

1 **Cerebrospinal fluid biomarkers of neurodegeneration, synaptic integrity**  
2 **and astroglial activation across the clinical Alzheimer's disease spectrum**

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1 **ABSTRACT**

2 **INTRODUCTION:** We investigated relations between amyloid- $\beta$  ( $A\beta$ ) status, *APOE*- $\epsilon 4$  and  
3 cognition, with cerebrospinal fluid (CSF) markers of Neurogranin (Ng), Neurofilament-light,  
4 (NFL), YKL-40 and Total tau (T-tau).

5  
6 **METHODS:** We included 770 individuals with normal cognition, MCI and AD-type-dementia  
7 from the EMIF-AD Multimodal Biomarker Discovery study. We tested the association of Ng,  
8 NFL, YKL-40 and T-tau with  $A\beta$  status ( $A\beta^-$  vs.  $A\beta^+$ ), clinical diagnosis *APOE*  $\epsilon 4$  carriership,  
9 baseline cognition and change in cognition.

10

11 **RESULTS:** Ng and T-tau distinguished between  $A\beta^+$  from  $A\beta^-$  individuals in each clinical  
12 group, while NFL and YKL-40 were associated with  $A\beta^+$  in non-demented individuals only.  
13 *APOE*  $\epsilon 4$  carriership did not influence NFL, Ng and YKL-40 in  $A\beta^+$  individuals. NFL was the  
14 best predictor of cognitive decline in  $A\beta^+$  individuals across the cognitive spectrum.

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16 **DISCUSSION:** Axonal degeneration, synaptic dysfunction, astroglial activation and altered  
17 tau metabolism are involved already in preclinical AD. NFL may be a useful prognostic  
18 marker.

19

20 **KEYWORDS**

21 Alzheimer's disease; amyloid-beta; neurofilament light; neurogranin; YKL-40; cognition;  
22 cerebrospinal fluid; *APOE*

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## 1 **1. Background**

2 Biomarkers have become increasingly important for the diagnosis of Alzheimer's disease (AD)  
3 [1, 2], and are contributing to an improved understanding of the temporal pattern of AD  
4 pathophysiology. It has been shown that amyloid-beta ( $A\beta$ ) deposition is one of the earliest  
5 detectable events in AD pathogenesis [3, 4], and that genetic risk for AD can be assessed by  
6 determining apolipoprotein E (*APOE*)  $\epsilon 4$  genotype. However, other pathophysiological  
7 mechanisms underlying AD and their relation to inter-individual variation in cognitive  
8 trajectories, are less well understood. By relating  $A\beta$ , *APOE* genotype and cognition to  
9 cerebrospinal fluid (CSF) biomarkers for AD-related processes including axonal degeneration,  
10 synaptic dysfunction and astroglial activation in individuals across the clinical AD spectrum,  
11 we will likely learn more about the temporal ordering of these pathological mechanisms. This  
12 may translate into improved diagnostic and prognostic algorithms, which, in turn, should help  
13 to develop and evaluate more targeted disease-modifying treatments.

14

15 Besides  $A\beta$ , a number of proteins in CSF have been found to be associated with AD. Both  
16 phosphorylated (P-tau) and total tau (T-tau) are well-established biomarkers for AD and  
17 cognitive decline [5, 6]. High concentrations of neurofilament-light (NFL) have been  
18 associated with axonal degeneration to, predominantly, subcortical brain areas [7, 8] and YKL-  
19 40 (also known as chitinase 3-like protein 1) concentrations were found to reflect astrocytic  
20 activation, an inflammatory response to neurodegenerative processes [9]. Neurogranin (Ng)  
21 has been identified as a candidate AD marker reflecting synaptic degeneration and cognitive  
22 decline in the early stages of AD [10, 11]. While NFL, YKL-40 and Ng have evolved over the  
23 last years as promising AD biomarkers and have been strongly associated with neuronal injury  
24 markers [11-13], data regarding their relation to  $A\beta$ , *APOE* and cognition have been  
25 inconsistent or inconclusive [10, 12, 14-16].

1

2 Hence, to unravel how NFL, Ng and YKL-40 relate to AD pathology, genetic risk and disease  
3 severity, we aimed to investigate their relationships with A $\beta$ , *APOE*  $\epsilon$ 4 carriership and  
4 cognition, in a large cohort consisting of individuals across the AD spectrum. To compare the  
5 relations regarding NFL, Ng and YKL-40 to those of an established neurodegenerative AD  
6 marker, we also examined the associations of T-tau with A $\beta$ , *APOE* genotype and cognition.

7

## 8 **2. Methods**

### 9 *2.1 Subjects*

10 We selected 770 individuals from the EMIF-AD Multimodal Biomarker Discovery (EMIF-AD  
11 MBD) study; a cross-cohort study consisting of collated data and samples from 11 European  
12 cohorts [17]. The EMIF-AD MBD includes a total of 1221 individuals across the cognitive  
13 spectrum: normal cognition (NC), Mild Cognitive Impairment (MCI) and AD-type dementia.  
14 Individuals were selected from prospective cohort studies based on the availability of plasma,  
15 DNA and CSF samples and MRI scans. Exclusion criteria for the EMIF-AD MBD study were  
16 the presence of neurological, psychiatric or somatic disorders that could cause cognitive  
17 impairment [17]. Written informed consent was obtained from all participants before inclusion  
18 in the study. The medical ethics committee at each site approved the study (Supplemental Table  
19 1).

20

21 For the current study we selected all participants from whom CSF samples were available for  
22 central analyses (n=770). Participants were included from three multicenter studies:  
23 DESCRIPA (n=29) [18], EDAR (n=197) [19] and IMI PharmaCog (n=146) [20], and four  
24 single center studies: Amsterdam (n=170) [21], Antwerp (n=148) [22], San Sebastian GAP  
25 (n=40) [23] and Lausanne (n=40) [24].

## 1    2.2 *Clinical diagnosis and assessment*

2    Normal cognition (NC) was defined as normal performance on neuropsychological assessment  
3    (within 1.5 SD of the average for age, gender and education). MCI was defined as having  
4    performance below 1.5 SD of the average on at least one neuropsychological test [25]. AD-  
5    type dementia was defined based on a clinical diagnosis, using the National Institute of  
6    Neurological and Communicative Disorders and Stroke – Alzheimer’s Disease and Related  
7    Disorders Association (NINCDS-ADRDA) criteria [26].

8

9    The clinical assessment is described in a previous publication [17]. In short, clinical data were  
10    collected using local routine protocol at each site and thereafter harmonized and stored onto  
11    the EMIF-AD online data platform for pooled analyses. We used the Mini Mental State  
12    Examination (MMSE) [27] as our main cognitive outcome measure, which was available in  
13    99% of the subjects at baseline and in 68% at follow-up. In general, baseline clinical assessment  
14    and CSF collection were conducted within a one year window. For a subgroup, the length of  
15    this time window was unknown (n=21) or longer than one year (n=2).

16

## 17    2.3 *CSF analyses*

18    Central CSF analyses were conducted at Gothenburg University, Sweden. NFL concentrations  
19    were measured using a commercial ELISA (NF-light® ELISA, Uman Diagnostics, Umeå,  
20    Sweden; [7]). Ng was measured using an in-house immunoassay for Ng [10]. YKL-40 was  
21    determined by a human chitinase-3 quantikine ELISA kit (R&D systems, Inc, Minneapolis,  
22    MN; [28]). A $\beta$ <sub>38</sub>, A $\beta$ <sub>40</sub>, and A $\beta$ <sub>42</sub> were measured using the V-PLEX Plus A $\beta$  Peptide Panel 1  
23    (6E10) Kit from Meso Scale Discovery (MSD, Rockville, MD). All analyses were performed  
24    according to the manufacturer’s instructions by board-certified laboratory technicians who  
25    were blinded to clinical information. All measurement were performed on one occasion using

1 one batch of reagents, except for n=8 samples from the EDAR cohort that were analysed  
2 beforehand in the same laboratory, but in a different batch. For phosphorylated tau (P-tau) and  
3 total tau (T-tau), we used available measures from the local cohorts (P-tau n=630; T-tau n=621)  
4 derived in clinical laboratory practice using INNOTEST ELISAs (Fujirebio, Ghent, Belgium).

