BRAIN COMMUNICATIONS

SCIENTIFIC COMMENTARY

In the blood: biomarkers for amyloid pathology and neurodegeneration in Alzheimer's disease

This scientific commentary refers to 'Plasma total-tau, neurofilament light chain and amyloid- β levels and risk of dementia: a population-based study' by de Wolf *et al.* (https://doi.org/10.1093/brain/awaa054), and 'Relationship of amyloid-b1–42 in blood and brain amyloid: Ginkgo Evaluation of Memory Study' by Lopez *et al.* (https://doi.org/10.1093/braincomms/fcz038), two papers that illustrate these latest developments.

Introduction

Among the most impactful and fast developing aspects in neurodegeneration research and clinical practice over the last 30 years have been the development of biomarkers. A biomarker is a measurable indicator of a biological pathological condition state or (Zetterberg, 2019). Frustratingly, the field finds itself in tension between the increasing power and role of biomarkers to detect and predict the pathologies that underlie dementia versus stagnation in prophylactic, therapeutic or reparative interventions. Reduction in the cost of trials by cheap, accurate and accessible pre-screening and better understanding of how biomarkers relate to brain pathology are important to resolve this problem. Here, we discuss the role blood-based biomarkers can play in the context of two recent papers by de Wolf et al. (2020) and Lopez et al. (2020).

Blood biomarkers in context

Both papers have emerged in the context of a burgeoning paradigm shift in the potential of fluid biomarkers. Over the past decade, cerebrospinal fluid (CSF) and positron emission tomography (PET) biomarkers have dominated neurodegeneration research and guided drug design. CSF amyloid beta 1-42 (A β 42), total tau (T-tau) and phosphorylated tau₁₈₁ (P-tau₁₈₁) and ¹¹C Pittsburgh Compound B, florbetapir and florbetaben PET for AB pathology are now well validated for Alzheimer's disease with 85-95% sensitivity and specificity (Zetterberg and Bendlin, 2020). Important recent developments are the introduction of certified reference methods and materials for Aβ42, and second-generation tau PET. However, collection of CSF is sometimes regarded as a minor surgical procedure, requiring specialist training, and brain imaging techniques are costly, also require specialist training, and employ radioactive tracers in the case of PET. These issues limit scalable testing and who can access them.

Blood-based biomarkers have the potential to circumvent or diminish many of these limitations. Phlebotomy is a comparatively cheap, routine and un-invasive procedure, and so fluid biomarker analysis from blood is highly scalable. Much scepticism surrounded early data from blood because of poor reproducibility (reasons portrayed in Fig. 1). Recent improvements in instrument sensitivity have rapidly begun to change the state of play. Sub-femtomolar concentration detection afforded by single molecule array (Simoa) technology, and improvements in immunoprecipitation mass spectrometry platforms enable accurate, consistent and high sensitivity measurement even after the extensive dilution of plasma and serum.

Amyloid beta

Of candidate biomarkers in plasma, A β 42 and A β 40 have been the most extensively studied (Olsson *et al.*, 2016). Many early studies showed no change or even increases in blood A β in Alzheimer's disease versus control samples (Olsson *et al.*, 2016); however, recent work using high sensitivity assays have begun to show the expected decrease and correlation between CSF and plasma A β 42/40 ratios (Janelidze *et al.*, 2016; Ovod *et al.*, 2017; Nakamura *et al.*, 2018).

The work of de Wolf *et al.* contributes one of the largest longitudinal analyses of plasma A β 42 and A β 40 to date. Measured by Simoa, baseline A β 42 and A β 42/40 levels, but not A β 40, were predictive of conversion to dementia. Hazard ratios (HR) showed that low plasma A β 42 was significantly associated with conversion to all-cause dementia and Alzheimer's disease.

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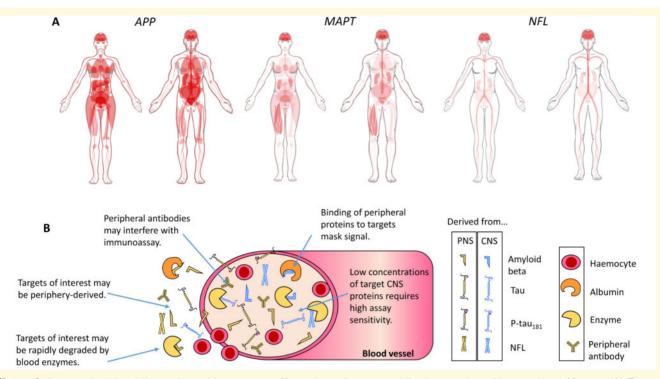


