



Figures and figure supplements

Neurocalcin regulates nighttime sleep and arousal in *Drosophila*

Ko-Fan Chen et al

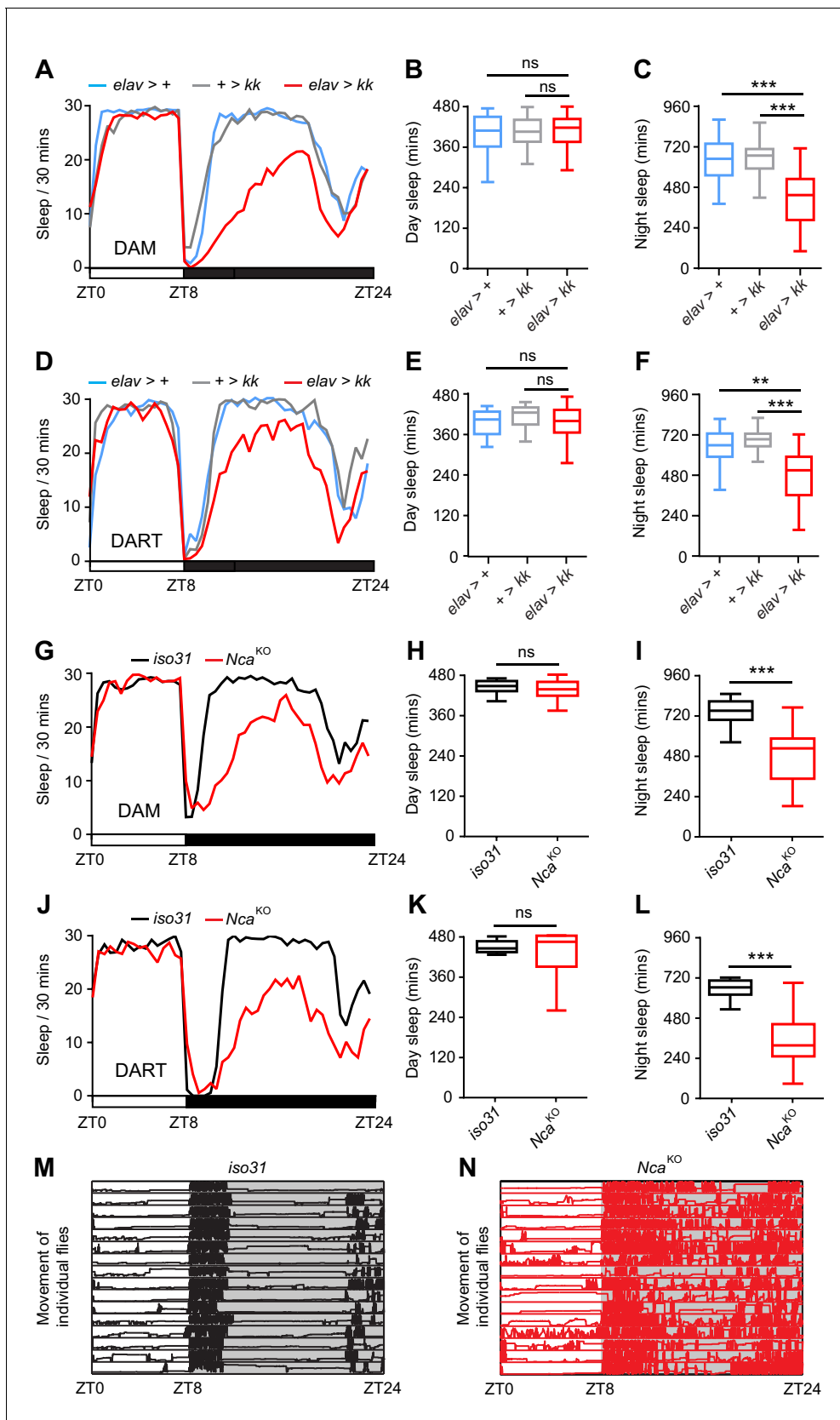


Figure 1. Neurocalcin promotes night sleep. (A) Mean sleep levels measured using the DAM system under 8L: 16D conditions for adult male pan-neuronal *Nca* knockdown flies (*elav > kk*) and associated controls (*elav-Gal4* driver or *kk* RNAi transgene heterozygotes). (B–C) Median day (B) and night (C) sleep levels for *elav > +*, *+ > kk*, and *elav > kk* genotypes. (D–F) Similar to (A–C), but using the DART system. (G–I) Median day (G) and night (I) sleep levels for *iso31* and *Nca^{KO}* genotypes. (J–L) Similar to (D–F), but using the DART system. (M) Movement heatmaps for *iso31* flies. (N) Movement heatmaps for *Nca^{KO}* flies. Error bars represent SEM. Statistical significance is indicated by asterisks: * p < 0.05, ** p < 0.01, *** p < 0.001, ns = not significant.

Figure 1 continued

(C) sleep levels in the above genotypes. $n = 54\text{--}55$. Data are presented as Tukey box plots. The 25th, Median, and 75th percentiles are shown. Whiskers represent 1.5 x the interquartile range. Identical representations are used in all subsequent box plots. (D) Mean sleep levels measured using the DART system in 8L: 16D conditions for male adult pan-neuronal *Nca* knockdown flies (*elav > kk*) and associated controls. (E–F) Median day (E) and night (F) sleep levels in the above genotypes. $n = 20$ per genotype. (G) Mean sleep levels in 8L: 16D conditions for *Nca*^{KO} adult males and *iso31* controls measured using the DAM. (H–I) Median day (H) and night (I) sleep levels in the above genotypes. $n = 32$ per genotype. (J) Mean sleep levels in 8L: 16D conditions for *Nca*^{KO} adult males and *iso31* controls measured by DART. (K–L) Median day (K) and night (L) sleep levels in the above genotypes. $n = 16$ per genotype. (M–N) The longitudinal movement for individual *iso31* (M) and *Nca*^{KO} (N) flies are shown as rows of traces plotting vertical position (Y-axis) over 24 hr (X-axis) under 8L: 16D condition. ns (not significant) - $p > 0.05$, ** $p < 0.01$, *** $p < 0.001$, Kruskal-Wallis test with Dunn's post-hoc test (B–C, E–F) or Mann-Whitney U-test (H–I, K–L).

DOI: <https://doi.org/10.7554/eLife.38114.002>

<i>Hs</i> HPCA	1	MGKQNSKLRPEMLQDLRENTFSELELQEWYKGFLLKDCPTGILNVDEFKKIYANFFPYGD
<i>Dm</i> NCA	1	MGKQNSKLRPEVLEDLKQNTFETDAELQEWYKGFLLKDCPSGHLSVEEFKKIYGNFFPYGD
		EF Hand 1
<i>Hs</i> HPCA	61	ASKFAEHVFRFTDINS DGTIDFREFTI ALSVTSRGRLEQKLMWAFSMYDL DGN GYISREE
<i>Dm</i> NCA	61	ASKFAEHVFRFTDANG DGTIDFREFLC ALSVTSRGRLEQKLMWAFSMYDL DGN GYISRQE
		EF Hand 2
<i>Hs</i> HPCA	121	MLEIVQAIYKMVS SVMKMPEDESTPEKRTDKIFRQMDTNN D G K L S L E E F I R G A K S D P S I V
<i>Dm</i> NCA	121	MLEIVTAIYKMVG SVMKMPEDESTPEKRTDKIFRQMDR N K D G K L S L E E F I E G A K S D P S I V
		EF Hand 3
<i>Hs</i> HPCA	181	RLLQCDEPSSASQF
<i>Dm</i> NCA	181	RLLQCDEQSH---

Figure 1—figure supplement 1. Human Hippocalcin and *Drosophila* Neurocalcin are highly homologous neuronal calcium sensors. Amino-acid alignment of human Hippocalcin (*Hs* HPCA) and *Drosophila* Neurocalcin (*Dm* NCA) is shown. Blue boxes: location of the calcium-binding EF-hand domains of Hippocalcin and Neurocalcin. Black boxes represent full amino-acid conservation, grey boxes represent functional conservation.

DOI: <https://doi.org/10.7554/eLife.38114.003>

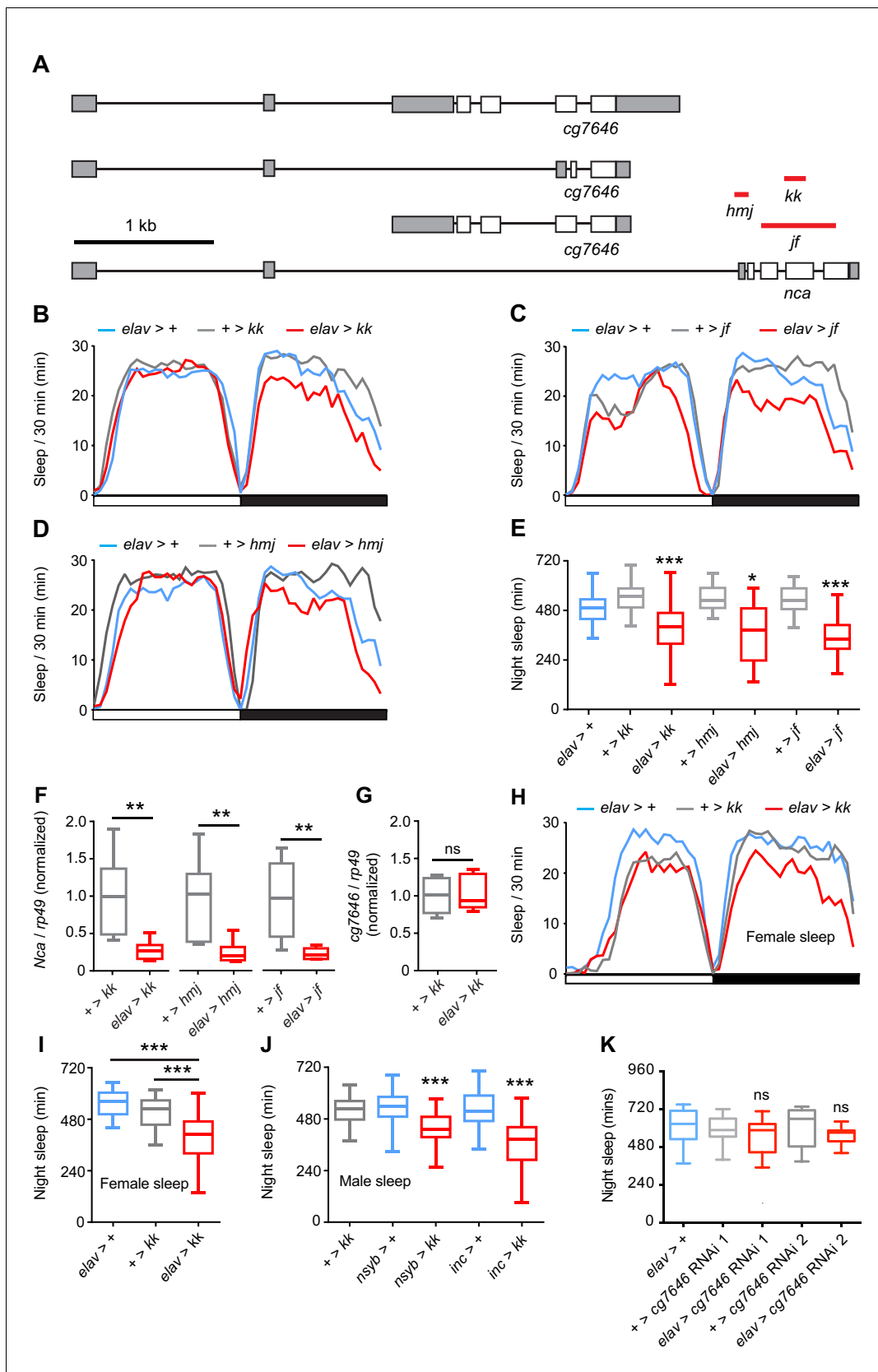


Figure 1—figure supplement 2. Pan-neuronal knockdown of *Nca* using independent RNAi lines causes night sleep loss. (A) Schematic showing transcripts derived from the *Nca* locus alongside transcripts derived from the *cg7646* locus, which shares common 5' untranslated regions with *Nca*. Figure 1—figure supplement 2 continued on next page

