

Coadaptation of mitochondrial and nuclear genes, and the cost of mother's curse

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ABSTRACT

Strict maternal inheritance renders the mitochondrial genome susceptible to accumulating mutations that harm males, but are otherwise benign or beneficial for females. This “mother’s curse” effect can degrade male survival and fertility if unopposed by counteracting evolutionary processes. Coadaptation between nuclear and mitochondrial genomes – with nuclear genes evolving to compensate for male-harming mitochondrial substitutions – may ultimately resolve mother’s curse. However, males are still expected to incur a transient fitness cost during mito-nuclear coevolution, and it remains unclear how severe such costs should be. We present a population genetic analysis of mito-nuclear coadaptation to resolve mother’s curse effects, and show that the magnitude of the “male mitochondrial load” – the negative impact of mitochondrial substitutions on male fitness components – may be large, even when genetic variation for compensatory evolution is abundant. We also find that the male load is surprisingly sensitive to population size: male fitness costs of mito-nuclear coevolution are particularly pronounced in both small and large populations, and minimized in populations of intermediate size. Our results reveal complex interactions between demography and genetic constraints during the resolution of mother’s curse, suggesting potentially widespread species differences in susceptibility to mother’s curse effects.

Keywords: epistasis, sex-specific selection, sexual conflict, adaptation, compensatory evolution

INTRODUCTION

With rare exceptions, mitochondria are maternally inherited, with little-to-no transmission from fathers to offspring (Birky 1995). This unique inheritance pattern may lead to unusual evolutionary dynamics of mitochondrial genes, compared to genes that are encoded within the nuclear genome (Frank 1989; Frank and Hurst 1996; Gemmell et al. 2004). Whereas strict maternal transmission ensures that the mitochondrial genome is evolutionarily responsive to natural selection in females, it inhibits evolutionary responses to selection in males. Mitochondrial DNA (mtDNA) mutations that reduce the survival, mating success, or fertility of males may therefore accumulate within a population, as long as these mutations do not reduce female fitness (*i.e.*, they may be benign or beneficial for females; Frank 1989; Frank and Hurst 1996; Gemmell et al. 2004; Dowling 2014; Beekman et al. 2014). The evolutionary accumulation of mtDNA mutations with male-harming effects, attributable to the maternal inheritance of mitochondria, has been dubbed “mother’s curse” (Gemmell et al. 2004).

Several observations suggest that mother’s curse influences the evolution of mitochondrial genomes, the genetic basis of female and male adaptations, and the manifestation of disease. In plants, cytoplasmic male sterility (CMS) is widespread, typically has a mitochondrial genetic basis, and is important in both the evolution of plant breeding systems and production of hybrid seeds in commercial agriculture (Frank 1989; Schnable and Wise 1998; Chase 2007). Sex-specific fitness effects of mitochondrial genetic variation have received less attention in animal systems, yet even so, some mtDNA mutations are known to have male-biased effects on animal fertility (*e.g.*, Nakada et al. 2006; Smith et al. 2010; Patel et al. 2016; Martikainen et al. 2017), and neurodegenerative disease (*e.g.*, *Leber’s Optical Hereditary Neuropathy*: Wallace et al. 1988; Milot et al. 2017). Studies in *Drosophila* have identified signals of mother’s curse in patterns of: (1) male-biased effects of mitochondrial

haplotypes on the transcription of nuclear-encoded genes (Innocenti et al. 2011; but see Mossman et al. 2016a, 2017); (2) male-biased genetic variance in ageing (Camus et al. 2012, 2015); and (3) male-limited genetic variation for sterility (Clancy et al. 2011; Patel et al. 2016). Overall, the pervasiveness of mother's curse effects among species, populations, and environmental contexts remains unclear (*e.g.*, Mossman et al. 2016b; Eyre-Walker 2017). Nevertheless, current examples suggest that at least a fraction of mtDNA mutations have male-biased or male-limited fitness costs (*e.g.*, Beekman et al. 2014; Wolff et al. 2016; Patel et al. 2016). The evolutionary logic of the mother's curse hypothesis predicts that such mutations are likely to accumulate within populations and depress male fitness components, unless they are countered by evolutionary mechanisms that oppose their spread, or compensate for their negative effects in males (Frank and Hurst 1996; Yee et al. 2013; Reinhardt et al. 2013).

