

Abstract

MRI plays an important role in the clinical management of pancreatic disorders and interpretation is reliant on qualitative assessment of anatomy. Conventional sequences capturing pancreatic structure can however be adapted to yield quantitative measures which provide more diagnostic information, with a view to increasing diagnostic accuracy, improving patient stratification, providing robust non-invasive outcome measures for therapeutic trials and ultimately personalising patient care.

In this review, we evaluate the use of established techniques such as secretin-enhanced MRCP, diffusion-weighted imaging, T1, T2* and fat fraction mapping, but also more experimental methods such as MR elastography and arterial spin labelling, and their application to the assessment of diffuse pancreatic disease (including chronic, acute and autoimmune pancreatitis/IgG4 disease, metabolic disease and iron deposition disorders) and cystic/solid focal pancreatic masses. Finally, we explore some of the broader challenges to their implementation and future directions in this promising area.

Introduction

Pancreatic MR protocols employed in clinical practice are generally optimised for anatomical imaging¹. High native T1 signal of normal pancreatic parenchyma, high T2 signal of the main pancreatic duct, pancreatic secretions and cystic pancreatic lesions, use of fat suppressed T2 weighted imaging for the assessment of regional inflammation, avid pancreatic parenchymal contrast enhancement (compared to low T1/hypoenhancement of pancreatic adenocarcinoma) and increased conspicuity of many pancreatic lesions on diffusion-weighted imaging (DWI)² have all rendered widespread use of pancreatic MRI in routine clinical pancreatico-biliary practice³.

Many of the signal changes underpinning structural pancreatic MR are amenable to quantification (supplementary material, [table 1](#)). MR sequences designed to quantify these signal changes can complement anatomical imaging, improve diagnostic accuracy and enable the development of new quantitative biomarkers, with the broader objective of better stratifying patients and developing more personalised treatment regimens. In this review we define this potential, appraise current evidence and discuss how methods could be used to expand the utility of MR evaluation of (a) chronic pancreatitis, (b) acute pancreatitis and pancreatic/peripancreatic collections, (c) autoimmune pancreatitis and IgG4 disease, (d) non-alcoholic fatty pancreatic disease and metabolic disease, (e) iron deposition disorders involving the pancreas, and (f) cystic and solid focal pancreatic lesions. As this review is pathology focussed, we have included a brief glossary of MR methods referenced in the text as supplementary material.

DIFFUSE PANCREATIC DISEASE

Chronic pancreatitis

Chronic pancreatitis (CP) is driven by recurrent pancreatic inflammation causing irreversible pancreatic structural damage and progressive endocrine/exocrine dysfunction. Proposed pathophysiological mechanisms include stagnant pancreatic juice forming protein plugs that undergo eventual calcification and cause ductal obstruction, ectasia, chronic inflammation, and periductal fibrosis/scarring. Toxic metabolites and reactive oxygen species are also thought to induce cellular inflammation eventually leading to fibrosis⁴.

MR assessment of CP typically involves assessment of pancreatic morphology, parenchymal T1 signal (as a marker of fibrosis) and where indicated, secretin-enhanced MR cholangio-pancreatography (sMRCP). Unfortunately, qualitative assessment is not particularly sensitive for early disease or assessment of disease severity⁵.

Several quantitative MR studies have developed methods for quantification of post-secretin fluid release^{6,7}, specifically measuring pancreatic flow rate (ml/minute), total excreted volume (mls) or time to peak fluid released (minutes). Quantification has been shown to have good inter-observer agreement and measurements are robust to differing doses of secretin⁸⁻¹⁰. Validation studies have typically been based on phantom studies, but correlations between faecal elastase and pancreatic flow rate ($r=0.79$)¹¹ and total excreted volume at 10 minutes ($r=0.573$, $p<0.001$)¹² have been encouraging. Correlation with invasive endoscopic post-secretin maximal bicarbonate concentration has been less convincing¹³.

In patients with severe CP, pancreatic flow rates (5.3 ± 2.4 ml/min) are lower relative to those with moderate chronic pancreatitis (7.0 ± 3.0 ml/min, $p=0.03$) or normal controls (7.4 ± 2.9 ml/min, $p=0.018$)¹⁴ and time to peak fluid release increases with progressive disease severity (normal controls 5.8 ± 1.7 mins vs mild CP 7.7 ± 2.6 mins vs moderate CP 9.1 ± 3.0 mins vs severe CP 12.3 ± 1.6 mins, $p=0.0001$)¹¹. Following endotherapy (sphincterotomy/stone extraction/stenting) in patients with CP, increases in pancreatic flow output and total excreted volume have been demonstrated¹⁰.

Applications to other conditions associated with pancreatic insufficiency such as cystic fibrosis have found limited value in the use of post-secretin main pancreatic duct diameter as a marker of disease severity¹⁵, but demonstrated qualitatively reduced post-secretin excreted volume

in patients with more advanced disease¹⁶. Quantitative MRCP studies in this cohort have not been reported.

As an alternative, a carefully positioned spatially selective inversion-recovery slab followed by high temporal resolution single-shot MRCP imaging can be used to derive a 'cine' loop of signal refilling the distal pancreatic duct. This has been used to develop a five point grading system that correlates with Cambridge CP scores ($r=-0.698$, $P<0.001$)¹⁷.

The use of secretin MRCP is unfortunately limited by supply shortages and its significant cost¹⁸. While a number of quantitative metrics have been proposed, a consensus around methodology, metrics and cut off levels is lacking, and will need to be defined for multicentre formal evaluation studies and more widespread adoption.

DWI signal reflects water movement within tissue, driven by random (thermal) motion but also water diffusion between tissue compartments across cell membranes (supplementary material, [table 2](#)). Water diffusion between the intracellular, intravascular, interstitial and intraductal compartments present in normal pancreatic tissue are each likely to be influenced by ductal stagnation/obstruction, and fibrosis/scarring occurring in CP.

Reduced and delayed increases in pancreatic apparent diffusion coefficient (ADC) following secretin have been described with more severe CP¹⁹, although findings have not been reproduced²⁰. Baseline ADC values have also been reported to be lower in CP and correlate strongly with endoscopic post-secretin maximal bicarbonate concentration ($r=0.771$, $p<0.0001$)^{20,21}.

Intravoxel incoherent motion (IVIM), an extension of DWI (supplementary material, [table 2](#)) has been applied to CP to show that perfusion fractions tend to be higher in CP, relative to pancreatic adenocarcinoma²³. In a study including 165 patients, patients with moderate and severe pancreatic fibrosis tended to have lower parenchymal pseudodiffusion coefficient (D^*) and perfusion fraction compared to those with mild or no fibrosis²⁴. Interestingly, intra-operative measurements of pancreatic hardness were not correlated with pancreatic ADC values²⁵.

DWI and IVIM are promising approaches, but much like in other areas of the body the lack of consensus on which and how many b-values to use and the approach to modelling ADC/IVIM parameters continue to hamper the development of DWI-derived biomarkers.

