

1 **Differential intensity projection (DIP) for visualisation and quantification of plaque**  
2 **neovascularisation in CEUS images of carotid arteries**

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30 **Abstract**

31 Studies have shown that intra-plaque neovascularisation (IPN) is closely correlated with plaque  
32 vulnerability. In this study, a new image processing approach, differential intensity projection  
33 (DIP), was developed to visualise and quantify IPN in contrast enhanced ultrasound (CEUS)  
34 image sequences of carotid arteries. DIP used the difference between the local temporal  
35 maximum and the local temporal average signals to identify bubbles against tissue background  
36 and noise. The total absolute and relative areas occupied by bubbles within each plaque were  
37 calculated to quantify IPN. In vitro measurements on a laboratory phantom were made,  
38 followed by in vivo measurements where twenty-four CEUS image sequences of carotid  
39 arteries from 48 patients were acquired. The results using DIP were compared with those  
40 obtained by maximum intensity projection (MIP) and visual assessment. The results show that  
41 DIP can significantly reduce nonlinear propagation tissue artefacts and is much more specific  
42 in detecting bubble signals than MIP, being able to reveal microbubble signals which are buried  
43 in tissue artefacts in the corresponding MIP image. A good correlation was found between  
44 microvascular area (MVA) ( $r = 0.83, p < 0.001$ ) / microvascular density (MVD) ( $r = 0.77, p <$   
45  $0.001$ ) obtained using DIP and the corresponding expert visual grades, comparing favourably  
46 to  $r = 0.26$  and  $0.23$  obtained using MIP on the same data. In conclusion, the proposed method  
47 shows great potential in quantification of IPN in contrast enhanced ultrasound of carotid  
48 arteries.

49

50 **Key words:** differential intensity projection, contrast enhanced ultrasound, carotid artery,  
51 intraplaque neovascularisation, perfusion quantification

52

53

## 54 **Introduction**

55 Stroke is a leading cause of death in the world-wide (Fuster and Voûte 2005). The formation  
56 of vulnerable atherosclerotic plaque in the carotid artery increases the risk of stroke (U-King-  
57 Im et al. 2009; Mughal et al. 2011). Several studies have reported that intraplaque  
58 neovascularisation (IPN) is a precursor of intraplaque haemorrhage (IPH) and IPN could thus  
59 be a surrogate biomarker of unstable plaque (Feinstein 2006; Virmani et al. 2006; Hellings et  
60 al. 2010). Therefore, quantification of IPN can be used for the early detection and clinical  
61 management of unstable atherosclerotic plaques and hence minimise the risk of stroke.

62

63 Recently, contrast-enhanced ultrasound (CEUS) imaging with microbubble contrast agents has  
64 provided a unique tool for visualizing and quantifying IPN. It has shown promise for imaging  
65 plaque vasculature. Several groups (Feinstein 2004; Coli et al. 2008; Giannoni et al. 2009; Lee  
66 et al. 2010) have established correlations between CEUS imaging results and histological  
67 plaque neovascularisation and the risk of plaque rupture. However, in these studies, only  
68 subjective visual assessment was used to quantify the findings. Furthermore, although several  
69 computer algorithms (Hoogi et al. 2012; Akkus et al. 2013) are available to assist in the  
70 quantitative analysis of the images, they have some limitations. Hoogi et al. proposed a method  
71 for segmenting the contrast spots within atherosclerotic plaques in individual images by  
72 tracking individual microbubbles. The main advantage of this approach is that it utilises the  
73 temporal behaviour of bubble flow can be demonstrated. This makes it robust to noise and  
74 allows differentiation between blood vessels and artefacts. However, several parameters of the  
75 algorithm were determined empirically from a few sequences, which may be a variable to  
76 quantitative results. Akkus et al. developed a statistical segmentation of carotid plaque  
77 neovascularisation. An iterative expectation-maximisation algorithm was employed to solve a

78 mixture estimation problem to identify contrast microbubble signals. But, this technique has  
79 difficulties quantifying IPN reliably for plaques located on the far wall of the carotid artery due  
80 to nonlinear propagation artefacts (also called pseudo-enhancement artefact) (Tang and  
81 Eckersley 2006; Tang et al. 2010). Non-linear propagation of ultrasound creates artefacts in  
82 CEUS images that could significantly affect both qualitative and quantitative IPN assessments  
83 (ten Kate et al. 2012). Although there are correction methods (Renaud et al. 2012; Yildiz et al.  
84 2015) to remove non-linear artefact, they are not available on current commercial scanners.

