

The effect of the DISC1 Ser704Cys polymorphism on striatal dopamine synthesis capacity: an [¹⁸F]-DOPA PET study

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Abstract

Whilst the role of the Disrupted-in-Schizophrenia 1 (*DISC1*) gene in the aetiology of major mental illnesses is debated, the characterisation of its function lends it credibility as a candidate. A key aspect of this functional characterisation is the determination of the role of common non-synonymous polymorphisms on normal variation within these functions. The common allele (A) of the *DISC1* SNP rs821616 encodes a serine at the Ser704Cys polymorphism, and has been shown to increase the phosphorylation of extracellular signal-regulated protein Kinases 1 and 2 (ERK1/2) which stimulate the phosphorylation of tyrosine hydroxylase, the rate-limiting enzyme for dopamine biosynthesis. We therefore set out to test the hypothesis that human serine (A) homozygotes would show elevated dopamine synthesis capacity compared to cysteine homozygotes and heterozygotes (TT and AT) for rs821616. [¹⁸F]-DOPA PET was used to index striatal dopamine synthesis capacity as the influx rate constant K_i^{cer} in healthy volunteers *DISC1* rs821616 serine homozygotes (N=46) and healthy volunteers *DISC1* rs821616 cysteine homozygotes and heterozygotes (N=56), matched for age, gender, ethnicity and using three scanners. We found *DISC1* rs821616 serine homozygotes exhibited a significantly higher striatal K_i^{cer} compared to cysteine homozygotes and heterozygotes (p=0.012) explaining 6.4% of the variance (partial eta squared=0.064). Our finding is consistent with its previous association with heightened activation of ERK1/2, which stimulates tyrosine hydroxylase activity for dopamine synthesis. This could be a potential mechanism mediating risk for psychosis, lending further credibility to the fact that *DISC1* is of functional interest in the aetiology of major mental illness.

Introduction

The dopamine hypothesis has been a leading theory underlying the neurobiology of schizophrenia for the last four decades (1, 2). The hypothesis was initially based on evidence showing that antipsychotic medications block dopamine receptors (3-5) and that drugs increasing dopamine levels elicit psychotic symptoms in healthy people (6-8) and people with schizophrenia (9, 10). Using [¹⁸F] fluoro-3,4-dihydroxyphenyl-L-alanine (F-DOPA) Positron Emission Tomography (PET), increased presynaptic dopamine synthesis capacity has been found in schizophrenia (11), people with prodromal psychotic symptoms (12, 13) and those with clinical progression to psychosis (14). Whilst a substantial body of evidence supports the role of increased presynaptic dopamine synthesis capacity in the pathoetiology of psychosis, little is known about how genetic factors affect the implicated dopamine system(s) (15).

The *Disrupted-in-Schizophrenia 1 (DISC1)* gene was originally discovered at the breakpoint of a balanced t(1;11) (q42;q14.3) translocation in a Scottish family with a high-prevalence of psychiatric disorders including schizophrenia (16-18). Further evidence for a link between *DISC1* and psychotic and affective disorders emerged from the follow-up of families displaying rare *DISC1* mutations (19, 20) and large family-based studies in the population isolate of Finland (21-23) although a large meta-analysis of families did not observe linkage at this region (24). Furthermore, evidence from individual population-based cohorts has been inconsistent (25, 26) leading to ongoing debate on its involvement in schizophrenia (27, 28). Whilst this controversy remains unresolved, there is value in seeking convergent evidence via studies elucidating the functional impact of the gene and its variations (29-32). *DISC1* is a scaffold protein involved in a wide range of neuronal functions including neuro-signalling (30, 33). Preclinical studies show that *DISC1* variant models exhibit increased amphetamine-induced dopamine release in the ventral striatum (see (34-37) reviewed in (38), indicating that *DISC1* variations might affect presynaptic dopamine synthesis capacity.

26 One of the most studied *DISC1* single nucleotide polymorphisms (SNPs) is rs821616 which is a non-
27 synonymous mutation leading to the translation of a serine (A allele) or a cysteine (T allele) at codon
28 704 in exon 11 (39). Importantly, this polymorphism represents therefore not only a variation at the
29 genetic sequence level but also at the protein sequence level of DISC1. At a molecular level,
30 Hashimoto et al. (2006) found that overexpression of the serine variant of codon 704 by viral
31 transduction resulted in a significant increase in phosphorylated ERK1/2, the more biologically active
32 form (40). ERK1/2 in turn regulates the state of phosphorylation of tyrosine hydroxylase, the rate-
33 limiting enzyme for dopamine biosynthesis, to increase its activity and subsequent dopamine synthesis
34 by up to two-fold (41-44). Dopamine is synthesized by converting first tyrosine into dihydroxyphenyl-
35 L-alanine (L-DOPA) by tyrosine hydroxylase, and second dihydroxyphenyl-L-alanine (L-DOPA) into
36 dopamine by aromatic acid decarboxylase (45). [¹⁸F]-DOPA PET signal reflects aromatic acid
37 decarboxylase function and dopamine storage capacity (45), but not directly tyrosine hydroxylase
38 function. However, it should be noted that 1) tyrosine hydroxylase is the rate limiting step for
39 dopamine synthesis capacity (43) and 2) the topological distribution of the [¹⁸F]-DOPA signal
40 correlates highly with tyrosine hydroxylase immunostaining in unilaterally 6-hydroxydopamine (6-
41 OHDA)-lesioned rats, thus indicating that the [¹⁸F]-DOPA signal is strongly influenced by
42 endogenous dopamine formed by tyrosine hydroxylase (46).

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44 In summary, preclinical findings suggest that the Ser704Cys variation affects dopamine synthesis by
45 regulating ERK1/2 and its control over tyrosine hydroxylase activity. However, it remains unknown
46 whether the Ser704Cys variation is associated with altered dopamine synthesis in humans. The aim of
47 this study was therefore to test the hypothesis that serine homozygotes would exhibit increased striatal
48 dopamine synthesis capacity relative to cysteine homozygotes and heterozygotes.

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Results

Demographics, scan parameters including the injected dose and substance use characteristics are shown in table 1. A total of 46 serine homozygotes and 56 cysteine homozygotes and heterozygotes (which encompass 45 heterozygotes and 11 cysteine homozygotes) were included in the study. The genotype frequencies (shown in table 1) did not significantly deviate from Hardy–Weinberg equilibrium ($\chi^2 = 1.422$ with $p = 0.233$), with a Minor Allele Frequency (T allele) of 0.335. Age (year) and K_i^{cer} (1/min) in the striatum were normally distributed across the two groups whereas injected dose (MBq) was not. There was no significant difference in age between groups $t(100) = 1.588$, $p = 0.115$ (independent t test) and no significant difference in injected dose $p = 0.408$ (Mann Whitney test). Levene’s test indicated no difference between the variances in the two groups, $F = 0.398$, $p = 0.529$. The univariate ANCOVA showed that the main effect of the *DISC1* SNP rs821616 on the dopamine synthesis capacity in the striatum was significant, $F(1,96) = 6.555$, $p = 0.012$, partial eta squared = 0.064. The effects of the covariates were: for scanner, $F(1,96) = 16.573$, $p < 0.01$, age, $F(1,96) = 1.056$, $p = 0.307$, gender, $F(1,96) = 0.114$, $p = 0.736$ and ethnicity, $F(1,96) = 0.061$, $p = 0.805$.