Reduction in pancreatic parenchymal T1 signal has been linked with pancreatic fibrosis²⁴ ([figure 1](#)). T1 mapping refers to the formal quantification of T1 signal (supplementary material, [table 3](#)). Increasing T1 with progressive disease has been demonstrated using both multi-flip angle (normal controls 1099 ms vs mild CP 979 ms, $p<0.0001$)²⁶ and modified Look-Locker inversion recovery methods (normal controls 865 ± 220 ms vs mild CP 1075 ± 221 ms vs severe CP 1350 ± 139 ms, $p<0.0001$)²⁷.

<Figure 1>

MR elastography (MRE, supplementary material [table 4](#)) has shown increasing pancreatic parenchymal stiffness with progressive disease, and was superior to T1 mapping in the diagnosis of moderate and severe CP²⁷. Pancreatic perfusion measured using Dynamic Contrast Enhanced (DCE) MRI (reliant on intravenous gadolinium-based contrast agent, supplementary material [table 5](#)) and arterial spin labelling (supplementary material, [table 5](#)) have both been used to measure increases in pancreatic perfusion post-secretin in normal volunteers, albeit both with poor repeatability^{28,29}.

Acute pancreatitis

Although diagnosis is based on clinical parameters, CT in the subacute phase is used to exclude acute complications, provide alternative diagnoses and give an early assessment of classification into 'interstitial' or 'necrotising' morphologies (as per the 2012 Revised Atlanta Classification)³⁰. Latent cross-sectional imaging in the early/late phase (before/after 4 weeks respectively) can subsequently

be used to assess fluid collections, necrosis and formation of pseudocysts/walled-off necrosis. For patients who are able to tolerate MR imaging, better characterisation of fluid/solid/haemorrhagic components of collections, identification of aetiology (such as gallstones) and the potential to derive prognostic information from quantitative imaging sequences justify its use over CT³¹.

Pancreatic parenchymal based phenomena such as acinar cell death, leucocytic infiltration, interstitial fibrin deposition and microvascular thrombosis each have the potential to affect the diffusion of water molecules. DWI has comparable sensitivity to CT for the detection of acute pancreatic inflammation³², with significantly lower parenchymal ADC values and diffusion tensor imaging derived fractional anisotropic measures (supplementary material, [table 2](#)) relative to normal controls^{33,34}, both of which have been correlated with MR severity index scores ($r=-0.63$ and $r=-0.65$, $p<0.01$)³⁴. Histogram analysis of pancreatic parenchymal ADC maps have linked reduced skewness and higher kurtosis (supplementary material, [tables 1 and 2](#)) with increased risk of future complications, although overall sensitivity was low (AUC=0.784)³⁵. Parenchymal T2* measurements can increase in acute pancreatitis (the cause of this is uncertain, but may be secondary to increased perfusion), although these reduce in the presence of superadded haemorrhage³⁶. Finally, DWI can also be useful in the evaluation of pancreatic/peripancreatic collections, as lower ADC values (as a result of cellular debris) can be indicative of superadded infection^{32,37}.

Quantification of T2 signal using T2 mapping (supplementary material, [table 3](#)) in acute pancreatitis has not been reported to date, but typically increased pancreatic/peri-pancreatic T2 signal ([figure 2](#)) has the potential to yield useful quantitative markers.

<Figure 2>

Autoimmune pancreatitis and IgG4 disease

Pathological phenomena in autoimmune pancreatitis (AIP) including acute inflammation, infiltration with IgG4 positive plasma cells, fibrosis and obliterative phlebitis all have the potential to influence quantitative MR parameters³⁸. Published data has focussed on DWI, where pancreatic parenchymal ADC values in AIP have been shown to be lower than in CP^{39,40}, pancreatic adenocarcinoma or normal pancreatic tissue^{41,42}. Pancreatic ADC values have also been significantly correlated with serum IgG4 levels ($r=-0.8$, $p<0.05$)³⁹, shown to be significantly lower in patients with symptoms⁴³ and have been reported to increase following steroid treatment³⁹. Association of low pancreatic parenchymal T1 signal with AIP⁴⁴ ([figure 3](#)) underscores the potential for T1 mapping, which to date has not been reported.

<Figure 3>

Non-Alcoholic Fatty Pancreas Disease and Metabolic Disease

Non-alcoholic fatty pancreas disease (NAFPD) refers to pancreatic steatosis in association with obesity and the metabolic syndrome. Eventual sequelae of intracellular lipid accumulation and proliferation of pancreatic adipocytes include inflammation, pancreatitis, malignancy and the development of type 2 diabetes mellitus⁴⁵. Fatty infiltration and pancreatic atrophy are common morphological findings, but changes in intralobular, interlobular and extralobular fat and their significance are poorly understood⁴⁶.

MR spectroscopy (MRS, supplementary material table 4) is the gold standard approach to fat quantification but is technically challenging in the pancreas because of organ size and respiratory motion artefact. Increased pancreatic fat in obesity, diabetes⁴⁷ and impaired glucose tolerance have been reported but correlation with measures of beta-cell function have been less clear^{48,49}.

Anatomical imaging methods such as the multipoint Dixon sequences have limitations, but have reported increased pancreatic fat in type 2 diabetes mellitus^{50,51} and associations with age⁵², obesity, hypertriglyceridaemia and insulin resistance^{53,54}, with reported reductions in pancreatic fat following low calorie diet regimes in diabetic patients⁵⁵. Proton density fat fraction (PDFF,

supplementary material table 4) mapping methods are currently the most robust method for pancreatic fat quantification but have to date been optimised for the liver and not the pancreas⁵⁶(figure 4). There is conflicting data on the association between pancreatic PDF fat and diabetes, with some reporting pancreatic fat increases^{57,58} or no relationship^{59,60}. Association with insulin resistance has been suggested⁶¹.

MRE has been shown to be repeatable in the pancreas, with increasing stiffness with age⁶². Reductions in pancreatic stiffness have also been reported in obese patients following glycaemic stress⁶³. Post-glycaemic load changes in pancreatic blood-oxygen level dependent (BOLD, supplementary material table 5) MRI based T2* measurements have also been shown in normal volunteers⁶⁴.

<Figure 4 >

Iron deposition disorders involving the Pancreas

Iron deposition in the pancreas remains poorly understood but occurs alongside atrophy and fibrosis and is associated with impaired exocrine/endocrine function. It is thought to account for the high rates of diabetes seen in patients with thalassaemia major⁶⁵.

T2*/R2* mapping (supplementary material, **table 3**) is sensitive to parenchymal iron content, although formal invasive validation studies have not been published. Quantitative studies have focussed on thalassaemia major, but reports of correlations between cardiac, liver and pancreatic T2*/R2* have been conflicting^{66–68}. Correlations between logarithmic pancreatic T2* values and beta-cell function⁶⁹, together with differences in pancreatic T2* between those with normal and abnormal (impaired glucose tolerance/diabetes) glucose metabolism are promising⁷⁰. Pancreatic R2* cut-off values for HFE gene positivity in hereditary haemochromatosis also raise the possibility of applications in other iron deposition disorders⁷¹.