Figure 1—figure supplement 2 continued

Regions of *Nca* mRNA targeted by the *kk108825*, *hmj21533* and *jf03398* RNAi lines (termed *kk*, *hmj* and *jf* respectively) are shown as red bars. (B–D) Mean sleep levels measured using the DAM system under 12L: 12D conditions for male adult pan-neuronal *Nca* knockdown flies (B: *elav > kk*, C: *elav > jf*; D: *elav > hmj*) and associated controls (*elav*-Gal4 driver or RNAi transgene heterozygotes). $n = 17–48$. (E) Median sleep levels in the above genotypes. Night sleep is significantly reduced in all knockdown backgrounds compared to both transgene and driver alone controls. (F) qPCR verification of *Nca* knockdown by the *kk*, *hmj* and *jf* RNAi constructs. Transgene insertions lacking the *elav*-Gal4 driver were used as controls. (G) Knockdown of *Nca* had no effect on expression of *cg7646*. Expression levels of *Nca* or *cg7646* were normalised to the *ribosomal protein 49* (*rp49*) control transcript and are displayed as the ratio to the mean level of the respective RNAi alone controls ($+ > kk$, $+ > hmj$ or $+ > jf$). $n = 6–9$ for all qPCRs (2–3 independent biological repetitions of RNA extraction with triplicated qPCR reactions for each genotype). (H–I) Pan-neuronal *Nca* knockdown in adult *Drosophila* females reduces night sleep. Mean sleep patterns of *Nca*^{KD} females and associated controls in 12L: 12D conditions are shown in (H). Median night sleep levels are shown in (I). Day sleep levels are unaffected relative to heterozygous *kk* RNAi insertion controls (H). $n = 31–32$. (J) Pan-neuronal or broad *Nca* knockdown in adult males using either *nsyb*- or *insomniac* (*inc*)-Gal4 also reduced total night sleep levels in 12L: 12D conditions compared to both transgene and driver alone controls. $n = 38–53$. (K) Pan-neuronal expression of RNAi targeting *cg7646* mRNA did not alter night sleep in *Drosophila* males compared to both transgene and driver alone controls. Two different chromosomal insertions of the same RNAi hairpin were used (RNAi one and RNAi 2). $n = 12–15$. ns - $p > 0.05$, * $p < 0.05$, ** $p < 0.01$, *** $p < 0.001$, Kruskal-Wallis test with Dunn's post-hoc test (E, I, J, K) or Mann-Whitney U-test (F, G).

DOI: <https://doi.org/10.7554/eLife.38114.004>

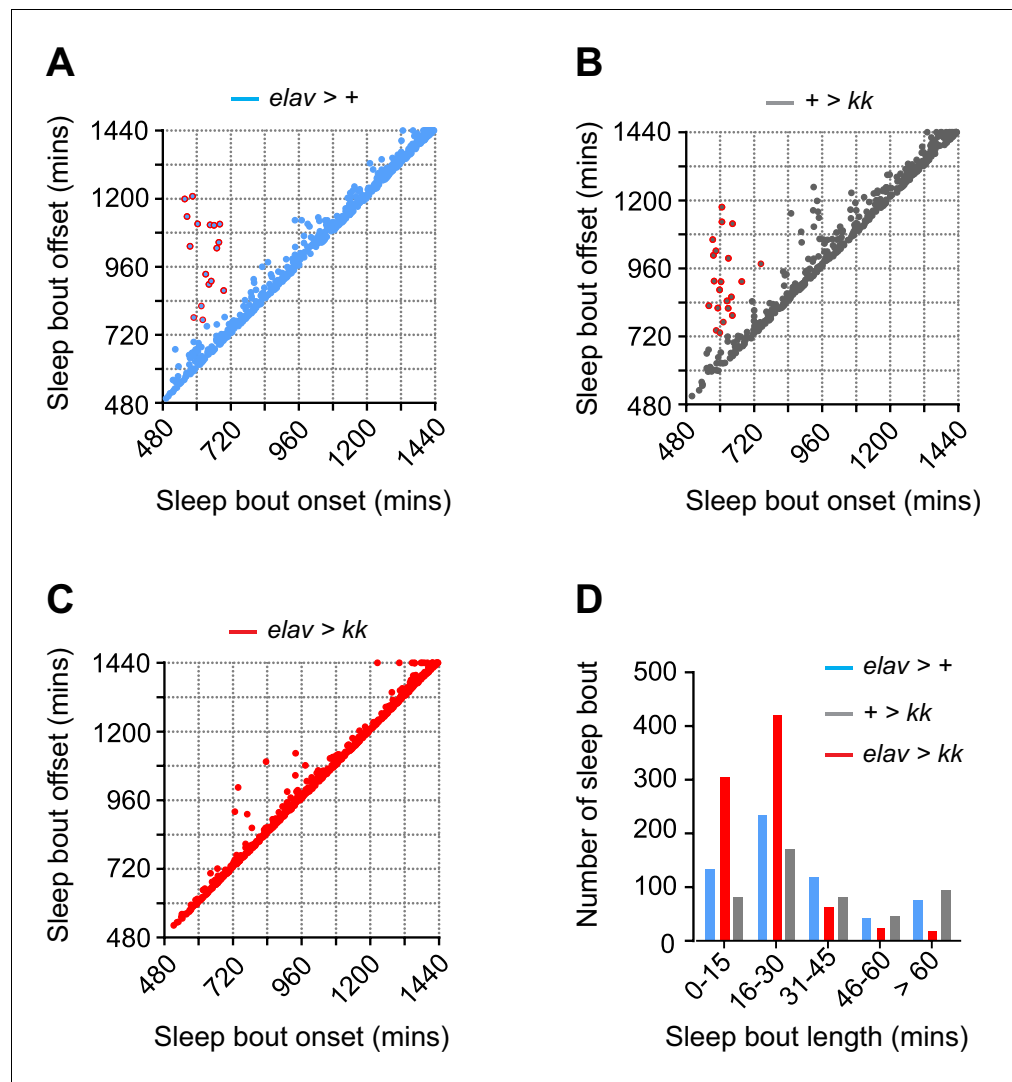


Figure 1—figure supplement 3. Reduced consolidated sleep in *Nca^{KD}* flies. (A–C) Individual sleep bout durations were measured using a custom-made R program and visualised by plotting sleep bout onset against offset for sleep bouts in control and *Nca^{KD}* adult males. In control flies (*elav > +* and *+ > kk*), longer sleep bouts initiated early during the night are highlighted in red (A, B), which are largely absent in *Nca^{KD}* adult males (C). $n = 48$ for each genotype. (D) Distribution of sleep bout lengths in *Nca^{KD}* and control adult males. Note the significant shift towards shorter sleep bout lengths in *Nca^{KD}* flies (*Nca^{KD}* vs. driver alone control, χ^2 , df : 142.0, 4, $p < 0.0001$; vs. RNAi alone control, χ^2 , df : 2112.0, 4, $p < 0.0001$).

DOI: <https://doi.org/10.7554/eLife.38114.005>

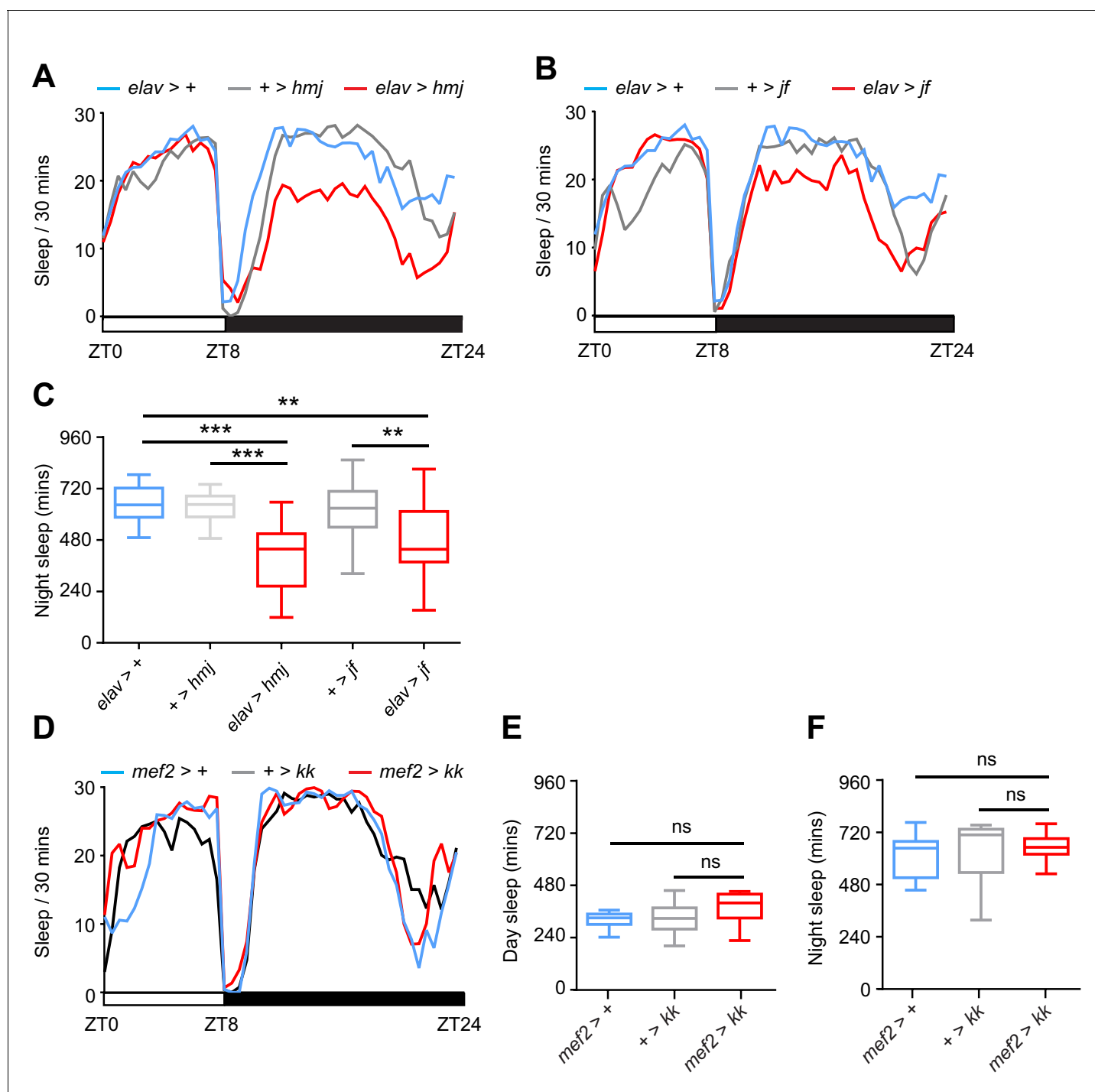


Figure 1—figure supplement 4. Pan-neuronal expression of independent *Nca* RNAi lines results in night sleep loss in 8L: 16D conditions. (A–B) Mean sleep profiles under 8L: 16D conditions for *elav*-Gal4 driven *hmj* (A) or *jf* (B) *Nca* RNAi. (C) Median night sleep amounts for genotypes shown in (A–B). *elav > +*: n = 32; *+ > hmj*: n = 26; *elav > hmj*: n = 17; *+ > jf*: n = 32; *elav > jf*: n = 32. (D–F) *Nca* knockdown in muscle cells does not affect sleep in *Drosophila*. (D) Mean sleep patterns of adult male flies with muscle-specific *Nca* knockdown via *mef2*-Gal4 (*mef2 > kk*) and associated controls under 8L: 16D. (E–F) Median day (E) and night (F) sleep levels are unaffected relative to controls. n = 16 per genotype. ns – p > 0.05, Kruskal-Wallis test with Dunn's post-hoc test. **p < 0.01, ***p < 0.001, ns – p > 0.05, Kruskal-Wallis test with Dunn's post-hoc test.

