

Contents lists available at ScienceDirect

EClinicalMedicine

journal homepage: https://www.journals.elsevier.com/ eclinicalmedicine



Research Paper

Trends in Pretreatment HIV-1 Drug Resistance in Antiretroviral Therapy-naive Adults in South Africa, 2000–2016: A Pooled Sequence Analysis

Benjamin Chimukangara ^{a,b,c,*}, Richard J. Lessells ^{a,b}, Soo-Yon Rhee ^d, Jennifer Giandhari ^a, Ayesha B.M. Kharsany ^b, Kogieleum Naidoo ^{b,e}, Lara Lewis ^b, Cherie Cawood ^f, David Khanyile ^f, Kassahun A. Ayalew ^g, Karidia Diallo ^g, Reshmi Samuel ^c, Gillian Hunt ^{h,i}, Alain Vandormael ^{a,j}, Babill Stray-Pedersen ^k, Michelle Gordon ^a, Tariro Makadzange ^l, Photini Kiepiela ^m, Gita Ramjee ^m, Johanna Ledwaba ^h, Monalisa Kalimashe ^h, Lynn Morris ^{b,h,i}, Urvi M. Parikh ⁿ, John W. Mellors ⁿ, Robert W. Shafer ^d, David Katzenstein ^d, Pravi Moodley ^c, Ravindra K. Gupta ^{o,p}, Deenan Pillay ^{o,p}, Salim S. Abdool Karim ^b, Tulio de Oliveira ^{a,b,*}

- a KwaZulu-Natal Research Innovation and Sequencing Platform (KRISP), Department of Laboratory Medicine & Medical Sciences, University of KwaZulu-Natal, Durban, South Africa
- b Centre for the AIDS Programme of Research in South Africa (CAPRISA), Doris Duke Medical Research Institute, Nelson R Mandela School of Medicine, University of KwaZulu-Natal, Durban, South Africa
- ^c Department of Virology, National Health Laboratory Service, University of KwaZulu-Natal, Durban, South Africa
- ^d Department of Medicine, Stanford University, Stanford, CA, United States of America
- e South African Medical Research Council (SAMRC)-CAPRISA HIV-TB Pathogenesis and Treatment Research Unit, Durban, South Africa
- ^f Epicentre AIDS Risk Management (Pty) Limited, PO Box 3484, Paarl, Cape Town, South Africa
- ^g Centers for Disease Control and Prevention, Pretoria, South Africa
- ^h Centre for HIV and STIs, National Institute for Communicable Diseases (NICD), Johannesburg, South Africa
- ⁱ Faculty of Health Sciences, University of the Witwatersrand, Johannesburg, South Africa
- ^j School of Nursing and Public Health, University of KwaZulu-Natal, Durban, South Africa
- k Institute of Clinical Medicine, University of Oslo, Oslo University Hospital, Oslo, Norway
- 1 Ragon Institute of Massachusetts General Hospital, Massachusetts Institute of Technology and Harvard, Cambridge, MA, United States of America
- m HIV Prevention Research Unit, Medical Research Council, Durban, South Africa
- ⁿ Department of Infectious Diseases, University of Pittsburgh School of Medicine, Pittsburgh, PA, United States of America
- ° Department of Infection, University College London, United Kingdom of Great Britain and Northern Ireland
- ^p Africa Health Research Institute, University of KwaZulu-Natal, Durban, South Africa

ARTICLE INFO

Article history: Received 10 September 2018 Received in revised form 1 March 2019 Accepted 5 March 2019 Available online 18 March 2019

Keywords: HIV Pre-treatment drug resistance Antiretroviral therapy Surveillance Molecular epidemiology Pooled sequence analysis South Africa

ABSTRACT

Background: South Africa has the largest public antiretroviral therapy (ART) programme in the world. We assessed temporal trends in pretreatment HIV-1 drug resistance (PDR) in ART-naïve adults from South Africa. *Methods:* We included datasets from studies conducted between 2000 and 2016, with HIV-1 *pol* sequences from more than ten ART-naïve adults. We analysed sequences for the presence of 101 drug resistance mutations. We pooled sequences by sampling year and performed a sequence-level analysis using a generalized linear mixed model, including the dataset as a random effect.

Findings: We identified 38 datasets, and retrieved 6880 HIV-1 pol sequences for analysis. The pooled annual prevalence of PDR remained below 5% until 2009, then increased to a peak of $11 \cdot 9\%$ (95% confidence interval (CI) $9 \cdot 2 - 15 \cdot 0$) in 2015. The pooled annual prevalence of non-nucleoside reverse-transcriptase inhibitor (NNRTI) PDR remained below 5% until 2011, then increased to 10.0% (95% CI 8.4 - 11.8) by 2014. Between 2000 and 2016, there was a 1.18-fold (95% CI 1.13 - 1.23) annual increase in NNRTI PDR (p < 0.001), and a 1.10-fold (95% CI 1.05 - 1.16) annual increase in nucleoside reverse-transcriptase inhibitor PDR (p = 0.001).

Interpretation: Increasing PDR in South Africa presents a threat to the efforts to end the HIV/AIDS epidemic. These findings support the recent decision to modify the standard first-line ART regimen, but also highlights the need for broader public health action to prevent the further emergence and transmission of drug-resistant HIV.

Source of Funding: This research project was funded by the South African Medical Research Council (MRC) with funds from National Treasury under its Economic Competitiveness and Support Package.

Disclaimer: The contents of this publication are solely the responsibility of the authors and do not necessarily represent the official views of CDC.

© 2019 Published by Elsevier Ltd. This is an open access article under the CC BY-NC-ND license (http://creativecommons.org/licenses/by-nc-nd/4.0/).

E-mail addresses: benjiechim@yahoo.com (B. Chimukangara), deoliveira@ukzn.ac.za (T. de Oliveira).

^{*} Corresponding authors at: KwaZulu-Natal Research Innovation and Sequencing Platform (KRISP), Department of Laboratory Medicine & Medical Science, University of KwaZulu-Natal, 719 Umbilo Road, Durban 4001, South Africa.

Research in context

Evidence before this study

We searched PubMed for systematic reviews and meta-analyses of pretreatment or transmitted HIV drug resistance in South Africa. We used the search terms "HIV" AND "South Africa" AND "drug resistance" AND "(systematic review OR meta-analysis)". We found two meta-analyses exploring regional prevalence of pretreatment or transmitted HIV drug resistance, where data from South Africa were combined with data from other countries in a regional analysis (southern Africa or sub-Saharan Africa). We found a meta-analysis of pretreatment HIV drug resistance in children younger than 12 years, which included data from South Africa. We also had a systematic review from our own group which analysed transmitted drug resistance in South Africa up to 2010. We did not identify any studies that focused on South Africa and incorporated sequences collected since 2010, when scale-up of antiretroviral therapy accelerated.