Segmentation, particularly in the presence of fatty infiltration/parenchymal atrophy and errors introduced by multi-echo measurements in the presence of low iron concentrations⁷² pose important challenges.

FOCAL PANCREATIC MASSES

Cystic pancreatic lesions

T2-weighted tissue contrast from fluid content within pancreatic cystic lesions enables differentiation from solid lesions using standard MRI. Improvements in MR and CT image quality, increasing obesity, diabetes and an aging population have resulted in a 19.6% rise in the prevalence of incidental indeterminate non-pseudocyst pancreatic cystic lesions on cross-sectional imaging⁷³. Cystic pathology can range from non-neoplastic cysts to pancreatic ductal adenocarcinomas, therefore there is a genuine need for imaging methods that can distinguish between malignant and benign pathology (figure 5).

The most common potentially malignant cystic pancreatic lesions - intraductal mucinous neoplasms (IPMNs) – can range from slow-growing indolent to aggressive forms, committing patients to extended follow-up regimes with significant resource implications⁷⁴. Several DWI studies have demonstrated that aggressive IPMNs tend to have lower ADC values than more benign lesions, supporting the use of DWI as a predictor malignant risk^{75–77}. Malignant IPMNs have also been associated with lower IVIM D_{slow} and higher D_{fast} values relative to more benign IPMNs⁷⁸.

Current prognostication guidelines are based entirely on anatomical features – distinction between mucinous content and serous content (and therefore differentiation between mucinous cystic neoplasms and benign serous cystadenomas) would significantly aid diagnosis and prognostication.

Simple cysts tend to have higher ADC values due to T2 shine through⁷⁹, while the presence of mucin tends to lower ADC values⁸⁰. Small scale studies have also demonstrated that ADC values of abscess tend to be lower than neoplastic cysts⁸¹.

Finally, genetic characterisation of excised pancreatic cystic lesions have suggested hyperpolarised ^{13}C -pyruvate MRS maybe useful in the distinction between malignant and benign cysts⁸².

<Figure 5 >

Solid pancreatic lesions

Unlike their cystic counterparts, solid pancreatic lesions are more commonly malignant (i.e. pancreatic ductal adenocarcinoma or neuroendocrine tumours, NETs; [figure 6](#)). Important benign differentials include solid inflammatory masses such as focal/mass-forming pancreatitis, which can be difficult to exclude without extended follow-up, often at the expense of a worsened prognosis if malignancy is eventually diagnosed. Improved diagnostic sensitivity, better characterisation, early exclusion of benign differentials and prognostic/therapeutic stratification are therefore important areas where quantitative MRI has the potential to improve patient outcomes.

Limited differences between lesion and pancreatic parenchymal T1/T2 signal intensity, gland atrophy and fatty infiltration can make the detection of small solid pancreatic lesions difficult. B-value optimised DWI can be used to increase lesion conspicuity⁸³ and sensitivity for small NETs⁸⁴.

Pancreatic malignancy is associated with reduced ADC^{85–88} and reduced fractional anisotropy⁸⁵ relative to adjacent normal pancreatic tissue (supplementary material, [table 2](#)). Studies using DWI to distinguish inflammatory and malignant solid pancreatic lesions, have been less conclusive with some demonstrating lower ADC in malignancy^{89–93} and others showing no difference^{79,94–96}. IVIM studies (supplementary material, [table 2](#)) have shown lower perfusion fractions and D_{fast} in adenocarcinoma⁹⁷, and that differences in D_{slow} could be used to separate pancreatic ductal adenocarcinoma and NETs⁷⁸.

Evaluation of NETs has been more promising with tumour differentiation linked to ADC values, although correlations with mitotic counts and Ki-67 have been poor⁹⁸. Whole lesion segmentation and measures of ADC entropy and kurtosis (supplementary material, [table 1](#)) are associated with more advanced NETs⁹⁹. ADC values of normal pancreatic tissue upstream to infiltrated tissue are also an independent prognostic factor in overall patient survival¹⁰⁰.

Quantitative DCE MRI (supplementary material, [table 5](#)), using Tofts pharmacokinetic modelling to T1-corrected enhancement data has demonstrated lower K_{trans} and higher extracellular volume in pancreatic ductal adenocarcinoma compared with normal pancreas^{101,102}, with enhancement patterns differing between inflammatory and malignant masses⁹³. Combining DCE and ADC data, sensitivity and specificity levels of 97% and 94% have been reported⁹³, but lesion DCE parameter variability is high and repeatability is poor¹⁰³.

Finally, a small study has demonstrated altered pancreatic adenocarcinoma T2* relative to normal pancreas, potentially explained by altered oxygenation (and vascularity) within the lesion¹⁰³.

<Figure 6 >

Challenges and future directions

Adoption of quantitative pancreatic MR methods within the wider clinical community requires validation against robust gold-standards, which are currently lacking. Percutaneous biopsy, with the attendant risks of haemorrhage and visceral damage is seldom justified, and histological characterisation of diffuse pancreatic disease or solid pancreatic masses is only likely if endoscopic ultrasound (EUS) guided or intraoperative aspiration/biopsy/excision takes place¹⁰⁴. Assessment of secretory function requires endoscopy and the method is prone to inaccuracies, particularly in early disease¹⁰⁵. Markers such as faecal elastase can be useful for validation but represent a competing biomarker – justifying the use of MRI given cost and patient convenience in this setting can be difficult.

Many of the quantitative MRI methods that have been applied in the pancreas have been developed and optimised for other organs (e.g. fat quantification using PDFF, T2*/R2* mapping for iron quantification). This is challenging because signal changes in the pancreas though measurable are small⁴⁶. Dedicated studies optimising methods such as DWI may have utility, but a lack of consensus in standardising acquisition parameters (e.g. number of b-values and modelling methods for ADC/IVIM¹⁰⁶) remain a major barrier to deriving diagnostic quantitative cut-off values that can be applied across different sites and scanners¹⁰⁷. The lack of standardised protocols is a common barrier for methods such as T1 mapping (where different methods can yield small but significant differences in quantification) and DCE MRI. Finally, quantitative MR sequences must be time-efficient to enable them to be included within normal anatomical imaging protocols.

Delivery of quantitative imaging to the end-user radiologist is also a challenge. Automated generation of parametric maps that are amenable to simple region-of-interest analysis on standard PACS systems is essential to the acceptance of quantitative techniques within the wider radiological community. Creative methods of displaying quantitative data, such as overlay of colour-rendered DWI on anatomical images have the potential to improve diagnostic utility⁸⁴. Improvements in image registration methods can address motion and misregistration artefacts particularly where co-registered acquisitions are needed¹⁰⁸. Additionally, extraction of quantitative data relies on image segmentation which remains cumbersome and controversial. For example, in the presence of diffuse disease, the pancreas becomes atrophic and irregular – little is known about the differing pathological implications of intralobular, interlobular and extralobular portions of the pancreas⁴⁶. The application of new machine-learning based segmentation methods promises to help address some of these challenges¹⁰⁹.