Figure 1 Caveats for blood biomarkers. Measurement of brain-derived proteins in blood is complicated by a number of factors. (**A**) Tissue expression of genes that encode proteins relevant to AD. Proteolysis of amyloid precursor protein (APP), encoded by the *APP* gene, generates A β . Microtubule-associated protein tau (*MAPT*) encodes tau protein. Neurofilament light chain (NFL) encodes the protein of the same name. Diagrams for APP (www.proteinatlas.org/ENSG00000142192-APP/tissue), MAPT (www.proteinatlas.org/ENSG00000186868-MAPT/tissue), and NFL (www.proteinatlas.org/ENSG00000277586-NEFL/tissue) generated by Human Protein Atlas (Uhlen *et al.*, 2015). Red indicates gene expression. From the expression patterns, it is clear that NFL is the most neuronal-specific of the biomarkers, which may explain why its plasma concentration associates the best with neurodegeneration. (**B**) A simplistic diagram of brain biomarkers in blood, highlighting the key issues for accurate detection.

Dividing the dataset into quartile groups delineated the relationship between lower A β 42 and increased risk further, and showed that the association was stable over time.

Trajectory analysis demonstrated that over 13 years AB42 declined at similar rates in both Alzheimer's disease and non-Alzheimer's disease converters. AB42/40 was significantly altered over time in Alzheimer's disease converters, but the effect size was negligible and driven by AB40. Similarly, Lopez et al. found that changes in Aβ42 over an eight year period were marginal and unrelated to brain amyloid deposition. The mean age in both studies was relatively advanced [71.9 years (de Wolf et al., 2020) and 85 years (Lopez et al., 2020)], and it may be that samples

were taken too late to catch longitudinal intra-individual AB changes. It must also be noted that Lopez et al. excluded dementia converters from their analysis. Nevertheless, this raises the important question of what AB measured from blood means biologically. Plasma A β is influenced by a number of factors (Fig. 1). If plasma Aβ concentrations are periphery driven, whether or not they are predictive of dementia, it would be difficult to use them to make inferences about, or therapeutically target, processes occurring in the brain. It would also raise questions about the extent to which dementia arises from CNS pathology in relative isolation.

Results from Lopez *et al.* showed that ¹¹C-PiB PET positivity (measured in 2009) correlated significantly with

decreased plasma Aβ42 (measured by enzyme-linked immunosorbent assay) in individuals whose blood was collected in the 2000-02 group of their study. All brain regions studied, particularly the anterior cingulate gyrus and frontal cortex, were significant contributors. However, the story was not straight-forward and data from the 2008-09 group only showed a non-significant tendency for correlation of plasma AB with brain amyloid deposition, with an inconsistent pattern of regional contribution. Other studies have identified correlation between plasma AB42/40 and amyloid PET-based deposition (Janelidze et al., 2016; Nakamura et al., 2018), as well as CSF concentrations (Janelidze et al., 2016). Thus, whilst likely brain reservoir

contributes to plasma $A\beta 42$ concentrations, the relationship of plasma $A\beta$ to neurodegenerative pathology remains in question, especially if not measured by the latest ultra-sensitive techniques.

Neurofilament light chain

Plasma/serum concentrations of NFL correlate well with CSF (Zetterberg et al., 2016) and have been among the most consistent blood markers for allcause neurodegeneration (Mattsson et al., 2019; Janelidze et al., 2020; Thijssen et al., 2020). Data presented by de Wolf et al. add more positive data to this story. Baseline plasma NFL concentration was strongly predictive of conversion to all-cause dementia, and particularly Alzheimer's and disease vascular dementia. Quartile analysis highlighted this pattern further and when the highest and lowest quartiles of AB42 and NFL concentration were combined and compared, HRs increased substantially (all-dementia HR: 9.5, Alzheimer's disease HR: 15.7). Of particular interest, cumulative incidence analysis showed that plasma NFL concentration increased 3.4 times faster in participants who developed Alzheimer's disease versus no dementia. These changes were detectable 9.6 years before Alzheimer's disease diagnosis.

Results from a single biomarker test must be interpreted in relation to cross-sectionally determined cut-points from sample populations, which may or may not have high relevance to the individual. Multiple measurements taken longitudinally from the same individual would provide an internal reference point and thus circumvent such issues. If pre-symptomatic disease-associated changes in a blood-based biomarker, such as NFL, could be detected over a clinically relevant time interval this could be a powerful tool. A decade may not be such a time frame, however some data suggest that plasma NFL changes may track certain aspects of underlying pathology over 15-30 months (Mielke et al., 2019).

