1	Green and simple approach for low-cost bioproducts preparation and \mathbf{CO}_2
2	capture
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27 Abstract

This study has demonstrated, for the first time, a simple, fast and flexible microwave processing 28 method for the simultaneous preparation of bio-products (bio-oil, bio-gas and biochar) using a 29 methodology that avoids any form of catalyst or chemical activation. The dielectric properties 30 of biomass and physicochemical characterisation such as TGA, elemental and proximate 31 analysis, XRD, SEM/EDX and textural properties, showed that 8 kJ g⁻¹ of microwave energy 32 can produce superior biochars for applications in CO₂ capture. The maximum CO₂ uptake 33 capacity for biochar produced was 2.5 mmol g⁻¹ and 2.0 mmol g⁻¹ at 0 and 25 °C and 1 bar, 34 which and also exhibited high gas selectivity compared with N₂, fast kinetics of adsorption 35 (<10 min) and desirable reusability (>95 %) after 20 cycles. GC-MS analysis of generated bio-36 oil products revealed that higher microwave energies (>8 kJ g⁻¹) significantly enhanced the 37 amount of bio-oil produced (39%) and specifically the formation of levoglucosan, furfural and 38 39 phenolics compounds, and bio-gas analysis identified trace levels of H₂ and CH₄. The results from this study confirm a green, inexpensive and efficient approach for biomass valorisation 40 41 which can easily be embedded within bio-refinery process, and also demonstrates the potential 42 of biochars for post-combustion CO₂ uptake.

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51 KEYWORDS: Lignocellulosic Biomass, Microwave pyrolysis, Bio-oil, Biochar, CO₂
52 capture

53 1 Introduction

The issues regarding climate change have attracted considerable attention, particularly since 54 55 the average global temperature has been estimated to increase due to the greenhouse gas emissions. In 2011, the European Union reaffirmed its objective to reduce carbon dioxide 56 57 (CO₂) emissions by 80-90 % ("Nations, U. Paris Agreement to the United Nations Framework Convention on Climate Change," 2015)(Zappa et al., 2019). In order to achieve these goals, 58 research efforts have focussed on the sustainable valorisation of biomass feedstocks and their 59 conversion into energy, fuels and bio-products; with emphasis on processes that can reduce 60 emissions and promotes circular bio-economy (Acevedo-García et al., 2020)(Olabi, 2019). 61

The transformation of biomass residues to chemicals has the potential to produce bio-products 62 base for bio-refinery, adhesives, surfactants and plasticisers using technologies such as 63 64 fermentation, extraction, hydrolysis, gasification and pyrolysis. In this sense, it is estimated that more than 422,800 tons of pecan nut shell (NS) (Carya illinoinensis) are produced yearly, 65 representing a significant volume of waste available for the generation of added value bio-66 67 products (Agustin-Salazar et al., 2018). Recent reports highlight its use for the preparation of reduced sugars (Santos et al., 2020), as reinforcing filler in poly (lactic acid) biocomposites 68 (Agustin-Salazar et al., 2018), gasification for syngas generation (Aldana et al., 2015) (Lozano 69 and Lozano, 2018) and as precursor of adsorbent for energy storage (Martínez-Casillas et al., 70 2019). 71

Amongst the available thermochemical processes, microwave pyrolysis has proved to be a very attractive processing technology, due to its nature and volumetric heating which offers the possibility to heat selectively and therefore maximize the production of desired products (Durán-Jiménez et al., 2015)(Kostas et al., 2020)(Kostas et al., 2017)(Beneroso et al., 2017). The heating effect resulting from the interaction of microwave energy with biomass

predominantly occurs through coupling of both permanent and induced dipoles, as well as ionic 77 components of the material to the oscillating electromagnetic field. This motion then induces 78 molecular friction and generation of heat within the volume of the material (Kostas et al., 2019). 79 The energy is transferred from inside to outside of the particle which results in an inverted 80 temperature gradient (Haeldermans et al., 2019)(Kostas et al., 2017). Compared with 81 conventional heating, this technology offers a variety of advantages, such as greener 82 83 instantaneous heating and the elimination of catalyst or other auxiliary chemicals, which means that wastes are not generated (Durán-Jiménez et al., 2020)(Yin, 2012). By the evaluation of 84 85 dielectric properties, microwave have proved to be compatible with biomass feedstock pyrolysis processes to generate a range of bio-based products including bio-oils and biochars 86 (Kostas et al., 2019)(Yin, 2012)(J. Li et al., 2016)(Kostas et al., 2020)(Sears et al., 87 2006)(Durán-Jiménez et al., 2016)(Ferrera-Lorenzo et al., 2014). Bio-oils typically contain a 88 mixture of oxygenated compounds (Kan et al., 2016), and have the potential to be catalytically 89 upgraded to fuel. Biochar is a porous carbon-based material used in several applications 90 including batteries, catalyst, soil remediation and as adsorbent of pollutants in liquid and gas 91 systems (Li et al., 2017)(Kan et al., 2016). 92

93 Nowadays, several materials such as fly ashes (Bui Viet et al., 2020) biomass, cement kiln dust, 94 paper sludge, sewage sludge (Sanna et al., 2012), activated carbons (Srinivas et al., 2014)(Hoseinzadeh Hesas et al., 2015)(Yang et al., 2017), zeolites and metal organic 95 frameworks (Lin et al., 2014)(Liang et al., 2009)(Saha et al., 2010) have been investigated for 96 97 CO₂ capture. Particular interest in the preparation of biochar for CO₂ capture has also been reported due to its simplicity and low cost (Huang et al., 2015). However, research has focused 98 on the modification of biochar by chemical and physical activation (Li and Xiao, 2019)(Y. Li 99 et al., 2016)(C. Zhang et al., 2016) or the incorporation of metals into the skeleton of biochars 100 101 (Lahijani et al., 2018) to enhance their CO_2 adsorption capacity. This modification has resulted in the increase of the costs and difficulties in the regeneration limiting its industrial application.