5

#### 6 *2.4 Genetic analyses*

7 For the entire EMIF-AD BMD cohort *APOE* genotyping data from the local genetic analyses  
8 was available for n=1121 (91%) individuals. For central analyses, 805 DNA and 148 whole  
9 blood samples were transferred to Lübeck University, Germany. From the blood samples, DNA  
10 was extracted using QIAamp® DNA Blood Mini Kit (QIAGEN GmbH, Hilden, Germany)  
11 resulting in 953 DNA samples, of which 926 passed quality control. All samples were subjected  
12 to genome-wide SNP genotyping using the Infinium Global Screening Array (GSA) with  
13 Shared Custom Content (Illumina Inc.). From these genome-wide data, *APOE* genotypes were  
14 determined either directly (rs7412) or by imputation (rs429358) in all 926 samples. For 80  
15 samples for which no local *APOE* genotype was available, and for 45 mismatches between  
16 locally and GSA derived genotypes (4.8%), *APOE* genotype was determined using TaqMan  
17 assays (ThermoFisher Scientific, Foster City, CA) on a QuantStudio-12K-Flex system in 384-  
18 well format. We classified individuals as *APOE*  $\epsilon 4$  carriers ( $\epsilon 4+$ ) or non-carriers ( $\epsilon 4-$ )  
19 according to their genotype status at rs429358 (C-allele =  $\epsilon 4$ ).

20

#### 21 *2.5 Biomarker classifications*

22  $A\beta$  status was defined by the CSF  $A\beta_{42/40}$  ratio, using a cut-off of  $<0.063$  to determine  
23 abnormality. This cut-off was defined using mixture model analyses in the current dataset [29,  
24 30], showing a clear binomial distribution (Supplemental Figure 1). Abnormality based on this  
25 cut-off showed a high concordance rate with abnormality based on the local  $A\beta_{42}$  measures



1 and cut-offs (82%). For the analyses regarding the influence of NFL, Ng and YKL-40 on  
2 cognition, a median-split was used to divide the sample (Cut-off values: NFL: 869 pg/ml; Ng:  
3 103 pg/ml; YKL-40: 163 ng/ml) as there are no well-established cut-offs or approaches yet to  
4 define abnormality and the use of tertiles or quartiles to divide the data would limit statistical  
5 power.”. Dichotomous T-tau values (normal vs. abnormal) was available in n=762 individuals  
6 and was determined using local cut-off points (Supplemental Table 2).

7

## 8 *2.6 Statistical analyses*

9 Baseline characteristics were compared by A $\beta$  status and diagnostic group using Chi-square  
10 for categorical variables and general linear mixed (GLM) models with study as a random effect  
11 for continuous variables. We also tested whether the influence of A $\beta$  on NFL, Ng and YKL-  
12 40 was different across diagnostic groups and age, by examining the diagnostic group by A $\beta$ ,  
13 and age by A $\beta$  interactions. Prior to the comparisons, A $\beta$ <sub>42</sub>, NFL, Ng, YKL-40, P-tau and T-  
14 tau values were log-transformed to approximate a normal distribution. Spearman’s correlations  
15 were used to assess the correlations between biomarker values. GLM models with random  
16 intercepts and slopes by study were used to examine the influence of A $\beta$  status and low/high  
17 or normal/abnormal biomarker levels on MMSE performance and decline over time, adjusted  
18 for age, gender, years of education and baseline diagnosis. Lastly, we tested the independent  
19 influence of all markers on cognitive decline by adding all dichotomous markers (high/low or  
20 normal/abnormal) in one GLM model with MMSE scores over time as outcome measure,  
21 stratified by A $\beta$  status. Missing values for *APOE*  $\epsilon$ 4 status (n=12) and years of education  
22 (n=105) were imputed using regression analyses within study, based on significant predictors  
23 (i.e. age, gender, MMSE, cognitive scores) for these variables. All analyses were repeated after  
24 exclusion of individuals with a long or unknown interval between clinical assessment and CSF  
25 collection (n=23). Statistical analyses were performed using R Statistical Software (version

1 3.3.3) and SPSS (version 24). We used two-sided  $p < 0.05$  to define statistical significance. Due  
2 to the exploratory nature of the study we did not adjust for multiple comparisons.

3

### 4 **3. Results**

5 We assessed 770 individuals who were on average 69.3 (SD 8.3) years old and had an average  
6 of 10.9 (SD 3.9) years of education. Three hundred ninety-nine (52%) were female. Clinical  
7 follow-up data was available for 557 (73%) individuals, with an average follow-up length of  
8 2.3 (SD 1.3) years. At baseline 140 (18%) individuals were considered cognitively normal  
9 (CN), 450 (58%) were diagnosed as having mild cognitive impairment (MCI) and 180 (23%)  
10 were clinically diagnosed as having AD-type dementia. Despite a clinical diagnosis of AD-  
11 type dementia, 23 (13%) individuals did not show evidence of amyloid pathology.

12

#### 13 *3.1 Demographics and biomarker values*

14 Table 1 shows the baseline characteristics and biomarker values per diagnostic group, stratified  
15 by  $A\beta$  status. As expected, in the whole sample,  $A\beta+$  individuals were older, more frequently  
16 *APOE-ε4* carrier and had lower MMSE scores compared to  $A\beta-$  individuals. When stratified  
17 by baseline diagnosis, we found that  $A\beta+$  individuals were older compared to the  $A\beta-$   
18 individuals in the CN and MCI groups, but not in the AD-type dementia group. Only in MCI  
19 we found a difference in MMSE score between groups by  $A\beta$  status. Other comparisons are  
20 shown in Table 1.

21

#### 22 *3.2 NFL, Ng, YKL-40 and T-tau by $A\beta$ status and baseline diagnosis*

23 Comparisons by  $A\beta$  status and baseline diagnoses of NFL, Ng, YKL-40 and T-tau  
24 concentrations are shown in Table 1. Figure 1 shows the comparisons by  $A\beta$  status within the  
25 diagnostic groups. When comparing by  $A\beta$  status, NFL and YKL-40 values were differentially

1 increased in A $\beta$ + CN and MCI individuals, while in the dementia stage NFL and YKL-40  
2 levels were elevated regardless of A $\beta$  status. T-tau and Ng values were stably increased in A $\beta$ +  
3 individuals across the cognitive spectrum. For NFL we found that the influence of A $\beta$  on NFL  
4 was different across diagnoses (interaction A $\beta$ \*diagnosis p=0.027). NFL concentrations  
5 increased in A $\beta$ - individuals with advancing clinical stage, while they were stable in the A $\beta$ +  
6 CN and MCI groups but increased further in the A $\beta$ + AD-type dementia group (Figure 1). The  
7 influence of A $\beta$  on YKL-40 levels was similar as for NFL (interaction A $\beta$ \*diagnosis p=0.001).  
8 For Ng and T-tau we found that influence of A $\beta$  was similar across diagnoses (interaction  
9 A $\beta$ \*diagnosis T-tau: p=0.771;Ng: p=0.580). A $\beta$ + did have a stronger effect on Ng and T-tau  
10 concentrations in younger individuals than in older individuals (interaction A $\beta$ \*age Ng:  
11 p=0.006; T-tau: p<0.001), while there was no age effect for NFL and YKL-40 (data not shown).

