Cerebrospinal Fluid Biomarkers of Neurodegeneration, Synaptic Dysfunction, and Axonal Injury Relate to Atrophy in Structural Brain Regions Specific to Alzheimer's Disease

Elizabeth E. Moore^{a,*}, Katherine A. Gifford, PsyD^{a,*}, Omair A. Khan, MS^b, Dandan Liu, PhD^b, Kimberly R. Pechman, PhD^a, Lealani Mae Y. Acosta, MD, MPH^a, Susan P. Bell, MBBS, MSCI^{a,c}, Maxim Turchan^a, Bennett A. Landman, PhD^{d,e,f}, Kaj Blennow, MD, PhD^{g,h}, Henrik Zetterberg, MD, PhD^{g,h,i,j}, Timothy J. Hohman, PhD^a, & Angela L. Jefferson, PhD^{a,c}

^aVanderbilt Memory & Alzheimer's Center, Department of Neurology, Vanderbilt University Medical Center, Nashville, TN, 37212, USA

^bDepartment of Biostatistics, Vanderbilt University Medical Center, Nashville, TN, 37212, USA

^cDivision of Cardiovascular Medicine, Department of Medicine, Vanderbilt University Medical Center, Nashville, TN, 37212, USA

^dDepartment of Radiology & Radiological Sciences, Vanderbilt University Medical Center, Nashville, TN, 37212, USA

^eDepartment of Biomedical Engineering, Vanderbilt University, Nashville, TN, 37212, USA

^fDepartment of Electrical Engineering and Computer Science, Vanderbilt University, Nashville, TN, 37212, USA

^gDepartment of Psychiatry and Neurochemistry, Institute of Neuroscience and Physiology, The Sahlgrenska Academy at University of Gothenburg, Mölndal, Sweden

^hClinical Neurochemistry Lab, Sahlgrenska University Hospital, Mölndal, Sweden

ⁱDepartment of Neurodegenerative Disease, UCL Institute of Neurology, Queen Square, London, UK

^jUK Dementia Research Institute at UCL, London, UK

*Elizabeth E. Moore and Katherine A. Gifford contributed equally.

Total word count: 3690 Abstract word count: 249 Introduction word count: 481 Discussion word count: 1349 Number of tables: 8 Number of figures: 3 Number of pages: 44 Abbreviated Title: CSF Biomarkers & AD Signature

Acknowledgements: The authors would like to thank the dedicated Vanderbilt Memory & Aging Project participants, their loved ones, and the devoted staff and trainees who contributed to recruitment, screening, and enrollment of the baseline cohort. The authors would also like to thank the dedicated and skilled laboratory technicians at the Clinical Neurochemistry Laboratory at Sahlgrenska University Hospital, Sweden, who performed all the CSF analyses. This research was supported by Alzheimer's Association IIRG-08-88733 (ALJ), R01-AG034962 (ALJ), R01-NS100980 (ALJ), R01-AG056534 (ALJ), T32-AG058524 (ALJ), K24-AG046373 (ALJ), R21-AG059941 (TJH), Paul B. Beeson Career Development Award in Aging K23-AG045966 (KAG), K23-AG030962 (ALJ), K01-AG049164 (TJH), T32-GM007347 (EEM), UL1-TR000445 (Vanderbilt Clinical Translational Science Award), S10-OD023680 (Vanderbilt's High-Performance Computer Cluster for Biomedical Research), and the Vanderbilt Memory & Alzheimer's Center. HZ is a Wallenberg Academy Fellow supported by grants from the Swedish Research Council (#2018-02532), the European Research Council (#681712) and Swedish State Support for Clinical Research (#ALFGBG-720931) and the UK Dementia Research Institute at UCL.

Conflicts of interest: HZ has served at scientific advisory boards for Roche Diagnostics, Samumed, CogRx and Wave, has given lectures in symposia sponsored by Biogen and Alzecure, and is a co-founder of Brain Biomarker Solutions in Gothenburg AB, a GU Ventures-based platform company at the University of Gothenburg. The other authors declare no competing financial interests.

Address for correspondence:

Angela L. Jefferson, PhD Vanderbilt Memory & Alzheimer's Center 1207 17th Avenue South, Suite 204 Nashville, TN 37212 Phone: 615-322-8676 Fax: 615-343-1302 Email: angela.jefferson@vumc.org

Abstract

Introduction: Patterns of atrophy can distinguish normal cognition from Alzheimer's disease (AD), but neuropathological drivers of this pattern are unknown. This study examined associations between cerebrospinal fluid biomarkers of AD pathology, synaptic dysfunction, and neuroaxonal injury with two AD imaging signatures.

Methods: Signatures were calculated using published guidelines. Linear regressions related each biomarker to both signatures, adjusting for demographic factors. Bootstrapped analyses tested if associations were stronger with one signature versus the other.

Results: Increased phosphorylated tau (p-tau), total tau, and neurofilament light (Pvalues <.045) related to smaller signatures (indicating greater atrophy). Diagnosis and sex modified associations between p-tau and neurogranin (P-values<.05) and signatures, such that associations were stronger among participants with mild cognitive impairment and female participants. The strength of associations did not differ between signatures.

Discussion: Increased evidence of neurodegeneration, axonopathy, and tau phosphorylation relate to greater AD-related atrophy. Tau phosphorylation and synaptic dysfunction may be more prominent in AD-affected regions in females.

Key Words: AD signature, cortical thickness, cerebrospinal fluid, sex differences

Significance Statement: Certain patterns of brain atrophy on MRI distinguish individuals with normal cognition from clinical Alzheimer's disease (AD), but the neuropathological drivers of this atrophy pattern are unknown. We present novel neuroimaging and biomarker evidence suggesting that in addition to phosphorylated tau, axonopathy uniquely contributes to AD-related atrophy. Sex modifies associations linking both phosphorylated tau and synaptic dysfunction with atrophy in regions susceptible to AD-related neurodegeneration, such that associations are only present in females. Results suggest neuronal dysfunction may be more prominent in these regions susceptible to AD-related neurodegeneration among females and underscore the importance of examining sex-specific drivers of neuropathology and structural brain changes in AD.

1. Introduction

Alzheimer's disease (AD) is traditionally considered a disease of grey matter neurodegeneration, resulting from the accumulation of amyloid beta (A β) plaques and phosphorylated tau (p-tau).¹ As pathology evolves, atrophy occurs in a regionally specific pattern beginning in the temporal lobe and hippocampus² and spreading in concentric circles to the surrounding cortices. Recent efforts have identified magnetic resonance imaging (MRI) patterns of neurodegeneration that are specific to AD-related changes. Prior work shows that cortical thinning in the temporal and inferior parietal lobes quantified on T_{7} -weighted MRI can reliably predict cognitive decline³ and conversion to AD.^{4,5} Two identified patterns (the Schwarz AD signature⁶ and the McEvoy AD signature⁷) reliably distinguish individuals with normal cognition from clinical AD. However, the neuropathological drivers of this regionally specific atrophy remain uncharacterized.

Prior work has found neurodegeneration within AD signature regions relate to amyloid deposition,⁸ *in vivo* cerebrospinal fluid (CSF) measurements of p-tau and total tau (t-tau),⁹ and tau burden at autopsy.⁶ However, beyond core AD pathology, it is unknown how concomitant pathways to neurodegeneration, such as synaptic dysfunction or axonal injury, relate to atrophy in AD signature regions. Previous work has implicated neurogranin (reflecting synaptic degeneration¹⁰) and neurofilament light (NFL, a marker of axonal injury¹¹) in AD, and each is associated with smaller hippocampal volumes.^{11,12} Thus, these pathologies may contribute to atrophy detected in the AD signature. Further, evidence suggests that AD risk factors, such as female sex,¹³ apolipoprotein E (*APOE*) ε 4 carrier status,¹⁴ and older age,¹⁵ may lead to

heterogeneity in AD-related atrophy and must be considered when examining neuropathologic correlates of atrophy. For example, females with AD pathology have greater hippocampal atrophy compared to males,¹³ and *APOE*-ɛ4 carriers have smaller hippocampal volumes compared to non-carriers.¹⁴ To optimize the diagnostic and prognostic utility of AD signatures, associations of co-occurring pathologies, as well as sex-specific, genetic, and normal aging contributions, to clinically relevant atrophy must be elucidated.

The aim of this study is to investigate associations between CSF biomarkers of core AD pathology (Aβ and p-tau), neurodegeneration (t-tau), synaptic dysfunction (neurogranin), and axonal injury (NFL) with two publicly available AD neuroimaging signatures among older adults.^{6,7} We hypothesize that increased CSF evidence of core AD pathology will relate to cortical thinning in AD signature regions, consistent with prior work,^{9,16} and concomitant pathologies (i.e., neurodegeneration, synaptic dysfunction, and axonal injury) will independently contribute to this atrophy pattern beyond AD pathology. Given the imaging signature can differentiate individuals with and without cognitive impairment^{6,7} and the known modifying effects of sex,¹³ APOE- ε 4 carrier status,¹⁴ and age¹⁵ on neurodegeneration, associations will be tested for interactions with diagnosis, sex, APOE- ε 4 carrier status, and age. We hypothesize that associations between CSF biomarkers and the AD signature will be stronger in MCI participants. females, APOE-E4 carriers, and older adults. Finally, we formally tested if the pattern of neurodegeneration associated with each CSF biomarker was more strongly related to one AD signature versus the other.