103 Currently, there is a huge need in the study of more environmentally friendly processes for the synthesis of biochars with effective adsorption properties. However, the global biomass 104 valorisation (liquid, solid and gas fraction) and the detailed assessment of CO₂ adsorption 105 capacities of biochars obtained without chemical modification and microwave heating, have 106 not been evaluated. The conversion of waste biomass feedstocks into added-value products will 107 108 address issues related to its disposal, which is essential for a sustainable global circular bioeconomy (Acevedo-García et al., 2020). Furthermore, the application of products generated in 109 one step that also avoids the use of catalysts is highly appealing from an environmental and 110 111 energy consumption perspective.

To our knowledge, this is the first study that employs nut shell as a renewable source that complies within a bio-refinery setting for multiple generation of added value products. This study does not only focus on the characterisation of generated bio-oil from microwave pyrolysis process, but also in the application of biochar. A detailed discussion of the physicochemical characterisation and evaluation of the simple-to-produce biochars for postcombustion CO_2 capture is presented, with assessments focussing on the kinetics, selectivity, reusability and equilibrium of adsorption of CO_2 at relevant temperatures (0 and 25 °C).

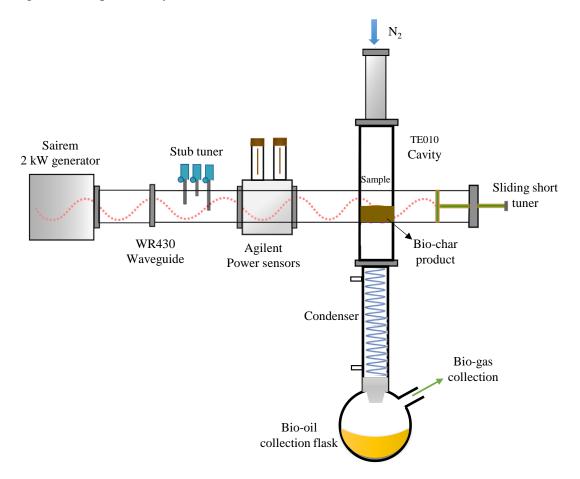
- 119 2 Materials and methods
- 120 **2.1 Materials**

In this study pecan nut shell (NS) was used as raw material. Prior to pyrolysis experiments, NS
was washed and dried at 70 °C.

123 **2.2 Biomass pyrolysis experiments**

124 The system used in the microwave pyrolysis is shown in Fig. 1, and is composed by one mode

- applicator, 2 kW microwave generator (2.45 GHz), an automatic tuner (S-TEAM STHD v1.5)
 and sliding short to maximise the power density in the sample. (Durán-Jiménez et al., 2020).
 The sample was placed in a quartz reactor in nitrogen atmosphere (2 L min⁻¹).
- The microwave processing conditions were in the range of 300-400 W input power and 2 to 6 min duration time as shown in Table S1. The liquid fraction was stored in a pre-weighed vials at 4 °C. The non-condensable gases were passed into a gas line and collected using a tedlar gas bag for subsequent analysis.



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Fig. 1. Microwave pyrolysis system used in this work

It should be noted that it is unfeasible to accurately measure the sample temperature as
microwave heating causes volumetric and instantaneous heating (Nagahata and Takeuchi,
2019).

- 137 The biochar and bio oil yield were quantified by their weights and the yield of the non-
- 138 condensable product was calculated by difference as follow:

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$$Y_{Bio-char} = \frac{M_{Bio-char}}{M_0} \times 100$$
(1)

140
$$Y_{Bio-oil} = \frac{M_{Bio-oil}}{M_0} \times 100$$
 (2)

141
$$Y_{Bio-gas} = 100 - [Y_{Bio-char} + Y_{Bio-oil}]$$
 (3)

143 2.3 Characterisation techniques

144 2.3.1 Dielectric properties

The dielectric response of a material is commonly presented as permittivity (ε) which can begiven by:

147
$$\epsilon = \epsilon_0 \epsilon_r = (\epsilon_r' - j \epsilon_r'') \tag{4}$$

where ε_0 is permittivity of the free space (8.854x10-12 F m⁻¹), ε_r is complex relative 148 permittivity, and j is imaginary unit (j^2 =-1). The complex relative permittivity is composed by 149 the real part, called relative dielectric constant (ε_r), and it is a measure of the ability of the 150 material to store electrical energy; and the imaginary part, (ε_r'') known as relative dielectric loss 151 factor, represents the energy dissipated. Dielectric properties of NS were obtained using the 152 cavity perturbation technique for a temperature range of 20 - 650 °C at 2450 MHz (Durán-153 Jiménez et al., 2020). The sample was placed in the system using a quartz tube of (ID) 3 mm 154 that is moved by an automated motor to the furnace and cylindrical TM0n0 mode cavity. The 155 dielectric properties were calculated using the cavity response between a load and empty tube 156 measured by a HP 8753 vector network analyzer (VNA). The results presented in this work are 157 the averages of 3 replications. 158

159 2.3.2 Bio-gas and Bio-oil Analysis

160 The bio-gas generated during the pyrolysis experiments were analysed as described somewhere161 else (Almustapha et al., 2017), using a Clarus 580 Gas Chromatograph (GC) system.

