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3	Stem cells as an emerging paradigm in stroke (STEPS) 4:
4	advancing and accelerating preclinical research
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42 Introduction

The scientific community has continuously advanced promising treatment concepts for cell-based therapies in stroke. These approaches principally aim to modulate post-ischemic immune responses and augment endogenous repair. Another aim currently under study at the bench level is to transplant new tissue and restore neural circuits. Many stem and non-stem cell populations have shown encouraging efficacy in preclinical models, and selected types of cell therapies are currently undergoing testing in clinical trials.¹⁻⁴

Recent mechanistic studies have tremendously advanced our understanding of the 49 different parameters that influence experimental stroke therapies. While cell therapies offer 50 unprecedented therapeutic time window expansions of days to weeks (and possibly even 51 52 months to years after stroke), there are several potential factors that may affect their impact. These include age^5 , comorbidities^{6,7}, concurrent medications⁸, and even chronobiological 53 54 mechanisms.⁹ In theory, thorough preclinical research should take into account all of these factors or at least their most relevant combinations. However, budgetary constraints, the lack 55 of adequate in vitro and in vivo models, and the enormous amount of time required to address 56 the multitude of relevant factors severely impairs such attempts in research practice. This 57 dilemma affects current and future translational work and thus requires careful consideration. 58

59 The Stem Cell Therapies as an Emerging Paradigm in Stroke (STEPS) meetings have regularly brought together academic and industry leaders and experts from regulatory 60 authorities to discuss the latest developments in cell therapies for stroke and to publish 61 recommendations for preclinical and clinical research.¹⁰⁻¹² The fourth STEPS meeting aimed to 62 update previous preclinical guidelines with respect to novel stroke models, biomaterials, and 63 advanced approaches combining cell therapies with biomaterials, drugs, or neurorehabilitation. 64 STEPS delegates further provide new recommendation on preclinical study designs including 65 multi-center preclinical trials (MCPTs) and suggest a strategy to accelerate and improve clinical 66 67 translation of cell therapies for stroke without sacrificing scientific rigor and patient safety. This

can be achieved by a close interlink of preclinical and clinical studies while targeting particular
stroke patient subpopulations. Main recommendations are summarized at the end of the STEPS
4 report.

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72 Part I: Updated preclinical guidelines

73 Stroke model selection in the era of recanalization therapies

We recommend selecting models that best represent the clinical population targeted with 74 a particular cell therapy. The recent advent of mechanical thrombectomy has changed the 75 clinical landscape, and the application of cell therapies are discussed directly after 76 recanalization.¹³ Transient models should be selected when investigating this scenario. The 77 filament model is widely used to represent mechanical recanalization¹⁴; however, its use for 78 long-term studies poses some limitations due to large infarcts associated with high mortality.¹⁵ 79 Thromboembolization followed by thrombolysis is a clinically important model for testing cell 80 therapies in the context of thrombolysis.¹⁶ Moreover, reperfusion is often incomplete in patients 81 undergoing thrombolytic therapy or spontaneous recanalization. This is also observed in 82 spontaneously hypertensive rats that can serve as a model for these conditions¹⁷ while also 83 exhibiting other important stroke comorbidities. Total reperfusion failure or persistent 84 occlusion can be modelled by permanent MCAO. 85

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87 *Large animal models*

The gyrencephalic brain featured by large animal modes (LAMs) is bigger than the rodent brain and more suitable for sophisticated clinical imaging approaches.^{18,19} Grey-towhite-matter ratio approximates that of humans.⁹ LAMs allow more realistic and precise testing of cell delivery techniques including stereotaxic and intra-arterial cell administration, and dose translation to human clinical trials. Cell migration and paracrine effects, as in the human brain, are challenged by larger anatomic distances. LAMs are also suitable to investigate stroke 94 sequelae such as cognitive impairment and decline²⁰, and are further recommended to assess
95 the value of potential biomarkers indicating cell therapy safety and efficacy.

On the other hand, LAM studies typically involve smaller sample sizes as they are more 96 expensive and require dedicated infrastructure. Major endpoints including functional outcome 97 and lesion size tend to be more variable than in standardized rodent studies. Although 98 resembling the situation in human patient cohorts, these issues can significantly reduce study 99 power.²¹ LAMs are therefore of limited use in exploratory cell therapy studies. Meaningful 100 LAM experiments require a precise understanding of the addressed endpoint(s), as well as of 101 sample and effect sizes. Nevertheless, LAMs are highly valuable translational tools when 102 considering their limitations and employing them in well-planned confirmative studies.¹¹ 103 104 Funding bodies are encouraged to support research using LAMs in such scenarios, particularly when critical information on patient safety and delivery route efficiency can be obtained. 105

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107 Sex differences, age, and comorbidities