2. Materials & Methods

2.1 Study Cohort

The Vanderbilt Memory & Aging Project (VMAP) is a longitudinal observational study investigating vascular health and brain aging,¹⁷ enriched with older adults with mild cognitive impairment (MCI). Inclusion required participants be ≥ 60 years, speak English, have adequate auditory and visual acuity, and have a reliable study partner. As part of a comprehensive screening, participants were excluded for a cognitive diagnosis of dementia,^{18,19} MRI contraindication, history of neurological disease (e.g., multiple sclerosis, stroke), heart failure, major psychiatric illness, head injury with loss of consciousness>5 minutes, or a systemic or terminal illness affecting follow-up participation. At enrollment, participants completed a comprehensive examination, including (but not limited to) fasting blood draw, physical examination, clinical interview, medication review, echocardiogram, cardiac magnetic resonance, multi-modal brain MRI, and optional lumbar puncture. Participants were excluded from the current study for missing usable CSF, brain MRI, or covariate data. See **Figure 1** for inclusion/exclusion details. The protocol was approved by the Vanderbilt University Medical Center Institutional Review Board. Written informed consent was obtained from participants prior to data collection.

2.2 Lumbar Puncture & Biochemical Analysis

A subset of participants completed an optional fasting lumbar puncture at enrollment (n=153, **Figure 1**). CSF was collected with polypropylene syringes using a Sprotte 25-gauge spinal needle in an intervertebral lumbar space. Samples were immediately mixed and centrifuged. Supernatants were aliquoted in 0.5 mL polypropylene tubes and stored at -80°C. Samples were analyzed in single batch using a commercially available enzyme-linked immunosorbent assays (Fujirebio, Ghent, Belgium) to measure CSF concentrations of Aβ42 (INNOTEST® β-AMYLOID₍₁₋₄₂₎), p-tau (INNOTEST® PHOSPHO-TAU_(181P)), and t-tau (INNOTEST® hTAU). Additional enzyme-linked immunosorbent assays measured CSF neurogranin²⁰ and CSF NFL (UmanDiagnostics, Umeå, Sweden) concentrations. Board-certified laboratory technicians processed data blinded to clinical information. Intra-assay coefficients of variation were <10%.²⁰

2.3 APOE-ε4 Genotyping

APOE-ε4 genotyping was performed on deoxyribonucleic acid extracted from whole blood with a Taqman single-nucleotide polymorphism genotyping assay from Applied Biosystems (Foster City, California), as previously published.¹⁷ Real-time polymerase chain reaction in 5 µl aliquots was performed on the Life Technologies 7900HT machine and results were analyzed using Life Technologies SDS 2.4.1 software. *APOE*-ε4 status was defined as carrier (ε2/ε4, ε3/ε4, ε4/ε4) or non-carrier (ε2/ε2, ε2/ε3, ε3/ε3).

2.4 Brain MRI Acquisition & Post-Processing

VMAP participants were scanned at the Vanderbilt University Institute of Imaging Science on a 3T Philips Achieva system (Best, the Netherlands) using an 8-channel SENSE reception coil array. T_1 -weighted images (repetition time=8.9ms, echo

time=4.6ms, spatial resolution=1x1x1mm³) were acquired as part of a larger multi-modal protocol and post-processed using FreeSurfer (http://surfer.nmr.mgh.harvard.edu/).²¹ T_1 -weighted images were registered to MNI space, intensity corrected, and skull stripped. Subcortical structures, cortical structures, and white matter were segmented, and white and grey matter surfaces were constructed for each hemisphere. These surfaces were used to calculate cortical thickness and inflated for visualization. Surfaces were manually inspected and corrected for registration, topological, and segmentation defects. After manual correction, images were reprocessed to update the transformation template and segmentation information. The Schwarz AD signature was calculated by summing bilateral cortical thickness measurements from regions shown to distinguish individuals with AD from normal cognition, including the entorhinal cortex, middle temporal cortex, inferior parietal cortex, fusiform gyrus, and precuneus.⁶ The McEvoy AD signature was calculated by a weighted sum of volumes and cortical thickness measurements from regions shown to predict conversion to AD, including the hippocampus, entorhinal cortex, middle temporal cortex, superior temporal cortex, cingulate gyrus, and orbitofrontal gyrus.⁷ Consistent with prior literature, contributions of intracranial volume were regressed out of the hippocampal volume term and contributions of age and sex were regressed out of the McEvoy signature.⁷

2.5 Experimental Design & Statistical Analyses

Linear regression models with ordinary least square estimates related continuous CSF measures of A β , p-tau, t-tau, neurogranin, and NFL concentrations (pg/mL) to each AD signature, adjusting for age, sex, race/ethnicity, education, *APOE*- ϵ 4 status, and

cognitive diagnosis. Models examining the McEvoy AD signature also adjusted for intracranial volume. Linear regression models also related those CSF biomarkers with significant associations to either AD signature as competing predictors using identical covariates. This competitive model approach assessed the extent to which variance accounted for by an individual biomarker was unique or overlapping with another biomarker. Secondary models evaluated *CSF biomarker x cognitive diagnosis*, *CSF biomarker x sex*, *CSF biomarker x APOE-ɛ4 status*, and *CSF biomarker x age* interactions on the AD signature. Models were repeated stratified by diagnosis (NC, MCI), sex (male, female), *APOE-ɛ*4 status (carrier, non-carrier), and age group based on the sample median (<73, \geq 73).

For all models, follow-up bootstrapped analyses of partial correlations tested if the pattern of neurodegeneration associated with each CSF biomarker was more strongly related to one signature versus the other. The threshold for statistical significance was set *a priori* at p<0.05. For all significant models, sensitivity analyses were performed repeating models excluding participants with predictor or outcome values >4 standard deviations from the group mean. Analyses were conducted using R version 3.5.2 (www.r-project.org).

3. Results

3.1. Participant Characteristics

Participants included 153 adults age 60 to 92 (72±6 years, 67% male, 93% non-Hispanic White, 33% *APOE*- ϵ 4 carriers). CSF A β concentrations ranged 289 to 1195 pg/mL, p-tau concentrations ranged 13 to 157 pg/mL, t-tau concentrations ranged 77 to 1542 pg/mL, neurogranin concentrations ranged 57 to 462 pg/mL, and NFL concentrations ranged 268 to 4025 pg/mL. The Schwarz AD signature ranged 1.93 to 2.65 and the McEvoy AD signature ranged -9.71 to 6.53. See **Table 1** for participant characteristics for the entire sample and stratified by diagnosis. See **Supplemental Table 1** for participants characteristics stratified by sex.

3.2. CSF Biomarkers & the AD Signatures

Among all participants, higher CSF t-tau (β =-0.0001, p=0.005) and NFL concentrations (β =-0.0001, p=0.001) were associated with smaller Schwarz AD signature values only. Higher CSF p-tau was associated with smaller McEvoy AD signature values (β =-0.0163, p=0.04) and with smaller Schwarz AD signature values (β =-0.0008, p=0.06) but the latter observation did not meet the *a priori* significance threshold. Neither CSF A β nor CSF neurogranin were associated with either AD signature (p-values>0.14). See **Table 2** and **Figure 2** for details. Sensitivity models excluding outliers yielded similar results (data not shown). In bootstrapped analyses, the variance explained by CSF p-tau (95% confidence interval (CI) -0.04, 0.02), t-tau (95% CI -0.05, 0.03), and NFL (95% CI -0.08, 0.02) on the Schwarz signature was not statistically difference from the variance explained on the McEvoy signature for each

biomarker. See Table 3 for details.

3.3. CSF Biomarkers as Competing Predictors of Atrophy in AD Signature Regions

To determine whether the CSF biomarkers represent a common pathological pathway to atrophy in AD signature regions, a linear regression model was fit including all statistically significant CSF biomarker predictors for each signature. For the Schwarz AD signature, higher CSF NFL remained associated with lower Schwarz signature values (β =-0.00005, p=0.01), while the association with CSF t-tau was slightly attenuated (β =-0.0001, p=0.09). See **Table 4** for details. To compare the effect size of associations across CSF biomarkers, a change in R² was calculated for the competing models. The addition of CSF NFL to the model contributed 5.6% of variance beyond covariates and the addition of CSF t-tau contributed 3.5% of variance beyond covariates. Together, CSF NFL and t-tau contributed 6.6% of variance beyond covariates.

For the McEvoy AD signature, higher CSF NFL (β =-0.0011, p=0.005) remained associated with lower McEvoy signature values in a combined model with t-tau. Similarly, when CSF NFL was combined with p-tau, only NFL remained associated with McEvoy signature values (β =-0.0012, p<0.002). See **Table 4** for details. To compare the effect size of associations across CSF biomarkers, a change in R² was calculated for the competing models. The addition of CSF NFL to the model contributed 8.5% of variance beyond covariates, the addition of CSF t-tau contributed 4.5% of variance beyond covariates, and the addition of CSF p-tau contributed 1.6% of variance beyond covariates. Together, CSF NFL, t-tau and p-tau contributed 14.6% of variance beyond covariates.