DOI: <https://doi.org/10.7554/eLife.38114.006>

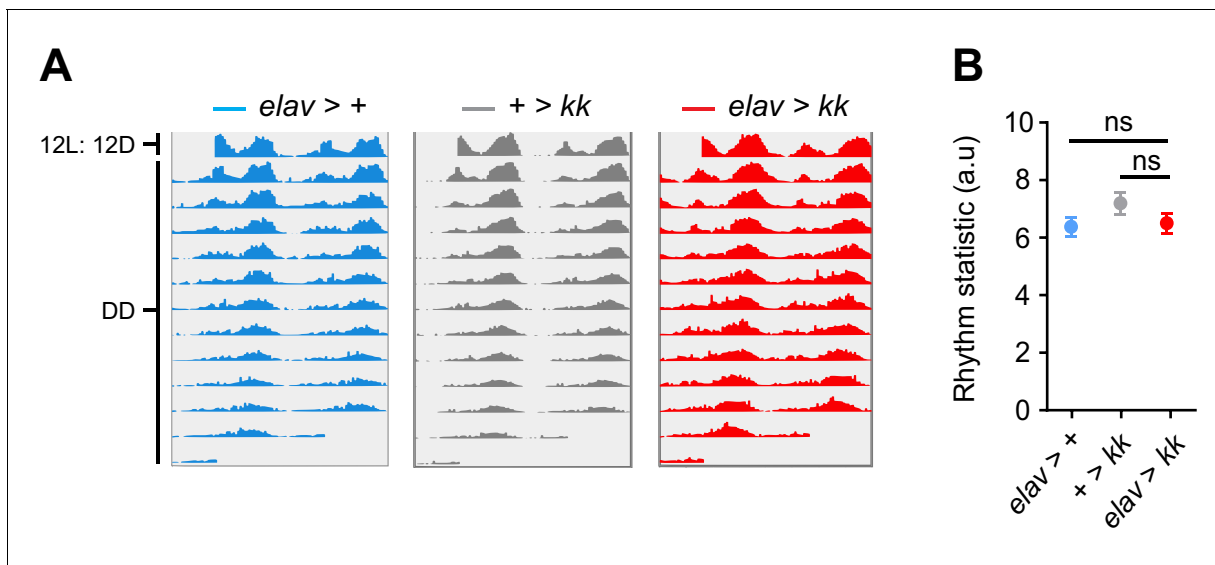


Figure 1—figure supplement 5. *Nca* knockdown does not alter circadian rhythmicity. **(A)** Actograms showing representative individual patterns of locomotor activity in one day of 12L: 12D conditions followed by 11 days of free-running activity in constant dark (DD) conditions. **(B)** Mean locomotor rhythm strength in *Nca*^{KD} adult males and controls. Robust circadian patterns of locomotor activity were still observed following in adult males expressing *Nca* RNAi (*kk*) under *elav*-Gal4 relative to controls. Error bars represent standard error of the mean. $n = 14$ –15.

DOI: <https://doi.org/10.7554/eLife.38114.007>

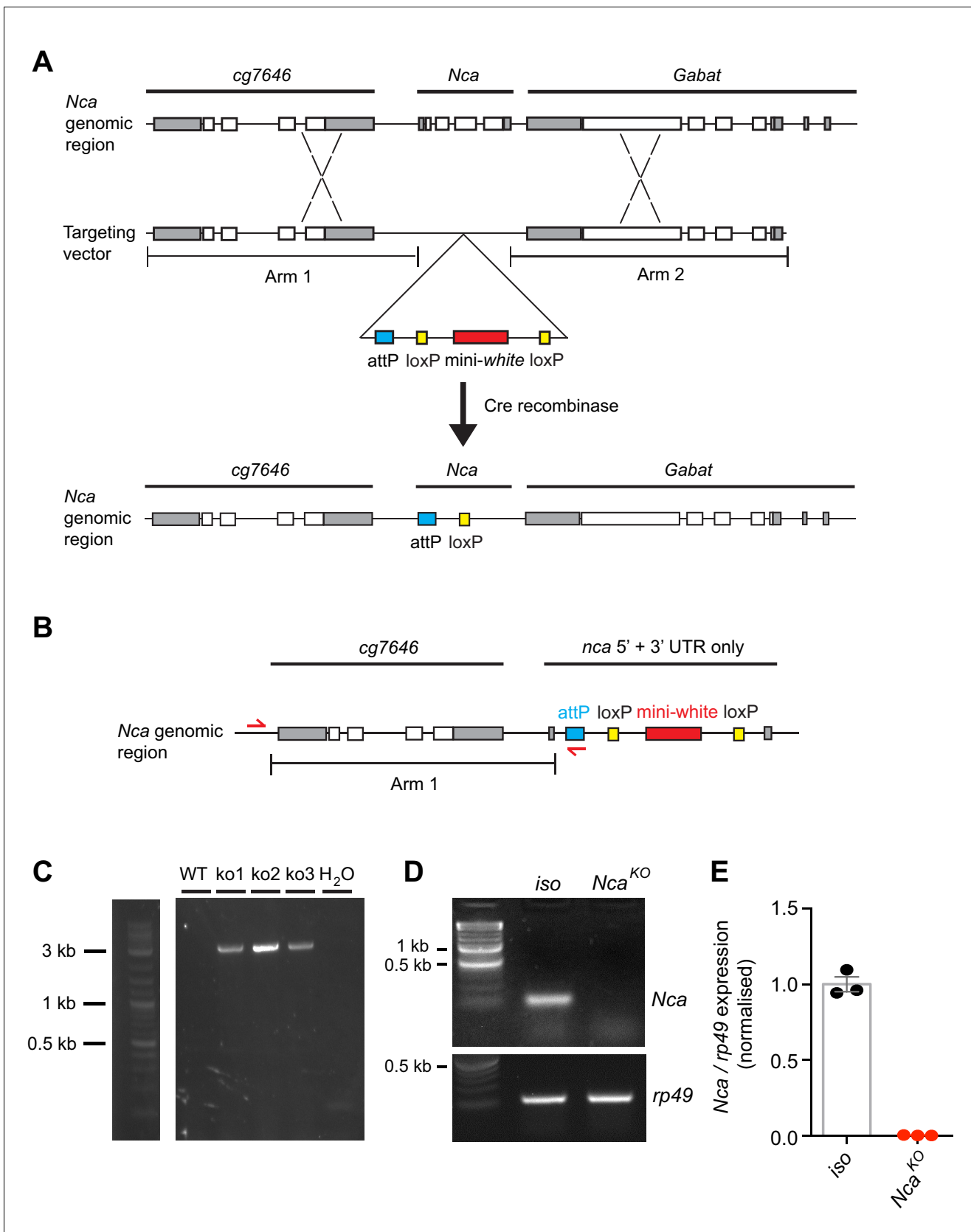


Figure 1—figure supplement 6. Generation of *Nca* null alleles using ends-out homologous recombination. (A) Schematic illustration of the procedure used to generate *Nca* knockout alleles. Homologous arms upstream (Arm 1) and downstream (Arm 2) of the *Nca* locus are indicated. Upstream 5' Figure 1—figure supplement 6 continued on next page

Figure 1—figure supplement 6 continued

promoter regions shared by the *cg7646* and *Nca* loci (see **Figure 1—figure supplement 2A**) are external to the homologous arm sequences and are not shown. Following homologous recombination, the endogenous *Nca* locus is replaced by a cassette containing the mini-*white* selection marker (red bar), and attP (blue bar) and loxP sites (yellow bars). The mini-*white* cassette was subsequently removed via Cre-loxP recombination. (**B–C**) PCR validation of homologous recombination events. Correct recombination was verified using primers designed to the attP site and upstream of the *cg7646* coding regions (**B**), which will only generate a ~ 3 kb product following homologous recombination between the targeting vector and the *Nca* locus (**C**). Three independent targeting events (ko1-3) were validated by genomic PCR. WT: wild-type genome lacking an attP site 3' of *cg7646*. (**D–E**) No *Nca* mRNA was detected in *Nca*^{KO1} using either standard RT-PCR (**D**) or quantitative RT-PCR (**E**; n = 3 qPCR reactions for *iso31* control and *Nca*^{KO1} flies).

DOI: <https://doi.org/10.7554/eLife.38114.008>

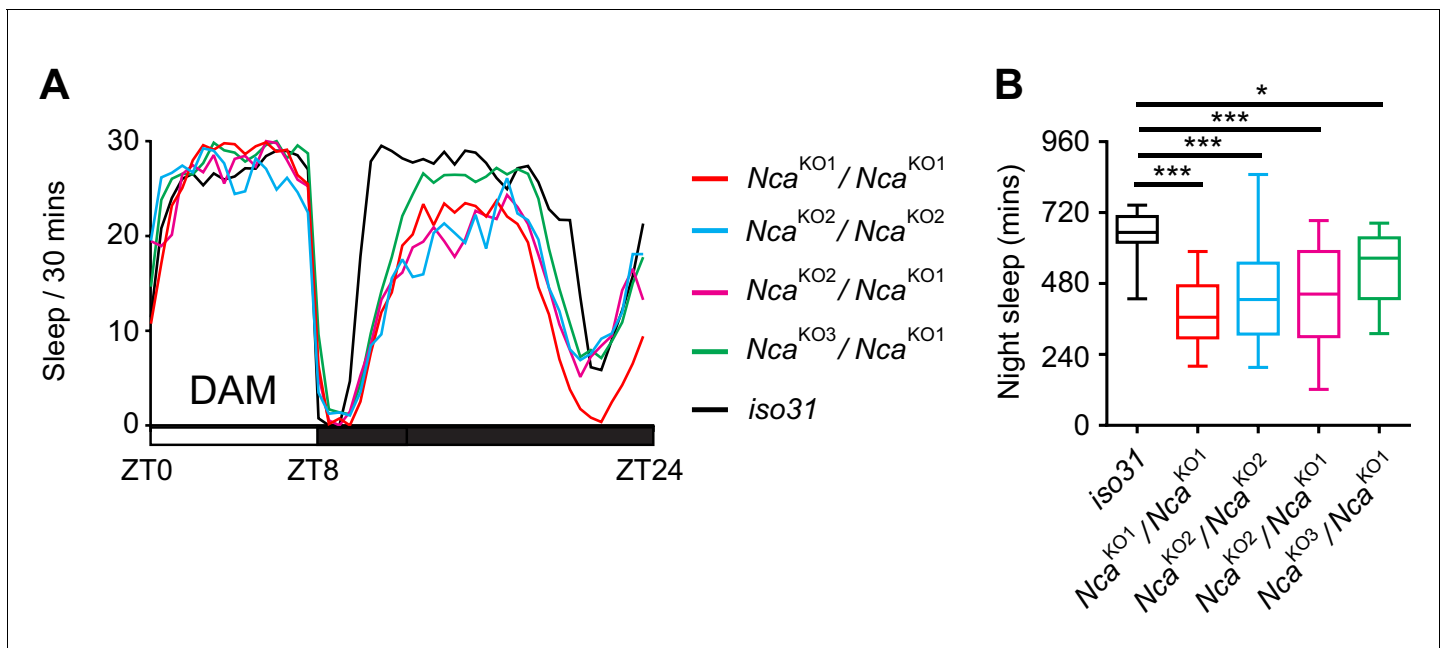


Figure 1—figure supplement 7. Independent combinations of *Nca* knockout alleles exhibit night sleep loss. (A–B) Trans-heterozygotic combinations of the *Nca*^{KO1-3} alleles, as well as homozygotes for the *Nca*^{KO2} allele, all result in significant night sleep loss compared to *iso31* controls. Night sleep levels in *Nca*^{KO1} homozygotes are also shown. (A) Mean sleep levels in the above genotypes in 8L: 16D conditions. (B) Median night sleep. n = 15–26. *p<0.05, **p<0.01, ***p<0.001 compared to *iso31* controls, Kruskal-Wallis test with Dunn’s post-hoc test.

