Research paper

Molecular mechanisms underlying altered neurobehavioural development of female offspring of mothers with polycystic ovary syndrome: FOS-mediated regulation of neurotrophins in placenta

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ABSTRACT

Background: This study explored the mechanisms underlying altered neurobehavioural development of female offspring born to mothers with polycystic ovary syndrome (PCOS).

Methods: In total, 20 women with PCOS and 32 healthy women who underwent caesarean deliveries with a single female foetus were recruited. Infants were assessed with Dubowitz scoring. Swan71 cell line with stable FOS overexpression was used to verify the regulatory effects of FOS on brain-derived neurotrophic factor (BDNF) and nerve growth factor (NGF) expression. Learning and memory in female first-generation (F1) and second-generation (F2) offspring in a rat model of PCOS was tested using the Morris water maze at puberty and adulthood. Transcriptome analysis of pubertal hippocampi and hypothalami of female F1 offspring was conducted.

Findings: Total score and behaviour subscales of Dubowitz scoring were significantly lower in female infants of women with PCOS. FOS and NGF protein levels were downregulated in placental villi of the PCOS group. FOS played a key role in BDNF inhibition and enhancing NGF in Swan71 cells. PCOS female F1 rats exhibited lower target crossing times during puberty when compared to controls. Transcriptome analysis revealed significant changes in hippocampal and hypothalamic neuronal pathways in female F1 rats at puberty.

Interpretation: FOS regulation of neurotrophins in the placenta negatively affects neurobehavioural development of female offspring of PCOS mothers.

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1. Introduction

Polycystic ovarian syndrome (PCOS) is characterised by hyperandrogenism, irregular ovulation, and polycystic ovaries. It is the most common endocrine disorder observed in women of reproductive age. This heterogeneous condition affects multiple aspects of women’s health including reproductive, metabolic, and psychological functions [1]. Moreover, maternal PCOS can alter the foetal endocrine environment, leading to adverse outcomes in offspring [2–4], which are more closely associated with maternal PCOS rather than pregnancy complications [5]. Offspring of PCOS mothers exhibit altered cardio-metabolic health relative to that of controls; these differences are predominantly observed in female offspring aged between 1 and 18 years [6]. Female offspring of women with PCOS have a higher risk for developmental delay [7]. PCOS-exposed offspring also have an increased risk of neurobehavioural disorders, involving attention-deficit/hyperactivity disorder (ADHD), autism spectrum disorders (ASD), Tourette’s disorder, chronic tic disorders (TD/CTD). In particular, the risk of ADHD and ASD is higher in female offspring born to PCOS women [8–11].

FOS belongs to the activator protein 1 (AP-1) superfamily of transcription factors and is responsible for diverse cellular processes, including proliferation, differentiation, apoptosis, hypoxia, angiogenesis, and steroidogenesis,[12, 13], which has been confirmed in
Research in context

Evidence before this study

As the most common endocrine disorder observed in women of reproductive age, polycystic ovarian syndrome (PCOS) can affect multiple aspects of women’s health and even alter the foetal endocrine environment, leading to adverse outcomes in offspring including altered cardiometabolic health as well as higher risk for developmental delay and neurobehavioural disorders. FOS, as a marker of stimulation-related neural activation, belongs to the activator protein 1 superfamily of transcription factors. Brain-derived neurotrophic factor (BDNF) and nerve growth factor (NGF) are essential for brain development and plasticity, which have been widely implicated in neurobehavioural disorders. BDNF and NGF in umbilical cord blood, placenta and amniotic fluid may have impact on foetal neurodevelopment. Despite the well-established association between maternal PCOS and the altered neurobehavioural development in female offspring of women with PCOS, the underlying mechanisms remain obscure.

Added value of this study

The total score and behaviour subscales of Dubowitz scoring were significantly lower in female infants of women with PCOS. We found negative alternations of proliferation and apoptosis of trophoblasts, vascularity of villi, endocrine functions and nutrient transfer in placenta of women with PCOS. In trophoblasts of PCOS group, the protein levels of FOS and NGF were downregulated, while BDNF unchanged. A cluster of neural function related genes was identified with integrative analysis of methylome and transcriptome. With respect to neural function, the enriched Gene Ontology (GO) terms in differential expressed gene cluster of PCOS group involved brain development, the proliferation and differentiation of neural precursor cell, neuron and neuroblast. Consistently, the enriched GO terms in differential methylated gene cluster of PCOS group were associated with neuropeptide and synaptic transmission, as well as cognition and learning or memory. We found that FOS gene probably plays a core role and predicted the potential sites in neurotrophin genes of FOS transcriptional modification. FOS could inhibit the mRNA expression of BDNF and enhance the mRNA expression of NGF in SWAN71 cells. In Morris water maze test of PCOS rat model, the target crossing times of female F1 offspring in puberty were greatly lower than the controls, however, no significant difference existed in adulthood, neither in puberty nor in adulthood of female F2 offspring. The transcriptome analysis showed that there were significant changes in the neuronal life processes and pathways in the puberty hippocampus and hypothalamus of the female F1 offspring of PCOS model rats.

Implications of all the available evidence

Our study demonstrates that FOS-mediated regulation of neurotrophins in the placenta may underpin the negatively effects of maternal PCOS on neurobehavioural development of female offspring.

trophoblasts [14]. FOS is a marker of stimulation-related neural activation [15]. As a switch that converts short-term stimuli into long-term responses, FOS activity in the brain is closely associated with learning, memory, hyperactivity, locomotion, anxiety-related behaviours, and impaired social behaviour [16–18]. In PCOS, FOS is decreased in adipose tissue and the hypothalamus, which may be associated with insulin resistance and neuroendocrine dysfunction [19, 20]. As FOS protein in ovarian cells regulates androgen production, impaired FOS activity may underpin the pathogenesis of hyperandrogenism in PCOS [21, 22].