The bio-oil were analysed by GC-MS using a Varian CP-3800 GC incorporated to a Varian 163 1200 MS (70 eV, EI mode) (Kostas et al., 2019). The identification of the compounds in the 164 bio-oils was determined according to the NIST Mass Spectral library from the National 165 Institute of Standards and Technology, Maryland, USA.

166 **2.3**.

2.3.3 Biochar and NS characterisation

Proximate analysis was conducted by the method previously described by Donahue (Donahue and Rais, 2009). The sample was placed in TGA Q500 TA Instrument and heated to 900 °C under a N₂ atmosphere (100 ml min⁻¹, 1 bar) and then held at 900 °C for 15 min, the atmosphere was then switched to air and held for a further 15 min. The ultimate analysis was determined using a LECO CHN-628 elemental analyzer. A Philips XL 30 microscope was used to analyze the morphology and the X-ray diffractograms were obtained by using a Bruker D8 Advance Da Vinci.

The textural parameters were calculated from the N₂ adsorption isotherms at -196 °C using a 174 Micromeritics ASAP 2420 apparatus. Brunauer–Emmett–Teller (BET) theory was used to 175 calculate the surface area while the micropore, total pore volume and size distributions were 176 determined by Non-Local Density Functional Theory (NLDFT) (Thommes et al., 2015) 177 (Thommes, M., Kaneko, K., Neimark, A. V., 2015)(Thommes, M., Kaneko, K., Neimark, A. 178 V., 2015)(Thommes, M., Kaneko, K., Neimark, A. V., 2015)[42](Thommes, M., Kaneko, K., 179 Neimark, A. V. 2015)(Thommes, M., Kaneko, K., Neimark, A. V., 2015)(Thommes, M., 180 Kaneko, K., Neimark, A. V., 2015)(Thommes, M., Kaneko, K., Neimark, A. V., 181 2015)(Adeniran et al., 2014) by combining a CO₂ adsorption isotherm at 0 °C to a N₂ 182 adsorption isotherm. 183

184 2.4 CO₂ capture experiments

185 The CO₂ uptake of the generated biochars were determined by thermogravimetric and

volumetric analysis. The volumetric experiments were conducted in a Micromeritics ASAP 186 2420 analyser, where approximately 500 mg of biochar was degassed at 120 °C in N₂ for 15 h. 187 The amount of CO₂ adsorbed was determined using 100 % CO₂ in the range of 0.001 to 1.2 188 bar. The thermogravimetric uptake was performed using a TGA Q500 TA instrument. 189 Approximately 25 mg of biochar was placed in a pan and dried for 30 min at 120 °C and 190 atmospheric pressure. After the physiosorbed moisture was removed, the sample was cooled 191 192 down to 25 °C and the gas was switched at 100% CO₂. Once the adsorption reached equilibrium, the adsorbed amount of CO₂ was recorded vs time for 60 min, then the sample 193 194 was heated up to 120 °C in N₂ for 15 min to complete the CO₂ desorption. In total, 20 adsorption-desorption cycles were conducted to determine stability and recyclability of the 195 biochar. The selectivity was determined by thermogravimetric analysis at 25 °C using N₂ (100 196 ml min⁻¹) for the adsorption stage and Ar in the desorption stage. All experiments were 197 performed in triplicate and the averages are presented. In general, standard deviations were 198 below 5 % of average values. 199

200 **3 Results and Discussion**

201 3.1 Microwave pyrolysis and bio-product formation mechanism

The TG and DTG curves obtained from the pecan NS and the dielectric characterisation are shown in Fig. 2. Based on the thermograms, the decomposition of the biomass occurs in three main stages. The first stage involves the removal of water at temperatures up to 150 °C, whereas the second and third stages describe the hemicellulose and cellulose decomposition (up to 400 ° C), and the lignin transformation into char (beyond 400 °C).

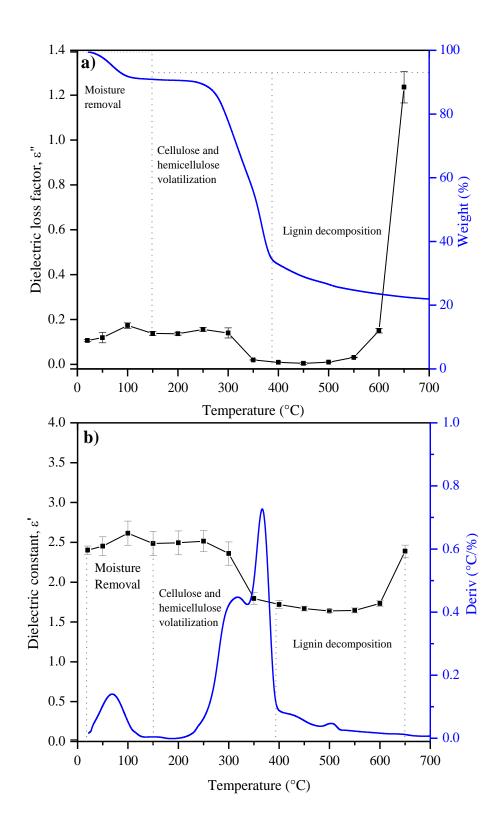
In microwave heating, the dielectric constant and loss factor determine the capability of the biomass to be heated by microwave heating. The results of dielectric characterisation illustrate that at temperatures below 150 °C, is the water the main responsible for the microwave absorption due to dipolar mechanisms. When the water is removed the dipolar movements
decreased leading to the reduction the dielectric constant and loss factor. However, at
temperatures above 600 °C the formation of char is promoted and hence an exponential
increase in the dielectric properties is observed, as results of the increase in the conductivity of
the sample. Similar behaviour for lignocellulosic biomasses have been previously reported
(Miura et al., 2004)(Namazi et al., 2015).