In line with previous recommendations^{11,12}, the STEPS group recommends testing cell 108 therapies in animal models of different age, sex, and comorbidities. However, we also recognize 109 110 that modeling these variables, especially comorbidities, has limitations due their multitude and 111 complexity. The impact of these factors might be better investigated in large phase III clinical trials allowing for sub-hoc analyses of patient populations with respective comorbidity profiles, 112 or in MCPTs combining the capacities of many labs. An alternative approach (outlined in part 113 III) is to focus on stroke patient subpopulations with particular stroke configuration and 114 comorbidity profiles, and to design preclinical studies accordingly. 115

116

117 Dose escalation studies: novel implications

118 In line with previous recommendations^{10,11} and in light of the neutral results from the 119 MASTERS (multipotent adult progenitor cells given intravenously, NCT01436487) and

ACTISSIMA (SB623 administered intracerebrally, NCT02448641) trials that may partially be 120 121 related to dosing issues, the STEPS group continues to recommend efficacy-focused preclinical dose escalation studies for all routes of administration. Intra-arterial administration of cells may 122 cause microvascular obstruction under certain circumstances.²² Hence, dose escalation studies 123 are not only important for preclinical efficacy assessments, but are highly recommended when 124 assessing safety aspects. This particularly accounts for intra-arterial or more invasive 125 application routes. Methods capable of predicting the target territory of cell infusions may help 126 to optimize the safety profile. LAMs may be suitable to simulate clinical transplantation 127 scenarios regarding vessel dimensions and imaging-based surveillance.²³ 128

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130 Drug-cell interactions

It is likely in clinical scenarios that patients receiving cell therapy also receive 131 medications for stroke comorbidities and for secondary prevention. Cell therapies may further 132 be combined with pharmacological treatments to enhance their therapeutic impact.²⁴ Given the 133 paracrine effects of many cell therapies, interactions between drugs and cells cannot be 134 excluded. This important aspect requires careful consideration when moving towards the clinic, 135 but little is known about these potential interactions. Detrimental effects were seen when 136 137 combining granulocyte-colony stimulating factor and bone marrow mononuclear cells, each of which is effective as a stand-alone treatment in rodents.^{25,26} On the other hand, synergistic 138 effects have been reported for the combination of cell therapies with other commonly prescribed 139 medication such as statins.²⁷ 140

The STEPS 4 group recommends more research on potential drug-cell interactions in appropriate *in vitro* and in *in vivo* test systems. Drug classes being predominantly used in stroke patients, such as antiplatelets, anti-hypertensive, and statins, should be the main focus. We further suggest testing on autologous cell preparations when applied in patients receiving multiple medications. These tests can be tailored to the medication profile of individual patients.

147 Biomaterials

Biomaterials are increasingly being incorporated for the delivery of cells to reduce shear 148 stress induced by needle injections^{28,29} but also to provide factors that improve post-149 transplantation cell survival.^{30,31} Scaffolds can support transplanted cells inside the lesion 150 cavity³² by providing structural cues and biochemical signals.^{33,34} Post-stroke tissue 151 restoration³⁵, and a guided neuronal differentiation³⁶ can be achieved using biomaterials 152 engineered to release growth factors, mediators of angiogenesis, or immunomodulators in a 153 temporal sequence and without exerting systemic side effects.³⁷⁻³⁹ A systematic optimization of 154 a hydrogel, for instance, improved the survival of human neural stem cells implanted into the 155 156 stroke-damaged brain and controlled their differentiation. However, it remains unclear if the combined use of biomaterials and cells will transfer to further improvements in functional 157 recovery. To date, most studies combining biomaterials and cells for transplantation are of an 158 exploratory rather than definitive/confirmative nature. We therefore recommend long-term 159 studies to investigate the safety and efficacy profile of biomaterial applications once a basic 160 therapeutic benefit has been shown. LAMs may help to optimize application procedures. Early 161 involvement of regulatory authorities, ideally already during early-stage preclinical research, is 162 163 recommended, as biomaterial-cell combinations are challenging from a regulatory perspective.

164

165 *Neurorehabilitation*

Most stroke survivors receive some form of rehabilitation. Thus, neurorehabilitation is important to consider when developing cellular therapies for stroke. Indeed, treadmill running and intravenous delivery of mesenchymal stem cells together improve behavioral recovery in animals with ischemic stroke.^{40,41} Timing of such combination therapy is crucial when targeting stroke recovery as there is a sensitive phase for neurorehabilitation (Fig. 1A). It is possible that some cell therapies might re-open a plasticity time window in chronic stroke, and

neurorehabilitation may be beneficial in such scenarios by stabilizing the recovered functions. 172 The recent Stroke Recovery and Rehabilitation Roundtable (SRRR)-115 and SRRR-242 173 recommendations are valuable in designing preclinical rehabilitation studies and in improving 174 clinical translation. However, as in the case of comorbidities, including rehabilitation renders 175 176 study designs complex and difficult to implement. Also, the effects of add-on neurorehabilitation should be discriminated from stand-alone cell therapies, which may be 177 challenging as shown recently with adipose tissue-derived stem cells and enriched 178 environments.⁴³ Routine investigation of cell therapy in combination with neurorehabilitation 179 is recommended when significant additional therapeutic effects are expected from this 180 combination, or when the combination is a central mode of action. 181