DOI: <https://doi.org/10.7554/eLife.38114.009>

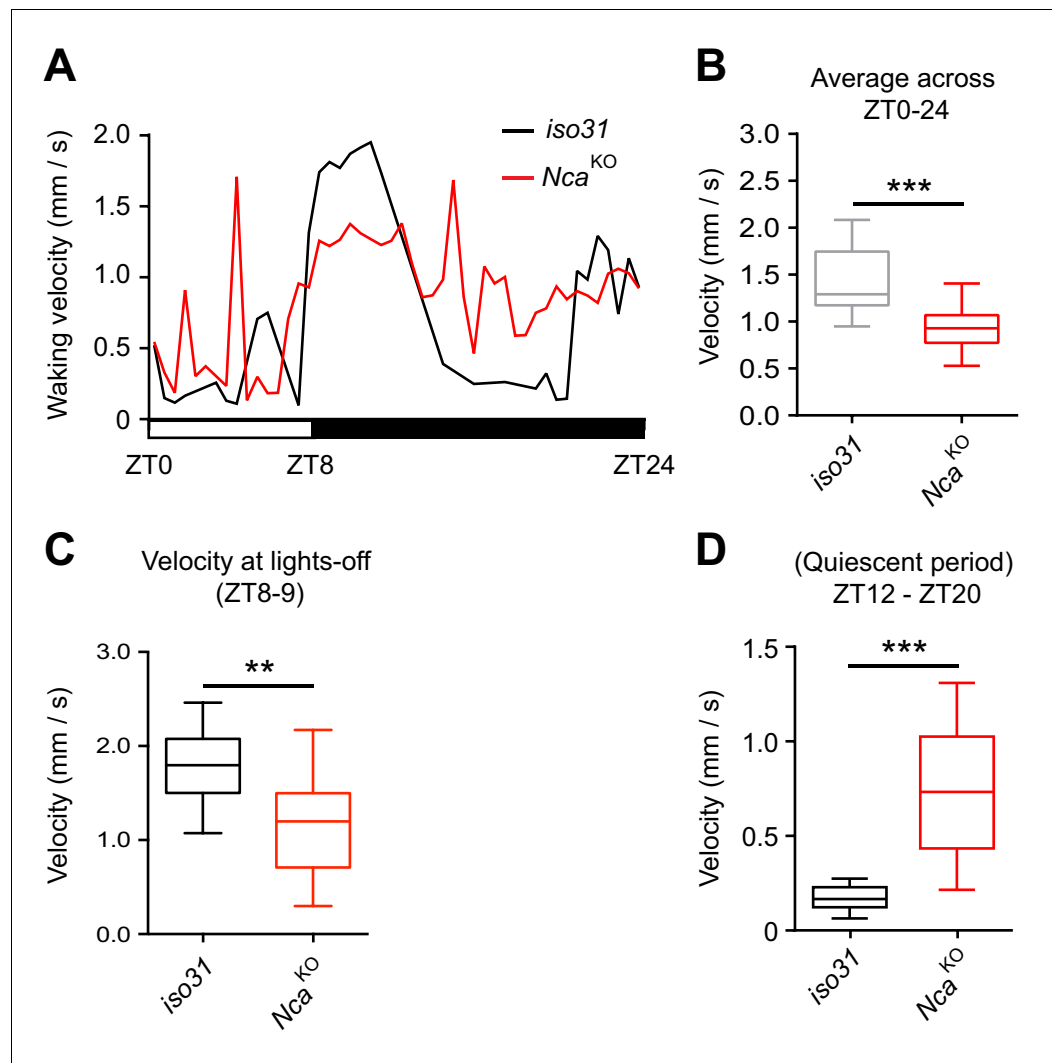


Figure 1—figure supplement 8. Locomotor velocities in *Nca* knockout flies. (A) Mean locomotor velocities across a 24 hr period in 8L: 16D conditions in *Nca^{KO}* adult males and *iso31* controls, measured via the DART system, reveals increased locomotor velocity during the night and reduced velocity during the evening activity peak *Nca^{KO}* flies. (B) Median locomotor velocities across 24 hr in 8L: 16D conditions in *Nca^{KO}* adult males and *iso31* controls. (C) Median locomotor velocity is significantly reduced in *Nca^{KO}* flies during the evening activity peak (ZT8-ZT9). (D) *Nca^{KO}* flies exhibit a significant increase in locomotor velocity between ZT12-ZT20 compared to controls – a normally quiescent period. n = 16 per genotype. **p<0.01, ***p<0.001, Mann-Whitney U-test.

DOI: <https://doi.org/10.7554/eLife.38114.010>

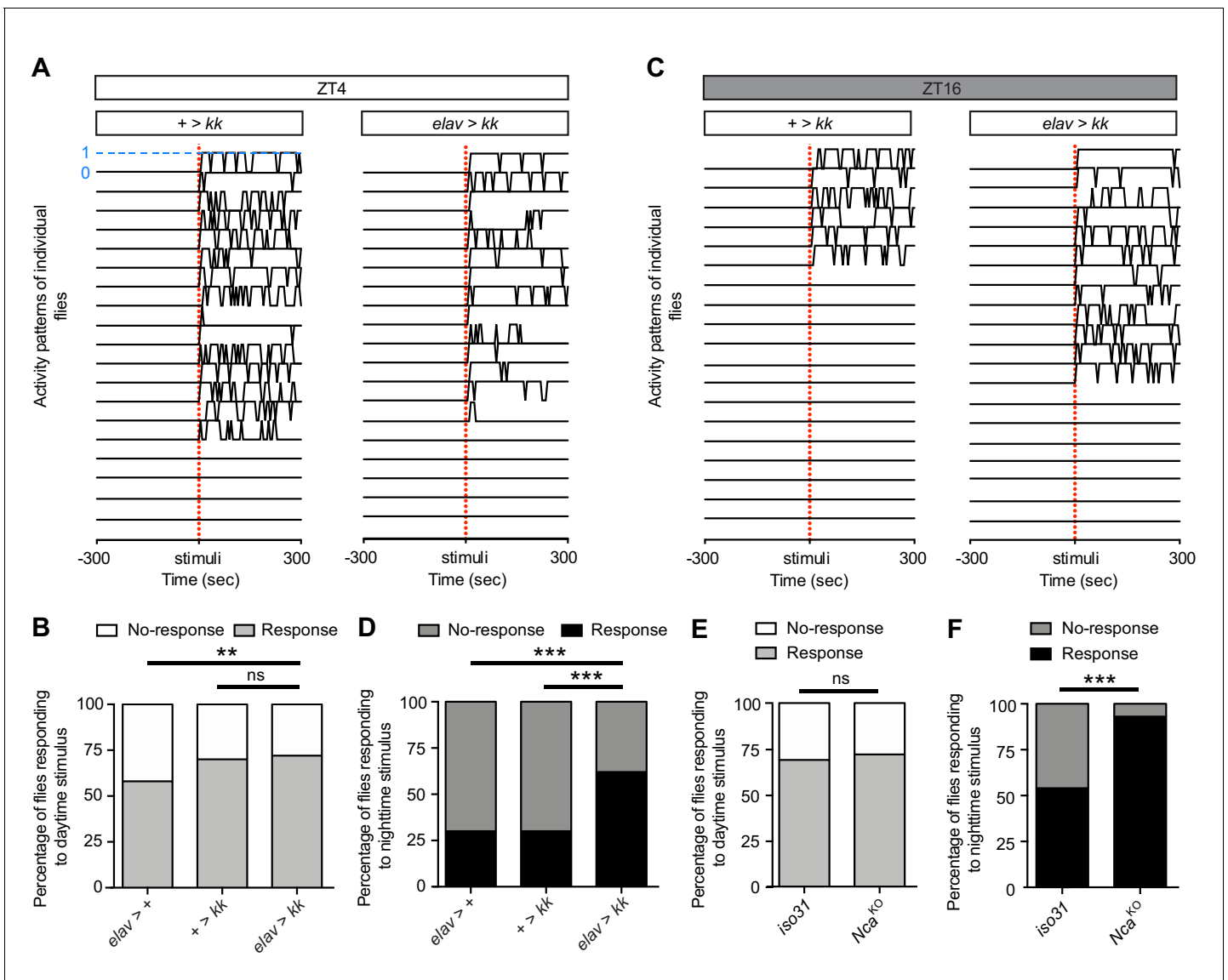


Figure 2. NCA reduces responsiveness to stimuli at night under 8L:16D conditions. (A, C) Locomotor activity in twenty representative control (+ > *kk*) and *Nca*^{KD} (*elav* > *kk*) adult male flies at either ZT4 (A) or ZT16 (C), as measured using the DART system. X-axis denotes 300 s before and after a vibration stimulus (red dotted line). Y-axis represents movement of individual flies in a binary manner (1 = movement, marked by blue dotted line for one fly; 0 = immobility). Only flies that were immobile for five mins preceding the stimulus were selected for analysis. (B, D) Percentage of *Nca*^{KD} and control flies responding or not responding to vibration stimulus at either ZT4 (B) or ZT16 (D). ZT4: *elav* > +: n = 24, + > *kk*: n = 33, *elav* > *kk*: n = 33. ZT16: *elav* > +: n = 23, + > *kk*: n = 30, *elav* > *kk*: n = 29. (E, F) Percentage of *Nca*^{KO} and *iso31* control flies responding or not responding to vibration stimulus at either ZT4 (E) or ZT16 (F). ZT4: *iso31*: n = 48, *Nca*^{KO}: n = 53. ZT16: *iso31*: n = 48, *Nca*^{KO}: n = 44. ns – p>0.05, **p<0.01, ***p<0.001, Binomial test with Bonferonni correction for multiple comparisons.

