

## DISPATCH

### Organelles: The Emerging Signalling Chart of Mitochondrial Dynamics

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**Many molecular and functional details of single events in mitochondrial dynamics have been reported, but little is known about their coordination (AU: ). A recent study describes how cellular Ca<sup>2+</sup> signals, via remodelling the actin cytoskeleton, synchronise the formation of endoplasmic reticulum–mitochondria contacts with inner and outer mitochondrial membrane fission.**

Our understanding of individual functional modules of mitochondrial dynamics, such as mitochondrial movement, mitochondrial shape and size (fusion and fission), and mitochondria–organelle interactions (i.e., with the endoplasmic reticulum, plasma membrane and lysosome), has increased massively during the last decade (Figure 1; for recent reviews see [1–3]). The overall picture is complex, but homeostasis in these modules is clearly an absolute requirement for normal cellular function, since perturbation of the system leads to altered cellular phenotypes, loss of function (AU: please clarify what you mean here by ‘loss of fuon’) and often results in cell death [3]. Intriguingly, recent discoveries have also led to the suggestion that the distinct modules are coordinated, so it has been anticipated that cellular signals co-regulate the individual functional elements underpinning mitochondrial dynamics.

The role of contacts between the endoplasmic reticulum (ER) and mitochondria in marking the sites of mitochondrial fission and in triggering the process was recently established, with these two modules thus co-regulating downstream events, such as the segregation and maintenance of mitochondrial DNA, which are required for long-term cellular energetic homeostasis [4]. Moreover, it has been proposed that the dynamics of ER–mitochondria contacts depend on mitochondrial

motility, a process possibly involving the metazoan Miro GTPases and their yeast orthologue Gem1, which shows a genetic interaction with components of the ER–mitochondria encounter structure (ERMES) (AU:OK?) [5]. Importantly,  $\text{Ca}^{2+}$  transfer is a central component of ER–mitochondria interactions, and  $\text{Ca}^{2+}$  has been suggested to regulate the extent of the interaction surface, at least under stress conditions [6]. Similarly,  $\text{Ca}^{2+}$  also triggers fission of both the outer and inner mitochondrial membranes [3,7].

In addition, the role of the cytoskeleton as modulator of mitochondrial dynamics has been extensively mapped. The key functions of the microtubule network in metazoans and the actin cytoskeleton in yeast for mitochondrial movement and positioning are well documented [1,8]. Moreover, actin and myosin in mammalian cells have been repeatedly shown to participate in the fission process [9,10]. The role of  $\text{Ca}^{2+}$  in the mitochondrial matrix and in metabolism (AU: please clarify how the role of  $\text{Ca}^{2+}$  in metabolism relates to fission as it is not mentioned in the part of the sentence that follows the colon) has also been highlighted in this interaction:  $\text{Ca}^{2+}$  mediated constrictions of the inner mitochondrial membrane were shown to precede the completion of fission by the outer mitochondrial membrane. However, how these complex interactions are coordinated by cellular signalling has not yet been shown.

Now, in a recent study, Chakrabarti *et al.* [11] have connected the dots, describing a signalling network by which  $\text{Ca}^{2+}$ , a global cellular signal, triggers a coordinated response involving all the functional modules of mitochondrial dynamics. Their key observation is that  $\text{Ca}^{2+}$  and the formin INF2 mediate actin polymerization on the ER surface that then expands the ER–mitochondria contact site (AU:OK?).  $\text{Ca}^{2+}$  mobilization from the ER was previously shown to trigger actin redistribution around mitochondria [12]. Strikingly, however, in the new work (AU:OK?)s mobilization is shown to be sufficiently fast to promote efficient  $\text{Ca}^{2+}$  transfer between the ER and mitochondria during  $\text{Ca}^{2+}$  transients, ensuring the activation of mitochondrial  $\text{Ca}^{2+}$  uptake through the mitochondrial  $\text{Ca}^{2+}$  uniporter complex. So far, a few examples of *in situ* dynamic regulation of ER–mitochondria contacts have been described [2,6], but such a coordinated regulation of different modules in mitochondrial dynamics has not been previously demonstrated. From the work by Chakrabarti *et al.* [11] (AU:OK?) actin nucleation on the ER emerges as a highly dynamic regulator of the contact sites, promoting fast  $\text{Ca}^{2+}$  accumulation in the mitochondria. To close the  $\text{Ca}^{2+}$ -mediated circuit, these authors also confirm that increased levels of  $\text{Ca}^{2+}$  in the matrix trigger inner mitochondrial membrane constrictions [7]. These precede and induce Drp1-dependent fission, which is also promoted by actin- and  $\text{Ca}^{2+}$ -dependent recruitment of Drp1 to the outer mitochondrial

