Tau positron emission tomography ligands provide the novel possibility to image tau pathology in vivo. However, little is known about how in vivo brain uptake of tau positron emission tomography ligands relates to tau aggregates observed post-mortem. We performed tau positron emission tomography imaging with $^{18}$F-AV-1451 in three patients harbouring a p.R406W mutation in the MAPT gene, encoding tau. This mutation results in 3- and 4-repeat tau aggregates similar to those in Alzheimer’s disease, and many of the mutation carriers initially suffer from memory impairment and temporal lobe atrophy. Two patients with short disease duration and isolated memory impairment exhibited $^{18}$F-AV-1451 uptake mainly in the hippocampus and adjacent temporal lobe regions, correlating with glucose hypometabolism in corresponding regions. One patient died after 26 years of disease duration with dementia and behavioural deficits. Pre-mortem, there was $^{18}$F-AV-1451 uptake in the temporal and frontal lobes, as well as in the basal ganglia, which strongly correlated with the regional extent and amount of tau pathology in post-mortem brain sections. Amyloid-β ($^{18}$F-flutemetamol) positron emission tomography scans were negative in all cases, as were stainings of brain sections for amyloid. This provides strong evidence that $^{18}$F-AV-1451 positron emission tomography can be used to accurately quantify in vivo the regional distribution of hyperphosphorylated tau protein.
Introduction

Tau pathology is closely related to neuronal dysfunction and degeneration in Alzheimer’s disease and other tauopathies. In these disorders, tau is abnormally hyperphosphorylated and accumulates intracellularly, forming tangles of paired helical filaments (PHF), twisted ribbons and/or straight filaments (lqbal et al., 2016). Tau aggregates in Alzheimer’s disease consist predominantly of PHF, which contain all six isoforms of tau, mixed with straight filaments. The recent development of PET tracers for tau aggregates opens up the possibility to assess tau pathology in vivo (Ossenkoppele et al., 2015; Villemagne and Okamura, 2015; Harada et al., 2016; Johnson et al., 2016; Scholl et al., 2016; Smith et al., 2016). These tracers, along with PET tracers revealing extracellular plaques containing amyloid-β fibrils, will be essential for improving the early diagnosis of Alzheimer’s disease in clinical practice as well as in the evaluation of novel disease-modifying therapies (Dani et al., 2016).

Studies using the most common PET tracer for tau pathology to date, 18F-AV-1451, demonstrated increased uptake in tempo-parietal regions in patients with Alzheimer’s disease (Chien et al., 2013; Johnson et al., 2016; Ossenkoppele et al., 2016; Smith et al., 2016). In post-mortem brain tissue, this ligand was screened to detect PHFs of tau, exhibiting a >25-fold higher affinity for PHF-tau compared to amyloid-β fibrils (Xia et al., 2013). Recently, an autoradiography study reported preferential in vitro binding of 18F-AV-1451 to PHFs in Alzheimer’s disease, not to straight filaments resulting from other tauopathies (Marquie et al., 2015). However, no study has hitherto compared regional in vivo binding of a tau PET tracer with the distribution and the degree of tau pathology using post-mortem immunohistochemistry in the same individual.

Individuals exhibiting only tau pathology, not amyloid-β fibrils, are an ideal group to further explore the binding properties of a tau PET tracer. The R406W mutation in the gene encoding the tau protein (MAPT C.1216C>T; R406W) on chromosome 17 results in neurofibrillary tangles that are similar to the neurofibrillary tangles in Alzheimer’s disease at an ultrastructural level (Ghetti et al., 2015). Many cases with this mutation present with slowly progressive episodic memory impairment, which in many ways clinically resembles Alzheimer’s disease (Ostojc et al., 2004; Passant et al., 2004). Previous neuropathological examination of R406W mutation carriers revealed tau-positive neurofibrillary tangles and neurites. These PHF-tau changes were mainly localized in temporal and frontal lobe areas as well as the basal ganglia, sparing the parietal and occipital lobes (Lindquist et al., 2008; Ghetti et al., 2015).

Here we report the first results of 18F-AV-1451 (tau), 18F-flutemetamol (amyloid-β) and 18F-fluorodeoxyglucose (FDG, glucose metabolism) PET imaging in three patients carrying the MAPT R406W mutation at different disease stages. In addition, we report results from the neuropathological examination of one of these subjects who died 2 weeks after the PET scans, as well as the relationship between in vivo and post-mortem assessment of tau pathology in this case. Finally, autoradiography using 3H-AV1451 was performed on cortical tissue sections from two additional deceased patients with the MAPT R406W mutation.

