- 1 Microglia tweak retinogeniculate pathways during visual circuit
- 2 refinement
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9 Abstract: Cheadle et al. reveal that microglia expressing TWEAK facilitate synapse
10 elimination through a novel, non-phagocytic mechanism in the retinogeniculate

- 11 pathway during visual circuit development. This novel mechanism is experience-
- dependent and occurs through the local binding of TWEAK to postsynaptic Fn14.
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14 Sensory-dependent plasticity plays a critical role in the development of the visual

- 15 system. While molecular cues and spontaneous activity are essential for establishing
- 16 connections early in development, visual experience is critical for the precise
- 17 refinement of these circuits in the later stages (Hooks and Chen, 2006). The
- retinogeniculate pathway has been used extensively as a model to examine both
- 19 components of circuit development. To date, a number of mechanisms have been
- 20 described for the early stages of development (Huberman, 2007), but the molecular

21 mechanisms associated with experience-dependent refinement are less clear.

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23 To examine the molecules that regulate synaptic refinement, previous work from

- 24 Cheadle et al. used single-nucleus RNA sequencing to demonstrate that the cytokine
- receptor fibroblast growth factor-inducible protein 14kDa (Fn14) was induced in
- 26 excitatory thalamocortical neurons in response to visual stimulation during
- 27 experience-dependent visual development (age p20-p27) (Cheadle et al., 2018).
- 28 Using electrophysiology and ultrastructural methods, they showed that the
- 29 strengthening of connections between retinal ganglion cells (RGC) and
- 30 thalamocortical neurons required Fn14. Synapses were not strengthened as
- 31 expected either in Fn14 knockout (KO) mice or following sensory deprivation, which
- resulted in a reduction of Fn14 induction (Cheadle et al., 2018). Furthermore, the
- 33 absence of Fn14 through genetic manipulation or deprivation also reduced the

34 elimination of weak synapses that occurs during circuit refinement (Lin et al., 2014).

The effects of manipulating Fn14 were only present during the later period of visual 35

development (between p20 and p27), but not prior to p20. While this study 36

highlighted the important role of Fn14 in experience-dependent visual refinement, 37

- 38 how Fn14 induction differentially regulated synapse strengthening and elimination
- 39 remained unclear.
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In this issue, the authors build on their previous work to describe the mechanism 41 42 underpinning the opposing Fn14 effects on the retinogeniculate connections (Cheadle, 2020). They used a novel form of semi-automated serial transmission 43 electron microscopy (TEM) to examine the structural properties of synapses during 44 this period of experience-dependent visual development. Consistent with their 45 previous work (Cheadle et al., 2018), the authors found that synaptic strengthening 46 47 and elimination is dependent on Fn14. In the absence of Fn14, the total percentage of bulbous spines (mushroom-like spines with a spine head that is at least twice as 48 49 wide as the neck) receiving RGC inputs decreased from 60% to 30%. Furthermore, in wildtype mice, bulbous spines on average have four synapses per spine, 50 51 compared to one to two synapses per spine in thin and non-bulbous spines. In mice where Fn14 was eliminated via genetic manipulation or sensory deprivation, the 52 53 bulbous spines had 40% fewer synapses per spine. This Fn14-dependent effect was largely confined to bulbous spines, with little effect on the number of synapses in thin 54 55 or non-bulbous spines. Together, this decrease in both the number of bulbous spines 56 and the number of synapses on each bulbous spine in the absence of Fn14 57 corresponds to a strong decrease in the total RGC input to these neurons when 58 Fn14 function is disrupted. Additionally, the authors developed an Fn14 conditional 59 KO mouse, which allowed them to spatially localize the effect of Fn14. They found that postsynaptic expression of Fn14 combined with sensory experience was 60 necessary for the development of bulbous spines, with limited contribution from 61 presynaptic effects. 62

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64 Having characterized the effects of Fn14 on dendritic spines, the authors examined the molecular mechanisms facilitating this effect. The cytokine tumor necrosis factor 65 (TNF)-like weak inducer of apoptosis (TWEAK), is a member of the TNF superfamily 66 and binds to Fn14 (Wiley and Winkles, 2003). The authors first showed that TWEAK 67

68 is expressed in a subset of microglia in the thalamus in an experience-dependent

manner. Seventy percent of microglia express TWEAK after visual stimulation, 69

70 whereas only 8% express TWEAK in unstimulated mice. Furthermore, TWEAK was

shown to bind in a highly localized way to retinogeniculate synapses expressing 71

72 Fn14. In the absence of Fn14, TWEAK was no longer reliably detectable at

- 73 synapses.
- 74

75 The authors then examined the effect of the TWEAK-Fn14 interaction on spines.

76 They found that overexpression of soluble TWEAK via an adeno-associated virus

(AAV) injection resulted in a 25% decrease in the bulbous spine population. 77

78 Conversely, in TWEAK KO mice, the bulbous spine population was increased by

30%. The increase in spine percentage in the KO mice could be reduced to wildtype 79

levels through re-expression of TWEAK via viral injection. Together, these results 80

suggest that TWEAK acts locally through Fn14 to facilitate the elimination of bulbous 81

82 spines.