Two general evolutionary scenarios can potentially resolve mother's curse, and rescue male fitness declines caused by mtDNA mutation accumulation. First, some forms of inbreeding, kin selection, assortative mating, and paternal mitochondrial transmission generate direct purifying selection against mtDNA mutations with male-limited costs (Wade and Brandvain 2009; Unckless and Herren 2009; Hedrick 2012; Zhang et al. 2012; Kuijper et al. 2015). These processes provide scope for the elimination of male-harming mtDNA mutations, although purifying selection through males is likely to remain inefficient (Hedrick 2012; Engelstädter and Charlat 2006), particularly when male-harming alleles have small fitness effects (see Wade and Brandvain 2009; Unckless and Herron 2009). Second, changes to the nuclear genome may compensate for the fitness costs of mtDNA alleles by masking their male fitness consequences (Yee et al. 2013; Wade 2014; Beekman et al. 2014). Mitochondrial and nuclear genomes are functionally coupled through their shared roles in encoding the cellular machinery of energy production (see Rand et al. 2004; Burton and

Baretto 2012; Wolff et al. 2014), and this close coupling provides a mechanistic basis for mito-nuclear genetic interactions. Mito-nuclear interactions for fitness – including sex-specific fitness components – have been reported in several animal systems (*e.g.*, Rand et al. 2001; James and Ballard 2003; Roubertoux et al. 2003; Ellison and Burton 2008; Dowling et al. 2010; Burton and Barreto 2012; Meiklejohn et al. 2013; Yee et al. 2013, 2015; Dobler et al. 2014 Immonen et al. 2016). Likewise, in plant systems that harbor mtDNA alleles that cause CMS, nuclear genes play central roles in restoring male fertility (Chase 2007).

Whether evolutionary changes in the nuclear genome can compensate for male-harming mtDNA mutants is not controversial. However, the speed and efficiency with which nuclear genome evolution resolves mother’s curse remains an open question (see Wade 2014). Several models have considered the coevolutionary dynamics of male-harming mitochondrial mutations and compensatory alleles in the nuclear genome (*e.g.*, Charlesworth 1981; Gregorius and Ross 1984; Frank 1989; Babcock and Asmussen 1996; Rand et al. 2001; Wade 2014). Theory of mito-nuclear coevolution is particularly well developed in contexts of CMS, where conditions for the evolutionary invasion, maintenance, and fixation of male-sterility genotypes are well characterized (reviewed in Jacobs and Wade 2003). On the other hand, prior theory largely focuses on the deterministic dynamics and long-term evolutionary equilibria of mitochondrial and nuclear genes. Much less attention has been given to non-equilibrium dynamics of mito-nuclear coevolution, where mito-nuclear dynamics should generate transient reductions in male fitness components prior to the ultimate resolution of mother’s curse. There is currently no clear expectation for the severity of male fitness costs that arise from bouts of mito-nuclear coevolution, nor expectations for the impacts of genetic drift and population size on the manifestation of mother’s curse effects.

Here, we present a theoretical population genetic analysis of mito-nuclear coevolution in the context of mother’s curse. We specifically quantify the “male mitochondrial load” –

the reduction in male fitness components during coevolutionary cycles between male-harming mtDNA substitutions and nuclear compensatory substitutions that restore the fitness components. Our formulation of load is conceptually similar to “substitution loads” or “lag loads” that arise during adaptation to a new environment (*e.g.*, Haldane 1957; Crow 1970; Maynard Smith 1976), with mtDNA divergence serving as an analog of environmental change. The theoretical framework also parallels a long tradition of compensatory evolution models in population genetics (Haldane 1931; Kimura 1985; Stephan 1996). Our results consider male-harming mtDNA substitutions that are fixed by positive selection in females (*i.e.*, sexually antagonistic mutations), as well as substitutions that are neutral in females and fixed by genetic drift. We show that the magnitude of the male mitochondrial load can be large relative to classical genetic loads. We also demonstrate that the load is highly sensitive to the effective population size of the species: the male mitochondrial load exhibits a non-monotonic relation with population size, which is minimized in intermediate-sized populations. These results shed new light on the contexts of selection and demography that are likely to elevate or reduce the impact of mother’s curse effects.

