

Cerebellar ataxia, neuropathy, vestibular areflexia syndrome: genetic and clinical insights

Roisin Sullivan^{a,*}, Rauan Kaiyrzhanov^{a,b,*}, and Henry Houlden^{a,b}

Purpose of review

This review aims to summarise the present cerebellar ataxia, neuropathy, vestibular ataxia syndrome (CANVAS) literature, providing both clinical and genetic insights that might facilitate the timely clinical and genetic diagnosis of this disease.

Recent findings

Recent advancements in the range of the clinical features of CANVAS have aided the development of a broader, more well-defined clinical diagnostic criteria. Additionally, the identification of a biallelic repeat expansion in *RFC1* as the cause of CANVAS and a common cause of late-onset ataxia has opened the door to the potential discovery of a pathogenic mechanism, which in turn, may lead to therapeutic advancements and improved patient care.

Summary

The developments in the clinical and genetic understanding of CANVAS will aid the correct and timely diagnosis of CANVAS, which continues to prove challenging within the clinic. The insights detailed within this review will raise the awareness of the phenotypic spectrum and currently known genetics. We also speculate on the future directions of research into CANVAS.

Keywords

bilateral vestibular failure, cerebellar ataxia with neuropathy and vestibular areflexia syndrome, cerebellar ataxia, neuropathy, repeat expansion, *RFC1*

INTRODUCTION

Cerebellar ataxia with neuropathy and vestibular areflexia syndrome (CANVAS) is a late-onset slowly progressive neurodegenerative ataxic disorder. There have been numerous publications defining the clinical spectrum and the pathological and genetic basis of this disease. These multidisciplinary and successional studies have expanded our understanding of CANVAS as a multisystem disease with a pattern of spatial progression and variable phenotype. Despite the accumulated knowledge, it seems that the suspicion and diagnosis of CANVAS still remain challenging in nonspecialist clinic settings and regions with no access to genetic testing. This review aims to integrate the important findings of the present CAN-VAS literature and provide clinico-genetic insights that might aid the timely diagnosis of CANVAS.

CHRONOLOGICAL OVERVIEW OF CEREBELLAR ATAXIA WITH NEUROPATHY AND VESTIBULAR AREFLEXIA SYNDROME LITERATURE

From a chronologic perspective, CANVAS literature can be divided into 3 periods. The initial period

includes papers published by neuro-otologists between 1991 and 1998, primarily addressing the role of the cerebellum in the compensatory ocular reflexes after bilateral vestibular failure (BVF) [1–3]. The following period from 2004 until 2018, when CANVAS had evolved into a distinct clinical entity with described neuropathology, proposed clinical criteria, and suspected but unknown genetic cause [4–10]. The recent period starting from 2019 discovers the common genetic cause of CANVAS and further expands our understanding of this disease

Correspondence to Roisin Sullivan, Department of Neuromuscular Disease, University College London, London, UK. E-mail: r.sullivan@ucl.ac.uk

*These authors contributed equally.

mese autions contributed equally.

^aDepartment of Neuromuscular Disease, University College London and ^bThe National Hospital for Neurology and Neurosurgery, London, UK

Curr Opin Neurol 2021, 34:556–564

DOI:10.1097/WCO.000000000000961

This is an open access article distributed under the terms of the Creative Commons Attribution-Non Commercial License 4.0 (CCBY-NC), where it is permissible to download, share, remix, transform, and buildup the work provided it is properly cited. The work cannot be used commercially without permission from the journal.

KEY POINTS

- The clinical diagnostic criteria of CANVAS has become better defined in the past few years, but correct diagnosis still proves a challenge within the clinic, highlighting a continued need to raise awareness of CANVAS to clinicians.
- The identification of a biallelic repeat expansion in *RFC1* as a common genetic cause of CANVAS gives an unparalleled advantage in the timely diagnosis of CANVAS and opens the door to the discovery of future novel genes involved in non-*RFC1* ataxia.
- The broadening phenotypic scope of RFC1-associated disease could give further clues to the pathological mechanism and pinpoint which pathways, and therefore symptoms, are caused by the *RFC1* expansion, potentially providing therapeutic targets.

[11^{••},12[•],13–15]. The early publications (1991– 2004) provided a unique contribution by going further than the highlighted association between BVF and cerebellar ataxia (CA). Despite the limited number of cases, the early reports picked up and described the currently known important features of CANVAS as peripheral neuropathy and downbeat nystagmus, albeit no association with the CA-BVF was made. Nevertheless, this has probably played a role in the definition of full CANVAS as subsequent studies deliberately sought the presence of peripheral neuropathy in larger cohorts. We anticipate that the fourth period of publications will address other genetic and mechanistic causes of CANVAS. Figure 1 depicts the available CANVAS literature in chronological order and provides a brief summary of the findings.

