Advances in the treatment of mitochondrial epilepsies

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Abstract

Epilepsy is frequently a severe and sinister symptom in primary mitochondrial diseases, a group of more than 350 different genetic disorders characterised by mitochondrial dysfunction and extreme clinical and biochemical heterogeneity. Mitochondrial epilepsy is notoriously difficult to manage, principally because the vast majority of primary mitochondrial diseases currently lack effective therapies. Treating the underlying mitochondrial disorder is likely to be a more effective strategy than using traditional antiepileptic drugs. This review, initially presented at the 7th London-Innsbruck Colloquium on Status Epilepticus and Acute Seizures at the Francis Crick Institute in London, summarises the currently available and emerging therapies for mitochondrial epilepsy. Potentially treatable mitochondrial diseases include disorders of coenzyme Q_{10} biosynthesis and a group of mitochondrial respiratory chain complex I subunit and assembly factor defects that respond to riboflavin (vitamin B2). Approaches that have been adopted in actively recruiting clinical trials include redox modulation, harnessing mitochondrial biogenesis, using rapamycin to target mitophagy, nucleoside supplementation, and gene and cell therapies. Most of the clinical trials are at an early stage (Phase 1 or 2) and none of the currently active trials is specifically targeting mitochondrial epilepsy.
**Introduction**

Primary mitochondrial diseases have recently been redefined as disorders caused by mutations that “primarily or secondarily lead to oxidative phosphorylation (OXPHOS) dysfunction or other disturbances of mitochondrial structure and function including perturbed mitochondrial ultrastructure, aberrant synthesis of cofactors and vitamins, or other impaired metabolic processes within the mitochondrion” [1]. Epilepsy is reported to affect up to 40% of patients with primary mitochondrial disease [2] and is often a late and sinister clinical feature; 50% of affected patients died within 9 months after seizure onset in one cohort [3]. Unfortunately there are currently no effective therapies for the vast majority of mitochondrial diseases, and this includes the mitochondrial epilepsies. This brief review, originally presented on 9 April 2019 at the 7th London-Innsbruck Colloquium on Status Epilepticus and Acute Seizures at the Francis Crick Institute in London, summarises the currently available treatments for mitochondrial epilepsy and emerging therapies either undergoing clinical trial or on the horizon.

**Antiepileptic drugs and supportive management**

Many mitochondrial epilepsies are notoriously resistant to antiepileptic drugs (AEDs), and patients have been reported to be resistant to many different combinations of drugs. There continue to be anecdotal case reports and small case series reporting benefit of specific AEDs in patients with mitochondrial epilepsies (e.g. [4, 5]), but the clinical, biochemical and genetic heterogeneity of mitochondrial epilepsies, and the unpredictable natural course, makes interpretation of such reports extremely challenging. In an ideal world the primary treatment should clearly be directed at the underlying cause rather than waiting to treat seizures when they occur. However at present there are no disease-modifying therapies for the vast majority of mitochondrial diseases, and supportive treatments remain the principal therapies offered to affected patients. These include AEDs and treatment of multisystemic disease complications when they occur [6].

**Cofactor and vitamin responsive disorders**

There are some notable exceptions of primary mitochondrial disorders where disease-modifying therapies do exist, and it is important to recognise these disorders promptly. The most well-known subgroup comprises disorders of coenzyme Q\(_{10}\) (ubiquinone) biosynthesis, which at least in theory are likely to be ameliorated by supplementation with pharmacological doses of coenzyme Q\(_{10}\) [7]. Coenzyme Q\(_{10}\) functions as a mobile electron carrier within the respiratory chain, as well as being a potent antioxidant. Coenzyme Q\(_{10}\) at 30mg/kg/day was reported to prevent renal impairment in a child with COQ2 mutations whose sibling had end-stage renal failure, seizures, stroke-like episodes and cognitive impairment [8]. However a more recent
report described progressive neurological dysfunction, including seizures and encephalopathy, in patients with COQ2 deficiency whose initial symptoms of diabetes and nephrotic syndrome had responded to coenzyme Q10 supplementation in the neonatal period [9]. Progression of neurological disease may be because of limited intracerebral bioavailability of coenzyme Q10. Another issue is that other coenzyme Q10 biosynthesis defects, particularly COQ4 and COQ9 mutations, appear to have prenatal onset and are associated with severe disease progression and early death, despite prompt initiation of coenzyme Q10 supplementation [10-13], so other treatment strategies may be needed for this group of disorders.