This review has also highlighted methods which have theoretical potential but as yet have not been investigated – the role of T1 mapping in autoimmune pancreatitis or T2 mapping in acute pancreatitis are ripe for well-designed imaging-outcome based studies.

Finally, the advent of new quantitative MRI methods in the pancreas has delivered a number of measures for which the pathological implications are either undefined or better defined in other organs. In combination with validation studies, development of hypotheses for mechanistic studies will help define a clear role for pancreatic quantitative MR and ultimately deliver robust non-invasive biomarkers that can be used as primary outcome measures in much-needed therapeutic trials.

References

1. Sandrasegaran K, Lin C, Akisik FM, Tann M. State-of-the-art pancreatic MRI. *AJR Am J Roentgenol.* 2010;195(1):42-53. doi:10.2214/AJR.10.442110.2214/ajr.195.3_supplement.0s42
2. Park HJ, Jang KM, Song KD, et al. Value of unenhanced MRI with diffusion-weighted imaging for detection of primary small (≤ 20 mm) solid pancreatic tumours and prediction of pancreatic ductal adenocarcinoma. *Clin Radiol.* 2017;72(12):1076-1084. doi:10.1016/J.CRAD.2017.07.009
3. Roth CG, Marzio DH-D, Guglielmo FF. Contributions of Magnetic Resonance Imaging to Gastroenterological Practice: MRIs for GIs. *Dig Dis Sci.* 2018;63(5):1102-1122. doi:10.1007/s10620-018-4991-x
4. Steer ML, Waxman I, Freedman S. Chronic Pancreatitis. *N Engl J Med.* 1995;332(22):1482-1490. doi:10.1056/NEJM199506013322206
5. Anaizi A, Hart PA, Conwell DL. Diagnosing Chronic Pancreatitis. *Dig Dis Sci.* 2017;62(7):1713-1720. doi:10.1007/s10620-017-4493-2
6. Heverhagen JT, Müller D, Battmann A, et al. MR Hydrometry to Assess Exocrine Function of the Pancreas: Initial Results of Noninvasive Quantification of Secretion. *Radiology.* 2001;218(1):61-67. doi:10.1148/radiology.218.1.r01ja2061
7. Punwani S, Gillams AR, Lees WR. Non-invasive quantification of pancreatic exocrine function using secretin-stimulated MRCP. *Eur Radiol.* 2003;13:273-276. doi:10.1007/s00330-002-1605-

8. Trout AT, Wallihan DB, Serai S, Abu-El-Haija M. Secretin-Enhanced Magnetic Resonance Cholangiopancreatography for Assessing Pancreatic Secretory Function in Children. *J Pediatr*. 2017;188:186-191. doi:10.1016/j.jpeds.2017.06.031
9. Bali MA, Sontou R, Arvanitakis M, Metens T, Devière J, Matos C. Evaluation of the stimulating effect of a low dose of secretin compared to the standard dose on the exocrine pancreas with MRCP: Preliminary results in normal subjects (MRCP quantification of secretin stimulation). *Abdom Imaging*. 2007;32(6):743-748. doi:10.1007/s00261-006-9164-2
10. Bali MA, Sztantics A, Metens T, et al. Quantification of pancreatic exocrine function with secretin-enhanced magnetic resonance cholangiopancreatography: normal values and short-term effects of pancreatic duct drainage procedures in chronic pancreatitis. Initial results. *Eur Radiol*. 2005;15(10):2110-2121. doi:10.1007/s00330-005-2819-5
11. Bian Y, Wang L, Chen C, et al. Quantification of pancreatic exocrine function of chronic pancreatitis with secretin-enhanced MRCP. *World J Gastroenterol*. 2013;19(41):7177-7182. doi:10.3748/wjg.v19.i41.7177
12. Manfredi R, Perandini S, Mantovani W, Frulloni L, Faccioli N, Pozzi Mucelli R. Quantitative MRCP assessment of pancreatic exocrine reserve and its correlation with faecal elastase-1 in patients with chronic pancreatitis. *Radiol Med*. 2012;117(2):282-292. doi:10.1007/s11547-011-0774-6
13. Balci NC, Smith A, Momtahn AJ, et al. MRI and S-MRCP findings in patients with suspected chronic pancreatitis: Correlation with endoscopic pancreatic function testing (ePFT). *J Magn Reson Imaging*. 2010;31(3):601-606. doi:10.1002/jmri.22085
14. Gillams AR, Lees WR. Quantitative secretin MRCP (MRCPQ): results in 215 patients with known or suspected pancreatic pathology. *Eur Radiol*. 2007;17(11):2984-2990. doi:10.1007/s00330-007-0708-9
15. Jonczyk-Potoczna K, Nowak JK, Madry E, et al. Secretin-enhanced Magnetic Resonance Cholangio-pancreatography in Pancreatic Insufficient and Pancreatic Sufficient Cystic Fibrosis Patients. *J Gastrointest Liver Dis*. 2016;25(1):57-62. <http://www.ncbi.nlm.nih.gov/pubmed/27014754>. Accessed June 21, 2018.
16. Engjom T, Erchinger F, Tjora E, Lærum BN, Georg D, Gilja OH. Diagnostic accuracy of secretin-stimulated ultrasonography of the pancreas assessing exocrine pancreatic failure in cystic fibrosis and chronic pancreatitis. *Scand J Gastroenterol*. 2015;50(5):601-610. doi:10.3109/00365521.2015.1004363
17. Yasokawa K, Ito K, Kanki A, et al. Evaluation of pancreatic exocrine insufficiency by cine-dynamic MRCP using spatially selective inversion-recovery (IR) pulse: Correlation with severity of chronic pancreatitis based on morphological changes of pancreatic duct. *Magn Reson Imaging*. 2018;48:70-73. doi:10.1016/j.mri.2017.12.007
18. Chamokova B, Bastati N, Poetter-Lang S, et al. The clinical value of secretin-enhanced MRCP in the functional and morphological assessment of pancreatic diseases. *Br J Radiol*. 2018;91(1084):20170677. doi:10.1259/bjr.20170677
19. Erturk SM, Ichikawa T, Motosugi U, Sou H, Araki T. Diffusion-Weighted MR Imaging in the Evaluation of Pancreatic Exocrine Function Before and After Secretin Stimulation. *Am J Gastroenterol*. 2006;101(1):133-136. doi:10.1111/j.1572-0241.2006.00406.x
20. Akisik MF, Aisen AM, Sandrasegaran K, et al. Assessment of Chronic Pancreatitis: Utility of Diffusion-weighted MR Imaging with Secretin Enhancement. *Radiology*. 2009;250(1):103-109. doi:10.1148/radiol.2493080160
21. Balci NC, Momtahn AJ, Akduman EI, Alkaade S, Bilgin M, Burton FR. Diffusion-weighted MRI of the Pancreas. Correlation with Secretin Endoscopic Pancreatic Function Test (ePFT). *Acad Radiol*. 2008;15(10):1264-1268. doi:10.1016/j.acra.2008.05.002
22. Madzak A, Olesen SS, Haldorsen IS, Drewes AM, Frøkjær JB. Secretin-stimulated MRI characterization of pancreatic morphology and function in patients with chronic pancreatitis.