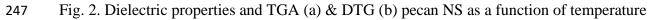
The distribution of the three pyrogenic products generated from the pyrolysis of NS using microwave heating, are presented in Table S1. The yields of biochar are in the range of 16 – 18 %. It has been reported the bio-oil yields for lignocellulosic biomasses such as pine wood and corn stover ranges between 20- 40 % (Kan et al., 2016) (Jung et al., 2019). Similar yields were found in this study (21.5 - 40 %).

The results indicate the higher bio-oil yields are a result of the faster processing times achieved 221 in microwaves, and also confirm that microwave pyrolysis could be thermochemically faster 222 223 than slow pyrolysis in a conventionally heated process (Huang et al., 2015). The bio-oil yields 224 were correlated to the specific energy, which is a measure of the real absorbed energy by the sample under processing. It appeared that at higher energies but low intensities (300 W) is 225 possible to obtain higher bio-oil yields, which can be as result of a more controlled volatile 226 matter evolution in the pyrolysis process. For example, the samples prepared at 300 W showed 227 a positive trend where the highest bio-oil yield (39.0%) corresponds to the higher specific 228 energy (8 kJ g⁻¹). This behaviour, was different for bio-oils produced using incident power of 229 400 W where the yield decreased for samples prepared at higher energy (24.8% at 8.4 kJ g^{-1}) 230 231 This phenomena can be explained in terms of heating rate and power density. For samples IV-N (300 W - 6 min) and VII-NS (400 W - 4 min), similar specific energy was reported, however 232 in VII-NS the heating rate and power density were higher, leading to the preferred gas 233 234 production and lower bio-oil yields.

This study confirms that the NS bio-oil yield is highly governed for the energy, but also for the power density used in the pyrolysis. This is of great relevance and establish the basis for microwave processing where the maximisation of bio-oil is priority.

The complex transformation of biomass into biochar and bio-oil has been proposed as result of 238 several depolymerisation reactions. The model describes groups in cellulose and hemicellulose 239 such as glucomannans and containing glycosidic bond that broke down to into levoglucosan 240 that further undergoes dehydration and intramolecular arrays to produce furfural and furan 241 (Singh et al., 2019). Lignin is a complex cross- linked polysaccharide and its decomposition 242 results in the formation of the biochar network (J. Li et al., 2016). The polymerisation and 243 244 aromatisation mechanisms produce the formation of wide-range phenols in the liquid phase and rich carbon content solid (biochar). 245





248 3.2 Bio-oil and bio-gas characterisation

249 The most prominent compounds identified in bio-oils were semi-quantitatively analysed and

are shown in Table 1. Fig. 3 illustrates a typical GC-MS total ion chromatogram, and all 250 samples are shown in Fig. S1. It can be seen that bio-oils are a mixture of numerous organic 251 compounds that have resulted from the pyrolytic breakdown of lignin, cellulose and 252 hemicellulose as per in Fig. S2, which include water, vanillins, furan carboxaldehydes, 253 pyrones, acetic acid, hydroxy-aldehydes and phenolics. A range of phenolic based compounds 254 such as methoxyphenol, dimethylphenol, ethylmethoxyphenol, methoxyvinylphenol, 255 256 methoxypropenylphenol were detected with totalling concentrations ranging between 28 to 44.5 % of the identified compounds. The presence of the aforementioned phenol derivatives 257 258 have resulted from the depolymerisation of lignin and its monomeric constituents, which according with the TGA analysis in Fig. 2, typically occur at temperatures between 400 and 259 600 °C (Mohan et al., 2006). Phenolic compounds can be used in the synthesis of bio-plastics, 260 resins and epoxy polyurethane materials (Mamaeva et al., 2016). 261

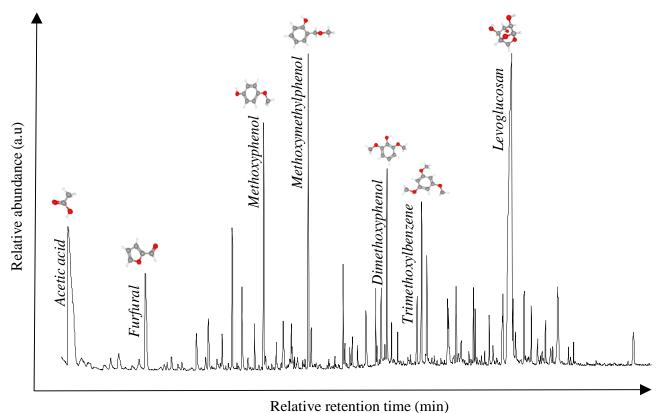
262 Cellulose degradation is expected to occur at lower temperatures (usually between 240 - 350 °C) and results in the formation of anhydrocellulose and levoglucosan (Mohan et al., 263 2006)(Kostas et al., 2020)(Wu et al., 2009). Around 8.6 – 23.1 % of the total compounds 264 identified in the generated bio-oils from this study was indeed levoglucosan, representing a 265 valuable precursor fraction for conversion into plastics, surfactants, and bio-polymers (Rover 266 267 et al., 2019). On the contrary, the main monomeric products from hemicellulose decomposition led to the formation of furfural and acetic acids, ranging between 13.2 to 25.2 %. Other 268 compounds present in the bio-oils were aldehydes (6.5 - 12.4 %), methyl, vanillins and benzene 269 270 and its derivatives (21.3 - 29.3 %).