CLINICAL PRESENTATION

Clinically defined and recently genetically confirmed series of CANVAS has helped to delineate its phenotype over time. Currently, we know that CANVAS clinically expresses predominantly lateonset, slowly progressive, severe imbalance with absent visually enhanced vestibulo-ocular reflex (VVOR) (a hallmark sign) suggesting combined bilateral vestibular and cerebellar dysfunction in the setting of nonlength dependent sensory deficits (a diagnostic triad). CANVAS has a number of additional symptoms, the most frequent ones being an autonomic failure and chronic spasmodic cough, which could start 15–29 years before gait imbalance and can even form 'the ataxia chronic cough' phenotypic cluster [16[•],17] (Table 1). A caveat is that a subgroup of CANVAS cases could remain with an

incomplete triad for many years due to phenotypic variability [8,12[•]]. Evidence from a 100 genetically confirmed CANVAS cohort has suggested that the complete triad might be present in only 2/3 of cases and that neuronopathy is a ubiquitous sign of CAN-VAS [12[•]].

According to the genetic series, the mean onset for any neurological sign of CANVAS (cough excluded) is 52 years, albeit the onset might range between 19 and 76 years [12[•]]. Thus, young adults could also develop CANVAS symptoms. Half of CANVAS cases might manifest with a progressive motion-dependent postural imbalance, typically exacerbating in darkness and causing dizziness and/or falls. Although, the onset with isolated sensory symptoms or oscillopsia is possible [12[•]].

Each component of the CANVAS triad can cause postural imbalance independently but when combined, they result in impaired VVOR and severe imbalance leading to significant disability. All CAN-VAS symptoms have an insidious onset, particularly BVF due to simultaneous compensation mechanisms. Oculomotor testing is very important for CANVAS diagnosis as it can reveal the signs of BVF and/or cerebellar dysfunction. VVOR reflects a combined failure of all three compensatory eye movement systems, namely, vestibulo-ocular reflex (VOR), smooth pursuit, and optokinetic reflex [18]. It can be demonstrated at the bedside by turning a patient's head slowly from side to side. The test is considered positive if compensatory eye movements are saccadic rather than smooth. Video-oculography, videonystagmography, or rotational chair testing can determine the degree of VVOR impairment [9]. Studies of vestibular evoked myogenic potentials measuring saccular and utricular function showed inconsistent results in CANVAS [19,20].

CA in CANVAS typically manifests with cerebellar-type ocular motor impairment including downbeat nystagmus, broken pursuit, gaze-evoked nystagmus, and saccadic dysmetria [4,7,8,18, 21,22]. Appendicular incoordination, truncal ataxia, and cerebellar-type dysarthria can precipitate typically 2–6 years after the onset of gait impairment [12"]. A pattern of spatial progression from sensory neuronal involvement early on, to later vestibular and cerebellar dysfunction has been suggested [12"].

Neuropathy in CANVAS clinically presents with a multimodal nonlength-dependant sensory deficit, frequently associated with normo- or hyperreflexia. Electrophysiology studies reveal severe axonal sensory neuropathy with a reduced or absent sensory nerve action potentials (SNAP) in upper- and lower limbs (neuronopathy), relatively intact motor conduction and F-wave latencies but reduced H-reflexes.

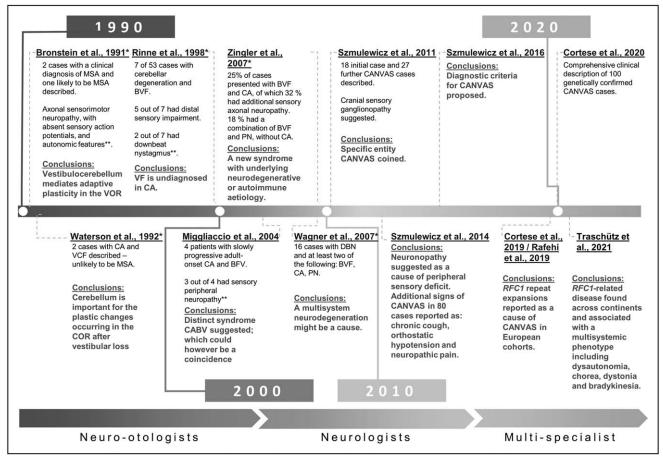


FIGURE 1. Timeline of CANVAS literature, from initial reports through to clinical diagnostic criteria and genetic causation. BVF, bilateral vestibular failure; CA, cerebellar ataxia; CABV, cerebellar ataxia with bilateral vestibulopathy; CA-BVF, cerebellar ataxia with bilateral vestibular atellaxia vestibular failure; CANVAS, cerebellar ataxia, neuronopathy, vestibular areflexia syndrome; COR, cervico-ocular reflex; DBN, downbeat nystagmus; MSA, Multiple systems atrophy; PN, peripheral neuropathy; RFC1, replication factor C subunit 1; VOR, vestibular ocular reflex. *Study objectives: Bronstein *et al.* 1991: To address which structures and/or mechanisms help to restore gaze stability after BFV; Waterson *et al.* 1992: To explore the mechanism that compensate for the loss of the VOR in patients with isolated bilateral BVF; Rinne at al., 1998: A retrospective review of the clinical presentations and aetiologies of 53 cases of BFV; Zingler *et al.*, 2007: To retrospectively determine causative factors and epidemiology of BVF in 255 cases; Wagner *et al.*, 2007: To review the aetiology and comorbidity of DBN in 117 patients. **Findings compatible with CANVAS.