DOI: <https://doi.org/10.7554/eLife.38114.012>

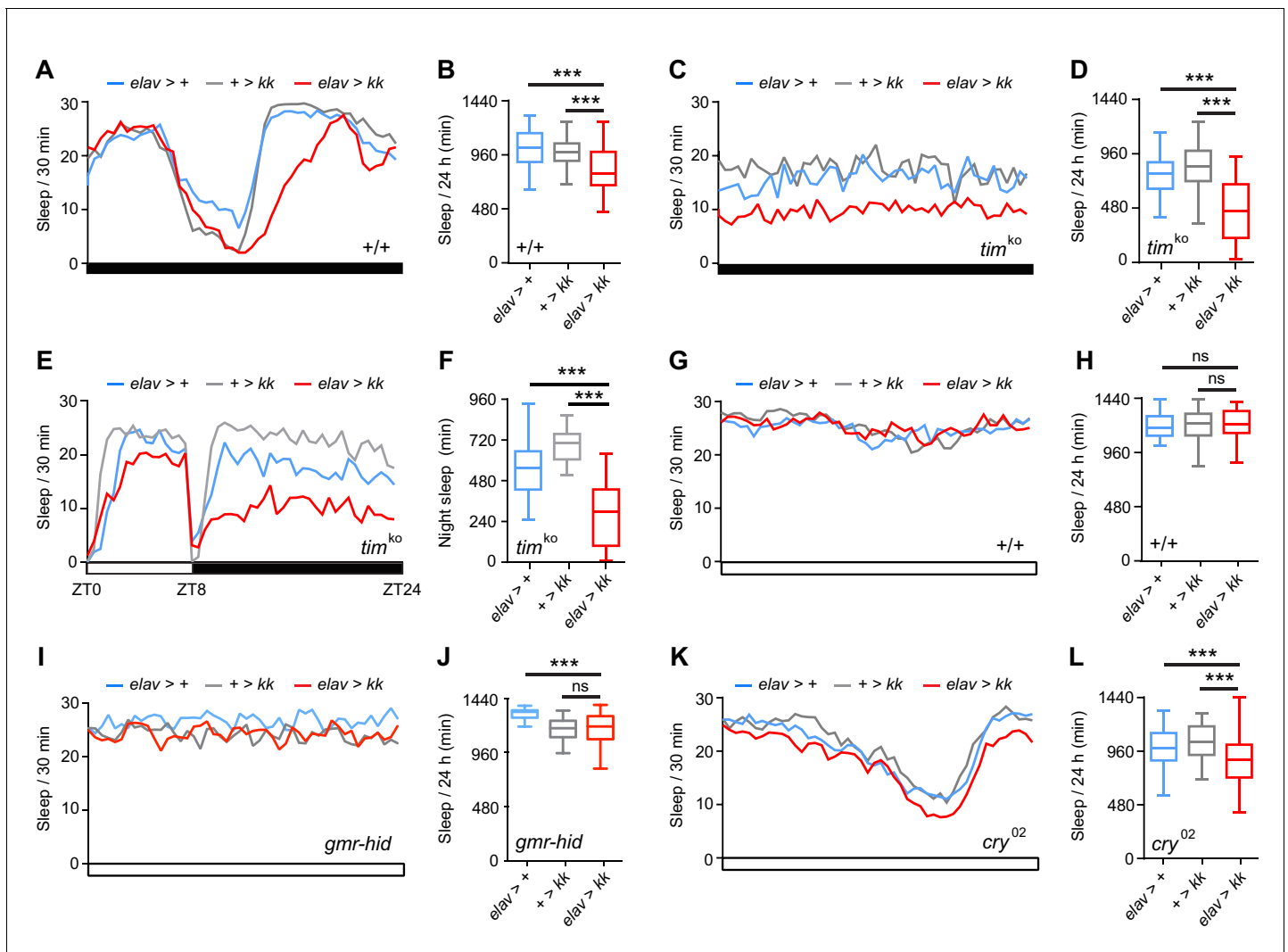


Figure 3. Circadian clock and light-sensing pathways define when NCA promotes sleep. (A–B) Mean sleep levels in *Nca^{KD}* and control adult males across 24 hr in constant-dark (DD) conditions (A), and total median sleep levels in the above genotypes (B). $n = 44\text{--}47$. Note the reduced sleep in the subjective night in *Nca^{KD}* relative to control adult males, but not the day. (C–D) Mean sleep levels in *Nca^{KD}* and control adult males across 24 hr in DD conditions in a *timeless* knockout (*tim^{KO}*) background (C), and total median sleep levels (D). $n = 32\text{--}39$. (E–F) Mean sleep levels in *Nca^{KD}* and control adult males across 24 hr in 8L: 16D conditions in a *tim^{KO}* background (E), median night sleep levels (F). $n = 22\text{--}26$. (G–H) Mean sleep levels in *Nca^{KD}* and control adult males across 24 hr in constant-light (LL) conditions (G), and total median sleep levels (H). $n = 44\text{--}47$. (I–J) Mean sleep levels in *Nca^{KD}* and control adult males across 24 hr in LL conditions in a *gmr-hid* background (I), and total median sleep levels (J). *elav > kk*, *gmr-hid/+*: $n = 51$; *+ > kk*, *gmr-hid/+*: $n = 48$; *elav > +*, *gmr-hid/+*: $n = 24$. (K–L) Mean sleep levels in *Nca^{KD}* and control adult males across 24 hr in LL conditions in a *cryptochrome* null (*cry⁰²*) background (K), and total median sleep levels in the above genotypes (L). $n = 61\text{--}72$. Note the small but consistent reduction in sleep in *Nca^{KD}*, *cry⁰²* males (K), leading to a significant decrease in total median sleep levels relative to controls (L). ns - $p > 0.05$, *** $p < 0.001$, as compared to driver and RNAi alone controls via Kruskal-Wallis test with Dunn’s post-hoc test.

DOI: <https://doi.org/10.7554/eLife.38114.014>

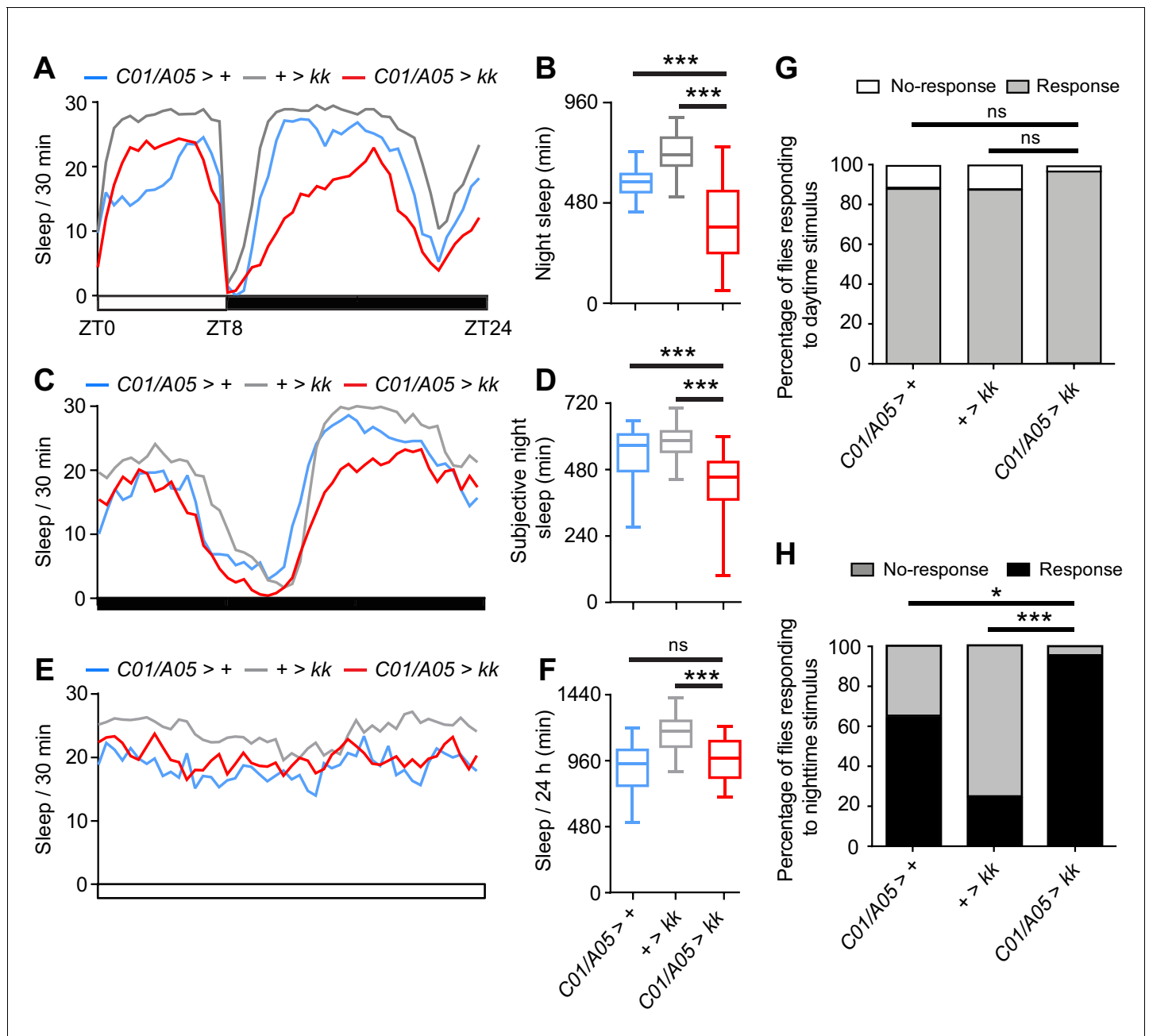


Figure 4. NCA acts in two distinct neural subpopulations to regulate night sleep. (A–F) Sleep patterns in adult male flies with *Nca* knockdown (using the *kk* *Nca* RNAi) in two neural domains defined by the *A05*- and *C01*-Gal4 drivers in varying light/dark regimes, compared to controls. (A–B) A: mean sleep patterns in 8L: 16D conditions. B: median night sleep in *Nca* knockdown flies compared to heterozygote drivers and transgene alone controls. *+ > kk*: n = 80; *C01/A05 > +*: n = 42; *C01/A05 > kk*: n = 71. (C–D) Mean sleep patterns (C) and median subjective night sleep (D) in constant dark (DD) conditions. *+ > kk*: n = 64; *C01/A05 > +*: n = 47; *C01/A05 > kk*: n = 51. (E–F) Mean sleep patterns (E) and median total sleep (F) in constant light (LL) conditions. *+ > kk*: n = 76; *C01/A05 > +*: n = 26; *C01/A05 > kk*: n = 28. (G–H) Percentage of *C01/A05 > kk* and control flies responding or not responding to vibration stimuli at either ZT4 (G; *C01/A05 > kk*, n = 38, *+ > kk*, n = 61 and *C01/A05 > +*, n = 26) or ZT16 (H; *C01/A05 > kk*, n = 24, *+ > kk*, n = 54 and *C01/A05 > +*, n = 28) under 8L: 16D conditions. ns = $p > 0.05$, * $p < 0.05$, ** $p < 0.01$, *** $p < 0.001$, compared to driver and RNAi alone controls, Kruskal-Wallis test with Dunn’s post-hoc test (B, D, F) or Binomial test with Bonferonni correction for multiple comparisons (G–H).

DOI: <https://doi.org/10.7554/eLife.38114.016>

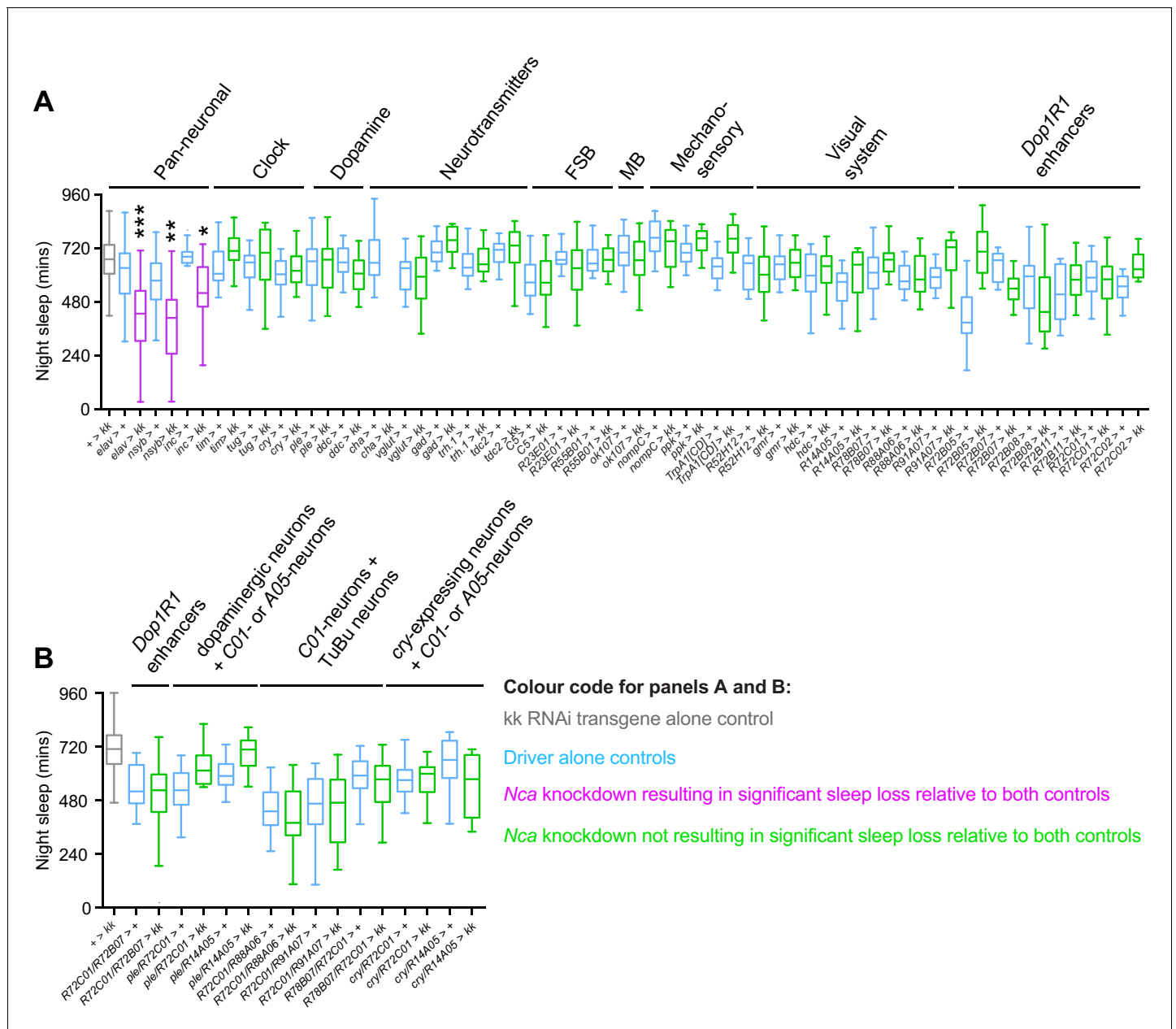


Figure 4—figure supplement 1. Transgenic RNAi-based mini-screen to identify key NCA-expressing neurons. (A) *Nca* knockdown with broadly expressed drivers resulted in reduced night sleep in adult males under 8L: 16D conditions. In contrast, *Nca* knockdown in previously defined sleep-regulatory centers, clock neurons, the visual system or subsets of *Dop1R1*-expressing neurons did not impact night sleep. FSB: fan-shaped body. MB: mushroom body. Grey and blue box plots: control lines. Magenta box plots: experimental lines showing reduced night sleep relative to controls. Green box plots: experimental lines failing to show reduced night sleep relative to one or both controls. Grey box plot: *kk* *Nca* RNAi alone (+ > *kk*) controls. Blue box plots: Gal4 driver heterozygotes. (B) *Nca* knockdown using combinations of driver lines labelling *C01*- and *A05*-neurons in addition to neurons in the dopaminergic pathway (dopamine-release and *Dop1R1*-expressing neurons), the anterior visual pathway (tubercular-bulbar (TuBu) neurons), and *cryptochrome* (*cry*)-expressing neurons. See **Figure 4** Source Data for n-values and additional statistical comparisons. * $p < 0.05$, ** $p < 0.01$, *** $p < 0.001$, as compared to driver and RNAi alone controls via Kruskal-Wallis test with Dunn's post-hoc test.