Added value of this study

In this pooled analysis of 6880 HIV-1 sequences from 38 datasets, we provide up-to-date estimates of the prevalence of pretreatment HIV drug resistance (PDR) in South Africa. We present evidence of increasing PDR, particularly since the acceleration of ART scale-up in 2010. We demonstrate that the increase is largely driven by non-nucleoside reverse-transcriptase inhibitor (NNRTI) PDR, but that levels of nucleoside reverse-transcriptase inhibitor (NRTI) PDR are also rising. In particular, we note a concerning increase in the prevalence of tenofovir resistance-associated mutations (TRAMs), which could have important implications for current treatment and prevention strategies.

Implications of all the available evidence

Our findings provide clear evidence that PDR in South Africa has reached the threshold at which the World Health Organization recommends urgent public health action (NNRTI PDR > 10%). Whilst our data provide support for the decision to move to a new dolutegravir-based first-line regimen, they also highlight the broader need to improve quality of HIV treatment and prevention if South Africa is to achieve the Joint United Nations Programme on HIV/AIDS goal of ending AIDS by 2030.

1. Introduction

The roll-out of antiretroviral therapy (ART) has been a major breakthrough in the global response to HIV, helping to reduce HIV-related deaths by 48% between 2005 and 2016, and new HIV infections by 11% between 2010 and 2016 [1]. Despite these impressive public health gains, substantial expansion of access to ART will be required to achieve the target of ending the HIV epidemic by 2030 [1]. The emergence and transmission of HIV drug resistance (HIVDR) pose a threat to the successful treatment and prevention of HIV, and there is now strong evidence that levels of HIVDR are increasing substantially in southern Africa [2], the region that faces the greatest challenges to ending the HIV epidemic.

Pretreatment HIV drug resistance (PDR) is drug resistance in a person initiating or re-initiating ART (i.e. with or without prior ART exposure) [3,4]. PDR can arise in one of three ways: transmission of drug-resistant HIV from a person with acquired drug resistance (ADR); transmission of primary drug-resistant HIV from another ART-naïve

person; or ADR resulting from prior exposure to antiretroviral drugs for treatment or prevention. The presence of PDR is associated with poorer virological outcomes on first-line ART [5.6].

South Africa, with over seven million people living with HIV (PLHIV) in 2016, accounts for almost one in five PLHIV globally [1]. The country has the largest public ART programme in the world, with more than four million people on ART by early 2018 [7]. In the first few years of ART rollout, the levels of PDR were low (<5%) [8]. More recent studies, conducted since the accelerated expansion of ART coverage in 2010, have suggested higher levels of PDR [9,10].

Given this evidence of rising levels of PDR in the country and the wider region, and the continued expansion of ART for treatment and prevention, we performed a pooled analysis of HIV sequence data from South Africa, firstly to determine the annual trends in PDR and secondly to explore in detail the patterns of observed drug resistance mutations (DRMs).

2. Methods

2.1. Search Strategy and Selection Criteria

This study was a systematic review and pooled analysis aimed at determining trends in PDR amongst ART-naïve adults in South Africa. We conducted and reported this in line with the Preferred Reporting Items for Systematic Reviews and Meta-Analyses statement (checklist included in Appendix, p 1) [11]. To identify relevant studies we first searched for published articles in MEDLINE using the OvidSP interface on 12 September 2017 (Appendix, p 3). We then scanned the reference lists of all articles selected for inclusion and conducted forward citation searches using Google Scholar. Finally, we searched South African HIV-1 sequence datasets not linked to a published article, using the PopSet database on the National Center for Biotechnology Information website [12].

We included studies involving adults (defined for the purpose of this analysis as 15 years or older) in South Africa with recent or chronic HIV infection and no documented prior ART exposure. We obtained information about prior ART exposure from either the article or the sequence annotation in GenBank. We excluded studies that enrolled women with documented exposure to antiretrovirals for prevention of mother-tochild transmission (pMTCT). We excluded studies with fewer than ten HIV-1 pol sequences; and studies where the sequences were generated from samples collected prior to 2000. Where articles reported on multiple separate cross-sectional studies (for example a series of annual antenatal surveys), we separated the sequences into individual datasets according to the sampling year. If results from the same study were presented in more than one publication, we pooled the sequences into a single dataset. We included sequences from one multi-national study [13], as South African sequences could be identified through the sequence annotation in GenBank.

From the articles, we retrieved a core set of information, including the year(s) of sample collection, province, study type, study population, proportion of participants that were female, and method for determining prior ART exposure.

2.2. Sequence Analysis

We downloaded publicly available sequences for the included studies from GenBank [12]. Where sequences were not publicly accessible, we contacted the study authors to request the sequences. We aligned and visually inspected the sequences in AliView v1.18 (http://ormbunkar.se/aliview/) [14]. We manually edited the sequences until perfect codon-based alignments were produced. We assessed sequences for their completeness and quality using the Calibrated Population Resistance (CPR) tool (http://cpr.stanford.edu/cpr.cgi) [15]. Stop codons, frameshift mutations, APOBEC3G/F hyper-mutations, highly unusual mutations and highly ambiguous nucleotides (B, D, H, V and N), were all used as indicators of poor sequence quality. We excluded from the analysis any

sequence that did not meet the sequence inclusion criteria of the CPR tool [14]. We included all sequences that had complete reverse transcriptase gene (RT) sequences (codons 40 to 240), with or without complete protease (PR) sequences. Where multiple sequences were identified from the same study participant (for example in cohort studies), we only included the sequence from the earliest time point. Most sequences were not annotated with information about participant sex or age, so we did not include this information in the datasets.

We defined PDR as the presence of any of 101 DRMs. The mutation list included the 93 mutations from the World Health Organization (WHO) 2009 list of surveillance drug-resistance mutations (SDRMs) [16]; and eight additional tenofovir (TDF) resistance-associated mutations (TRAMs) characterised in a recent international collaborative analysis (A62V, K65N, S68GDN, K70QT, and V75L) [17], (Appendix, p 4). Overall, the mutation list encompassed 42 nucleoside reverse-transcriptase inhibitor (NRTI)-resistance mutations at 17 RT positions, 19 non-nucleoside reverse-transcriptase inhibitor (NNRTI)-resistance mutations at ten RT positions, and 40 protease inhibitor (Pl)-resistance mutations at 18 PR positions. We used the CPR tool to calculate the proportion of sequences with overall and drug class-specific PDR [15].

2.3. Trends in Pretreatment Drug Resistance

To assess the annual increase in overall and drug class-specific PDR, we pooled sequences from different studies by year of sample collection and performed a generalized linear mixed regression model using the

R package (v3.3.1) lme4. We used the presence or absence of PDR (or drug class-specific PDR) as the binary outcome variable and the sampling year as the explanatory variable. Where samples from the same study had been collected over more than one year and where the sequence annotation did not include year of sample collection, we allocated the sequences to the median sampling year. To account for heterogeneity between studies, we included the dataset as a random effect in the model. Given the relatively small number of sequences with specific mutations, we also pooled the sequences into three periods (2000–2008, 2009–2012, and 2013–2016) and checked for any trend in prevalence of specific NRTI- and NNRTI-resistance mutations using the chi-squared test for trend.