MODEL

Model structure and assumptions

Our baseline model follows the evolution of a pair of interacting mito-nuclear loci that affect male fitness. Derived (mutant) alleles at the mitochondrial locus are assumed to reduce male fitness relative to the ancestral genotype; these alleles may be neutral or beneficial for females. For simplicity, we assume that the nuclear (*i.e.*, compensatory) locus is male-limited in expression (alleles are neutral in females); we later revisit this assumption. Generations are discrete with a life-cycle of: (1) birth, (2) selection, (3) meiosis and mutation, followed by random mating, and (4) death of the adults.

Interacting loci have two alleles each. The mitochondrial locus has alleles M and m , and the nuclear locus has alleles A and a . M and A represent the ancestral (wild-type) alleles, whereas m and a are derived alleles. We assume that mitochondrial inheritance is strictly maternal (there is no paternal transmission). The nuclear locus is diploid, with biparental inheritance. Following prior models (Frank and Hurst 1996; Wade and Brandvain 2009; Unckless and Herren 2009; Kuijper et al. 2015), we assume that each individual in the population is homoplasmic for one of the two mitochondrial alleles. This simplifying assumption is reasonable under strict maternal inheritance, with mitochondria strongly bottlenecked during oogenesis; we return to this issue in the Discussion. Following the tradition of compensatory evolution models (*e.g.*, Stephan 1996), we assume that derived mutations are individually deleterious (fitness costs of s_m and t_m are associated with the m and a alleles, respectively), but beneficial for males in combination (see Table 1). Thus, AA/M and aa/m genotypes yield maximal fitness, and the remaining genotypic combinations impose a cost to males.

Each evolutionary cycle between a mitochondrial and nuclear locus initiates with the invasion – from a single, initial copy – of a derived, male-harming mitochondrial allele, m . Allele m evolves under positive selection in females or genetic drift until it is eventually fixed in the population. The nuclear locus evolves under recurrent mutation and selection in males. The population is initially fixed for the ancestral allele, A , which is beneficial for males in combination with the ancestral mitochondrial allele, M . For simplicity, we assume that mutations from A to a are unidirectional, with a rate of ν , per meiosis; introduction of back-mutation is trivial, and simply allows for the maintenance of rare A alleles in populations that become fixed for the m cytotype. The evolutionary cycle completes when both derived alleles (a and m) are fixed.

Quantifying the male mitochondrial load

Each cycle of mito-nuclear coevolution contributes to the male genetic load, as follows. Let t ($t \geq 0$) represent the number of generations following the initial appearance of a mitochondrial allele destined to eventually fix. Following the standard genetic load definition, the male load at generation t is:

$$l(t) = \frac{w_{\max} - \bar{w}_m(t)}{w_{\max}} = 1 - \bar{w}_m(t)$$

(1),

where $\bar{w}_m(t)$ is the mean of relative male fitness at generation t , and w_{\max} is the relative fitness of the best genotype for males (*i.e.*, *AAM* and *aam* from Table 1, where w_{\max} is scaled to one in our model). The cumulative contribution of the coevolutionary cycle to the male load is: $L = \sum_{t=0}^{\tau} l(t)$, where τ is the duration, in generations, of the cycle (*i.e.*, the time between the origin of the male-harming allele, m , to the eventual fixation of the derived mito-nuclear combination, am). From this, the average contribution of a single coevolutionary cycle to the male load is:

$$E[L] = E \left[\sum_{t=0}^{\tau} l(t) \right]$$

(2),

where $E[x]$ denotes the expectation. Biologically, $(N/2)E[L]$ represents the average number of males that are selectively eliminated during a typical bout of mito-nuclear coevolution, where $N/2$ is the number of males born in each generation (see Crow 1970; Ewens 2004, pp. 81-82). This concept of the male load parallels other forms of genetic load that arise during adaptation, particularly Haldane's (1957) "cost of selection", and Maynard Smith's (1976) "lag load" (see Charlesworth and Charlesworth 2010, p. 169). We return to this comparison further below.