Overall, the range of compounds identified in the bio-oil fractions generated in this study are comparable to bio-oils typically generated from lignocellulosic feedstock materials, offering wide opportunity for upgrading and incorporation bio-product manufacture (Pinheiro Pires et al., 2019).

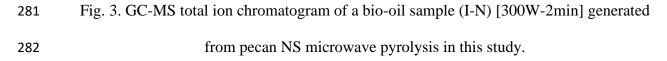
Table 1. Semi-quantitative bio-oils composition of NS biomass.

	RetentionPercent of total identified (%)							
Compound	Time (min)	I-N [300-3]	II-N [300-4]	III-N [300-5]	IV-N [300-6]	V-N [400-2]	VI-N [400-3]	VII-N [400-4]
Acetic acid	7.60	4.91	6.28	2.93	5.65	8.69	5.76	13.23
Furfural	12.64	8.28	6.99	4.47	8.18	13.47	11.48	11.95
Cyclopentanedione	15.99	1.30	1.32	1.36	1.27	1.27	1.23	1.49
Furanone	17.67	1.33	1.45	1.38	1.37	1.85	1.41	1.48
Oxazolidine, 2,2-diethyl-3-methyl-	18.34	5.04	4.58	5.50	4.69	5.88	5.35	7.98
Cyclopentanedione	18.97	3.46	2.98	2.95	2.99	3.68	3.10	3.44
Phenol	19.81	1.30	1.38	1.61	1.70	2.53	2.08	2.68
Methoxyphenol	20.40	7.87	5.23	5.42	4.64	5.68	4.94	6.66
Methylphenol	22.22	1.36	0.70	0.78	0.94	1.23	1.12	1.42
Methoxymethylphenol	23.33	9.85	6.39	7.66	6.14	6.81	6.57	8.09
Dimethylphenol	23.51	1.42	1.37	1.74	1.27	1.54	1.17	1.60
Ethylmethoxyphenol	25.62	2.75	1.45	1.76	0.32	0.40	1.57	1.77
Methoxyvinylphenol	27.10	2.86	2.19	3.16	3.04	2.81	3.20	3.42
Methoxypropenylphenol	27.77	2.09	1.26	1.46	1.28	1.40	1.42	1.70
Hydroxymethylfurancarboxaldehyde	28.11	3.13	1.88	2.41	2.04	2.59	2.01	2.48
Dimethoxyphenol	28.49	6.16	3.67	5.01	3.70	4.25	3.67	3.98
Methoxypropenylphenol	29.19	1.04	0.58	0.83	0.73	0.63	0.82	0.57
Methoxypropenylphenol	30.46	2.25	1.50	2.76	2.61	2.15	2.81	1.71
Trimethoxylbenzene	30.76	4.85	2.31	4.67	4.27	3.04	4.12	2.80
Vanillin	31.10	3.50	1.93	2.54	2.15	3.20	2.39	2.37
Methoxypropylphenol	32.47	1.93	1.01	1.42	1.05	0.64	0.11	0.74
Methyltrimethoxylbenzene	32.52	1.49	1.12	1.82	1.23	1.11	1.16	0.85
Hydroxymethyoxyphenyl	33.03	2.02	1.13	1.62	1.22	1.58	1.29	1.16
Hydroxymethoxyphenylpropanone	34.17	1.96	0.86	1.23	0.88	0.97	0.82	0.64
Dimethoxypropenylphenol	34.29	1.69	1.00	1.39	1.22	1.33	1.16	0.83
Hydroxymethoxyphenylpropanone	35.20	1.41	0.75	1.02	0.78	0.79	0.79	0.22
Levoglucosan	36.50	2.28	32.12	19.68	15.69	10.41	9.91	8.61
Dimethoxypropenylphenol	36.76	1.96	1.01	2.50	10.89	1.53	12.16	1.27
Hydroxydimethoxybenzealdehyde	37.49	2.10	1.32	1.91	1.57	1.80	1.46	1.19
Hydroxydimethoxyethanone	38.90	1.25	0.79	1.14	0.85	0.96	0.76	0.61
Hydroxymethoxycinnaanaldehyde	39.70	5.52	2.83	4.58	4.35	4.10	3.00	2.39
Dimethoxyhydroxycinnanaldehyde	44.66	1.64	0.62	1.30	1.29	1.68	1.16	0.68

277 *Preparation conditions [Power (W)-time (min)]



Relative retention time (IIIII)



Although the synthesis of bio-gas is beyond the scope of this work, attempts were made to 283 analyse the non-condensable fraction obtained from the pyrolysis of biomass and it was found 284 trace amounts of hydrogen (H₂) and methane (CH₄) (Fig. S3). Reasons for the trace levels could 285 be due to the fact non catalyst were used in the process to maximize the bio-gas production 286 (Weihong et al., 2019), but also due to that a relatively high nitrogen gas flow was applied in 287 order to maintain an oxygen-free atmosphere and therefore aimed the dilution of gases 288 289 produced. Despite the synthesis of bio-gas was not the main objective of this study, is important to highlight that studies have demonstrated the potential of NS for syngas production (Aldana 290 et al., 2015), confirming the versatility and opportunities that this biomass offers for its 291 292 valorisation.