Brain-derived neurotrophic factor (BDNF) and nerve growth factor (NGF) are essential for brain development and plasticity, and have been widely implicated in neurobehavioural disorders [23, 24]. BDNF levels in umbilical cord blood reflect the degree of neural maturity in infants [25]. Reduced placental BDNF negatively affects foetal neurodevelopment, as placental BDNF enters the foetal brain to contribute to its development [26]. As a possible source of neurotrophins to the foetus, the placenta plays a more prominent role than maternal passage and endogenous foetal synthesis [25, 27–31]. BDNF and NGF are expressed in the placenta and serve diverse roles in differentiation, proliferation, survival, angiogenesis, and transplacental nutrient transport, thus impacting on the development of the materno-foetal-placental unit [32–38]. BDNF has been implicated in “developmental programming” via the regulation of metabolism and energy balance from early life to adulthood [39]. BDNF and NGF in amniotic fluid are markers for the presence of foetal central nervous system abnormalities in uterus [40]. With regards to PCOS, neurotrophins in peripheral blood or foetalic fluid may affect follicular and embryonic development [41, 42]. Despite the well-established association between maternal PCOS and altered neurobehavioural development in female offspring of women with PCOS, the underlying mechanisms remain obscure. The present study aimed to address this gap in knowledge using RNA sequencing (RNA-seq), reduced representation bisulphite sequencing (RRBS), and subsequent bioinformatics analysis, involving Gene Ontology (GO), Kyoto Encyclopedia of Genes and Genomes (KEGG) enrichment, protein-protein interaction (PPI) network, and in silico analysis.

2. Methods

2.1. Subjects and sampling

In total, 20 women with PCOS and 32 healthy control women who underwent caesarean deliveries with a single female foetus in Women's Hospital, School of Medicine, Zhejiang University between June 2016 and June 2017 were recruited. Patients were diagnosed with PCOS according to the Rotterdam Consensus (European Society for Human Reproduction and Embryology/American Society for Reproductive Medicine criteria) [43]. The control women had regular menstrual cycles and normal sex hormone levels prior to pregnancy. Uteri and ovaries of all women were structurally normal. Exclusion criteria were other disorders with a similar clinical presentation, such as congenital adrenal hyperplasias, (ovarian or adrenal) androgen-secreting tumours, Cushing’s syndrome, thyroid dysfunction, hyperprolactinemia, primary premature ovarian insufficiency, premature ovarian failure, and/or functional hypothalamic amenorrhea. The study was approved by the Ethics Committee of Women’s Hospital, School of Medicine, Zhejiang University. Peripheral blood, umbilical cord blood, and placenta were collected during caesarean section. Anthropometric parameters including age, cycle length, pregestational body mass index (BMI), BMI at delivery, and foetal digit ratio of the right hand were measured using questionnaires and standard equipment. Serum biochemical indexes including total testosterone (TT) (80979, 0.05 ng/mL, Crystal Chem Inc, USA), sex hormone-binding globulin (SHBG) (RAB0734, 1.2 pmol/L, Sigma-Aldrich, USA), BDNF (RAB0026, 80 pg/mL, Sigma-Aldrich, USA), NGF (CSB-E04683h, 6.86pg/mL, Cusabio, USA), interleukin 6 (IL6) (BMS213-2, 0.92 pg/mL, Invitrogen, USA), and interleukin 15 (IL15) (BMS2106, 3.4 pg/mL, Invitrogen, USA) were measured with enzyme-linked immunosorbent assay (ELISA) in maternal peripheral blood and umbilical cord
blood. Free androgen index (FAI) was calculated as TT (nmol/l) divided by SHBG (nmol/l) × 100.

2.2. Neurological examination of the infants

Infants were examined using the Dubowitz neurologic examination method on the day after birth [44]. Trained researchers performed the neurological examinations, and results were recorded. In total, 34 items were classified into six categories, including tone, tone patterns, reflexes, movements, abnormal signs, and behaviour. Written informed consent was obtained from all women included in the present study.

2.3. Human tissue collection

After obtaining informed consent, the term placentas (38-40 weeks of gestation) from non-labour women who underwent elective caesarean section were collected. Placental tissue and villous biopsies were gently separated from the foetal side as described previously [45]. Samples of placental tissue were collected and stored at −80°C after flash freezing in liquid nitrogen. Samples of placental tissue were fixed in 4% paraformaldehyde for haematoxylin and eosin (H&E) staining and immunofluorescence.

2.4. Immunofluorescence

Placental villi were immunostained with primary antibodies against BDNF (ab205067, Abcam, UK, RRID: AB_205067), NGF (ab52918, Abcam, UK, RRID: AB_502154), FOS (GTX25794, GeneTex, USA, RRID: AB_369382), beta-humna chorionic gonadotropin (β-hCG) (11615-1-AP, Proteintech, China), glucose transporter 1 (GLUT-1) (1:200, 21829-1-AP, Proteintech, China RRID: AB_10837075), and glucose transporter 3 (GLUT-3) (20403-1-AP, Proteintech, China RRID: AB_10694437) using a standard protocol as previously described [46]. Cell nuclei were stained with 4',6-diamidino-2-phenylindole (DAPI) (32670-5MG-F, Merck, Germany). Staining was visualized with a laser-scanning confocal microscope at × 400 magnification (LSM 780, Zeiss, Germany). The integrated optical density (IOD) and area of trophoblasts were quantified using Image-pro plus 6.0 (Media Cybernetics, Inc., Rockville, MD, USA).