3. Results

We initially identified 856 articles through our database search and nine articles through other sources. After removing duplicate publications, we screened 790 abstracts and assessed 46 full-text articles for eligibility. We excluded 14 articles on the basis of our eligibility criteria: eight contained fewer than 10 HIV-1 *pol* sequences; two had only PR sequences with no RT sequences; one reported on a duplicate sequence dataset; one contained only sequences generated from samples collected prior to 2000; one was based on targeted sequencing for a single mutation (K65R); and sequences were unavailable for one study (Appendix, p 5). From the 32 articles, we identified 38 datasets with at least ten HIV-1 *pol* sequences from ART-naïve adults (Fig. 1, Table 1, Appendix,

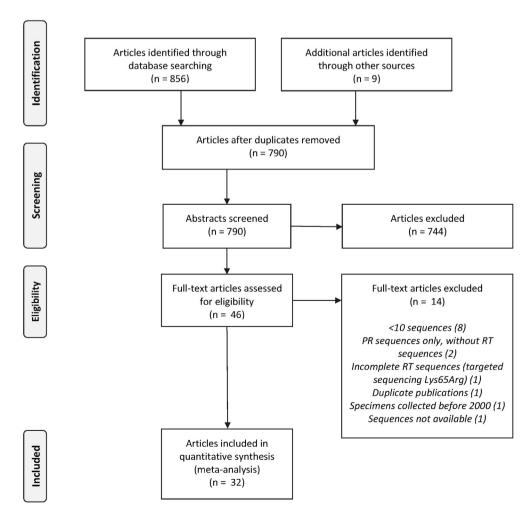


Fig. 1. Flow diagram of articles and datasets identified and selected for the pooled sequence analysis of PDR in antiretroviral therapy-naive adults in South Africa.

Table 1Characteristics of included datasets with ten or more RT sequences from ART-naïve adults.

Dataset ID	Source	Sampling years	Province(s)	Study type	Study population	Proportion females	Method for determining prior ART use	Met criteria for WHO TDR/PDR survey
1	Bessong	2001	LP	Genetic diversity	ART-naïve adults	79%	NS	No
2	Bessong	2001-2004	GT, LP	TDR	ART-naïve adults	68%	NS	Yes
3	Chimukangara	2013	KZN	Population HIV surveillance	HIV-positive adults > 15 years	73%	Linkage to public sector records	
4	Chimukangara	2014	KZN	Population HIV surveillance	HIV-positive adults > 15 years	75%	Linkage to public sector records	No
5	Chimukangara	2014–2015	KZN	Population HIV surveillance	HIV-positive adults 15–49 years	66%	Self-report	No
6	Gordon	2001-2002	KZN	Genetic diversity	ART-naïve adults	66%	NS	No
7	Hamers	2007-2008	GT, MP	PDR	Adults eligible for ART	62%	Self-report	Yes
3	Huang	2006	FS	TDR	ART-naïve adults	NS	Self-report	Yes
9	Hunt	2005	GT, KZN	ANC survey	Primigravid female <25 years	100%	NS	Yes
10	Hunt	2006	GT, KZN	ANC survey	Primigravid female <25 years	100%	NS	Yes
11	Hunt	2007	GT, KZN	ANC survey	Primigravid female <25 years	100%	NS	Yes
12	Hunt	2008	GT, KZN	ANC survey	Primigravid female <25 years	100%	NS	Yes
13	Hunt	2009	GT, KZN	ANC survey	Primigravid female	100%	NS	Yes
14	Hunt	2010	GT, KZN	ANC survey	<25 years Primigravid females	100%	NS	Yes
15	Hunt	2011	EC, FS, GT, KZN,	ANC survey	≤21 years Primigravid females	100%	NS	Yes
16	Hunt	2012	WC EC, FS, GT, KZN, LP,	ANC survey	≤25 years Primigravid females	100%	NS	Yes
17	Ianiahan	2007 2000	MP, NC, NW, WC	Comptin dissensites	≤21 years	00%	Calf mamont	Me
17	Iweriebor	2007–2008	LP	Genetic diversity	ART-naïve adults	90%	Self-report	No
18	Jacobs	2002–2004	WC	Genetic diversity	ART-naïve adults	66%	Self-report	No
19	Jacobs	2008-2010	WC	Neurocognitive study	ART-naïve females	100%	NS	No
20	Manasa	2010	KZN	Population HIV surveillance	HIV-positive adults >15 years	85%	NS	No
21	Manasa	2011	KZN	Population HIV surveillance	HIV-positive adults >15 years	76%	Linkage to public sector records	
22	Manasa	2012	KZN	Population HIV surveillance	HIV-positive adults >15 years	71%	Linkage to public sector records	No
23	Matthews	2000–2004	KZN	Chronic infection cohort	ART-naïve adults	92%	Self-report	No
24	Msimanga	2009	MP	Genetic diversity	ART-naïve adults	95%	Self-report	No
25	Musyoki	2007	GT	Genetic diversity	Adults initiating ART	NS	Self-report	No
26	Nwobegahay	2008	LP	TDR	ART-naïve adults	70-73%	Self-report	Yes
27 28	Papathanasopoulos Parboosing	2006–2007 2009	GT KZN	Genetic diversity TDR	ART-naïve adults Primigravid female	74% 100%	Self-report NS	No Yes
29	Parikh	2010-2011	KZN	Trial screening	<22 years Females 18–40 years	100%	NS	No
30	Pillay	2000	GT	(HIV prevention) Trial screening	first positive test ART-naïve pregnant	100%	NS	No
31	Pillay	2002	GT	(pMTCT) ANC survey	females Primigravid females	100%	NS	Yes
32	Pillay	2004	GT	ANC survey	<22 years Primigravid females	100%	NS	Yes
33	Seoighe	2003–2005	GT, KZN	Trial baseline (pMTCT)	<22 years Pregnant females	100%	NS	No
34	Steegen	2013–2014	EC, FS, GT, KZN, LP, MP, NC, NW, WC	PDR	Adults initiating ART or in pre-ART care	59%	Self-report	Yes
35	Treurnicht	2004–2005	KZN	Acute infection study	Females with documented acute infection	100%	NS	No
36	van Zyl	2016–2017	WC	PDR	ART-naïve adults initiating ART	52%	Self-report	Yes
37	Wilkinson	2000	WC	Phylogenetic study	ART-naïve patients	NS	NS	No
38	Wilkinson	2004	WC	Phylogenetic study	ART-naïve patients	NS	NS	No

ANC, antenatal care; ART, antiretroviral therapy; EC, Eastern Cape; FS, Free State; GT, Gauteng; KZN, KwaZulu-Natal; LP, Limpopo; MP, Mpumalanga; NC, Northern Cape; NS, not stated; NW, North West; PDR, pretreatment drug resistance; pMTCT, prevention of mother-to-child transmission; TDR, transmitted drug resistance; WC, Western Cape.

pp. 6–9) [8–10,13,18–44]. Seventeen datasets were from formal surveys of PDR or transmitted drug resistance.