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293 **3.3 Biochar characterisation**

The elemental composition of the generated biochar samples is presented in Table 2, and show 294 that regardless of the experimental conditions that were applied for the preparation of the 295 biochars, biomass conversion into biochar was fully achieved as indicated by the carbon 296 297 content ranging between 83.1 to 91.27 %. The hydrogen content (0.12 - 0.99 %), is associated with the small portion of compounds which remained in the solid carbon matrix. The biochars 298 produced contained a high number of aromatic groups and relatively small aliphatic chains as 299 indicated by a low H/C ratio, confirming that the aromaticity is due to dehydration, 300 decarboxylation and cyclization reactions (Coromina et al., 2015). For all biochar samples, the 301 carbon content increased for samples prepared at higher energies, for example VII-N has a 302 specific energy of 8.4 kJ g⁻¹ and a carbon content of 91.3 %, suggesting a higher level of 303 graphitisation (See Tables 1 and 3). 304

The thermogravimetric profiles (TGA) of biochars are illustrated in Fig. S4. The weight losses 305 observed from room temperature up to 110° C, are as result of the moisture removal. From 110 306 ° C all biochars showed a weight loss due to the volatile matter degradation. The proximate 307 analysis is summarised in Table 2. The pecan NS has fixed carbon (10.8 %), volatile matter 308 (84.4 %), moisture (3.6 %) and ash (1.3 %) (Durán-Jiménez et al., 2017). For the biochar, the 309 310 data reveals a significant increase in fixed carbon whereas the volatile content shows a reversed trend. An increase in the microwave pyrolysis energy led to an increasing conversion to char, 311 312 and reduction in the volatile matter. The increases observed in the fixed carbon and ash content as the pyrolysis energy increases could be due to the high temperature reached in the pyrolysis. 313 The ash contents of the biochars were between 2.75 and 3.78 % and are a result of the 314 315 transformation of the calcium oxalate in the precursor.

316 The elemental composition of biochar samples is consistent with the powder XRD patterns

shown in Fig. S5. An apparent increase in the intensity can be observed for the peaks at $2\theta =$ 317 23 ° and 2 θ = 43 °. Both peaks are ascribed to the diffraction of graphitic carbon domains (002) 318 and (101) (Lahijani et al., 2018). The wide peak at 23 ° is associated with the highly disordered 319 structure or amorphous carbon. The peak at $2\theta = 43^{\circ}$ correspond to the spacing between 320 aromatic layers (Serafin et al., 2019). Additionally, the pecan NS biomass is rich in calcium in 321 the form of oxalates (CaC₂O₄·H₂O), which progressively disappeared at higher microwave 322 323 energy and at the time new peaks of CaCO₃ were formed in the biochar. This behaviour can be due to higher energies promote the decomposition of the oxalates, which is expected to occur 324 325 at temperatures beyond 450 °C.

The morphology of the biochar prepared using a microwave power of 300 W for 6 min (IV-N) was analysed by SEM (Fig. S6 5 a-b) and EDX (Figs. S6 c-d). The morphology shows irregular cavities and pores (Fig. S6 a-b). The electron diffraction illustrates that the presence of superficial deposits composed of CaCO₃ (Fig. S6 c). The composition is validated by the XRD analysis (Fig. S5 b).

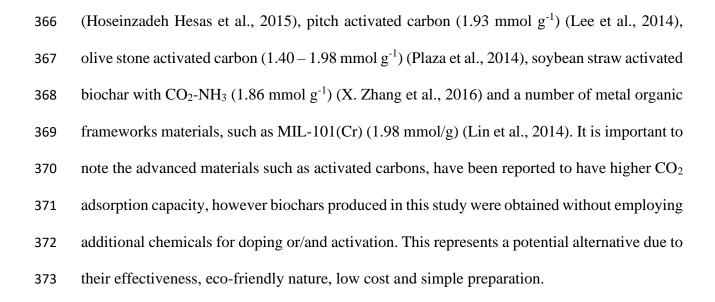
331 The textural parameters and the N₂ adsorption isotherms at -196 °C presented in Table 2 and Fig. S7. In general, the temperature required to devolatilise the biomass was successfully 332 achieved after 3 min, thus enhancing the surface area and porosity of the biochar, whereas the 333 largest specific surface area (187 m² g⁻¹) and the highest pore volume (0.075 cm³ g⁻¹) were 334 observed in the biochar prepared at 300 W for 6 min (300W -6 min). The results suggest that 335 by increasing the microwave processing time to 6 min, the degree of micropore fraction also 336 increases (85 %). The reduction in specific surface area and development of micropores along 337 with the increase of specific energies (beyond 8 kJ g⁻¹), can be related to the disintegration of 338 porous cavities as they are exposed to higher temperatures. Similar results have been previously 339 reported for biochar prepared from similar biomass feedstocks such as walnut shells (Lahijani 340 341 et al., 2018).