Eq. (2) quantifies the cumulative contribution of single mito-nuclear coevolutionary cycles to the male load, but it does not take into account the tempo of male-harming mtDNA substitutions that initiate each cycle. To quantify the fitness cost to males, per generation, we can scale the cumulative load per coevolutionary cycle by the average number of generations between cycles of mito-nuclear coevolution (denoted as T_M):

$$\frac{E[L]}{T_M} = K \cdot E[L]$$

(3)

(Ewens 2004, pp. 81-82), where $K = 1/T_M$ is the rate of substitution of male-harming alleles in the mitochondrial genome. Eq. (3) applies when the timescale for resolution of mother's curse is faster than the tempo of male-harming mtDNA substitution. Otherwise mitochondrial function in males will systematically degenerate over time – a situation more closely aligned with models of mutational meltdown (*e.g.*, Lande 1994, 1998).

Model analysis

Our analytical results assume that selection coefficients are small ($1 \gg s_m, t_m$), and population-scaled selection is strong ($Ns_m, Nt_m \gg 1$, where N is the size of a Wright-Fisher population). Under random mating, deviations from Hardy-Weinberg equilibrium and mito-nuclear linkage disequilibrium are negligible. We assume throughout that the effective size of a mtDNA locus is one-quarter the size of a diploid, nuclear locus ($2N$ for a diploid locus; $N/2$ for a mtDNA locus).

At the beginning of a coevolutionary cycle (arbitrarily labeled as generation $t = 0$), a male-harming m allele begins to spread within the population until it is eventually fixed. The frequency of m in the zygotes at generation t ($t \geq 0$) is q_{mt} . Following prior theory (*e.g.*, Maynard Smith 1971, 1976; Betancourt et al. 2004; Ewens 2004, p. 147; Takahasi 2009), we use deterministic models to approximate the expected evolutionary trajectory of the m allele,

conditioned on its eventual fixation (see the Supplementary Material). For the case of neutrally evolving mtDNA alleles ($s_f = 0$; Table 1), the expected frequency of m at generation t is:

$$q_{mt} = 1 - (1 - q_{m0})e^{-2t/N}$$

(4a)

(see Ewens 2004, p. 147; Takahasi 2009), where the initial frequency of m is $q_{m0} = 2/N$. For the case of a sexually antagonistic m allele ($s_f > 0$), the expected frequency of m at generation t is:

$$q_{mt} = \frac{q_{m0}e^{s_f t}}{1 - q_{m0} + q_{m0}e^{s_f t}}$$

(4b)

(e.g., Otto and Day 2007), where $q_{m0} = (2/N)(1/2s_f) = 1/Ns_f \ll 1$ is the “effective” initial frequency of the allele (see Maynard Smith 1971, 1976; Betancourt et al. 2004 and Orr and Unckless 2014 discuss the “effective” initial frequency of sweeping beneficial alleles).

The frequency of the m allele mediates selection at the interacting nuclear locus.

Consider an a allele that segregates at a frequency of q_{at} in zygotes of generation t . Assuming Hardy-Weinberg and linkage equilibrium in the zygotes (as stated above), the expected frequencies of the three nuclear genotypes in adult males (after selection in generation t) are:

$$f_{AA} = \frac{(1 - q_{at})^2(1 - q_{mt}s_m)}{\bar{w}_{nuc}}$$

$$f_{Aa} = \frac{q_{at}(1 - q_{at})(2 - q_{mt}s_m - (1 - q_{mt})t_m)}{\bar{w}_{nuc}}$$

$$f_{aa} = \frac{q_{at}^2(1 - (1 - q_{mt})t_m)}{\bar{w}_{nuc}}$$

(5),

where \bar{w}_{nuc} is the sum of the numerators of f_{AA} , f_{Aa} , and f_{aa} . The expected change in frequency of a due to selection in males is:

$$\Delta q_{sel}(\text{males}) = \frac{q_{at}(1 - q_{at})(q_{mt}s_m - (1 - q_{mt})t_m)}{2\bar{w}_{nuc}}$$

(6).

