

Plasma neurofilament light chain concentration is increased and correlates with the severity of neuropathy in hereditary transthyretin amyloidosis

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Abstract

Hereditary transthyretin amyloidosis (ATTRm) causes a disabling peripheral neuropathy as part of a multi-system disorder. The recent development of highly effective gene silencing therapies has highlighted the need for effective biomarkers of disease activity to guide the decision of when to start and stop treatment. In this study we measured plasma neurofilament light chain (pNfL) concentration in 73 patients with ATTR and found that pNfL was significantly raised in ATTRm patients with peripheral neuropathy compared to healthy controls. Furthermore, pNfL correlated with disease severity as defined by established clinical outcome measures in 26 patients for whom this information was available. These findings suggest a potential role of pNfL in monitoring clinical progression in ATTRm patients.

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Biomarkers

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Introduction

Hereditary transthyretin amyloidosis is an aggressive, multisystem disorder due to mutations in the *TTR* gene that promote the deposition of TTR amyloid with a propensity for the heart and peripheral and autonomic nervous systems¹. More than 90% of TTR is synthesised in the liver. As TTR is a relatively redundant protein, therapeutic strategies have focused on suppressing total TTR production from the liver in an effort to reduce the rate of amyloid deposition and disease progression. In 2018, two genetic therapies (an antisense oligonucleotide and a RNAi) both achieved their primary outcome measures in a trial of ATTRm peripheral neuropathy and have been approved by the FDA and EMA^{2,3}. As ATTRm has a wide range of age of onset with some mutations not being fully penetrant. One of the most pressing issues in the current management of ATTRm is both to diagnose ATTRm as early as possible and to define the optimum time to start gene silencing therapy. Clinical examination and outcome measures, such as the neuropathy impairment score (NIS), will detect the development of a significant large fibre neuropathy but unfortunately, by the time the neuropathy is clinically evident, the disease often progresses rapidly and irreversible disability is acquired⁴.

There is therefore a need for a biomarker that can detect the onset of peripheral neuropathy before the emergence of significant clinical disability. Neurofilaments are the major cytoskeletal proteins of neurons and comprise a light, medium and heavy chain. Following axonal damage, neurofilaments are released into the circulation and can be detected in the plasma using highly sensitive analytical methods such as the SIMOA platform⁵. We have previously shown that pNFL is raised in patients with Charcot-Marie-Tooth (CMT) disease and that it correlates with disease severity⁶. In this study we sought to determine whether pNFL was also raised in patients with ATTRm neuropathy compared to controls and if it correlates with disease severity.

Subjects and Methods

Blood samples were collected from neurologically symptomatic and asymptomatic ATTRm patients attending the UK National Amyloid Centre and the National Hospital for Neurology and Neurosurgery. Control samples were collected from spouses or friends of patients as part of a separate study. The severity of the neuropathy was recorded using the Rasch modified CMT examination and the neuropathy impairment scores within four months of plasma collection^{7,8,9}. This study was approved by the NHNN Research Ethics Committee (REC)/ Central London (09/H0716/61), the East London and the City REC (09/H0703/27) and Royal Free Hospital and University College Medical School REC (06/Q0501/42). Written informed consent was obtained from all participants in the study. Genetic testing was performed according to standard protocols (details are available upon request).

Laboratory Markers

Blood was collected into EDTA-containing tubes, centrifuged at 20°C at 3,500 rpm for 10 minutes and stored within 1 hour at -80°C until analysis. Plasma NfL concentration was determined using an in-house Simoa NfL assay, as previously described¹⁰.

Statistical Analysis

Plasma NfL concentration and age was compared using a 2-sided Mann-Whitney U test and correlations were determined using Pearson's and Spearman's correlation co-efficient, at a significance level of 5%. Multiple linear regression was performed to model the relationship between pNfL (dependent variable), age, NIS and CMTES-R score (independent variables). Statistical analysis was performed using SPSS version 24 (IBM; Armonk, NY) and GraphPad Prism 8.0.1 (GraphPad Inc., La Jolla, CA).

Results

A total of 73 patients with pathogenic *TTR* mutations (V30M n=49, T60A n=10, S77Y n=22) and 16 healthy controls were enrolled in to the study. There was a significant difference in the mean ages (TTR = 56 years, controls = 67 years; $p= 0.04$) of the 2 groups (Table 1). Five patients were taking diflunisal for a median time of 127 days (range 60-2379 days), 1 patient was on tafamidis. 26 patients had disease severity measured (MRC sum score, NIS and/or CMTES-R) within 4 months of blood collection (Table 1).

Plasma NfL concentration was significantly higher in the entire group of patients with ATTRm, and in the subgroup of ATTRm patients with a neuropathy compared to healthy controls ($p< 0.001$; Figure 1). An ANCOVA was run to determine whether pNfL differed between healthy controls and those ATTRm patients with no clinical neurological findings (six participants, NIS=0, MRC=70, CMTES-R=0) after controlling for age. After adjustment for age, there was no statistically significant difference in pNfL, $F(1, 23) = 0.70$, $p= 0.41$, partial $\eta^2 = .03$

Multiple regression analysis to predict pNfL concentration from age, NIS, and CMTES-R found that disease severity measured by NIS was a significant predictor ($\beta = 1.167$, $p = 0.006$), whereas age and CMTES-R were not (age: $\beta = 0.04$, $p = 0.94$; CMTES-R: $\beta = 2.767$, $p = 0.057$). A Spearman's rank-order correlation was run to assess the relationship between NIS and pNfL, and CMTES-R and pNfL. There was a statistically significant, positive correlation between NIS and pNfL ($r_s(30) = 0.65$, $p < 0.0001$, Figure 2a) and CMTES-R and pNfL ($r_s(35) = 0.69$, $p < 0.0001$, Figure 2b)). Correlation analysis between NIS and CMTES-R showed a tight interaction between these 2 scores ($r = 0.89$, $p = <0.0001$, Figure 3).

