



Since January 2020 Elsevier has created a COVID-19 resource centre with free information in English and Mandarin on the novel coronavirus COVID-19. The COVID-19 resource centre is hosted on Elsevier Connect, the company's public news and information website.

Elsevier hereby grants permission to make all its COVID-19-related research that is available on the COVID-19 resource centre - including this research content - immediately available in PubMed Central and other publicly funded repositories, such as the WHO COVID database with rights for unrestricted research re-use and analyses in any form or by any means with acknowledgement of the original source. These permissions are granted for free by Elsevier for as long as the COVID-19 resource centre remains active.

## Failure to seroconvert after two doses of BNT162b2 SARS-CoV-2 vaccine in a patient with uncontrolled HIV

SARS-CoV-2 infection elicits similar antibody responses in HIV-negative individuals and people living with HIV that is well controlled by antiretroviral therapy (ART).<sup>1</sup> SARS-CoV-2 vaccines, including the mRNA vaccine BNT162b2, are efficacious in clinical trials, inducing antibodies and T cells specific to the spike protein.<sup>2-4</sup> However, vaccine responses in people living with HIV have been assessed only in those stable on ART.<sup>5</sup> Immune function in people with HIV and uncontrolled HIV replication is impaired due to destruction of the CD4 T cells that help B cells during an antibody response. Immune responses to vaccines (eg, against influenza and hepatitis B) in people living with HIV are inferior to those in the general population.<sup>6</sup> We present a case of one individual with uncontrolled HIV replication who did not respond to two doses of the BNT162b2 SARS-CoV-2 vaccine.

The index patient, with advanced HIV, was recruited to a cohort study, along with 13 people living with HIV suppressed on ART (median CD4 count 590 cells per  $\mu\text{L}$  [SD 202; range 310–940]) and 43 HIV-negative controls, all of whom had received one or two doses of BNT162b2.<sup>1</sup> The index patient had no history of SARS-CoV-2 infection or AIDS-defining conditions and had already received two doses of BNT162b2 24 days apart. Blood samples were obtained 16 days and 44 days after the second dose (appendix p 2). At day 16, the patient's HIV viral load was 831 764 copies per mL and CD4 count was 20 cells per  $\mu\text{L}$  (CD4% 4.6%; CD4/CD8 0.05). ART with bicitgravir, emtricitabine, and tenofovir alafenamide and prophylaxis for opportunistic infections were initiated at day 16. At day 44 (29 days

after ART initiation), HIV viral load was undetectable and CD4 count was 70 cells per  $\mu\text{L}$ .

Post-vaccine samples from the index patient showed no IgG reactivity against the S1 subunit of the spike protein by an in-house ELISA (appendix pp 1–2).<sup>7</sup> By contrast, an HIV-negative, SARS-CoV-2-naive participant had an S1-specific IgG titre of 43.4  $\mu\text{g}/\text{mL}$  44 days after the second dose of BNT162b2, consistent with the binding titres across the wider cohort, in which all participants produced S1-specific IgG, even after only one dose (appendix pp 1–2). No SARS-CoV-2-specific neutralisation was observed at either timepoint for the index patient. By contrast, the HIV-negative, SARS-CoV-2-naive vaccine recipient had a neutralisation titre of 1/656 after the second dose. No quantifiable spike protein-specific T cells, evaluated via ELISpot (MAIPN4550; Merck Millipore, Darmstadt, Germany), were observed in the index patient compared with the HIV-negative control participant, sampled 44 days after the second dose, and a subsample of people living with well controlled, stable HIV after one dose of BNT162b2 (appendix p 2). Although no spike protein-specific T cells were detected via intracellular cytokine staining in the index patient, responses against cytomegalovirus pp65 were dominated by CD8 T cells, strikingly so after ART (appendix p 2). The profound peripheral immune cell perturbations in the index patient were accompanied by increased frequencies of CD8 T cells with a terminally differentiated effector memory phenotype (CCR-7<sup>-</sup> and CD45RA<sup>+</sup>) and an inverted CD4/CD8 ratio, which could influence the size of T-cell responses to SARS-CoV-2.<sup>1</sup>

In conclusion, an individual with profound HIV-related immune dysfunction did not seroconvert to a SARS-CoV-2 vaccine. We suggest monitoring SARS-CoV-2 seroconversion in people with advanced HIV and considering repeat vaccination upon HIV suppression and CD4 count improvement with

ART. This case highlights an urgent need to establish correlates of vaccine efficacy in people living with HIV, particularly in those with suboptimal viral suppression or ongoing perturbed immune function, to better inform clinical management and guidelines.