The total frequency change across a single generation, owing to selection, will be $\Delta q_{sel} = [\Delta q_{sel}(\text{males}) + \Delta q_{sel}(\text{females})]/2$, where $\Delta q_{sel}(\text{females})$ refers to the change in a over one generation as a consequence of selection in females ($\Delta q_{sel}(\text{females}) = 0$ when the a is neutral for females). From eq. (6), note that selection in males favors the a allele once m exceeds the critical frequency $q_{mt} = t_m/(s_m + t_m)$, and otherwise allele A is favored. Given the assumption of small selection coefficients and mutation rate of v (see above), the total expected change in the frequency of a will be: $E[\Delta q_{tot}] = \Delta q_{sel} + (1 - q_{at})v$.

Mathematical analysis of the model focuses on two idealized scenarios. First, under sufficiently weak mutation at the compensatory locus, the timescale to resolve mother's curse is slow relative to the time to fixation of m . The load is then dominated by a lag until a compensatory mutation arises and invades the population. The male fitness component effectively "jumps" between two states: a high state (relative fitness of one), and a low state (relative fitness of $1 - s_m$; see Fig. 1). Assuming that the waiting time for a compensatory substitution is smaller than the interval between male-harming mtDNA substitutions ($T_C < T_M$, where T_C is the mean time to compensation following the fixation of the male-harming allele; see above), the fraction of time spent in the low-fitness state will be $f_{low} = T_C/T_M$, and male load, scaled per generation (eq. (3)), is $E[L \mid \text{slow}]/T_M = s_m f_{low}$.

Second, with a sufficiently high compensatory mutation rate, a alleles will spread rapid once they are favoured. For the neutral model of mtDNA substitution, the evolutionary dynamics of the m allele are slow relative to the rate of spread of a , and the latter is therefore likely to fix before the former. In such cases, we can use a separation of timescales approximation (e.g., Otto and Day 2007), and model the substitution process at the nuclear locus as if a fixes instantaneously once selection favors it. The cumulative load becomes:

$$E[L | \text{fast}] \approx s_m \int_0^{t_{pos}} q_{mt} dt + t_m \int_{t_{pos}}^{\infty} (1 - q_{mt}) dt$$

(7a),

where q_{mt} is given by eq. (4a), and t_{pos} is the time until positive selection favors the compensatory allele (the value of t for which $q_{mt} > t_m/(s_m + t_m)$, based on eq. (4a)).

When mtDNA mutations are positively selected in females, a compensatory allele is unlikely to invade until the m cytotype approaches fixation, but nevertheless, the load may be dominated by the time to fixation of the compensatory allele rather than the time until its initial appearance. In this case, the total load will be:

$$E[L] \approx E[L | \text{slow}] + s_m \int_0^{\infty} (1 - q_{at}) dt$$

(7b),

where q_{at} tracks the frequency of a compensatory allele that eventually becomes fixed.

Simulations

Analytical results were verified using forward simulations that incorporate selection, mutation, and multinomial sampling in a Wright-Fisher population of effective size N . The simulations track all six possible genotypes – AAM , AAm , AaM , Aam , aaM , aam – in each sex, allowing for the buildup of linkage disequilibrium between mitochondrial and nuclear loci. Each simulation run was initiated with the population fixed for the A allele, and initial frequencies of $2/N$ and $1 - 2/N$ for M and m alleles, respectively. Each run completes with the fixation of both a and m . For a given parameter set of s_f , s_m , t_m , N , and v , the average over simulation runs provides an estimate the exact cumulative contribution of mito-nuclear coevolution to the male load (eq. (2), above). All simulations were run in R, version 3.3.0 (R Core Team 2016). Additional details can be found in the Supplementary Material.

RESULTS

Male-harming mtDNA substitutions with neutral effects in females

When male-harming mtDNA substitutions are neutral for females ($s_f = 0$), the evolutionary dynamics of mitochondrial and nuclear substitutions depend on the magnitude of the compound parameter, N^2vs_m . When $N^2vs_m \ll 1$, each male-harming mtDNA allele fixes rapidly relative to the waiting time to invasion and fixation of its corresponding compensatory allele. Intuitively, this is because the mean time to fixation of a neutral mtDNA mutation is N generations, whereas the time to invasion of a favoured compensatory allele is roughly $T_C = 1/(Nvs_m)$ generations. Thus, when $N \ll T_C$ (or $N^2vs_m \ll 1$), the contribution to the male load will be:

$$E[L|\text{slow}] \approx \frac{1}{Nv}$$

(8a).