Discussion

In line with our hypothesis, we found that participants serine homozygotes (AA genotype) for the Ser704Cys functional DISC1 polymorphism exhibited a significantly greater K_i^{cer} value in the striatum, indicating greater dopamine synthesis capacity compared to cysteine homozygotes and heterozygotes (AT or TT genotype). This result is in accordance with preclinical evidence showing that the serine 704 DISC1 variant increases the activity of ERK1/2, which in turn enhances the phosphorylation of tyrosine hydroxylase, the rate limiting step in dopamine synthesis (41, 47).

Limitations

The main limitation of this study was that we used data from three different PET scanners, which could add error variance. However, scanner was included as a covariate to adjust for this. Furthermore, the effect of the Ser704Cys polymorphism remained significant when we only included subjects from PET scanner 2 ($F(1,28) = 5.273$, $p=0.029$ (N=16 cysteine homozygotes and heterozygotes, N=17 serine homozygotes)), but not PET scanner 1 only ($F(1,30) = 0.766$, $p=0.388$, (N=19 cysteine homozygotes and heterozygotes, N=16 serine homozygotes)) and PET scanner 3 only ($F(1,29) = 0.426$, $p=0.519$, (N=21 cysteine homozygotes and heterozygotes, N=13 serine homozygotes)). It is important to recognise that we measured the final step in the synthesis of dopamine, the conversion of L-DOPA into dopamine via aromatic acid decarboxylase (AADC). However, the parameter measured could be affected by other variables including the uptake of L-DOPA into the brain, although this should be controlled for by the reference region and there is no *a priori* reason to consider that this should be affected by the DISC1 protein. Importantly, this polymorphism was chosen based on a specific prior hypothesis. Although there was evidence to reject the null hypothesis, the p-value would not survive genome-wide correction and therefore the result requires replication.

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Implications for mental disorders

The Ser704Cys polymorphism has been associated with schizophrenia with an odds ratio in the range of 1.3 – 4.18 in various populations including European (48), mixed European/African-American (49), and Chinese Han (50-52). Inconsistencies have been found, with some studies indicating increased risk associated with the serine (A) allele (48, 51), whilst others the cysteine (T) (allele) (50, 52) and no association found (25) mainly in the Japanese population (53-55). A recent meta-analysis has also reported association of the serine allele with schizophrenia in Chinese (OR=1.338) and Japanese populations (OR=1.524), as well as in the overall mixed race sample (56). The inconsistencies in these results might be due to different ethnic populations. It should be noted that ever expanding studies of European ancestry population level genetic variants in schizophrenia continually demonstrate no significant associations at the entire *DISC1* locus (57, 58), although there is evidence implicating the *DISC1* interactor phosphodiesterase 4B (*PDE4B*) as a genome-wide significant single gene locus in a recent large schizophrenia genome-wide association study (GWAS) (58). Whilst GWAS have made crucial advances in the understanding of the genetic of schizophrenia, the biological mechanisms directly underlying the disorder remain yet poorly elucidated (59-61). In this context, the *DISC1* protein has been suggested as a biological candidate of interest for investigating molecular mechanisms of mental illnesses at the protein levels (33, 62). Beyond studies of dichotomous diagnoses, the serine allele has also been associated with increased risk for poor concentration among Korean patients with schizophrenia (63), increased severity of positive symptoms and hallucinations in European patients with First-Episode Psychosis (64) and increased lifetime severity of delusions in European patients with schizophrenia (65). A potential mechanism for the increased risk could be by dysregulating the control of dopamine to lead to increased dopamine synthesis. Findings in prodromal populations show that increased dopamine synthesis is associated with increased risk for psychosis (12, 13). The difference in dopamine synthesis capacity we observe here between serine homozygotes and carriers of the alternative allele is much smaller than the differences seen in at risk subjects (14,

112 66). It is therefore likely that the Ser704Cys variant interacts with other genetic changes to mediate
113 risk, potentially by affecting dopamine synthesis.

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115 The fact that the common serine allele has been described as the risk allele is compatible with
116 schizophrenia GWAS, in which approximately 50% of the implicated index SNPs are the more
117 common alleles (67). At the population level, the genetic susceptibility to schizophrenia is caused by a
118 few rare variants of high penetrance (mainly copy number variants and translocations) and many
119 common variants of small penetrance (SNPs and variable number of tandem repeats) (68). As each
120 SNP very minimally impacts schizophrenia risk and is compatible with modern models of natural
121 selection (67), it is expected that other genetic factors are needed, in the same individual, to increase
122 the liability to a point of schizophrenia onset. For example, the Ser704Cys site affects interaction with
123 nuclear distribution element-like 1 (NDEL1) and its homolog Nuclear Distribution Element 1 (NDE1,
124 also known as NudE) (69, 70), and there is evidence for an interaction between NDEL1 rs1391768
125 and the Ser704 allele and the NDE1 rs3784859 and the Cys704 allele on the risk for schizophrenia in
126 European participants (71). Ser704Cys is also the binding site for proteins such as kendrin (also
127 known as pericentrin PCNT) and Pericentriolar material 1 (PCM1) (72), which have been both
128 described as risk factor genes for schizophrenia (73). Furthermore, environmental factors such as
129 exposure to psychosocial stress may also interact with the Ser704Cys polymorphism to affect
130 dopamine function and mediate risk for schizophrenia (15). Interestingly, using a transgenic
131 expression of truncated human *Disc1* protein with dominant-negative effect, Niwa et al. have shown
132 that an interaction between *DISC1* and stress exposure, as a 3 week social isolation paradigm,
133 increased dopamine release after amphetamine challenge (34) and induced alterations in DNA
134 methylation of the tyrosine hydroxylase gene (74).

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136 Evidence also suggests that the Ser704Cys polymorphism is a risk factor for affective disorders. The
137 cysteine allele has been associated with major depression in Japanese population (47), and shown to
138 form a protective haplotype for bipolar spectrum disorder with two others *DISC1* SNPs (rs1411771

139 and rs980989) in Finnish population (75), whereas a higher serine allele rate has been found in South
140 Indian population with bipolar disorder (76). Interestingly, increased dopamine synthesis capacity is
141 seen in both mania (77) and bipolar psychosis (78), whilst major depression with affective flattening is
142 characterized by a decreased synthesis capacity (79, 80).

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144 The Ser704Cys SNP has also been shown to have a functional impact at the brain level (39).
145 Compared to healthy cysteine homozygotes and heterozygotes, serine homozygotes display increased
146 (for the same level of performance, thus putatively inefficient) prefrontal cortex activation in the left
147 middle and left superior frontal gyri and in the homologous right superior frontal gyrus, the left
148 inferior frontal and cingulate cortex, the thalamus and the caudate nucleus in a verbal fluency task
149 (81), as well as an effect on thalamic-prefrontal connectivity (82). Ser704Cys SNP has also been
150 shown to affect activation during declarative memory task with inconsistent findings. Callicott et al
151 (48) found decreased activation bilaterally in the hippocampal formation during a declarative
152 memory task and increased activation bilaterally in the hippocampal formation in an N-back task in
153 Ser704 homozygotes controls compared to cysteine homozygotes and heterozygotes, whereas Di
154 Giorgio et al (83) found increased hippocampal formation/dorsolateral prefrontal cortex coupling
155 during memory encoding in a declarative memory task in serine homozygotes compared to healthy
156 cysteine homozygotes and heterozygotes.