2.5. Cell proliferation and apoptosis detection

Ki67 staining and terminal deoxynucleotidyl transferase dUTP nick end labelling (TUNEL) assay were performed to evaluate proliferation and apoptosis using an anti-Ki67 primary antibody (GB14102, Servicebio, China) and fluorescein-based cell death detection kit (11684795910, Roche, Switzerland). The number of Ki67-positive cells, TUNEL-positive cells, total number of DAPI-stained cells, and IOD were measured with Image-pro plus 6.0 (Media Cybernetics, Inc., Rockville, MD, USA). The proliferation and apoptotic indexes were calculated as the percentage of Ki67-positive and TUNEL-positive nuclei divided by the total number of DAPI-stained cells in each section, respectively.

2.6. RNA extraction and real-time PCR

Vascular endothelial growth factor (VEGF), BDNF, NGF, FOS, Cyclin Dependent Kinase Inhibitor 1A (CDKN1A), Cyclin A2 (CCNA2), lin-7 homolog A (LIN7A), microtubule associated protein 1B (MAP1B), and transcription factor 4 (TCF4) mRNA expression levels in the foetal side of placenta from women with PCOS and controls were analysed with real-time polymerase chain reaction (real-time PCR). Isolation of total RNA was performed using Trizol Reagent (15596026, Invitrogen, USA), and cDNA was synthesised using the RevertAid First Strand cDNA Synthesis Kit (K1621, Thermo Fisher Scientific, USA) according to the manufacturer’s instructions. Real-time PCR was performed using the Step One Plus Real-Time PCR System (4376600, Applied Biosystems, USA). The relative quantification of gene expression was performed using the comparative ΔΔCt method. The primer sequences used for RT-PCR are listed in Table S1.

2.7. Bisulphite sequencing PCR

A methylation assay was performed at the promoter region of BDNF, NGF, and FOS genes in the foetal side of placenta from PCOS and control women using the bisulphite genomic sequencing (BSP) method as previously described [47]. Total genomic DNA was extracted using the TIANamp Genomic DNA Kit (DP304-02, Tiangen Biotech, China). Bisulphite treatment was conducted using EZ DNA Methylation-Gold Kit (DS005, ZYMO Research, USA). The methylation status of cytosine phosphate guanine (CpG) sites located on gene promoters were analysed by cloning and sequencing bisulphite-treated DNA. The modified DNA was amplified by PCR, and the products were purified and cloned into a pMD 19-T vector (3271, Takara, Japan). Ten colonies of each subject were randomly selected for the plasmid DNA and sequenced with 3730 DNA Analyzer Polymers (3730S, Applied Biosystems, USA, RRID: SCR_018052).

2.8. RRBS

RRBS libraries of genomic DNA were prepared using TruSeq DNA LT Sample Prep Kit v2 (Illumina, USA). The libraries were quantified with Quant-IT PicoGreen dsDNA Assay Kit (P7589, Life technologies, USA). All libraries were sequenced with Illumina HiSeq 2500 platform. Clean reads were obtained by removing the reads containing sequencing adaptors and primers or low quality reads. Contaminated and short sequences were filtered. Differentially methylated promoters (DMPs) were obtained with a P-value threshold of 0.05 and two-fold change.

2.9. RNA-seq

Total RNA was extracted using Trizol reagent (15596026, Invitrogen, USA) according to the manufacturer’s instructions. RNA quantity and purity were measured with Bioanalyzer 2100 (G2939BA, Agilent, USA) and RNA 6000 Nano LabChip Kit (5065-4476, Agilent, USA). RNA-seq libraries were obtained using TruSeq RNA LT Sample Prep Kit v2 (Illumina, USA). Paired-end sequencing was performed on the Illumina HiSeq 2500 platform (Genery Biotech, China). The expression values were obtained by cuffnorm based on fragments per kilo-base of exon per million reads mapped (FPKM) value [48]. Differentially expressed genes (DEGs) were screened out based on the criteria of two-fold change and P-value less than 0.05 using DESeq2 [49]. The raw data of RRBS and RNA-seq have been deposited in NCBI’s Gene Expression Omnibus and are accessible through GEO Series accession number GSE154274 (https://www.ncbi.nlm.nih.gov/geo/query/acc.cgi?acc=GSE154274) [50].

2.10. Differential alternative splicing analysis

Replicate multivariate analysis of transcript splicing (rMATS) v3.2.5 was used to detect differential alternative splicing (DAS) genes [51]. Five alternative splicing events were obtained, including skipped exons, retained introns, alternative 5’ splice sites, alternative 3’ splice sites, and mutually exclusive exons.

2.11. GO, KEGG enrichment analysis, and PPI network analysis

For RNA-seq of placenta, significantly enriched GO terms were achieved with DEGs and DMP-related genes by TopGO at the significance level set (P < 0.05) [52]. PPI network analysis was performed
with DEGs using the Search Tool for the Retrieval of Interacting Genes (STRING) v.9.05 [53]. Cytoscape software was used to visualise these associations [54]. For RNA-seq of the hippocampus and hypothalamus, GO and KEGG enrichment analysis were conducted with DAS genes using KOABs v 2.0 software [55].

2.12. In silico analysis

Putative motifs and potential transcription factor binding sites in genomic DNA 5 kb upstream and downstream were analysed using Genomatix MatInspector (Genomatix Software GmbH, Germany).

2.13. Overexpression of FOS in stable clonal lines

The first-trimester human trophoblast cell line, Swan71, was originally provided by Professor Charles H Graham, Queen’s University, Kingston, Ontario, Canada and maintained in the Zhejiang Province Key Laboratory of Female Reproductive Health Research. Swan71 cells were cultured in Dulbecco’s Modified Eagle’s Medium/Ham’s Nutrient Mixture F-12 (DMEM/F-12) medium (11320033, Gibco, USA) supplemented with 10% foetal bovine serum (FBS) (10099141C, Gibco, USA) and 1% penicillin–streptomycin (15140148, Invitrogen, USA) at 37°C and 5% CO₂.