Electromyography is typically normal [6,9,12[•]]. Normoreflexia is explained by the sparing of muscle spindle afferents (Ia fibers) [23,24]. Peripheral nerve ultrasound studies showed that nerves of CANVAS patients are significantly smaller than their age- and gender-matched controls, which is consistent with neuronopathy [25,26].

Brain MRI data has suggested a pattern of cerebellar atrophy, corresponding to the neuropathological findings. This includes predominant hemispheric atrophy in the posterosuperior and horizontal fissures, delimiting Crus 1, and associated with this anterior and dorsal vermial atrophy (lobules VI, VIIa, and VIIB) [8,18]. Mild brain stem atrophy and posterior cord atrophy are also seen on brain and spinal cord MRI respectively [12[•]]. Despite the proposed clinical criteria, even full CANVAS seems to escape clinical suspicion and/or diagnosis, exemplified by only 23 out of 66 genetically confirmed patients with full CANVAS having received the initial diagnosis of CANVAS [12[•]]. This might suggest that further work needs to be done to increase the awareness of CANVAS among medical professionals.

NEUROPATHOLOGY OF CEREBELLAR ATAXIA WITH NEUROPATHY AND VESTIBULAR AREFLEXIA SYNDROME

Oto-and neuropathology studies of CANVAS have suggested that widespread ganglionopathy (neuronopathy) is a hallmark of CANVAS that underlies

Table 1. CANVAS/RFC1 CANV	AS associated features
Symptoms	Frequency (references)
Downbeat nystagmus	65% [21]
Spasmodic cough	64% [13]
Normoreflexia	50% [13,24]
Neuropathic pain	42/100 [13]
Oscillopsia	32% [13]
Dysphagia	27% [13]
Autonomic features	32% (50% of those undergoing specific investigations), 91%, 44%. [5,21,16 [•]]
Cold and pale feet	78% [21]
Light-headedness	65% [21]
Xerostomia/xerophthalmia	52% [21]
Postural instability with retropulsion	49% [16"]
REM sleep behaviour disorder	40–70% [14]
Gastroesophageal reflux disease	31% [16"]
Erectile dysfunction	7%,78%, 44% [13,16",21]
Persistent constipation	7%, 65%, 24% [13,16•,21]
Reduced feet sweating	48% [21]
Mild cognitive impairment	25% [16"]
Postural hypotension	13-30% [13,21,16"]
Urinary leakage	8%, 39% [13,21]
Bradikinesia	28% [16"]
Nausea, vomiting, and bloating after small meal	26% [21]
Slowing of vertical saccades	17% [16"]
Hyperkinetic movement disorders	5% [16 "]
Sudomotor dysfunction	[17]
Neurotrophic Keratopathy	[21]

CANVAS, cerebellar ataxia, neuronopathy, vestibular areflexia syndrome; REM, rapid eye movement.

peripheral sensory deficit, vestibular areflexia, and dysautonomia [7,27]. This was shown by sub-total neuronal loss of the sensory nerve ganglia of the dorsal root of the spinal cord with resultant posterior column atrophy and associated axonal neuropathy. The anterior and lateral horns of spinal cord appeared normal. Additionally, a multiple cranial nerve neuronopathy affecting the vestibular, facial, and trigeminal nerves has been demonstrated [27]. Selective atrophy of the vestibular nerves and marked loss in Scarpa's ganglion cells with intact auditory nerve and the spiral (cochlear) ganglion were also observed. The normal population of hair cells and supporting cells were demonstrated in the cristae and maculae [27]. The most significant cerebellar pathology was a neuronal loss in the Purkinje cells with the preferential atrophy of the anterior and dorsal vermis (lobules VI, VIIa, and VIIb) and hemispheric crus I. The cerebellar dentate nuclei,

basal ganglia, and diencephalon appeared intact [7]. Currently available oto-and neuropathology studies derive from four cases with a clinically based and one case of a genetically confirmed diagnosis of full CANVAS.

CEREBELLAR ATAXIA WITH NEUROPATHY AND VESTIBULAR AREFLEXIA SYNDROME LOOK-ALIKES AND CLUES TO THE DIAGNOSIS OF CEREBELLAR ATAXIA WITH NEUROPATHY AND VESTIBULAR AREFLEXIA SYNDROME

Progressive ataxia resulting from variably combined impairments in proprioceptive, vestibular, and cerebellar functions is not specific to CANVAS. The diagnostic workup should start by excluding curable disorders with underlying nutritional, immune-mediated, and paraneoplastic causes. Afterward, the main differential diagnosis should include multisystem atrophy cerebellar variant (MCA-C) and genetic disorders such as Friedreich ataxia, SCA3, SCA1, SCA6, and the nuclear POLG1 gene. More details on the suggested differential diagnosis are given in Table 2. With the discovery of the genetics cause of CANVAS it is plausible to test cases suggestive of CANVAS for *RFC1* expansions first, if genetic testing is possible, then if negative to investigate for other genetic causes.