Note that eq. (8a) is inversely proportional to population size, reflecting adaptation limited by the availability of compensatory mutations. At the opposite extreme, when compensatory evolution is fast relative to the timescale of fixation for a mtDNA allele ($N^2vs_m \gg 1$, justifying eq. (7a)), the contribution to the male load is:

$$E[L|\text{fast}] \approx \frac{Ns_m}{2} \ln(1 + t_m/s_m)$$

(8b)

(see the Supplementary Material). From eq. (8b), we see that in the limit of fast compensatory change, the male load now increases with population size. This positive relation between N and load reflects the impact of the fixation time of male harming mtDNA alleles on the male load. Coevolution is no longer limited by the availability of genetic variation at compensatory loci; rather, the resolution of the male load is limited by the time until substitution of the mtDNA allele, which is proportional to N .

Forward simulations across a spectrum of N^2vs_m show that eqs. (8a) and (8b) work well within their relevant regions of parameter space (Fig. 2A; Supplementary Material), and their sum provides a general approximation for the male load:

$$E[L] \approx \frac{1}{Nv} + \frac{Ns_m}{2} \ln(1 + t_m/s_m)$$

(9).

Eq. (9) implies that for a given strength of selection (s_m, t_m) and compensatory mutation rate (v), there is a critical effective population size that minimizes the male mitochondrial load.

This critical population size corresponds to:

$$N_{crit} = \sqrt{\frac{2}{s_m v \ln(1 + t_m/s_m)}}$$

(10).

Below the threshold, there is a negative relation between N and the male load; above the threshold, the relation is positive (Fig. 2B). Intuitively, the critical population size increases as the compensatory mutation rate decreases, reflecting the greater influx of compensatory alleles in larger populations. A similar result applies if we rescale $E[L]$ relative to the tempo at which male-harming/female-neutral mutations become fixed (*i.e.*, eq. (3)). Following standard population genetics theory (*e.g.*, Kimura and Ohta 1971), the average interval between neutrally evolving mtDNA substitutions is simply $T_M = 1/U_N$, where U_N is the mitochondrial genomic mutation rate to male-harming alleles that are neutral to females. Consequently, the scaled load is $E[L]/T_M = U_N E[L]$, which is minimized at the N_{crit} in eq. (10).

Sexually antagonistic mtDNA substitutions

When male-harming mtDNA substitutions are sexually antagonistic (*i.e.*, they benefit females), mito-nuclear substitutions are much more likely to fix sequentially because positive

selection on each mtDNA mutation speeds the time to its fixation. When the strength of selection in females is of similar order to selection in males (s_f/s_m is not too small), compensatory alleles rarely begin to spread before the male-harming mtDNA allele is fixed (or nearly fixed) in the population. From eq. (7b), the contribution to the male load becomes:

$$E[L] \approx \frac{1}{Nv} + 4\ln(Ns_m)$$

(11)

(see the Supplementary Material). Eq. (11) closely approximates the male load when s_m and s_f have a similar order of magnitude (see Fig. 3), and it underestimates the true load when compensatory mutation is strong and selection in females is weak (Nv is large; $s_m \gg s_f$; see the Supplementary Material).

From eq. (11), the impact of individual bouts of mito-nuclear coevolution on the male load is minimized at an intermediate population size: $N_{\text{crit}} = 1/4v$. The overall relation between population size and the male load also depends the rate of male-harming mtDNA substitutions (K), which may also be dependent on N (see Gillespie 2004; McCandlish and Stoltzfus 2014; Lanfear et al. 2014). Most models of adaptive substitution predict a positive relation between population size and the rate of evolution. Some models predict a positive, linear association between K and N (e.g.: Kimura 1979; McCandlish and Stoltzfus 2014), while others predict that K is, at most, a diminishing returns function of N (see Maynard Smith 1976; Gillespie 2004). We can generalize between these extremes by supposing that $K \propto N^b$, where b is a constant in $0 < b < 1$ that defines the strength of the diminishing return (e.g., K saturates at modest N when with b is small). The net load is therefore proportional to $N^b E[L]$, and the critical population size falls within the range:

$$\frac{1}{s_m e} < N_{\text{crit}} < \frac{1}{4v}$$

(12),

where $e \sim 2.718$ (note that $b = 1$ sets the lower bound of N_{crit} , whereas $b = 0$ sets the upper bound). As before, the male load is minimized at an intermediate population size, with N_{crit} decreasing as b increases.