To establish stable cell lines that overexpressed FOS, we transfected Swan71 cells with the lentiviral gene expression vector of FOS (pLenti-EF1a-EGFP-P2A-CMV-FOS-3Flag, H13612) and a control vector (pLenti-EF1a-EGFP-P2A-CMV-MCS-3Flag, GL107) using polybrene (H9268, Sigma, USA). Swan71 cells were infected with lentiviral particles at a multiplicity of infection of 60. After infection for 72 hours, transfected cells were selected by culturing with puromycin (P8833, Sigma, USA) for 2 weeks.

RNA samples were isolated from Swan71-GL107 and Swan71-H13612 to validate BDNF and NGF expression levels using real-time PCR. FOS protein expression levels in Swan71 cells were detected with western blot using a standard protocol as previously described [56]. Monoclonal anti-flag M2 antibody (F18045, Sigma, USA, RRID: AB_262044), goat anti-mouse IgG (A0216, Beyotime, China, RRID: AB_2860575), GAPDH polyclonal antibody (AP0063, Bioworld, USA, RRID: AB_2651132), and goat anti-rabbit IgG (A0208, Beyotime, China) were used.

2.14. Establishment of PCOS rat model

A total of 20 neonatal female Sprague-Dawley rats were provided by the Laboratory Animal Research Center, Zhejiang Chinese Medical University, Hangzhou, China. Animals were randomly divided into PCOS model and control groups (n=10 per group). On postnatal day nine, a PCOS rat model was established as previously described [47, 57]. The study was performed according to the Care and Use of Laboratory Animals protocol of National Research Council of China, following the ARRIVE guidelines, and was approved by Zhejiang University Ethics Committee.

On postnatal day nine, a PCOS rat model was induced by subcutaneous injection of testosterone propionate (57-85-2, Solarbio, China) at a dose of 0.1 mg/0.004 mL olive oil per g of body weight [57]. The control group received olive oil only. Animals were raised and housed in a temperature-controlled room (25°C) under a 12-hour light/dark cycle at a constant humidity of 55% humidity, and were provided ad libitum access to food and water. At 8 weeks of age, oestrous cycles were determined by analysing the cell types in vaginal smears for 14 consecutive days [58]. In the PCOS model group, only rats with irregular vaginal smears were used for subsequent experiments. Oestrous cycle, serum level of FAI, and ovarian histology were assessed to validate the PCOS rat model [47]. All rats were sacrificed in the dioestrous stage. Serum samples were collected after blood centrifugation and stored for hormone assays. Ovaries were removed from sacrificed rats and fixed in 4% paraformaldehyde. After embedding in paraffin, 4-μm sections were mounted on glass slides and stained with H&E for morphological observation under light microscopy.

2.15. Offspring acquisition

Rats in the PCOS model and control groups were designated as the parental generation (F0). At 12 weeks of age, F0 dams were injected intraperitoneally with 20 IU pregnant mare serum (PMSG, Ningbo Second Hormone Factory, China) at 5:00 pm followed by 20 IU of human chorionic gonadotropin (HCG, Ningbo Second Hormone Factory) 48 hours later and immediately paired with proven male breeders. Vaginal sperm plugs were assessed the following morning [55]. Pups born to F0 dams were designated as the F1 generation. At 12 weeks of age, female F1 rats were used to generate F2 by breeding them with proven male breeders.

2.16. Learning and memory testing with the Morris water maze in female offspring

The Morris water maze test was performed at 5 weeks (puberty) and 13 weeks of age (adulthood) to evaluate learning and memory in female offspring. The test was conducted and recorded between 8:00 am and 2:00 pm under poor lighting conditions. The water maze apparatus consisted of a circular water tank 1.5 m in diameter and was filled to a depth of 60 cm with water (22 ± 2°C). The tank was divided into four virtual quadrants. A circular platform, 8 cm in diameter, was placed in one of the quadrants (target quadrant) 2 cm below the water surface. Rats were subjected to four training sessions per day for 2 days and a memory retention test (probe test) on the third day. In the training sessions, rats were placed at the same starting position in each quadrant and were allowed to swim freely to find the hidden platform (target) for 60 s. The time spent from being placed in the water to reaching the platform was recorded as latency to target. If the rat did not locate the platform within 60 s, it was guided to the platform and allowed to stay there for 10 s, and its latency to target was recorded as 60 s. On the third day of the probe test, the platform was removed, and rats were allowed to swim freely for 60 s. The latency and latency to target in the training sessions; distance, time, and entries in the target zone; and distance, time, and entries in the target quadrant during the probe test were recorded using a camera linked to a computerized video tracking system (SMART v3.0, Panlab SL, Spain).

2.17. Brain tissue collection

After completion of the Morris water maze test, female offspring were deeply anesthetised with sodium pentobarbital (50 mg/kg, Sigma, P3761, USA) in the dioestrus stage. The hippocampus and hypothalamus were collected and stored at −80°C after flash freezing in liquid nitrogen.