Another group of potentially treatable mitochondrial disorders are those that respond to supplementation with riboflavin (vitamin B2) [14]. These include some complex I deficiencies, particularly mutations of ACAD9, encoding a flavoprotein required for complex I assembly. Affected patients typically have a good clinical response to riboflavin supplementation [15]. The most frequent clinical features associated with ACAD9 deficiency are hypertrophic cardiomyopathy, lactic acidosis and exercise intolerance, but seizures have been reported in occasional cases [16, 17]. Mutations of AIFM1, encoding apoptosis inducing factor mitochondria associated 1, another flavoprotein required for complex I assembly, have been reported to cause riboflavin-responsive ataxia in two patients [18]. AIFM1 mutations have also been associated with seizures, but riboflavin therapy was not reported in these individuals [19]. Riboflavin may also be beneficial in other complex I deficiencies, particularly mutations of the NDUFV1 and NDUFV2 subunits which are flavoproteins interacting with the flavin mononucleotide (FMN) active cofactor of the complex I holoenzyme [14].

The role of vitamins and cofactors in other mitochondrial disorders is uncertain. There have been very few randomised clinical trials in primary mitochondrial diseases, and a Cochrane review published in 2012 concluded that further research is needed in this area [20].

Development of novel therapies

Development of novel therapies for mitochondrial disease is a growth industry. However, despite more than 2000 publications per year on mitochondrial disease treatment there are currently no licensed treatments for systemic mitochondrial disease. Why is it so difficult to treat mitochondrial disease? The multi-layered complexity of mitochondrial disease, with extreme clinical, biochemical and genetic heterogeneity, provides an enormous barrier to therapy development. However, despite these difficulties, a large number of candidate therapies is currently in development [1, 21].

Novel therapies in development for mitochondrial disease may be divided broadly into two groups: pharmacological and genetic treatments. Many of the pharmacological approaches are generic 'disease
agnostic’ approaches and include targeting reactive oxygen species, stabilising the mitochondrial membrane, harnessing mitochondrial biogenesis and targeting mitophagy (Figure 1).

**Antioxidant approaches**

Antioxidants have played a central role in the (attempted) treatment of mitochondrial disease for decades, although evidence for their efficacy remains limited at present [20]. The central tenet underlying their use is that impairment of mitochondrial function, especially of complexes I and III, leads to excessive generation of reactive oxygen species (ROS), which in turn leads to a vicious cycle of further mitochondrial damage, including lipid, protein and DNA peroxidation. ROS include the superoxide radical and hydrogen peroxide. Accumulating evidence implicating ROS in seizure generation outside the context of primary mitochondrial disease [22, 23] supports the continuing use of antioxidants in mitochondrial epilepsies, in the absence of more specific disease-modifying therapies. A word of caution is that ROS are not merely damage-inducing molecules but rather have important physiological roles in the mitochondrion and in the cell, especially in inter-organellar signalling [24]. One study suggested that excessive antioxidant exposure could be harmful in a mouse model of mitochondrial disease; a muscle-specific COX15 knockout had worse survival following N-acetylcysteine (NAC) therapy [25]. The antioxidants used in clinical practice include coenzyme Q10, NAC and vitamins E and C. Newer antioxidants that have been investigated in preclinical studies include EPI-743 and KH176. Open label studies of EPI-743 in the mitochondrial encephalomyelopathy Leigh syndrome did not conclusively demonstrate benefit [26, 27]. The KHENERGY study, a double-blind, randomized, placebo-controlled, two-way crossover phase IIA study of KH176, demonstrated tolerability and safety in adult patients with relatively mild multisystemic disease related to the m.3243A>G mutation (average heteroplasmy 61% in urinary epithelial cells, 18% in leukocytes) [28]. A phase III study is planned but not yet active.