- Pancreatology*. 2017;17(2):228-236. doi:10.1016/j.pan.2017.01.009
23. Klau M, Lemke A, Grünberg K, et al. Intravoxel Incoherent Motion MRI for the Differentiation Between Mass Forming Chronic Pancreatitis and Pancreatic Carcinoma. *Invest Radiol*. 2011;46(1):57-63. doi:10.1097/RLI.0b013e3181fb3bf2
 24. Yoon JH, Lee JM, Lee KB, et al. Pancreatic Steatosis and Fibrosis: Quantitative Assessment with Preoperative Multiparametric MR Imaging. *Radiology*. 2016;279(1):140-150. doi:10.1148/radiol.2015142254
 25. Hong TH, Choi J-I, Park MY, et al. Pancreatic hardness: Correlation of surgeon's palpation, durometer measurement and preoperative magnetic resonance imaging features. *World J Gastroenterol*. 2017;23(11):2044-2051. doi:10.3748/wjg.v23.i11.2044
 26. Tirkes T, Lin C, Fogel EL, Sherman SS, Wang Q, Sandrasegaran K. T1 mapping for diagnosis of mild chronic pancreatitis. *J Magn Reson Imaging*. 2017;45(4):1171-1176. doi:10.1002/jmri.25428
 27. Wang M, Gao F, Wang X, et al. Magnetic resonance elastography and T₁ mapping for early diagnosis and classification of chronic pancreatitis. *J Magn Reson Imaging*. March 2018. doi:10.1002/jmri.26008
 28. Bali MA, Devie J. Pancreatic Perfusion : Noninvasive Quantitative Assessment with Dynamic Contrast-enhanced MR Imaging Healthy Volunteers — Initial Results 1 Purpose : Methods : Results : Conclusion : 2008;247(1):115-121.
 29. Schawkat K, Ith M, Christe A, et al. Dynamic non-invasive ASL perfusion imaging of a normal pancreas with secretin augmented MR imaging. *Eur Radiol*. 2018;28(6):2389-2396. doi:10.1007/s00330-017-5227-8
 30. Bollen TL. Acute pancreatitis: international classification and nomenclature. *Clin Radiol*. 2016;71(2):121-133. doi:10.1016/J.CRAD.2015.09.013
 31. McPherson SJ, O'Reilly DA, Sinclair MT, Smith N. The use of imaging in acute pancreatitis in United Kingdom hospitals: findings from a national quality of care study. *Br J Radiol*. 2017;90(1080):20170224. doi:10.1259/bjr.20170224
 32. Islim F, Salik AE, Bayramoglu S, Guven K, Alis H, Turhan AN. Non-invasive detection of infection in acute pancreatic and acute necrotic collections with diffusion-weighted magnetic resonance imaging: preliminary findings. *Abdom Imaging*. 2014;39(3):472-481. doi:10.1007/s00261-014-0076-2
 33. Thomas S, Kayhan A, Lakadamyali H, Oto A. Diffusion MRI of acute pancreatitis and comparison with normal individuals using ADC values. *Emerg Radiol*. 2012;19(1):5-9. doi:10.1007/s10140-011-0983-2
 34. Li X, Zhuang L, Zhang X, et al. Preliminary Study of MR Diffusion Tensor Imaging of Pancreas for the Diagnosis of Acute Pancreatitis. *PLoS One*. 2016;11(9):e0160115. doi:10.1371/journal.pone.0160115
 35. Iranmahboob AK, Kierans AS, Huang C, Ream JM, Rosenkrantz AB. Preliminary investigation of whole-pancreas 3D histogram ADC metrics for predicting progression of acute pancreatitis. *Clin Imaging*. 2017;42:172-177. doi:10.1016/J.CLINIMAG.2016.12.007
 36. Tang MY, Chen TW, Huang XH, et al. Acute pancreatitis with gradient echo T2*-weighted magnetic resonance imaging. *Quant Imaging Med Surg*. 2016;6(2):157-167. doi:10.21037/qims.2016.04.03
 37. Borens B, Arvanitakis M, Absil J, et al. Added value of diffusion-weighted magnetic resonance imaging for the detection of pancreatic fluid collection infection. *Eur Radiol*. 2017;27(3):1064-1073. doi:10.1007/s00330-016-4462-8
 38. Chari ST, Kloepfel G, Zhang L, et al. Histopathologic and Clinical Subtypes of Autoimmune Pancreatitis. *Pancreas*. 2010;39(5):549-554. doi:10.1097/MPA.0b013e3181e4d9e5
 39. Taniguchi T, Kobayashi H, Nishikawa K, et al. Diffusion-weighted magnetic resonance imaging in autoimmune pancreatitis. *Jpn J Radiol*. 2009;27(3):138-142. doi:10.1007/s11604-008-0311-2