membrane from the cytoplasm (Figure 1) [10]. This view unveils actin and  $\text{Ca}^{2+}$  as central players in integrating mitochondrial fission and ER–mitochondria contact formation, through an intra-mitochondrial  $\text{Ca}^{2+}$ -mediated loop, to ensure complete fission of both mitochondrial membranes. This intriguing mechanism aligns well with the previously known roles of  $\text{Ca}^{2+}$  in the individual functional modules (Figure 1), but also raises a series of outstanding questions.

First, how does acute actin redistribution affect mitochondrial movement and overall positioning, and via what mechanism? (AU:?) Mitochondrial distribution in mammalian cells so far has been attributed to microtubule-associated motor activity, and inhibition of mitochondrial movement along microtubules by  $\text{Ca}^{2+}$  and reactive oxygen species has been shown to contribute to the positioning of these organelles [13]. On the other hand, the role of the actin network in mitochondrial distribution is critical in yeast [8,14] and is starting to be recognized also in metazoans [15], but the effect of  $\text{Ca}^{2+}$ -mediated actin redistribution on mitochondrial positioning remains to be established.

Second, actin redistribution, according to Chakrabarti *et al.* [11], is a critical factor regulating mitochondrial  $\text{Ca}^{2+}$  uptake. While mitochondrial shape has been previously proposed to affect  $\text{Ca}^{2+}$  uptake [16,17], the question now arises as to whether cell shape and the cytoskeleton also have a role in regulating the mitochondrial  $\text{Ca}^{2+}$  signal.

Finally, how actin dynamics contribute to stress-mediated expansion of ER–mitochondria contacts [6], increased  $\text{Ca}^{2+}$  transfer to mitochondria, and mitochondrial fission [18] remains to be determined. ER–mitochondrial  $\text{Ca}^{2+}$  transfer can result in either metabolic adaptation to stress [19] or cell death due to mitochondrial  $\text{Ca}^{2+}$  overload (AU:?) [6]. Thus, remodelling of the actin cytoskeleton can affect cell fate decisions, signalling either survival or death. Metabolic activity is required for  $\text{Ca}^{2+}$ - and actin-induced mitochondrial fragmentation [11], suggesting cooperation with the adaptive response to stress. However, fragmentation of the mitochondrial network and Drp1 recruitment have also often been associated with cell death [18], and changes in the fusion–fission balance interfere with metabolic function [20]. Most likely, answers to these unresolved issues are not too far away.