DISCUSSION

Strict maternal inheritance of mitochondrial genes facilitates accumulation of male-harming mtDNA mutations, which poses a problem for the long-run viability and fertility of males (Frank and Hurst 1996; Beekman et al. 2014). This is the dilemma of mother's curse (Gemmell et al. 2004). Although compensatory evolution in the nuclear genome may ultimately resolve mother's curse, our theoretical analysis suggests that transient reductions in male fitness components are far from trivial – even in cases where the genetic architecture of compensatory adaptation is evolutionarily permissive.

Our results build upon a recent theoretical argument by Wade (2014), who reasoned that compensatory evolution at nuclear genes provides an evolutionarily inefficient means for resolving mother's curse. By quantifying this inefficiency, we show that costs of mother's curse – though ultimately resolved – may nevertheless be substantial, particularly when benchmarked against other forms of genetic load. Haldane's (1957) “cost of selection” (or substitution load; see Crow 1970) – the transient load that arises during the substitution of a beneficial mutation – provides the most straightforward point of contrast with our results. Following standard theory (Haldane 1957; Crow and Kimura 1970, p. 247; Charlesworth and Charlesworth 2010, p. 169; see the Supplementary Material), the load contributed by a mitochondrial substitution that equally benefits both sexes is $C = -\ln(p_0)$, where p_0 is the initial frequency of the beneficial allele. When adaptation uses new, beneficial mutations, then $p_0 \sim 1/Ns$ for a mitochondrial mutant that eventually fixes (Maynard Smith 1971, 1976; see above), and the substitution load becomes $C = \ln(Ns)$. The male mitochondrial load easily

exceeds the substitution load (C) over a wide range of parameter conditions (see Figs. 2 and 3). For example, sexually antagonistic mtDNA mutations contribute to a male load of, at minimum, $4\ln(Ns_m)$ (see eq. (11)), which is four-fold higher than Haldane's substitution load, given a similar order of selection ($s \sim s_m$). The effect is amplified for male-harming mtDNA mutations that are fixed by drift: while C increases logarithmically with Ns , the baseline load due to male-limited mtDNA substitutions increases linearly with Ns_m (eq. (9)), so that the latter may be orders of magnitude greater than the former.

Scope of the model and potential for additional costs of mtDNA inheritance

Our model focuses on an idealized scenario of mito-nuclear coadaptation in which: (1) compensatory loci are male-limited with additive expression in males (Table 1); (2) nuclear loci are diploid; and (3) individuals are homoplasmic. We also focused on the fitness costs of mtDNA mutations that eventually fix, ignoring fitness costs of segregating mutations that are eventually lost due to purifying selection or genetic drift (but see Appendix V of the Supplementary Material). As we discuss below, these assumptions are generally conservative, and should – if anything – underestimate the true magnitude of the load arising from mother's curse.

Effects of pleiotropy at compensatory loci. Our model provides baseline predictions for the male load when compensatory loci are completely unconstrained by pleiotropic fitness effects in females. When compensatory alleles are expressed in both sexes, their effects are likely to be sexually antagonistic (reducing female fitness). This sexual antagonism in the nuclear genome should dampen the evolutionary capacity of nuclear loci to quickly resolve mother's curse, by reducing the proportion of compensatory alleles that are favoured by selection (effectively reducing ν), and dampening the rate at which favoured alleles spread to fixation (*e.g.*, Otto 2004; Connallon and Clark 2013).

Effects of dominance at nuclear genes. Although additivity is assumed in our models, genetic incompatibilities may instead exhibit dominance, as they often do in contexts of inter-specific genetic incompatibilities (Coyne and Orr 2004). Non-additive effects of incompatibilities should have two consequences for mito-nuclear coadaptation and male fitness. First, to the extent that compensatory mutations are partially or completely recessive, their fixation probabilities will decline relative to the additive model (*e.g.*, Haldane 1932, p. 201). This should increase the waiting time to compensatory adaptation, and elevate the male load. Deviations from additivity may also bias the relative contributions of sex-linked and autosomal (diploid) genes to the resolution of mother's curse. When compensatory alleles are recessive and evolution is mutation-limited, the waiting time to compensatory substitution should be lower under X-linked compared to autosomal inheritance; the converse is true for dominant compensatory alleles (Charlesworth et al. 1987; Meisel and Connallon 2013). Such considerations may impact the chromosomal distribution of nuclear-encoded genes that are expressed in mitochondria (see Dean et al. 2015, and references therein).