**Harnessing mitochondrial biogenesis**

The process of making new mitochondria, known as mitochondrial biogenesis, is ultimately regulated by the master transcriptional co-activator PGC1α, which interacts with multiple transcription factors including the nuclear respiratory factors NRF1 and NRF2 and the peroxisome proliferator-activated receptors (PPARs), leading to increased transcription of hundreds of genes encoding mitochondrial components including OXPHOS subunits and assembly factors and fatty acid oxidation enzymes [29]. Theoretically, increasing mitochondrial mass by stimulating mitochondrial biogenesis should result in a net increase in energy production and thus potentially ameliorate at least some of the effects of mitochondrial dysfunction. Several therapeutic approaches have attempted to manipulate this process by activating PGC1α by modulating its
acetylation (e.g. by activating the SIRT1 deacetylase) or phosphorylation (e.g. via the AMP-activated protein kinase) or by increasing PGC1α target molecules such as the PPARs (e.g. with decanoic acid). As with antioxidant therapies, evidence for efficacy of these approaches is currently limited. Initial studies of bezafibrate, resveratrol and AICAR in animal models yielded conflicting data [30]. A clinical trial of resveratrol in mitochondrial myopathies is ongoing (NCT03728777, Table 1) and a trial of bezafibrate has recently completed but is yet to report its results (NCT02398201, clinicaltrials.gov). We observed promising preclinical data in human patient cell models of complex I deficiency treated with decanoic acid (C10) [31], but this has not yet been translated into a clinical trial for primary mitochondrial disease. Increasing NAD+, either by supplementing with the vitamin B3 derivative nicotinamide riboside or by inhibiting poly(ADP-ribose) polymerase 1 (PARP1, which consumes NAD+), may also promote mitochondrial biogenesis by activating SIRT1. Increasing NAD+ availability ameliorated the phenotypes of two mouse models of mitochondrial disease [32, 33], and improved mitochondrial function in human complex I deficient fibroblasts [34]. Two actively recruiting trials are investigating an NAD+ modulator KL 1333 (NCT03888716) and nicotinamide riboside (NCT03432871) whilst a third aims to use a novel PPAR agonist REN001 (NCT03862846) to increase mitochondrial biogenesis (Table 1). It is interesting to note that all of the active trials investigating mitochondrial biogenesis as a therapeutic strategy are targeting mitochondrial myopathies; none are aiming to treat mitochondrial epilepsies, possibly because of the challenges inherent in targeting molecules to the central nervous system.

Targeting mitophagy

Inhibition of the mTOR pathway by rapamycin has been used to target mitophagy, the process of selective elimination of dysfunctional mitochondria. Several recent preclinical studies have reported beneficial effects of rapamycin in treating mitochondrial disease; high-dose intraperitoneal rapamycin increased survival and attenuated the neurological phenotype and brain lesions of \textit{Ndufs4}-/- mice, a murine model of Leigh syndrome, and also improved the phenotype of myopathic ‘deletor’ mice harbouring a mutation in the twinkle DNA helicase, whilst low dose rapamycin increased survival of Tk2 deficient mice [35-37]. In contrast, there was no evidence of benefit from rapamycin in an encephalomyopathic mouse model harbouring a homozygous nonsense mutation in the coenzyme Q\textsubscript{10} biosynthetic factor Coq9 [38]. Switching immunosuppression from calcineurin inhibitors to mTOR inhibitors was reported to lead to clinical improvement in four renal transplant patients with mitochondrial disease related to the m.3243A>G mutation (associated with mitochondrial encephalomyopathy with lactic acidosis and stroke-like episodes, MELAS) [39]. Recently investigators in New York treated two patients with mitochondrial seizures with everolimus, a rapamycin analogue. The authors reported apparent benefit of everolimus in the first child, who had Leigh
syndrome caused by a homozygous NDUFS4 mutation, but the second child, who had m.3243A>G MELAS, had progressive disease despite everolimus therapy and subsequently died [40]. The mechanisms underlying the potential efficacy of rapamycin in mitochondrial disease remain unknown. Modulation of mitophagy is the most favoured mechanism but other possibilities include increased lysosomal biogenesis and metabolic reprogramming to reduce the dependence on OXPHOS [40]. The mTOR pathway has also been linked to folate availability, which is interesting since folate is a key player in one-carbon metabolism, a metabolic pathway implicated in the pathogenesis of mitochondrial disease [36, 41, 42]. The variable findings of the preclinical and compassionate use data suggest that rapamycin may not be a universal panacea for mitochondrial disease. A formal clinical trial is needed to determine which patients may benefit from rapamycin or everolimus and to establish the lowest efficacious dose, in view of the known adverse effects associated with these drugs, including immunosuppression and hyperlipidaemia. A clinical trial of NAB-sirolimus (nanoparticle albumin-bound rapamycin) in genetically confirmed Leigh syndrome is currently active although not yet recruiting patients (NCT03747328, Table 1).