40. Muhi A, Ichikawa T, Motosugi U, et al. Mass-forming autoimmune pancreatitis and pancreatic carcinoma: Differential diagnosis on the basis of computed tomography and magnetic resonance cholangiopancreatography, and diffusion-weighted imaging findings. *J Magn Reson Imaging*. 2012;35(4):827-836. doi:10.1002/jmri.22881
41. Kamisawa T, Takuma K, Anjiki H, et al. Differentiation of Autoimmune Pancreatitis From Pancreatic Cancer by Diffusion-Weighted MRI. *Am J Gastroenterol*. 2010;105(8):1870-1875. doi:10.1038/ajg.2010.87
42. Hur BY, Lee JM, Lee JE, et al. Magnetic resonance imaging findings of the mass-forming type of autoimmune pancreatitis: Comparison with pancreatic adenocarcinoma. *J Magn Reson Imaging*. 2012;36(1):188-197. doi:10.1002/jmri.23609
43. Oki H, Hayashida Y, Oki H, et al. DWI findings of autoimmune pancreatitis: Comparison between symptomatic and asymptomatic patients. *J Magn Reson Imaging*. 2015;41(1):125-131. doi:10.1002/jmri.24508
44. Hansen TM, Nilsson M, Gram M, Frokjaer JB. Morphological and functional evaluation of chronic pancreatitis with magnetic resonance imaging. *World J Gastroenterol*. 2013;19(42):7241-7246. doi:10.3748/wjg.v19.i42.7241
45. Tariq H, Nayudu S, Akella S, Glandt M, Chilimuri S. Non-Alcoholic Fatty Pancreatic Disease: A Review of Literature. *Gastroenterol Res*. 2016;9(6):87-91. doi:10.14740/gr731w
46. Sakai NS, Taylor SA, Chouhan MD. Obesity, metabolic disease and the pancreas—Quantitative imaging of pancreatic fat. *Br J Radiol*. June 2018:20180267. doi:10.1259/bjr.20180267
47. Tushuizen ME, Bunck MC, Pouwels PJ, et al. Pancreatic Fat Content and beta-Cell Function in Men With and Without Type 2 Diabetes. *Diabet Care*. 2007;30:2916-2921.
48. van der Zijl NJ, Goossens GH, Moors CCM, et al. Ectopic Fat Storage in the Pancreas, Liver, and Abdominal Fat Depots: Impact on β -Cell Function in Individuals with Impaired Glucose Metabolism. *J Clin Endocrinol Metab*. 2011;96(2):459-467. doi:10.1210/jc.2010-1722
49. Komada H, Sakaguchi K, Hirota Y, et al. Pancreatic fat content assessed by ¹H magnetic resonance spectroscopy is correlated with insulin resistance, but not with insulin secretion, in Japanese individuals with normal glucose tolerance. *J Diabetes Investig*. 2017;9(3):505. doi:10.1111/jdi.12720
50. Begovatz P, Koliaki C, Weber K, et al. Pancreatic adipose tissue infiltration, parenchymal steatosis and beta cell function in humans. *Diabetologia*. 2015;58(7):1646-1655. doi:10.1007/s00125-015-3544-5
51. Raeder H, Haldorsen IS, Ersland L, et al. Pancreatic lipomatosis is a structural marker in nondiabetic children with mutations in carboxyl-ester lipase. *Diabetes*. 2007;56(2):444-449. doi:10.2337/db06-0859
52. Le K-M, Ventura E, Fisher J, et al. Ethnic Differences in Pancreatic Fat Accumulation and Its Relationship With Other Fat Depots and Inflammatory Markers. *Diabetes Care*. 2011;34(1):485-490. doi:10.2337/dc10-0760.
53. Staaf J, Labmayr V, Paulmichl K, et al. Pancreatic Fat Is Associated With Metabolic Syndrome and Visceral Fat but Not Beta-Cell Function or Body Mass Index in Pediatric Obesity. *Pancreas*. 2017;46(3):358-365. doi:10.1097/MPA.0000000000000771
54. Wong VW, Wong GL, Yeung DK, et al. Fatty pancreas, insulin resistance, and beta-cell function: a population study using fat-water magnetic resonance imaging. *Am J Gastroenterol*. 2014;109(4):589-597. doi:10.1038/ajg.2014.1
55. Lim EL, Hollingsworth KG, Aribisala BS, Chen MJ, Mathers JC, Taylor R. Reversal of type 2 diabetes: Normalisation of beta cell function in association with decreased pancreas and liver triacylglycerol. *Diabetologia*. 2011;54(10):2506-2514. doi:10.1007/s00125-011-2204-7
56. Bray TJ, Chouhan MD, Punwani S, Bainbridge A, Hall-Craggs MA. Fat fraction mapping using magnetic resonance imaging: insight into pathophysiology. *Br J Radiol*. 2017:20170344. doi:10.1259/bjr.20170344
57. Heber SD, Hetterich H, Lorbeer R, et al. Pancreatic fat content by magnetic resonance

- imaging in subjects with prediabetes, diabetes, and controls from a general population without cardiovascular disease. *PLoS One*. 2017;12(5):e0177154. doi:10.1371/journal.pone.0177154
58. Idilman IS, Tuzun A, Savas B, et al. Quantification of liver, pancreas, kidney, and vertebral body MRI-PDFF in non-alcoholic fatty liver disease. *Abdom Imaging*. 2015;40(6):1512-1519. doi:10.1007/s00261-015-0385-0
 59. Kuhn JP, Berthold F, Mayerle J, et al. Pancreatic Steatosis Demonstrated at MR Imaging in the General Population: Clinical Relevance. *Radiology*. 2015;276(1):129-136.
 60. Patel NS, Peterson MR, Brenner DA, Heba E, Sirlin C, Loomba R. Association between novel MRI-estimated pancreatic fat and liver histology-determined steatosis and fibrosis in non-alcoholic fatty liver disease. *Aliment Pharmacol Ther*. 2013;37(6):630-639. doi:10.1111/apt.12237
 61. Patel NS, Peterson MR, Lin GY, et al. Insulin Resistance Increases MRI-Estimated Pancreatic Fat in Nonalcoholic Fatty Liver Disease and Normal Controls. *Gastroenterol Res Pr*. 2013;2013:498296. doi:10.1155/2013/498296
 62. Kolipaka A, Schroeder S, Mo X, Shah Z, Hart PA, Conwell DL. Magnetic resonance elastography of the pancreas: Measurement reproducibility and relationship with age. *Magn Reson Imaging*. 2017;42:1-7. doi:10.1016/J.MRI.2017.04.015
 63. Ji R, Li J, Yin Z, et al. Pancreatic stiffness response to an oral glucose load in obese adults measured by magnetic resonance elastography. *Magn Reson Imaging*. 2018;51:113-119. doi:10.1016/j.mri.2018.04.019
 64. Chen B, Chen W, Chan Q, Zhou N, He J, Zhou Z. Functional MRI of human pancreas using BOLD contrast: Responses following glucose ingestion. *J Magn Reson Imaging*. 2017;46(3):831-836. doi:10.1002/jmri.25640
 65. Chatterjee R, Bajoria R. New Concept in Natural History and Management of Diabetes Mellitus in Thalassemia Major. *Hemoglobin*. 2009;33(sup1):S127-S130. doi:10.3109/09553000903347880
 66. de Assis RA, Ribeiro AAF, Kay FU, et al. Pancreatic iron stores assessed by magnetic resonance imaging (MRI) in beta thalassemic patients. *Eur J Radiol*. 2012;81(7):1465-1470. doi:10.1016/J.EJRAD.2011.03.077
 67. Pfeifer CD, Schoennagel BP, Grosse R, et al. Pancreatic iron and fat assessment by MRI-R2* in patients with iron overload diseases. *J Magn Reson Imaging*. 2015;42(1):196-203. doi:10.1002/jmri.24752
 68. Pinto VM, Bacigalupo L, Gianesin B, et al. Lack of correlation between heart, liver and pancreas MRI-R2*: Results from long-term follow-up in a cohort of adult β -thalassemia major patients. *Am J Hematol*. 2018;93(3):E79-E82. doi:10.1002/ajh.25009
 69. Au W-Y, Lam WW-M, Chu W, et al. A T2* magnetic resonance imaging study of pancreatic iron overload in thalassemia major. *Haematologica*. 2008;93(1):116-119. doi:10.3324/haematol.11768
 70. Kosaryan M, Rahimi M, Darvishi-Khezri H, Gholizadeh N, Akbarzadeh R, Aliasgharian A. Correlation of Pancreatic Iron Overload Measured by T2*-Weighted Magnetic Resonance Imaging in Diabetic Patients with β -Thalassemia Major. *Hemoglobin*. 2017;41(3):151-156. doi:10.1080/03630269.2017.1340306
 71. Henninger B, Rauch S, Zoller H, Plaikner M, Jaschke W, Kremser C. R2*-relaxometry of the pancreas in patients with human hemochromatosis protein associated hereditary hemochromatosis. *Eur J Radiol*. 2017;89:149-155. doi:10.1016/j.ejrad.2017.02.006
 72. Meloni A, De Marchi D, Positano V, et al. Accurate estimate of pancreatic T2* values: how to deal with fat infiltration. *Abdom Imaging*. 2015;40(8):3129-3136. doi:10.1007/s00261-015-0522-9
 73. Stark A, Donahue TR, Reber HA, Hines OJ. Pancreatic Cyst Disease. *JAMA*. 2016;315(17):1882. doi:10.1001/jama.2016.4690