We retrieved 7025 *RT* sequences and 6501 *PR* sequences. We excluded 145 RT sequences and 207 PR sequences that did not meet sequence quality criteria. Therefore, we included 6880 *RT* sequences and 6294 *PR* sequences in the analysis (i.e. 6294 sequences with combined

PR and *RT* and 586 with *RT* only) (Appendix, pp. 10, 11). The majority of sequences were subtype C (99.2%). Overall, 478 of 6880 sequences (6.9%) had at least one DRM. The majority of these sequences had only NNRTI-resistance mutations (289/478, 60.5%); dual class NRTI and NNRTI PDR were present in 79/478 (16.5%) (Appendix, p 12). The prevalence of overall and drug class-specific PDR in each dataset is displayed

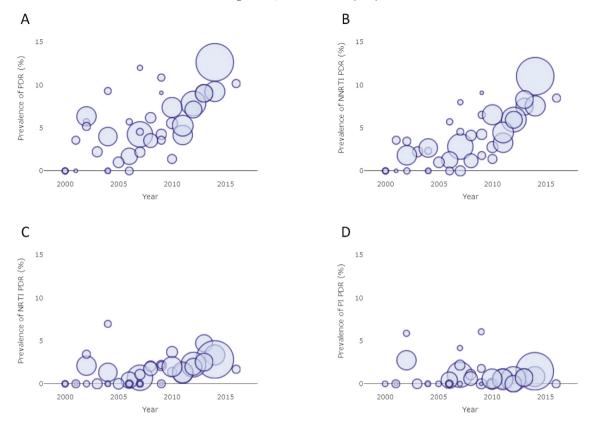


Fig. 2. Prevalence of pretreatment HIV drug resistance by year of sampling. A) Overall, B) non-nucleoside reverse-transcriptase inhibitor, C) nucleoside reverse-transcriptase inhibitor, D) protease inhibitor. Each bubble represents a dataset and the size of the bubble is proportional to the number of sequences in the dataset. The sampling year is shown on the horizontal axis and the percentage PDR on the vertical axis. PDR, pretreatment HIV drug resistance.

in Fig. 2, and the crude pooled prevalence of overall and drug class-specific PDR by year is shown in Table 2. The prevalence of NNRTI PDR remained below 5% until 2011 and then increased rapidly to above 10% by 2014. The pooled prevalence of NRTI PDR and PI PDR remained below 5% across all years. Over the entire study period (2000–2016), there was a 1.10-fold yearly increase in the odds of PDR (95% confidence interval (CI) 1.06–1.15), which was driven by increasing NNRTI PDR (odds ratio (OR) 1.18, 95% CI 1.13–1.23) and NRTI PDR (OR 1.10, 95% CI 1.05–1.16) (Table 3).

Overall, 374 sequences (5.4%) had at least one NNRTI DRM (Appendix, p 13). The most prevalent mutation was K103NS, occurring in 278

sequences (58.2% of sequences with any DRM; 4.0% of all sequences) (Fig. 3). In the majority of these sequences (218/278), K103NS was the only DRM. Other common NNRTI-resistance mutations included V106AM (n = 47), Y181C (n = 34), K101EP (n = 29) and G190ASE (n = 27). Overall, 77/374 (20.6%) had more than one NNRTI DRM, most commonly K103N + P225H (n = 16) and K103N + V106M (n = 12). The prevalence of some specific NNRTI-resistance mutations increased over time. This trend was most marked for the K103NS and V106AM mutations, and less so for the K101EP mutations. There was no evidence of changing prevalence of Y181C or G190ASE (Appendix, p 14).

Table 2Pooled prevalence of pretreatment HIV drug resistance (PDR), NNRTI PDR, and NRTI PDR, by year.

Year	Number of RT sequences	Any DRM	Any PDR (95% CI)	NNRTI DRM	NNRTI PDR (95% CI)	NRTI DRM	NRTI PDR (95% CI)
2000	66	0	=	0	-	0	-
2001	69	2	2.9 (0.4-10.1)	2	2.9 (0.4-10.1)	0	_
2002	424	26	6.1 (4.0-8.9)	8	1.9 (0.8-3.7)	9	2.1 (1.0-4.0)
2003	90	2	2.2 (0.3-7.8)	2	2.2 (0.3-7.8)	0	_
2004	377	16	4.2 (2.4-6.8)	9	2.4 (1.1-4.5)	7	1.9 (0.7-3.8)
2005	113	1	0.9 (0-4.8)	1	0.9 (0-4.8)	0	_
2006	303	5	1.7 (0.5-3.8)	4	1.3 (0.4-3.3)	1	0.3 (0-1.8)
2007	748	32	4.3 (2.9-6.0)	21	2.8 (1.7-4.3)	5	0.7 (0.2-1.6)
2008	290	13	4.5 (2.4-7.5)	7	2.4 (1.0-4.9)	5	1.7 (0.6-4.0)
2009	172	7	4.1 (1.7-8.2)	6	3.5 (1.3-7.4)	2	1.2 (0.1-4.1)
2010	306	17	5.6 (3.3-8.7)	12	3.9 (2.0-6.7)	6	2.0 (0.7-4.2)
2011	953	54	5.7 (4.3-7.3)	45	4.7 (3.5-6.3)	16	1.7 (1.0-2.7)
2012	788	60	7.6 (5.9-9.7)	47	6.0 (4.4-7.9)	17	2.2 (1.3-3.4)
2013	370	36	9.7 (6.9-13.2)	31	8.4 (5.8-11.7)	16	4.3 (2.5-6.9)
2014	1255	142	11.3 (9.6-13.2)	126	10.0 (8.4-11.8)	38	3.0 (2.2-4.1)
2015	497	59	11.9 (9.2–15.0)	48	9.7 (7.2–12.6)	12	2.4 (1.3-4.2)
2016	59	6	10.2 (3.8-20.8)	5	8.5 (2.8-18.7)	1	1.7 (0-9.1)

CI, confidence interval; DRM, drug resistance mutation; NRTI, nucleoside reverse-transcriptase inhibitor; NNRTI, non-nucleoside reverse-transcriptase inhibitor; PDR, pretreatment drug resistance; RT, reverse transcriptase.

Table 3Annual change in odds of pretreatment HIV drug resistance, 2000–2016.

Drug class	Odds ratio (95% CI)	p value
NRTI	1.10 (1.05–1.16)	0.0001
NNRTI	1.18 (1.13–1.23)	< 0.0001
PI	0.96 (0.89-1.04)	0.3650
Overall	1.10 (1.06-1.15)	< 0.0001

CI, confidence interval; NRTI, nucleoside reverse-transcriptase inhibitor; NNRTI, non-nucleoside reverse-transcriptase inhibitor; PI, protease inhibitor.