12

### 13 3.3 APOE $\epsilon$ 4 carriership

14 In A $\beta$ + individuals, no effect was found of APOE  $\epsilon$ 4 carriership on NFL, Ng and YKL-40  
15 levels, regardless of clinical diagnosis (Table 2). In A $\beta$ - individuals, APOE  $\epsilon$ 4 carriership was  
16 associated with lower levels of NFL in the total group and in individuals with MCI, as well as  
17 with lower Ng levels in the MCI and AD-type dementia groups, but with higher Ng levels in  
18 the total group (Table 2). We found no influence of APOE  $\epsilon$ 4 carriership on YKL-40 and T-  
19 tau levels when comparing within A $\beta$  status, stratified by diagnosis. However, compared to the  
20 CN A $\beta$ - APOE  $\epsilon$ 4 non-carriers, T-tau and YKL-40 levels were elevated in A $\beta$ + individuals  
21 regardless of clinical diagnosis (Table 2).

22

### 23 3.4 Correlations

1 The A $\beta$  isoforms were highly positively correlated and a more abnormal A $\beta_{42/40}$  ratio was  
2 correlated with higher NFL, Ng and YKL-40 levels. P-tau and t-tau were highly correlated,  
3 and were both associated with all three emerging biomarkers (Supplemental Figure 2).

4

### 5 *3.5 Baseline cognition and change in cognition over time*

6 Cross-sectional analyses showed that in A $\beta$ <sup>+</sup> individuals, high NFL, Ng and T-tau levels were  
7 associated with lower MMSE scores in the total group (Table 3, Figure 2). When stratifying by  
8 diagnostic group within the A $\beta$ <sup>+</sup> individuals, high NFL levels were associated with low MMSE  
9 scores in the MCI and AD-type dementia groups, and high T-tau levels with low MMSE scores  
10 in the MCI group (Table 3). In A $\beta$ <sup>-</sup> individuals, high NFL levels were associated with lower  
11 MMSE scores in the total group, and high T-tau levels with lower scores in the AD-type  
12 dementia group. In addition, high Ng levels were associated with higher MMSE scores in the  
13 AD-type dementia group in A $\beta$ <sup>-</sup> individuals.

14

15 Longitudinal analyses showed that in A $\beta$ <sup>+</sup> individuals, high baseline levels of NFL and T-tau  
16 were associated with an increased rate of cognitive decline in the total sample. High baseline  
17 levels of NFL and Ng were also associated with increased rate of decline in the AD-type  
18 dementia group. In A $\beta$ <sup>-</sup> individuals, high baseline levels of NFL, YKL-40 and T-tau were  
19 associated with an increased rate of cognitive decline in the total group, as well as in the MCI  
20 and AD-type dementia groups (Table 3). In A $\beta$ <sup>-</sup> individuals, high Ng levels were associated  
21 with a decreased rate of decline in the MCI group, but with an increased rate of decline in the  
22 AD-type dementia group (Table 3).

23

24 Next, we combined NFL, YKL-40, Ng, and T-tau in the longitudinal analyses and stratified by  
25 baseline diagnosis (Table 4). In CN A $\beta$ <sup>+</sup> individuals, only high baseline NFL levels predicted

1 decline. In A $\beta$ + individuals with MCI, increased baseline NFL and T-tau and decreased Ng  
2 levels independently predicted cognitive decline. In A $\beta$ + individuals with AD-type dementia,  
3 increased baseline NFL and Ng levels predicted decline. Among A $\beta$ - individuals, increased  
4 baseline NFL and tau levels predicted decline only in individuals with MCI (Table 4).

5

6 When repeating all analyses without the individuals for whom the interval between CSF  
7 collection and cognition was longer than one year or unknown (n=23), results remained similar.

8 Exclusion of an individual with very high Ng concentrations also yielded similar results. In

9 addition, outcomes were also similar when using P-tau instead of T-tau in the analyses

10 regarding APOE  $\epsilon$ 4 carriership and cognition.

11

#### 12 **4. Discussion**

13 We investigated the relations between A $\beta$  status, *APOE*  $\epsilon$ 4 carriership and cognition, with CSF

14 concentrations of NFL, Ng, YKL-40 and T-tau, in a large cohort of individuals across the

15 clinical AD spectrum. The main findings were: (1) CSF NFL, Ng, YKL-40 and T-tau levels

16 were associated with A $\beta$  already in the preclinical stage; (2) A $\beta$ - *APOE*  $\epsilon$ 4 carriers with MCI

17 or AD-type dementia had lower concentrations of NFL and Ng compared to non-carriers; (3)

18 High baseline NFL levels predicted cognitive decline in A $\beta$ + individuals with normal

19 cognition, MCI and AD-type dementia, independent of the other markers.

20

21 NFL, Ng, YKL-40 and T-tau concentrations were all associated with A $\beta$ +. In A $\beta$ + individuals,

22 NFL levels were higher in the dementia stage compared to the MCI stage, whereas Ng and

23 YKL-40 levels stayed relatively stable over time. Yet in A $\beta$ - individuals, we found an increase

24 of both NFL and YKL-40 levels in MCI individuals compared to CN individuals, while Ng

25 levels in A $\beta$ - individuals remained low with increasing disease severity. T-tau levels increased

1 with disease severity regardless of A $\beta$  status, albeit the rate of increase was faster in A $\beta$ +  
2 individuals. These findings confirm that synaptic dysfunction – as measured by Ng – plays an  
3 important role in AD pathophysiology in all clinical stages [31, 32]. In addition, our data  
4 verifies that axonal degeneration and neuroinflammation - as respectively measured by NFL  
5 and YKL-40 – are less specific to AD [9, 33], but their temporal pattern across the clinical  
6 stages is AD specific: in AD, NFL and YKL-40 levels are already increased in the preclinical  
7 stage, while in A $\beta$ - individuals concentrations merely start to increase from the MCI stage  
8 onwards. Our findings regarding T-tau levels, confirm the association of altered neuronal tau  
9 metabolism with A $\beta$  pathology [6, 34], and support the notion this process also occurs in A $\beta$ -  
10 individuals, although to a lesser extent [35]. Together these results provide novel insights into  
11 the temporal pattern of AD pathophysiology, which should be validated by longitudinal  
12 biomarker studies.

13

14 The APOE genotype did not influence NFL, Ng, YKL-40 and T-tau levels in A $\beta$ + individuals  
15 in all clinical stages, suggesting that these markers reflect a generic reaction to amyloid  
16 aggregation regardless of APOE genotype. In A $\beta$ - individuals, APOE  $\epsilon$ 4 carriers with MCI or  
17 AD-type dementia had lower NFL and Ng levels compared to non-carriers. This suggests that  
18 the A $\beta$ - APOE  $\epsilon$ 4 non-carriers with MCI or AD-type dementia might have other pathologies  
19 not related to A $\beta$  and APOE  $\epsilon$ 4 carriership that are causing cognitive impairment, axonal  
20 degeneration, and to a lesser extent also synaptic dysfunction. Regarding T-tau and YKL-40  
21 levels, we found similar concentrations in APOE  $\epsilon$ 4 carriers and non-carriers, which is in line  
22 with previous studies [36-38], but in contrast with a previous study in which a modest  
23 association of APOE  $\epsilon$ 4 carriership on YKL-40 levels was found in individuals with MCI due  
24 to AD [39]. Besides the inconsistency with the latter study, possibly due to heterogeneity in