CANVAS can usually be diagnosed clinically. However, the diagnosis may be more challenging in those cases where a significant delay in the manifestation of all three cardinal features exists, e.g., CANVAS in evolution, and the onset age of the first signs is < 60 years, particularly in sporadic patients. The final component of the diagnostic triad may take more than 10 years [8]. Ataxia is usually an overt sign of CANVAS, whereas neuropathy and BVF could be subclinical requiring their deliberate and regular search using specialised tests. There are certain clues to the diagnosis of CANVAS that should prompt a purposeful search for other components of the diagnostic triad (Fig. 2A). Invaluable help in the clinical diagnosis of CANVAS comes from a description of large genetically confirmed CANVAS cases and the proposed clinical diagnostic criteria. Considering a range of CANVAS-associated features that are not reflected in the current diagnostic criteria, might also be important in the clinical diagnosis.

GENETICS OF CEREBELLAR ATAXIA WITH NEUROPATHY AND VESTIBULAR AREFLEXIA SYNDROME: *RFC1* EXPANSION

CANVAS has largely been thought of as a sporadic disease; however, affected sibships suggest a genetic

			Genetic disorders	orders			Oth	Other conditions
	CANVAS	FRDA	SCA3	SCA1	SCA6	ANS-POLG disorders	MSA	Wernicke's encephalopathy
Gene	RFC 1	FXN	ATXN3	ATXN1	CACNAIA	POIG	NA	AA
Typical age of ons	Typical age of onset Late adulthood (mean 52yrs)	< 25 yrs Rarely late onset	30-40yrs	30–40yrs	43 -52 years	2nd-4th decade	Pick onset 55–60 (range 30–90)	Variable
Progression	Slow	Slow	Could be rapid depending on the number of CAG repeats	Could be rapid depending on the number of CAG repeats	Slow	Slow	Typically rapid	Subacute anset
Cerebellar ataxia	Progressive	Progressive	Progressive	Progressive	Progressive	Progressive	Progressive	Yes Part of the classic triad
Neuropathy	Very frequent, Progressive axonal sensory neuronopathy, Severe	Frequent, Progressive sensory neuronopathy	Frequent, mixed motor and sensory	Frequent, Mixed sensory-motor	Rare, axonal sensary-mator	Sensory, motor, or mixed. Severe	°Z	Sensory or motor
BVF	Progressive, typically severe	Mild	Yes, Mild to moderate	Yes, Mild	Yes, Mild	oZ	Very rare	Yes
Chronic cough	Frequent	No	No	No	No	No	No	No
Dysautonomia	Postganglianic AF, Frequent, Mild, Late presentation	Rare, Late presentation, mild	Frequent (67–70%), Mild to moderate	Frequent (22– 100%), Mild to moderate	Ŷ	Very rare	Preganglionic AF Frequent Severe Progressive, early presentation	Yes
DBN	Frequent	Rare	No	No	Frequent	No	Rare	Yes
Brain MRI features	Cerebellar atrophy involving anterior and dorsal vermis, as well as the hemispheric crus	Significant grey and white matter loss in the deep cerebellar nuclei and brainstem, significant atrophy in the superior cerebellar peduncles	Cerebellar, cerebral cortical, and pontine atrophy, 'Hot cross bun sign'	Olivopontocerebellar atrophy (OPCA) and white matter (WM) atrophy. 'Hot cross bun sign'	Pure cerebellar atrophy, 'Hot cross bun sign'	Atrophy or abnormal signal intensity in the cerebellum brainstem, and other brain structures	Cerebellar, brainstem and pontine atrophy, putaminal rim sign, 'Hot cross bun sign'	T2/FLAIR: symmetrically increased signal intensity in the mammillary bodies, dorsomedial thalami, tectal plate, periaqueductal grey matter, around the third ventricle.
Other distinguishing features	Impaired VVOR Normoreflexia	Frequent muscle weakness, pyramidal involvement (Babinski signs) & skeletal deformities (pes cavus, scoliosis) Assoc cardionyopathy & diabetes Vision & hearing loss	Pyramidal and extrapyramidal signs, Amyotrophy fasciculations	Pyramidal signs, cognitive decline	Slow progression	Encephalopathy with seizures Ophthalmoplegia Multisystem involvement	Parkinsonism, RBD (frequent)	Acute confusion, impaired vision and hearing, psychosis

Movement disorders

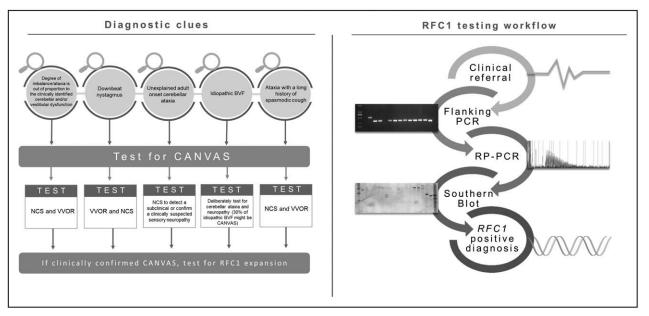


FIGURE 2. Diagnostic clues and RFC1 testing workflow for clinical and molecular diagnosis of CANVAS. 2A presents diagnostic clues suggestive of clinical CANVAS and associated tests for clinical confirmation. 2B presents the testing workflow for molecularly diagnosing RFC1-caused CANVAS. BVF, bilateral vestibular areflexia; CANVAS, cerebellar ataxia, neuronopathy, vestibular areflexia syndrome; NCS, nerve conduction study; PCR, polymerase-chain reaction; RFC1, replication factor C subunit 1; RPPCR, repeat-primed PCR; VVOR, visually enhanced vestibulo-ocular reflex.

component [11^{••}]. Recently a biallelic intronic expansion in *RFC1* has been identified as a cause of sporadic late-onset ataxia and CANVAS, with a core ancestral haplotype that most likely arose 25,000 years ago in Europe [11^{••},13].

