Drying process	Age of leaves	Moisture removal	Dried leaf color		
		(%)	Lightness	Red	Yellow
			(L^*)	(<i>a*</i>)	(b*)
Freeze dry	Young	92.1 ± 0.2^{b}			
	Developing	$92.2{\pm}0.1^{ab}$	64.43 ± 0.01^{a}	-4.54±0.01 ^c	25.08±0.01 ^a
	Mature	92.6±0.3 ^{ab}			
Microwave	Young	92.7±0.5 ^{ab}			
	Developing	92.8 ± 0.3^{ab}	$44.75 {\pm} 0.01^{b}$	-0.75 ± 0.02^{b}	$22.75{\pm}0.01^{b}$
	Mature	93.1±0.4 ^{ab}			
Oven	Young	92.8 ± 0.2^{ab}			
	Developing	92.9 ± 0.4^{ab}	37.17±0.02 ^c	$2.79{\pm}0.02^{a}$	18.17±0.01 ^c
	Mature	93.2±0.4 ^a			

Table 1 Colour and moisture removal (%) of G. pseudochina leaves with various leaf ages and drying methods.

Different letter(s) (a-c) in same column are significant differences according to Scheffe's test (p < 0.05). Data are given as means $\pm SD$ (n = 3).



Fig. 1 TPC of *G. pseudochina* leaf extracts per g dry weight (a) and per g crude extract (b), TFC of *G. pseudochina* leaf extracts per g dry weight (c) and per g crude extract (d), crude content (e) and 50% of FRSA activity (IC₅₀) (f) prepared with different drying processes, leaf ages and polarity of solvents. Abbreviations of sample names: first letter, F is freeze dry, M is microwave and O is oven; second letter, Y is young leaf, D is developing leaf and M is mature leaf. Different letter (s) (a-p) are significant differences according to Scheffe's test (p < 0.05). Data are presented as the means $\pm SD$ (n = 3).

Table 2 Summary of TPC and TFC in *G. pseudochina* leaf extracts obtained through different drying processes and leaf ages.

Drying	Leaf age	Content/ g dry weight			Content/ g crude extract		
process		TPC	TFC	Crude content	TPC	TFC	
		(µmol CAE)	(µmol ECE)	(mg)	(mmol CAE)	(mmol ECE)	
	Young	122.41±0.83 ^b	91.78±0.61 ^b	192.50±1.08 ^{cd}	2.14 ± 0.02^{b}	1.73±0.04 ^c	
Freeze dry	Developing	114.78 ± 0.22^{c}	$92.74{\pm}0.64^{b}$	$206.67 {\pm} 0.36^{b}$	2.06 ± 0.00^{bc}	1.82±0.01 ^c	
	Mature	95.60 ± 1.81^{d}	$70.39{\pm}1.26^{d}$	186.25±2.86 ^{cd}	1.77 ± 0.02^{d}	1.36 ± 0.02^{d}	
	Young	108.99±0.25 ^c	83.05±0.97°	216.46±0.72 ^a	1.75 ± 0.00^{d}	1.71±0.01 ^c	
Microwave	Developing	122.49 ± 1.24^{b}	81.01±0.52 ^c	224.17±4.61 ^a	1.96±0.05 ^c	$1.81{\pm}0.02^{b}$	
	Mature	157.18±3.25 ^a	110.85±0.83 ^a	217.71±0.95 ^a	2.70±0.06 ^a	2.00 ± 0.02^{a}	
	Young	35.77 ± 0.27^{f}	26.17±0.73 ^e	160.83±3.15 ^e	0.63 ± 0.02^{f}	0.86±0.04 ^e	
Oven	Developing	38.14 ± 0.68^{f}	27.19 ± 0.34^{e}	194.17±2.37°	$0.71 {\pm} 0.01^{\rm f}$	$0.94{\pm}0.03^{e}$	
	Mature	48.27±0.69 ^e	28.32 ± 0.57^{e}	$184.58{\pm}1.57^{d}$	0.88 ± 0.01^{e}	$0.69{\pm}0.01^{\rm f}$	

Different letter(s) (a-f) in same column are significant differences according to Scheffe's test (p < 0.05). Data are given as means $\pm SD$ (n = 3)



Fig. 2 Normalized HPLC chromatograms with retention times of (a, b) standards of phenolic compounds, and *G. pseudochina* extracts from continuous extracts with ethyl acetate, ethanol and 50% methanol of (c-e) the freeze-dried leaves, (f-h) microwave-dried leaves and (i-k) oven-dried leaves at various leaf ages.

Table 3 TPC, TFC, 50% free radical scavenging activity (IC₅₀), CGA, CA and RUT content of the MLM extracts from separate extractions with various ethanol concentrations of 25, 50, 75 and 100%.