85

86 Moreover, the maximum intensity projection (MIP) is a common intensity-based bubble  
87 imaging method. It can visualise bubble paths (i.e. vessel trajectories) by displaying the  
88 maximum intensity over time for each pixel in CEUS images (Suri et al. 2002; van Ooijen et  
89 al. 2003; Hoogi et al. 2011). While this approach is sensitive, simple and fast, the disadvantage  
90 is that this method has low specificity to bubbles. In particular, it is difficult to distinguish  
91 between tissue artefact due to nonlinear propagation and blood vessels, and therefore it could  
92 generate over-estimated vessel paths and affect quantification results.

93

94 The objective of this study was to develop and evaluate a sensitive, specific, simple and fast  
95 microbubble detection technique for CEUS carotid artery imaging by using differential  
96 intensity projection (DIP). This technique was demonstrated *in vivo*, and applied to the  
97 quantification of intraplaque neovascularisation *in vivo*.

98

99

## 100 **Methods**

101 *Differential intensity projection*

102 The proposed algorithm worked at a pixel level to detect microbubble signals. The CEUS  
103 images contained primarily three components: tissue artefact, noise, and microbubble signals.  
104 The differential intensity projection (DIP) was defined as below to capture the microbubble  
105 signals.

$$DIP(x_i, y_i) = \text{maximun}(I(t, x_i, y_i)) - \langle I(t, x_i, y_i) \rangle \quad (1)$$

106

107 where  $DIP(x_i, y_i)$  is the differential image intensity at the  $i$ th pixel between the temporal peak  
108 signal  $I(t, x_i, y_i)$  and the temporal average intensity  $\langle I(t, x_i, y_i) \rangle$ . For a given bubble  
109 occasionally passing an otherwise dark image pixel, the peak intensity was expected to be much  
110 higher than the average intensity. On the other hand, the peak intensity and the average intensity  
111 were expected to be similar for tissue signal. For noise both the peak and average intensity are  
112 expected to be relatively low. As a result, the differential intensity of pixels containing  
113 microbubble signals is expected to be higher than that of tissue or noise.

114

115 *Threshold selection*

116 A threshold in differential intensity was required to separate microbubble signals from tissue  
117 and noise. It was estimated from the histogram of differential intensity projection, an example  
118 of which is shown in Figure 1. It should be noted that the threshold is automatically adjusted  
119 for each patient based on the entire image. The intensity histogram of differential intensity  
120 projection is constructed (see Figure 2). The threshold is determined at the intersection point  
121 of microbubble and tissue distributions.

122

123

124 *Microvascular area and density*

125 The ROI in plaque was selected manually. The number of pixels identified as containing bubble  
126 signal was defined as the microvascular area (MVA), which can then be normalized by the  
127 total number of pixels within the plaque ROI to obtain the microvascular density (MVD)  
128 measure for the ROI.

$$MVD = \frac{MVA}{\text{area of ROI}} \quad (2)$$

129

130 *In-vitro study*

131 The DIP algorithm was validated on a simple laboratory phantom constructed in-house and  
132 shown in Figure 1A. It consisted of a piece of tissue-mimicking material, above which a highly  
133 diluted microbubble suspension was gently stirred to simulate individual bubbles moving  
134 within the phantom.