2.18. Electron microscopy

Samples were post-fixed in 2.5% glutaraldehyde for 2 hours at room temperature. After fixation, samples were rinsed with BP buffer, post-fixed with 2% osmium tetroxide for 1-2 hours, rinsed again with BP buffer, dehydrated in an ethanol series, infiltrated in a mixture of Spurr and acetone, and embedded in 100% Spurr embedding agent overnight and polymerised for 24 hours. The specimens were cut into blocks and serially sliced into 70-nm thick slices for transmission electron microscope (H-7650, Hitachi, Japan) analysis. Micrographs of neurons and synapses were obtained using a side-mounted CCD camera (SC200, Gatan, USA).
2.19. Statistical analysis

Data are presented as mean ± SEM (n indicates the number of tissue preparations, cells, or separate experiments, as indicated in the figure legends). Statistical analysis was performed using unpaired two-tailed Student’s t-tests, one-way analysis of variance with post-hoc tests, or their equivalent nonparametric tests for continuous variables. Chi-squared test or Fisher’s exact test was performed for categorical variables (version 21.0; SPSS). P < 0.05 was considered statistically significant.

3. Results

3.1. Clinical characteristics

Of 20 PCOS patients, 17 presented with all three diagnostic criteria items (oligo- or anovulation, clinical or biochemical signs of hyperandrogenism, and PCO morphology); two did not present with signs of hyperandrogenism, and one did not present with PCO morphology. No significant differences were observed in maternal age at delivery, pregestational BMI, and BMI at delivery between PCOS patients and controls, with the exception of a longer cycle length in PCOS patients (Table 1). Relative to serum FAI levels in the control group at delivery, serum FAI levels in maternal peripheral blood were higher in the PCOS group, but those in umbilical cord blood of the corresponding newborns of women with PCOS had comparable IL6 and IL15 levels compared to controls, women with PCOS had higher IL6 levels and comparable IL15 levels to controls, corroborating the data from umbilical cord blood. Comparatively, newborns of women with PCOS had higher IL6 levels and comparable IL15 levels to controls. Foetal digit ratios (2D:4D) of the right hand were similar in female newborns of women with PCOS compared to controls, women with PCOS had higher IL6 levels and comparable IL15 levels to controls, corroborating the data from umbilical cord blood. Com- pared to controls, women with PCOS had higher IL6 levels and comparable levels of IL15, BDNF, and NGF in peripheral blood. Female newborns of women with PCOS had comparable IL6 and IL15 levels but significantly lower levels of BDNF and NGF in umbilical cord blood compared to those of control women, which is in accordance with a previous study that reported that neural factors are unlikely to traffic between foetus and mother [60].

Table 1
Clinical characteristics of subjects.

<table>
<thead>
<tr>
<th>Characteristics</th>
<th>Control (n = 32)</th>
<th>PCOS (n = 20)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Pregestational BMI (kg/m²)</td>
<td>20.81 ± 0.32</td>
<td>21.44 ± 0.58</td>
</tr>
<tr>
<td>Cycle length (days)</td>
<td>29.16 ± 0.20</td>
<td>55.25 ± 3.56*</td>
</tr>
<tr>
<td>Maternal age at delivery (years)</td>
<td>31.78 ± 0.57</td>
<td>32.55 ± 0.89</td>
</tr>
<tr>
<td>BMI at delivery (kg/m²)</td>
<td>26.53 ± 0.51</td>
<td>27.18 ± 0.51</td>
</tr>
<tr>
<td>Serum FAI in maternal peripheral blood at delivery (nmol/L)</td>
<td>0.73 ± 0.06</td>
<td>1.34 ± 0.15*</td>
</tr>
<tr>
<td>Serum BDNF in maternal peripheral blood at delivery (ng/mL)</td>
<td>178.52 ± 10.53</td>
<td>173.02 ± 13.92</td>
</tr>
<tr>
<td>Serum NGF in maternal peripheral blood at delivery (pg/mL)</td>
<td>20.28 ± 1.24</td>
<td>20.58 ± 1.18</td>
</tr>
<tr>
<td>Serum IL-6 in maternal peripheral blood at delivery (pg/ml)</td>
<td>6.59 ± 0.58</td>
<td>9.25 ± 0.98*</td>
</tr>
<tr>
<td>Serum IL-15 in maternal peripheral blood at delivery (pg/ml)</td>
<td>16.25 ± 1.43</td>
<td>19.21 ± 2.20</td>
</tr>
<tr>
<td>Gestational age at delivery (weeks)</td>
<td>38.34 ± 0.10</td>
<td>38.45 ± 0.11</td>
</tr>
<tr>
<td>Newborn digit ratio of the right hand</td>
<td>0.93 ± 0.005</td>
<td>0.92 ± 0.005</td>
</tr>
<tr>
<td>Serum FAI in umbilical cord blood</td>
<td>1.33 ± 0.09</td>
<td>1.79 ± 0.21</td>
</tr>
<tr>
<td>Serum BDNF in umbilical cord blood (ng/mL)</td>
<td>137.64 ± 11.82</td>
<td>96.42 ± 7.18*</td>
</tr>
<tr>
<td>Serum NGF in umbilical cord blood (pg/mL)</td>
<td>13.91 ± 0.87</td>
<td>9.41 ± 0.65*</td>
</tr>
<tr>
<td>Serum IL-6 in umbilical cord blood (pg/ml)</td>
<td>7.24 ± 0.74</td>
<td>9.14 ± 0.95</td>
</tr>
<tr>
<td>Serum IL-15 in umbilical cord blood (pg/ml)</td>
<td>10.18 ± 1.16</td>
<td>9.87 ± 1.40</td>
</tr>
<tr>
<td>Birth weight (kg)</td>
<td>3.56 ± 0.07</td>
<td>3.37 ± 0.09</td>
</tr>
</tbody>
</table>

Note: PCOS, Polycystic ovarian syndrome; BMI, body mass index; FAI, free androgen index; BDNF, brain-derived neurotrophic factor; NGF, nerve growth factor; IL-6, interleukin 6; IL-15, interleukin 15. Data were presented as mean ± SEM. *P < 0.05.