74. Megibow AJ, Baker ME, Morgan DE, et al. Management of Incidental Pancreatic Cysts: A White Paper of the ACR Incidental Findings Committee. *J Am Coll Radiol*. 2017;14(7):911-923. doi:10.1016/J.JACR.2017.03.010
75. Kang KM, Lee JM, Shin C Il, et al. Added value of diffusion-weighted imaging to MR cholangiopancreatography with unenhanced mr imaging for predicting malignancy or invasiveness of intraductal papillary mucinous neoplasm of the pancreas. *J Magn Reson Imaging*. 2013;38(3):555-563. doi:10.1002/jmri.24022
76. Zhang L, Rao SX, Xu XF, Wang DS, Jin DY, Zeng MS. Value of apparent diffusion coefficient for predicting malignancy of intraductal papillary mucinous neoplasms of the pancreas. *Diagn Interv Radiol*. 2016;22(4):308-313. doi:10.5152/dir.2016.15354
77. Kim M, Mi Jang K, Kim SH, et al. Diagnostic accuracy of diffusion restriction in intraductal papillary mucinous neoplasm of the pancreas in comparison with "high-risk stigmata" of the 2012 international consensus guidelines for prediction of the malignancy and invasiveness. *Acta radiol*. 2017;58(10):1157-1166. doi:10.1177/0284185116685921
78. Kang KM, Lee JM, Yoon JH, Kiefer B, Han JK, Choi BI. Intravoxel Incoherent Motion Diffusion-weighted MR Imaging for Characterization of Focal Pancreatic Lesions. *Radiology*. 2014;270(2):444-453. doi:10.1148/radiol.13122712
79. Wang Y, Miller FH, Chen ZE, et al. Diffusion-weighted MR Imaging of Solid and Cystic Lesions of the Pancreas. *RadioGraphics*. 2011;31(3):E47-E64. doi:10.1148/rg.313105174
80. Yamashita Y, Namimoto T, Mitsuzaki K, et al. Mucin-producing tumor of the pancreas: diagnostic value of diffusion-weighted echo-planar MR imaging. *Radiology*. 1998;208(3):605-609. doi:10.1148/radiology.208.3.9722835
81. Inan N, Arslan A, Akansel G, Anik Y, Demirci A. Diffusion-weighted imaging in the differential diagnosis of cystic lesions of the pancreas. *Am J Roentgenol*. 2008;191(4):1115-1121. doi:10.2214/AJR.07.3754
82. Penheiter AR, Deelchand DK, Kittelson E, et al. Identification of a pyruvate-to-lactate signature in pancreatic intraductal papillary mucinous neoplasms. *Pancreatology*. 2018;18(1):46-53. doi:10.1016/j.pan.2017.11.006
83. Fukukura Y, Shindo T, Hakamada H, et al. Diffusion-weighted MR imaging of the pancreas: optimizing b-value for visualization of pancreatic adenocarcinoma. *Eur Radiol*. 2016;26(10):3419-3427. doi:10.1007/s00330-015-4174-5
84. Brenner R, Metens T, Bali M, Demetter P, Matos C. Pancreatic neuroendocrine tumor: Added value of fusion of T2-weighted imaging and high b-value diffusion-weighted imaging for tumor detection. *Eur J Radiol*. 2012;81(5):e746-e749. doi:10.1016/j.ejrad.2012.01.032
85. Nissan N, Golan T, Furman-Haran E, et al. Diffusion tensor magnetic resonance imaging of the pancreas. *PLoS One*. 2014;9(12):e115783. doi:10.1371/journal.pone.0115783
86. Muraoka N, Uematsu H, Kimura H, et al. Apparent diffusion coefficient in pancreatic cancer: Characterization and histopathological correlations. *J Magn Reson Imaging*. 2008;27(6):1302-1308. doi:10.1002/jmri.21340
87. Rosenkrantz AB, Matza BW, Sabach A, Hajdu CH, Hindman N. Pancreatic cancer: Lack of association between apparent diffusion coefficient values and adverse pathological features. *Clin Radiol*. 2013;68(4):e191-e197. doi:10.1016/J.CRAD.2012.11.006
88. Kartalis N, Manikis GC, Loizou L, et al. Diffusion-weighted MR imaging of pancreatic cancer: A comparison of mono-exponential, bi-exponential and non-Gaussian kurtosis models. *Eur J Radiol open*. 2016;3:79-85. doi:10.1016/j.ejro.2016.04.002
89. Seung SL, Jae HB, Beom JP, et al. Quantitative analysis of diffusion-weighted magnetic resonance imaging of the pancreas: Usefulness in characterizing solid pancreatic masses. *J Magn Reson Imaging*. 2008;28(4):928-936. doi:10.1002/jmri.21508
90. Fattahi R, Balci NC, Perman WH, et al. Pancreatic diffusion-weighted imaging (DWI): Comparison between mass-forming focal pancreatitis (FP), pancreatic cancer (PC), and normal pancreas. *J Magn Reson Imaging*. 2009;29(2):350-356. doi:10.1002/jmri.21651