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158 In summary, our results provide unprecedented preliminary evidence that DISC1 Ser704Cys has an
159 impact on the dopamine synthesis capacity, in a large sample of 102 healthy volunteers. Further
160 studies should aim at 1) replicating this result in different cohorts; 2) investigating potential epistatic
161 interactions with *DISC1* and other risk genes. Genetic studies based on molecular evidence could help
162 identify the molecular mechanism that underlies the pathoetiology of dopamine-related disorders
163 such as psychotic disorders, and help identify novel potential treatment targets (15).

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Conclusion

165 We found that the serine allele of DISC1 Ser704Cys (rs821616) was associated with significantly
166 higher striatal dopamine synthesis capacity, consistently with its previous association with heightened
167 activation of ERK1/2 which stimulates tyrosine hydroxylase activity for dopamine synthesis. This
168 implicates the DISC1 polymorphism in altering a psychosis relevant mechanism in the brain i.e. the
169 facilitation of greater dopamine synthesis capacity. Although, this effect of rs821616 may be of too
170 small effect to be identified in population-based studies of end state diagnoses at their current large
171 size, it continues to implicate the functional role of DISC1. Firstly by highlighting the role of this
172 polymorphism at this gene in creating variation within the normal functioning of the brain, but also by
173 indicating this function as a potential mechanism through which other rare or familial mutations for
174 major mental illnesses could disrupt functioning and increase risk to these devastating disorders.

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Material and Methods

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Overview

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All participants gave informed written consent to take part after full description of the study. All

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studies were approved by the institutional review board and the local research ethics committee.

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Participants

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Participants were recruited via advertisement in local media based in London. One hundred and

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twenty-three participants underwent a [¹⁸F]-DOPA PET scan. For all participants the inclusion criteria

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were 1) age above 18 years; 2) capacity to give written informed consent. The exclusion criteria were

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1) any current medical conditions or history of medical condition (past minor self-limiting conditions

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were permitted); 2) history of a psychiatric disorder as determined by the Structured Clinical Interview

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for DSM-IV Axis 1 Disorders, Clinician Version (SCID-CV) (84); 3) history of substance

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abuse/dependence as determined by the Structured Clinical Interview for DSM-IV Axis 1 Disorders,

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Clinician Version (SCID-CV) (84); 4) history of head injury with a loss of consciousness; 5) a family

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history of any psychotic disorder in first- or second-degree relatives; 6) contraindications to positron

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emission tomography (PET) scanning (significant prior exposure to radiation, pregnancy or breast

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feeding). All participants provided urine samples prior to the scan to screen for drug use and

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pregnancy test in women. Six participants were excluded due to positive urine THC screening, 12

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participants were excluded to contamination of samples and 3 participants were excluded due to

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current psychotropic medication use. This resulted in the final inclusion of 102 participants (46

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females/56 males, age: 30.2±9.3 years (mean±Standard Deviation). Both scanning and imaging

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analysis were done blind to the genotype status.

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[¹⁸F]-FDOPA PET

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PET data were acquired using three different PET scanners. PET scanner 1 was an ECAT HR+ 962 PET scanner (CTI/Siemens, Knoxville, Tennessee). The dynamic images were acquired in 3D mode with an axial field of view of 15.5 cm and reconstructed using filterback projection. PET scanners 2 and 3 were two Siemens Biograph HiRez XVI PET-CT scanner (Siemens Healthcare, Erlangen, Germany) at Imanova, Centre for Imaging Sciences. PET scanner 1 and PET scanner 2-3 were identical with the only exception of the axial field of view: 16.2 cm vs 21.6 cm respectively. The dynamic images were also reconstructed using a 3D filtered back-projection algorithm (discrete inverse Fourier transform, DIFT) with a 128 matrix, a zoom of 2.6 and a 5mm isotropic Gaussian smoothing. Participants were scanned at various times of the day. Some of the imaging data has been included in prior reports but not for genetic analysis (85-88). For attenuation and model-based scatter correction, a 10 min transmission scan was performed using a 150-MBq cesium-137 rotating point source for the ECAT HR+ 962 PET scanner and a computed tomography scan (effective dose=0.36 mSv) for the Siemens Biograph HiRez XVI PET-CT scanners were acquired prior to each PET scan. Experimental protocol was consistent for all the participants (85). Participants were asked to fast and abstain from smoking from midnight on the day of the scan as tobacco use has been associated with increased striatal dopamine synthesis capacity (89) although this has not been replicated (85). Oral doses of carbidopa (150mg) and entacapone (400mg) were administrated 1 hour before scanning. While the first reduces the peripheral metabolism of the tracer (90), the latter minimizes the formation of radiolabeled [¹⁸F]-FDOPA metabolites, which can cross the blood-brain barrier (91). Head movement was monitored and minimized with a light head strap. If participants moved extensively during the acquisition or got out of the scanner a second attenuation correction image was acquired at the end of the acquisition. PET data were acquired dynamically during 95 minutes after bolus injection of the radioactive tracer [¹⁸F]-DOPA through a cannula inserted into a vein. Dynamic data were binned into 26 frames (PET scanner 1) and 32 frames (PET scanner 2 and 3).

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Image Analysis

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Head movement was corrected using a frame-by-frame realignment and denoising algorithm (92) with a level 2 order 64 Battle-Lemarie wavelet filter applied on the non-attenuation-corrected dynamic images. These images were used because they include a significant scalp signal compared to attenuation-corrected images (93). Frames were realigned to a reference frame corresponding to the frame with the highest number of counts, i.e. obtained 7 minutes (for the ECAT HR+ 962 PET scanner-CTI/Siemens, Knoxville, Tennessee) and 17 minutes (for the Siemens Biograph HiRez XVI PET-CT scanners-Siemens Healthcare, Erlangen, Germany) after the radiotracer injection using a mutual information algorithm (94). The transformation parameters were then applied to the corresponding attenuation-corrected dynamic images. These realigned frames were summed, creating a movement-corrected dynamic image from which to extract the Time Activity Curves (TAC) for graphical analysis quantification. Standardized regions in Montreal Neurologic Institute (MNI) space were defined in the striatum delineated as previously described to create a Region of Interest (ROI) map (95) and in the cerebellum using the probabilistic Martinez atlas (95, 96). The cerebellum was used as a reference region as it is largely devoid of dopaminergic neurons or projections (45). A nonlinear transformation procedure on SPM8 (<http://www.fil.ion.ucl.ac.uk/spm>) was used to normalize the ROI map together with the [¹⁸F]-DOPA template to each individual PET summation image, in order to place the ROI automatically on individual [¹⁸F]-DOPA PET dynamic images. Influx constant K_i^{cer} value, (min^{-1}) for the striatum was calculated relative to uptake in the reference region using a graphical approach (97), a method which has been shown to have good reliability (95).

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Genetic analysis

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DNA was extracted from blood or cheek swabs using standard methods (98). Genotyping of the rs821616 A>T SNP, was performed by KBioscience (Herts, UK, <http://www.kbioscience.co.uk>) using

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245 a competitive allele specific Polymerase Chain Reaction system (CASP). Quality control procedures
246 included negative control (water) wells and duplicate wells.

247 **Statistical analysis**

248 The normality of the distribution for all variables was examined using the Shapiro Wilk test,
249 inspection of Q-Q plots and skewness and kurtosis values within range of ± 2 . Homogeneity of
250 variance was assessed with Levene's Test for Equality of Variances. An alpha threshold was set at
251 0.05 (two-tailed) for significance for all statistical comparisons. Statistical Package for the Social
252 Sciences (SPSS) version 24 was used for all statistical analysis (IBM, Armonk, N.Y.). All data are
253 shown as mean \pm SD. An univariate analysis of covariance (ANCOVA) was performed on 102 healthy
254 controls, with the DISC1 SNP Ser704Cys variation (serine homozygotes versus cysteine homozygotes
255 and heterozygotes) as the independent variable, K_i^{cer} in the striatum as the dependent variable and age,
256 gender, ethnicity (table 1) and the three PET scanners separately as covariates as these variables have
257 been previously found to influence dopamine synthesis capacity (99, 100). Effect sizes are reported as
258 partial eta squared. Independent t test and Mann-Whitney test were used to compare age and injected
259 dose.