**Hypoxia**

A genome-wide CRISPR screen identified the Von Hippel Lindau (VHL) ubiquitin ligase as a key factor rescuing mitochondrial dysfunction in a cellular model, thus implicating the hypoxia inducible factor (HIF) transcriptional pathway as a therapeutic target in mitochondrial disease [43]. The authors went on to demonstrate that maintaining the Ndufs4-/- Leigh syndrome mice in chronic hypoxic conditions (11% O₂) prolonged survival and attenuated the brain lesions observed in these mutant mice [43]. A more recent study by the same group has shown that the mechanism of neuroprotection by hypoxia in the Ndufs4-/- mouse is by prevention of brain tissue hyperoxia rather than by activation of the HIF pathway [44]. Chronic hypoxia is obviously not a therapy of choice for human patients, so clinical translation of this treatment will be challenging. However, an immediate clinical message is that hyperoxia should be avoided in mitochondrial patients in the intensive care unit [45, 46], including those with mitochondrial status epilepticus.

**Other pharmacological approaches**

Another pharmacological approach is to stabilise the inner mitochondrial membrane lipid milieu in which the OXPHOS complexes and supercomplexes are embedded, in order to preserve mitochondrial function. Elamipretide (SS-31, MTP-131) is a Szeto-Schiller tetrapeptide (D-Arg-dimethylTyr-Lys-Phe-NH2) that appears to stabilise cardiolipin, the major lipid component of the inner mitochondrial membrane. A recently completed clinical trial of elamipretide in adults with primary mitochondrial myopathy, the MMPOWER study, reported
improvements in the 6 minute walk test after 5 days of treatment [47]. A phase 3 double-blind placebo-controlled trial of elamipretide in mitochondrial myopathy is due to start recruiting patients shortly (NCT03323749, Table 1). Intriguingly, the small molecule nature of elamipretide means that it is able to cross the blood brain barrier (BBB) [48], which makes it potentially a relevant molecule to treat mitochondrial epilepsies.

Stroke-like episodes (SLEs) are increasingly recognised to represent seizure activity. Management of the canonical mitochondrial stroke syndrome MELAS is extremely challenging [49]. Agents which have aimed to address SLEs in MELAS include L-arginine, L-citrulline, L-taurine and succinate, but high level evidence of efficacy is currently lacking. Nitric oxide deficiency has been implicated in the pathogenesis of SLEs and there was apparent benefit of the nitric oxide precursor L-arginine in both prevention and amelioration of SLEs in a series of open-label studies performed in Japan [50, 51]. As a consequence of these open-label studies, L-arginine use in MELAS has become widespread across the globe even though a randomised placebo-controlled trial has never been performed. A dose-finding study of L-citrulline, another precursor of nitric oxide, in MELAS is currently in progress (NCT03952234, Table 1).

Nucleosides have shown efficacy in preclinical trials to treat myopathic mitochondrial DNA depletion syndrome (MDDS) caused by thymidine kinase 2 (TK2) deficiency [52], and a compassionate use study recently reported beneficial effects in affected children and adults [53]. A Phase 2 open label extension of combination pyrimidine nucleosides in TK2 deficiency is ongoing (NCT03845712). The ability of nucleosides to penetrate the BBB is not known, and whether these molecules would successfully treat seizures in encephalomyopathic forms of MDDS remains controversial.

Genetic and cellular therapies

Genetic therapies need to target the mtDNA or nuclear genome, depending on the specific mitochondrial disorder. Adeno-associated viral (AAV) vector mediated gene therapy has been reported in several preclinical models of nuclear-encoded mitochondrial disease, including mouse models of mitochondrial neurogastrointestinal encephalomyopathy (MNGIE) and ETHE1, MPV17 and NDUFS4 deficiencies [54-57]. These have shown generally positive effects and could potentially be a useful therapeutic strategy for all nuclear-encoded mitochondrial diseases. However, the limiting factor at present is the enormous expense and time requirement to develop and treat preclinical models for >300 different gene defects. Unless new methods are developed to scale up these treatments, or regulatory agencies relax the preclinical therapy requirements in some way, it is unrealistic to expect that gene therapy will ever be available for the large number of ultra-rare mitochondrial gene defects affecting only a few individuals each worldwide. CRISPR-based gene editing
was used to rescue mitochondrial dysfunction in an induced pluripotent stem cell model of coenzyme Q\textsubscript{10} deficiency [58], and this may be another therapeutic strategy for nuclear-encoded mitochondrial diseases going forwards.