3.4. CSF Biomarkers x Cognitive Diagnosis Interactions & the AD Signatures

CSF p-tau (p=0.002), t-tau (p=0.003), and neurogranin (p=0.03) interacted with cognitive diagnosis on the Schwarz AD signature. CSF A β and NFL did not interact with diagnosis (p-values>0.10). In stratified analyses, higher CSF p-tau (β =-0.0025, p=0.009), t-tau (β =-0.0002, p=0.002), and NFL (β =-0.0001, p=0.001) were associated with smaller Schwarz AD signature values among MCI participants only. CSF A β (p=0.04), p-tau (p=0.002), t-tau (p=0.001), and neurogranin (p=0.03) interacted with cognitive diagnosis on the McEvoy AD signature but NFL did not (p=0.14). Similarly, stratified analyses revealed that higher CSF p-tau (β =-0.0303, p=0.02), t-tau (β =-0.0040, p=0.004), and NFL (β =-0.0016, p=0.003) were associated with smaller McEvoy AD signature values among MCI participants only. In bootstrapped analyses, interactions and stratified associations did not differ in variance explained between the two signatures. See **Table 3** for details.

3.5. CSF Biomarkers x Sex Interactions & the AD Signatures

CSF p-tau (p=0.05) and neurogranin (p=0.04) interacted with sex on the Schwarz AD signature. Stratified models revealed that higher CSF p-tau (β =-0.0015, p=0.04) and t-tau (β =-0.0001, p=0.01) related to lower Schwarz AD signature values in females only, while the association with neurogranin approached significance (β =-0.0005, p=0.06). CSF neurogranin interacted with sex on the McEvoy AD signature (p=0.04), such that

associations were present in females only (β =-0.0073, p=0.04). Stratified analyses also revealed that higher CSF t-tau (β =-0.0027, p=0.02) related to lower McEvoy AD signature values in females only. CSF A β and NFL did not interact with sex on either AD signature (p-values>0.07). See **Table 6** and **Figure 3** for details. Sensitivity models excluding outliers yielded similar results (data not shown). In bootstrapped analyses, interactions and stratified associations did not differ in variance explained between the two signatures. See **Table 3** for details.

3.6. CSF Biomarkers x APOE- ε 4 Carrier Status Interactions & the AD Signatures

No CSF biomarker interacted with *APOE*- ε 4 status on either AD signature (p-values>0.11). In models stratified by *APOE*- ε 4 status, higher CSF t-tau (β =-0.0001, p=0.02) and NFL (β =-0.0001, p=0.006) related to smaller Schwarz AD signature values among *APOE*- ε 4 non-carriers. CSF NFL (β =-0.0014, p<0.0001) also related to smaller McEvoy AD signature values among *APOE*- ε 4 non-carriers. See **Table 7** for details. Sensitivity models excluding outliers yielded similar results (data not shown). In bootstrapped analyses, stratified associations did not differ in variance explained between the two signatures. See **Table 3** for details.

3.7. CSF Biomarkers x Age Interactions & the AD Signatures

No CSF biomarker interacted with age on either AD signature (p-values>0.11). However, stratified analyses revealed that higher CSF t-tau (β =-0.0001, p=0.04) and NFL (β =-0.0001, p=0.02) related to lower Schwarz AD signature values among individuals over age 73. Similarly, higher CSF t-tau (β =-0.0030, p=0.02) and NFL (β =- 0.0014, p=0.001) related to lower McEvoy AD signature values among individuals over age 73. See **Table 8** for details. Sensitivity models excluding outliers yielded similar results (data not shown). In bootstrapped analyses, stratified associations did not differ in variance explained between the two signatures. See **Table 3** for details.

4. Discussion

Among community-dwelling older adults, higher concentrations of CSF p-tau, ttau, and NFL were associated with lower AD signature values, indicating greater atrophy in grey matter regions affected by AD pathology. As expected, cognitive diagnosis modified the associations between CSF p-tau, t-tau, and neurogranin and AD signature values, such that associations were stronger among participants with MCI. Sex also modified associations between CSF p-tau and neurogranin, such that associations were stronger in females. Collectively, results suggest that CSF biomarkers of p-tau, t-tau, synaptic dysfunction, and axonal injury are associated with AD-related atrophy measured on MRI, and p-tau and synaptic dysfunction may be more prominent in AD-specific regions in females.

This study is among the first to show associations linking *in vivo* molecular biomarkers of core AD and concomitant pathologies with atrophy in AD-specific regions. As expected, higher levels of CSF p-tau and t-tau are associated with atrophy in the AD signatures among all participants, extending prior work focusing only on individuals with MCI.^{6,9} As tau becomes more phosphorylated, it is no longer able to bind to microtubules within the axon,²² compromising its ability to maintain cell structure and leading to cell death.²³ Additionally, it is well established that tau tangles first develop in anatomical regions included in the AD signatures²⁴ and *in vivo* atrophy in the Schwarz AD signature is associated with greater tangle deposition at autopsy.⁶ Thus, increased levels of CSF p-tau and t-tau may indicate clinically relevant atrophy in AD-specific regions. Despite some previous work demonstrating Aβ relates to cortical thinning in AD signature regions,⁸ we did not detect an association between CSF Aβ and AD-related atrophy. It is notable this prior study included individuals with clinical AD, suggesting A β may not contribute to neurodegeneration in these regions until later disease states.

We also found an association between CSF NFL, a biomarker of axonal injury,²⁵ and AD-related atrophy. Importantly, CSF NFL explained variance in the AD signatures beyond that of CSF p-tau or t-tau, suggesting that axonal injury uniquely contributes to atrophy in these regions. NFL is a support protein within large axons²⁵ that is released into the CSF in a variety of pathological states, including vascular disease,²⁶ inflammation,²⁷ and neurodegeneration.¹¹ Regions included in the AD signatures, such as the entorhinal cortex and hippocampus, are vulnerable to ischemic small vessel disease²⁸ and chronic inflammation,^{29,30} possibly contributing to axonopathy in these regions. Though increased CSF NFL is not specific to AD, axonal injury does contribute to AD-related atrophy, highlighting the importance of pathologies other than Aβ and tau in the development of AD-related atrophy and axonal injury as a potential therapeutic target in AD.

Results suggest that diagnosis modifies CSF p-tau, t-tau, and neurogranin associations with the AD signatures, such that associations are only present in participants with MCI. Additionally, though a formal interaction was not detected, CSF NFL was associated with the AD signatures only among participants with MCI. These findings are consistent with prior work showing that CSF p-tau, t-tau,³¹ NFL,¹¹ and neurogranin³² are elevated in individuals with MCI, likely leading to greater MRI evidence of neurodegeneration in AD-specific regions. Notably, though neurogranin was not associated with either AD signature among the entire sample, the diagnostic interaction and stratified results suggest that increased CSF neurogranin has a greater contribution to AD-related atrophy in MCI compared to NC participants. Neurogranin is a post-synaptic protein prominent in the hippocampus³³ critical for long-term potentiation and synaptic plasticity.³⁴ While neurogranin tissue levels are decreased in AD,³⁵ CSF levels rise, presumably leaking into the CSF as synapses become disrupted and misfire. As synaptic dysfunction begins prior to overt tau tangle formation,³² apoptotic cascades are initiated,³⁶ followed by neurodegeneration and cognitive impairment.³⁷ Given the importance and relative overexpression of neurogranin in regions vulnerable to AD pathology,^{35,38} synaptic dysfunction in MCI, indicated by increased CSF neurogranin, may be an etiology underlying AD-related atrophy. CSF neurogranin is also elevated in conditions other than AD, such as stroke,³⁹ suggesting synaptic dysfunction may serve as one pathway by which vascular disease directly contributes to AD-related atrophy.