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Conflicts of interest

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D.P. is a co-founder of the neuroimaging services company NeuroPsyAI, Ltd. O.D.H. has received

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investigator-initiated research funding from and/or participated in advisory/ speaker meetings

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organised by Angellini, Astra-Zeneca, Autifony, Biogen, BMS, Eli Lilly, Heptares, Jansenn,

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Lundbeck, Lyden-Delta, Otsuka, Servier, Sunovion, Rand and Roche. Neither Dr Howes or his family

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have been employed by or have holdings / a financial stake in any biomedical company. The views

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expressed are those of the authors and not necessarily those of the NHS, the NIHR or the Department

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of Health. All other authors do not declare any conflict of interest.

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References

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293

1 Meltzer, H.Y. and Stahl, S.M. (1976) The dopamine hypothesis of schizophrenia: a review. *Schizophrenia bulletin*, **2**, 19-76.

294
295

2 Howes, O.D., McCutcheon, R. and Stone, J. (2015) Glutamate and dopamine in schizophrenia: an update for the 21st century. *Journal of psychopharmacology*, **29**, 97-115.

296
297

3 Seeman, P., Lee, T., Chau-Wong, M. and Wong, K. (1976) Antipsychotic drug doses and neuroleptic/dopamine receptors. *Nature*, **261**, 717-719.

298
299

4 Creese, I., Burt, D.R. and Snyder, S.H. (1976) Dopamine receptors and average clinical doses. *Science*, **194**, 546.

300
301
302

5 van Rossum, J.M. (1966) The significance of dopamine-receptor blockade for the mechanism of action of neuroleptic drugs. *Archives internationales de pharmacodynamie et de therapie*, **160**, 492-494.

303
304
305

6 Berman, S.M., Kuczenski, R., McCracken, J.T. and London, E.D. (2009) Potential adverse effects of amphetamine treatment on brain and behavior: a review. *Molecular psychiatry*, **14**, 123-142.

306
307
308
309

7 Grant, K.M., LeVan, T.D., Wells, S.M., Li, M., Stoltenberg, S.F., Gendelman, H.E., Carlo, G. and Bevins, R.A. (2012) Methamphetamine-associated psychosis. *Journal of neuroimmune pharmacology : the official journal of the Society on NeuroImmune Pharmacology*, **7**, 113-139.

310

8 Connell, P.H. (1957) Amphetamine Psychosis. *Br Med J*, **1**, 582.

311
312

9 Curran, C., Byrappa, N. and McBride, A. (2004) Stimulant psychosis: systematic review. *The British journal of psychiatry : the journal of mental science*, **185**, 196-204.

313
314

10 Lieberman, J.A., Kane, J.M. and Alvir, J. (1987) Provocative tests with psychostimulant drugs in schizophrenia. *Psychopharmacology*, **91**, 415-433.

315
316
317

11 Howes, O.D., Kambeitz, J., Kim, E., Stahl, D., Slifstein, M., Abi-Dargham, A. and Kapur, S. (2012) The nature of dopamine dysfunction in schizophrenia and what this means for treatment. *Archives of general psychiatry*, **69**, 776-786.

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319
320
321

12 Howes, O.D., Montgomery, A.J., Asselin, M.C., Murray, R.M., Valli, I., Tabraham, P., Bramon-Bosch, E., Valmaggia, L., Johns, L., Broome, M. *et al.* (2009) Elevated striatal dopamine function linked to prodromal signs of schizophrenia. *Archives of general psychiatry*, **66**, 13-20.

322
323
324
325

13 Egerton, A., Chaddock, C.A., Winton-Brown, T.T., Bloomfield, M.A., Bhattacharyya, S., Allen, P., McGuire, P.K. and Howes, O.D. (2013) Presynaptic striatal dopamine dysfunction in people at ultra-high risk for psychosis: findings in a second cohort. *Biological psychiatry*, **74**, 106-112.

326
327
328
329

14 Howes, O.D., Bose, S., Turkheimer, F., Valli, I., Egerton, A., Stahl, D., Valmaggia, L., Allen, P., Murray, R. and McGuire, P. (2011) Progressive increase in striatal dopamine synthesis capacity as patients develop psychosis: a PET study. *Molecular psychiatry*, **16**, 885-886.

- 330 15 Howes, O.D., McCutcheon, R., Owen, M.J. and Murray, R.M. (2017) The Role of
331 Genes, Stress, and Dopamine in the Development of Schizophrenia. *Biological psychiatry*,
332 **81**, 9-20.
- 333 16 St Clair, D., Blackwood, D., Muir, W., Carothers, A., Walker, M., Spowart, G.,
334 Gosden, C. and Evans, H.J. (1990) Association within a family of a balanced autosomal
335 translocation with major mental illness. *Lancet*, **336**, 13-16.
- 336 17 Jacobs, P., Brunton, M., Frackiewicz, A., Newton, M., Cook, P. and Robson, E.
337 (1970) Studies on a family with three cytogenetic markers. *Annals of Human Genetics*
338 (*Lond*), **33**, 325–336.
- 339 18 Millar, J.K., Wilson-Annan, J.C., Anderson, S., Christie, S., Taylor, M.S., Semple,
340 C.A., Devon, R.S., St Clair, D.M., Muir, W.J., Blackwood, D.H. *et al.* (2000) Disruption of two
341 novel genes by a translocation co-segregating with schizophrenia. *Hum Mol Genet*, **9**, 1415-
342 1423.
- 343 19 Blackwood, D.H., Fordyce, A., Walker, M.T., St Clair, D.M., Porteous, D.J. and Muir,
344 W.J. (2001) Schizophrenia and affective disorders--cosegregation with a translocation at
345 chromosome 1q42 that directly disrupts brain-expressed genes: clinical and P300 findings in
346 a family. *American journal of human genetics*, **69**, 428-433.
- 347 20 Sachs, N.A., Sawa, A., Holmes, S.E., Ross, C.A., DeLisi, L.E. and Margolis, R.L.
348 (2005) A frameshift mutation in Disrupted in Schizophrenia 1 in an American family with
349 schizophrenia and schizoaffective disorder. *Molecular psychiatry*, **10**, 758-764.
- 350 21 Ekelund, J., Hennah, W., Hiekkalinna, T., Parker, A., Meyer, J., Lonqvist, J. and
351 Peltonen, L. (2004) Replication of 1q42 linkage in Finnish schizophrenia pedigrees.
352 *Molecular psychiatry*, **9**, 1037-1041.
- 353 22 Ekelund, J., Hovatta, I., Parker, A., Paunio, T., Varilo, T., Martin, R., Suhonen, J.,
354 Ellonen, P., Chan, G., Sinsheimer, J.S. *et al.* (2001) Chromosome 1 loci in Finnish
355 schizophrenia families. *Hum Mol Genet*, **10**, 1611-1617.
- 356 23 Hennah, W., Varilo, T., Kestila, M., Paunio, T., Arajärvi, R., Haukka, J., Parker, A.,
357 Martin, R., Levitzky, S., Partonen, T. *et al.* (2003) Haplotype transmission analysis provides
358 evidence of association for DISC1 to schizophrenia and suggests sex-dependent effects.
359 *Hum Mol Genet*, **12**, 3151-3159.
- 360 24 Lewis, C.M., Levinson, D.F., Wise, L.H., DeLisi, L.E., Straub, R.E., Hovatta, I.,
361 Williams, N.M., Schwab, S.G., Pulver, A.E., Faraone, S.V. *et al.* (2003) Genome scan meta-
362 analysis of schizophrenia and bipolar disorder, part II: Schizophrenia. *American journal of*
363 *human genetics*, **73**, 34-48.
- 364 25 Mathieson, I., Munafo, M.R. and Flint, J. (2012) Meta-analysis indicates that common
365 variants at the DISC1 locus are not associated with schizophrenia. *Molecular psychiatry*, **17**,
366 634-641.
- 367 26 Farrell, M.S., Werge, T., Sklar, P., Owen, M.J., Ophoff, R.A., O'Donovan, M.C.,
368 Corvin, A., Cichon, S. and Sullivan, P.F. (2015) Evaluating historical candidate genes for
369 schizophrenia. *Molecular psychiatry*, **20**, 555-562.
- 370 27 Sullivan, P.F. (2013) Questions about DISC1 as a genetic risk factor for
371 schizophrenia. *Molecular psychiatry*, **18**, 1050-1052.