DOI: <https://doi.org/10.7554/eLife.38114.017>

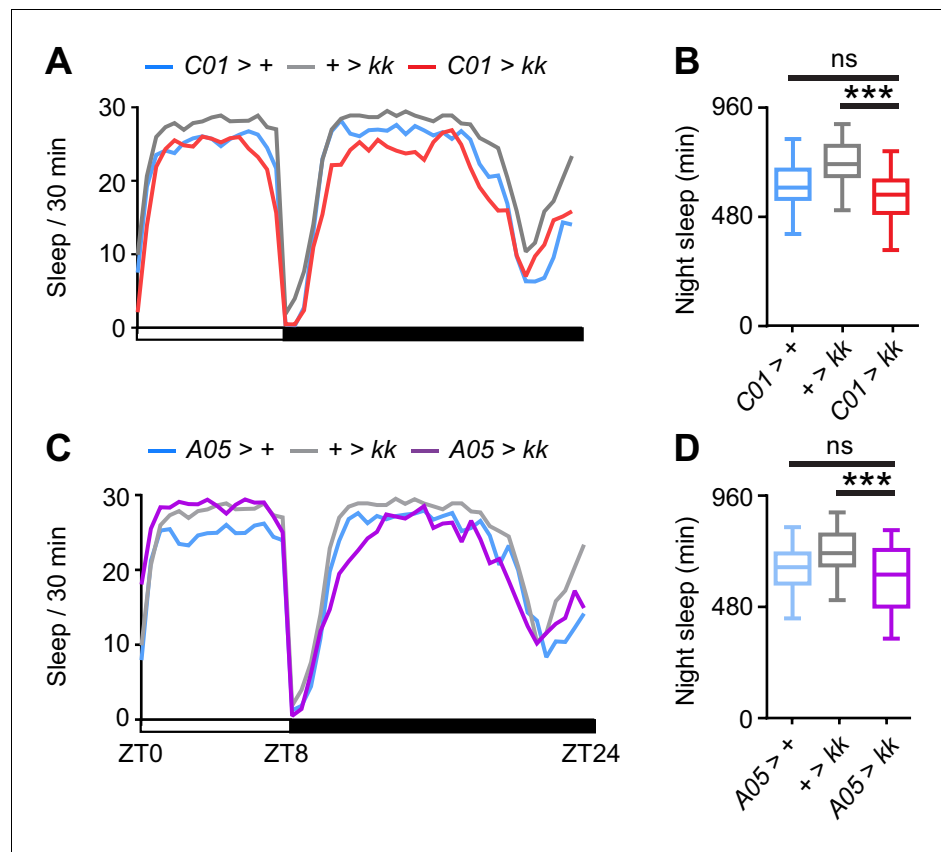


Figure 4—figure supplement 2. *Nca* knockdown in *C01*- or *A05*-neurons alone does not significantly alter sleep. (A–B) Mean sleep patterns (A) and median night sleep (B) of adult males with expressing *Nca* RNAi (*kk*) in *C01*-neurons compared to heterozygote driver and transgene alone controls in 8L: 16D conditions. + > *kk*: $n = 80$, *C01* > +: $n = 64$, *C01* > *kk*: $n = 80$. (C–D) Mean sleep patterns (A) and median night sleep (B) of adult males with expressing *Nca* RNAi (*kk*) in *A05*-neurons compared to controls in 8L: 16D conditions. + > *kk*: $n = 80$, *A05* > +: $n = 31$, *A05* > *kk*: $n = 31$. Note that the same population of + > *kk* control males was used in **Figure 4A–B**, as the combined *C01*- and *A05*-Gal4 experiments were performed in parallel. ns – $p > 0.05$, *** $p < 0.001$ compared to driver and RNAi alone controls, Kruskal-Wallis test with Dunn’s post-hoc test.

DOI: <https://doi.org/10.7554/eLife.38114.018>

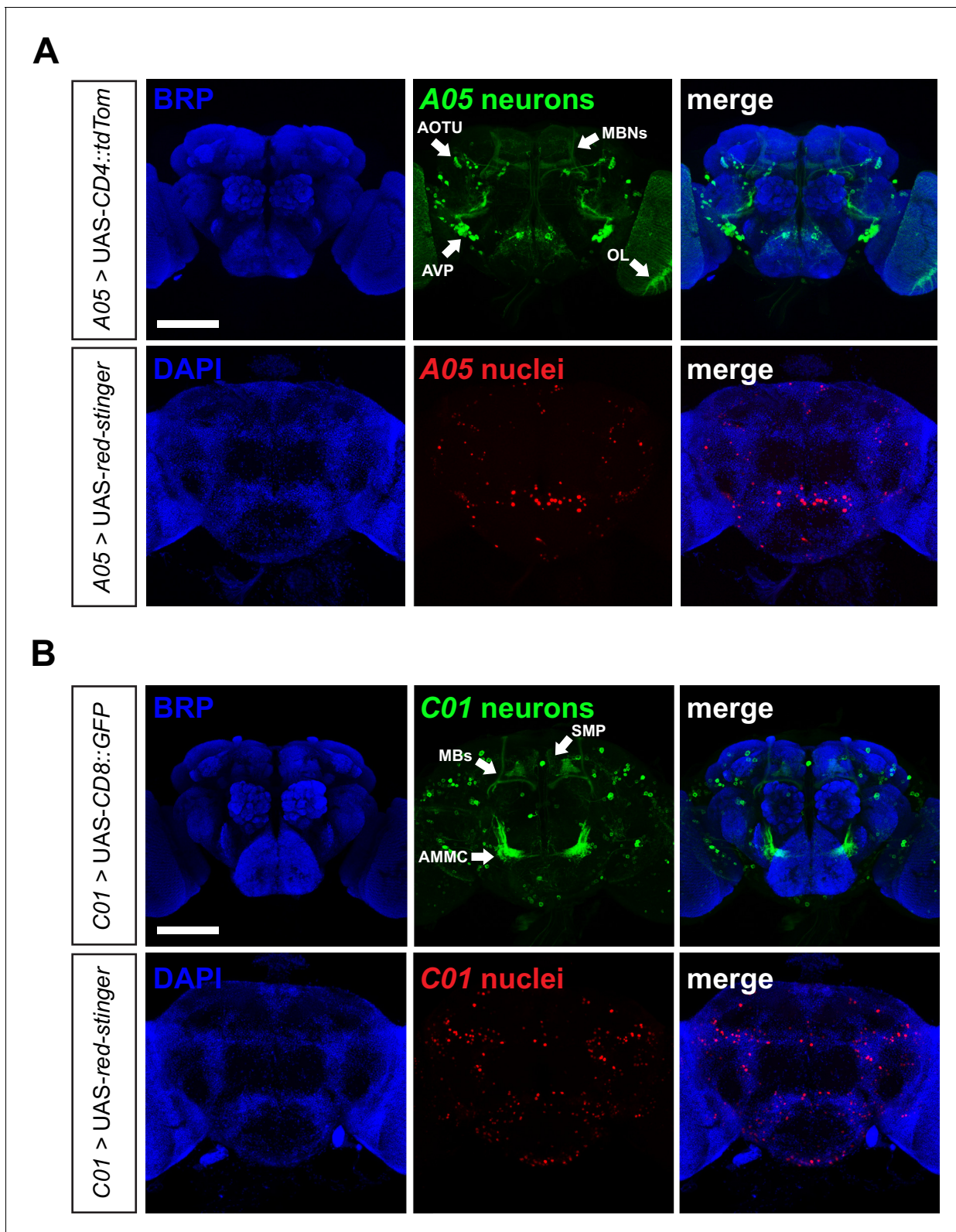


Figure 5. Distribution of A05- and C01-neurons in the adult *Drosophila* brain. (A–B) Confocal z-stacks of adult male brains expressing genetically-encoded fluorophores labelling either neuronal processes (CD4::TdTom or CD8::GFP) or nuclei (Red-stinger) under the A05- (A) or C01-Gal4 (B) drivers. Neuropil regions are labelled with anti-Bruchpilot (BRP). Nuclei are co-labelled with DAPI. Scale bars, 100 μ m. Arrows point to neuropil centers. AOTU: Figure 5 continued on next page

Figure 5 continued

anterior optic tubercle. MBNs: mushroom body neurons. OL: optic lobe. AMMC: antennal mechanosensory and motor center. AVP: anterior ventrolateral protocerebrum. SMP: superior medial protocerebrum.