Materials and methods

Participants

Three subjects carrying the MAPT R406W mutation, aged 56, 60 and 76 years, (Patients A, B and C, respectively) from a family with hereditary dementia from Southern Sweden were investigated. Four control subjects and five patients with Alzheimer’s disease were recruited from the ongoing Swedish Biofinder Study (www.biofinder.se). Further information on the clinical assessments is given in the Supplementary material. All clinical data are summarized in Table 1. Written informed consent was obtained from the participants, and approval of this study was obtained from the Regional Ethical Review Board in Lund, Sweden.

MRI and CT scan

Patients A and B underwent a 3 T MRI scan using a Siemens Skyra scanner. T1-weighted magnetization-prepared rapid gradient echo (T1-MPRAGE) was acquired at a resolution of 1 x 1 x 1 mm. Patient C could not undergo MRI due to claustrophobia and consequently underwent a CT scan, using a Philips Brilliance 64 scanner. Images were reconstructed to 5 mm axial and 3 mm coronal slices.

PET

18F-AV-1451 PET scans were performed on a GE Discovery 690 PET scanner (General Electric Medical Systems) as dynamic scans using LIST-mode 100–120 min after a bolus injection of 365 ± 14 MBq of 18F-AV-1451. 18F-FDG scans were acquired on the same scanner in a static scan 60–80 min after injection of 198 ± 2 MBq of FDG. Images of 18F-flutemetamol (Nelissen et al., 2009) were obtained as dynamic scans 90–110 min after injection of 193 ± 2 MBq on a Philips Gemini TF PET-CT scanner (Philips Medical Systems). Low dose CT scans for attenuation correction were performed immediately prior to the PET scans. PET data were reconstructed into 5 min frames using an iterative Vue Point HD algorithm with six subsets, 18 iterations with 3 mm filter and no time-of-flight correction. 18F-flutemetamol data were processed using NeuroMarQ software (GE Healthcare), analyses of 18F-AV-1451 and 18F-FDG data were performed using PMOD 3.603 software (Pmod, Zurich, Switzerland). PET images were co-registered to their corresponding magnetic resonance T1-MPRAGE or CT images. We chose to analyse the 18F-AV-1451 standardized uptake value ratios (SUVRs) from the same areas that were processed for neuropathological analysis in Patient C, with the addition of a few areas in the
temporal and frontal lobes and the basal ganglia, areas known to be affected by pathology in R406W mutation carriers. Regions of interest were delineated manually for the data presented in Figs 1 and 3. Regions of interest were delineated on the magnetic resonance images (Patients A and B, as well as control subjects and cases with Alzheimer’s disease) or CT image (Patient C) and transformed into PET space using transformations derived from co-registrations. The cerebellar cortex was used as a reference region for the calculation of SUVRs.

For the analysis of correlations between regional 18F-FDG SUVRs and 18F-AV-1451 SUVRs in Patients A and B (Fig. 2), the MRI was segmented into grey and white matter and regions of interest were defined using FreeSurfer (version 5.3 and its Desikan-Killiany Atlas). PET images were co-registered to the MRIs and SUVRs calculated in PET space using transformations derived from co-registrations. The cerebellar cortex was also used as a reference region for the calculation of SUVRs. Further, the SUVRs of the 18F-FDG scans were normalized to the mean control 18F-FDG value for each region of interest. The correlation between neurite density, the neuropathological grade of tau pathology and 18F-AV-1451 SUVR values as well as the correlation between 18F-FDG and 18F-AV-1451 SUVR were analysed with non-parametric Spearman’s Rank-Order Correlation using Graph Pad Prism 6. A P-value of < 0.05 was considered statistically significant.

### Results

#### Study subjects

The three examined patients belonged to two generations of a family with slowly progressive dementia; all carried the MAPT R406W mutation. Cognitive testing of Patients A and B revealed an isolated memory impairment (Table 1), and both fulfilled the criteria of mild cognitive impairment (Petersen, 2004). Patient A had a 10-year history of isolated short-term memory loss and irritability and Patient B had developed mild memory impairment and increased anxiety.
4 years prior to the clinical examination. MRI in both cases showed no or slight general brain atrophy with moderate atrophy in the medial temporal lobes, most pronounced in the parahippocampal gyri with pronounced widening of the collateral sulci. Medial temporal atrophy corresponded to Scheltens score 1. Patient C exhibited memory impairment 26 years prior to this examination and had developed marked dementia, behavioural disturbances, mutism, dysphagia and mild parkinsonism by the time of this study.