M184VI was the most common NRTI-resistance mutation, present in 71 sequences (14.9% of sequences with any DRM; 1.0% of all sequences) (Appendix, p 15). Most of the sequences with M184VI had at least one NNRTI DRM (66/71) and just under half had additional NRTI DRMs (31/71). The other NRTI DRMs accompanying M184VI included thymidine analogue mutations (TAMs, n = 11), TRAMs (n = 11), L74VI and/or Y115F (n = 7), and other multi-NRTI mutations (n = 2). Classical TAMs (M41L, D67N, K70R, L210W, T215FY, and K219EQ) were detected in 36 sequences (7.5% of sequences with any DRM; 0.5% of all sequences). The majority of these (30/36) had a single TAM; and eleven sequences had the M41L mutation alone without other DRMs. Overall, TRAMs were detected in 37 sequences (7.7% of sequences with any DRM; 0.5% of all sequences). The TRAM most frequently detected was K65R (n = 21). Twelve sequences had a TRAM not on the WHO SDRM list (A62V, n = 10; K70T, n = 2), although in four of these sequences the mutation was present with the K65R mutation. The prevalence of TRAMs increased in later time periods: 0.1% (3/2480) in 2000–2008, 0.5% (11/2219) in 2009–2012, and 1.1% (23/2181) in 2013–2016, and for the M184VI mutation: 0.2% (4/2480) in 2000-2008, 0.9% (20/2219) in 2009-2012, and 2.2% (47/2181) in 2013–2016 (p < 0.001, χ^2 test for trend) (Appendix, p 14).

Fifty-six sequences (0.9%) had at least one PI DRM. The most frequently observed mutation was the relatively non-polymorphic M46IL mutation, which was detected in 35 sequences (0.6%) (Appendix, p 16).

4. Discussion

In this pooled analysis with more than 6000 HIV-1 sequences from ART-naïve adults in South Africa, we observed a sustained increase in pretreatment HIV drug resistance between 2000 and 2016, driven primarily by NNRTI-resistance. The increase in PDR seems to have accelerated since 2010, which coincides with the rapid expansion of ART coverage in the country from just 20% in 2010 to 56% in 2016 [45]. By 2014, the pooled

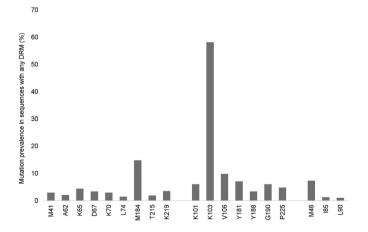


Fig. 3. Prevalence of specific mutations in HIV-1 sequences with any drug resistance mutation. Mutations shown on the horizontal axis include all mutations observed in >1% of the sequences with any drug resistance mutation. DRM, drug resistance mutation.

prevalence of NNRTI PDR had reached 10%, the threshold at which the WHO now recommends urgent public health action [46]. There was also some evidence of increasing NRTI PDR, particularly TDF-associated resistance and the M184VI mutation associated with lamivudine (3TC) and emtricitabine (FTC) resistance. However, the pooled prevalence of NRTI-resistance remained low (<5%) in each sampling year.

These findings are consistent with those from recent meta-analyses exploring drug resistance across Africa, which showed levels of resistance rising to moderate levels about ten years into the scale-up of ART in the region [2,47]. The overall 11% annual increase in odds of PDR between 2000 and 2016 in South Africa is comparable to the 12% increase in odds of transmitted drug resistance across sub-Saharan Africa between 2000 and 2013 [47]. The 18% annual increase in odds of NNRTI PDR is somewhat lower than the 24% reported for the southern Africa region in a more recent meta-analysis [2]. That could be explained by the fact that we only included ART-naïve adults, whereas the regional meta-analysis included a small number of sequences from people with prior ART exposure. Alternatively, it could be that the higher rate of increase in PDR in the regional meta-analysis was reflective of higher levels of PDR in other southern African countries.

Our analysis was restricted to ART-naïve individuals and our assumption is therefore that transmitted drug resistance is the primary driver of the increasing PDR prevalence. There are limitations to this assumption, best illustrated by the most prevalent DRM, the K103NS mutation. This mutation, selected by efavirenz (EFV) and nevirapine (NVP), is the most common acquired NNRTI DRM in people with virological failure on standard first-line ART regimens in South Africa [48]. Viruses with the K103NS mutation have transmission fitness similar to wild-type virus [49,50], and can persist for years in the infected host [51]. It's therefore entirely plausible that the high prevalence of this mutation is a consequence of frequent transmission. However, K103NS is also the most common mutation to emerge in women who receive single-dose NVP for the prevention of mother-to-child transmission and, in this context too, the mutation can persist for years in the absence of antiretroviral therapy [52,53]. Although we restricted the analysis to ART-naïve individuals, we could not be certain that participants in the individual studies were truly ART naïve. Most studies relied on selfreport of antiretroviral use, which can be unreliable [54–59]. Given that the majority of sequences were from women, it is possible that some of the NNRTI-resistance arose from prior exposure to NVP for pMTCT rather than from transmitted drug resistance.

We also revealed evidence of increasing NRTI-resistance, at a rate similar to that observed in the larger regional meta-analyses [2,47]. We specifically demonstrated increasing prevalence of TRAMs and the M184VI mutation, which is of some concern as TDF and FTC/3TC remain the NRTI backbone of choice for first-line ART regimens. In the latter years (2013–2016), the pooled prevalence of the M184VI mutation was approximately 2% and the prevalence of TRAMs was 1%. TDF and FTC/3TC have been part of the standard first-line ART regimen in South Africa since 2010. The national drug resistance survey in 2013-14 showed that most people with virological failure on first-line NNRTI-based ART harboured the M184VI mutation and about half had TRAMs [48]. Whilst our findings could be a signal of increasing transmission of NRTI-resistant virus, we urge some caution in interpretation. Viruses with the M184VI and K65R mutations are thought to be infrequently transmitted due to low transmission fitness [49,50]. If they are transmitted, the mutations revert rapidly in the absence of drug pressure [51,60]. It is possible that some of the sequences with NRTI resistance were obtained from people who reported themselves to be ART naïve but who had previously been exposed to NRTIs. This is certainly plausible as there is an increasing frequency of cyclical engagement in care as ART programmes have matured [61]. Somewhat against that was the observation that the prevalence of TAMs did not change and remained very low (<1%) throughout the study period, although this may be a reflection of the diminished use of stavudine and zidovudine in first-line regimens.

We included a number of TRAMs that are not currently in the WHO SDRM list, but that are associated with TDF selection pressure [17]. We did identify sequences with these TRAMs, in particular the A62V mutation, which was present both with and without the signature K65R mutation. Further work is required to understand the significance of these mutations and their effect on response to TDF-based regimens.

Without appropriate action, PDR at the levels we have documented would be likely to have a significant impact on the HIV epidemic in South Africa. One mathematical model suggested that with PDR prevalence ≥10% and no change in the rates of resistance acquisition and transmission, 16% more AIDS deaths each year, 9% higher HIV incidence, and 8% higher ART costs would be attributable to drug resistance in Africa between 2016 and 2030 [62]. Once prevalence of NNRTI PDR exceeds 10%, the WHO recommends that national programmes consider switching to an alternative non-NNRTI first-line ART regimen [46]. Many countries, including South Africa, have taken the decision to transition to a new first-line regimen of co-formulated generic TDF, 3TC and dolutegravir (DTG) [63]. This is the option that mathematical models have predicted will mitigate the effects of HIVDR, will produce the greatest health benefits and reduce overall programme costs [64,65]. However, there remain unanswered questions around DTG in the South African context, and strengthening of HIVDR surveillance and response systems will still be important to maximise the impact of the new regimen [66,67].