We found that sex modifies CSF p-tau and neurogranin associations with the AD signatures, such that associations are only present in females. While there is not a significant interaction, stratified results suggest that associations between CSF t-tau and the AD signatures are also female specific. This observation is consistent with prior work showing that the association between CSF t-tau and hippocampal volume is stronger in females.¹³ Though atrophy in AD signature regions distinguishes NC from AD in both males and females,^{6,7} our results suggest that the underlying pathology may differ. Estrogen is protective of tau hyperphosphorylation⁴⁰ and modulates dendritic spine density in the hippocampus,^{41,42} so the decline in estrogen in aging females may lead to increased p-tau, synaptic dysfunction, and subsequent neurodegeneration. Additionally, women generally have fewer synapses in the temporal lobe,⁴³ suggesting that a given degree of synaptic dysfunction in aging in the temporal lobe may exert a

much greater effect on neuronal health in women compared to men. Thus, p-tau and synaptic dysfunction may be more prominent etiologies of AD-related atrophy in women, adding to the growing body of literature showing sex differences in pathology and structural brain changes in AD.^{13,44} Interestingly, CSF NFL did not interact with sex on either AD signature, despite men having higher concentrations. It is possible that the effect of axonal injury on atrophy is more robust than other pathologies,³⁷ such that any sex-specific drivers of axonal injury are not strong enough to alter the association with atrophy. Future work is needed to better understand the mechanisms underlying sex-specific associations reported here and elsewhere.^{13,44,45}

APOE- ε 4 carrier status did not modify associations between CSF biomarkers and the AD signatures. Prior work suggests that the effects of *APOE*- ε 4 on brain health are modified by sex and biomarker status.^{13,45} Thus, it is possible the effects of *APOE*- ε 4 on AD-related atrophy are also modified by these variables. Future work is needed with replication in larger samples to better understand these complicated three-way interactions and elucidate mechanisms by which *APOE*- ε 4 affects neurodegeneration. Additionally, age did not modify any of the associations reported here, likely because the AD signature regions were originally selected based on their ability to separate agerelated neurodegeneration from AD-specific neurodegeneration.^{6,7}

Finally, one observation worth noting is that none of the associations reported here were significantly different between the Schwarz⁶ and McEvoy⁷ AD signatures. Despite the McEvoy signature including hippocampal volume and cortical thickness measurements that differ from the brain regions in the Schwarz signature, the two measures do not provide different information regarding the CSF biomarkers assessed here. That is, one signature is not more sensitive to detecting atrophy associated with AD pathology, neurodegeneration, synaptic dysfunction, or axonopathy compared to the other. Future research is needed to further delineate differences in the two signatures (and others not examined here)⁴⁶⁻⁴⁸ and determine if specific regions within the signatures are more or less vulnerable to certain pathologies.

The current study has several strengths, including a clinically well characterized cohort with excellent methods for quantifying cortical thickness and CSF biomarkers of AD and concomitant pathology. Additional strengths include comprehensive ascertainment of potential confounders, a comprehensive investigation of concomitant pathways to neurodegeneration beyond primary AD pathology, and the use of core laboratories using quality control procedures to analyze all CSF and brain MRI measurements in batch with technicians blinded to participant clinical information. However, limitations include the cross-sectional methods, which cannot address causality. Longitudinal studies are needed to understand how AD pathology, synaptic dysfunction, and axonal injury affect AD-related neurodegeneration over time. Additionally, the cohort was older and predominantly non-Hispanic White, thus limiting generalizability to other races, ethnicities, and age groups.

This study demonstrates novel associations linking CSF biomarkers of neurodegeneration and axonopathy with atrophy in regions vulnerable to AD-related neurodegeneration. Additionally, sex modified associations, such that p-tau, t-tau, and neurogranin were associated with AD-related atrophy in females only. These findings suggest that axonal injury driving AD-related neurodegeneration and neuronal dysfunction may be more prominent in these regions in females. Future research is needed to better understand how these pathologies regionally interact to precipitate cognitive decline and sex-specific etiologies of neurodegeneration.

References

- Braak H, Braak E, Bohl J, Reintjes R. Age, neurofibrillary changes, a betaamyloid and the onset of alzheimer's disease. *Neuroscience Letters*. 1996;210:87-90
- Rabinovici GD, Seeley WW, Kim EJ, Gorno-Tempini ML, Rascovsky K, Pagliaro TA, Allison SC, Halabi C, Kramer JH, Johnson JK, Weiner MW, Forman MS, Trojanowski JQ, Dearmond SJ, Miller BL, Rosen HJ. Distinct mri atrophy patterns in autopsy-proven alzheimer's disease and frontotemporal lobar degeneration. *American journal of Alzheimer's disease and other dementias*. 2007;22:474-488
- Busovaca E, Zimmerman ME, Meier IB, Griffith EY, Grieve SM, Korgaonkar MS, Williams LM, Brickman AM. Is the alzheimer's disease cortical thickness signature a biological marker for memory? *Brain imaging and behavior*. 2016;10:517-523
- Gross AL, Manly JJ, Pa J, Johnson JK, Park LQ, Mitchell MB, Melrose RJ, Inouye SK, McLaren DG. Cortical signatures of cognition and their relationship to alzheimer's disease. *Brain imaging and behavior*. 2012;6:584-598
- Bakkour A, Morris JC, Dickerson BC. The cortical signature of prodromal ad: Regional thinning predicts mild ad dementia. *Neurology*. 2009;72:1048-1055
- Schwarz CG, Gunter JL, Wiste HJ, Przybelski SA, Weigand SD, Ward CP, Senjem ML, Vemuri P, Murray ME, Dickson DW, Parisi JE, Kantarci K, Weiner MW, Petersen RC, Jack CR, Jr. A large-scale comparison of cortical thickness and volume methods for measuring alzheimer's disease severity. *NeuroImage. Clinical.* 2016;11:802-812

- McEvoy LK, Fennema-Notestine C, Roddey JC, Hagler DJ, Jr., Holland D, Karow DS, Pung CJ, Brewer JB, Dale AM. Alzheimer disease: Quantitative structural neuroimaging for detection and prediction of clinical and structural changes in mild cognitive impairment. *Radiology*. 2009;251:195-205
- Wang L, Benzinger TL, Su Y, Christensen J, Friedrichsen K, Aldea P, McConathy J, Cairns NJ, Fagan AM, Morris JC, Ances BM. Evaluation of tau imaging in staging alzheimer disease and revealing interactions between betaamyloid and tauopathy. *JAMA Neurol.* 2016;73:1070-1077
- Dickerson BC, Wolk DA. Biomarker-based prediction of progression in mci: Comparison of ad signature and hippocampal volume with spinal fluid amyloidbeta and tau. *Front Aging Neurosci*. 2013;5:55
- Thorsell A, Bjerke M, Gobom J, Brunhage E, Vanmechelen E, Andreasen N, Hansson O, Minthon L, Zetterberg H, Blennow K. Neurogranin in cerebrospinal fluid as a marker of synaptic degeneration in alzheimer's disease. *Brain research*. 2010;1362:13-22
- Zetterberg H, Skillback T, Mattsson N, Trojanowski JQ, Portelius E, Shaw LM, Weiner MW, Blennow K. Association of cerebrospinal fluid neurofilament light concentration with alzheimer disease progression. *JAMA Neurol.* 2016;73:60-67
- Portelius E, Zetterberg H, Skillback T, Tornqvist U, Andreasson U, Trojanowski JQ, Weiner MW, Shaw LM, Mattsson N, Blennow K. Cerebrospinal fluid neurogranin: Relation to cognition and neurodegeneration in alzheimer's disease. *Brain.* 2015;138:3373-3385

- Koran MEI, Wagener M, Hohman TJ. Sex differences in the association between ad biomarkers and cognitive decline. *Brain imaging and behavior*. 2017;11:205-213
- 14. Agosta F, Vossel KA, Miller BL, Migliaccio R, Bonasera SJ, Filippi M, Boxer AL, Karydas A, Possin KL, Gorno-Tempini ML. Apolipoprotein e epsilon4 is associated with disease-specific effects on brain atrophy in alzheimer's disease and frontotemporal dementia. *Proc Natl Acad Sci U S A*. 2009;106:2018-2022
- Oh H, Madison C, Villeneuve S, Markley C, Jagust WJ. Association of gray matter atrophy with age, beta-amyloid, and cognition in aging. *Cereb Cortex*. 2014;24:1609-1618
- Knopman DS, Lundt ES, Therneau TM, Vemuri P, Lowe VJ, Kantarci K, Gunter JL, Senjem ML, Mielke MM, Machulda MM, Roberts RO, Boeve BF, Jones DT, Petersen RC, Jack CR, Jr. Joint associations of beta-amyloidosis and cortical thickness with cognition. *Neurobiol Aging*. 2018;65:121-131
- 17. Jefferson AL, Gifford KA, Acosta LM, Bell SP, Donahue MJ, Taylor Davis L, Gottlieb J, Gupta DK, Hohman TJ, Lane EM, Libon DJ, Mendes LA, Niswender K, Pechman KR, Rane S, Ruberg FL, Ru Su Y, Zetterberg H, Liu D. The vanderbilt memory & aging project: Study design and baseline cohort overview. *Journal of Alzheimer's Disease*. 2016;52:539-559
- Aisen PS, Petersen RC, Donohue MC, Gamst A, Raman R, Thomas RG, Walter S, Trojanowski JQ, Shaw LM, Beckett LA, Jack CR, Jr., Jagust W, Toga AW, Saykin AJ, Morris JC, Green RC, Weiner MW, Alzheimer's Disease

Neuroimaging Initiative. Clinical core of the alzheimer's disease neuroimaging initiative: Progress and plans. *Alzheimers Dement*. 2010;6:239-246