3.2. Dubowitz neurological examination of female newborns

The total score and behaviour subscale of the Dubowitz scoring system were significantly lower in female infants of women with PCOS compared to those of controls. All remaining subscales (including tone, tone pattern, reflexes, movements, and abnormal signs) were comparable between these two populations (Fig. 1).

3.3. Altered placental function in PCOS patients

No significant difference in placental weight was noted between PCOS and control groups (Fig. 2a). The morphological features of placental villi in PCOS and control groups are shown in Fig. 2a. With regards to placental vascularisation, we observed substantially decreased vascularisation of villi and elevated levels of vascular endothelial growth factor (VEGF) in placental trophoblasts of PCOS women (Fig. 2c). Ki67 and TUNEL staining revealed suppressed cell proliferation and slightly enhanced apoptosis of trophoblasts in placental villi of the PCOS group (Fig. 2d). With regards to cell-cycle related gene expression, CCNA2 expression was significantly decreased and CDKN1A expression was increased in placenta of the PCOS group (Fig. S1). With regards to placental endocrine function and nutrient transfer in PCOS, reduced levels of β-HCG, GLUT1, and GLUT3 were observed in placenta of the PCOS group (Fig. 2e), suggesting attenuated hormone secretion and carrier-mediated glucose transport in pathological trophoblasts of PCOS.

To investigate the placental production of neurotrophins, including BDNF and NGF, real-time PCR and immunofluorescence were performed. In the PCOS group, NGF mRNA levels were significantly downregulated in placental villi, and NGF protein expression in trophoblasts was reduced. BDNF protein expression in trophoblasts remained unchanged, while BDNF mRNA levels were significantly reduced in placental villi of the PCOS group (Fig. 3a). These results were consistent with the decreased levels of NGF (but not BDNF) in umbilical cord blood of the PCOS group (Table 1), underscoring the potential involvement of neurotrophins in PCOS offspring development.

To identify the potential role of DNA methylation in transcriptional regulation, we analysed CpG islands of BDNF and NGF genes in placenta using bisulphite genomic sequencing PCR. No significant differences between PCOS individuals and controls were identified. However, the examined promoter region of all examined genes was hypomethylated (Fig. 3b and S2).

3.4. Genome-wide DNA methylation and expression patterns in placenta of PCOS patients

Global analysis of methylomic and transcriptomic data revealed an altered pattern of DNA methylation of promoters (Fig. 4a, b) and gene

Fig. 1. Neurological examination of female offspring in polycystic ovary syndrome (PCOS) and control group with Dubowitz optimality score. PCOS: n = 20; control: n = 32. Data were presented as mean ± SEM. *P < 0.05. PCOS, polycystic ovary syndrome.
A cluster of neural function-related genes, including \textit{LIN7A} and \textit{MAP1B}, were identified with integrative analysis of the methylome and transcriptome. mRNA expression levels were confirmed with real-time PCR (Fig. S3).

Inflammatory disease and response processes were identified using GO enrichment analysis of differentially transcribed gene clusters in PCOS. Cellular growth, and development and function of nervous and endocrine systems were also closely related to PCOS (Fig. 4d). Enriched GO terms in differentially expressed gene clusters of the PCOS group involved brain development, and proliferation and differentiation of neural precursor cells, neurons, and neuroblast. The enriched GO terms in differentially methylated gene clusters of the PCOS group were consistently associated with neuromodulation and synaptic transmission, cognition, learning, and, memory (Fig. 4e).

To investigate possible protein-protein interactions among these genes, we performed transcriptome analysis with STRING, which identified the FOS gene as having a core role (Fig. S4). mRNA and protein expression levels were significantly downregulated in women
with PCOS (Fig. 4f). The methylation level of FOS was detected by bisulphite genomic sequencing PCR (Fig. 4g, S2). However, no significant differences between PCOS individuals and controls were identified.

Next, we employed bioinformatic analysis to elucidate the molecular basis of transcriptional modification of BDNF and NGF in the placenta. The putative motifs of transcriptional factor FOS were identified through in silico analysis of genomic DNA 5 kb upstream and downstream of BDNF and NGF placenta-specific transcription start sites (TSS). The predicted proximal binding sites of FOS were 2.2 kb upstream and 32 bp downstream of BDNF and NGF TSS, respectively (Fig. 4h).

3.5. Expression levels of NGF and BDNF in FOS-overexpressing Swan71 cells

In order to further verify the regulatory effects of FOS on BDNF and NGF expression, we constructed a human placental trophoblast cell line Swan71 with stable overexpression of FOS. Western blot analysis indicated that FOS protein expression levels were higher in the overexpression group (Fig. 4i, S5). BDNF mRNA levels in the overexpression group were lower, while NGF levels were significantly higher than those of controls (Fig. 4j, k), indicating that FOS inhibited BDNF expression and enhanced NGF expression.
3.6. Morris water maze test in female F1 and F2 offspring of PCOS model rats

The PCOS rat model was successfully established, indicated by irregular oestrous cycle, elevated serum FAI levels (Fig. S6a), multiple cystic follicles, and decreased numbers of GCs and corpora lutea observed with H&E staining (Fig. S6b). The target crossing times of PCOS female F1 animals at puberty were lower than those of controls, with fold change as the abscissa and −log10(P value) as the ordinate. Peacock blue and purple splashes represent genes that significantly up or down regulated respectively. Gray splashes mean genes without significantly different expression. FOS gene is marked as red. (d) Gene Ontology (GO) terms enriched in differently expressed genes in placenta from women with PCOS. (e) Neural GO terms enriched in differentially expressed gene (top) and methylated DNA (bottom) in placenta from women with PCOS. (f) FOS gene mRNA and protein expression levels in human placenta, as assessed by real-time polymerase chain reaction (real-time PCR) (PCOS: n = 20; control: n = 32) and immunofluorescence with FOS (green), and nuclear DNA (blue) labeling (n = 5 in each group). All scale bars are 50 μm. (g) The summary data of methylation status at the promoter region of FOS gene in the placental villi using bisulphite genomic sequencing (BSP) (n = 3 in each group, significance was determined by Chi-square test). (h) In silico analysis of the genomic DNA 5 kb upstream and downstream of BDNF (pIV) and NGF (pI) transcription start site (TSS). Detection of FOS (i) protein expression levels, BDNF (j) and NGF (k) mRNA expression levels in overexpression group (n = 3) and control group (n = 3). Data were presented as mean ± SEM. *P < 0.05, unpaired two-tailed Student’s t tests.