1 sample sizes or biomarker classifications, our results confirm that YKL-40 concentrations are  
2 independent of *APOE*  $\epsilon 4$  carriership.  
3  
4 Higher levels of NFL and T-tau were associated with a lower cognitive performance and an  
5 increased rate of decline regardless of  $A\beta$  status. As both NFL and T-tau are markers of axonal  
6 degeneration [5, 12], these findings imply that axonal loss may be an important driver of  
7 cognitive decline in both  $A\beta+$  and  $A\beta-$  individuals [33, 40]. Concerning Ng, we found that  
8 only in the dementia stage, higher concentrations were associated with a faster rate of decline,  
9 regardless of  $A\beta$ . This is congruent with previous CSF biomarker studies suggesting that Ng  
10 might be strongly associated with cognition, irrespective of amyloid plaque pathology [40-42].  
11 However, Ng changes have also been associated with cognitive decline in preclinical AD [11],  
12 a finding we could not confirm with our analyses possibly due to a lower sensitivity of the  
13 cognitive outcome measure we used (i.e. MMSE) or because we used a median-split instead of  
14 tertiles to define low and high Ng levels. Posthoc, we explored the influence of the cognitive  
15 outcome measure by repeating the analyses in a subgroup (n=615) with a pooled standardized  
16 memory score [17]. These posthoc analyses showed that high Ng levels tended to be associated  
17 with a faster decline in memory performance in CN  $A\beta+$ , but not in CN  $A\beta-$  individuals (data  
18 not shown). The negative impact of high YKL-40 levels on cognition seems to only relate to  
19  $A\beta-$  individuals or the influence is masked by  $A\beta$  pathology in  $A\beta+$  individuals. These findings  
20 suggest that YKL-40 may be a prognostic marker for individuals with MCI but without  
21 evidence of  $A\beta$  pathology, for instance those with Suspected Non-Alzheimer's Disease  
22 Pathophysiology (SNAP) [43]. When all markers were combined in one model we found that  
23 NFL, and from the MCI stage onwards also T-tau, were independent predictors of cognitive  
24 decline in  $A\beta+$  individuals. Remarkably high Ng levels were associated with a slower rate of  
25 decline in  $A\beta+$  individuals with MCI and a faster rate of decline in  $A\beta+$  individuals with AD-

1 type dementia. Although a similar finding was described in a previous study [42], it remains  
2 uncertain what the underlying mechanism is. Possibly, Ng is not a direct contributor to  
3 cognitive decline in the pre-dementia stages or the relation between Ng and cognition is again  
4 dependent on the cognitive outcome measure used (global cognition vs. memory).

5

6 This study has several limitations. First, data was collected at different centers using routine  
7 local protocols. However, the CSF samples were analyzed centrally for most outcome measures  
8 – A $\beta$ <sub>38</sub>, A $\beta$ <sub>40</sub>, A $\beta$ <sub>42</sub>, NFL, Ng and YKL-40 - and clinical data was harmonized using validated  
9 methods like standardization and dichotomization. Second, our AD-type dementia group  
10 contained A $\beta$ - individuals; a consequence of using a clinical diagnosis for classification,  
11 instead of a biomarker-based diagnosis. Although this makes our demented group more  
12 heterogeneous, it does reflect current clinical practice and is in line with earlier research  
13 showing that ~20% of individuals with AD dementia are A $\beta$ - [44]. Third, our clinical follow-  
14 up may have been too short to obtain an accurate view of cognitive trajectories over time. And  
15 lastly, we chose the MMSE to assess cognition as this data was available in nearly all  
16 individuals, but it might not be sensitive enough to detect subtle cognitive decline and decline  
17 in specific cognitive domains. Future studies with longer follow-up and employing other  
18 cognitive measures should therefore validate our results regarding cognitive decline.

19

20 In conclusion, we found that NFL, Ng and YKL-40 were associated with A $\beta$  pathology,  
21 showing that axonal degeneration, synaptic dysfunction and neuroinflammation are all to some  
22 extent involved in AD pathophysiology. Furthermore, we found that NFL is a generic  
23 prognostic marker which is elevated early in AD, and has a profound influence on cognition.  
24 Ng is a useful AD marker as it is closely related to A $\beta$  and tau in all cognitive stages and is  
25 associated with cognition. YKL-40 has an influence on cognitive decline in absence of A $\beta$ , and



1 thereby may be of value to increase the accuracy of the prognosis of individuals with SNAP.  
2 Lastly, our data identifies NFL as the strongest predictor of cognitive decline in A $\beta$ +  
3 individuals across the cognitive stages. Altogether, our findings improve prognostic accuracy  
4 and increase our knowledge of biomarker changes in relation to disease evolution.

#### 6 **Authors' contributions**

7 Study concept and design: IB, SV, HZ & PJV. Acquisition and/or interpretation of data or  
8 samples: all authors. Statistical analysis and drafting the manuscript: IB, SV, & PJV. Critical  
9 revision of final draft of manuscript: all authors.

#### 10 **Funding**

11 The present study was conducted as part of the EMIF-AD project which has received support  
12 from the Innovative Medicines Initiative Joint Undertaking under EMIF grant agreement n $^{\circ}$   
13 115372, resources of which are composed of financial contribution from the European Union's  
14 Seventh Framework Program (FP7/2007-2013) and EFPIA companies' in kind contribution.  
15 The DESCRIPA study was funded by the European Commission within the 5th framework  
16 program (QLRT-2001- 2455). The EDAR study was funded by the European Commission  
17 within the 5th framework program (contract # 37670). The San Sebastian GAP study is  
18 partially funded by the Department of Health of the Basque Government (allocation  
19 17.0.1.08.12.0000.2.454.01.41142.001.H). The Lausanne center study was supported by a  
20 grant from the Swiss National Research Foundation to JP (SNF 320030\_141179). KS is  
21 supported by the University of Antwerp Research Fund, Belgium. FB is supported by the NIHR  
22 UCLH biomedical research centre. KB holds the Torsten Söderberg Professorship in medicine  
23 at the Swedish Royal Academy of Science. HZ is a Wallenberg Academy Fellow. The GAP  
24 study is supported by grants from the Department of Economic Promotion, Rural Areas and  
25 Territorial Balance of the Provincial Government of Gipuzkoa (124/16); the Department of

1 Health of the Basque Government (2016111096); and by the Carlos III Institute of Health  
2 (PI15/00919, PN de I+D+I 2013-2016); Obra Social Kutxa-Fundazioa and anonymous private  
3 donors. CT received grants from the European Commission, the Dutch Research Council  
4 (ZonMW), Association of Frontotemporal Dementia/Alzheimer's Drug Discovery Foundation,  
5 Alzheimer Netherlands.

6

### 7 **Competing interests**

8 Dr. Teunissen has functioned in advisory boards of Fujirebio and Roche, received non-financial  
9 support in the form of research consumables from ADxNeurosciences and Euroimmun,  
10 performed contract research or received grants from Probiodrug, Janssen prevention center,  
11 Boehringer, Brainsonline, AxonNeurosciences, EIP farma and Roche. Dr. Martinez-Lage  
12 reports personal fees from Lilly, Axon, General Electric and Nutricia for advisory boards, and  
13 lecturing fees from Lilly, Nutricia, Piramal. Dr. Blennow has served as a consultant or at  
14 advisory boards for Fujirebio Europe, IBL International, and Roche Diagnostics, and is a co-  
15 founder of Brain Biomarker Solutions in Gothenburg AB, a GU Venture-based platform  
16 company at the University of Gothenburg. The other authors declare no conflict of interest.