- Albert MS, DeKosky ST, Dickson D, Dubois B, Feldman HH, Fox NC, Gamst A, Holtzman DM, Jagust WJ, Petersen RC, Snyder PJ, Carrillo MC, Thies B, Phelps CH. The diagnosis of mild cognitive impairment due to alzheimer's disease: Recommendations from the national institute on aging-alzheimer's association workgroups on diagnostic guidelines for alzheimer's disease. *Alzheimers Dement*. 2011;7:270-279
- Kvartsberg H, Duits FH, Ingelsson M, Andreasen N, Ohrfelt A, Andersson K, Brinkmalm G, Lannfelt L, Minthon L, Hansson O, Andreasson U, Teunissen CE, Scheltens P, Van der Flier WM, Zetterberg H, Portelius E, Blennow K. Cerebrospinal fluid levels of the synaptic protein neurogranin correlates with cognitive decline in prodromal alzheimer's disease. *Alzheimers Dement*. 2015;11:1180-1190
- 21. Fischl B, Dale AM. Measuring the thickness of the human cerebral cortex from magnetic resonance images. *Proc Natl Acad Sci U S A*. 2000;97:11050-11055
- Bramblett GT, Goedert M, Jakes R, Merrick SE, Trojanowski JQ, Lee VM.
 Abnormal tau phosphorylation at ser396 in alzheimer's disease recapitulates development and contributes to reduced microtubule binding. *Neuron*.
 1993;10:1089-1099
- Tarawneh R, Head D, Allison S, Buckles V, Fagan AM, Ladenson JH, Morris JC, Holtzman DM. Cerebrospinal fluid markers of neurodegeneration and rates of brain atrophy in early alzheimer disease. *JAMA Neurol.* 2015;72:656-665

- 24. Braak H, Braak E. Staging of alzheimer's disease-related neurofibrillary changes. *Neurobiol Aging*. 1995;16:271-278; discussion 278-284
- Friede RL, Samorajski T. Axon caliber related to neurofilaments and microtubules in sciatic nerve fibers of rats and mice. *The Anatomical Record*. 1970;167:379-387
- Skillback T, Farahmand B, Bartlett JW, Rosen C, Mattsson N, Nagga K, Kilander L, Religa D, Wimo A, Winblad B, Rosengren L, Schott JM, Blennow K, Eriksdotter M, Zetterberg H. Csf neurofilament light differs in neurodegenerative diseases and predicts severity and survival. *Neurology*. 2014;83:1945-1953
- 27. Melah KE, Lu SY, Hoscheidt SM, Alexander AL, Adluru N, Destiche DJ, Carlsson CM, Zetterberg H, Blennow K, Okonkwo OC, Gleason CE, Dowling NM, Bratzke LC, Rowley HA, Sager MA, Asthana S, Johnson SC, Bendlin BB. Cerebrospinal fluid markers of alzheimer's disease pathology and microglial activation are associated with altered white matter microstructure in asymptomatic adults at risk for alzheimer's disease. *Journal of Alzheimer's Disease*. 2016;50:873-886
- Du AT, Schuff N, Laakso MP, Zhu XP, Jagust WJ, Yaffe K, Kramer JH, Miller BL, Reed BR, Norman D, Chui HC, Weiner MW. Effects of subcortical ischemic vascular dementia and ad on entorhinal cortex and hippocampus. *Neurology*. 2002;58:1635-1641
- 29. Hauss-Wegrzyniak B, Lynch MA, Vraniak PD, Wenk GL. Chronic brain inflammation results in cell loss in the entorhinal cortex and impaired ltp in perforant path-granule cell synapses. *Experimental neurology*. 2002;176:336-341

- Lue LF, Brachova L, Civin WH, Rogers J. Inflammation, a beta deposition, and neurofibrillary tangle formation as correlates of alzheimer's disease neurodegeneration. *Journal of neuropathology and experimental neurology*. 1996;55:1083-1088
- 31. Mattsson N, Zetterberg H, Hansson O, Andreasen N, Parnetti L, Jonsson M, Herukka SK, van der Flier WM, Blankenstein MA, Ewers M, Rich K, Kaiser E, Verbeek M, Tsolaki M, Mulugeta E, Rosen E, Aarsland D, Visser PJ, Schroder J, Marcusson J, de Leon M, Hampel H, Scheltens P, Pirttila T, Wallin A, Jonhagen ME, Minthon L, Winblad B, Blennow K. Csf biomarkers and incipient alzheimer disease in patients with mild cognitive impairment. *JAMA*. 2009;302:385-393
- 32. Kester MI, Teunissen CE, Crimmins DL, Herries EM, Ladenson JH, Scheltens P, van der Flier WM, Morris JC, Holtzman DM, Fagan AM. Neurogranin as a cerebrospinal fluid biomarker for synaptic loss in symptomatic alzheimer disease. JAMA Neurol. 2015;72:1275-1280
- Scheff SW, Price DA, Schmitt FA, DeKosky ST, Mufson EJ. Synaptic alterations in ca1 in mild alzheimer disease and mild cognitive impairment. *Neurology*. 2007;68:1501-1508
- 34. Huang KP, Huang FL, Jager T, Li J, Reymann KG, Balschun D. Neurogranin/rc3 enhances long-term potentiation and learning by promoting calcium-mediated signaling. *J Neurosci.* 2004;24:10660-10669
- 35. Davidsson P, Blennow K. Neurochemical dissection of synaptic pathology in alzheimer's disease. *Int Psychogeriatr*. 1998;10:11-23

- 36. D'Amelio M, Cavallucci V, Middei S, Marchetti C, Pacioni S, Ferri A, Diamantini A, De Zio D, Carrara P, Battistini L, Moreno S, Bacci A, Ammassari-Teule M, Marie H, Cecconi F. Caspase-3 triggers early synaptic dysfunction in a mouse model of alzheimer's disease. *Nature neuroscience*. 2011;14:69-76
- Mattsson N, Insel PS, Palmqvist S, Portelius E, Zetterberg H, Weiner M, Blennow K, Hansson O. Cerebrospinal fluid tau, neurogranin, and neurofilament light in alzheimer's disease. *EMBO molecular medicine*. 2016;8:1184-1196
- Represa A, Deloulme JC, Sensenbrenner M, Ben-Ari Y, Baudier J. Neurogranin: Immunocytochemical localization of a brain-specific protein kinase c substrate. J Neurosci. 1990;10:3782-3792
- De Vos A, Bjerke M, Brouns R, De Roeck N, Jacobs D, Van den Abbeele L, Guldolf K, Zetterberg H, Blennow K, Engelborghs S, Vanmechelen E. Neurogranin and tau in cerebrospinal fluid and plasma of patients with acute ischemic stroke. *BMC Neurol*. 2017;17:170
- Goodenough S, Schleusner D, Pietrzik C, Skutella T, Behl C. Glycogen synthase kinase 3beta links neuroprotection by 17beta-estradiol to key alzheimer processes. *Neuroscience*. 2005;132:581-589
- Gould E, Woolley CS, Frankfurt M, McEwen BS. Gonadal steroids regulate dendritic spine density in hippocampal pyramidal cells in adulthood. *J Neurosci*. 1990;10:1286-1291
- 42. Alvarez-de-la-Rosa M, Silva I, Nilsen J, Perez MM, Garcia-Segura LM, Avila J, Naftolin F. Estradiol prevents neural tau hyperphosphorylation characteristic of

alzheimer's disease. *Annals of the New York Academy of Sciences*. 2005;1052:210-224

- Alonso-Nanclares L, Gonzalez-Soriano J, Rodriguez JR, DeFelipe J. Gender differences in human cortical synaptic density. *Proc Natl Acad Sci U S A*. 2008;105:14615-14619
- 44. Buckley RF, Mormino EC, Rabin JS, Hohman TJ, Landau S, Hanseeuw BJ, Jacobs HIL, Papp KV, Amariglio RE, Properzi MJ, Schultz AP, Kirn D, Scott MR, Hedden T, Farrell M, Price J, Chhatwal J, Rentz DM, Villemagne VL, Johnson KA, Sperling RA. Sex differences in the association of global amyloid and regional tau deposition measured by positron emission tomography in clinically normal older adults. *JAMA Neurol.* 2019
- 45. Hohman TJ, Dumitrescu L, Barnes LL, Thambisetty M, Beecham G, Kunkle B, Gifford KA, Bush WS, Chibnik LB, Mukherjee S, De Jager PL, Kukull W, Crane PK, Resnick SM, Keene CD, Montine TJ, Schellenberg GD, Haines JL, Zetterberg H, Blennow K, Larson EB, Johnson SC, Albert M, Bennett DA, Schneider JA, Jefferson AL. Sex-specific association of apolipoprotein e with cerebrospinal fluid levels of tau. *JAMA Neurol.* 2018
- 46. Dickerson BC, Bakkour A, Salat DH, Feczko E, Pacheco J, Greve DN, Grodstein F, Wright CI, Blacker D, Rosas HD, Sperling RA, Atri A, Growdon JH, Hyman BT, Morris JC, Fischl B, Buckner RL. The cortical signature of alzheimer's disease: Regionally specific cortical thinning relates to symptom severity in very mild to mild ad dementia and is detectable in asymptomatic amyloid-positive individuals. *Cereb Cortex*. 2009;19:497-510