An alternative approach to the modified first-line ART regimen would be to introduce pretreatment HIVDR testing and shift towards individualised drug regimens [46]. Whilst there is some evidence that HIVDR testing can be implemented in a research setting in South Africa [68], there is no evidence that it can be delivered cost-effectively through the public health system. The shift towards more rapid initiation of ART (including same-day initiation) would make it particularly challenging to deliver pretreatment HIVDR testing. We still lack simple, rapid, and inexpensive HIVDR assays, although there are promising technologies in development [69]. Given the increasing complexity of HIV care and the uncertainty about the long-term effectiveness of DTG-based regimens, there is still a need to develop and evaluate HIVDR assays and pretreatment HIVDR testing strategies.

We believe it would be a mistake to think that modifying the firstline ART regimen is an adequate response on its own to the rising levels of PDR. Whilst there will clearly be a reduced risk of drug resistance emergence with DTG-based regimens, the public health approach to ART creates scenarios where the risk may be higher, particularly where DTG is the only fully active agent in the regimen [66,67]. The increasing prevalence of PDR reflects weaknesses in prevention, treatment, and care. Although South Africa implements routine viral load monitoring for people on ART, there are critical gaps in the viral load testing cascade and long delays in switching people with virological failure to second-line regimens [70]. This means there is probably an expanding pool of people with acquired HIVDR who can then transmit drug-resistant virus to susceptible individuals. Our findings therefore support calls to focus on improving the quality of HIV services [71]. This needs to be rooted within a broader multisectoral response, informed by high quality transdisciplinary research, that addresses the social and structural drivers of the epidemic [72].

Interpretation of our findings should be subject to some limitations beyond those already discussed. Firstly, certain provinces were overrepresented in our analysis, particularly KwaZulu-Natal and Gauteng, and estimates from the latter years were dominated by two large population-based surveillance studies from KwaZulu-Natal. Findings from the national PDR survey in 2013–14 suggested substantial heterogeneity between the provinces in levels of PDR, and therefore our estimates may not reflect the situation throughout the country [9]. Secondly, we pooled results from a number of individual studies, not all of which were designed to evaluate PDR. We did not account for individual study design in our analysis and derived only pooled crude estimates of prevalence. Our estimates should therefore not be taken to represent

population prevalence. Lastly, we analysed only sequence data and were unable to explore differences by sex, age, CD4+ cell count, and duration of infection, as this information was not available for the majority of sequences.

In conclusion, we present evidence that the prevalence of PDR has risen substantially in South Africa in the past few years. Whilst this is predominantly NNRTI-resistance, there is also evidence of rising levels of resistance to TDF and FTC/3TC, although the absolute prevalence of PDR to these drugs remains low. Our findings support the decision to transition to a new, DTG-based first-line ART regimen. If the association between neural tube defects and DTG is confirmed, and NNRTIs continue to be recommended for women of childbearing age [73], this evidence would suggest the need for additional interventions, such as pre-treatment genotypic resistance testing or early VL testing. These findings also highlight the need for broader strengthening of HIV services within the public health system if we are to eliminate HIV/AIDS as a public health threat by 2030.

Contributors

BC, RJL, S-YR and TDO were responsible for the study conception and design; BC, RJL, S-YR, AK, GH, PK, JM, and TDO were responsible for acquisition of data; BC, RJL, S-YR, JG, KN, LL, RS, AV, and TDO were responsible for data analysis; BC, RJL, S-YR, JG, AK, KN, LL, CC, DK, KAA, KD, RS, GH, AV, BS-P, MG, TM, PK, GR, JL, MK, LM, UMP, JWM, RWS, DK, PM, RKG, DP, SSAK and TDO were responsible for data interpretation, and critically revising the manuscript for important intellectual content. All authors approved the final version of the manuscript.

Declaration of Interests

We declare no competing interests.

Acknowledgments

We acknowledge support from the Stanford–SPARK program (Stanford University Medical Center), Letten Foundation, Norway, the Poliomyelitis Research Foundation, and the National Health Laboratory Services, Department of Virology, at the University of KwaZulu-Natal. The funder had no role in study design, data collection, data analysis, data interpretation, or writing of the report. The corresponding authors had full access to all the data and had final responsibility for the decision to submit the manuscript for publication.

Appendix A. Supplementary data

Supplementary data to this article can be found online at https://doi.org/10.1016/j.eclinm.2019.03.006.

References

- Joint United Nations Programme on HIV/AIDS (UNAIDS). Ending AIDS: progress towards the 90-90-90 targets. Geneva, Switzerland. http://www.unaids.org/sites/ default/files/media_asset/Global_AIDS_update_2017_en.pdf; 2017.
- [2] Gupta RK, Gregson J, Parkin N, et al. HIV-1 drug resistance before initiation or reinitiation of first-line antiretroviral therapy in low-income and middle-income countries: a systematic review and meta-regression analysis. Lancet Infect Dis 2018:18:346–55.
- [3] World Health Organization. The HIV drug resistance report 2017. Geneva, Switzerland. http://apps.who.int/iris/bitstream/10665/255896/1/9789241512831-eng.pdf?ua=1; 2017.
- [4] World Health Organization. Global action plan on HIV drug resistance 2017–2021. Geneva, Switzerland. http://apps.who.int/iris/bitstream/10665/ 255883/1/9789241512848-eng.pdf; 2017.
- [5] Wittkop L, Günthard HF, de Wolf F, et al. Effect of transmitted drug resistance on virological and immunological response to initial combination antiretroviral therapy for HIV (EuroCoord-CHAIN joint project): a European multicohort study. Lancet Infect Dis 2017:11:363–71.
- [6] Hamers RL, Schuurman R, Sigaloff KCE, et al. Effect of pretreatment HIV-1 drug resistance on immunological, virological, and drug-resistance outcomes of first-line