LEM has received honorarium for lectures from the London School of Hygiene and Tropical Medicine. SR-J has received honorarium for lectures from the London School of Hygiene and Tropical Medicine, declares additional grant support from UK Research and Innovation, EDCTP, Kumamoto University, the Rosetrees Trust, and the Global Challenges Research Fund, and is a trustee and past president of the Royal Society of Tropical Medicine and Hygiene. RKG has received consulting fees from UMOVIS Lab and Janssen. FB has received payment for teaching research methods from Gilead Sciences. LW has received honorarium for participation in advisory boards, speaker fees, or both from Gilead Sciences, ViiV, Janssen, MSD, Theratechnologies, Mylan, and Cipla, and has also received conference attendance support from ViiV. All other authors declare no competing interests. ET and AA contributed equally to this Correspondence. This Correspondence was supported by a Medical Research Council (MRC) grant to LEM (MR/R008698/1), a MRC studentship (MR/N013867/1), and a MRC grant (MR/M008614) and a National Institutes of Health grant (R01AI55182) to DP. We thank Peter Cherepanov of the Francis Crick Institute (London, UK) for supplying recombinant S1 antigen. We are grateful to Rebecca Matthews, Paulina Prymas, Marzia Fiorino, Thomas Fernandez, Nnenna Ngwu, the clinical research teams at the Mortimer Market Centre (London, UK) and the Ian Charleson Day Centre (London, UK), and all the clinic staff and participants.

*Emma Touizer, Aljawharah Alrubayyi, Chloe Rees-Spear, Natasha Fisher-Pearson, Sarah A Griffith, Luke Muir, Pierre Pellegrino, Laura Waters, Fiona Burns, Sabine Kinloch, Sarah Rowland-Jones, Ravindra K Gupta, Richard Gilson, \*Dimitra Peppas, \*Laura E McCoy*  
 dimitra.peppas@ndm.ox.ac.uk;  
 l.mccoy@ucl.ac.uk

Division of Infection and Immunity (ET, C-RS, SAG, LM, LEM), Department of Immunology, Royal Free Campus (SK), and Institute for Global Health (FB, RG), University College London, London WC1E 6BT, UK; Nuffield Department of Clinical Medicine, University of Oxford, Oxford, UK (AA, NF-P, SR-J, DP); The Mortimer Market Centre, Department of HIV, CNWL NHS Foundation Trust, London, UK (PP, LW, DP); Department of Infection and Immunity (SK) and Department of HIV Medicine (FB), Royal Free NHS Foundation Trust, London, UK; Department of Medicine, University of Cambridge, Cambridge, UK (RKG)



See Online for appendix

- 1 Alrubayyi A, Gea-Mallorquí E, Touizer E, et al. Characterization of humoral and SARS-CoV-2 specific T cell responses in people living with HIV. *bioRxiv* 2021; published online Feb 16. <https://doi.org/10.1101/2021.02.15.431215> (preprint).
- 2 Polack FP, Thomas SJ, Kitchin N, et al. Safety and efficacy of the BNT162b2 mRNA Covid-19 vaccine. *N Engl J Med* 2020; **383**: 2603–15.
- 3 Sahin U, Muik A, Derhovanessian E, et al. COVID-19 vaccine BNT162b1 elicits human antibody and TH1 T cell responses. *Nature* 2020; **586**: 594–99.
- 4 Collier DA, De Marco A, Ferreira IATM, et al. Sensitivity of SARS-CoV-2 B.1.1.7 to mRNA vaccine-elicited antibodies. *Nature* 2021; published online March 11. <https://doi.org/10.1038/s41586-021-03412-7>.
- 5 Frater K, Ewer KJ, Ogbé A, et al. Safety and immunogenicity of the ChAdox1 nCoV-19 (AZD1222) vaccine against SARS-CoV-2 in HIV infection. *SSRN* 2021; published online April 19. [https://papers.ssrn.com/sol3/papers.cfm?abstract\\_id=3829931](https://papers.ssrn.com/sol3/papers.cfm?abstract_id=3829931) (preprint).
- 6 El Chaer F, El Sahly HM. Vaccination in the adult patient infected with HIV: a review of vaccine efficacy and immunogenicity. *Am J Med* 2019; **132**: 437–46.
- 7 Ng KW, Faulkner N, Cornish GH, et al. Preexisting and de novo humoral immunity to SARS-CoV-2 in humans. *Science* 2020; **370**: 1339–43.

## Need for transgender-specific data from Asia

In *The Lancet HIV*, Venkatesan Chakrapani<sup>1</sup> commented on the study by Adrian D Smith and colleagues<sup>2</sup> and suggested transgender-specific data collection to inform the redress of health inequalities. We believe heeding this call for transgender-specific data could benefit efforts in Asia to identify gaps in HIV response for transgender people.