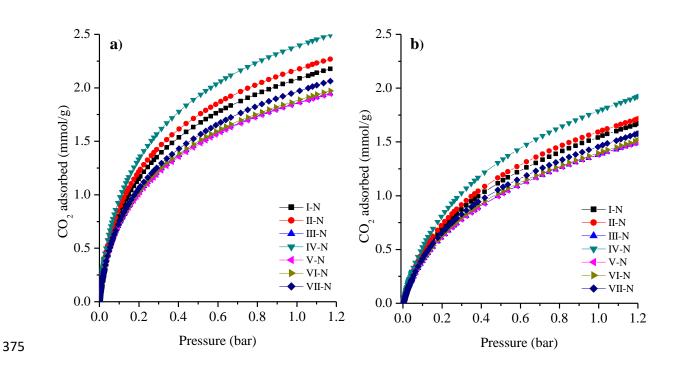
All biochars prepared exhibited a type I isotherm based on the IUPAC classification 342 (Thommes, M., Kaneko, K., Neimark, A. V., 2015) (See Fig. S7) with high nitrogen uptake at 343 low relative pressure (P/Po < 0.01); a feature that is characteristic of microporous materials 344 (Choi et al., 2019). The widening of the knee of the isotherm in sample IV-N indicates a slight 345 broadening of the micropore size range. The pore size distributions illustrate the higher 346 micropore volume in the range of 0.33 to 0.44 nm and a small portion of 1.3 and 1.8 nm pores. 347 348 The widening of the pore distribution in sample IV-N may be related to the controlled gasification of this particular biochar during pyrolysis at 6 min and 300 W. At powers of 400 349 350 W, a decrease in the textural properties was observed and may be related to the formation of hot spots which could lead to the detriment of pores by collapse of the pore walls (Huang et 351 al., 2015). 352

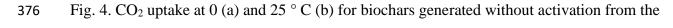
353 **3.4** Evaluation of biochar properties for CO₂ capture

The CO₂ uptake of the biochar produced was explored by volumetric analysis at 0 and 25 °C 354 (Fig. 4). The simple-to-produce biochar in this study, present moderate CO₂ adsorption 355 capacity at atmospheric pressure. The highest adsorption capacity of 2.5 and 2 mmol g⁻¹ at 0 356 and 25 °C, respectively (Fig. 4) was obtained by the biochar (IV-N) produced at 300 W for 6 357 min of microwave heating making it superior to previously published biochars (Table S2, 358 atmospheric pressure), such as rice straw (1.75 mmol g⁻¹) (Huang et al., 2015), sugar cane 359 bagasse and hickory wood (1.67 & 1.32 mmol g⁻¹) (Creamer et al., 2014), walnut shell (1.65 360 mmol g⁻¹) (Lahijani et al., 2018), hickory chips (1.10 mmol g⁻¹) (Xu et al., 2019), pig manure 361 and wheat straw (0.53 & 0.78 mmol g⁻¹) (Xu et al., 2016), mesquite wood and manure chicken 362 $(1.92 \& 1.60 \text{ mmol g}^{-1})$ (Dissanayake et al., 2020). 363

The CO₂ adsorption results also indicate that biochars have higher CO₂ adsorption capacities than some activated carbons such as oil palm shell activated carbon (1.7 mmol g^{-1})







microwave pyrolysis

	Parameter (%)								Textural parameters				
Sample	С	Н	N	аО	H/C	Moisture	Volatile Matter	Fixed Carbon	Ash	^b S _{BET} (m ² /g)	^c V _p (cm ³ /g)	^d V _{mic} (cm ³ /g)	V _{mes} (cm ³ /g)
I-N [300-3]	83.10	0.89	0.62	15.40	0.011	6.3	12.4	77.5	3.8	122	0.051	0.042	0.009
II-N [300-4]	86.58	0.83	0.80	11.79	0.010	5.2	8.6	82.4	3.7	151	0.062	0.05	0.012
III-N [300-5]	89.84	0.20	0.88	9.08	0.002	6.2	6.7	84.2	3.0	115	0.05	0.039	0.01
IV-N [300-6]	87.41	0.66	0.85	11.08	0.008	5.1	9.4	82.0	3.5	187	0.075	0.066	0.009
V-N [400-2]	83.56	0.99	0.51	14.94	0.012	6.0	7.1	84.2	2.8	42	0.02	0.017	0.003
VI-N [400-3]	87.1	0.37	0.82	11.71	0.004	4.7	9.0	82.5	3.7	134	0.057	0.047	0.01
VII-N [400-4]	91.27	0.12	0.97	7.64	0.001	4.1	6.1	86.6	3.3	112	0.048	0.038	0.01
NS	47.27	6.41	0.18	46.14	0.136	3.6	84.4	10.8	1.3	-	-	-	-

380 Preparation conditions [Power (W)-time (min)]

381 ^a Calculated by difference (%O=100-%C-%N-%H)

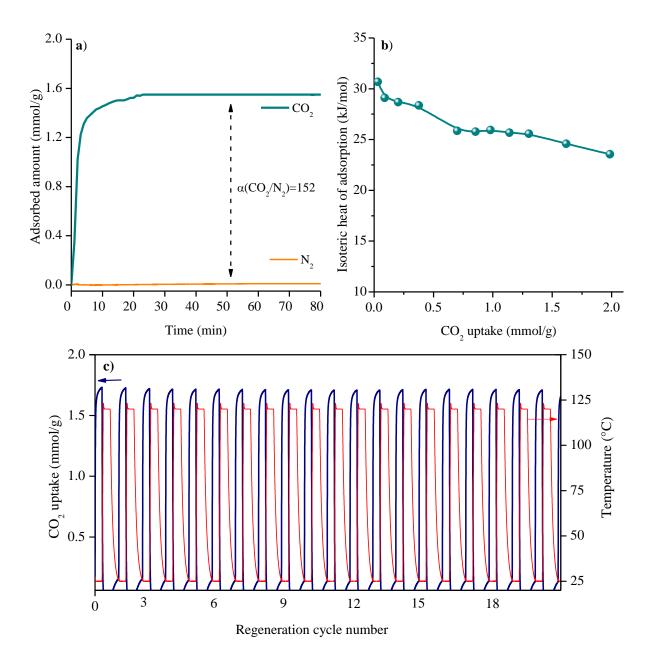
382 ^b BET surface area calculated by the BET method.