91. Balci NC, Perman WH, Saglam S, Akisik F, Fattahi R, Bilgin M. Diffusion-weighted magnetic resonance imaging of the pancreas. *Top Magn Reson Imaging*. 2009;20(1):43-47. doi:10.1097/RMR.0b013e3181b48667
92. Yao X-Z, Yun H, Zeng M-S, et al. Evaluation of ADC measurements among solid pancreatic masses by respiratory-triggered diffusion-weighted MR imaging with inversion-recovery fat-suppression technique at 3.0 T. *Magn Reson Imaging*. 2013;31(4):524-528. doi:10.1016/J.MRI.2012.09.006
93. Zhang T-T, Wang L, Liu H-H, et al. Differentiation of pancreatic carcinoma and mass-forming focal pancreatitis: qualitative and quantitative assessment by dynamic contrast-enhanced MRI combined with diffusion-weighted imaging. *Oncotarget*. 2017;8(1):1744-1759. doi:10.18632/oncotarget.12120
94. Barral M, Taouli B, Guiu B, et al. Diffusion-weighted MR Imaging of the Pancreas: Current Status and Recommendations. *Radiology*. 2015;274(1):45-63. doi:10.1148/radiol.14130778
95. Momtahn AJ, Balci NC, Alkaade S, Akduman EI, Burton FR. Focal pancreatitis mimicking pancreatic mass: Magnetic resonance imaging (MRI)/magnetic resonance cholangiopancreatography (MRCP) findings including diffusion-weighted MRI. *Acta radiol*. 2008;49(5):490-497. doi:10.1080/02841850802014602
96. Wiggermann P, Grützmänn R, Weissenböck A, Kamusella P, Dittert DD, Stroszczynski C. Apparent diffusion coefficient measurements of the pancreas, pancreas carcinoma, and mass-forming focal pancreatitis. *Acta radiol*. 2012;53(2):135-139. doi:10.1258/ar.2011.100252
97. Concia M, Sprinkart AM, Penner A-H, et al. Diffusion-Weighted Magnetic Resonance Imaging of the Pancreas. *Invest Radiol*. 2014;49(2):93-100. doi:10.1097/RLI.0b013e3182a71cc3
98. Guo C, Zhuge X, Chen X, Wang Z, Xiao W, Wang Q. Value of diffusion-weighted magnetic resonance imaging in predicting World Health Organization grade in G1/G2 pancreatic neuroendocrine tumors. *Oncol Lett*. 2017;13(6):4141-4146. doi:10.3892/ol.2017.6029
99. De Robertis R, Maris B, Cardobi N, et al. Can histogram analysis of MR images predict aggressiveness in pancreatic neuroendocrine tumors? *Eur Radiol*. 2018;28(6):2582-2591. doi:10.1007/s00330-017-5236-7
100. Hayano K, Miura F, Wada K, et al. Diffusion-weighted MR imaging of pancreatic cancer and inflammation: Prognostic significance of pancreatic inflammation in pancreatic cancer patients. *Pancreatology*. 2016;16(1):121-126. doi:10.1016/J.PAN.2015.10.004
101. Maria A, Bali M, Thierry Metens P, Vincent Denolin P, et al. Tumoral and Nontumoral Pancreas : Correlation between Quantitative Dynamic Contrast- enhanced MR Imaging and Purpose : Methods : Results : *Radiology*. 2011;261(2):456-466. doi:10.1148/radiol.11103515/-/DC1
102. Ma W, Li N, Zhao W, et al. Apparent Diffusion Coefficient and Dynamic Contrast-Enhanced Magnetic Resonance Imaging in Pancreatic Cancer. *J Comput Assist Tomogr*. 2016;40(5):709-716. doi:10.1097/RCT.0000000000000434
103. Klaassen R, Gurney-Champion OJ, Wilmink JW, et al. Repeatability and correlations of dynamic contrast enhanced and T2* MRI in patients with advanced pancreatic ductal adenocarcinoma. *Magn Reson Imaging*. 2018;50:1-9. doi:10.1016/J.MRI.2018.02.005
104. Levy MJ. Know when to biopsy 'em, know when to walk away. *Gastrointest Endosc*. 2006;63(4):630-634. doi:10.1016/j.gie.2005.06.021
105. Schibli S, Corey M, Gaskin KJ, Ellis L, Durie PR. Towards the ideal quantitative pancreatic function test: analysis of test variables that influence validity. *Clin Gastroenterol Hepatol*. 2006;4(1):90-97. <http://www.ncbi.nlm.nih.gov/pubmed/16431310>. Accessed July 9, 2018.
106. Park HJ, Sung YS, Lee SS, et al. Intravoxel incoherent motion diffusion-weighted MRI of the abdomen: The effect of fitting algorithms on the accuracy and reliability of the parameters. *J Magn Reson Imaging*. 2017;45(6):1637-1647. doi:10.1002/jmri.25535
107. Nissan N. Modifications of pancreatic diffusion MRI by tissue characteristics: what are we

- weighting for? *NMR Biomed.* 2017;30(8):e3728. doi:10.1002/nbm.3728
108. Jin J, McKenzie E, Fan Z, et al. Nonlocal Means Denoising of Self-Gated and k-Space Sorted 4-Dimensional Magnetic Resonance Imaging Using Block-Matching and 3-Dimensional Filtering: Implications for Pancreatic Tumor Registration and Segmentation. *Int J Radiat Oncol Biol Phys.* 2016;95(3):1058-1066. doi:10.1016/j.ijrobp.2016.02.006
109. Cai J, Lu L, Zhang Z, Xing F, Yang L, Yin Q. Pancreas Segmentation in MRI using Graph-Based Decision Fusion on Convolutional Neural Networks. *Med Image Comput Comput Assist Interv.* 2016;9901:442-450. doi:10.1007/978-3-319-46723-8_51

Figure legends:

Figure 1:

Axial T1 weighted images of the pancreas (white dashed line) in a normal volunteer (a) and patient with chronic pancreatitis (b). Chronic inflammation and fibrosis are thought to account for low parenchymal T1 and reduced volume in (b). Also note the presence of high T1 signal dependent pigment gallstones (solid white arrow) in (b).

Figure 2:

Axial T2 fat saturated image through the pancreas in a patient with acute pancreatitis. Note the presence of high peripancreatic T2 signal (solid white arrows) indicative of peripancreatic free fluid and the presence of patchy areas of increased pancreatic parenchymal T2 signal (clear white arrow) indicative of acute interstitial oedema. Perihepatic and perisplenic high T2 signal free fluid is also noted.

Figure 3:

Axial T1 weighted image through the pancreas of a patient with a flare of autoimmune pancreatitis. The pancreas is bulky, with a rind of low T1 signal (solid white arrows) and there is diffusely reduced pancreatic parenchymal signal with patchy areas of further reductions in T1 signal (clear white arrow) likely secondary to acute inflammation.

Figure 4:

Axial proton-density fat fraction image of the pancreas (black dashed line). Preserved pancreatic parenchyma is low signal but the gland is irregular and there is interlobular fatty infiltration suggestive of pancreatic steatosis.

Figure 5:

Axial T2 weighted image through the pancreas. The lobulated cystic lesion in the pancreatic tail (solid white arrow), demonstrated some concerning features (e.g. 2.7 cm size, thickened internal septations) but also some reassuring features (such as no associated main pancreatic ductal dilatation or enhancing components). Endoscopic US guided fine needle aspiration confirmed an intraductal papillary mucinous neoplasm.

Figure 6:

Coronal (a) and axial (b) T1 weighted fat saturated post-contrast images of the pancreas in two different patients. The hypervascular lesion in the superior head of pancreas (a), is associated with two large ill-defined right-sided hepatic metastases (solid white arrows) and was later confirmed as a neuroendocrine tumour. The irregular hypovascular mass in the head of the pancreas (b), was later confirmed to be a pancreatic ductal adenocarcinoma.