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183 Part II: New considerations on preclinical study designs

184 Potential new models and targets: lacunar, white matter, and hemorrhagic strokes

Most preclinical studies model large territorial infarcts. However, other important 185 clinical target populations are patients with smaller infarcts in the subcortical grey and white 186 matter. Importantly, the smaller volume of the infarct and the preservation of some anatomical 187 tissue structures may foster repair.⁴⁴ Small deep white matter infarcts may be particularly 188 suitable for cells (e.g. glial progenitors) capable of or intended for tissue restoration⁴⁵ and might 189 be responsive to cell-borne local paracrine mechanisms. We recommend to consider such stroke 190 types (see supplementary table) as alternative targets to large territorial infarcts and/or when 191 192 working on tissue-restorative cell therapies.

193 Intracerebral hemorrhage (ICH)⁴⁶ involves pathogenic mechanisms that may provide 194 novel cell therapy targets. Hemoglobin breakdown products (HBPs), such as hemin, damage 195 axons and induce ferroptosis and necroptosis in distant, primarily intact neuronal somata.⁴⁷ 196 These processes might be mitigated or reversed by factors released from therapeutic cells. 197 Smaller hemorrhagic lesions or damage caused by HBPs may also be promising targets for 198 tissue regeneration approaches. Furthermore, peripheral and central inflammatory processes 199 also contribute to further brain injury after ICH and these mechanisms might make excellent 200 targets for some cell-based therapies.

201

202 Preconditioning of cell transplants

Long-term survival of transplanted cells is an important aspect for approaches that target long-term engraftment of neural stem cells to support or repair damaged neuronal circuits, or for which long-term trophic support is required. While cell survival has been poor in most previous studies, recent advantages were made in the field of cell preconditioning.^{48,49} These techniques can significantly enhance and/or prolong survival of transplanted cells and should be considered for approaches that may benefit thereof.

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210 Behavioral readout parameter selection

Functional tests should be sensitive to detect long-term impairment and treatments effects, but not be affected by repeated testing or compensation.²⁰ Various reaching tasks, foot fault, cylinder and adhesive tests provide quantitative and objective assessment in efficacy studies.¹⁵ Simpler tasks can overestimate treatment effects but are valuable for exclusion of stroke animals with no/minor impairment, stratification regarding impairment severity, and treatment assessment during the acute phase. Appropriate tests should be selected for the respective stroke model, species, scenario, and study duration (Fig. 1B).

Smaller lesions require particularly sensitive and precise behavioral outcome measures. These lesions are more sensitive for functional compensation/spontaneous recovery and impairments may be masked. Automated readout systems carry high specificity and sensitivity and are being increasingly used in neurodegenerative disorders with initial subtle motor deficits.⁵⁰ The supplementary table summarizes information on specific deficits and their

223 measurement in lacunar lesions. Lastly, cognitive impairment and depression are common 224 stroke complications, but at present there is no consensus on which tests to use in animals.

225

226 Safety assessments as a focus

Definitive demonstration of safety across multiple preclinical endpoints will be an invaluable resource when advancing cellular therapies for stroke treatment. The cell administration site should be evaluated for signs of inflammation or edema as well as acute respiratory problems for intravenous delivery to ensure the cell therapy is not inducing local or systemic immune responses. This may include animals with a humanized immune system. When performing repetitive administration of a cells, recipient sensitization (e.g., by lymphocyte proliferation assays), indicating adaptive immune system activation, should be contemplated.

Short- and long-term biodistribution and possible cell engraftment should be evaluated 234 to determine cell persistence, particularly if the intended goal is engraftment. However, cell 235 types exerting paracrine and immunomodulatory mechanisms, or exogenous cells may not 236 persist which is viewed as an attractive component of approaches for which cell survival is not 237 necessarily required. Complete endpoint evaluations of tissues and organ systems should be 238 performed to definitively demonstrate that the cell administration does not have any off-target 239 240 effects. Abnormal tissue growth, tumorigenesis or aberrant ectopic fiber sprouting should be excluded when using pluripotent stem cells or other cell types with high proliferation, 241 differentiation, and fiber projection capabilities. 242

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244 Multicenter trials

Innovative preclinical study designs including MCPTs have been proposed since the last STEPS recommendations. MCPTs mimic the design of large scale, efficacy-centered clinical trials with rigorous implementation of quality assurance measures as performed in clinical research.⁵¹ MCPTs are believed to enhance predictive value and statistical power in preclinical

research, and to provide a close-to-practice assessment of the potential treatment. They may be 249 250 of particular value when assessing cell therapies with mild to moderate impact on stroke (i.e. improvements of 10 to $20\%)^{52}$ or when assessing the impact of multiple therapy-influencing 251 factors. MCPTs can also help to verify the benefit of combination therapies. This requires 252 greatly enhanced statistical power to discriminate the effect of the combination from the impact 253 of the individual therapies (e.g., rehabilitation plus cell therapy). The MCPT concept has been 254 well received throughout the stroke community^{53,54}, and first MCPTs revealed effect sizes being 255 considerably lower than what would have been expected from standard single center preclinical 256 studies.55 257