Lopez et al. did not analyse the relationship between NFL and brain pathology, but other recent studies have addressed this question. Despite finding elevated plasma NFL in cortical-basal syndrome, progressive supranuclear palsy, behavioural variant frontotemporal dementia and Alzheimer's disease compared to healthy controls, Thijssen et al. (2020) found that plasma NFL was not related to either tau PET (flortaucipir) or Aß PET (PiB/florbetapir). Mielke et al. observed a similar relationship between baseline NFL concentration, PiB PET and fluorodeoxyglucose (FDG)-PET (a biomarker for synaptic dysfunction/degeneration) (Mielke et al., 2019). However, longitudinal elevation of plasma NFL were associated declines in hippocampal volume, cortical thickness, FDG-PET, corpus callosum fractional anisotropy and global cognitive z scores, as well as with increasing amyloid PET positivity (Mielke et al., 2019). In summary, it is not yet clear exactly how well peripherally measured NFL reflects CNS neurodegeneration (see also Fig. 1), but indications are that brain reservoir has a net contribution over time.

Tau

Recent tau kinetics data suggest that Alzheimer's disease-related increase of CSF T-tau and P-tau may be a neuronal response to AB pathology (Sato et al., 2018), rather than a direct reflection of neurodegeneration and tangle pathology. Despite this, de Wolf et al. found plasma T-tau to be unchanged in Alzheimer's disease versus cognitively normal individuals, consistent with the observations of Verberk et al. (2018), but in contrast to others (Mattsson et al., 2016; Olsson et al., 2016; Mielke et al., 2018; Pase et al., 2019). Such conflicting evidence likely stem from the poor correlation of plasma T-tau with CSF T-tau (Mattsson et al., 2016), and factors portrayed in Fig. 1.

A notable gap in the work of de Wolf and Lopez *et al.* is that of plasma P-tau₁₈₁. CSF P-tau₁₈₁ is a core Alzheimer's disease-specific biomarker, found to be increased relatively early in the disease course, potentially as part of a neuronal response to AB pathology. In four recent studies, baseline plasma P-tau181 was significantly increased, associated specifically with conversion of cognitively normal individuals to Alzheimer's disease (Mielke et al., 2019; Janelidze et al., 2020; Karikari et al., 2020; Thijssen et al., 2020). Plasma P-tau₁₈₁ accurately discriminated Alzheimer's disease dementia from non- Alzheimer's disease neurodegenerative diseases with sensitivity and specificity similar to CSF AB42/AB40 and CSF T-tau, and slightly worse than CSF P-tau181 combined with Tau PET. As a point of comparison with NFL data from de Wolf et al., a longitudinal study found that baseline plasma P-tau₁₈₁ concentration was a significant predictor of Alzheimer's disease conversion (Janelidze et al., 2020). When data were thresholded based on a 1.81 pg ml^{-1} cut-point, associated risk increased markedly (HR = 10.9) for those with higher versus lower concentrations (Janelidze et al., 2020).

Plasma P-tau₁₈₁ correlates well with CSF P-tau₁₈₁, as well as both A β PET and tau PET (Mielke *et al.*, 2019; Thijssen *et al.*, 2020). Furthermore, plasma P-tau₁₈₁ has been shown to be significantly associated with Braak stage (Karikari *et al.*, 2020), whilst NFL is not (Thijssen *et al.*, 2020). Interestingly, a decreased ratio of Ptau₁₈₁/NFL was able to distinguish FTLD from Alzheimer's disease, likely as a result of non-A β driven (and therefore not involving P-tau₁₈₁) neuronal damage (Janelidze *et al.*, 2020).

Conclusion

The studies of de Wolf *et al.* and Lopez *et al.* have contributed important longitudinal data to the emerging picture of blood-based biomarkers for neurodegenerative disease. With further validation, blood biomarker analysis has the potential to synergise with polygenic risk score screening and open up a paradigm where one might go to a general practitioner, and receive multiple biomarker measurements monitored longitudinally. This could guide pre-emptive therapy aimed at managing concentrations relative to an individual's own normal.

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Competing interests

H.Z. has served at scientific advisory boards for Denali, Roche Diagnostics, Wave, Samumed and CogRx, has given lectures in symposia sponsored by Fujirebio, Alzecure and Biogen, and is a co-founder of Brain Biomarker Solutions in Gothenburg AB (BBS), which is a part of the GU Ventures Incubator Program. J.T. reports no disclosures.