- 372 28 Porteous, D.J., Thomson, P.A., Millar, J.K., Evans, K.L., Hennah, W., Soares, D.C.,
373 McCarthy, S., McCombie, W.R., Clapcote, S.J., Korth, C. *et al.* (2014) DISC1 as a genetic
374 risk factor for schizophrenia and related major mental illness: response to Sullivan. *Molecular*
375 *psychiatry*, **19**, 141-143.
- 376 29 Brandon, N.J., Millar, J.K., Korth, C., Sive, H., Singh, K.K. and Sawa, A. (2009)
377 Understanding the role of DISC1 in psychiatric disease and during normal development. *The*
378 *Journal of neuroscience : the official journal of the Society for Neuroscience*, **29**, 12768-
379 12775.
- 380 30 Porteous, D.J., Millar, J.K., Brandon, N.J. and Sawa, A. (2011) DISC1 at 10:
381 connecting psychiatric genetics and neuroscience. *Trends in molecular medicine*, **17**, 699-
382 706.
- 383 31 Hennah, W., Thomson, P., McQuillin, A., Bass, N., Loukola, A., Anjorin, A.,
384 Blackwood, D., Curtis, D., Deary, I.J., Harris, S.E. *et al.* (2009) DISC1 association,
385 heterogeneity and interplay in schizophrenia and bipolar disorder. *Molecular psychiatry*, **14**,
386 865-873.
- 387 32 Tomppo, L., Hennah, W., Miettunen, J., Jarvelin, M.R., Veijola, J., Ripatti, S.,
388 Lahermo, P., Lichtermann, D., Peltonen, L. and Ekelund, J. (2009) Association of variants in
389 DISC1 with psychosis-related traits in a large population cohort. *Archives of general*
390 *psychiatry*, **66**, 134-141.
- 391 33 Brandon, N.J. and Sawa, A. (2011) Linking neurodevelopmental and synaptic
392 theories of mental illness through DISC1. *Nature reviews. Neuroscience*, **12**, 707-722.
- 393 34 Niwa, M., Jaaro-Peled, H., Tankou, S., Seshadri, S., Hikida, T., Matsumoto, Y.,
394 Cascella, N.G., Kano, S., Ozaki, N., Nabeshima, T. *et al.* (2013) Adolescent stress-induced
395 epigenetic control of dopaminergic neurons via glucocorticoids. *Science*, **339**, 335-339.
- 396 35 Jaaro-Peled, H., Niwa, M., Foss, C.A., Murai, R., de Los Reyes, S., Kamiya, A.,
397 Mateo, Y., O'Donnell, P., Cascella, N.G., Nabeshima, T. *et al.* (2013) Subcortical
398 dopaminergic deficits in a DISC1 mutant model: a study in direct reference to human
399 molecular brain imaging. *Hum Mol Genet*, **22**, 1574-1580.
- 400 36 Niwa, M., Kamiya, A., Murai, R., Kubo, K., Gruber, A.J., Tomita, K., Lu, L., Tomisato,
401 S., Jaaro-Peled, H., Seshadri, S. *et al.* (2010) Knockdown of DISC1 by in utero gene transfer
402 disturbs postnatal dopaminergic maturation in the frontal cortex and leads to adult behavioral
403 deficits. *Neuron*, **65**, 480-489.
- 404 37 Nakai, T., Nagai, T., Wang, R., Yamada, S., Kuroda, K., Kaibuchi, K. and Yamada, K.
405 (2014) Alterations of GABAergic and dopaminergic systems in mutant mice with disruption of
406 exons 2 and 3 of the Disc1 gene. *Neurochemistry international*, **74**, 74-83.
- 407 38 Dahoun, T., Trossbach, S.V., Brandon, N.J., Korth, C. and Howes, O.D. (2017) The
408 impact of Disrupted-in-Schizophrenia 1 (DISC1) on the dopaminergic system: a systematic
409 review. *Translational psychiatry*, **7**, e1015.
- 410 39 Duff, B.J., Macritchie, K.A., Moorhead, T.W., Lawrie, S.M. and Blackwood, D.H.
411 (2013) Human brain imaging studies of DISC1 in schizophrenia, bipolar disorder and
412 depression: a systematic review. *Schizophr Res*, **147**, 1-13.
- 413 40 Roskoski, R., Jr. (2012) ERK1/2 MAP kinases: structure, function, and regulation.
414 *Pharmacol Res*, **66**, 105-143.