First, transgender-specific HIV care cascade data are scarce. This scarcity could be because the data are usually merged with data from men who have sex with men (MSM) or not reported due to negligence or not enough attention being paid to this group of people.<sup>3</sup> Among the countries where data were available, HIV prevalence among transgender people reached 24.8% in 2019 in Indonesia, and increased in countries that used integrated biological and behavioural surveillance (eg, Cambodia, Malaysia, and Thailand).<sup>3,4</sup> The percentage of transgender people

reporting HIV testing in the past year ranged from 15% (Philippines) to 89% (Nepal). However, the percentage of transgender people with HIV receiving antiretroviral therapy was not reported, or low if available.<sup>4</sup> Hence, Asian countries need to include transgender people as a separate group in HIV surveillance programmes and expand current programmes to include smaller cities and rural areas.

Second, use of HIV prevention services is low among transgender people in Asia. As of 2019, less than 50% of transgender people reportedly received a combined set of HIV prevention interventions in Bangladesh, Nepal, Pakistan, Philippines, and Sri Lanka.<sup>4</sup> Data on the use of pre-exposure prophylaxis (PrEP) were consistently included with data from MSM in Asia.<sup>3</sup> Our cross-sectional study reported in 2020 also showed a low PrEP uptake among transfeminine people in China (24 [8.7%] of 277 people).<sup>5</sup> Additionally, societal stigma and discrimination consistently hindered transgender people's access to PrEP and other HIV services in Asia.<sup>3,6</sup> Therefore, it is crucial to consider the socioecological systems that dictate access to HIV care and to plan data collection strategies accordingly. We suggest researchers collect interpersonal and structural-level data through both quantitative and qualitative methods when considering health inequalities in transgender people.

Third, few data exist on mental health and gender-affirming interventions among transgender people in Asia.<sup>7</sup> Using the gender minority stress framework, our study in China found that gender-identity-related stress can effect transgender people's engagement with gender-affirming interventions and HIV prevention services.<sup>5</sup> As access to HIV care services is interconnected with mental health and gender affirmation,<sup>8</sup> we recommend collecting data related to these areas to address health inequalities.

Now is the time to tackle health inequalities for transgender people.

We declare no competing interests. This work was supported by the National Nature Science Foundation of China (81903371), the National Key Research and Development Program of China (2017YFE0103800), the National Institutes of Health (NIAID K24AI143471), the University of North Carolina Center for AIDS Research (NIAID 5P30AI050410), and the National Institute of Mental Health (R34MH119963).

Yongjie Sha, Gifty Marley,

\*Weiming Tang

[weiming\\_tang@med.unc.edu](mailto:weiming_tang@med.unc.edu)

Guangdong Second Provincial General Hospital, Guangzhou 510095, China (YS, WT); University of North Carolina Project-China, Guangzhou, China (YS, GM, WT); School of Public Health of Nanjing Medical University, Nanjing, China (GM)

- 1 Chakrapani V. Need for transgender-specific data from Africa and elsewhere. *Lancet HIV* 2021; **8**: e249–50.
- 2 Smith AD, Kimani J, Kabuti R, Weatherburn P, Fearon E, Bourne A. HIV burden and correlates of infection among transfeminine people and cisgender men who have sex with men in Nairobi, Kenya: an observational study. *Lancet HIV* 2021; **8**: e274–83.
- 3 van Griensven F, de Lind van Wijngaarden JW, Eustaquio PC, et al. The continuing HIV epidemic among men who have sex with men and transgender women in the ASEAN region: implications for HIV policy and service programming. *Sex Health* 2021; **18**: 21–30.
- 4 UNAIDS. HIV and AIDS data hub for Asia Pacific. 2019. <http://aphub.unaids.org/> (accessed March 15, 2021).
- 5 Sha Y, Dong W, Zheng L, Muessig K, Tang W, Tucker JD. Unmet health needs and gender minority stress among transgender individuals: a cross-sectional study in China. *UNC Project-China Annual Meeting*; Guangzhou, China; 2020 (abstr 12).
- 6 Tang W, Dong W, Huang X. Addressing unmet health needs among Chinese transgender individuals. *Sex Health* 2021; published online March 5. <https://doi.org/10.1071/SH20213>.
- 7 Zhu X, Gao Y, Gillespie A, et al. Health care and mental wellbeing in the transgender and gender-diverse Chinese population. *Lancet Diabetes Endocrinol* 2019; **7**: 339–41.
- 8 Shaikh S, Mburu G, Arumugam V, et al. Empowering communities and strengthening systems to improve transgender health: outcomes from the Pehchan programme in India. *J Int AIDS Soc* 2016; **19** (suppl 2): 20809.