Targeting the mitochondrial genome is more challenging, since it is relatively inaccessible owing to its protected position within the mitochondrion, encased by two lipid membranes. However, recent approaches using mitochondrial TALENs and zinc finger nucleases to target and selectively eliminate mutant mtDNA sequences have shown benefit in both cell and animal models of heteroplasmic mtDNA disease [59-62]. A concern is that selective destruction of mutant mtDNA at high heteroplasmy level may lead to a temporary state of mtDNA depletion before the wild-type mtDNA recovers to normal copy numbers, and patients would be extremely vulnerable during this recovery period. Strategies to tackle this problem are awaited, but it is theoretically possible that nucleoside supplementation may hasten the recovery of the wild-type mtDNA in this situation.

Another gene therapy approach, currently in clinical trial, is to treat LHON by intra-ocular injection of AAV2-ND4 recoded in the nuclear genetic code so that it can be expressed from the nucleus, the recombinant protein being synthesised on cytosolic ribosomes and subsequently imported into the mitochondrion (NCT02161380, Table 1) [63]. How this could be adapted for treatment of mitochondrial epilepsies, which would need at least systemic gene therapy if not intrathecal or intra-cerebroventricular injection, remains to be determined.

Organ transplantation as a genetic rescue for mitochondrial disease has been explored for MNGIE using either allogeneic haematopoietic stem cell transplantation (AHSCT - NCT02427178, Table 1) or liver transplantation, and for ETHE1 deficiency (liver transplantation) [64-66]. A trial of erythrocyte-encapsulated thymidine phosphorylase enzyme replacement therapy in MNGIE is about to start recruiting patients (NCT03866954, Table 1) [67]. Another study is examining the effects of transplantation of autologous CD34+ haematopoietic stem cells enriched with maternal blood-derived mitochondria (MNV-BLD) in children with the Pearson marrow pancreas syndrome (NCT03384420).

**Concluding remarks**

Recent years have seen a dramatic increase in the number of novel therapies under development for primary mitochondrial disease. At the time of writing (24 August 2019) a search for ‘mitochondrial diseases’ identified 215 trials in clinicaltrials.gov, of which at least 8 are interventional studies actively recruiting patients (Table 1). Although none of the active trials are specifically targeting mitochondrial epilepsy, this is an exciting time for mitochondrial disease, and the possibility of effective therapies for these devastating disorders is finally on the horizon.
Acknowledgements

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References


Figure legend

Figure 1: Emerging therapies for mitochondrial disease

Approaches to treat primary mitochondrial diseases that are currently in development include redox modulation using antioxidants, harnessing mitochondrial biogenesis, stabilising the mitochondrial membrane lipid cardiolipin with elamipretide, targeting mitophagy using rapamycin, nucleoside supplementation to bypass the molecular defect in thymidine kinase 2 deficiency, heteroplasmy shifting using MitoTALENs and zinc finger nucleases, and gene and cell therapies.
Table 1: Selection of currently active interventional trials for primary mitochondrial disorders