- 415 41 Lindgren, N., Goiny, M., Herrera-Marschitz, M., Haycock, J.W., Hokfelt, T. and
416 Fisone, G. (2002) Activation of extracellular signal-regulated kinases 1 and 2 by
417 depolarization stimulates tyrosine hydroxylase phosphorylation and dopamine synthesis in
418 rat brain. *The European journal of neuroscience*, **15**, 769-773.
- 419 42 Guo, Z., Du, X. and Iacovitti, L. (1998) Regulation of tyrosine hydroxylase gene
420 expression during transdifferentiation of striatal neurons: changes in transcription factors
421 binding the AP-1 site. *The Journal of neuroscience : the official journal of the Society for
422 Neuroscience*, **18**, 8163-8174.
- 423 43 Daubner, S.C., Le, T. and Wang, S. (2011) Tyrosine hydroxylase and regulation of
424 dopamine synthesis. *Archives of biochemistry and biophysics*, **508**, 1-12.
- 425 44 Haycock, J.W. (2002) Peptide substrates for ERK1/2: structure-function studies of
426 serine 31 in tyrosine hydroxylase. *Journal of neuroscience methods*, **116**, 29-34.
- 427 45 Kumakura, Y. and Cumming, P. (2009) PET studies of cerebral levodopa metabolism:
428 a review of clinical findings and modeling approaches. *The Neuroscientist : a review journal
429 bringing neurobiology, neurology and psychiatry*, **15**, 635-650.
- 430 46 Kyono, K., Takashima, T., Katayama, Y., Kawasaki, T., Zochi, R., Gouda, M.,
431 Kuwahara, Y., Takahashi, K., Wada, Y., Onoe, H. *et al.* (2011) Use of [18F]FDOPA-PET for
432 in vivo evaluation of dopaminergic dysfunction in unilaterally 6-OHDA-lesioned rats. *EJNMMI
433 Res*, **1**, 25.
- 434 47 Hashimoto, R., Numakawa, T., Ohnishi, T., Kumamaru, E., Yagasaki, Y., Ishimoto, T.,
435 Mori, T., Nemoto, K., Adachi, N., Izumi, A. *et al.* (2006) Impact of the DISC1 Ser704Cys
436 polymorphism on risk for major depression, brain morphology and ERK signaling. *Hum Mol
437 Genet*, **15**, 3024-3033.
- 438 48 Callicott, J.H., Straub, R.E., Pezawas, L., Egan, M.F., Mattay, V.S., Hariri, A.R.,
439 Verchinski, B.A., Meyer-Lindenberg, A., Balkissoon, R., Kolachana, B. *et al.* (2005) Variation
440 in DISC1 affects hippocampal structure and function and increases risk for schizophrenia.
441 *Proceedings of the National Academy of Sciences of the United States of America*, **102**,
442 8627-8632.
- 443 49 Song, W., Li, W., Feng, J., Heston, L.L., Scaringe, W.A. and Sommer, S.S. (2008)
444 Identification of high risk DISC1 structural variants with a 2% attributable risk for
445 schizophrenia. *Biochemical and biophysical research communications*, **367**, 700-706.
- 446 50 Qu, M., Tang, F., Yue, W., Ruan, Y., Lu, T., Liu, Z., Zhang, H., Han, Y., Zhang, D.,
447 Wang, F. *et al.* (2007) Positive association of the Disrupted-in-Schizophrenia-1 gene (DISC1)
448 with schizophrenia in the Chinese Han population. *American journal of medical genetics. Part
449 B, Neuropsychiatric genetics : the official publication of the International Society of
450 Psychiatric Genetics*, **144B**, 266-270.
- 451 51 Luo, X., Jin, C., Zhou, Z., Liu, X., Zhang, F., Zhang, F., Zhu, J., Wang, Y., Cheng, Z.
452 and Shugart, Y.Y. (2015) New findings support the association of DISC1 genetic variants
453 with susceptibility to schizophrenia in the Han Chinese population. *Psychiatry research*, **228**,
454 966-968.
- 455 52 He, B.S., Zhang, L.Y., Pan, Y.Q., Lin, K., Zhang, L.L., Sun, H.L., Gao, T.Y., Su, T.Q.,
456 Wang, S.K. and Zhu, C.B. (2016) Association of the DISC1 and NRG1 genetic
457 polymorphisms with schizophrenia in a Chinese population. *Gene*, **590**, 293-297.

- 458 53 Schumacher, J., Laje, G., Abou Jamra, R., Becker, T., Muhleisen, T.W., Vasilescu,
459 C., Mattheisen, M., Herms, S., Hoffmann, P., Hillmer, A.M. *et al.* (2009) The DISC locus and
460 schizophrenia: evidence from an association study in a central European sample and from a
461 meta-analysis across different European populations. *Hum Mol Genet*, **18**, 2719-2727.
- 462 54 Kinoshita, M., Numata, S., Tajima, A., Ohi, K., Hashimoto, R., Shimodera, S., Imoto,
463 I., Itakura, M., Takeda, M. and Ohmori, T. (2012) Meta-analysis of association studies
464 between DISC1 missense variants and schizophrenia in the Japanese population. *Schizophr*
465 *Res*, **141**, 271-273.
- 466 55 Ratta-Apha, W., Hishimoto, A., Mouri, K., Shiroywa, K., Sasada, T., Yoshida, M.,
467 Supriyanto, I., Ueno, Y., Asano, M., Shirakawa, O. *et al.* (2013) Association analysis of the
468 DISC1 gene with schizophrenia in the Japanese population and DISC1 immunoreactivity in
469 the postmortem brain. *Neuroscience research*, **77**, 222-227.
- 470 56 Wang, H.Y., Liu, Y., Yan, J.W., Hu, X.L., Zhu, D.M., Xu, X.T. and Li, X.S. (2017)
471 Gene polymorphisms of DISC1 is associated with schizophrenia: Evidence from a meta-
472 analysis. *Progress in neuro-psychopharmacology & biological psychiatry*, **81**, 64-73.
- 473 57 Schizophrenia Working Group of the Psychiatric Genomics, C. (2014) Biological
474 insights from 108 schizophrenia-associated genetic loci. *Nature*, **511**, 421-427.
- 475 58 Pardiñas, A.F., Holmans, P., Pocklington, A.J., Escott-Price, V., Ripke, S., Carrera,
476 N., Legge, S.E., Bishop, S., Cameron, D., Hamshere, M.L. *et al.* (2016) Common
477 schizophrenia alleles are enriched in mutation-intolerant genes and maintained by
478 background selection. *bioRxiv*, in press.
- 479 59 Harrison, P.J. (2014) Recent genetic findings in schizophrenia and their therapeutic
480 relevance. *Journal of psychopharmacology*, in press.
- 481 60 Corvin, A. and Sullivan, P.F. (2016) What Next in Schizophrenia Genetics for the
482 Psychiatric Genomics Consortium? *Schizophrenia bulletin*, **42**, 538-541.
- 483 61 McClellan, J. and King, M.C. (2010) Genomic analysis of mental illness: a changing
484 landscape. *Jama*, **303**, 2523-2524.
- 485 62 Niwa, M., Cash-Padgett, T., Kubo, K.I., Saito, A., Ishii, K., Sumitomo, A., Taniguchi,
486 Y., Ishizuka, K., Jaaro-Peled, H., Tomoda, T. *et al.* (2016) DISC1 a key molecular lead in
487 psychiatry and neurodevelopment: No-More Disrupted-in-Schizophrenia 1. *Molecular*
488 *psychiatry*, **21**, 1488-1489.
- 489 63 Kim, H.J., Park, H.J., Jung, K.H., Ban, J.Y., Ra, J., Kim, J.W., Park, J.K., Choe, B.K.,
490 Yim, S.V., Kwon, Y.K. *et al.* (2008) Association study of polymorphisms between DISC1 and
491 schizophrenia in a Korean population. *Neuroscience letters*, **430**, 60-63.
- 492 64 Vazquez-Bourgon, J., Mata, I., Roiz-Santianez, R., Ayesa-Arriola, R., Suarez Pinilla,
493 P., Tordesillas-Gutierrez, D., Vazquez-Barquero, J.L. and Crespo-Facorro, B. (2014) A
494 Disrupted-in-Schizophrenia 1 Gene Variant is Associated with Clinical Symptomatology in
495 Patients with First-Episode Psychosis. *Psychiatry investigation*, **11**, 186-191.
- 496 65 DeRosse, P., Hodgkinson, C.A., Lencz, T., Burdick, K.E., Kane, J.M., Goldman, D.
497 and Malhotra, A.K. (2007) Disrupted in schizophrenia 1 genotype and positive symptoms in
498 schizophrenia. *Biological psychiatry*, **61**, 1208-1210.