- 47. Lerch JP, Pruessner JC, Zijdenbos A, Hampel H, Teipel SJ, Evans AC. Focal decline of cortical thickness in alzheimer's disease identified by computational neuroanatomy. *Cereb Cortex*. 2005;15:995-1001
- 48. Hua X, Lee S, Yanovsky I, Leow AD, Chou YY, Ho AJ, Gutman B, Toga AW, Jack CR, Jr., Bernstein MA, Reiman EM, Harvey DJ, Kornak J, Schuff N, Alexander GE, Weiner MW, Thompson PM. Optimizing power to track brain degeneration in alzheimer's disease and mild cognitive impairment with tensorbased morphometry: An adni study of 515 subjects. *Neuroimage*. 2009;48:668-681

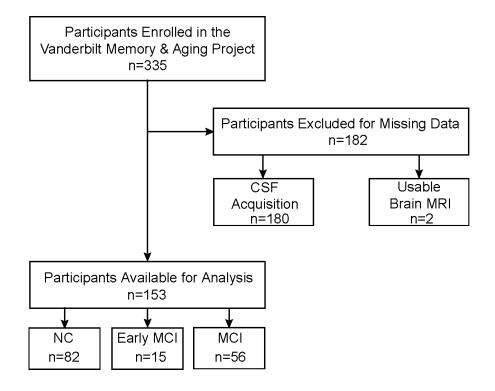
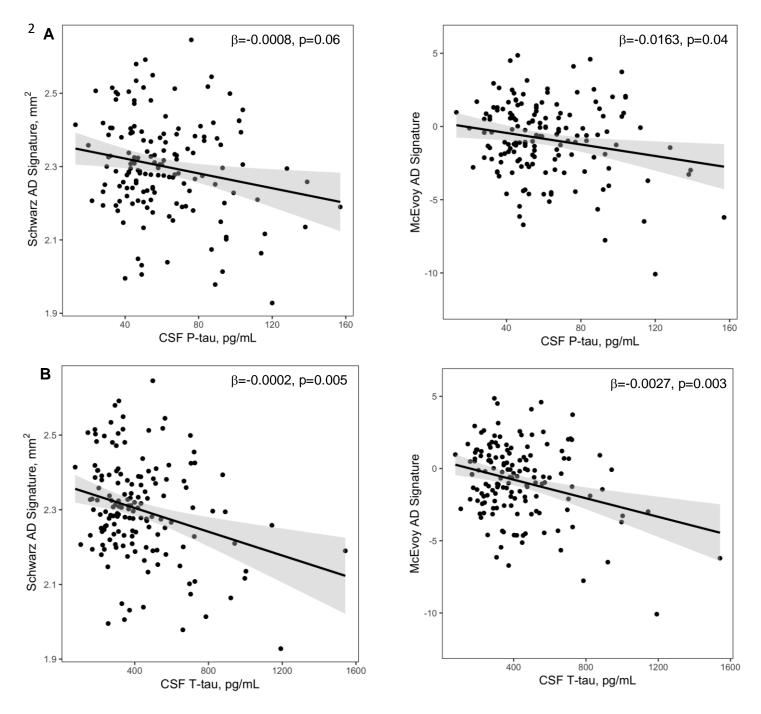


Figure 1. Participant Inclusion/Exclusion Details

Figure 1. Missing data categories are mutually exclusive. An additional 5 participants were excluded from analyses examining CSF neurofilament light. CSF=cerebrospinal fluid; MCI=mild cognitive impairment; MRI=magnetic resonance imaging; NC=normal cognition.



1 Figure 2. CSF Biomarker Associations with the AD Signature

CSF Biomarkers & AD Signature 33

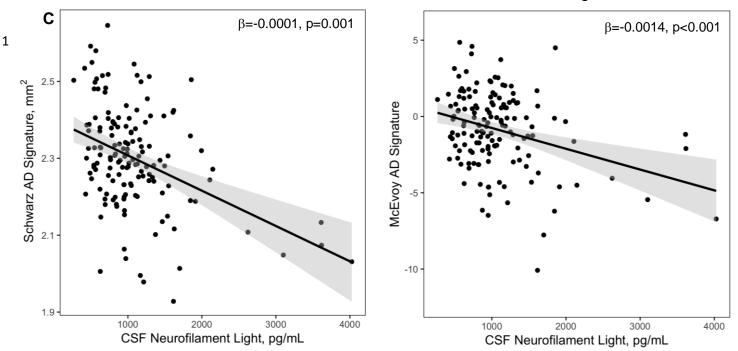


Figure 2. Solid lines reflect values of the AD signature corresponding to CSF biomarkers. Shading reflects 95% confidence interval. **A)** Associations between CSF p-tau and the AD signatures; **B)** Associations between CSF t-tau and the AD signatures; **C)** Associations between CSF NFL and the AD signatures. AD=Alzheimer's disease; CSF=cerebrospinal fluid; NFL=neurofilament light; P-tau=phosphorylated tau; T-tau=total tau.

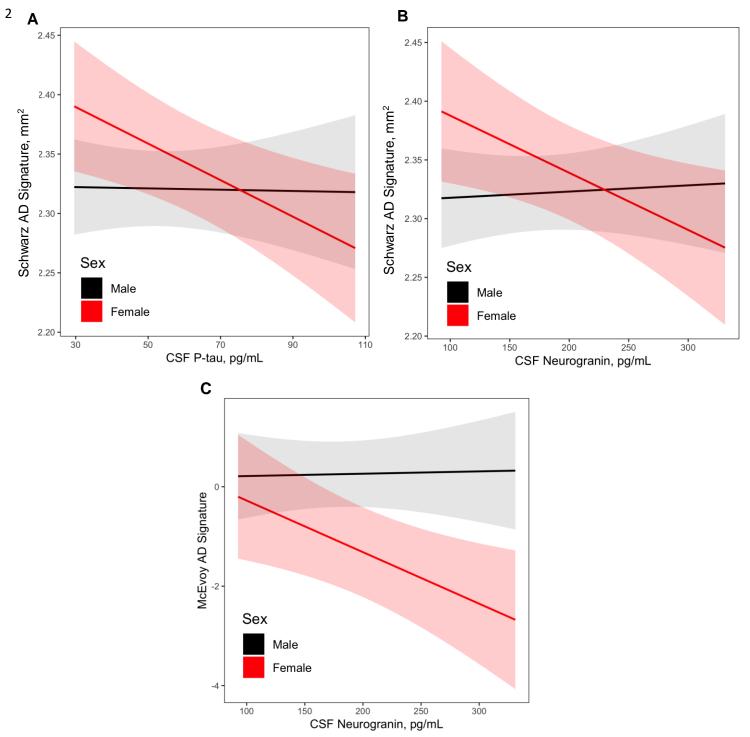


Figure 3. CSF Biomarker x Sex Interactions with the AD Signatures

Figure 3. Lines reflect values of the AD signatures corresponding to CSF biomarkers. Shading reflects 95% confidence interval. **A)** Associations between CSF p-tau and the Schwarz AD signature, stratified by sex; male β =-0.0004, p=0.44; female β =-0.0011, p=0.04; **B)** Associations between CSF neurogranin and the Schwarz AD signature, stratified by sex; male β =-0.00002, p=0.91; female β =-0.0004, p=0.06; **C)** Associations between CSF neurogranin and the McEvoy AD signature, stratified by sex; male β =-0.0073; p=0.04; AD=Alzheimer's disease; CSF=cerebrospinal fluid; P-tau=phosphorylated tau.

Demographic Characteristics	Total n=153	NC n=82	eMCI n=15	MCI n=56	p-value
Age, years	72±6	72±7	73±6	73±6	0.90
Sex, % male	67	70	80	61	0.30
Race, % Non-Hispanic White	93	94	93	91	0.82
Education, years	16±3	17±2	16±3	15±3	0.002*
<i>APOE</i> -ε4, % carrier	33	28	13	45	0.03
Schwarz AD signature, mm ²	2.3±0.13	2.3±0.12	2.3±0.09	2.2±0.14	<0.001*†
McEvoy AD signature	-0.85±2.5	-0.09±2.1	-0.87±2.2	-1.96±2.9	<0.001*
CSF Biomarkers, pg/mL					
Αβ	718±245	765±226	817±282	622 ± 234	<0.001*†
P-tau	61±26	56±22	63±17	67±31	0.06
T-tau	425±227	373±175	429±125	500±290	0.02*
Neurogranin	196±77	185±74	213±73	206±82	0.21
Neurofilament Light	1070±582	939±453	1088±465	1250±718	0.004*

Table 1. Participant Characteristics

Note. Values denoted as mean±standard deviation or frequency. P-values were generated using a Kruskal-Wallis test for continuous variables and a Pearson test for categorical variables. *NC is different than MCI. †eMCI is different than MCI. Aβ=amyloid beta; AD=Alzheimer's disease; APOE=apolipoprotein E; CSF=cerebrospinal fluid; eMCI=early MCI; MCI=mild cognitive impairment; NC=normal cognition; Ptau=phosphorylated tau; T-tau=total tau.