DOI: <https://doi.org/10.7554/eLife.38114.020>

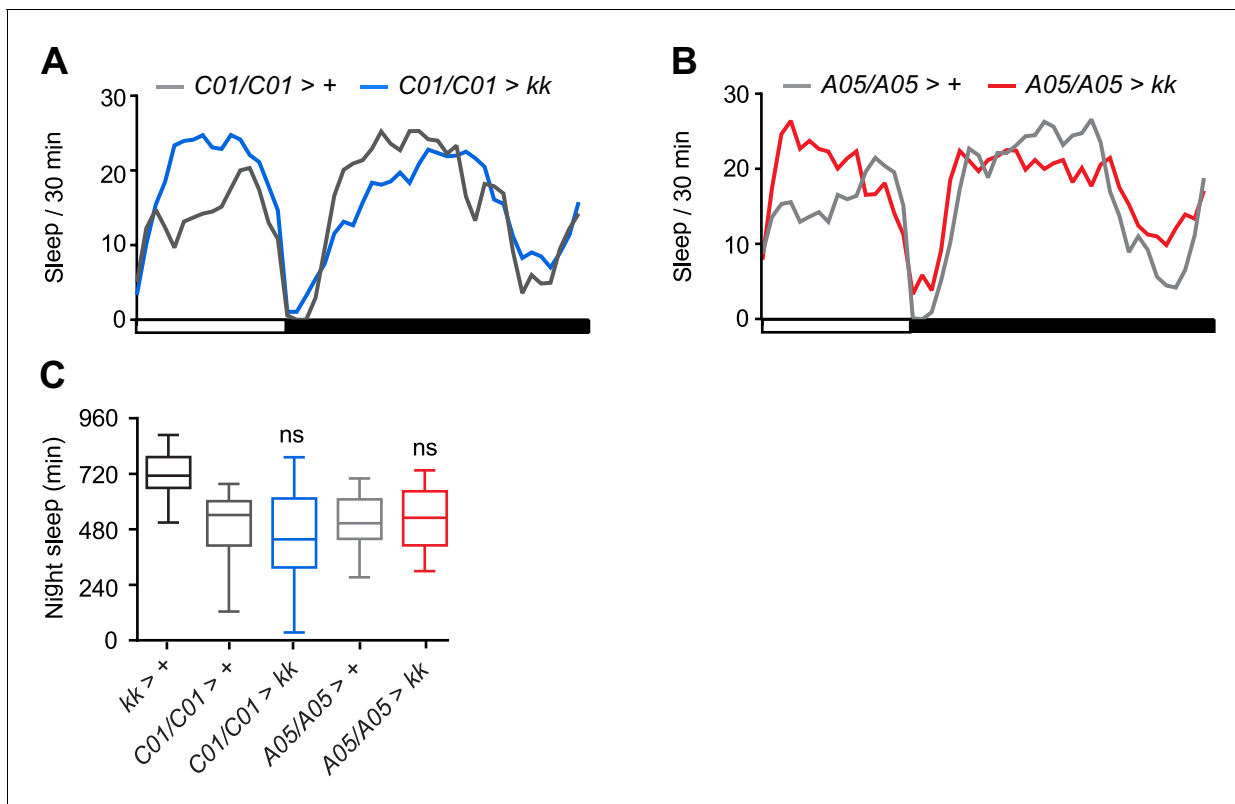


Figure 5—figure supplement 1. *Nca* knockdown using homozygous *C01*- or *A05*-Gal4 drivers does not affect night sleep. (A) Mean sleep patterns of adult males homozygous for the *C01*-Gal4 driver with and without the *kk Nca* RNAi insertion. (B) Mean sleep patterns of adult males homozygous for the *A05*-Gal4 driver with and without the *kk Nca* RNAi insertion. (C) Median night sleep levels for heterozygous RNAi transgene and homozygous driver controls, and males expressing *Nca* RNAi with two Gal4 driver copies. No night sleep loss was observed using two copies of either driver relative to controls. + > *kk*: $n = 80$ (the same population was used in **Figure 4A–B**, as the experiments were performed in parallel), $C01/C01 > +$: $n = 24$, $C01/C01 > kk$: $n = 39$; $A05/A05 > +$: $n = 22$, $A05/A05 > kk$: $n = 23$. ns – $p > 0.05$, Kruskal-Wallis test with Dunn’s post-hoc test.

DOI: <https://doi.org/10.7554/eLife.38114.021>

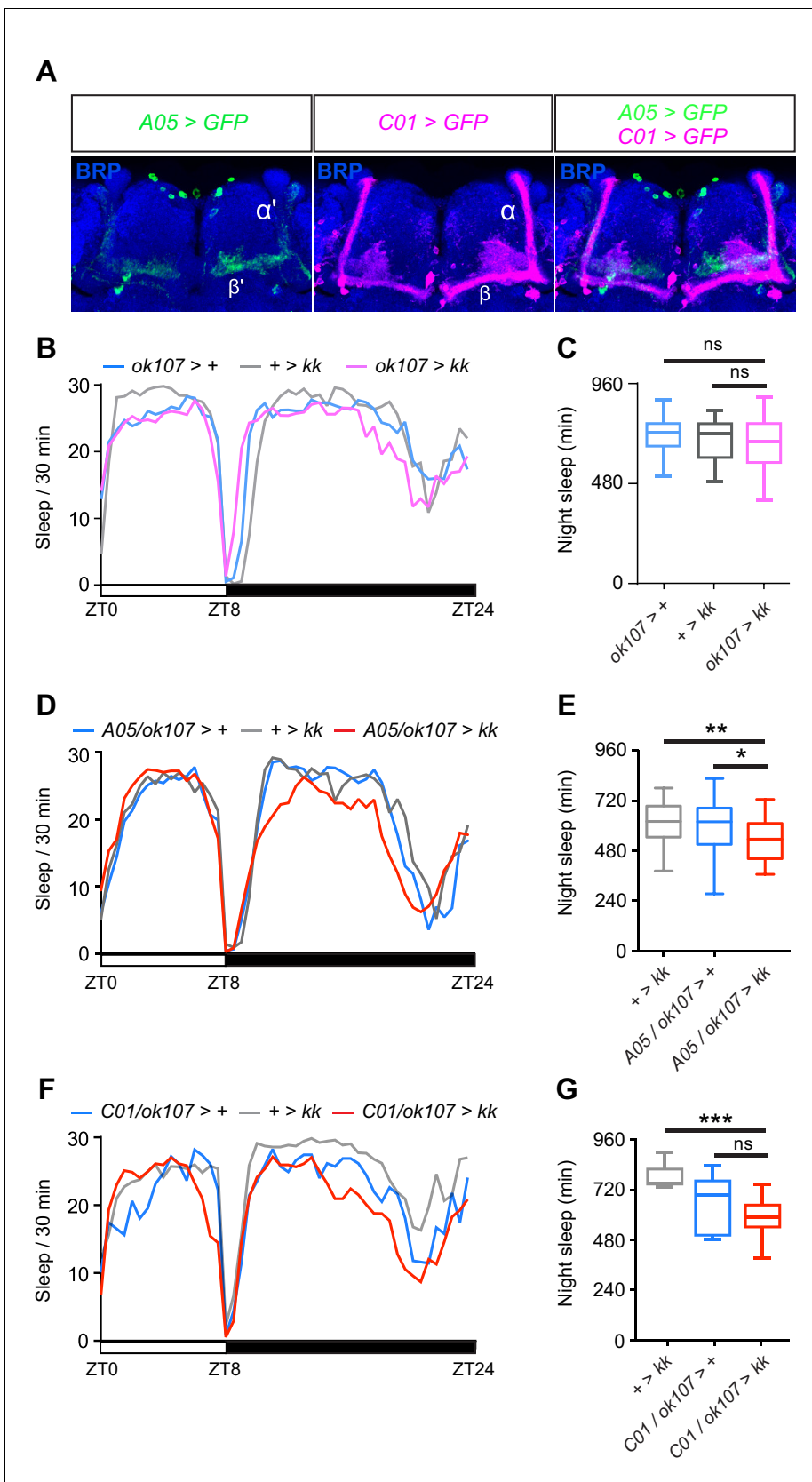


Figure 5—figure supplement 2. The mushroom bodies are a sleep-relevant subdomain within *C01*-neurons. (A) Standardised confocal stacks labelling *R14A05* (*A05 > GFP*, green) and *R72C01* (*C01 > GFP*, magenta) positive neurons. Images from *Jenett et al. (2012)*, deposited at the Virtual Fly Brain, *Figure 5—figure supplement 2 continued on next page*

Figure 5—figure supplement 2 continued

www.virtualflybrain.org), were downloaded and digitally superimposed (right panel) onto the universal fly brain (the image source data is distributed under a CC BY-NC-SA 4.0 license). BRP, Bruchpilot. (B–C) Mean sleep pattern (B) and median night sleep levels (C) in adult male flies expressing *Nca* RNAi (*kk*) in MB-KCs using *ok107-gal4* in 8L: 16D. *ok107 > +*, *n* = 55, *+ > kk*, *n* = 47, *ok107 > kk*, *n* = 65. (D–E) *Nca* knockdown in both A05 and MB-KC neurons results in reduced night sleep. Mean sleep patterns in 8L: 16D are shown in (D), median night sleep levels are shown in (E). *A05/ok107 > +*: *n* = 33, *+ > kk*: *n* = 31, *A05/ok107 > kk*: *n* = 42. (F–G) Knocking down *Nca* in MB-KC and the *C01*-neurons does not result in significant night sleep loss in 8L: 16D. Mean sleep patterns are shown in (F), median night sleep levels are shown in (G). *C01/ok107 > +*: *n* = 11, *+ > kk*: *n* = 15, *C01/ok107 > kk*: *n* = 21. ns – *p* > 0.05, **p* < 0.05, ***p* < 0.01, ****p* < 0.001, Kruskal-Wallis test with Dunn’s post-hoc test.

DOI: <https://doi.org/10.7554/eLife.38114.022>

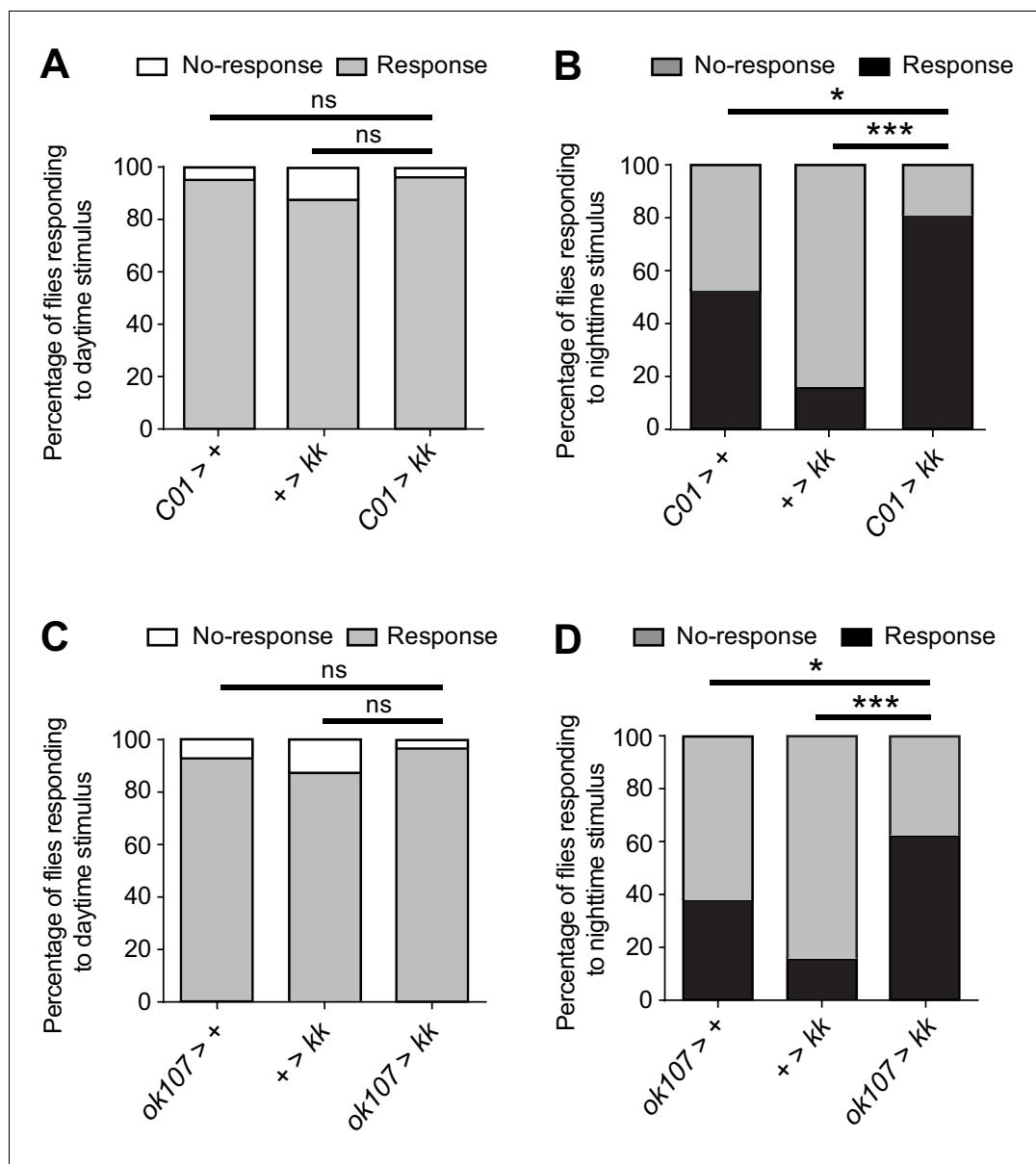


Figure 6. NCA acts in the mushroom bodies to regulate nocturnal arousal. (A–B) Percentage of adult male flies expressing *Nca* RNAi (*kk*) in *C01*-neurons (*C01 > kk*) and control flies responding or not responding to vibration stimulus at either ZT4 (day; A) or ZT16 (night; B). ZT4: *C01 > +*, *n* = 22, *+ > kk*, *n* = 61, *C01 > kk*, *n* = 27. ZT16: *C01 > +*, *n* = 19, *+ > kk*, *n* = 54, *C01 > kk*, *n* = 21. (C–D) Percentage of adult male flies expressing *Nca* RNAi (*kk*) in MB-KCs (*ok107 > kk*) and control flies responding or not responding to vibration stimulus at either ZT4 (day; C) or ZT16 (night; D). ZT4: *ok107 > +*, *n* = 26, *+ > kk*, *n* = 47, *ok107 > kk*, *n* = 28. ZT16: *ok107 > +*, *n* = 26, *+ > kk*, *n* = 44, *ok107 > kk*, *n* = 27. ns – *p* > 0.05, **p* < 0.05, ****p* < 0.001, Binomial test with Bonferonni correction for multiple comparisons.

