinergic signalling in the control of respiration. *Neuroscience* **107**, 481-490.

Thomas T, Ralevic V, Gadd CA, & Spyer KM (1999). Central CO₂ chemoreception: A mechanism involving P2 purinoceptors localized in the ventrolateral medulla of the anaesthetized rat. *Journal of Physiology* **517**, 899-905.

Thomas T & Spyer KM (2000). ATP as a mediator of mammalian central CO₂ chemoreception. *J Physiol* **523 Pt 2**, 441-447.

Turovsky E, Karagiannis A, Abdala AP, & Gourine AV (2015). Impaired CO₂ sensitivity of astrocytes in a mouse model of Rett syndrome. *J Physiol* **593**, 3159-3168.

Turovsky E, Theparambil SM, Kasymov V, Deitmer JW, Del Arroyo AG, Ackland GL, Corneveaux JJ, Allen AN, Huentelman MJ, Kasparov S, Marina N, & Gourine AV (2016). Mechanisms of CO₂/H⁺ sensitivity of astrocytes. *J Neurosci* **36**, 10750-10758.

Turovsky EA, Braga A, Yu Y, Esteras N, Korsak A, Theparambil SM, Hadjihambi A, Hosford PS, Teschemacher AG, Marina N, Lythgoe MF, Haydon PG, & Gourine AV (2020). Mechanosensory signaling in astrocytes. *J Neurosci* **40**, 9364-9371.

Van de Wiel J, Meigh L, Bhandare A, Cook J, Nijjar S, Huckstepp R, & Dale N (2020). Connexin26 mediates CO₂-dependent regulation of breathing via glial cells of the medulla oblongata. *Commun Biol* **3**, 521.

Vlaskovska M, Kasakov L, Rong W, Bodin P, Bardini M, Cockayne DA, Ford AP, & Burnstock G (2001). P2X₃ knock-out mice reveal a major sensory role for urothelially released ATP. *J Neurosci* **21**, 5670-5677.

Wall MJ & Dale N (2013). Neuronal transporter and astrocytic ATP exocytosis underlie activity-dependent adenosine release in the hippocampus. *J Physiol* **591**, 3853-3871.

Wells JA, Christie IN, Hosford PS, Huckstepp RT, Angelova PR, Vihko P, Cork SC, Abramov AY, Teschemacher AG, Kasparov S, Lythgoe MF, & Gourine AV (2015). A critical role for purinergic

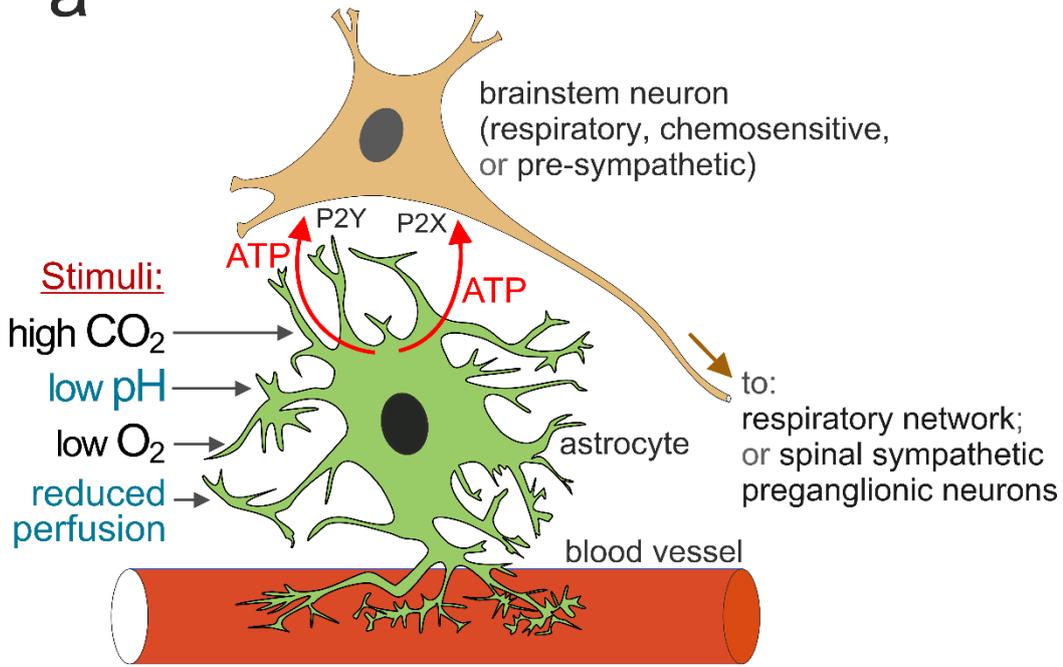
signalling in the mechanisms underlying generation of BOLD fMRI responses. *J Neurosci* **35**, 5284-5292.

Wenker IC, Kreneisz O, Nishiyama A, & Mulkey DK (2010). Astrocytes in the retrotrapezoid nucleus sense H⁺ by inhibition of a Kir4.1-Kir5.1-like current and may contribute to chemoreception by a purinergic mechanism. *J Neurophysiol* **104**, 3042-3052.

Wenker IC, Sobrinho CR, Takakura AC, Moreira TS, & Mulkey DK (2012). Regulation of ventral surface CO₂/H⁺-sensitive neurons by purinergic signalling. *J Physiol* **590**, 2137-2150.

Zwicker JD, Rajani V, Hahn LB, & Funk GD (2011). Purinergic modulation of preBotzinger complex inspiratory rhythm in rodents: the interaction between ATP and adenosine. *J Physiol* **589**, 4583-4600.

a



b