- antiretroviral treatment in sub-Saharan Africa: a multicentre cohort study. Lancet Infect Dis 2012;12:307–17.
- [7] Department of Health Republic of South Africa. Debate on the health budget vote. http://www.health.gov.za/index.php/2014-03-17-09-48-36/speeches?download= 2756:budget-vote-speech: 2018.
- [8] Manasa J, Katzenstein D, Cassol S, Newell M-L, de Oliveira T. Primary drug resistance in South Africa: data from 10 years of surveys. AIDS Res Hum Retroviruses 2012;28: 558-65.
- [9] Steegen K, Carmona S, Bronze M, et al. Moderate levels of pre-treatment HIV-1 antiretroviral drug resistance detected in the first South African National Survey. PLoS One 2016:11:e0166305.
- [10] Manasa J, Danaviah S, Lessells R, et al. Increasing HIV-1 drug resistance between 2010 and 2012 in adults participating in population-based HIV surveillance in rural KwaZulu-Natal. South Africa. AIDS Res Hum Retroviruses 2016;32:763–9.
- [11] Moher D, Liberati A, Tetzlaff J, Altman DG, Group and the P. Preferred reporting items for systematic reviews and meta-analyses: the PRISMA statement. Ann Intern Med 2009:151:264–9
- [12] Clark K, Karsch-Mizrachi I, Lipman DJ, Ostell J, Sayers EW. GenBank. Nucleic Acids Res 2016:44:D67–72.
- [13] Hamers RL, Wallis CL, Kityo C, et al. HIV-1 drug resistance in antiretroviral-naive individuals in sub-Saharan Africa after rollout of antiretroviral therapy: a multicentre observational study. Lancet Infect Dis 2011;11:750–9.
- [14] Larsson A. AliView: a fast and lightweight alignment viewer and editor for large datasets. Bioinformatics 2014;30:3276–8.
- [15] Gifford RJ, Liu TF, Rhee S-Y, et al. The calibrated population resistance tool: standardized genotypic estimation of transmitted HIV-1 drug resistance. Bioinformatics 2009:25:1197–8.
- [16] Bennett DE, Camacho RJ, Otelea D, et al. Drug resistance mutations for surveillance of transmitted HIV-1 drug-resistance: 2009 update. PLoS One 2009;4:e4724.
- [17] Rhee SY, Varghese V, Holmes SP, et al. Mutational correlates of virological failure in individuals receiving a WHO-recommended Tenofovir-containing first-line regimen: an international collaboration. EBioMedicine 2017;18:225–35.
- [18] Hunt G, Ledwaba J, Salimo A, et al. Surveillance of transmitted HIV-1 drug resistance in five provinces in South Africa in 2011. Commun Dis Surveill Bull 2013; 11:122–5.
- [19] Hunt G, Ledwaba J, Kalimashe M, et al. National surveillance of transmitted HIV-1 drug resistance in 2012. Commun Dis Surveill Bull 2015;13:30–2.
- [20] Iweriebor BC, Mavhandu LG, Masebe T, et al. Molecular epidemiology of HIV in two highly endemic areas of northeastern South Africa. Arch Virol 2012;157: 455–65.
- [21] Jacobs GB, Laten A, van Rensburg EJ, et al. Phylogenetic diversity and low level antiretroviral resistance mutations in HIV type 1 treatment-naive patients from Cape Town, South Africa. AIDS Res Hum Retroviruses 2008;24:1009–12.
- [22] Jacobs GB, Wilkinson E, Isaacs S, et al. HIV-1 subtypes B and C unique recombinant forms (URFs) and transmitted drug resistance identified in the Western Cape Province, South Africa. PLoS One 2014;9:e90845.
- [23] Matthews PC, Prendergast A, Leslie A, et al. Central role of reverting mutations in HLA associations with human immunodeficiency virus set point. J Virol 2008;82: 8548–59.
- [24] Msimanga PW, Vardas E, Engelbrecht S. HIV-1 diversity in an antiretroviral treatment naïve cohort from Bushbuckridge, Mpumalanga Province, South Africa. Virol I 2015:12:24.
- [25] Musyoki AM, Rakgole JN, Selabe G, Mphahlele J. Identification and genetic characterization of unique HIV-1 A1/C recombinant strain in South Africa. AIDS Res Hum Retroviruses 2015;31:347–52.
- [26] Nwobegahay J, Bessong P, Masebe T, et al. Prevalence of drug-resistant mutations in newly diagnosed drug-naïve HIV-1-infected individuals in a treatment site in the Waterberg district, Limpopo province. S Afr Med J 2011;101:335–7.
- [27] Nwobegahay JM, Bessong PO, Masebe TM, Mavhandu LG, Iweriebor BC, Selabe G. Prevalence of antiretroviral drug resistance mutations and HIV-1 subtypes among newly-diagnosed drug-naïve persons visiting a voluntary testing and counselling centre in Northeastern South Africa. J Health Popul Nutr 2011;29:303–9.
- [28] Nwobegahay J, Selabe G, Ndjeka NO, Manhaeve C, Bessong PO. Low prevalence of transmitted genetic drug resistance in a cohort of HIV infected naive patients entering antiretroviral treatment programs at two sites in northern South Africa. J Med Virol 2012;84:1839–43.
- [29] Papathanasopoulos MA, Vardas E, Wallis C, et al. Characterization of HIV type 1 genetic diversity among south African participants enrolled in the AIDS Vaccine Integrated Project (AVIP) study. AIDS Res Hum Retroviruses 2010;26:705–9.
- [30] Parboosing R, Naidoo A, Gordon M, Taylor M, Vella V. Resistance to antiretroviral drugs in newly diagnosed, young treatment-naive HIV-positive pregnant women in the province of KwaZulu-Natal, South Africa. J Med Virol 2011;83:1508–13.
- [31] Parikh UM, Kiepiela P, Ganesh S, et al. Prevalence of HIV-1 drug resistance among women screening for HIV prevention trials in KwaZulu-Natal, South Africa (MTN-009). PLoS One 2013;8:e59787.
- [32] Pillay C, Bredell H, McIntyre J, Gray G, Morris L. HIV-1 subtype C reverse transcriptase sequences from drug-naive pregnant women in South Africa. AIDS Res Hum Retroviruses 2002;18:605–10.
- [33] Pillay V, Ledwaba J, Hunt G, et al. Antiretroviral drug resistance surveillance among drug-naive HIV-1-infected individuals in Gauteng Province, South Africa in 2002 and 2004. Antivir Ther 2008:13:101–7.
- [34] Seoighe C, Ketwaroo F, Pillay V, et al. A model of directional selection applied to the evolution of drug resistance in HIV-1. Mol Biol Evol 2007;24:1025–31.
- [35] Treurnicht FK, Seoighe C, Martin DP, et al. Adaptive changes in HIV-1 subtype C proteins during early infection are driven by changes in HLA-associated immune pressure. Virology 2010;396:213–25.