However, MCPTs are more challenging to harmonize and carry much higher costs than 258 standard study designs. Industry may benefit from MCPTs prior to initiating a clinical study.⁵⁶ 259 The STEPS 4 consortium recommends considering MCPTs as an option when planning a 260 translational research program in cell therapy for stroke. Importantly, NIH recently supported 261 the creation of MCPTs and has launched the Stroke Preclinical Assessment Network (SPAN) 262 program currently focused on multicenter evaluations of acute neuroprotectants as a 263 complementary treatment to recanalization. Industry participation is highly encouraged in 264 SPAN. Experience from the program will be invaluable to learn how MCPTs can be organized 265 266 best to fully benefit from the enhanced power in assessing complex treatments, and how the complex logistics of MCPTs can be mastered. Ideally, successful SPAN activities will serve as 267 a role model for MCPTs in cell therapies. 268

269

270 *Potency assay development and qualification*

A new recommendation from the STEPS group is the development of surrogate potency assays. Demonstrating a direct measurable correlation between a cell therapy and a biomarker or another quantifiable biological process with a beneficial outcome is critical to monitor the hypothesized mechanism of action. Biomarkers for putative mechanisms of action are also

275 critical to regulators for late stage clinical trial authorization. Biomarkers might be used to 276 develop potency assays that should be robust, specific, informative, and reproducible in describing a fundamental biological effect of the expected benefit. Qualified potency assays are 277 "locked down" as part of phase III clinical testing. They need to be transferred and performed 278 279 under Good Manufacturing Practice (GMP) conditions before officially filing for product approval with the Food and Drug Administration in the United States. The development of 280 potency assays during preclinical animal testing is therefore paramount prior to moving cellular 281 therapies into advanced stages of clinical trials. As hypotheses change to reflect advances in the 282 fundamental understanding of how cellular therapies provide benefit, new potency assays 283 should be developed to parallel our understanding of cell-mediated benefits. For example, given 284 285 increasing studies showing how many cell therapies target immune responses after stroke, immunomodulation may be an important potency assay for some cell therapies.⁵⁷ 286

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288 Part III: Concepts for accelerating and improving preclinical research

289 Rethinking content and sequence of preclinical and clinical trials

State of the art preclinical research on cell therapy safety and efficacy takes significant 290 291 time and resources. The broad and increasing spectrum of potential confounders is expected to 292 engender additional budgetary and temporal demands that may severely hamper clinical translation. STEPS 4 discussed options to accelerate preclinical research while giving 293 consideration to the complexity of potential confounding factors. A promising concept is to 294 more clearly discriminate exploratory and confirmatory preclinical research⁵⁸, and to rigorously 295 distinguish the primary goals of phase I/II clinical trials (safety) from later phases (efficacy). 296 This allows a well-orchestrated sequence of preclinical and clinical tests with partially parallel 297 workflows (Fig. 2). 298

299 Once a cell therapeutic paradigm is identified in initial exploratory studies, research 300 activities are divided into two parallel tracks. First, exploratory research in standard rodent

stroke models confirms basic efficacy. Second, confirmative research investigates safety. This 301 302 should also consider the most important comorbidities in the expected patient population, risks exhibited by the approach and the intended route of administration.⁵⁹ Thorough confirmation 303 of safety and basic efficacy then allows proceeding to a phase I/IIa clinical trial which should 304 not have a major focus on efficacy endpoints, but would be powered to confirm safety. 305 Moreover, it should identify predominant profile characteristics of the targeted patient 306 population such as type and frequency of comorbidities, infarct location and size, and co-307 medications. 308

This information is used to design advanced preclinical efficacy tests tailored to the 309 target patient population profile. Ideally, these efficacy studies would be conducted in parallel 310 311 to the phase I/IIa study. They may also be designed to identify subgroups with a pronounced benefit from the particular cell therapy which can be considered in a subsequent phase IIb/III 312 clinical trial. 313

This approach has three major advantages: First, basic and enhanced preclinical efficacy 314 studies can be organized in parallel to preclinical or clinical safety tests, saving valuable time. 315 Second, the sequence of investigations in animal models and patients yields important data that 316 317 will help to identify the most suitable patient populations for efficacy-driven clinical trials. 318 Third, more thorough preclinical efficacy data can be used to design GMP potency assays with a higher predictive value than commonly applied ones. 319

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Cell therapy responders versus non-responders

The STEPS 4 working group recommends storage of tissues and samples from animals 322 that both respond and do not respond to cell therapy. As we learn more about the mechanisms 323 324 of action through which cell therapies provide benefit, we may be able to retrieve stored samples from previous experiments to compare if preclinical responders and non-responders differ 325 326 regarding newly identified or proposed biomarkers or pathways. This enables to refine our

clinical understanding of "responders" or "non-responders" and to better identify patients whocan optimally benefit.

