

Detecting Tuberculosis in Prisons: Switching Off the Disease at its Source

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Mycobacterium tuberculosis is estimated to have killed 1 billion people over the last 200 years (1) and remains the world's most deadly human pathogen (2).

In order to improve upon the current 2% annual decline in tuberculosis incidence and get anywhere near the 90% by 2035 reduction target of the World Health Organization (WHO) "End Tuberculosis Strategy" (3), tuberculosis must be stopped at source.

There are few sources of tuberculosis worthier of intervention than tuberculosis disease arising in prisons. In resource poor settings, the incidence of tuberculosis is on average 23 times greater inside prisons than in the surrounding community (4). The incidence in the Brazilian prisons in this study was 3,900 per 100,000, one hundred times that of the general population. Unless tuberculosis disease is diagnosed and treated within the prison, on release, prisoners are more likely to return to high risk transmission networks and fuel the epidemic further (5). Prison settings often compound the problem by facilitating transmission through overcrowding, inadequate ventilation, incomplete treatment, late case detection and high prisoner turnover (6). Thus, the case for prison intervention is compelling.

It is naïve to think that tuberculosis disease is contained within the walls of the prison. As a case in point, Sarita Colonia prison in the province of Callao, Peru according to publicly available figures is overcrowded by 483% (7). In 2018, it was shown that living in close proximity to the prison significantly increased the probability of sharing identical tuberculosis pathogen genotypes with those inside the prison (8). Prison intervention will therefore diminish the incidence of tuberculosis cases in the surrounding community. Arguably even more concerning; due to prisoner exchange between countries, new cases of identical transmitted strains from Sarita Colonia prison in Peru have now emerged in both Florence and Madrid (9). Therefore, prison intervention may even act to contain international tuberculosis spread.

Although much research time and many publications have focussed on the benefits of intervening in prisons to diagnose and treat tuberculosis, there remains a lack of evidence to determine which - of all the options available - is the best employ (10). Santos and co-authors in this edition of *Clinical Infectious Diseases* present the results of an ambitious yet well delivered study that helps to address the gaps in this important field of research.

The authors intensively and prospectively screened consenting prisoners in three Brazilian prisons for tuberculosis using symptom screening, GeneXpert, sputum culture and chest radiography with Computer-Aided Detection for Tuberculosis (CAD4TB). They then retrospectively applied four alternative less comprehensive screening algorithms to the data to determine which was most cost effective per case detected. They found that 84% of tuberculosis cases were detectable by a single sputum sample for Xpert MTB/RIF, and that systematic screening with this method had a cost per case of US\$234. By comparison, symptom screening had a similar cost per case detected at US\$235, but missed twice as many cases, while algorithms involving CXR screening were more expensive and did not increase overall yield compared to testing with sputum Xpert/MTBRIF alone.

This is a clear, well written paper drawn from research undertaken in extremely difficult field conditions that has generated clinically applicable results. It demonstrates that the most sensitive and effective algorithm - of those that were tested - to detect tuberculosis disease in prisons is to apply GeneXpert to any prisoner who is able to produce sputum. Considering the challenging environment that the authors were working in, the screening participation rate of 89.9% is impressive and adds to the generalizability of the study.

Only 31% of patients were able to produce a sputum sample, which, as the authors conclude themselves likely underestimates the true burden of tuberculosis disease.

This raises the question of whether interferon gamma release assays and/or tuberculin skin testing together with chest radiography could have a role in detecting these cases that were potentially missed. While many studies have demonstrated the utility of GeneXpert as a point of care test and it is therefore expected that it would improve screening algorithms wherever it is applied, there are few studies that examine its use head to head with other algorithms in prisons.

Overall, the merit of this paper rests on the quality of its intensive screening strategy and the fact that this was applied to all prisoners in an extremely high burden setting. It clearly demonstrates the utility of GeneXpert in prisons - one of the major sources of new tuberculosis disease - and argues correctly for the implementation of this test in similar settings worldwide. If the WHO End TB targets are to be achieved then well-informed, well-funded and widespread scaling-up tuberculosis control in prisons is a good place to start.

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References

1. Paulson T. Epidemiology: A mortal foe. *Nature*. 2013 Oct 10;502(7470):S2-3.
2. Gordon SV, Parish T. Microbe Profile: Mycobacterium tuberculosis: Humanity's deadly microbial foe. *Microbiol Read Engl*. 2018;164(4):437–9.
3. Uplekar M, Weil D, Lonnroth K, Jaramillo E, Lienhardt C, Dias HM, et al. WHO's new end TB strategy. *Lancet Lond Engl*. 2015 May 2;385(9979):1799–801.
4. Baussano I, Williams BG, Nunn P, Beggiato M, Fedeli U, Scano F. Tuberculosis Incidence in Prisons: A Systematic Review. *PLoS Med* [Internet]. 2010 Dec 21 [cited 2020 Jan 31];7(12). Available from: <https://www.ncbi.nlm.nih.gov/pmc/articles/PMC3006353/>
5. Altice FL, Azbel L, Stone J, Brooks-Pollock E, Smyrnov P, Dvoriak S, et al. The perfect storm: incarceration and the high-risk environment perpetuating transmission of HIV, hepatitis C virus, and tuberculosis in Eastern Europe and Central Asia. *Lancet Lond Engl*. 2016 Sep 17;388(10050):1228–48.
6. Escombe AR, Oeser CC, Gilman RH, Navincopa M, Ticona E, Pan W, et al. Natural ventilation for the prevention of airborne contagion. *PLoS Med*. 2007 Feb;4(2):e68.
7. INPE. Informe Estadístico Penitenciario [Internet]. Ministerio de Justicia y Derechos Humanos; 2016 [cited 2020 Jan 31]. Available from: <https://www.inpe.gob.pe/revistas/estadistica/2016/octubre2016/mobile/index.html>
8. Warren JL, Grandjean L, Moore DAJ, Lithgow A, Coronel J, Sheen P, et al. Investigating spillover of multidrug-resistant tuberculosis from a prison: a spatial and molecular epidemiological analysis. *BMC Med*. 2018 Aug 3;16(1):122.
9. Perez Garcia L. Tuberculosis en un escenario global: Nuevas estrategias transnacionales para optimizar la vigilancia de su transmisión. XV Congreso Argentino de Microbiología CAM 2019; 2019 Sep 26; Buenos Aires.
10. Systematic review on the diagnosis, treatment, care and prevention of tuberculosis in prison settings [Internet]. European Centre for Disease Prevention and Control. 2017 [cited 2020 Jan 31]. Available from: <https://www.ecdc.europa.eu/en/publications-data/systematic-review-diagnosis-treatment-care-and-prevention-tuberculosis-prison>