The CT scan showed generalized cortical atrophy with substantial atrophy of the medial temporal lobes, especially of the parahippocampal gyri with pronounced widening of the collateral sulci. Medial temporal atrophy corresponded to Scheltens score 3.

\[ ^{18} \text{F-AV-1451} \] and \[ ^{18} \text{F-flutemetamol} \] PET

In the two younger cases (Patients A and B; Fig. 1A, B and I), retention of \[ ^{18} \text{F-AV-1451} \] was most distinct in the temporal poles, the hippocampus and anterior aspects of the inferior temporal gyrus. Uptake was also detected in the basal regions of the frontal lobes. In the more affected case (Patient C, Fig. 1C and I), a more widespread retention pattern was observed in the temporal lobes, the basal ganglia and the frontal lobes. No significant tracer uptake was detected in the parietal or occipital lobes. Control cases showed no tracer retention in cortical areas (Fig. 1D and Supplementary Fig. 1).

The patients further underwent \[ ^{18} \text{F-flutemetamol} \] PET scans. Composite scores representing mean global cortical uptake were below a previously determined cut-off for amyloid-\( \beta \)-positivity (<1.42; Palmqvist et al., 2014) in the three patients who did not show visual signs of cortical \[ ^{18} \text{F-flutemetamol} \] retention either (Fig. 1E–G).

Correlation of \[ ^{18} \text{F-AV-1451} \] with \[ ^{18} \text{F-FDG} \]

Patients A and B underwent an additional \[ ^{18} \text{F-FDG} \] scan. The \[ ^{18} \text{F-FDG} \] SUVRs for each region were normalized to the controls’ regional SUVRs and correlated with the corresponding AV-1451 SUVRs (Fig. 2A–D). We found statistically significant inverse relationships of higher regional AV-1451 uptake with lower FDG retention in both patients (Patient A: \( r_s = -0.39, P < 0.01 \), Fig. 2E; Patient B: \( r_s = -0.37, P < 0.01 \), Fig. 2F).

Neuropathology

Patient C died 2 weeks after the PET examinations. Immunohistochemical staining for hyperphosphorylated tau protein showed extensive neuronal tau pathology in the temporo-limbic region, the anterior temporal lobe and hippocampus, where tau-positive neurofibrillary tangles were abundant along with tau-positive neurites. Lower-grade pathology was detected in frontal cortical areas, whereas posterior cortical areas were only affected to a minor degree. Tau pathology was assessed according to a four-point grading system (0, +, ++, +++), as described above, and results are summarized in Fig. 3A–J. Both 3R and 4R tau-positive neurites could be detected using isotype specific antibodies (data not shown). No amyloid-\( \beta \)-containing neuritic plaques or amyloid angiopathy were detected (data not shown).

Non-parametric correlation analysis showed a statistically highly significant positive correlation between
18F-AV-1451 SUVRs in different brain regions and the severity of tau-neuropathology according to immunohistochemistry in the same regions when using either neuropathological grading of tau pathology ($r = 0.93$, $P < 0.01$; Fig. 3I) or quantification of the density of tau-positive neurites ($r = 0.92$, $P < 0.01$; Fig. 3K). Interestingly, the region with the highest SUVR (2.85) was putamen, which did not exhibit higher tau pathology density than the inferior temporal lobe with an SUVR of 2.38.