329

330 Preclinical data sharing platforms

A complementary opportunity to handle the increasing complexity of preclinical data 331 are (open) sharing platforms. STEPS 4 participants unanimously agreed that such platforms, 332 also including information from cell therapy cases in patients, are beneficial. Data would be 333 available for benchmarking against other research programs, enhance study power, and 334 facilitate meta-analyses. A central registry and predefinition of common preclinical data 335 elements are required, but can be informed by existing clinical registries. The Collaborative 336 Approach to Meta-Analysis and Review of Animal Data from Experimental Studies 337 (CAMRADES) database is an excellent role model, although a cell therapy registry for stroke 338 must reflect the specific requirements of the community in detail. 339

Original data may be sensitive when related to pending intellectual property or commercial interests. Industry leaders among the STEPS 4 group stressed that such data should enjoy special protection, but is not necessarily excluded from sharing. For instance, the identity of a sensitive cell product could be concealed, but cell-treated subjects as well as all insensitive information on the cell product can be disclosed. Contributors using highly sensitive cell products may at least provide control cases.

Options to motivate contribution to data sharing platforms may be to allow access only to those who contribute and/or a general requirement that publically funded cell therapy research for stroke shall be publically. The STEPS 4 consortium suggests that decision makers at the NIH or the European Commission should consider funding schemes that help realizing data platforms tailored to cell therapies. Ideally, open data registries are organized internationally and provide connection hubs for industry and clinical cell therapy data.

352

353 *Novel collaboration formats and the role of industry*

354 The increasing complexity of preclinical stroke research and the parallel need for acceleration without sacrificing specificity and accuracy may not only require novel research 355 strategies but also novel research alliances. Providing methodological knowhow, flexibility, 356 357 and sufficient funds is required to meet the increasing need for rigor in preclinical research, raising the need for academic-industry alliances. Such alliances should not be restricted to 358 sponsored contract research but true collaboration.⁵⁶ Academic-industry collaborations are also 359 pivotal to sustainably utilize MCPTs. Finally, the experience of industry in meeting regulatory 360 demands, technical aspects of cell therapies, and related logistics as well as clinical trial design 361 is invaluable to inform preclinical research in order to advance the field. The STEPS 4 group 362 363 recommends long-term academic-industry partnerships to thoroughly develop cell therapeutics from bench to bedside through closer collaborations. 364

365

366 **Recommendation summary**

367 1. A stronger focus on safety rather than confirming efficacy in early preclinical
 368 research, followed by early, safety-oriented clinical research has the potential to accelerate
 369 translational research without sacrificing quality.

2. We recommend thorough and advanced safety assessments and sufficient (standard stroke model) efficacy testing to support phase I/II safety trials. Advanced preclinical efficacy testing should be tailored to match targeted patient populations. This approach addresses the increasing complexity of potential confounding factors in a reasonable time. Appropriate primary readout parameters should be chosen for subsequent phase IIb/III trials.

375 3. Specific stroke models should best mimic the targeted patient population. LAMs are376 recommended if they provide additional, crucial information for clinical translation.

4. High priority should be given to developing specific and validated potency assays.
Investigating drug-cell interactions and identifying cell therapy responders versus nonresponders is recommended.

380 5. Sharing preclinical and clinical data will help the community tackle more complex381 research questions (e.g., whether comorbidities affect efficacy or safety).

382 6. Confirmative MCPTs are a valuable confirmative research format, but larger research
 383 consortia including industry joint ventures are required for successful implementation. MCPTs
 384 are preferred prior to definitive efficacy trials

385

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394 **Disclosures**

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418 **References**

- Steinberg GK, Kondziolka D, Wechsler LR, Lunsford LD, Kim AS, Johnson JN, et al.
 Two-year safety and clinical outcomes in chronic ischemic stroke patients after
 implantation of modified bone marrow-derived mesenchymal stem cells (SB623): a
 phase 1/2a study. J Neurosurg. 2018;doi:10.3171/2018.5.JNS173147.
- 423 2. Hess DC, Wechsler LR, Clark WM, Savitz SI, Ford GA, Chiu D, et al. Safety and
 424 efficacy of multipotent adult progenitor cells in acute ischaemic stroke (MASTERS): a
 425 randomised, double-blind, placebo-controlled, phase 2 trial. Lancet Neurol.
 426 2017;16:360-368.
- Moniche F, Rosado-de-Castro PH, Escudero I, Zapata E, de la Torre Laviana FJ,
 Mendez-Otero R, et al. Increasing Dose of Autologous Bone Marrow Mononuclear
 Cells Transplantation Is Related to Stroke Outcome: Results from a Pooled Analysis of
 Two Clinical Trials. Stem Cells Int. 2016;2016:8657173.
- 4. Kalladka D, Sinden J, Pollock K, Haig C, McLean J, Smith W, et al. Human neural stem
 cells in patients with chronic ischaemic stroke (PISCES): a phase 1, first-in-man study.
 Lancet. 2016;388:787-796.
- 434 5. Sandu RE, Balseanu AT, Bogdan C, Slevin M, Petcu E, Popa-Wagner A. Stem cell
 435 therapies in preclinical models of stroke. Is the aged brain microenvironment refractory
 436 to cell therapy? Exp Gerontol. 2017;94:73-77.
- 437 6. Möller K, Pösel C, Kranz A, Schulz I, Scheibe J, Didwischus N, et al. Arterial
 438 Hypertension Aggravates Innate Immune Responses after Experimental Stroke. Front
 439 Cell Neurosci. 2015;9:461.
- Chen J, Ye X, Yan T, Zhang C, Yang XP, Cui X, et al. Adverse effects of bone marrow
 stromal cell treatment of stroke in diabetic rats. Stroke. 2011;42:3551-3558.