## References

1. Barnhart, E.L. (2016). Mechanics of mitochondrial motility in neurons. *Curr. Opin. Cell Biol.* 38, 90–99.

2. Murley, A., and Nunnari, J. (2016). The emerging network of mitochondria-organelle contacts. *Mol. Cell* 61, 648–653.
3. Pernas, L., and Scorrano, L. (2016). Mito-morphosis: mitochondrial fusion, fission, and cristae remodeling as key mediators of cellular function. *Annu. Rev. Physiol.* 78, 505–531.
4. Lewis, S.C., Uchiyama, L.F., and Nunnari, J. (2016). ER-mitochondria contacts couple mtDNA synthesis with mitochondrial division in human cells. *Science* 353, aaf5549.
5. Kornmann, B., Osman, C., and Walter, P. (2011). The conserved GTPase Gem1 regulates endoplasmic reticulum-mitochondria connections. *Proc. Natl. Acad. Sci. USA* 108, 14151–14156.
6. Chami, M., Oulès, B., Szabadkai, G., Tacine, R., Rizzuto, R., and Paterlini-Bréchet, P. (2008). Role of SERCA1 truncated isoform in the proapoptotic calcium transfer from ER to mitochondria during ER stress. *Mol. Cell* 32, 641–651.
7. Cho, B., Cho, H.M., Jo, Y., Kim, H.D., Song, M., Moon, C., Kim, H., Kim, K., Sesaki, H., Rhyu, I.J., *et al.* (2017). Constriction of the mitochondrial inner compartment is a priming event for mitochondrial division. *Nat. Commun.* 8, 15754.
8. Mishra, P., and Chan, D.C. (2014). Mitochondrial dynamics and inheritance during cell division, development and disease. *Nat. Rev. Mol. Cell Biol.* 15, 634–646.
9. Korobova, F., Ramabhadran, V., and Higgs, H.N. (2013). An actin-dependent step in mitochondrial fission mediated by the er-associated formin INF2. *Science* 339, 464–467.
10. Moore, A.S., Wong, Y.C., Simpson, C.L., and Holzbaur, E.L.F. (2016). Dynamic actin cycling through mitochondrial subpopulations locally regulates the fission–fusion balance within mitochondrial networks. *Nat. Commun.* 7, 12886.
11. Chakrabarti, R., Ji, W., Stan, R. V, Sanz, J.D.J., Ryan, T.A., and Higgs, H.N. (2017). INF2-mediated actin polymerization at the ER stimulates mitochondrial calcium uptake, inner membrane constriction, and division. *J. Cell Biol.* doi: 10.1083/jcb.201709111
12. Wales, P., Schuberth, C.E., Aufschnaiter, R., Fels, J., García-Aguilar, I., Janning, A., Dlugos, C.P., Schäfer-Herte, M., Klingner, C., Wälte, M., *et al.* (2016). Calcium-mediated actin reset (CaAR) mediates acute cell adaptations. *Elife* 5, 1–31.
13. Debattisti, V., Gerencser, A.A., Saotome, M., Das, S., and Hajnóczky, G. (2017). ROS control mitochondrial motility through p38 and the motor adaptor Miro/Trak. *Cell Rep.* 21, 1667–1680.
14. Frederick, R.L., and Shaw, J.M. (2007). Moving mitochondria: establishing distribution of an essential organelle. *Traffic* 8, 1668–1675.
15. Sheng, Z.-H., and Cai, Q. (2012). Mitochondrial transport in neurons: impact on synaptic

homeostasis and neurodegeneration. *Nat. Rev. Neurosci.* 13.

16. Szabadkai, G., Simoni, A.M., Chami, M., Wieckowski, M.R., Youle, R.J., and Rizzuto, R. (2004). Drp-1-dependent division of the mitochondrial network blocks intraorganellar Ca<sup>2+</sup> waves and protects against Ca<sup>2+</sup>-mediated apoptosis. *Mol. Cell* 16, 59–68.
17. Bianchi, K., Vandecasteele, G., Carli, C., Romagnoli, A., Szabadkai, G., and Rizzuto, R. (2006). Regulation of Ca<sup>2+</sup> signalling and Ca<sup>2+</sup>-mediated cell death by the transcriptional coactivator PGC-1alpha. *Cell Death Differ.* 13, 586–596.
18. Youle, R.J., and van der Bliek, A.M. (2012). Mitochondrial fission, fusion, and stress. *Science* 337, 1062–1065.
19. Bravo, R., Vicencio, J.M., Parra, V., Troncoso, R., Munoz, J.P., Bui, M., Quiroga, C., Rodriguez, A.E., Verdejo, H.E., Ferreira, J., *et al.* (2011). Increased ER-mitochondrial coupling promotes mitochondrial respiration and bioenergetics during early phases of ER stress. *J. Cell Sci.* 124, 2143–2152.
20. Hoitzing, H., Johnston, I.G., and Jones, N.S. (2015). What is the function of mitochondrial networks? A theoretical assessment of hypotheses and proposal for future research. *Bioessays* 37, 687–700.

**Figure 1. Regulation of functional mitochondrial dynamics modules by the Ca<sup>2+</sup>- and actin-mediated signalling pathway described by Chakrabarti *et al.* [11].**

1, Ca<sup>2+</sup>-mediated actin nucleation on the ER surface [9,11,12]; 1a, Ca<sup>2+</sup>-mediated arrest of mitochondrial movements [13]; 2, actin-mediated extension of ER–mitochondria contacts and outer mitochondrial membrane fission [9–11]; 3, enhanced ER–mitochondrial Ca<sup>2+</sup> transfer via the mitochondrial calcium uniporter [6,11]; 4, inner mitochondrial membrane fission mediated by mitochondrial matrix Ca<sup>2+</sup> [7,11]; 5, coupling of inner and outer mitochondrial membrane fission [7,11]. : what is the difference between the grey and black arrows? Please clarify what is denoted by the question marks.