3.7. Transcriptomic analysis of the hippocampus and hypothalamus of F1 pubertal female rats

As hypothalamic and hippocampal gene expression changes lead to persistent neurobehavioural consequences in rodents [61], we conducted a global analysis of transcriptomic data of F1 pubertal female rats. The top 20 GO terms and KEGG pathways were identified in brains of pubertal female offspring of PCOS model rats. Synapse- and neuron-related terms were significantly enriched in the hippocampus and hypothalamus (Fig. 5c, d). Nervous system-related pathways including long-term potentiation, amphetamine addiction, glutamatergic synapses, dopaminergic synapses, long-term
Fig. 5. Neurobehavioral and cerebral changes in female offspring of polycystic ovary syndrome (PCOS) model rats. (a) Morris water maze test in PCOS female F1 (n = 5 in 5 week, n = 6 in 13 week) and controls (n = 7 in 5 week, n = 6 in 13 week). (b) Morris water maze test in PCOS female F2 (n = 6 in 5 week and 13 week) and controls (n = 6 in 5 week and 13 week). Data are presented as mean ± SEM. *P < 0.05, unpaired two-tailed Student’s t tests. (c, d) The Gene Ontology (GO) and Kyoto Encyclopedia of Genes and Genomes (KEGG) enrichment scatter plot of differential alternative splicing (DAS) genes in hippocampus (c) and hypothalamus (d) of F1 pubertal female offspring. The x-axis indicates the rich factor, and the y-axis indicates the name of the term or pathway. The dot size means the gene number, and the dot color represents the P value. The top 20 GO terms and KEGG pathways were shown. (e) The morphological and ultrastructure study of hippocampus neurons and synapses of F1 pubertal female offspring with electron microscopy. The nuclear chromatin (red arrows), rough endoplasmic reticulum (blue arrows), mitochondria (green arrows) and synapses (yellow arrows) in hippocampus were observed.
depression, and neurotrophin signalling were enriched in the hippocampus (Fig. 5c). Pathways involving amphetamine addiction, gluta-
matergic synapses, dopaminergic synapses, long-term potentiation, and neurotrophin signalling were enriched in the hypothalamus (Fig. 5d). We identified nine significantly enriched DNA stability-
related GO terms in analysis of the DEGs in the hypothalamus of F1
pubertal female offspring (Fig. S7); however, none were identified in the hippocampus (data not shown).

3.8. Electron microscopic examination of the pubertal hippocampus of female F1 rats

In the pubertal hippocampus of female F1 rats, nuclear chromatin
was uneven. Part of the cytoplasm was dissolved and presented as
vacuole-like denaturation. Mitochondria were swollen and dilated with
vacuolisation and ruptured mitochondrial cristae. A decrease in
rough endoplasmic reticulum was also identified in neurons. Synaptic
structures were partially dissolved, and synaptic number was
decreased. Aberrant synaptic vesicles and blurred synaptic clefts
were observed (Fig. 5e).

4. Discussion

A major finding of this study was impaired neurobehavioural
development of female infants of women with PCOS. In trophoblasts
of the PCOS group, FOS and NGF protein levels were downregulated.
A cluster of neural function-related genes was identified using inte-
grative analysis of the methylome and transcriptome. The FOS gene
plays a core role in inhibiting BDNF and enhancing NGF in Swan71
cells. The target crossing times in the Morris water maze test were
lower in female F1 offspring of PCOS model rats at puberty than in
controls; moreover, significant changes were noted in neuronal pro-
cesses and pathways in the pubertal hippocampus and hypothalamus
based on transcriptomic analysis.

FOS inhibits placental trophoblast proliferation, migration, and
invasion [14]. FOS is also critical for regulating placental development
and controlling trophoblast function. Dysfunction in FOS may lead to
placental-related pregnancy complications involving pre eclampsia
and abortion [62–64]. As placental FOS expression is diminished and
FOS inhibits placental cell proliferation, diminished FOS expression
in PCOS may be the result of a compensatory placental response to a
sub-optimal maternal environment. BDNF potentiates placental
development and plays an important role in cytrophoblast differ-
entiation, proliferation, and survival [65]. Neurotrophins may also
modulate metabolism or nutrient flux to the foetus [66]. BDNF signal-
ing impacts placental responses to adverse maternal effects [67, 68].
The down-regulation of placental BDNF may be implicated in altered
foetal programming in these infants [69, 70]. Adequate NGF levels at
the fetomaternal interface are essential for the progression of preg-
nancy [71]. Nevertheless, evidence for the interaction between FOS
and neurotrophins in human placenta is limited. Here, we observed
that the expression of the neurotrophins NGF and BDNF were regu-
lated by FOS in both human placental trophoblasts and trophoblast
cell lines, which may contribute to the maternal impact to the foetus.
FOS may regulate BDNF in zebrafish brain [56]. Attenuated FOS
expression indirectly decreases NGF in stromal cells from patients
with endometriosis [72]. AP-1 transcription factors, including FOS
family members, participate in the induction of BDNF exon I, III, and
VI transcripts in rat neurons [73]. In addition, we predicted potential
sites of FOS transcriptional modification in neurotrophin genes.
Although the methylation levels of NGF and BDNF promoters did not
significantly differ according to PCOS status, other pathways may
underpin FOS-mediated regulation of NGF and BDNF.