- Kui X, Chopp M, Zacharek A, Roberts C, Lu M, Savant-Bhonsale S, et al. Chemokine,
 vascular and therapeutic effects of combination Simvastatin and BMSC treatment of
 stroke. Neurobiol Dis. 2009;36:35-41.
- Boltze J, Nitzsche F, Jolkkonen J, Weise G, Pösel C, Nitzsche B, et al. Concise Review:
 Increasing the Validity of Cerebrovascular Disease Models and Experimental Methods
 for Translational Stem Cell Research. Stem Cells. 2017;35:1141-1153.
- Savitz SI, Cramer SC, Wechsler L, STEPS 3 Consortium. Stem cells as an emerging
 paradigm in stroke 3: enhancing the development of clinical trials. Stroke. 2014;45:634639.
- 451 11. Savitz SI, Chopp M, Deans R, Carmichael T, Phinney D, Wechsler L, et al. Stem Cell
 452 Therapy as an Emerging Paradigm for Stroke (STEPS) II. Stroke. 2011;42:825-829.
- 453 12. Stem Cell Therapies as an Emerging Paradigm in Stroke (STEPS): bridging basic and
 454 clinical science for cellular and neurogenic factor therapy in treating stroke. Stem Cell
 455 Therapies as an Emerging Paradigm in Stroke Participants. Stroke. 2009;40:510-515.
- 456 13. Guzman R, Janowski M, Walczak P. Intra-Arterial Delivery of Cell Therapies for
 457 Stroke. Stroke. 2018;49:1075-1082.
- 458 14. Sutherland BA, Neuhaus AA, Couch Y, Balami JS, DeLuca GC, Hadley G, et al. The
 459 transient intraluminal filament middle cerebral artery occlusion model as a model of
 460 endovascular thrombectomy in stroke. J Cereb Blood Flow Metab. 2016;36:363-369.
- 461 15. Corbett D, Carmichael ST, Murphy TH, Jones TA, Schwab ME, Jolkkonen J, et al.
 462 Enhancing the Alignment of the Preclinical and Clinical Stroke Recovery Research
 463 Pipeline: Consensus-Based Core Recommendations From the Stroke Recovery and
 464 Rehabilitation Roundtable Translational Working Group. Neurorehabil Neural Repair.
 465 2017;31:699-707.
- 466 16. Orset C, Macrez R, Young AR, Panthou D, Angles-Cano E, Maubert E, et al. Mouse
 467 model of in situ thromboembolic stroke and reperfusion. Stroke. 2007;38:2771-2778.

468 17. Chan SL, Bishop N, Li Z1, Cipolla M. Inhibition of PAI (Plasminogen Activator
469 Inhibitor)-1 Improves Brain Collateral Perfusion and Injury After Acute Ischemic
470 Stroke in Aged Hypertensive Rats. Stroke. 2018;49:1969-1976.

- 472 18. Boltze J, Förschler A, Nitzsche B, Waldmin D, Hoffmann A, Boltze CM, et al.
 473 Permanent middle cerebral artery occlusion in sheep: a novel large animal model of
 474 focal cerebral ischemia. J Cereb Blood Flow Metab. 2008;28:1951-1964.
- 475 19. Werner P, Saur D, Zeisig V, Ettrich B, Patt M, Sattler B, et al. Simultaneous PET/MRI
 476 in stroke: a case series. J Cereb Blood Flow Metab. 2015;35:1421-1425.
- 477 20. Hainsworth AH, Allan SM, Boltze J, Cunningham C, Farris C, Head E, et al.
 478 Translational models for vascular cognitive impairment: a review including larger
 479 species. BMC Med. 2017;15:16.
- 480 21. Balkaya MG, Trueman RC, Boltze J, Corbett D, Jolkkonen J. Behavioral outcome
 481 measures to improve experimental stroke research. Behav Brain Res. 2018;352:161482 171.
- 483 22. Cui LL, Kerkelä E, Bakreen A, Nitzsche F, Andrzejewska A, Nowakowski A, et al. The
 484 cerebral embolism evoked by intra-arterial delivery of allogeneic bone marrow
 485 mesenchymal stem cells in rats is related to cell dose and infusion velocity. Stem Cell
 486 Res Ther. 2015;6:11.
- Walczak P, Wojtkiewicz J, Nowakowski A, Habich A, Holak P, Xu J, et al. Real-time
 MRI for precise and predictable intra-arterial stem cell delivery to the central nervous
 system. J Cereb Blood Flow Metab. 2017;37:2346-2358.
- 490 24. Sommer CJ, Schäbitz WR. Fostering Poststroke Recovery: Towards Combination
 491 Treatments. Stroke. 2017;48:1112-1119.
- 492 25. Balseanu AT, Buga AM, Catalin B, Wagner DC, Boltze J, Zagrean AM, et al.
 493 Multimodal Approaches for Regenerative Stroke Therapies: Combination of