References

- Janelidze S, Mattsson N, Palmqvist S, Smith R, Beach TG, Serrano GE, et al. Plasma Ptau181 in Alzheimer's disease: relationship to other biomarkers, differential diagnosis, neuropathology and longitudinal progression to Alzheimer's dementia. Nat Med 2020; 26: 379–86.
- Janelidze S, Stomrud E, Palmqvist S, Zetterberg H, Van Westen D, Jeromin A, et al. Plasma β-amyloid in Alzheimer's disease and vascular disease. Sci Rep 2016; 6: 26801.
- Karikari TK, Pascoal TA, Ashton NJ, Janelidze S, Benedet AL, Rodriguez JL, et al. Blood phosphorylated tau 181 as a biomarker for Alzheimer's disease: a modelling study using data from four prospective cohorts. Lancet Neurol 2020; 2: fcz038.
- Lopez OL, Klunk WE, Mathis CA, Snitz BE, Chang Y, Tracy RP, et al. Relationship of amyloid-β1–42 in blood and brain amyloid: Ginkgo Evaluation of Memory Study. Brain Commun 2020.
- Mattsson N, Cullen NC, Andreasson U, Zetterberg H, Blennow K. Association between longitudinal plasma neurofilament light and neurodegeneration in patients with Alzheimer Disease. JAMA Neurol 2019; 76: 791–9.
- Mattsson N, Zetterberg H, Janelidze S, Insel PS, Andreasson U, Stomrud E, et al. Plasma tau in Alzheimer disease. Neurology 2016; 87: 1827–35.
- Mielke MM, Hagen CE, Xu J, Chai X, Vemuri P, Lowe VJ, et al. Plasma phospho-tau181 increases with Alzheimer's disease clinical severity and is associated with tau- and amyloid-positron emission tomography. Alzheimer's Dement 2018; 14: 989–97.
- Mielke MM, Syrjanen JA, Blennow K, Zetterberg H, Vemuri P, Skoog I, et al. Plasma and CSF neurofilament light. Neurology 2019; 93: e252–e260.
- Nakamura A, Kaneko N, Villemagne VL, Kato T, Doecke J, Doré V, et al. High

performance plasma amyloid-β biomarkers for Alzheimer's disease. Nature 2018; 554: 249–54.

- Olsson B, Lautner R, Andreasson U, Öhrfelt A, Portelius E, Bjerke M, et al. CSF and blood biomarkers for the diagnosis of Alzheimer's disease: a systematic review and meta-analysis. Lancet Neurol 2016; 15: 673–84.
- Ovod V, Ramsey KN, Mawuenyega KG, Bollinger JG, Hicks T, Schneider T, et al. Amyloid β concentrations and stable isotope labeling kinetics of human plasma specific to central nervous system amyloidosis. Alzheimer's Dement 2017; 13: 841–9.
- Pase MP, Beiser AS, Himali JJ, Satizabal CL, Aparicio HJ, Decarli C, et al. Assessment of plasma total tau level as a predictive biomarker for dementia and related endophenotypes. JAMA Neurol 2019; 76: 598–606.
- Sato C, Barthélemy NR, Mawuenyega KG, Patterson BW, Gordon BA, Jockel-Balsarotti J, et al. Tau kinetics in neurons and the human central nervous system. Neuron 2018; 97: 1284–98.e7.
- Thijssen EH, La Joie R, Wolf A, Strom A, Wang P, Iaccarino L, et al.; Advancing Research and Treatment for Frontotemporal Lobar Degeneration (ARTFL) investigators. Diagnostic value of plasma phosphorylated tau181 in Alzheimer's disease and frontotemporal lobar degeneration. Nat Med 2020; 26: 387-97.
- Uhlen M, Fagerberg L, Hallstrom BM, Lindskog C, Oksvold P, Mardinoglu A, et al. Tissue-based map of the human proteome. Science 2015; 347: 1260419. doi: 10.1126/science.1260419.
- Verberk IMW, Slot RE, Verfaillie SCJ, Heijst H, Prins ND, van Berckel BNM, et al. Plasma amyloid as prescreener for the earliest Alzheimer pathological changes. Ann Neurol 2018; 84: 648–58.
- de Wolf F, Ghanbari M, Licher S, McRae-McKee K, Gras L, Weverling GJ, et al. Plasma tau, neurofilament light chain and amyloid-β levels and risk of dementia; a population-based cohort study. Brain 2020; 143: 1220–32.
- Zetterberg H. Blood-based biomarkers for Alzheimer's disease—an update. J Neurosci Methods 2019; 319: 2–6.
- Zetterberg H, Bendlin BB. Biomarkers for Alzheimer's disease—preparing for a new era of disease-modifying therapies. Mol Psychiatry 2020; 1–13.
- Zetterberg H, Skillbäck T, Mattsson N, Trojanowski JQ, Portelius E, Shaw LM, et al.; Alzheimer's Disease Neuroimaging Initiative. Association of cerebrospinal fluid neurofilament light concentration with Alzheimer disease progression. JAMA Neurol 2016; 73: 60–7.