	β	95% Confidence Intervals	p-value
Schwarz AD Signature			
Αβ	0.00004	-0.00005, 0.0001	0.36
P-tau	-0.0008	-0.0015, 0.00003	0.06
T-tau	-0.0001	-0.0002, -0.00004	0.005
Neurogranin	-0.0002	-0.0004, 0.0001	0.23
NFL	-0.00006	-0.0001, -0.00002	0.001
McEvoy AD Signature			
Αβ	0.0009	-0.0010, 0.0029	0.39
P-tau	-0.0163	-0.0319, -0.0007	0.04
T-tau	-0.0027	-0.0045, -0.0009	0.003
Neurogranin	-0.0036	-0.0082, 0.0011	0.14
NFL	-0.0014	-0.0021, -0.0007	<0.001

Table 2. CSF Biomarker Associations with the AD Signatures

Note. Models were adjusted for age, sex, education, race/ethnicity, diagnosis, and APOE- $\varepsilon 4$ status, Models examining the McEvoy AD signature were also covaried for intracranial volume. An additional 5 participants were excluded from the CSF NFL analyses. β indicates the degree of change in volume of AD signature per 1 pg/mL increase in each CSF biomarker. A β = amyloid beta; AD=Alzheimer's Disease; APOE=apolipoprotein E; NFL=neurofilament light; P-tau=phosphorylated tau; T-tau=total tau.

	Schwarz Incremental R ²	McEvoy Incremental R ²	Difference in Incremental R ²	95% Confidence Intervals
Main Effects				
P-tau	0.019	0.023	-0.005	-0.038, 0.019
T-tau	0.041	0.048	-0.007	-0.050, 0.030
NFL	0.057	0.084	-0.027	-0.078, 0.017
Diagnostic Interactions				
Αβ	0.016	0.035	-0.019	-0.071, 0.010
P-tau	0.053	0.078	-0.025	-0.085, 0.016
T-tau	0.048	0.079	-0.031	-0.093, 0.009
Neurogranin	0.029	0.050	-0.021	-0.084, 0.011
MCI Participants Only				
P-tau	0.103	0.088	0.015	-0.065, 0.083
T-tau	0.134	0.131	0.004	-0.064, 0.078
NFL	0.156	0.139	0.017	-0.079, 0.104
Sex Interactions				
P-tau	0.020	0.047	-0.027	-0.095, 0.007
Neurogranin	0.022	0.056	-0.033	-0.108, 0.005
Female Participants Only				
P-tau	0.050	0.048	0.003	-0.076, 0.056
T-tau	0.067	0.070	-0.003	-0.082, 0.065
Neurogranin	0.040	0.053	-0.012	-0.076, 0.042
NFL	0.063	0.082	-0.019	-0.092, 0.046
Male Participants Only				
NFL	0.068	0.100	-0.032	-0.101, 0.034
APOE-ε4 Non-Carriers Only				
T-tau	0.043	0.025	0.018	-0.017, 0.071
NFL	0.059	0.102	0.231	-0.121, 0.015
Participants >73 Only				
T-tau	0.046	0.058	-0.020	-0.055, 0.057
NFL	0.062	0.109	-0.047	-0.121, 0.035

Table 3. Statistical Comparison Between CSF Biomarkers Associations with theAD Signatures

Note. Confidence intervals crossing 0 indicate that there is not a statistically significant difference in the variance explained by the predictor on the two signatures. Difference in incremental R^2 calculated as Schwarz incremental R^2 – McEvoy Incremental R^2 . A β = amyloid beta; *APOE*=apolipoprotein E; NFL=neurofilament light; P-tau=phosphorylated tau; T-tau=total tau.

	β	95% Confidence Intervals	p-value						
Schwarz AD Signature (T-tau + NFL)									
T-tau	-0.0001	-0.0002, 0.00001	0.09						
NFL	-0.00005	-0.0001, -0.00001	0.01						
McEvoy AD Signature (T-tau + NFL)									
T-tau	-0.002	-0.0036, 0.0003	0.09						
NFL	-0.0011	-0.0018, -0.0003	0.005						
McEvoy AD Signature (P-tau + NFL)									
P-tau	-0.007	-0.0231, 0.0096	0.42						
NFL	-0.0012	-0.0020, -0.0005	0.002						

Table 4. Combined Models of CSF Biomarkers in Relation to the AD Signatures

Note. Analyses performed on n=148. Combined models only included biomarkers significantly associated with each signature when analyzed individually (**Table 2**). Models were adjusted for age, sex, education, race/ethnicity, diagnosis, and *APOE*- ϵ 4 status, Models examining the McEvoy AD signature were also covaried for intracranial volume. β indicates the degree of change in volume of AD signature per 1 pg/mL increase in each CSF biomarker. AD=Alzheimer's Disease; *APOE*=apolipoprotein E; CSF=cerebrospinal fluid; NFL=neurofilament light; P-tau=phosphorylated tau; T-tau=total tau.

	CSF Biomarker x Diagnosis Interaction n=138*			NC n=82 [†]			MCI n=56 [†]		
Schwarz AD Signature	β	95% CI	<i>p</i> -value	β	95% CI	p-value	β	95% CI	<i>p</i> -value
Αβ	0.0002	-0.00003, 0.0003	0.10	0.00004	-0.0001, 0.0002	0.53	0.0001	-0.0001, 0.0003	0.47
P-tau	-0.0025	-0.0041, -0.0009	0.002	0.0004	-0.0008, 0.0016	0.51	-0.0016	-0.0027, -0.0004	0.009
T-tau	-0.0003	-0.0005, 0.0001	0.003	0.00003	-0.0001, 0.0002	0.73	-0.0002	-0.0003, 0.0001	0.002
Neurogranin	-0.0006	-0.0011, -0.0001	0.03	0.0001	-0.0003, 0.0004	0.71	-0.0003	-0.0008, 0.0001	0.16
NFL	-0.0001	-0.0001, 0.00001	0.11	-0.00001	-0.0001, 0.0001	0.74	-0.0001	-0.0001, -0.00003	0.001
	CSF Bion	narker x Diagnosis lı n=138*	nteraction	NC n=82 [†]			MCI n=56 [†]		
McEvoy AD Signature	β	95% CI	<i>p</i> -value	β	95% CI	<i>p</i> -value	β	95% CI	<i>p</i> -value
Αβ	0.0039	0.0003, 0.0076	0.04	0.0009	-0.0016, 0.0033	0.48	0.0027	-0.0017, 0.0072	0.22
P-tau	-0.0493	-0.0802, -0.0802	0.002	0.0077	-0.0142, 0.0296	0.49	-0.0303	-0.0555, -0.0052	0.02
T-tau	-0.006	-0.0096, -0.0024	0.001	0.0008	-0.0020, 0.0036	0.58	-0.0040	-0.0066, -0.0014	0.004
Neurogranin	-0.0118	-0.0224, -0.0012	0.03	0.0010	-0.0054, 0.0074	0.76	-0.0066	-0.0170, 0.0038	0.21
NFL	-0.0011	-0.0026, 0.0004	0.14	-0.0004	-0.0017, 0.0008	0.51	-0.0016	-0.0026, -0.0006	0.003

Table 5. CSF Biomarker x Diagnosis Interactions on the AD Signatures

Note: *Models were adjusted for age, sex, education, race/ethnicity, diagnosis, and $APOE \epsilon 4$ status. [†]Models were adjusted for age, sex, education, race/ethnicity, and $APOE \epsilon 4$ status. Models examining the McEvoy AD signature were also covaried for intracranial volume. An additional 5 participants were excluded from the CSF NFL analyses. β indicates the degree of change in volume per 1 pg/mL increase in each CSF biomarkers. A β =amyloid beta; AD=Alzheimer's disease; APOE=apolipoprotein E; CI=confidence interval; CSF=cerebrospinal fluid; MCI=mild cognitive impairment; NC=normal cognition; NFL=neurofilament light; P-tau=phosphorylated tau; T-tau=total tau.

	CSF Biomarker x Sex Interaction n=153*			Female n=50 [†]			Male n=103 [†]		
Schwarz AD Signature	β	95% CI	<i>p</i> - value	β	95% CI	p-value	β	95% CI	<i>p</i> -value
Αβ	0.0001	-0.00005, 0.0003	0.14	0.00003	-0.0001, 0.0002	0.73	0.00007	-0.00005, 0.0002	0.26
P-tau	-0.0015	-0.0030, -0.000001	0.05	-0.0011	-0.0022, -0.0001	0.04	-0.0004	-0.0015, 0.0007	0.44
T-tau	-0.0001	-0.0003, 0.00002	0.08	-0.0001	-0.0003, -0.00003	0.01	-0.0001	-0.0002, 0.00004	0.16
Neurogranin	-0.0005	-0.0011, -0.00003	0.04	-0.0004	-0.0008, 0.00002	0.06	-0.00002	-0.0004, 0.0003	0.91
NFL	-0.00004	-0.0001, 0.00003	0.26	-0.0001	-0.0001, -0.00001	0.02	-0.0001	-0.0001, -0.00002	0.005
	CSF B	iomarker x Sex Intera n=153*	ction	Female n=50) [†]			Male n=103 [†]		
McEvoy AD Signature	β	95% CI	<i>p</i> - value	β	95% CI	<i>p</i> -value	β	95% Cl	<i>p</i> -value
Αβ	0.0032	-0.0003, 0.0068	0.07	0.0001	-0.0042, 0.0044	0.96	0.0014	-0.0011, 0.0038	0.27
P-tau	-0.0246	-0.0543, 0.0051	0.10	-0.0201	-0.0411, 0.0008	0.06	-0.0112	-0.0332, 0.0108	0.31
T-tau	-0.0026	-0.0060, 0.0008	0.13	-0.0027	-0.0049, -0.0004	0.02	-0.0021	-0.0048, 0.0006	0.12
Neurogranin	-0.0098	-0.0190, -0.0006	0.04	-0.0073	-0.0144, -0.0002	0.04	-0.0009	-0.0069, 0.0052	0.78
NFL	-0.0006	-0.0021, 0.0021	0.39	-0.0016	-0.0028, -0.0003	0.01	-0.0015	-0.0024, 0.0006	0.001