	TPC	TFC	Crude	IC ₅₀ of FRSA	CGA	CA	RUT
Solvent	(µmol CAE/ g	(µmol ECE/ g	(g/ g dry	(µg crude	(mg/ g crude	(mg/ g crude	(mg/ g crude
	dry weight)	dry weight)	weight)	extract/ ml)	extract)	extract)	extract)
25% Ethanol	59.17±6.36 ^a	70.44±6.92 ^b	0.18±0.02 ^a	83.66±2.51ª	7.49±0.13°	4.23±0.14 ^a	< LOQ*
50% Ethanol	67.01±5.32ª	94.76±4.39ª	0.09 ± 0.01^{b}	76.07±8.21ª	15.04 ± 0.74^{b}	2.22±0.17 ^b	7.16±0.15°
75% Ethanol	60.28±4.80ª	89.26±6.50 ^a	0.09 ± 0.01^{b}	82.58 ± 4.65^{a}	16.85±0.35 ^a	1.33±0.03°	8.69 ± 0.75^{b}
100% Ethanol	27.02±3.97 ^b	20.18±1.20 ^c	0.04±0.00°	102.49±4.51 ^b	5.67 ± 0.14^{d}	1.01±0.04°	15.71±0.25 ^a

Different letter (s) (a-d) in the same column are significant differences according to Scheffe's test (p < 0.05). Data are presented as the means $\pm SD$ (n = 3). * The LOQ of RUT is 0.46 µg/ml.



Fig. 3 LC-ESI base peak chromatograms (BPC) of the MLM extracts from separate extractions with (a) 25% ethanol and (b) 100% ethanol. For major peak assignments, see Table 4.

Peak RT		T ESI-MS m/z		Tentative identification	Formula	Error
no.	(min)	IN) [M-H] MS/MS fragment		-		(ppm)
1	9.58	385.04	277.03,204.99,73.02	Unidentified	-	-
2	9.89	353.08	191.05,135.04	Caffeoyl quinic acid isomer1	$C_{16}H_{18}O_9$	1.15
3	10.02	423.15	363.12,113.02	(+)-Tephropurpurin	$C_{24}H_{24}O_7$	-13.37
4	11.08	353.08	191.05,135.04	Caffeoyl quinic acid isomer2 (CGA)*	$C_{16}H_{18}O_9$	1.72
5	11.32	439.18	393.17,163.05,205.06	1,3,8-Trihydroxy-4-methyl- 2,7 diprenylxanthone	$C_{24}H_{26}O_5$	-12.09
6	11.68	421.16	341.11,213.04	2-(2,4-Dihydroxyphenyl)-5- hydroxy-8-methyl-8-(4- methyl-3-penten-1-yl)-2,3- dihydro-4H,8H-pyrano[2,3- f]chromen-4-one	C ₂₅ H ₂₆ O ₆	9.17
7	11.91	353.08	191.05	Caffeoyl quinic acid isomer3	$C_{16}H_{18}O_9$	1.15
8	12.57	179.03	135.04	CA*	$C_9H_8O_4$	8.28
9	12.99	609.14	463.08,300.02,178.99,151.00	Quercetin rutinoside (RUT)*	$C_{27}H_{30}O_{16}$	-1.14
10	13.45	367.10	179.03,135.04,99.01	3-O-Caffeoyl-1-O- methylquinic acid	$C_{17}H_{20}O_9$	6.14
11	13.76	593.15	285.03,327.04,535.21,417.24	Kaempferol rutinoside	$C_{27}H_{30}O_{15}$	-0.35
12	13.94	713.47	677.49,313.06,147.04	Unknown-C-glycoside	$C_{35}H_{70}O_{14}$	-3.51
13	14.29	826.55	790.57,656.96	Unidentified	-	-
14	14.76	515.11	353.08,173.04,179.03	Dicaffeoyl quinic acid isomer1	$C_{25}H_{24}O_{12}$	0.97
15	15.02	515.11	353.08,173.04,179.03	Dicaffeoyl quinic acid isomer2	$C_{25}H_{24}O_{12}$	0.97
16	15.55	313.07	313.07	3,4-Dihydroxycinnamoyl- (Z)-2-(3,4-dihydroxyphenyl) ethenol	$C_{17}H_{14}O_6$	4.99
17	16.49	463.25	417.24,161.04	1-(9Z-octadecenoyl)-sn- glycero-2,3-cyclic phosphate	$C_{21}H_{39}O_6P$	-13.30
18	17.16	497.21	429.20,249.14,119.0313,59.01	Unidentified	-	-
19	18.17	301.03	151.00, 121.02	Quercetin	$C_{15}H_{10}O_7$	4.23
20	18.54	623.11	311.05, 265.04, 147.04, 109.02	5-Hydroxy-2'-methoxy-6,7- methylenedioxyisoflavone	$C_{17}H_{12}O_6$	1.97

Table 4 LC-ESI-QTOF-MS/MS analysis of phenolic compounds from the MLM extracts from separateextractions with 25 and 100% ethanol.