The biallelic *RFC1* expansion has been confirmed in a range of populations and ethnicities. The pentanucleotide AAGGG_(n) carrier frequency within the normal population ranges between 0.7 and 6.8% [11^{••},13,28–30]. Several studies have identified the recessive RFC1 expansion in 90% of patients with full CANVAS, 20–22% of patients with late-onset ataxia and/or peripheral neuropathy, around 60% with cerebellar involvement and sensory neuropathy and 3.2% with undiagnosed CA [5,13,21]. Non-European, unselected late-onset ataxia cohorts of Turkish, West-Asian, and Indonesian ethnicity were found to have an RFC1 frequency rate of 14%, showing that RFC1related disease is extending globally, which could also be influenced by higher consanguinity rates [16]. Contrastingly, several studies have reported lower or absent rates of the *RFC1* repeat expansion in certain populations including Canadian and Brazilian lateonset ataxia, Japanese familial and sporadic late-onset ataxia, and Chinese Han ataxia cohorts [28,29,31].

REPEAT MOTIFS

The *RFC1* locus is highly polymorphic, spanning the pathogenic conformation $AAGGG_{(n)}$, the reference

 $AAAAG_{(11)}$, expanded $AAAAG_{(n)}$ and $AAAGG_{(n)}$. The allelic distribution of these has been identified as 75.5%, 13.0%, and 7.9%, respectively, within the control population [11**]. Three percent were undetermined and most likely correspond to novel conformations that include $AAGAG_{(n)}$ and $AGAGG_{(n)}$, that have, as yet, uncertainty concerning their possible pathogenicity [28]. Two novel pathogenic conformations were identified in Asian Pacific cohorts. A likely pathogenic recessive $ACAGG_{(n)}$ motif was identified in Asian Pacific and Japanese cohorts [15,31]. An alternate pathogenic allele configuration (AAAGG)₁₀₋₂₅ AAGGG_(n) AAAGG_(n) was found to be specific to the Māori population, that had no phenotypic differences to European cohorts [14].

The various motif conformations follow a pattern in expansion size; with pathogenic $AAGGG_{(n)}$ ranging from 400 to 2000 repeating units, $AAAGG_{(n)}$ from 40 to 1000 and $AAAAG_{(n)}$ with 15 to 200 repeats [11^{••}]. There is a positive correlation between *RFC1* expansion size and GC content, with each listed motif containing 20%, 40%, and 60% GC content, respectively [15]. There has been no correlation observed between the size of expansion and age of onset, symptom onset (cough excluded) and severity of disease [12[•]]. There are likely to be further novel repeat motifs that have evaded detection due to high GC content and consequential low coverage, exemplified by the historic challenge of sequencing the GC-rich GGGGCC repeat expansion in C9*orf*72 and discovery of the *VWA1* repeat expansion in recessive hereditary motor neuropathy; both of which have GC content implicated in pathogenicity [32,33]. GC content has been shown to increase mutation rates, with an altered yeast gene with 63% GC content showing a > 6-fold increased mutation rate [34].

RFC1 PATHOLOGICAL MECHANISM

The pathological mechanism underlying the biallelic repeat expansion in RFC1 is currently unknown. RFC1 is a large subunit of the RFC complex that loads and unloads proliferating cell nuclear antigen onto DNA [35,36]. The biallelic expansion in *RFC1* is located at the 3'-end of a deep intronic AluSx3 element. Alu elements undergo insertions into new genomic locations and contribute to genetic instability, potentially contributing to age-related degeneration [37,38]. The polyA tail at the 3' end of the Alu element is theorized to be a microsatellite expansion initiation site and undergoes A-tail shortening which has resulted in the majority being inactive [37,39]. The shared core haplotype and highly polymorphic polyA tail supports evidence of an original genetic event predicted to have occurred during the Alu element polyA-tail shortening and degradation process; with AAAAG changing to AAAGG or AAAGG in a more recent common ancestor 25,000 years ago [13].

The *RFC1* expansion does not cause an associated loss of function of the *RFC1* protein and RNAseq did not implicate any bordering or distant genomic regions [11^{••}]. *RFC1* has been associated with DNA damage recognition, DNA repair enzyme interaction, DNA enzyme recruitment and pathways that include nucleotide excision repair [40]. Ataxia and neuropathy are features of many neurodegenerative diseases suggesting that the peripheral nerves and cerebellum are particularly vulnerable to DNA damage [41]. However, response to DNA repair was found to be normal within fibroblast lines with the biallelic *RFC1* repeat expansion [11^{••}].