Table 6. CSF Biomarker x Sex Interactions on the AD Signatures

Note: *Models were adjusted for age, sex, education, race/ethnicity, diagnosis, and APOE- ϵ 4 status. [†]Models were adjusted for age, education, race/ethnicity, diagnosis, and APOE- ϵ 4 status. [†]Models were adjusted for age, education, race/ethnicity, diagnosis, and APOE- ϵ 4 status. Models examining the McEvoy AD signature were also covaried for intracranial volume. An additional 5 participants were excluded from the CSF NFL analyses. β indicates the degree of change in volume per 1 pg/mL increase in each CSF biomarkers. A β =amyloid beta; AD=Alzheimer's disease; APOE=apolipoprotein E; CI=confidence interval; CSF=cerebrospinal fluid; MCI=mild cognitive impairment; NC=normal cognition; NFL=neurofilament light; P-tau=phosphorylated tau; T-tau=total tau.

	CSF Biomarker x APOE- <i>ɛ</i> 4 Interaction n=153*			APOE-ε4 Carrier n=50 [†]			APOE-ε4 Non-Carrier n=103 [†]		
Schwarz AD Signature	β	95% CI	<i>p</i> -value	β	95% CI	p-value	β	95% CI	<i>p</i> -value
Αβ	0.0001	-0.0001, 0.0003	0.29	0.0001	-0.0001, 0.0003	0.46	0.00004	-0.0001, 0.0001	0.53
P-tau	0.0001	-0.0015, 0.0017	0.87	-0.0001	-0.0015, 0.0013	0.88	-0.0009	-0.0019, 0.0001	0.08
T-tau	0.00001	-0.0002, 0.0002	0.96	-0.0001	-0.0002, 0.0001	0.50	-0.0001	-0.0003, -0.00002	0.02
Neurogranin	-0.0001	-0.0006, 0.0005	0.86	0.00004	-0.0005, 0.0006	0.89	-0.0002	-0.0005, 0.0001	0.32
NFL	0.00001	-0.0001, -0.0001	0.91	-0.0001	-0.0001, 0.00003	0.19	-0.0001	-0.0001, -0.00002	0.006
	CSF Bior	<i>narker x APOE-ɛ4</i> Ir n=153*	nteraction	APOE-ε4 Carrier n=50 [†]			APOE-ε4 Non-Carrier n=103 [†]		
McEvoy AD Signature	β	95% CI	<i>p</i> -value	β	95% CI	<i>p</i> -value	β	95% CI	<i>p</i> -value
Αβ	0.0033	-0.0008, 0.0073	0.11	0.0029	-0.0009, 0.0067	0.13	-0.00004	-0.0024, 0.0023	0.97
P-tau	-0.0126	-0.0448, 0.0195	0.44	-0.0065	-0.0329, 0.0198	0.62	-0.0124	-0.0324, 0.0076	0.22
T-tau	-0.0018	-0.0053, 0.0018	0.33	-0.0015	-0.0044, 0.0014	0.30	-0.0020	-0.0044, 0.0003	0.09
Neurogranin	-0.0046	-0.0161, 0.0069	0.43	-0.0004	-0.0103, 0.0094	0.93	-0.0028	-0.0088, 0.0033	0.37
NFL	0.0001	-0.0014, 0.0016	0.94	-0.0008	-0.0023, 0.0007	0.30	-0.0014	-0.0022, -0.0006	<0.001

Table 7. CSF Biomarker x APOE-_E4 Carrier Status Interactions on the AD Signatures

Note: *Models were adjusted for age, sex, education, race/ethnicity, diagnosis, and APOE- ϵ 4 status. [†]Models were adjusted for age, sex, education, race/ethnicity, and diagnosis. Models examining the McEvoy AD signature were also covaried for intracranial volume. An additional 5 participants were excluded from the CSF NFL analyses. β indicates the degree of change in volume per 1 pg/mL increase in each CSF biomarkers. A β =amyloid beta; AD=Alzheimer's disease; APOE=apolipoprotein E; CI=confidence interval; CSF=cerebrospinal fluid; MCI=mild cognitive impairment; NC=normal cognition; NFL=neurofilament light; P-tau=phosphorylated tau; T-tau=total tau.

	CSF Biomarker x Age Interaction n=153*			<73 years old n=73 [†]			≥73 years old n=80 [†]		
Schwarz AD Signature	β	95% CI	<i>p</i> -value	β	95% CI	p-value	β	95% CI	<i>p</i> -value
Αβ	-0.0001	-0.0003, 0.00003	0.11	0.0001	-0.00001, 0.0003	0.07	-0.00002	-0.0002, 0.0001	0.73
P-tau	-0.0001	-0.0016, 0.0014	0.86	-0.0007	-0.0019, 0.0004	0.19	-0.0008	-0.0019, 0.0004	0.17
T-tau	-0.00004	-0.0002, 0.0001	0.66	-0.0001	-0.0002, 0.00001	0.07	-0.0001	-0.0003, -0.00004	0.04
Neurogranin	0.0001	-0.0004, 0.0006	0.74	-0.0002	-0.0006, 0.0001	0.22	-0.0001	-0.0005, 0.0003	0.74
NFL	0.00001	-0.0001, 0.0001	0.74	-0.0001	-0.0001, 0.00001	0.08	-0.0001	-0.00001, -0.00001	0.02
	CSF Bi	omarker x Age Inter n=153*	action	<73 years old n=73 [†]			≥73 years old n=80 [†]		
McEvoy AD Signature	β	95% CI	<i>p</i> -value	β	95% CI	<i>p</i> -value	β	95% CI	<i>p</i> -value
Αβ	-0.0026	-0.0057, 0.0006	0.11	0.0024	-0.0010, 0.0058	0.16	-0.0001	-0.0028, 0.0025	0.92
P-tau	-0.0080	-0.0378, 0.0218	0.60	-0.0123	-0.0365, 0.0120	0.31	-0.0192	-0.0410, 0.0026	0.08
T-tau	-0.0011	-0.0044, 0.0023	0.52	-0.0022	-0.0049, 0.0006	0.12	-0.0030	-0.0055, -0.0005	0.02
Neurogranin	-0.0014	-0.0116, 0.0089	0.79	-0.0030	-0.0108, 0.0048	0.45	-0.0033	-0.0110, 0.0043	0.39
NFL	-0.0001	-0.0016, 0.0014	0.86	-0.0012	-0.0027, 0.0003	0.12	-0.0014	-0.0023, -0.0006	0.001

Table 8. CSF Biomarker x Age Interactions on the AD Signatures

Note: *Models were adjusted for age, sex, education, race/ethnicity, diagnosis, and APOE- ϵ 4 status. [†]Models were adjusted for sex, education, race/ethnicity, APOE- ϵ 4 status, and diagnosis. Models examining the McEvoy AD signature were also covaried for intracranial volume. An additional 5 participants were excluded from the CSF NFL analyses. β indicates the degree of change in volume per 1 pg/mL increase in each CSF biomarkers. A β =amyloid beta; AD=Alzheimer's disease; APOE=apolipoprotein E; CI=confidence interval; CSF=cerebrospinal fluid; MCI=mild cognitive impairment; NC=normal cognition; NFL=neurofilament light; P-tau=phosphorylated tau; T-tau=total tau.

Demographic Characteristics	Female n=50	Male n=103	p-value
Age, years	72±7	72±6	0.69
APOE-ε4, % carrier	32	33	1
Race, % Non-Hispanic White	85	94	0.55
Education, years	15±3	17±3	<0.001
MCI, %	42	33	0.30
Schwarz AD Signature, mm ²	2.3±0.14	2.3±0.13	0.82
McEvoy AD Signature	-1.63±2.7	-0.47±2.4	0.02
CSF Biomarkers, pg/mL			
Αβ	636±228	756±244	0.005
P-tau	65±30	59±23	0.22
T-tau	471±285	403±191	0.20
Neurogranin	203±83	190±74	0.20
NFL	975±549	1114±594	0.05

Supplemental Table 1. Participant Characteristics Stratified by Sex

Note. Values denoted as mean±standard deviation or frequency. P-values were generated using a Kruskal-Wallis test for continuous variables and a Pearson test for categorical variables. AD=Alzheimer's disease; APOE=apolipoprotein E; CSF=cerebrospinal fluid; MCI=mild cognitive impairment; NFL=neurofilament light; P-tau=phosphorylated tau; T-tau=total tau.