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## 2 **References**

- 3 [1] Zetterberg H. Applying fluid biomarkers to Alzheimer's disease. *Am J Physiol Cell*  
4 *Physiol.* 2017;313:C3-C10.
- 5 [2] Albert MS, DeKosky ST, Dickson D, Dubois B, Feldman HH, Fox NC, et al. The  
6 diagnosis of mild cognitive impairment due to Alzheimer's disease: recommendations from  
7 the National Institute on Aging-Alzheimer's Association workgroups on diagnostic guidelines  
8 for Alzheimer's disease. *Alzheimer's & dementia : the journal of the Alzheimer's Association.*  
9 2011;7:270-9.
- 10 [3] Blennow K, Mattsson N, Scholl M, Hansson O, Zetterberg H. Amyloid biomarkers in  
11 Alzheimer's disease. *Trends Pharmacol Sci.* 2015;36:297-309.
- 12 [4] Lewczuk P, Matzen A, Blennow K, Parnetti L, Molinuevo JL, Eusebi P, et al.  
13 Cerebrospinal Fluid Aβ<sub>42</sub>/Aβ<sub>40</sub> Corresponds Better than Aβ<sub>42</sub> to Amyloid PET in  
14 Alzheimer's Disease. *Journal of Alzheimer's disease : JAD.* 2017;55:813-22.
- 15 [5] Blennow K, Wallin A, Agren H, Spenger C, Siegfried J, Vanmechelen E. Tau protein in  
16 cerebrospinal fluid: a biochemical marker for axonal degeneration in Alzheimer disease?  
17 *Molecular and chemical neuropathology.* 1995;26:231-45.
- 18 [6] Hampel H, Blennow K, Shaw LM, Hoessler YC, Zetterberg H, Trojanowski JQ. Total  
19 and phosphorylated tau protein as biological markers of Alzheimer's disease. *Experimental*  
20 *gerontology.* 2010;45:30-40.
- 21 [7] Zetterberg H, Skillback T, Mattsson N, Trojanowski JQ, Portelius E, Shaw LM, et al.  
22 Association of Cerebrospinal Fluid Neurofilament Light Concentration With Alzheimer  
23 Disease Progression. *JAMA Neurol.* 2016;73:60-7.
- 24 [8] Bruno D, Pomara N, Nierenberg J, Ritchie JC, Lutz MW, Zetterberg H, et al. Levels of  
25 cerebrospinal fluid neurofilament light protein in healthy elderly vary as a function of  
26 TOMM40 variants. *Experimental gerontology.* 2012;47:347-52.
- 27 [9] Antonell A, Mansilla A, Rami L, Llado A, Iranzo A, Olives J, et al. Cerebrospinal fluid  
28 level of YKL-40 protein in preclinical and prodromal Alzheimer's disease. *Journal of*  
29 *Alzheimer's disease : JAD.* 2014;42:901-8.
- 30 [10] Portelius E, Zetterberg H, Skillback T, Tornqvist U, Andreasson U, Trojanowski JQ, et  
31 al. Cerebrospinal fluid neurogranin: relation to cognition and neurodegeneration in  
32 Alzheimer's disease. *Brain : a journal of neurology.* 2015;138:3373-85.
- 33 [11] Kester MI, Teunissen CE, Crimmins DL, Herries EM, Ladenson JH, Scheltens P, et al.  
34 Neurogranin as a Cerebrospinal Fluid Biomarker for Synaptic Loss in Symptomatic  
35 Alzheimer Disease. *JAMA Neurol.* 2015;72:1275-80.
- 36 [12] Zetterberg H, Skillback T, Mattsson N, Trojanowski JQ, Portelius E, Shaw LM, et al.  
37 Association of Cerebrospinal Fluid Neurofilament Light Concentration With Alzheimer  
38 Disease Progression. *JAMA Neurol.* 2016;73:60-7.
- 39 [13] Sala-Llonch R, Idland AV, Borza T, Watne LO, Wyller TB, Braekhus A, et al.  
40 Inflammation, Amyloid, and Atrophy in The Aging Brain: Relationships with Longitudinal  
41 Changes in Cognition. *Journal of Alzheimer's disease : JAD.* 2017;58:829-40.
- 42 [14] Janelidze S, Hertze J, Zetterberg H, Landqvist Waldo M, Santillo A, Blennow K, et al.  
43 Cerebrospinal fluid neurogranin and YKL-40 as biomarkers of Alzheimer's disease. *Ann Clin*  
44 *Transl Neurol.* 2016;3:12-20.
- 45 [15] Lista S, Toschi N, Baldacci F, Zetterberg H, Blennow K, Kilimann I, et al. Diagnostic  
46 accuracy of CSF neurofilament light chain protein in the biomarker-guided classification  
47 system for Alzheimer's disease. *Neurochem Int.* 2017;108:355-60.

- 1 [16] Thorsell A, Bjerke M, Gobom J, Brunhage E, Vanmechelen E, Andreasen N, et al.  
2 Neurogranin in cerebrospinal fluid as a marker of synaptic degeneration in Alzheimer's  
3 disease. *Brain research*. 2010;1362:13-22.
- 4 [17] Bos I, Vos, S.J.B., Vandenberghe, R., Scheltens, P., Engelborghs, S., Frisoni, G.,  
5 Molinuevo, J., Wallin, A., Lléo, A., Popp, J., Martinez-Lage, P., Baird, A., Dobson, R.,  
6 Legido-Quigley, C., Bertram, L., Slegers, K., Kate ten, M., Barkhof, F., Zetterberg, H.,  
7 Lovestone, S., Streffer, J., Visser, P. . The EMIF-AD Multimodal Biomarker Discovery  
8 Study: Design, methods and cohort characteristics. *Alzheimer's research & therapy*. 2018.
- 9 [18] Visser PJ, Verhey FR, Boada M, Bullock R, De Deyn PP, Frisoni GB, et al.  
10 Development of screening guidelines and clinical criteria for predementia Alzheimer's  
11 disease. The DESCRIPA Study. *Neuroepidemiology*. 2008;30:254-65.
- 12 [19] Reijs BLR, Ramakers I, Elias-Sonnenschein L, Teunissen CE, Koel-Simmelink M,  
13 Tsolaki M, et al. Relation of Odor Identification with Alzheimer's Disease Markers in  
14 Cerebrospinal Fluid and Cognition. *Journal of Alzheimer's disease : JAD*. 2017;60:1025-34.
- 15 [20] Galluzzi S, Marizzoni M, Babiloni C, Albani D, Antelmi L, Bagnoli C, et al. Clinical  
16 and biomarker profiling of prodromal Alzheimer's disease in workpackage 5 of the  
17 Innovative Medicines Initiative PharmaCog project: a 'European ADNI study'. *Journal of*  
18 *internal medicine*. 2016;279:576-91.
- 19 [21] van der Flier WM, Pijnenburg YA, Prins N, Lemstra AW, Bouwman FH, Teunissen CE,  
20 et al. Optimizing patient care and research: the Amsterdam Dementia Cohort. *Journal of*  
21 *Alzheimer's disease : JAD*. 2014;41:313-27.
- 22 [22] Somers C, Struyfs H, Goossens J, Niemantsverdriet E, Luyckx J, De Roeck N, et al. A  
23 Decade of Cerebrospinal Fluid Biomarkers for Alzheimer's Disease in Belgium. *Journal of*  
24 *Alzheimer's disease : JAD*. 2016;54:383-95.
- 25 [23] Estanga A, Ecay-Torres M, Ibanez A, Izagirre A, Villanua J, Garcia-Sebastian M, et al.  
26 Beneficial effect of bilingualism on Alzheimer's disease CSF biomarkers and cognition.  
27 *Neurobiology of aging*. 2017;50:144-51.
- 28 [24] Tautvydaite D, Kukreja D, Antonietti JP, Henry H, von Gunten A, Popp J. Interaction  
29 between personality traits and cerebrospinal fluid biomarkers of Alzheimer's disease  
30 pathology modulates cognitive performance. *Alzheimer's research & therapy*. 2017;9:6.
- 31 [25] Petersen RC. Mild cognitive impairment as a diagnostic entity. *Journal of internal*  
32 *medicine*. 2004;256:183-94.
- 33 [26] McKhann G, Drachman D, Folstein M, Katzman R, Price D, Stadlan EM. Clinical  
34 diagnosis of Alzheimer's disease: report of the NINCDS-ADRDA Work Group under the  
35 auspices of Department of Health and Human Services Task Force on Alzheimer's Disease.  
36 *Neurology*. 1984;34:939-44.
- 37 [27] Folstein MF, Folstein SE, McHugh PR. "Mini-mental state". A practical method for  
38 grading the cognitive state of patients for the clinician. *Journal of psychiatric research*.  
39 1975;12:189-98.
- 40 [28] Olsson B, Hertze J, Lautner R, Zetterberg H, Nagga K, Hoglund K, et al. Microglial  
41 markers are elevated in the prodromal phase of Alzheimer's disease and vascular dementia.  
42 *Journal of Alzheimer's disease : JAD*. 2013;33:45-53.
- 43 [29] Benaglia T, Chauveau D, Hunter D, Young D. mixtools: An R package for analyzing  
44 finite mixture models. *Journal of Statistical Software*. 2009;32:1-29.
- 45 [30] Palmqvist S, Zetterberg H, Blennow K, Vestberg S, Andreasson U, Brooks DJ, et al.  
46 Accuracy of brain amyloid detection in clinical practice using cerebrospinal fluid  $\beta$ -amyloid  
47 42: a cross-validation study against amyloid positron emission tomography. *JAMA*  
48 *neurology*. 2014;71:1282-9.
- 49 [31] Selkoe DJ. Alzheimer's disease is a synaptic failure. *Science*. 2002;298:789-91.