Discussion

In this study, we have demonstrated that plasma NFL concentration is increased in patients with ATTRm related polyneuropathy and that it correlates with disease severity. This suggests pNFL may be a useful biomarker for monitoring disease progression and treatment efficacy.

The role of pNFL as a diagnostic biomarker in ATTRm related polyneuropathy is uncertain as it is raised in many other acquired, genetic, peripheral and central nervous system disorders, and increases with age⁵. However, the absence of a difference in the mean pNFL concentration between neurologically asymptomatic patients with ATTRm and controls suggests that pNFL may have utility in detecting the onset of neuropathy before clinical signs develop. This is of clinical relevance as several current treatments are only licenced for ATTRm patients with evidence of peripheral neuropathy.

Several invasive and non-invasive biomarkers have been assessed in ATTRm amyloidosis to aid in early diagnosis. Intraepidermal nerve fibre density (IENFD) and in vivo laser scanning confocal microscopy (IVCM) of the cornea sensitively detect small fibre dysfunction in patients with ATTRm and correlate with clinical severity measures^{11,12}. IENFD is a quantitative, albeit invasive marker that would limit its use for repeated measurements and while IVCM is a non-invasive, rapid procedure, depending on the quantification method used, the results can be examiner dependent. High-resolution magnetic resonance neurography (MRN) has been shown to quantitatively identify peripheral nerve impairment in sural nerves of asymptomatic and symptomatic ATTRm patients. This is an encouraging finding but has not been correlated with clinical parameters¹³.

There are a number of limitations to our study. Firstly, the number of control subjects is small, however, the pNFL concentrations in the cohort are in keeping with historical control samples from our group⁶. Secondly, a small proportion of our patients were prescribed

diflusal which may have a disease modifying effect and might artificially reduce pNFL concentration in the ATTRm cohort, however, the statistically significant results of this study are promising as NIS and CMTES-R were only available in a subset of patients, with a smaller subgroup having a neuropathy.

In this study, we have shown that pNFL concentration is sensitive to detecting peripheral axonal damage in ATTRm related polyneuropathy and that it correlates with two commonly used measures of disease severity. Longitudinal studies to assess responsiveness of pNFL to treatments and disease onset are needed before it can be adopted into routine clinical practice.

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Competing interests:

MMR consults/serves on advisory boards for Alnylam Pharmaceuticals, Akcea Therapeutics, and Inflectis. AMR and MK have received support from Alnylam UK Limited to attend scientific meetings and an honorarium for speaking at a sponsored symposium. MMR has or has had consultancies with Alnylam Pharmaceuticals and Ionis Pharmaceuticals and MMR and ML were involved in the NEURO-TTR Trial (ClinicalTrials.gov Identifier: NCT01737398). HZ has served at advisory boards for CogRx, Samumed, Roche Diagnostics, Eli Lilly and Wave, has received travel support from Teva and is a co-founder of Brain Biomarker Solutions in Gothenburg AB, a GU Ventures-based platform company at the University of Gothenburg. JG serves on advisory boards for Alnylam Pharmaceuticals, Akcea Therapeutics, Pfizer Inc, and GlaxoSmithKline. The remaining authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Conception and design: AMR, MMR

Provision of study materials or patients: AMR, MMR, JG, AM, MPL

Sample processing: MF, AH, HZ

Data analysis and interpretation: AMR, MK

Manuscript writing: MK, AMR

Final approval of manuscript: All authors

Characteristics	Controls	All ATTRm	ATTRm without neurology	ATTRm with neurology
No.	16	85	6	20
Age in years (median (range))	67 (29.6- 80.2)	56.7 (21.3- 80.6)	46.0 (33.0- 60.6)	65.8 (30.4- 80.3)
Gender (F/M)	1/14	27/61	3/3	8/12
Plasma NfL (pg.ml) (mean (SD))	15.5 (11.6- 19.3)	58.1 (45.1- 71.0)	2.5 (0-5.2)	68.4 (35.1- 101.8)

Table 1: Descriptive Statistics of Demographic and Clinical Variables in ATTRm Cohort and Controls

Figure 1: Plasma NfL concentration for all patients with ATTRm (TTR), healthy controls, ATTRm patients with neuropathy (Neuropathy) and ATTRm patients without neuropathy (No Neuropathy). * $p < 0.05$. ns = not significant

Figure 2: Plasma NfL concentration plotted against disease severity as measured by the neuropathy impairment score (A), and the Rasch modified CMT examination score (B).

Figure 3: The neuropathy impairment score (NIS) plotted against the Rasch modified Charcot-Marie-Tooth (CMT) examination score shows a significant correlation.

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