DOI: <https://doi.org/10.7554/eLife.38114.024>

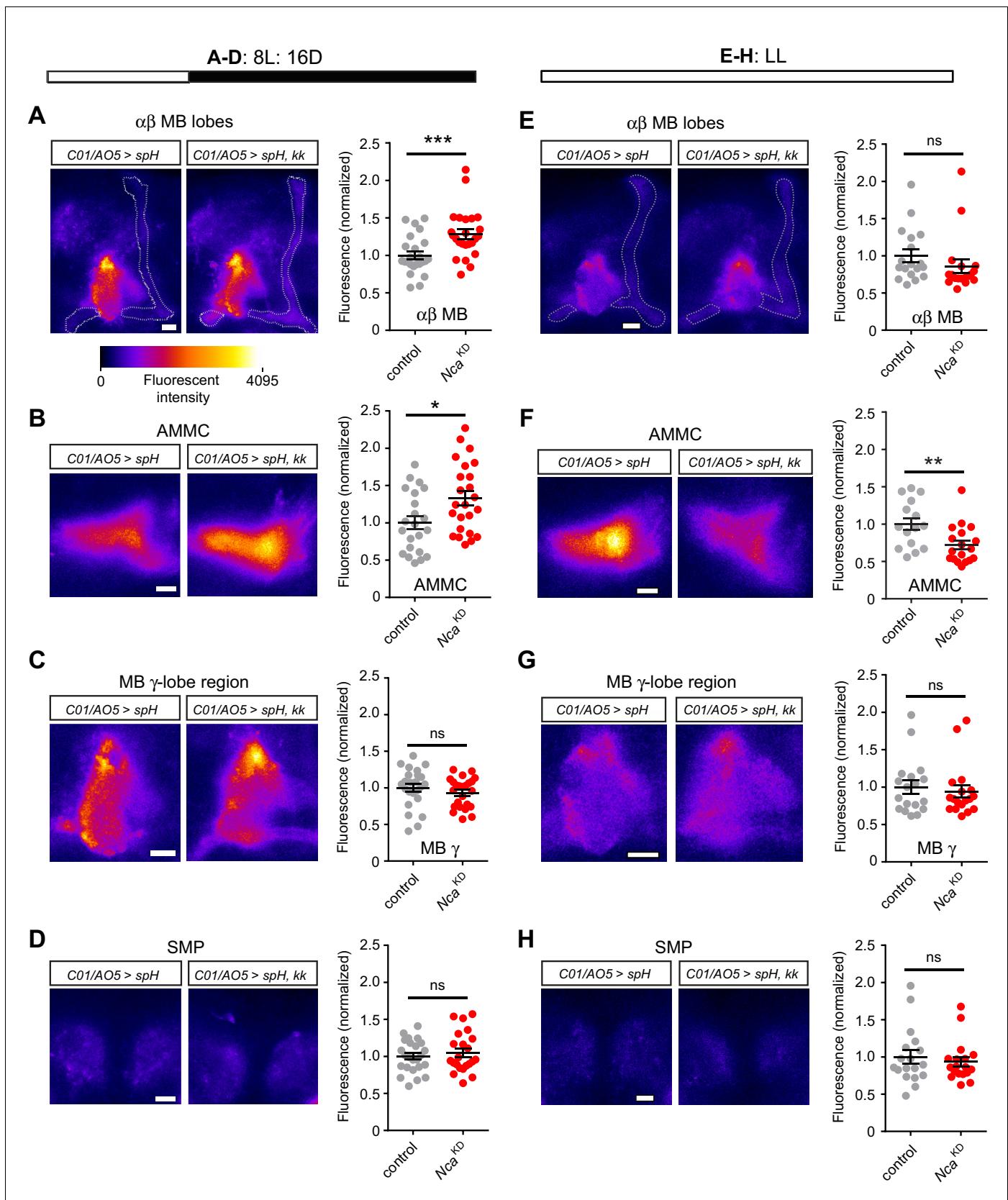


Figure 7. NCA suppresses synaptic release in subsets of *C01/A05*-neurons during darkness. (A–D) Fluorescence of an optical reporter of synaptic release (synapto-pHluorin, spH) in neuropil regions labelled by the *C01*- and *A05*-drivers, in control adult males (*C01/A05 > spH*) or following *Nca*

Figure 7 continued on next page

Figure 7 continued

knockdown in *C01*- and *A05*-neurons (*C01/A05 > spH, kk*). Flies were housed under 8L: 16D conditions, in which *Nca* knockdown in *C01*- and *A05*-neurons causes robust nighttime sleep loss. **(E–G)** spH fluorescence in control adult males or following *Nca* knockdown in *C01*- and *A05*-neurons (*C01/A05 > spH, kk*). Flies were housed in LL conditions, in which *Nca* knockdown in *C01*- and *A05*-neurons has no effect on sleep levels. In each panel, representative confocal images of spH fluorescence (left) and mean fluorescent intensity (right, normalized to the mean of *C01/A05 > spH* controls) are shown. Dots within dot plots represent individual brain hemisphere measurements. A–D: n = 22–24. E–H: n = 15–18. Neuropil regions are noted. MB: mushroom body. AMMC: antennal mechanosensory motor center. SMP: superior medial protocerebrum. ns – p>0.05, *p<0.05, **p<0.01, ***p<0.001, Mann-Whitney U-test.

DOI: <https://doi.org/10.7554/eLife.38114.026>

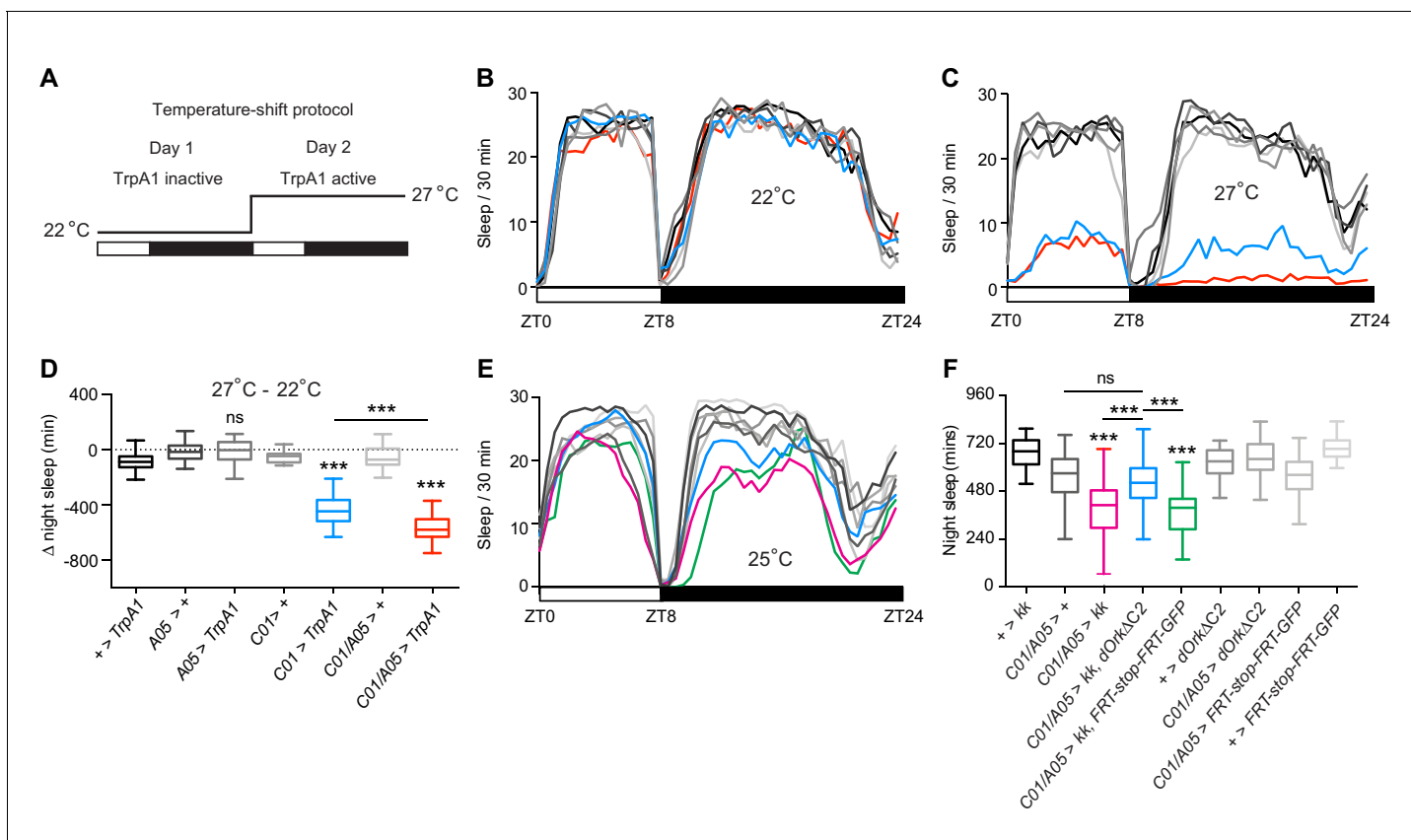


Figure 8. Sleep loss in *Nca* knockdown flies is caused by enhanced excitability of *C01/A05*-neurons. (A) Experimental paradigm for acute activation of *A05* or *C01*-neurons. 22°C: non-activating temperature for *TrpA1*. 27°C: activating temperature. Sleep levels were recorded over two days in 8L: 16D conditions. (B–C) Mean sleep levels across 8L: 16D following expression of *TrpA1* in *A05*-, *C01*- or *A05*- and *C01*-neurons (and associated controls) at 22°C (B) or 27°C (C). (D) Median change in night sleep levels (Δ night sleep) following the shift from 22°C on day 1 to 27°C on day 2. +> *TrpA1*: $n = 53$, *A05* > +: $n = 23$, *A05* > *TrpA1*: $n = 68$, *C01* > +: $n = 24$, *C01* > *TrpA1*: $n = 40$, *C01/A05* > +: $n = 33$, *C01/A05* > *TrpA1*: $n = 40$. ns – $p > 0.05$, *** $p < 0.001$, as compared to *TrpA1* or driver alone controls by Kruskal-Wallis test with Dunn’s post-hoc test (for *C01* > *TrpA1*, *A05* > *TrpA1*, or *C01/A05* > *TrpA1* compared to controls) or Mann-Whitney U-test (for *C01/A05* > *TrpA1* compared to *C01* > *TrpA1*). (E–F) Inhibition of *C01/A05*-neurons by expressing *dORK Δ C2* rescues sleep loss due to *Nca* knockdown, while expression of *dORK Δ C2* does not change baseline sleep. Mean sleep patterns in 8L: 16D conditions are shown in (E). Median night sleep levels are shown in (F). +> *kk*: $n = 72$, *C01/A05* > +: $n = 85$, *C01/A05* > *kk*: $n = 95$, *C01/A05* > *dORK Δ C2*, *kk*: $n = 77$, *C01/A05* > *kk*, *FRT-stop-FRT-GFP*: $n = 39$, +> *dORK Δ C2*: $n = 57$, *C01/A05* > *dORK Δ C2*: $n = 73$, *C01/A05* > *FRT-stop-FRT-GFP*: $n = 49$, +> *FRT-stop-FRT-GFP*: $n = 36$. ns – $p > 0.05$, *** $p < 0.001$, Kruskal-Wallis test with Dunn’s post-hoc test.

DOI: <https://doi.org/10.7554/eLife.38114.028>