^c Total pore volume recorded at 100 nm on cumulative pore volume by NLDFT carbon slit pore model.

^d Micropore volume obtained at 2 nm on cumulative pore volume by NLDFT carbon slit pore model.

The CO₂ uptake capacity was correlated to the higher specific surface area and micropore 383 volume. In our previous study, it was demonstrated that at pressures below to 1 bar, the CO₂ 384 uptake is highly controlled by micropores with size below 0.7 nm (Durán-Jiménez et al., 2020). 385 The results this work are also in agreement with the work published by Presser et al. (Presser 386 et al., 2011) where was found that high surface area (>2900 m² g⁻¹) with large total pore volume 387 $(> 1.5 \text{ cm}^3 \text{ g}^{-1})$ is negligible for high CO₂ uptake and that high CO₂ adsorption will result in 388 adsorbents with large narrow micropore volume. These findings are in correlation with the 389 results shown in Fig. S8 that indicate the highest fraction of CO₂ filled pore occurs in pores of 390 391 size of 0.7 nm, without further uptake increment for pores with size beyond 0.8 nm. This study confirms that pores larger than three times the molecular diameter of CO_2 (i.e. mesoporous 392 materials) does not govern the CO₂ uptake at pre combustion conditions. The correlation of 393 adsorbed amount with narrow micropores is fundamental in the design of effective and un-394 expensive adsorbents for industrial applications. Additionally, it is well-known the control of 395 narrow micropores can be favourable to increase the CO₂ kinetics. This attribute in adsorbents 396 is highly desirable for practical applications to reduce the required time of contact and energy 397 for regeneration. Fig. 5 (a) presents the adsorption kinetics of biochar IV-N at 25 °C. It shows 398 that levels of CO₂ adsorption up to 90 % can be achieved in less than 10 min, and fully 399 equilibrate after 25 min. 400

To explore the potential of biochar for industrial applications, not only do the CO₂ uptake kinetics and mechanisms need to be considered, but also the CO₂ over N₂ selectivity, the isosteric heat of adsorption, adsorbent stability and recyclability ought to be investigated. The adsorbent's selectivity determines if the CO₂ can be removed from the flue gas mixture with minimal interference from other major gaseous species. In this study the selectivity of sample IV-N for CO₂ over N₂ was determined by thermogravimetric single component analysis at 25 °C at 1 bar (Durán-Jiménez et al., 2020). The CO₂ and N₂ uptake are compared in Fig. 5 (a).



408

409 Fig. 5. CO₂ uptake kinetics & gas selectivity of biochar IV-N (a), Isoteric heat (Q_{st}) of CO₂
410 adsorption (b) and recyclability after 20 cycles of CO₂ IV-N at 25 °C (c)

The CO₂ uptake of IV-N (1.5 mmol g⁻¹) is distinctive higher than that of N₂ (0.001 mmol g⁻¹). The selectivity based on physisorption uptake, is dependent upon the different physical properties of gas molecules (Parshetti et al., 2015). Since the polarisability of CO₂ is higher than N₂, it is expected to have higher enthalpy of adsorption which leads to a higher affinity of the pore surface, and therefore a higher selectivity. It should also be noted that the selectivity

reported in this study is higher than recently reported porous carbon materials (Li and Xiao,
2019)(Lahijani et al., 2018)(Chen et al., 2018), indicating that the biochar produced in this
study could be implemented for gas separation in practical applications.

The high CO_2/N_2 selectivity of IV-N is also supported by the high isosteric heats of adsorption shown in Fig. 5 (b) calculated applying the Clausius-Clapeyron equation (Durán-Jiménez et al., 2020). The relatively high initial Q_{st} values in the low-pressure region, suggest the CO₂ molecules are being adsorbed on sites with the highest energy and narrowest micropores. At higher CO₂ uptake, the Q_{st} values decay below 30 kJ mol⁻¹ indicating the CO₂ adsorption is governed by the physisorption mechanism. The values reported for sample IV-N are similar to values reported for porous carbon materials (Chen et al., 2018) (Hong et al., 2016).

It is essential to evaluate the regeneration and reusability of the adsorbents to assess its further industrial implementation. Fig. 5c) shows the CO₂ adsorption - desorption cycles of the IV-N biochar at 100 % CO₂. It can be seen that after twenty consecutive cycles the CO₂ adsorption kinetics and equilibrium uptake remained unaffected, confirming the stability and easy regeneration. This result demonstrates the high chemical and physical stability of biochar and confirms the adsorption is reversible governed by mechanisms of the physi-sorption.

432 4 Conclusions

This work has demonstrated that a range of desireable bio-products with further applications can be effectively synthesised from pecan NS waste biomass employing only 6 min of microwave heating. The yields of the pyrolysis products were found to be greatly influenced by microwave energy and processing parameters, and 8 kJ g⁻¹ of energy was required to obtain 39 % of bio-oil that contains upgradeable chemical compounds such as levoglucosan, phenols, and furfural. The results also revealed unique characteristics of the generated biochars, such as pore size distribution and ultra-micropores being the main responsible parameter for the post440 combustion CO_2 uptake. The values reported herein (2.5 mmol CO_2 g⁻¹) are competitive or 441 higher than other values reported for other biochars, which include: significantly greater CO_2 442 uptake over N₂ and easy regeneration. These findings are of significant importance, not only 443 due to the competitive properties of biochar and due to its application in the CO_2 capture, but 444 also because it offers a greener and simple approach using microwave heating. Furthermore, 445 no doping agents were used in the synthesis methodology that was employed, making it a great 446 sustainable alternative for waste disposal.

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