* Peaks are compared with standard compounds.

Table 5 Total pyrrolizidine alkaloid content (TPAsC) in *G. pseudochina* leaf extracts prepared from different drying processes (freeze drying and microwave and oven drying) and serial extraction with 25% and 50% ethanol.

Solvent fraction and dried leaf sample	mmol MCTE/ g crude extract
25% Ethanol MLF	0.004±0.001 ^e
50% Ethanol MLF	0.142 ± 0.015^{d}
25% Ethanol MLM	0.003±0.002 ^e
50% Ethanol MLM	0.065±0.011°
25% Ethanol MLO	0.233±0.038 ^b
50% Ethanol MLO	0.684 ± 0.125^{a}

Different letter(s) (a-e) in same column are significant differences according to Scheffe's test (p < 0.05). Data are given as means $\pm SD$ (n = 3).

	a quantity of each marrier comp	
Samples	Cytotoxicity IC	₅₀ values (μg/ml)
	Non TNF-α	TNF-α
EMLM	680.80±23.98 ^{c*}	744.02±62.2 ^{c*}
CGA	$180.61 \pm 7.09^{d*}$	284.50±45.76 ^{d**}

 $36.18 \pm 2.78^{e^*}$

4393.41±436.11^{a*}

 $1682.78 \pm 107.96^{b^*}$

 $2.69 \pm 0.32^{f^*}$

CA

RUT

PCA

PTX (positive control)

Table 6 Cytotoxicity of the EMLM extract and marker compounds on HaCaT cells, non-stimulated and stimulated by TNF- α , and quantity of each marker compound in the EMLM extract.

Different letter(s) (a-f) in same column are significant differences according to Scheffe's test (p < 0.05). Different symbols (*, **) in same column are significant differences according to T-test (p < 0.05). Data are given as means $\pm SD$ (n = 3).

 $60.19{\pm}1.76^{e^{**}}$

 $1497.99 \pm 205.50^{b^{**}}$

 $2840.76 \pm 174.12^{a^{**}}$

 $1.84{\pm}0.49^{f^*}$

TNF-α stimulation	Sample for treatment	Concentration (µg/ml)	IL-8 contents (pg/ml)
Non TNF-α	Control	-	10.17±2.01 ^g
	Control non DMSO	-	342.80±34.62ª
	Control 0.7% DMSO	-	322.88±36.03 ^{ab}
	FMI M crudo ovtrocto	375	133.79±18.18 ^{ef}
	EMEM crude extracts	750	148.82±28.15 ^e
		140	284.82±35.92 ^{bc}
	CUA	240	90.10 ± 16.39^{f}
TNF-α	CA	30	254.60±42.63°
	CA	60	87.19 ± 11.76^{f}
	DUT	750	115.17±20.50 ^{ef}
	KU I	1500	93.68 ± 14.33^{f}
	DCA	1400	369.71±41.43 ^a
	r CA	2800	201.21±24.38 ^d
	CUR (positive control)	50	14.35±9.78 ^g

Table 7 Interleukin 8 (IL-8) content in HaCaT cell lysate after treatment with the EMLM extract and marker compounds.

Different letter(s) (a-f) in same column are significant differences according to Duncan's test (p < 0.05). Data are given as means $\pm SD$ (n = 3).

Treatment	Nucleus	RelA	RelB	Overlay	
(-)TNF-α Control	*** \$/ 6			** <i>(</i>)	
(+)TNF-α Control			1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1	1 0 0 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1	
(+)TNF-α EMLM 375 µg/ml	8 8			8 	
(+)TNF-α EMLM 750 µg/ml	\$*** 3 \$*		2° 1 31	9* ± 9	
(+)TNF-α CGA 140 μg/ml	Har I		10 5 × 11	1 2 2 4 4 4 4 4 4 4 4 4 4 4 4 4 4 4 4 4	
(+)TNF-α CGA 280 μg/ml	• • •	-	6. S. • •	e. E.	
(+)TNF-α CA 30 μg/ml	ten		Ser.	100	
(+)TNF-α CA 60 µg/ml	194 			1.4	
(+)TNF-α RUT 750 μg/ml					
(+)TNF-α RUT 1500 µg/ml	£		2) ·	8	
(+)TNF-α PCA 1400 μg/ml	4 - 🕅		4	4	
(+)TNF-α PCA 2800 μg/ml	1			4 8	
(+)TNF-α CUR 50 μg/ml	100			Contraction of the second seco	

Fig. 4 Localization of RelA and RelB on HaCaT cells due to TNF- α stimulation. HaCaT cells were pre-treated with 50 ng/ml of TNF- α for 12 h and treated with the EMLM extracts, marker compounds (CGA, CA, PCA and RUT) and CUR (positive control) for 24 h.