- 499 66 Howes, O.D., Bose, S.K., Turkheimer, F., Valli, I., Egerton, A., Valmaggia, L.R.,
500 Murray, R.M. and McGuire, P. (2011) Dopamine synthesis capacity before onset of
501 psychosis: a prospective [18F]-DOPA PET imaging study. *The American journal of*
502 *psychiatry*, **168**, 1311-1317.
- 503 67 Pardinas, A.F., Holmans, P., Pocklington, A.J., Escott-Price, V., Ripke, S., Carrera,
504 N., Legge, S.E., Bishop, S., Cameron, D., Hamshere, M.L. *et al.* (2018) Common
505 schizophrenia alleles are enriched in mutation-intolerant genes and in regions under strong
506 background selection. *Nature genetics*, **50**, 381-389.
- 507 68 Owen, M.J., Sawa, A. and Mortensen, P.B. (2016) Schizophrenia. *Lancet*, **388**, 86-
508 97.
- 509 69 Leliveld, S.R., Hendriks, P., Michel, M., Sajnani, G., Bader, V., Trossbach, S.,
510 Prikulis, I., Hartmann, R., Jonas, E., Willbold, D. *et al.* (2009) Oligomer assembly of the C-
511 terminal DISC1 domain (640-854) is controlled by self-association motifs and disease-
512 associated polymorphism S704C. *Biochemistry*, **48**, 7746-7755.
- 513 70 Kamiya, A., Tomoda, T., Chang, J., Takaki, M., Zhan, C., Morita, M., Cascio, M.B.,
514 Elashvili, S., Koizumi, H., Takanezawa, Y. *et al.* (2006) DISC1-NDEL1/NUDEL protein
515 interaction, an essential component for neurite outgrowth, is modulated by genetic variations
516 of DISC1. *Hum Mol Genet*, **15**, 3313-3323.
- 517 71 Burdick, K.E., Kamiya, A., Hodgkinson, C.A., Lencz, T., DeRosse, P., Ishizuka, K.,
518 Elashvili, S., Arai, H., Goldman, D., Sawa, A. *et al.* (2008) Elucidating the relationship
519 between DISC1, NDEL1 and NDE1 and the risk for schizophrenia: evidence of epistasis and
520 competitive binding. *Hum Mol Genet*, **17**, 2462-2473.
- 521 72 Soares, D.C., Carlyle, B.C., Bradshaw, N.J. and Porteous, D.J. (2011) DISC1:
522 Structure, Function, and Therapeutic Potential for Major Mental Illness. *ACS chemical*
523 *neuroscience*, **2**, 609-632.
- 524 73 Bradshaw, N.J. and Porteous, D.J. (2012) DISC1-binding proteins in neural
525 development, signalling and schizophrenia. *Neuropharmacology*, **62**, 1230-1241.
- 526 74 Niwa, M., Lee, R.S., Tanaka, T., Okada, K., Kano, S. and Sawa, A. (2016) A critical
527 period of vulnerability to adolescent stress: epigenetic mediators in mesocortical
528 dopaminergic neurons. *Hum Mol Genet*, **25**, 1370-1381.
- 529 75 Palo, O.M., Antila, M., Silander, K., Hennah, W., Kilpinen, H., Soronen, P., Tuulio-
530 Henriksson, A., Kieseppa, T., Partonen, T., Lonnqvist, J. *et al.* (2007) Association of distinct
531 allelic haplotypes of DISC1 with psychotic and bipolar spectrum disorders and with
532 underlying cognitive impairments. *Hum Mol Genet*, **16**, 2517-2528.
- 533 76 Ram Murthy, A., Purushottam, M., Kiran Kumar, H.B., ValliKiran, M., Krishna, N.,
534 Jayramu Sriharsha, K., Janardhan Reddy, Y.C., Ghosh, S. and Jain, S. (2012) Gender-
535 specific association of TSNAX/DISC1 locus for schizophrenia and bipolar affective disorder
536 in South Indian population. *J Hum Genet*, **57**, 523-530.
- 537 77 Ashok, A.H., Marques, T.R., Jauhar, S., Nour, M.M., Goodwin, G.M., Young, A.H. and
538 Howes, O.D. (2017) The dopamine hypothesis of bipolar affective disorder: the state of the
539 art and implications for treatment. *Molecular psychiatry*, **22**, 666-679.
- 540 78 Jauhar, S., Nour, M.M., Veronese, M., Rogdaki, M., Bonoldi, I., Azis, M., Turkheimer,
541 F., McGuire, P., Young, A.H. and Howes, O.D. (2017) A Test of the Transdiagnostic

- 542 Dopamine Hypothesis of Psychosis Using Positron Emission Tomographic Imaging in Bipolar
543 Affective Disorder and Schizophrenia. *JAMA psychiatry*, **74**, 1206-1213.
- 544 79 Bragulat, V., Paillere-Martinot, M.L., Artiges, E., Frouin, V., Poline, J.B. and Martinot,
545 J.L. (2007) Dopaminergic function in depressed patients with affective flattening or with
546 impulsivity: [18F]fluoro-L-dopa positron emission tomography study with voxel-based
547 analysis. *Psychiatry research*, **154**, 115-124.
- 548 80 Martinot, M., Bragulat, V., Artiges, E., Dolle, F., Hinnen, F., Jouvent, R. and Martinot,
549 J. (2001) Decreased presynaptic dopamine function in the left caudate of depressed patients
550 with affective flattening and psychomotor retardation. *The American journal of psychiatry*,
551 **158**, 314-316.
- 552 81 Prata, D.P., Mechelli, A., Fu, C.H., Picchioni, M., Kane, F., Kalidindi, S., McDonald,
553 C., Kravariti, E., Touloupoulou, T., Miorelli, A. *et al.* (2008) Effect of disrupted-in-
554 schizophrenia-1 on pre-frontal cortical function. *Molecular psychiatry*, **13**, 915-917, 909.
- 555 82 Liu, B., Fan, L., Cui, Y., Zhang, X., Hou, B., Li, Y., Qin, W., Wang, D., Yu, C. and
556 Jiang, T. (2015) DISC1 Ser704Cys impacts thalamic-prefrontal connectivity. *Brain structure*
557 *& function*, **220**, 91-100.
- 558 83 Di Giorgio, A., Blasi, G., Sambataro, F., Rampino, A., Papazacharias, A., Gambi, F.,
559 Romano, R., Caforio, G., Rizzo, M., Latorre, V. *et al.* (2008) Association of the SerCys
560 DISC1 polymorphism with human hippocampal formation gray matter and function during
561 memory encoding. *The European journal of neuroscience*, **28**, 2129-2136.
- 562 84 First, Michael B., Spitzer, Robert L, Gibbon Miriam and Williams, J.B.W. (1996)
563 Structured Clinical Interview for DSM-IV Axis I Disorders. *American Psychiatric Press, Inc.*, ,
564 in press.
- 565 85 Bloomfield, M.A., Pepper, F., Egerton, A., Demjaha, A., Tomasi, G., Mouchlianitis, E.,
566 Maximen, L., Veronese, M., Turkheimer, F., Selvaraj, S. *et al.* (2014) Dopamine function in
567 cigarette smokers: an [(1)(8)F]-DOPA PET study. *Neuropsychopharmacology : official*
568 *publication of the American College of Neuropsychopharmacology*, **39**, 2397-2404.
- 569 86 Bloomfield, M.A., Morgan, C.J., Egerton, A., Kapur, S., Curran, H.V. and Howes, O.D.
570 (2014) Dopaminergic function in cannabis users and its relationship to cannabis-induced
571 psychotic symptoms. *Biological psychiatry*, **75**, 470-478.
- 572 87 Jauhar, S., Veronese, M., Rogdaki, M., Bloomfield, M., Natesan, S., Turkheimer, F.,
573 Kapur, S. and Howes, O.D. (2017) Regulation of dopaminergic function: an [18F]-DOPA PET
574 apomorphine challenge study in humans. *Translational psychiatry*, **7**, e1027.
- 575 88 Froudish-Walsh, S., Bloomfield, M.A., Veronese, M., Kroll, J., Karolis, V.R., Jauhar,
576 S., Bonoldi, I., McGuire, P.K., Kapur, S., Murray, R.M. *et al.* (2017) The effect of perinatal
577 brain injury on dopaminergic function and hippocampal volume in adult life. *eLife*, **6**.
- 578 89 Salokangas, R.K., Vilkmann, H., Ilonen, T., Taiminen, T., Bergman, J., Haaparanta, M.,
579 Solin, O., Alanen, A., Syvalahti, E. and Hietala, J. (2000) High levels of dopamine activity in
580 the basal ganglia of cigarette smokers. *The American journal of psychiatry*, **157**, 632-634.
- 581 90 Garnett, E.S., Firnau, G. and Nahmias, C. (1983) Dopamine visualized in the basal
582 ganglia of living man. *Nature*, **305**, 137-138.