**Discussion**

In the present study we describe three members of a family with MAPT R406W mutation at different stages of a degenerative neurological disorder. *In vivo* imaging with 18F-AV-1451 PET indicated that tau pathology in these cases starts in the inferior temporal lobes and temporal poles, and at a later stage also involves the basal ganglia and the frontal cortex while sparing posterior cortical areas.
and the cerebellum. In one patient who died shortly after PET scanning and was examined neuropathologically, we found a strong positive correlation between the regional in vivo retention of 18F-AV-1451 and the density of tau aggregates visualized with immunohistochemistry in post-mortem brain tissue. Our study is the first to report 18F-AV-1451 PET signal being directly correlated to neuropathological tau burden in the same individual, which provides strong evidence that in vivo 18F-AV-1451 PET retention indeed reflects the density of tau pathology consisting of PHFs. However, it is known that unspecific retention of 18F-AV-1451 occurs in the striatum and choroid plexus, for example (Scholl et al., 2016). It is therefore interesting to note that the region with highest retention of 18F-AV-1451 in Patient C was the putamen. The neuropathological evaluation revealed relatively severe tau pathology in this region, but even greater pathological burden was observed in the inferior temporal lobe, which exhibited a slightly lower retention of 18F-AV-1451 (Fig. 3K), indicating that a proportion of the signal in the putamen might represent unspecific tracer retention in addition to the specific binding to tau aggregates.

The R406W mutation is located in the N-terminal exon 13 of the MAPT gene and causes the formation of aggregations containing six isoforms of tau (including 3R and 4R tau) highly similar to the aggregates found in Alzheimer’s disease (Hong et al., 1998; Ghetti et al., 2015), which was corroborated in the present study.
At an ultrastructural level the tau filaments are PHF and straight filaments with a diameter of 8–20 nm, structurally similar to filaments found in Alzheimer’s disease (Reed et al., 1997; Hutton et al., 1998). It has previously been shown that 18F-AV-1451 can detect tau aggregates in patients with Alzheimer’s disease (Ossenkoppele et al., 2015; Johnson et al., 2016; Smith et al., 2016). Our study provides proof of concept that it is also possible to visualize tau pathology in R406W tau mutation carriers using 18F-AV-1451 PET. This is in accordance with a recent study reporting highly concordant patterns of staining with tau-antibody AT8 and 18F-AV-1451 binding in post-mortem brain tissue of among others, Alzheimer’s disease patients and subjects carrying MAPT mutations (Sander et al., 2016), although this study also found evidence for off-target binding and lack of correlation between antibody staining and in vivo tracer binding in certain tau strains.

However, the present study indicates that the in vivo binding of 18F-AV-1451 correlates highly with the extent of tau aggregates containing PHF. With tau immunization therapies for Alzheimer’s disease currently emerging (Pedersen and Sigurdsson, 2015), tau mutation carriers constitute the ideal population for studying the effects of these therapies. Not only would it be possible to examine the effects of anti-tau treatment in the absence of concurrent amyloid-β pathology, but also to identify and recruit subjects to clinical trials at a presymptomatic disease stage. Our results also suggest that one study endpoint in such clinical trials could be 18F-AV-1451 retention.

Furthermore, we found an inverse relationship between the retention of 18F-AV-1451 and brain glucose metabolism as measured by 18F-FDG, indicative of decreased neuronal metabolism and dysfunction in areas affected by tau pathology. These results indicate that tau-pathology is tightly associated with neuronal dysfunction, which is in line with previous publications on tau and glucose hypometabolism in Alzheimer’s disease (Ossenkoppele et al., 2015; Smith et al., 2016). Similarly, cognitive testing in Patients A and B only showed deficits restricted to memory, likely reflecting the limited distribution of tau pathology and glucose hypometabolism to the inferior and medial temporal lobes in these two cases.

In conclusion, our results strongly support the notion that in vivo 18F-AV-1451 PET reflects the intensity of regional tau neuropathology. The level of 18F-AV-1451 retention correlates with glucose hypometabolism and neuronal dysfunction. Tau pathology in R406W mutation carriers can be visualized using 18F-AV-1451 PET, thus providing a useful tool for monitoring the effects of future therapies directed against the formation of tau-inclusions in these patients.

**Acknowledgements**

We wish to acknowledge the contribution of Dr Olof Strandberg, Dr Douglas Hägerström and Dr Jonas Jögi at Skåne University Hospital. The precursor of AV-1451 was provided by Avid Radiopharmaceuticals and [18F]flutemetamol was by GE Healthcare, but no funding was received from the companies. The data were analysed and the manuscript written independent from any company interests.

**Funding**

The study was funded by the Swedish Research Council, the European Research Council, the Swedish Brain Foundation, the Swedish Alzheimer foundation, the Trolle-Wachtmeister Foundation and the Swedish Federal Government under the ALF Agreement.

**Supplementary material**

Supplementary material is available at *Brain* online.

**References**