- 1 [32] Musardo S, Marcello E. Synaptic dysfunction in Alzheimer's disease: From the role of  
2 amyloid beta-peptide to the alpha-secretase ADAM10. *European journal of pharmacology*.  
3 2017;817:30-7.
- 4 [33] Skillback T, Farahmand B, Bartlett JW, Rosen C, Mattsson N, Nagga K, et al. CSF  
5 neurofilament light differs in neurodegenerative diseases and predicts severity and survival.  
6 *Neurology*. 2014;83:1945-53.
- 7 [34] Selkoe DJ. The molecular pathology of Alzheimer's disease. *Neuron*. 1991;6:487-98.
- 8 [35] Iqbal K, Alonso Adel C, Chen S, Chohan MO, El-Akkad E, Gong CX, et al. Tau  
9 pathology in Alzheimer disease and other tauopathies. *Biochimica et biophysica acta*.  
10 2005;1739:198-210.
- 11 [36] Craig-Schapiro R, Perrin RJ, Roe CM, Xiong C, Carter D, Cairns NJ, et al. YKL-40: a  
12 novel prognostic fluid biomarker for preclinical Alzheimer's disease. *Biological psychiatry*.  
13 2010;68:903-12.
- 14 [37] Sutphen CL, Jasielc MS, Shah AR, Macy EM, Xiong C, Vlassenko AG, et al.  
15 Longitudinal Cerebrospinal Fluid Biomarker Changes in Preclinical Alzheimer Disease  
16 During Middle Age. *JAMA Neurol*. 2015;72:1029-42.
- 17 [38] Morris JC, Roe CM, Xiong C, Fagan AM, Goate AM, Holtzman DM, et al. APOE  
18 predicts amyloid-beta but not tau Alzheimer pathology in cognitively normal aging. *Annals*  
19 *of neurology*. 2010;67:122-31.
- 20 [39] Gispert JD, Monte GC, Suarez-Calvet M, Falcon C, Tucholka A, Rojas S, et al. The  
21 APOE epsilon4 genotype modulates CSF YKL-40 levels and their structural brain correlates  
22 in the continuum of Alzheimer's disease but not those of sTREM2. *Alzheimers Dement*  
23 *(Amst)*. 2017;6:50-9.
- 24 [40] Kvartberg H, Duits FH, Ingelsson M, Andreasen N, Ohrfelt A, Andersson K, et al.  
25 Cerebrospinal fluid levels of the synaptic protein neurogranin correlates with cognitive  
26 decline in prodromal Alzheimer's disease. *Alzheimer's & dementia : the journal of the*  
27 *Alzheimer's Association*. 2015;11:1180-90.
- 28 [41] Masliah E, Mallory M, Alford M, DeTeresa R, Hansen LA, McKeel DW, Jr., et al.  
29 Altered expression of synaptic proteins occurs early during progression of Alzheimer's  
30 disease. *Neurology*. 2001;56:127-9.
- 31 [42] Mattsson N, Insel PS, Palmqvist S, Portelius E, Zetterberg H, Weiner M, et al.  
32 Cerebrospinal fluid tau, neurogranin, and neurofilament light in Alzheimer's disease. *EMBO*  
33 *Mol Med*. 2016;8:1184-96.
- 34 [43] Jack CR, Jr., Knopman DS, Chetelat G, Dickson D, Fagan AM, Frisoni GB, et al.  
35 Suspected non-Alzheimer disease pathophysiology--concept and controversy. *Nat Rev*  
36 *Neurol*. 2016;12:117-24.
- 37 [44] Ossenkoppele R, Jansen WJ, Rabinovici GD, Knol DL, van der Flier WM, van Berckel  
38 BN, et al. Prevalence of amyloid PET positivity in dementia syndromes: a meta-analysis.  
39 *Jama*. 2015;313:1939-49.
- 40

## Figure Legends

### Figure 1. CSF NFL, Ng, YKL-40 and T-tau levels by diagnostic groups and A $\beta$ status

Boxplots (displaying first quartile, median and third quartile) and scatterplots of CSF neurogranin (Ng), neurofilament (NFL) and YKL-40 by diagnostic groups and by A $\beta$  status (A $\beta$ -: green; A $\beta$ +: orange). \* $p < 0.05$ , \*\* $p < 0.01$ , \*\*\* $p < 0.001$  comparisons by A $\beta$  status within diagnostic group. Figure A shows log transformed NFL concentrations, Figure B shows log transformed Ng concentrations and Figure C shows log transformed YKL-40 concentrations. Figure D shows log transformed T-tau concentrations.

### Figure 2. Influence of CSF NFL, Ng, YKL-40 and T-tau on cognition in the total group.

The graphs show mean scores and 95% confidence intervals of cognitive performance over time for high (red) and low (blue) CSF biomarker levels and by A $\beta$  status (dashed lines: A $\beta$ -; solid lines: A $\beta$ +) . \* $p < 0.05$  comparisons within A $\beta$  group, \*\* $p < 0.01$  comparisons within A $\beta$  group, \*\*\* $p < 0.001$  comparisons within A $\beta$  group. Figure A shows the influence of NFL levels. Figure B shows the influence of Ng levels. Figure C shows the influence of YKL-40 levels. Figure D shows the influence of T-tau levels.

### Figure 3. Schematic overview of associations between NFL, Ng and YKL-40 with APOE $\epsilon 4$ positivity and cognition by diagnostic group and A $\beta$ status

This figure shows the various associations examined in this study. In the top panel the associations in cognitively normal are visualized. In the middle panel the associations in individuals with MCI are visualized and in the bottom panel the association in individuals with AD-type dementia. The green arrows represent association in A $\beta$ - individuals, the orange arrow represent association in A $\beta$ + individuals. Negative association are visualized with a minus (-) and positive association with a plus (+).