<table>
<thead>
<tr>
<th>Agent (other names)</th>
<th>Trial number</th>
<th>Mechanism of action</th>
<th>Disorder</th>
<th>Age range</th>
<th>Design</th>
<th>Primary outcome measures</th>
<th>Status</th>
<th>Sponsor</th>
<th>Type of study</th>
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<td>KL 1333</td>
<td>NCT03888716</td>
<td>NAD+ modulator (mitochondrial biogenesis)</td>
<td>Mitochondrial diseases</td>
<td>18-75y</td>
<td>Phase 1a/1b Randomised, placebo-controlled</td>
<td>Safety</td>
<td>Recruiting</td>
<td>NeuroVive</td>
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<td>Mitochondrial respiratory chain deficiencies</td>
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<td>Nicotinamide riboside</td>
<td>NCT03432871</td>
<td>Increase NAD+ (mitochondrial biogenesis)</td>
<td>Mitochondrial diseases</td>
<td>18-70y</td>
<td>Phase 1 Open label</td>
<td>Bioavailability, safety, mitochondrial biogenesis (assessed by muscle biopsy)</td>
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<td>REN001</td>
<td>NCT03862846</td>
<td>PPARδ agonist (mitochondrial biogenesis)</td>
<td>Primary mitochondrial myopathy</td>
<td>≥16y</td>
<td>Phase 1 Open label</td>
<td>Safety and tolerability (assessed by number of participants with adverse events)</td>
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<td>Resveratrol</td>
<td>NCT03728777</td>
<td>Mitochondrial biogenesis</td>
<td>Mitochondrial myopathies</td>
<td>18-80y</td>
<td>Double-blind, randomized, placebo-controlled,</td>
<td>Decrease in heart rate during constant load</td>
<td>Recruiting</td>
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<td>Sodium phenylbutyrate</td>
<td>NCT03734263</td>
<td>Stabilisation of PDHc enzyme</td>
<td>• PDHc deficiency</td>
<td>3m-18y</td>
<td>Phase 1/2 Open label</td>
<td>Blood lactate level</td>
<td>Recruiting</td>
<td>Fondazione Telethon</td>
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<td>Dichloroacetate</td>
<td>NCT02616484</td>
<td>Decrease inactivation of PDHc by inhibiting PDK</td>
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<td>6m-17y</td>
<td>Phase 3 Randomised, placebo-controlled, quadruple-masked, crossover study</td>
<td>Observer reported outcome (ObsRO) measure of health</td>
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<td>scAAV2-P1ND4v2</td>
<td>NCT02161380</td>
<td>Adeno-associated virus vector gene therapy</td>
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<td>≥15y</td>
<td>Phase 1 Open-label dose escalation study</td>
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<td>AHSCT</td>
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<td>Allogeneic hematopoietic stem cell transplant</td>
<td>• MNGIE</td>
<td>5-55y</td>
<td>Phase 1 Engraftment success (neutrophil count)</td>
<td>Recruiting</td>
<td>Michio Hirano, Cornell University, National Institute of Neurological Disorders and Stroke</td>
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<td>MT1621 (Combination pyrimidine nucleosides)</td>
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<td>Nucleoside supplementation</td>
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<td>‘Mitochondria augmentation therapy’</td>
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<td>1-18y</td>
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<td>Open label Long term effect on disease severity (NPMDS)</td>
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<td>Phase 2A</td>
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<td>Industry</td>
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</tr>
<tr>
<td><strong>Elamipretide</strong> (MTP-131)</td>
<td>NCT03323749</td>
<td>Stabilisation of cardiolipin</td>
<td>16-80y</td>
<td>Phase 3</td>
<td>Randomized, double-blind, parallel-group, placebo-controlled 6MWT, PMMSA fatigue score</td>
<td>Active, not yet recruiting</td>
<td>Stealth BioTherapeutics Inc.</td>
<td>Industry</td>
<td></td>
</tr>
<tr>
<td><strong>L-citrulline</strong></td>
<td>NCT03952234</td>
<td>Nitric oxide precursor</td>
<td>18-65y</td>
<td>Phase 1</td>
<td>Dose-finding Incidence of dose limiting toxicities, to establish</td>
<td>Active, not yet recruiting</td>
<td>Baylor College of Medicine, National Institutes of Medicine</td>
<td>Investigator led</td>
<td></td>
</tr>
<tr>
<td>Erythrocyte encapsulated thymidine phosphorylase (EE-TP)</td>
<td>NCT03866954</td>
<td>Enzyme replacement</td>
<td>MNGIE</td>
<td>≥12y</td>
<td>Phase 2 Open label (sequential assignment)</td>
<td>Safety</td>
<td>Active, not yet recruiting</td>
<td>St George's, University of London, The Clinical Trial Company, Orphan Technologies Ltd</td>
<td>Investigator led/Industry</td>
</tr>
</tbody>
</table>

**Key:** 6MWT six minute walk test, AHSCT allogeneic hematopoietic stem cell transplant, GMFM gross motor function measure, m months, LHON Leber hereditary optic neuropathy, MELAS mitochondrial encephalomyopathy with lactic acidosis and stroke-like episodes, MNGIE mitochondrial neurogastrointestinal encephalomyopathy, NPMDS Newcastle paediatric mitochondrial disease scale, PDHc pyruvate dehydrogenase complex, PDK pyruvate dehydrogenase kinase, PMMSA Primary Mitochondrial Disease Symptom Assessment, TK2 thymidine kinase 2, y years
Harnessing mitochondrial biogenesis

Gene & cell therapies

Antioxidant approaches

Vitamin & cofactors

Targeting mitophagy

Stabilisation of cardiolipin

Heteroplasmic shifting
MitoTALENs & ZFNs

Nucleoside supplementation