- 494 Granulocyte Colony-Stimulating Factor with Bone Marrow Mesenchymal Stem Cells is
 495 Not Superior to G-CSF Alone. Front Aging Neurosci. 2014;6:130.
- 496 26. Pösel C, Scheibe J, Kranz A, Bothe V, Quente E, Fröhlich W, et al. Bone marrow cell
 497 transplantation time-dependently abolishes efficacy of granulocyte colony-stimulating
 498 factor after stroke in hypertensive rats. Stroke. 2014;45(8):2431-2437.
- Cui X, Chopp M, Shehadah A, Zacharek A, Kuzmin-Nichols N, Sanberg CD, et al.
 Therapeutic benefit of treatment of stroke with simvastatin and human umbilical cord
 blood cells: neurogenesis, synaptic plasticity, and axon growth. Cell Transplant.
 2012;21:845-856.
- 503 28. Amer MH, Rose FRAJ, Shakesheff KM, Modo M, White LJ. Translational
 504 considerations in injectable cell-based therapeutics for neurological applications:
 505 concepts, progress and challenges. NPJ Regen Med. 2017;2:23.
- Aguado BA, Mulyasasmita W, Su J, Lampe KJ, Heilshorn SC. Improving viability of
 stem cells during syringe needle flow through the design of hydrogel cell carriers. Tissue
 Eng Part A. 2012;18:806-815.
- 509 30. Nih LR, Carmichael ST, Segura T. Hydrogels for brain repair after stroke: an emerging
 510 treatment option. Curr Opin Biotechnol. 2016;40:155-163.
- Moshayedi P, Nih LR, Llorente IL, Berg AR, Cinkornpumin J, Lowry WE, et al.
 Systematic optimization of an engineered hydrogel allows for selective control of
 human neural stem cell survival and differentiation after transplantation in the stroke
 brain. Biomaterials. 2016;105:145-155.
- 32. Bible E, Dell'Acqua F, Solanky B, Balducci A, Crapo PM, Badylak SF, et al. Noninvasive imaging of transplanted human neural stem cells and ECM scaffold remodeling
 in the stroke-damaged rat brain by (19)F- and diffusion-MRI. Biomaterials.
 2012;33:2858-2871.

- Nih LR, Sideris E, Carmichael ST, Segura T. Injection of Microporous Annealing 519 33. 520 Particle (MAP) Hydrogels in the Stroke Cavity Reduces Gliosis and Inflammation and NPC 521 Promotes Migration to the Lesion. Adv Mater. 2017;doi: 10.1002/adma.201606471. 522
- 523 34. Ghuman H, Massensini AR, Donnelly J, Kim SM, Medberry CJ, Badylak SF, et al. ECM
 524 hydrogel for the treatment of stroke: Characterization of the host cell infiltrate.
 525 Biomaterials. 2016;91:166-181.
- 35. Bible E, Chau DY, Alexander MR, Price J, Shakesheff KM, Modo M. The support of
 neural stem cells transplanted into stroke-induced brain cavities by PLGA particles.
 Biomaterials. 2009;30:2985-2994.
- 529 36. Conway A, Vazin T, Spelke DP, Rode NA, Healy KE, Kane RS, et al. Multivalent
 530 ligands control stem cell behaviour in vitro and in vivo. Nat Nanotechnol. 2013;8:831531 838.
- 37. Bible E, Qutachi O, Chau DY, Alexander MR, Shakesheff KM, Modo M. Neovascularization of the stroke cavity by implantation of human neural stem cells on
 VEGF-releasing PLGA microparticles. Biomaterials. 2012;33:7435-7446.
- 535 38. Cook DJ, Nguyen C, Chun HN, L Llorente I, Chiu AS, Machnicki M, et al. Hydrogel536 delivered brain-derived neurotrophic factor promotes tissue repair and recovery after
 537 stroke. J Cereb Blood Flow Metab. 2017;37:1030-1045.
- 538 39. Emerich DF, Silva E, Ali O, Mooney D, Bell W, Yu SJ, et al. Injectable VEGF hydrogels
 539 produce near complete neurological and anatomical protection following cerebral
 540 ischemia in rats. Cell Transplant. 2010;19:1063-1071.
- 40. Zhang YX, Yuan MZ, Cheng L, Lin LZ, Du HW, Chen RH, et al. Treadmill exercise
 enhances therapeutic potency of transplanted bone mesenchymal stem cells in cerebral
 ischemic rats via anti-apoptotic effects. BMC Neurosci. 2015;16:56.