- 583 91 Sawle, G.V., Burn, D.J., Morrish, P.K., Lammertsma, A.A., Snow, B.J., Luthra, S.,
584 Osman, S. and Brooks, D.J. (1994) The effect of entacapone (OR-611) on brain [18F]-6-L-
585 fluorodopa metabolism: implications for levodopa therapy of Parkinson's disease. *Neurology*,
586 **44**, 1292-1297.
- 587 92 Turkheimer, F.E., Brett, M., Visvikis, D. and Cunningham, V.J. (1999) Multiresolution
588 analysis of emission tomography images in the wavelet domain. *Journal of cerebral blood*
589 *flow and metabolism : official journal of the International Society of Cerebral Blood Flow and*
590 *Metabolism*, **19**, 1189-1208.
- 591 93 Bose, S.K., Turkheimer, F.E., Howes, O.D., Mehta, M.A., Cunliffe, R., Stokes, P.R.
592 and Grasby, P.M. (2008) Classification of schizophrenic patients and healthy controls using
593 [18F] fluorodopa PET imaging. *Schizophr Res*, **106**, 148-155.
- 594 94 Studholme, C., Hill, D.L. and Hawkes, D.J. (1997) Automated three-dimensional
595 registration of magnetic resonance and positron emission tomography brain images by
596 multiresolution optimization of voxel similarity measures. *Medical physics*, **24**, 25-35.
- 597 95 Egerton, A., Demjaha, A., McGuire, P., Mehta, M.A. and Howes, O.D. (2010) The
598 test-retest reliability of 18F-DOPA PET in assessing striatal and extrastriatal presynaptic
599 dopaminergic function. *NeuroImage*, **50**, 524-531.
- 600 96 Martinez, D., Slifstein, M., Broft, A., Mawlawi, O., Hwang, D.R., Huang, Y., Cooper,
601 T., Kegeles, L., Zarah, E., Abi-Dargham, A. *et al.* (2003) Imaging human mesolimbic
602 dopamine transmission with positron emission tomography. Part II: amphetamine-induced
603 dopamine release in the functional subdivisions of the striatum. *Journal of cerebral blood flow*
604 *and metabolism : official journal of the International Society of Cerebral Blood Flow and*
605 *Metabolism*, **23**, 285-300.
- 606 97 Patlak, C.S. and Blasberg, R.G. (1985) Graphical evaluation of blood-to-brain transfer
607 constants from multiple-time uptake data. Generalizations. *Journal of cerebral blood flow and*
608 *metabolism : official journal of the International Society of Cerebral Blood Flow and*
609 *Metabolism*, **5**, 584-590.
- 610 98 Freeman, B., Smith, N., Curtis, C., Hockett, L., Mill, J. and Craig, I.W. (2003) DNA
611 from buccal swabs recruited by mail: evaluation of storage effects on long-term stability and
612 suitability for multiplex polymerase chain reaction genotyping. *Behavior genetics*, **33**, 67-72.
- 613 99 Kumakura, Y., Vernaleken, I., Buchholz, H.G., Borghammer, P., Danielsen, E.,
614 Grunder, G., Heinz, A., Bartenstein, P. and Cumming, P. (2010) Age-dependent decline of
615 steady state dopamine storage capacity of human brain: an FDOPA PET study.
616 *Neurobiology of aging*, **31**, 447-463.
- 617 100 Egerton, A., Howes, O.D., Houle, S., McKenzie, K., Valmaggia, L.R., Bagby, M.R.,
618 Tseng, H.H., Bloomfield, M.A., Kenk, M., Bhattacharyya, S. *et al.* (2017) Elevated Striatal
619 Dopamine Function in Immigrants and Their Children: A Risk Mechanism for Psychosis.
620 *Schizophrenia bulletin*, **43**, 293-301.
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Legend to Figure

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Figure 1: Mean (SEM) striatal dopamine synthesis capacity (K_i^{cer} value, min^{-1}) in *DISC1* rs821616 cysteine homozygotes and

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heterozygotes (TT and TA, N=56) and *DISC1* rs821616 serine homozygotes (AA, N=46). Dopamine synthesis capacity was

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significantly increased in serine homozygotes compared with cysteine homozygotes and heterozygotes ($F(1,96)=6.555$,

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$p=0.012$).

628

Table

Table 1	DISCI SNP rs821616			
	Total	cysteine homozygotes and heterozygotes	serine homozygotes	P value
Total genotype counts	102	45 (AT) and 11 (TT)	46 (AA)	
Females	46	21	25	
PET scanner 1	35	19	16	0.549 ⁱⁱⁱ
PET scanner 2	33	16	17	
PET scanner 3	34	21	13	
Age	30.2 (9.3)	31.5 (9.9)	28.6 (8.4)	0.115 ⁱ
Tobacco smoking status (nonsmoker)	75	43	32	0.411 ⁱⁱ
Tobacco smoking status (smoker)	27	13	14	
Radioactivity injected (MBq)	157.7 (16.2)	156.6 (16.2)	159.2 (16.4)	0.529 ⁱⁱ
White European	70	35	35	0.503 ⁱⁱⁱ
Black British/other	22	15	7	
Asian British/other	5	3	2	
Mixed ethnicity	5	3	2	
All data ± SD. ⁱ Independent t test ⁱⁱ Mann-Whitney U test ⁱⁱⁱ Pearson Chi-Square				

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Abbreviations

633	Analysis of covariance (ANCOVA)
634	Competitive allele specific Polymerase Chain Reaction system (CASP)
635	Cysteine (cys)
636	Disrupted-in-Schizophrenia 1 (<i>DISC1</i>)
637	Extracellular signal-regulated protein Kinases 1 and 2 (ERK1/2)
638	Genome-wide association study (GWAS)
639	Montreal Neurologic Institute (MNI)
640	Nuclear distribution element-like 1 (NDEL1)
641	Nuclear Distribution Element 1 (NDE1)
642	Pericentrin (PCNT)
643	Pericentriolar material 1 (PCM1)
644	Phosphodiesterase 4B (PDE4B)
645	Positron Emission Tomography (PET)
646	Serine (ser)
647	Single-Nucleotide Polymorphism (SNP)
648	Statistical Package for the Social Sciences (SPSS)
649	Structured Clinical Interview for DSM-IV Axis 1 Disorders, Clinician Version (SCID-CV)
650	Time Activity Curves (TAC)
651	[¹⁸ F] fluoro-3,4-dihydroxyphenyl-L-alanine (F-DOPA)