135

136 *Clinical application (plaque)*

137 Forty-eight patients previously treated for head and neck cancer (HNC) with at least one risk  
138 factor for atherosclerosis were recruited from a cancer centre. These patients are asymptomatic  
139 for cardiovascular events. From this group, 24 videos with carotid plaque were selected for this  
140 study. The study was approved by the institutional research and ethics committee and each  
141 patient provided informed consent. CEUS image sequences were acquired on both sides of the  
142 neck with a clinical scanner (GE Vivid7 with a 9 MHz broadband linear array transducer). The  
143 GE scanner was used to scan the subject with the following settings: MI = 0.21, Gain = 0, DR

144 = 54, TGC = manually adjusted, Frequency = 3.2/6.4 MHz. The contrast mode is used to  
145 perform contrast enhanced imaging. Contrast-enhanced ultrasound video loops were taken  
146 using a commercially available ultrasound contrast agent, SonoVue™ (Bracco, Milan) given  
147 as an intravenous infusion via a peripheral vein at a rate of 1.2 mL/min. The infusion was  
148 delivered over a total of 5-7 minutes. Imaging was performed in real-time prior to the arrival  
149 of and following the saturation of the carotid artery with SonoVue.

150

151

### 152 *Visual assessment*

153 IPN was graded semi-quantitatively as absent (Grade 0), limited to the adventitia/plaque base  
154 (Grade 1) or extensive and/or extending into the plaque body (Grade 2) by a clinician (Dr.  
155 Shah).

156

### 157 *Motion compensation and DIP*

158 The motion of carotid artery was tracked and corrected by a dedicated motion correction  
159 algorithm (Stanziola et al. 2015). The algorithm consisted of three steps: (A) Pre-processing,  
160 (B) Lumen segmentation and (C) Registration. In the first step, large rigid motions were  
161 removed by a rigid registration. Then, the algorithm used the information of the cardiac cycle  
162 and the Gabor filter responses of the corresponding frames to obtain a mixture of frames where  
163 the fragmentation of the lumen signal was largely removed. In the second step, the lumen was  
164 segmented by using thresholding and level set methods. A binary mask of the lumen region  
165 was obtained for each frame. Finally, a non-rigid registration was performed to correct the  
166 motion effect on each frame based on minimising the energy functional of non-lumen region

167 of two consecutive images and the energy functional of segmented lumen region of two  
168 consecutive binary masks.

169 Then DIP images were calculated for each CEUS image sequence using Eqn (1). Maximum  
170 Intensity Projection (MIP) images were also obtained for comparison purpose.

171

### 172 *Regions of interest (ROI) analysis*

173 Analysis of CEUS video sequences was performed off-line using software developed in-house  
174 using MATLAB (The MathWorks, Natick, MA, USA). Carotid plaques were segmented  
175 manually as the regions of interest (ROIs) by a clinical expert (Dr Chahal) using both CEUS  
176 sequence and maximum intensity projection (MIP) (Figure 2, first and second columns). Both  
177 MVA and MVD were calculated for each plaque, and results compared with visual grading.

178

### 179 *Statistical analysis*

180 The sample size was small and not normally distributed. Therefore, non-parametric statistical  
181 analyses were used in this study. The correlation between the visual grade and the MVA/MVD  
182 derived from our method was tested by Spearman rank correlation. The differences between  
183 the mean rank of MVA/MVD and the visual grade groups were tested by Kruskal-Wallis test  
184 with alpha set at 0.05. Statistical analyses were performed using SPSS (IBM Corp. Released  
185 2013. IBM SPSS Statistics for Windows, Version 22.0. Armonk, New York, USA). Further, a  
186 comparison between MIP and DIP using patient data was performed

187

## 188 **Results**

189 *In-vitro study*

190 The CEUS image of the phantom, MIP and DIP images were shown in Figure 1. It can be seen  
191 that while the microbubble detection is similar between the MIP and DIP, the tissue linear  
192 artefact at the lower part of the image in the CEUS image and the MIP was completely removed  
193 in DIP image.

194

195 *Differential Intensity Histogram and Threshold selection*

196 By examining the histogram of differential intensity projection (0.25) (Figure 2, dotted line), it  
197 can be seen that there are peaks in the histogram corresponding to microbubble, tissue and  
198 noise.

199

200 *Visual Assessment and Differential intensity projection*

201 Among the 24 video sequences, Grade 0 IPN was seen in 12/24 videos, while Grade 1 IPN in  
202 8/24 videos and Grade 2 IPN in 4/24 videos. Examples of CEUS images and the MIP images  
203 with each visual grade are shown in Figure 3. The corresponding DIP images are shown in  
204 Figure 2 (third column).