Repeat expansions within transcripts are implicated in altered splicing machinery; RNA foci and RNA toxicity are features of myotonic dystrophy type 1 (DM1) and 2 (DM2) [42]. Repeat-associated non-ATG (RAN) proteins are also implicated in diseases such as C9*orf*72 Amyotrophic lateral sclerosis (ALS)/frontotemporal dementia (FTD), with their accumulation affecting downstream pathways [43]. However, CANVAS brain tissue with *RFC1* biallelic expansion showed no sense or antisense RNA foci. Similarly to other diseases with GC rich intronic expansions such as C9orfALS/FTD and DM2, intronic retention has been observed across all tissues in CANVAS patients, affecting intron 2 in *RFC1* premRNA [11^{••},44]. This finding could be key to elucidating the pathogenic mechanism underpinning the *RFC1* repeat expansion as intronic retention, and therefore impaired pre-mRNA processing, can affect nuclear retention and nucleocytoplasmic transport of pre-mRNA.

MOLECULAR DIAGNOSIS OF RFC1

Molecular diagnosis of CANVAS consists of a combination of flanking polymerase-chain reaction (PCR), repeat-primed PCR (RPPCR) and Southern blotting to confirm the presence of the biallelic RFC1 expansion (Fig. 2B). Patients positive for the biallelic repeat expansion will show no amplifiable product on flanking PCR, a positive trace on RPPCR (which resembles a hallmark sawtooth decremental peak pattern), and distinct bands on Southern blotting corresponding to the expanded alleles. Longread sequencing has also been shown to precisely identify the biallelic *RFC1* repeat expansion and has the potential to replace Southern blotting as the gold-standard in genetic diagnosis [45]. Despite the intronic location of the RFC1 repeat expansion, the AAGGG motif has also been identified using whole-genome- and exome-sequencing (WGS/ WES). If successfully validated it could prove a useful screening tool for existing WGS and WES datasets in the identification of currently undiagnosed RFC1 positive patients [34].

PHENOTYPIC SPECTRUM OF RFC1 EXPANSION

The phenotypic spectrum implicated with the *RFC1* expansion is broadening. Whilst the biallelic RFC1 repeat expansion has been found to be absent in pathologically confirmed MSA, positive cases have been reported with clinically confirmed MSA [29,46,47]. The biallelic *RFC1* expansion has been associated with unspecified, late-onset ataxia, doparesponsive parkinsonism, hereditary sensory neuropathy with cough and Sjögren syndrome and [48–50]. Furthermore, a resent series of *RFC1* CAN-VAS has suggested that *RFC1*-disease presents with multisystemic phenotypes overlapping with MSA-C, progressive supranuclear palsy, and hyperkinetic movement disorders [16[•]]. Further validation on these RFC1-related disorders is required to confirm causal RFC1-associated pathogenicity, but it suggests a broadening phenotypic scope.

CONCLUSION: FUTURE PERSPECTIVES IN CEREBELLAR ATAXIA WITH NEUROPATHY AND VESTIBULAR AREFLEXIA SYNDROME RESEARCH

BVF involves circumscribed atrophy of the hippocampus with impairments of spatial memory and navigation [15] which would be interesting to investigate in CANVAS patients. Although sensory neuropathy is established to be a prominent feature of the disease, whether it is the site of initial disease manifestation requires further confirmation [12[•]]. We anticipate that future work will center on identifying the mechanism underpinning the RFC1 repeat expansion. Although RFC1 pathology has been previously detailed on one CANVAS brain, further validation will be required [11^{•••}]. Additionally, no neuropathology has yet been reported on other *RFC1*-related disorders, which will be pivotal in understanding which phenotypes are directly caused by the RFC1 expansion. Finally, with the advancement of next-generation sequencing technology, we expect that novel genetic causes of non-*RFC1* CANVAS may soon come to light, furthering mechanistic understanding and treatment options for this disease.

Acknowledgements

We would like to thank all of our collaborators, both within the UK and internationally. We also would like to thank the Medical Research Council (MRC) who fund R.S and University College London Overseas Research Scholarships and Graduate Research Scholarships who fund R.K. We would also like to thank Ataxia UK, the Rosetrees Trust, The Wellcome Trust, the Multiple Systems Atrophy Trust, NIH Rare Diseases Clinical Research Network and Inherited Neuropathy Consortium who support our work.

Financial support and sponsorship

None.

Conflicts of interest

There are no conflicts of interest.

REFERENCES AND RECOMMENDED READING

Papers of particular interest, published within the annual period of review, have been highlighted as:

- of special interestof outstanding interest
- Bronstein AM, Mossman S, Luxon LM. The neck-eye reflex in patients with reduced vestibular and optokinetic function. Brain 1991: 114:1-11.
- Waterston JA, Barnes GR, Grealy MA, Luxon LM. Coordination of eye and head movements during smooth pursuit in patients with vestibular failure. J Neurol Neurosurg Psychiatry 1992; 55:1125–1131.
- **3.** Rinne T, Bronstein AM, Rudge P, *et al.* Bilateral loss of vestibular function: clinical findings in 53 patients. J Neurol 1998; 245:314–321.