- [36] Van Zyl GU, Grobbelaar CJ, Claassen M, Bock P, Preiser W. Moderate levels of preantiretroviral therapy drug resistance in a generalized epidemic: time for better first-line ART? Aids 2017;31:2387–91.
- [37] Wilkinson E, Engelbrecht S, de Oliveira T. Detection of transmission clusters of HIV-1 subtype C over a 21-year period in Cape Town, South Africa. PLoS One 2014;9: e109296.
- [38] Chimukangara B, Kharsany ABM, Lessells RJ, et al. Moderate-to-high levels of pretreatment HIV drug resistance in KwaZulu-Natal Province, South Africa. AIDS Res Hum Retroviruses 2019;35:129–38.
- [39] Bessong PO, Larry Obi C, Cilliers T, et al. Characterization of human immunodeficiency virus type 1 from a previously unexplored region of South Africa with a high HIV prevalence. AIDS Res Hum Retroviruses 2005;21:103–9.
- [40] Bessong PO, Mphahlele J, Choge IA, et al. Resistance mutational analysis of HIV type 1 subtype C among rural South African drug-naive patients prior to large-scale availability of antiretrovirals. AIDS Res Hum Retroviruses 2006;22:1306–12.
- [41] Gordon M, De Oliveira T, Bishop K, et al. Molecular characteristics of human immunodeficiency virus type 1 subtype C viruses from KwaZulu-Natal, South Africa: implications for vaccine and antiretroviral control strategies. I Virol 2003;77:2587–99.
- [42] Huang K, Goedhals D, Fryer H, et al. Prevalence of HIV type-1 drug-associated mutations in pre-therapy patients in the Free State, South Africa. Antivir Ther 2009;14: 975–84
- [43] Hunt GM, Ledwaba J, Basson AE, et al. Surveillance of transmitted HIV-1 drug resistance in Gauteng and KwaZulu-Natal provinces, South Africa, 2005–2009. Clin Infect Dis 2012;54:S334–8.
- [44] Hunt G, Ledwaba J, Basson A, et al. Surveillance of transmitted HIV-1 drug resistance in Gauteng and KwaZulu-Natal in 2010. Commun Dis Surveill Bull 2012;10:86–9.
- [45] Joint United Nations Programme on HIV/AIDS (UNAIDS). Country profile: South Africa. http://www.unaids.org/en/regionscountries/countries/southafrica; 2018.
- [46] World Health Organization. Guidelines on the public health response to pretreatment HIV drug resistance. Geneva, Switzerland. http://apps.who.int/iris/bitstream/ 10665/255880/1/9789241550055-eng.pdf; 2017.
- [47] Rhee S-Y, Blanco JL, Jordan MR, et al. Geographic and temporal trends in the molecular epidemiology and genetic mechanisms of transmitted HIV-1 drug resistance: an individual-patient and sequence-level meta-analysis. PLoS Med 2015;12:e1001810.
- [48] Steegen K, Bronze M, Papathanasopoulos MA, et al. HIV-1 antiretroviral drug resistance patterns in patients failing NNRTI-based treatment: results from a national survey in South Africa. J Antimicrob Chemother 2017;72:210–9.
- [49] Wertheim JO, Oster AM, Johnson JA, et al. Transmission fitness of drug-resistant HIV revealed in a surveillance system transmission network. Virus Evol 2017;3:2045–52.
- [50] Kühnert D, Kouyos R, Shirreff G, et al. Quantifying the fitness cost of HIV-1 drug resistance mutations through phylodynamics. PLoS Pathog 2018;14:1–16.
- [51] Castro H, Pillay D, Cane P, et al. Persistence of HIV-1 transmitted drug resistance mutations. J Infect Dis 2013;208:1459–63.
- [52] Coovadia A, Hunt G, Abrams EJ, et al. Persistent minority K103N mutations among women exposed to single-dose nevirapine and virologic response to nonnucleoside reverse-transcriptase inhibitor-based therapy. Clin Infect Dis 2009;48:462–72.
- [53] Flys TS, Donnell D, Mwatha A, et al. Persistence of K103N-containing HIV-1 variants after single-dose nevirapine for prevention of HIV-1 mother-to-child transmission. J Infect Dis 2007;195:711–5.
- [54] Kim AA, Mukui I, Young PW, et al. Undisclosed HIV infection and antiretroviral therapy use in the Kenya AIDS indicator survey 2012: relevance to national targets for HIV diagnosis and treatment. AIDS 2016;30:2685–95.
- [55] Manne-Goehler J, Rohr J, Montana L, et al. ART denial: results of a home-based study to validate self-reported antiretroviral use in rural South Africa. AIDS Behav 2018. https://doi.org/10.1007/s10461-018-2351-7.
- [56] Grabowski MK, Reynolds SJ, Kagaayi J, et al. The validity of self-reported antiretroviral use in persons living with HIV: a population-based study. Aids 2018;32:363–9.
- [57] Moyo S, Gaseitsiwe S, Powis KM, et al. Undisclosed antiretroviral drug use in Botswana: implication for national estimates. AIDS 2018;32:1543–6.
- [58] Fogel JM, Wang L, Parsons TL, et al. Undisclosed antiretroviral drug use in a multinational clinical trial (HIV prevention trials network 052). J Infect Dis 2013;208: 1624–8
- [59] Kahle EM, Kashuba A, Baeten JM, et al. Unreported antiretroviral use by HIV-1-infected participants enrolling in a prospective research study. J Acquir Immune Defic Syndr 2014;65:e90–4.
- [60] Yang WL, Kouyos RD, Böni J, et al. Persistence of transmitted HIV-1 drug resistance mutations associated with fitness costs and viral genetic backgrounds. PLoS Pathog 2015;11:1–13.
- [61] Nsanzimana S, Binagwaho A, Kanters S, Mills EJ. Churning in and out of HIV care. Lancet HIV 2014;1:e58–9.
- [62] Phillips AN, Stover J, Cambiano V, et al. Impact of HIV drug resistance on HIV/AIDSassociated mortality, new infections, and antiretroviral therapy program costs in sub-Saharan Africa. J Infect Dis 2017;215:1362–5.
- [63] Joint United Nations Programme on HIV/AIDS (UNAIDS). New high-quality antiretroviral therapy to be launched in South Africa, Kenya and over 90 low-and middle-income countries at reduced price; 2018http://www.unaids.org/en/resources/presscentre/pressreleaseandstatementarchive/2017/september/20170921_TLD.
- [64] Phillips AN, Cambiano V, Nakagawa F, et al. Cost-effectiveness of public-health policy options in the presence of pretreatment NNRTI drug resistance in sub-Saharan Africa: a modelling study. Lancet HIV 2018;5:e146–54.
- [65] Phillips AN, Venter F, Havlir D, et al. Risks and benefits of dolutegravir-based antiretroviral drug regimens in sub-Saharan Africa: a modelling study. Lancet HIV 2019. https://doi.org/10.1016/S2352-3018(18)30317-5 published online Jan.
- [66] Dorward J, Lessells R, Drain PK, et al. Dolutegravir for first-line antiretroviral therapy in low-income and middle-income countries: uncertainties and opportunities for implementation and research. Lancet HIV 2018;5:e400-4.

- [67] de Waal R, Lessells R, Hauser A, et al. HIV drug resistance in sub-Saharan Africa: public health questions and the potential role of real-world data and mathematical modelling. J Virus Erad 2018;4:55–8.

 [68] Lessells RJ, Stott KE, Manasa J, et al. Implementing antiretroviral resistance testing in a
- primary health care HIV treatment programme in rural KwaZulu-Natal, South Africa: early experiences, achievements and challenges. BMC Health Serv Res 2014;14:116.
- [69] Duarte HA, Panpradist N, Beck IA, et al. Current status of point-of-care testing for human immunodeficiency virus drug resistance. J Infect Dis 2017;216:S824–8.
 [70] Murphy RA, Court R, Maartens G, Sunpath H. Second-line antiretroviral therapy in sub-Saharan Africa: it is time to mind the gaps. AIDS Res Hum Retroviruses 2017; 33:1181-4.
- [71] Venter W, Ford N, Vitoria M, Stevens W. Diagnosis and monitoring of HIV programmes to support treatment initiation and follow up and improve programme quality. Curr Opin HIV AIDS 2017;12:117–22.
- Hopkins KL, Doherty T, Gray GE. Will the current National Strategic Plan enable South Africa to end AIDS, tuberculosis and sexually transmitted infections by 2022? South African J HIV Med 2018;19(1) DO - 104102/sajhivmed.v19i1796 2018; published online Oct 4. https://sajhivmed.org.za/index.php/hivmed/article/view/796.
 [73] Zash R, Makhema J, Shapiro RL. Neural-tube defects with dolutegravir treatment
- from the time of conception. N Engl J Med 2018;379:979–81.