- Sasaki Y, Sasaki M, Kataoka-Sasaki Y, Nakazaki M, Nagahama H, Suzuki J, et al.
 Synergic Effects of Rehabilitation and Intravenous Infusion of Mesenchymal Stem Cells
 After Stroke in Rats. Phys Ther. 2016;96(11):1791-1798.
- 547 42. Bernhardt J, Borschmann KN, Kwakkel G, Burridge JH, Eng JJ, Walker MF, et al.
 548 Setting the scene for the Second Stroke Recovery and Rehabilitation Roundtable. Int J
 549 Stroke. 2019;doi:10.1177/1747493019851287.
- Mu J, Bakreen A, Juntunen M, Korhonen P, Oinonen E, Cui L, et al. Combined Adipose
 Tissue-Derived Mesenchymal Stem Cell Therapy and Rehabilitation in Experimental
 Stroke. Front Neurol. 2019;10:235.50. Preisig DF, Kulic L, Krüger M, Wirth F,
 McAfoose J, Späni C, et al. High-speed video gait analysis reveals early and
 characteristic locomotor phenotypes in mouse models of neurodegenerative movement
 disorders. Behav Brain Res. 2016;311:340-353.
- 556 51. Dirnagl U, Fisher M. International, multicenter randomized preclinical trials in
 557 translational stroke research: it's time to act. J Cereb Blood Flow Metab. 2012;32:933558 935.
- 559 52. Macleod MR, van der Worp HB, Sena ES, Howells DW, Dirnagl U, Donnan GA.
 560 Evidence for the efficacy of NXY-059 in experimental focal cerebral ischaemia is
 561 confounded by study quality. Stroke. 2008;39:2824-2829.
- 562 53. Boltze J, Ayata C, Wagner DC, Plesnila N. Preclinical phase III trials in translational 563 stroke research: call for collective design of framework and guidelines. Stroke. 564 2014;45:357.
- 565 54. Boltze J, Wagner DC, Henninger N, Plesnila N, Ayata C. Phase III Preclinical Trials in
 566 Translational Stroke Research: Community Response on Framework and Guidelines.
 567 Transl Stroke Res. 2016;7:241-247.

- 568 55. Llovera G, Hofmann K, Roth S, Salas-Pérdomo A, Ferrer-Ferrer M, Perego C, et al.
 569 Results of a preclinical randomized controlled multicenter trial (pRCT): Anti-CD49d
 570 treatment for acute brain ischemia. Sci Transl Med. 2015;7:299ra121.
- 571 56. Boltze J, Wagner DC, Barthel H, Gounis MJ. Academic-industry Collaborations in
 572 Translational Stroke Research. Transl Stroke Res. 2016;7:343-353.
- 573 57. Mays RW, Savitz SI. Intravenous Cellular Therapies for Acute Ischemic Stroke. Stroke.
 574 2018;49:1058-1065.
- 575 58. Dirnagl U, Hakim A, Macleod M, Fisher M, Howells D, Alan SM, et al. A concerted
 576 appeal for international cooperation in preclinical stroke research. Stroke.
 577 2013;44:1754-1760.
- 578 59. Boltze J, Arnold A, Walczak P, Jolkkonen J, Cui L, Wagner DC. The Dark Side of the
 579 Force Constraints and Complications of Cell Therapies for Stroke. Front Neurol.
 580 2015;6:155.

584 Figure 1. Functional improvement by neurorehabilitation and recommended readout 585 parameters.

(A) Schematic time course of spontaneous functional recuperation (light grey line), 586 functional improvement with cell therapy alone (grey line), and with additional, appropriately 587 timed supportive rehabilitation (black line). The relatively small differences between the 588 therapy groups may require large sample sizes. (B) Behavioral tests differ with respect to 589 sensitivity and specificity. Simple tests detect relatively large deficits in the acute and subacute 590 591 stage. More sensitive tests address particular sensory and motor functions. Elaborated, often 592 highly automated tests reveal very fine motor and sensory differences, or mental/cognitive impairment following stroke. 593

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595 Figure 2. Proposed concept for accelerated clinical translation.

The basic suggestion of the concept is to initially focus on thorough and advanced safety 596 assessments. Exploratory (basic) efficacy results warrant entering an early stage, safety-597 oriented clinical trial (phase I/IIa). This trial should also retrieve important characteristics of 598 599 the target patient population, directly informing the design of more advanced, confirmative preclinical efficacy study (optionally followed by a multicenter preclinical trial) and of tailored 600 potency assays. Those allow moving forward to clinical efficacy studies (phase IIb/III) tailored 601 602 to the expected patient population, but in less time as would be required by sequential research programs. Regulatory authorities should be consulted regularly to ensure adequate planning of 603 each parallel step. 604