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84 The authors next examined how TWEAK-expressing microglia may facilitate synaptic 85 elimination. Previous work has described mechanisms by which microglia are involved in synaptic pruning, with phagocytic engulfment as one of the best-studied 86 87 mechanisms of microglial-synaptic interactions (Schafer et al., 2013). The authors 88 examined if the TWEAK-mediated elimination of synapses occurred through 89 phagocytic engulfment. They found that while presynaptic components of the 90 synapse did undergo phagocytic engulfment, postsynaptic spines did not. These 91 results suggest that TWEAK acts through a non-phagocytic mechanism. Additionally, 92 there was no difference in phagocytic engulfment in wildtype or TWEAK KO mice, 93 either during early (P7) or experience-dependent development (p27), suggesting that 94 TWEAK is not involved in phagocytic engulfment. Together these results point to microglia acting through a novel experience-dependent mechanism. 95 96

97 Postsynaptic Fn14 is necessary for synapse strengthening and, through interactions

with TWEAK, elimination. The authors next examined how the TWEAK-Fn14 98

interaction targets particular synapses for elimination. They found a spatial 99

100 organization of spines that are strengthened or eliminated. Spines that were in the

101 vicinity of microglia expressing TWEAK were eliminated. Specifically, within 200 nm 102 of a given microglial cell, there was an inverse correlation between TWEAK

103 expression levels and the number of bulbous spines. These results suggest that

there is a spatial element to synapse elimination. Whether additional genetic or

105 experience-dependent influences determine the position of TWEAK expressing

106 microglia with respect to the dendritic spines will require further study.

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108 In summary, Cheadle et al. (2020) show that microglia play a novel role in shaping synaptic organization during the experience-dependent phase of retinogeniculate 109 110 circuit refinement. In the early phase of visual development (prior to p20), synaptic changes occur that are independent of Fn14-TWEAK signalling. After p20, there is 111 112 an experience-dependent expression of Fn14 in the thalamic relay neurons and TWEAK in microglia. In this later phase, microglia employ an experience-dependent 113 114 process to sculpt postsynaptic spine morphology and pruning ((Cheadle, 2020) 115 Figure 8).

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117 While the results presented here occur during late visual development, many immune-related molecules have ongoing roles in plasticity throughout the brain 118 119 (Fourgeaud and Boulanger, 2010). Thus, the plasticity mechanism described here 120 may generalize to other developmental periods and brain regions. In line with this, 121 TNF-alpha, which is also from the TNF superfamily and released locally from microglia, is known to be critical for homeostatic synaptic strengthening throughout 122 123 development and adulthood (Barnes et al., 2017; Pribiag and Stellwagen, 2014). The degree to which the experience-dependent plasticity mechanisms described here 124 125 reflect homeostatic or Hebbian processes remains to be seen, but the work adds to 126 the literature suggesting that this class of immune-related molecules may have an 127 important role in plasticity. Overall, the work from Cheadle et al. suggests a novel role for microglia in shaping the synaptic connectivity and organization of neural 128 circuits, providing further evidence for the importance of this cell type in the 129 130 development and plasticity of neural circuitry.

131 References

Barnes, S.J., Franzoni, E., Jacobsen, R.I., Erdelyi, F., Szabo, G., Clopath, C., Keller, 132 133 G.B., and Keck, T. (2017). Deprivation-Induced Homeostatic Spine Scaling In Vivo Is Localized to Dendritic Branches that Have Undergone Recent Spine Loss. Neuron 134 135 96, 871-882 e875. 136 137 Cheadle, L., Tzeng, C.P., Kalish, B.T., Harmin, D.A., Rivera, S., Ling, E., Nagy, M.A., Hrvatin, S., Hu, L., Stroud, H., et al. (2018). Visual Experience-Dependent 138 139 Expression of Fn14 Is Required for Retinogeniculate Refinement. Neuron 99, 525-140 539 e510. 141 142 Cheadle, L., Rivera, S.A., Phelps, J.S., Ennis, K.A., Stevens, B., Burkly, L.C., Lee, W.C. and Greenberg, M.E. (2020). Sensory Experience Engages Microglia to Shape 143 Neural Connectivity through a Non-Phagocytic Mechanism. Neuron 144 145 https://doi.org/10.1016/j.neuron.2020.08.002. 146 147 Fourgeaud, L., and Boulanger, L.M. (2010). Role of immune molecules in the establishment and plasticity of glutamatergic synapses. Eur J Neurosci 32, 207-217. 148 149 Hooks, B.M., and Chen, C. (2006). Distinct roles for spontaneous and visual activity 150 in remodeling of the retinogeniculate synapse. Neuron 52, 281-291. 151 152 Huberman, A.D. (2007). Mechanisms of eye-specific visual circuit development. Curr 153 Opin Neurobiol 17, 73-80. 154 155 Lin, D.J., Kang, E., and Chen, C. (2014). Changes in input strength and number are 156 157 driven by distinct mechanisms at the retinogeniculate synapse. J Neurophysiol 112. 158 942-950. 159 160 Pribiag, H., and Stellwagen, D. (2014). Neuroimmune regulation of homeostatic 161 synaptic plasticity. Neuropharmacology 78, 13-22. 162 Schafer, D.P., Lehrman, E.K., and Stevens, B. (2013). The "guad-partite" synapse: 163 164 microglia-synapse interactions in the developing and mature CNS. Glia 61, 24-36. 165 Wiley, S.R., and Winkles, J.A. (2003). TWEAK, a member of the TNF superfamily, is 166 a multifunctional cytokine that binds the TweakR/Fn14 receptor. Cytokine Growth 167 Factor Rev 14, 241-249. 168 169

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