**Table 1. Baseline characteristics and CSF biomarker values across the diagnostic groups and by Aβ status**

	CN		MCI		AD-type dementia	
	Aβ- n=95 (A)	Aβ+ n=45 (B)	Aβ- n=187 (C)	Aβ+ n=263 (D)	Aβ- n=23 (E)	Aβ+ n=157 (F)
Age	62.7 ± 7.3 <sup>B,C,D,E,F</sup>	69.5 ± 8.1 <sup>A,E</sup>	68.6 ± 8.2 <sup>A,D,E</sup>	71.4 ± 7.1 <sup>A,C,F</sup>	74.2 ± 7.9 <sup>A,B,C,F</sup>	69.8 ± 8.8 <sup>A,D,E</sup>
Female, n	49 (52)	23 (51)	89 (48)	145 (55)	8 (34)	85 (54)
Education in years	12.6 ± 3.5 <sup>C,D,E,F</sup>	12.2 ± 3.9 <sup>C,D,E,F</sup>	10.4 ± 3.8 <sup>A,B,E</sup>	11.0 ± 3.6 <sup>A,B,E</sup>	8.6 ± 4.7 <sup>A,B,C,D,F</sup>	10.6 ± 3.6 <sup>A,B,E</sup>
<i>APOE</i> -ε4 carrier, n	28 (30) <sup>B,C,D,F</sup>	27 (60) <sup>A,C,E</sup>	38 (20) <sup>A,B,D,F</sup>	175 (67) <sup>A,C,E</sup>	5 (22) <sup>B,D,F</sup>	104 (66) <sup>A,C,E</sup>
MMSE	28.7 ± 1.2 <sup>C,D,E,F</sup>	28.7 ± 1.3 <sup>C,D,E,F</sup>	26.8 ± 2.4 <sup>A,B,D,E,F</sup>	25.8 ± 2.6 <sup>A,B,C,E,F</sup>	22.4 ± 4.5 <sup>A,B,C,D</sup>	21.3 ± 4.8 <sup>A,B,C,D</sup>
Aβ <sub>38</sub> , pg/ml	2245.7 ± 834.3	2405.5 ± 670.0 <sup>F</sup>	2247.3 ± 948.2 <sup>F</sup>	2160.2 ± 858.6 <sup>F</sup>	2447.4 ± 1248.2	2139.6 ± 834.8 <sup>B,C,D</sup>
Aβ <sub>40</sub> , pg/ml	5217.7 ± 1709.4	5585.8 ± 1470.9 <sup>F</sup>	5190.4 ± 1970.7 <sup>F</sup>	4939.9 ± 1824.2 <sup>F</sup>	5556.8 ± 2269.6	5078.1 ± 1801.5 <sup>B,C,D</sup>
Aβ <sub>42</sub> , pg/ml	466.2 ± 182.8 <sup>B,D,F</sup>	254.4 ± 75.0 <sup>A,C,E,F</sup>	467.2 ± 218.2 <sup>B,D,F</sup>	211.6 ± 88.8 <sup>A,C,E,F</sup>	461.4 ± 217.6 <sup>B,D,F</sup>	215.9 ± 89.4 <sup>A,B,C,D,E</sup>
Aβ <sub>42/40</sub> ratio	0.089 ± 0.01 <sup>B,D,E,F</sup>	0.045 ± 0.01 <sup>A,C,D,E</sup>	0.089 ± 0.02 <sup>B,D,F</sup>	0.04 ± 0.01 <sup>A,C,E</sup>	0.08 ± 0.01 <sup>B,D,F</sup>	0.04 ± 0.01 <sup>A,C,E</sup>
P-tau, pg/ml <sup>#</sup>	38.7 ± 12.4 <sup>B,C,D,F</sup>	61.5 ± 27.3 <sup>A,C,D,F</sup>	48.2 ± 18.6 <sup>A,B,D,F</sup>	80.3 ± 32.8 <sup>A,B,C,E</sup>	41.5 ± 17.4 <sup>D,F</sup>	86.2 ± 41.1 <sup>A,B,C,E</sup>
T-tau, pg/ml <sup>#</sup>	197.3 ± 72.5 <sup>B,C,D,F</sup>	405.2 ± 330.0 <sup>A,C,D,F</sup>	280.4 ± 134.2 <sup>A,B,D,F</sup>	572.3 ± 315.9 <sup>A,B,C,E</sup>	225.3 ± 82.7 <sup>D,F</sup>	708.0 ± 445.0 <sup>A,B,C,E</sup>
NFL, pg/ml	627.4 ± 293.3 <sup>B,C,D,E,F</sup>	983.13 ± 678.4 <sup>A,E,F</sup>	1031.2 ± 919.1 <sup>A,D,E,F</sup>	1242.3 ± 2556.1 <sup>A,C,F</sup>	1931.9 ± 1934.8 <sup>A,C</sup>	1742.2 ± 2893.2 <sup>A,B,C,D</sup>
Ng, pg/ml	110.8 ± 224 <sup>B,D,F</sup>	152.6 ± 149.6 <sup>A,C</sup>	99.2 ± 102.9 <sup>B,D,F</sup>	175.5 ± 217.8 <sup>A,C,E</sup>	118.3 ± 136.0 <sup>D,F</sup>	155.2 ± 121.4 <sup>A,C,E</sup>
YKL-40, ng/ml	127.0 ± 45.4 <sup>B,C,D,E,F</sup>	175.1 ± 63.6 <sup>A</sup>	162.2 ± 65.2 <sup>A,D,F</sup>	183.4 ± 60.5 <sup>A,C</sup>	184.2 ± 64.6 <sup>A</sup>	193.6 ± 68.7 <sup>A,C</sup>

Results are mean ± SD or number (%). Biomarker comparisons were done with the log transformed values for Aβ<sub>42</sub>, NFL, Ng, YKL-40, p-tau and t-tau, and adjusted for age, gender, *APOE*-ε4 carrier status and with study as a random effect. <sup>#</sup>P-tau and t-tau values were analyzed locally and available in a subgroup p-tau: CN n=103, MCI n=403, AD n=124; t-tau: CN n=103, MCI n=399, AD n=119. <sup>A</sup>p<0.05 compared to CN Aβ-, <sup>B</sup>p<0.05 compared to CN Aβ+, <sup>C</sup>p<0.05 compared to MCI Aβ-, <sup>D</sup>p<0.05 compared to MCI Aβ+, <sup>E</sup>p<0.05 compared to AD dementia Aβ-, <sup>F</sup>p<0.05 compared to AD dementia Aβ+. Abbreviations: Aβ= amyloid-beta; AD = Alzheimer's Disease; *APOE* = Apolipoprotein E; CN = cognitively normal; MCI = Mild Cognitive Impairment; NFL = neurofilament light; Ng = neurogranin; P-tau = phosphorylated tau; T-tau = total tau.

**Table 2. Comparisons of CSF NFL, Ng, YKL-40 and T-tau concentrations by APOE ε4 status within Aβ group**

Biomarker	Group	Aβ-			Aβ+		
		number (ε4-/ ε4+)	ε4-	ε4+	number (ε4-/ ε4+)	ε4-	ε4+
NFL, pg/ml	All	233/70	1042.5 ± 69.1	728.7 ± 50.1*	159/299	1460.5 ± 246.7 <sup>^</sup>	1349.7 ± 129.5 <sup>^</sup>
	CN	67/28	<b>627.1 ± 33.5</b>	628.2 ± 64.2	18/27	1044.2 ± 117.5	942.4 ± 150.6
	MCI	148/38	1091.9 ± 81.7 <sup>^</sup>	795.0 ± 77.0*	88/168	1509.8 ± 441.7 <sup>^</sup>	1102.2 ± 76.1 <sup>^</sup>
	AD-type dementia	18/4	2183.1 ± 485.7 <sup>^</sup>	801.7 ± 123.8	53/104	1519.9 ± 98.7 <sup>^</sup>	1855.4 ± 345.0 <sup>^</sup>
Ng, pg/ml	All	202/63	101.7 ± 6.8	111.7 ± 32.5**	149/292	167.3 ± 11.8 <sup>^</sup>	166.0 ± 11.8 <sup>^</sup>
	CN	54/24	<b>91.3 ± 11.2</b>	154.7 ± 79.3	15/27	194.4 ± 57.5 <sup>^</sup>	129.3 ± 16.2
	MCI	132/35	101.3 ± 8.2	91.4 ± 22.1*	81/169	169.1 ± 15.9 <sup>^</sup>	178.5 ± 18.9 <sup>^</sup>
	AD-type dementia	16/4	140.1 ± 36.1	31.1 ± 8.1*	53/96	156.8 ± 16.0 <sup>^</sup>	154.3 ± 12.7 <sup>^</sup>
YKL-40, ng/ml	All	234/71	156.0 ± 4.2	142.6 ± 6.9	158/305	192.9 ± 4.8 <sup>^</sup>	182.5 ± 3.7 <sup>^</sup>
	CN	67/28	<b>123.3 ± 4.8</b>	136.0 ± 11.0	18/27	180.8 ± 16.8 <sup>^</sup>	171.3 ± 11.4 <sup>^</sup>
	MCI	149/38	165.4 ± 5.5 <sup>^</sup>	149.6 ± 9.4 <sup>^</sup>	87/174	187.6 ± 5.9 <sup>^</sup>	181.3 ± 4.8 <sup>^</sup>
	AD-type dementia	18/5	200.2 ± 14.9 <sup>^</sup>	126.8 ± 11.1	53/104	205.7 ± 8.9 <sup>^</sup>	187.5 ± 6.9 <sup>^</sup>
T-tau, pg/ml	All	170/47	266.2 ± 10.1	221.3 ± 15.1	125/240	627.7 ± 39.9 <sup>^</sup>	576.8 ± 20.5 <sup>^</sup>
	CN	43/15	<b>198.3 ± 10.7</b>	194.6 ± 21.1	14/21	455.8 ± 131.8 <sup>^</sup>	371.5 ± 33.9 <sup>^</sup>
	MCI	119/29	292.2 ± 12.6 <sup>^</sup>	232.0 ± 20.3	78/150	578.5 ± 44.2 <sup>^</sup>	569.1 ± 22.1 <sup>^</sup>
	AD-type dementia	8/3	182.5 ± 36.5	332.3 ± 122.1	33/69	816.8 ± 87.3 <sup>^</sup>	656.0 ± 43.2 <sup>^</sup>