205

206 By examining MIP and DIP images of the same dataset against visually confirmed bubble  
207 signals by clinician experts (solid arrows in the images), DIP showed not only much less  
208 nonlinear tissue artefact and better image contrast, but also clearly revealed IPN signals which  
209 were buried by tissue artefacts in the corresponding MIP image (Figure 2B).

210

211 *Clinical evaluation (MVA and MVD)*

212 The average and the median of MVA for each visual grade are shown in Table 1. It can be  
213 observed that both mean and median of MVA increased with the visual grade. Furthermore,  
214 Table 2 shows the average and the median of MVD for each visual grade. It can be seen that  
215 both mean and median of MVD increased with the visual grade.

216 The box plots of visual grade vs MVA or MVD were displayed in Figure 4.

217

218 *Spearman's rank correlation coefficient*

219 Both MVA and MVD were significantly correlated with the visual grade ( $R = 0.83$  and  $0.77$   
220 respectively, ( $p < 0.001$  for both) for DIP. This is a significant improvement over those  
221 obtained by MIP (MVA:  $R = 0.26$ , MVD:  $R = -0.23$ ).

222

223

224 *Kruskal-Wallis Test*

225 Table 3 shows the visual grade groups comparison. For the group grade 0 vs grade 1, there was  
226 a significant difference in MVA or MVD at  $\alpha = 0.05$ . Similarly, there was a significant  
227 difference in MVA or MVD at  $\alpha = 0.05$  for the group grade 0 vs grade 2. However, for the  
228 group grade 1 vs grade 2, the difference in MVA or MVD was not significant at  $\alpha = 0.05$ , while  
229 the difference in MVD was significant at  $\alpha = 0.1$ .

230

231 **Discussion**

232 In this study a new image processing approach, differential intensity projection (DIP), was  
233 developed to visualise and quantify plaque IPN in CEUS image sequences of carotid arteries  
234 in vivo. Compared with existing method MIP, the proposed DIP can significantly reduce  
235 nonlinear propagation tissue artefacts and improve imaging specificity, as validated in the in  
236 vitro study where ground truth is available. Two quantitative measures, MVA which is related  
237 to the total vascular areas occupied by IPN in the plaque, and MVD which is a vascular density  
238 measure, were generated based on each DIP image. The in vivo data on human carotid artery  
239 analysed by DIP showed a strong and much higher correlation between MVA/MVD and visual  
240 IPN grade than that by MIP. There was also a significant difference in MVA/MVD between  
241 patient groups (i.e. grade 0 vs grade 1 or grade 0 vs grade 2).

242

243 Quantification of IPN as a novel surrogate marker for stroke risk can be highly valuable in  
244 clinical diagnosis. Recently, several groups have developed various methods for IPN  
245 quantification. Huang et al. (Huang et al. 2008) proposed a dynamic evaluation of the plaque  
246 enhancement by a time intensity curve analysis (TIC). TIC is commonly used in analysing  
247 large and well perfused organs, for example, the liver, prostate and heart. However, plaques in  
248 the carotid artery are often small and weakly perfused. Therefore, TIC analysis may not be  
249 appropriate to quantify microvessels in plaques. Hoogi et al. (Hoogi et al. 2012) adopted  
250 electrocardiogram (ECG) gating to correct for motion and only one CEUS image per cardiac  
251 cycle was used. Hence, the connection of microvessel paths after time integration may be lost.  
252 More importantly, these algorithms can be significantly affected by the nonlinear propagation  
253 tissue artefact. The DIP method has a unique advantage of being able to efficiently reduce such  
254 tissue artefacts.

255

256 One challenge of quantifying neovascularisation in plaque is tissue motion. It is caused by the  
257 expansion and contraction of blood vessels, breathing and swallowing. Our dedicated motion  
258 compensation algorithm (Stanziola et al. 2015) was applied to improve the quantification of  
259 IPN. The software performs better than other current available methods. It should be noted that  
260 even if motion compensation is applied, some out-of-plane motion could still affect the  
261 quantification. Any non-corrected motion will potentially introduce artefacts into DIP images.  
262 Further studies to take into account of out-of-plane motion could further improve the  
263 quantification results.