- Migliaccio AA, Halmagyi GM, McGarvie LA, Cremer PD. Cerebellar ataxia with bilateral vestibulopathy: description of a syndrome and its characteristic clinical sign. Brain 2004; 127:280–293.
- Zingler VC, Cnyrim C, Jahn K, et al. Causative factors and epidemiology of bilateral vestibulopathy in 255 patients. Ann Neurol 2007; 61:524–532.
- Szmulewicz DJ, Waterston JA, Halmagyi GM, *et al.* Sensory neuropathy as part of the cerebellar ataxia neuropathy vestibular areflexia syndrome. Neurology 2011; 76:1903–1910.
- Szmulewicz DJ, McLean CA, Rodriguez ML, et al. Dorsal root ganglionopathy is responsible for the sensory impairment in CANVAS. Neurology 2014; 82:1410-1415.
- Szmulewicz DJ, McLean CA, MacDougall HG, et al. CANVAS an update: clinical presentation, investigation and management. J Vestib Res 2014; 24:465-474.
- Szmulewicz DJ, Seiderer L, Halmagyi GM, et al. Neurophysiological evidence for generalized sensory neuronopathy in cerebellar ataxia with neuropathy and bilateral vestibular areflexia syndrome. Muscle Nerve 2015; 51:600-603.
- Wagner JN, Glaser M, Brandt T, Strupp M. Downbeat nystagmus: aetiology and comorbidity in 117 patients. J Neurol Neurosurg Psychiatry 2008; 79:672–677.
- 11. Cortese A, Simone R, Sullivan R, et al. Biallelic expansion of an intronic repeat
- In RFC1 is a common cause of late-onset ataxia. Nat Genet 2019; 51:649-658.

This is the first study to identify the biallelic intronic repeat expansion in RFC1 as a cause of CANVAS and late-onset ataxia and highlights the emergence of a potential pathological pathway.

 12. Cortese A, Tozza S, Yau WY, *et al.* Cerebellar ataxia, neuropathy, vestibular areflexia syndrome due to RFC1 repeat expansion. Brain 2020; 143:480-490.

This is a comprehensive study into the clinical phenotype associated with RFC1associated CANVAS and highlights the wide spectrum of phenotype linked to the disease.

- Rafehi H, Szmulewicz DJ, Bennett MF, et al. Bioinformatics-based identification of expanded repeats: a nonreference intronic pentamer expansion in RFC1 Causes CANVAS. Am J Hum Genet 2019; 105:151–165.
- Beecroft SJ, Cortese A, Sullivan R, *et al.* A Maori specific RFC1 pathogenic repeat configuration in CANVAS, likely due to a founder allele. Brain 2020; 143:2673-2680.
- Scriba CK, Beecroft SJ, Clayton JS, et al. A novel RFC1 repeat motif (ACAGG) in two Asia-Pacific CANVAS families. Brain 2020; 143:2904-2910.
- 16. Traschutz A, Cortese A, Reich S, *et al.* Natural history, phenotypic spectrum,
 and discriminative features of multisystemic rfc1-disease. Neurology 2021; 96:e1369-e1382.

This is a comprehensive study into RFC1-associated disease, highlighting additional phenotypes and demonstrating the first report of the use of whole-exome sequencing to successfully identify the RFC1 repeat expansion.

- Umeh CC, Polydefkis M, Chaudhry V, Zee DS. Sweat gland denervation in cerebellar ataxia with neuropathy and vestibular areflexia syndrome (CAN-VAS). Mov Disord Clin Pract 2017; 4:46–48.
- Szmulewicz DJ, Waterston JA, MacDougall HG, et al. Cerebellar ataxia, neuropathy, vestibular areflexia syndrome (CANVAS): a review of the clinical features and video-oculographic diagnosis. Ann NY Acad Sci 2011; 1233:139–147.
- Rust H, Peters N, Allum JHJ, et al. VEMPs in a patient with cerebellar ataxia, neuropathy and vestibular areflexia (CANVAS). J Neurol Sci 2017; 378:9-11.
- Moreno-Ajona D, Alvarez-Gomez L, Manrique-Huarte R, et al. VEMPs and dysautonomia assessment in definite cerebellar ataxia, neuropathy, vestibular areflexia syndrome (CANVAS): a Case Series Study. Cerebellum 2019.
- Wu TY, Taylor JM, Kilfoyle DH, *et al.* Autonomic dysfunction is a major feature of cerebellar ataxia, neuropathy, vestibular areflexia 'CANVAS' syndrome. Brain 2014; 137:2649-2656.
- Paley GL, Kung NH, Bucelli RC, et al. Cerebellar ataxia with neuropathy and vestibular areflexia syndrome presenting with neurotrophic keratopathy. J Neuroophthalmol 2018; 38:342–343.
- 23. Infante J, Garcia A, Serrano-Cardenas KM, et al. Cerebellar ataxia, neuropathy, vestibular areflexia syndrome (CANVAS) with chronic cough and preserved muscle stretch reflexes: evidence for selective sparing of afferent la fibres. J Neurol 2018; 265:1454–1462.
- Burke D, Halmagyi GM. Normal tendon reflexes despite absent sensory nerve action potentials in CANVAS: a neurophysiological study. J Neurol Sci 2018; 387:75–79.
- Pelosi L, Leadbetter R, Mulroy E, *et al.* Peripheral nerve ultrasound in cerebellar ataxia neuropathy vestibular areflexia syndrome (CANVAS). Muscle Nerve 2017; 56:160–162.
- Pelosi L, Mulroy E, Leadbetter R, *et al.* Peripheral nerves are pathologically small in cerebellar ataxia neuropathy vestibular areflexia syndrome: a controlled ultrasound study. Eur J Neurol 2018; 25:659–665.
- Szmulewicz DJ, Merchant SN, Halmagyi GM. Cerebellar ataxia with neuropathy and bilateral vestibular areflexia syndrome: a histopathologic case report. Otol Neurotol 2011; 32:e63-e65.