Results are mean ± SE. Comparisons were conducted between log-transformed biomarker concentrations and adjusted for age, gender and study. \*p<0.05, \*\*p<0.01, \*\*\*p<0.001 as compared to the ε4- within the Aβ group. <sup>^</sup>p<0.05 compared to the CN Aβ- ε4- group (in bold). Abbreviations: Aβ= amyloid-beta; AD = Alzheimer's Disease; CN = cognitively normal; MCI = Mild Cognitive Impairment; NFL = neurofilament light; Ng = neurogranin.



**Table 3. Influence of CSF NFL, Ng, YKL-40 and T-tau on cognitive performance and decline by A $\beta$  status**

Biomarker	Group	A $\beta$ -			A $\beta$ +		
		number (low/high) <sup>#</sup>	Baseline difference	Slope difference	number (low/high) <sup>#</sup>	Baseline difference	Slope difference
NFL	All	194/109	-0.98 $\pm$ 0.44*	-0.40 $\pm$ 0.13**	182/276	-1.89 $\pm$ 0.34***	-0.39 $\pm$ 0.10***
	CN	74/21	0.14 $\pm$ 0.78	0.40 $\pm$ 0.27	28/17	-0.36 $\pm$ 1.03	-0.40 $\pm$ 0.40
	MCI	112/74	-0.86 $\pm$ 0.45	-0.51 $\pm$ 0.14***	122/134	-0.72 $\pm$ 0.36*	0.04 $\pm$ 0.17
	AD-type dementia	8/14	-2.53 $\pm$ 1.39	-0.33 $\pm$ 0.71*	32/125	-1.71 $\pm$ 0.68*	-0.60 $\pm$ 0.25*
Ng	All	171/94	0.51 $\pm$ 0.45	0.21 $\pm$ 0.10	182/259	-0.58 $\pm$ 0.34*	-0.15 $\pm$ 0.11
	CN	52/26	0.45 $\pm$ 0.80	0.17 $\pm$ 0.25	17/25	0.49 $\pm$ 1.08	-0.29 $\pm$ 0.37
	MCI	108/59	0.10 $\pm$ 0.48	0.25 $\pm$ 0.12*	109/141	-0.52 $\pm$ 0.36	-0.24 $\pm$ 0.16
	AD-type dementia	11/9	4.90 $\pm$ 1.49**	-2.48 $\pm$ 0.74**	56/93	0.01 $\pm$ 0.62	-0.76 $\pm$ 0.22**
YKL-40	All	198/107	-0.45 $\pm$ 0.42	-0.44 $\pm$ 0.13**	186/277	0.07 $\pm$ 0.34	0.01 $\pm$ 0.10
	CN	74/21	-0.36 $\pm$ 0.82	0.29 $\pm$ 0.20	20/25	-0.32 $\pm$ 1.00	-0.32 $\pm$ 0.40
	MCI	113/74	0.07 $\pm$ 0.43	-0.60 $\pm$ 0.11***	111/150	0.18 $\pm$ 0.35	0.15 $\pm$ 0.16
	AD-type dementia	11/12	-2.12 $\pm$ 1.36	-1.40 $\pm$ 0.59*	55/102	0.79 $\pm$ 0.60	0.22 $\pm$ 0.23
T-tau	All	236/66	-0.67 $\pm$ 0.49	-0.77 $\pm$ 0.14***	106/355	-1.64 $\pm$ 0.37***	-0.38 $\pm$ 0.12**
	CN	85/10	0.71 $\pm$ 1.01	0.02 $\pm$ 0.36	23/21	-0.26 $\pm$ 1.01	0.01 $\pm$ 0.41
	MCI	141/43	-0.51 $\pm$ 0.51	-0.79 $\pm$ 0.12***	60/201	-0.87 $\pm$ 0.40*	-0.18 $\pm$ 0.21
	AD-type dementia	10/13	-2.96 $\pm$ 1.37*	-0.96 $\pm$ 0.56*	23/133	-0.41 $\pm$ 0.81	-0.41 $\pm$ 0.31

Baseline differences in MMSE scores are mean difference  $\pm$  standard error between low and high NFL, Ng and YKL-40 groups defined by median-split. Slopes are linear mixed model coefficient indicating annual decline  $\pm$  standard error, relative to group with low biomarker level with MMSE score as outcome. \*p<0.05, \*\*p<0.01, \*\*\*p<0.001 compared to group with low biomarker levels, adjusted for age, gender, education level and study. Comparisons in the total sample were also adjusted for baseline diagnosis. <sup>#</sup>Number with low and high biomarker levels at baseline, for t-tau number with normal and abnormal t-tau levels at baseline.

**Table 4. Independent influence of biomarkers on cognitive decline across the diagnostic groups**

		A $\beta$ -		A $\beta$ +	
		$\beta$	p-value	$\beta$	p-value
<b>CN</b>	High NFL	0.20 $\pm$ 0.31	0.508	-1.19 $\pm$ 0.39	<b>0.004</b>
	High Ng	0.27 $\pm$ 0.21	0.216	-0.54 $\pm$ 0.35	0.134
	High YKL-40	-0.09 $\pm$ 0.26	0.741	0.28 $\pm$ 0.31	0.367
	High T-tau	-0.10 $\pm$ 0.30	0.737	0.48 $\pm$ 0.38	0.219
<b>MCI</b>	High NFL	-0.30 $\pm$ 0.15	<b>0.045</b>	-0.74 $\pm$ 0.26	<b>0.001</b>
	High Ng	0.28 $\pm$ 0.14	0.060	0.46 $\pm$ 0.16	<b>0.005</b>
	High YKL-40	-0.19 $\pm$ 0.16	0.242	0.12 $\pm$ 0.15	0.430
	High T-tau	-0.43 $\pm$ 0.18	<b>0.017</b>	-0.58 $\pm$ 0.22	<b>0.009</b>
<b>AD-type dementia</b>	High NFL	2.83 $\pm$ 2.77	0.857	-0.91 $\pm$ 0.35	<b>0.009</b>
	High Ng	0.42 $\pm$ 2.76	0.993	-0.64 $\pm$ 0.27	<b>0.021</b>
	High YKL-40	-9.12 $\pm$ 3.77	0.939	0.32 $\pm$ 0.31	0.315
	High T-tau	4.48 $\pm$ 2.65	0.971	-0.74 $\pm$ 0.43	0.084

Numbers are linear mixed model coefficients  $\pm$  standard error with MMSE scores over time as dependent variable adjusted for age, gender and years of education. All CSF variables were entered at the same step. NFL, Ng and YKL-40 were dichotomized based on median-split, T-tau based on the local cut-off for abnormality. Abbreviations: A $\beta$  = amyloid-beta, CN = cognitively normal, MCI = Mild Cognitive Impairment, NFL = Neurofilament light, Ng = neurogranin, T-tau = Total tau.