A significant reduction in placental weight, thickness, density, and
volume; and a higher foetal/placental weight ratio are observed in
the PCOS population compared to that in healthy controls. Further,
irregular placental shape is more frequent in PCOS patients [74]. Pla-
cental pathophysiological examination has revealed that women
with PCOS have significantly more inflammatory and thrombotic
lesions as well as increased villous immaturity of the placenta when
compared with women from a reference group [75]. To date, the clin-
ic evidence on birth weights of offspring of mothers with PCOS
remains controversial [75]. The present study demonstrates altered
proliferation and apoptosis of trophoblasts, vascularisation of villi,
derocrine function, and nutrient transfer in placenta of PCOS women,
which may affect foetal development.

The influence of maternal PCOS on offspring has received consid-
erable attention in recent years [76–78]. Environmental factors dur-
ing foetal development may induce an epigenetic transgenerational
inheritance of adult-onset disease [79–81]. Female F0 mice with
PCOS-like traits induced by late-gestation injection of dihydrotestos-
terone resulted in female F1-F3 offspring with PCOS-like repro-
ductive and metabolic phenotypes. The sequencing of single metaphase
II oocytes from F1-F3 offspring revealed common and unique altered
gene expression across all generations [82]. Female F1 and F2 off-
spring of rats with ancestral dehydroepiandrosterone exposure
exhibited PCOS-like reproductive and metabolic phenotypes, includ-
ing disrupted oestrous cycles and polycystic ovaries, increased serum
levels of testosterone, impaired glucose tolerance, and widespread
metabolic abnormalities [83].

In the present study, we evaluated the neurobehavioural develop-
ment of female first and second filial generations of PCOS model rats
at puberty and adulthood using the Morris water maze, which pro-
vides insight into the effects of maternal PCOS on offspring. Maternal
PCOS significantly affected the neurobehavioural development of the
female first filial generation of PCOS model rats at puberty. Based on
these results, subsequent GO and KEGG enrichment analysis were
conducted. DAS genes were identified in the hippocampus and hypo-
thalmus of pubertal female offspring of PCOS model rats. As alterna-
tive splicing occurs at high frequency in brain tissue and contributes
to multiple aspects of nervous system development including cell-
fate decisions, neuronal migration, axon guidance, and synaptogene-
sis [67,72], these findings were consistent with those of the Morris
water maze test.

Epidemiologic, clinical, and experimental data indicate that the
maternal intrauterine environment plays a critical role in tissue and
organ development, and may induce aberrant responses later in life
by triggering a cascade of events [84]. PCOS induces gestational
hyperandrogenism [85], which was also observed in our study, con-
stituting a potential source of androgen excess for the foetus. To
address this issue, the present study assessed in utero androgen ex-
posure conditions in both gestational and perinatal periods in an indi-
rect manner, but no significant effects of intrauterine androgen
exposure on foetal tissue were noted. Similar phenomena were
observed in experimental animal studies, although the maternal levels of inflammatory
cytokines may compromise brain phenotypes [86]. Based on these
findings, other factors inducing placental alterations in PCOS may be
involved. Future work should explore other intrauterine variables
with higher-powered studies.

The present study has several limitations. First, the sample size
of women with PCOS and controls was relatively small which has
increased sampling bias in the study. More samples are needed to
confirm the present conclusions given the clinical heterogeneity of
PCOS across patients and populations. Second, we were unable to
conduct a long-term follow-up of the offspring. As our intention was
to compare PCOS offspring with controls to provide clinicians insight
into relevant neurobehavioural differences, neurological examination
of newborns was performed in our study. Third, the Swan71 cell line
likely originated from females and is now tetraploid with chromo-
somal and microsatellite instability, which may compromise the re-
levance of any in vitro findings. Further investigations are underway to
clarify the mechanisms underlying FOS-driven modifications of
neurotrophin genes. Fourth, the PCOS rat model used here reflects a restricted range of PCOS phenotypes in women. As F1 and F2 rats were randomly selected from the F0 generation, and six F0 PCOS rats delivered, the number of F1 and F2 rats for neurobehavioural testing was limited. A previous study reported that both mothers and fathers exposed to chronic social instability during adolescence and early adulthood transmit altered behaviours, including enhanced anxiety and social deficits, to their F1 offspring; however, only F1 fathers transmit all behaviours to their F2 and F3 daughters, indicating that maternal exposure may skip a generation and manifest only in F2 and F3 [87]. The low sample size of the study may have obscured differences in FAL. Future studies should follow offspring until F3 with larger sample sizes to investigate the transgenerational effects of PCOS. Finally, the role of placental FOS in neurotrophin regulation in the context of maternal impact on neurobehavioural alterations in PCOS offspring should be examined in the future.

In conclusion, our findings demonstrate that FOS-mediated regulation of neurotrophins in the placenta at least partly underpins the negative effects of maternal PCOS on neurobehavioural development of female offspring.

Data sharing
The raw data of RRBS and RNA-seq have been deposited in NCBI's Gene Expression Omnibus and are accessible through GEO Series accession number GSE154274 (https://www.ncbi.nlm.nih.gov/geo/query/acc.cgi?acc=GSE154274).

Declaration of Competing Interests
The authors have no conflict of interest to declare.

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Supplementary materials
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