- Akcimen F, Ross JP, Bourassa CV, *et al.* Investigation of the RFC1 repeat expansion in a Canadian and a Brazilian ataxia cohort: identification of novel conformations. Front Genet 2019; 10:1219.
- 29. Fan Y, Zhang S, Yang J, et al. No biallelic intronic AAGGG repeat expansion in RFC1 was found in patients with late-onset ataxia and MSA. Parkinsonism Relat Disord 2020; 73:1–2.
- Aboud Syriani D, Wong D, Andani S, et al. Prevalence of RFC1-mediated spinocerebellar ataxia in a North American ataxia cohort. Neurol Genet 2020; 6:e440.
- Tsuchiya M, Nan H, Koh K, et al. RFC1 repeat expansion in Japanese patients with late-onset cerebellar ataxia. J Hum Genet 2020; 65:1143–1147.
- Pagnamenta AT, Kaiyrzhanov R, Zou Y, et al. An ancestral 10-bp repeat expansion in VWA1 causes recessive hereditary motor neuropathy. Brain 2021; 144:584-600.
- Ebbert MTW, Farrugia SL, Sens JP, et al. Long-read sequencing across the C9orf72 'GGGGCC' repeat expansion: implications for clinical use and genetic discovery efforts in human disease. Mol Neurodegener 2018; 13:46.
- Kiktev DA, Sheng Z, Lobachev KS, Petes TD. GC content elevates mutation and recombination rates in the yeast Saccharomyces cerevisiae. Proc Natl Acad Sci USA 2018; 115:E7109–E7118.
- Cai J, Uhlmann F, Gibbs E, *et al.* Reconstitution of human replication factor C from its five subunits in baculovirus-infected insect cells. Proc Natl Acad Sci USA 1996; 93:12896–12901.
- 36. Yao N, Turner J, Kelman Z, et al. Clamp loading, unloading and intrinsic stability of the PCNA, beta and gp45 sliding clamps of human, E. coli and T4 replicases. Genes Cells 1996; 1:101–113.
- 37. Deininger P. Alu elements: know the SINEs. Genome Biol 2011; 12:236.
- 38. Rogers J. Retroposons defined. Nature 1983; 301:460.
- Arcot SS, Wang Z, Weber JL, et al. Alu repeats: a source for the genesis of primate microsatellites. Genomics 1995; 29:136–144.

- Overmeer RM, Gourdin AM, Giglia-Mari A, et al. Replication factor C recruits DNA polymerase delta to sites of nucleotide excision repair but is not required for PCNA recruitment. Mol Cell Biol 2010; 30:4828–4839.
- McKinnon PJ. Maintaining genome stability in the nervous system. Nat Neurosci 2013; 16:1523-1529.
- Philips AV, Timchenko LT, Cooper TA. Disruption of splicing regulated by a CUG-binding protein in myotonic dystrophy. Science 1998; 280:737-741.
- Cleary JD, Pattamatta A, Ranum LPW. Repeat-associated non-ATG (RAN) translation. J Biol Chem 2018; 293:16127–16141.
- 44. Sznajder LJ, Thomas JD, Carrell EM, et al. Intron retention induced by microsatellite expansions as a disease biomarker. Proc Natl Acad Sci U S A 2018; 115:4234-4239.
- 45. Nakamura H, Doi H, Mitsuhashi S, et al. Long-read sequencing identifies the pathogenic nucleotide repeat expansion in RFC1 in a Japanese case of CANVAS. J Hum Genet 2020; 65:475–480.
- Sullivan R, Yau WY, Chelban V, et al. RFC1 intronic repeat expansions absent in pathologically confirmed multiple systems atrophy. Mov Disord 2020; 35:1277-1279.
- Sullivan R, Yau WY, Chelban V, et al. RFC1-related ataxia is a mimic of early multiple system atrophy. J Neurol Neurosurg Psychiatry 2021.
- da Silva Schmitt G, Martinez ARM, da Graca FF, et al. Dopa-responsive Parkinsonism in a patient with homozygous RFC1 expansions. Mov Disord 2020; 35:1889-1890.
- Gisatulin M, Dobricic V, Zuhlke C, et al. Clinical spectrum of the pentanucleotide repeat expansion in the RFC1 gene in ataxia syndromes. Neurology 2020; 95:e2912-e2923.
- Kumar KR, Cortese A, Tomlinson SE, et al. RFC1 expansions can mimic hereditary sensory neuropathy with cough and Sjogren syndrome. Brain 2020; 143:e82.