264

265 Besides the nonlinear propagation tissue artefact and motion compensation, attenuation is also  
266 an important consideration that may affect quantification. Whilst it appeared in our study that  
267 quantification was not significantly affected by attenuation, it may not always be the case.  
268 Recently, Cheung et al (Cheung et al. 2015) have developed an automated attenuation  
269 correction and normalisation algorithm to improve the quantification of contrast enhancement  
270 in ultrasound images of carotid arteries. The algorithm firstly corrects for attenuation artefact  
271 and normalises intensity within the contrast agent-filled lumen and then extends the correction  
272 and normalisation to regions beyond the lumen.

273

274 The proposed method can generate more specific visualisation of vessels and more reliable IPN  
275 quantification. It could have important implications for clinical screening, diagnosis and  
276 management of this important disease. Specifically, such quantitative information on plaque  
277 vascularisation enables improved patient risk stratification and potentially improves drug  
278 treatment by providing a tool for monitoring treatment.

279

280 The DIP is simple and computationally efficient and can be implemented in real time, as it only  
281 involves simple mathematical operations. The quantification process is semi-automated, only  
282 requiring manual input for segmenting the plaques. Fully automated segmentation is possible  
283 but requires further studies.

284

285 It should be noted that there is some overlap in MVD between grade 1 and grade 2 plaques and  
286 the difference was not statistically significant. This is likely due to the the small sample size of  
287 the analysis (n=4 for grade 2). More patient data in future studies would help demonstrate any  
288 significance in quantification results between the two groups using our method.

289

290 In our clinical data only two out of the twenty four plaques are located in the near wall, while  
291 there are 22 plaques found in the far wall. Due to the low number of the near wall plaques it is  
292 not possible to draw any conclusion on how our method performs on plaques located at the  
293 different sides of the wall. However, it should be noted that the correlation of the far wall  
294 quantification by DIP improved significantly over MIP.

295

296

## 297 **Conclusions**

298 DIP is demonstrated to be a specific, simple and fast technique for visualisation and  
299 quantification of small vessels in CEUS images and has potential for clinical assessment of  
300 intraplaque neovascularisation.

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372 Figure 1: (A) The CEUS image of tissue mimicking phantom (B) Maximum intensity  
373 projection (C) Differential intensity projection.

374

375 Figure 2: Histogram of differential intensity projection

376

377 Figure 3: First column: CEUS image with ROI. Second column: Maximum intensity  
378 projection. Third column: Differential intensity projection. (A) plaque with grade 0 (B) plaque

379 with grade 1 (C) plaque with grade 2, tissue artefact is indicated by a dashed arrow and bubble  
380 signal is indicated by a solid arrow.

381

382 Figure 4: Box plot of visual grade versus (A) MVA (B) MVD, outlier is indicated by a circle  
383 with number.

384

385 [Video: A CEUS video sequence of a carotid artery with IPN \(Grade 2\), where microbubbles are seen](#)  
386 [passing through the plaque \(red arrows\).](#)

387

388 Table 1: The average and median of MVA for each visual grade

<i>Visual Grade</i>	<i>Mean <math>\pm</math> SD</i>	<i>Median</i>
Grade 0	1.42 $\pm$ 2.90	0
Grade 1	95.67 $\pm$ 105.58	48
Grade 2	538.50 $\pm$ 701.27	228.5

389

390 Table 2: The average and median of MVD for each visual grade

<i>Visual Grade</i>	<i>Mean <math>\pm</math> SD</i>	<i>Median</i>
Grade 0	0.08 $\pm$ 0.47 (%)	0%
Grade 1	1.21 $\pm$ 1.40 (%)	0.40%
Grade 2	8.26 $\pm$ 12.88 (%)	2.18%

391

392 Table 3: Visual grade groups comparison by Kruskal-Wallis test

<i>Visual Grade Groups Comparison</i>	<i>MVA</i>	<i>MVD</i>
Grade 0 vs Grade 1	$p = 0.001^*$	$p = 0.006^*$
Grade 1 vs Grade 2	$p = 0.126$	$p = 0.062$
Grade 0 vs Grade 2	$p = 0.001^*$	$p = 0.001^*$
* significant at $\alpha = 0.05$		

393