Effects of heteroplasmy. For simplicity, we modeled selection and evolution of mitochondrial genes under the assumption of homoplasmy (*i.e.*, each individual inherits a single mitochondrial cytotype). In reality, each individual inherits many mitochondria from their mother, leaving some potential for heteroplasmy (see Bergstrom and Pritchard 1998; Roze et al. 2005). In the absence of selection on mtDNA mutations (*i.e.*, under neutrality in females), transient heteroplasmy should have no effect on the evolutionary trajectories of male-harming substitutions (*e.g.*, Bergstrom and Pritchard 1998; Roze et al. 2005). Likewise, when mtDNA alleles are sexually antagonistic, approximations based on homoplasmy are robust to heteroplasmy when mitochondrial genotypes additively affect fitness (as shown in Kuijper et al. 2015). Deviations from our results could occur in models that allow for

heteroplasmy and strong dominance interactions among mitochondrial genotypes within an individual (see Kuijper et al. 2015).

Further contributions to the male genetic load. Our models ignore three additional factors that are likely to further elevate the male genetic load. First, we have ignored fitness costs of mtDNA alleles that segregate in the population before they are ultimately eliminated by purifying selection in females, or genetic drift. As pointed out elsewhere (Frank and Hurst 1996; Smith and Connallon 2017; Appendix V of the Supplementary Material), transient polymorphisms can also contribute substantially to fitness variance among males, and deleterious mutation load in the mitochondrial genome. Second, we have ignored mtDNA substitutions with mildly deleterious effects on female fitness, which may fix under genetic drift and elevate the load of both sexes. Third, we have assumed that the genetic architecture of compensatory evolution is simple: that a single genetic substitution in the nuclear genome is sufficient to completely resolve mother's curse. If individual bouts of compensation require fixation of multiple alleles in the nuclear genome, then costs of mtDNA substitution may linger beyond our theoretical expectations.

CONCLUSION

Mitochondrial genetic effects on the organismal phenotype can be sexually asymmetric, with mutations that are primarily male-harming representing a component of the mutational landscape of the mitochondrial genome. Our study provides a theoretical framework for the potential role of mito-nuclear coevolution in sex-specific adaptation and disease. In particular, by demonstrating that the male mitochondrial load can be much larger than genetic load predictions from classical population genetics, we provide a theoretical rationale to expect persistent mother's curse effects in gonochoristic species – even in cases where the resolution of mother's curse is evolutionarily permissive. Finally, by showing that

the male mitochondrial load is likely to exhibit a threshold-dependent relationship with population size, our model predicts that the fitness consequences of mother's curse may vary widely among species with different demographic histories. These predictions provide a strong rationale for systematically testing mother's curse predictions in species exhibiting pronounced differences in their effective population sizes.

DATA ACCESSIBILITY

Details of the model, and R code for the simulations, are included in the Supplementary Material.

AUTHORS' CONTRIBUTIONS

Conceived the project: TC, MFC, EHM, DKD; developed and analyzed the models: TC; drafted and edited the manuscript: TC, MFC, EHM, DKD.

COMPETING INTERESTS

We have no competing interests.

FUNDING

This research was supported by funds from the Australian Research Council, the School of Biological Sciences at Monash University, a Marie Skłodowska-Curie Fellowship (to MFC), and a Royal Society University Research Fellowship (to EHM)

ACKNOWLEDGEMENTS

We thank two anonymous reviewers, whose suggestions led to substantial improvements in the paper.

REFERENCES

- Babcock CS, Asmussen MA. 1996. Effects of differential selection in the sexes on cytonuclear polymorphism and disequilibria. *Genetics* 144:839-853.
- Beekman, M., Dowling, D. K. & Aanen, D. K. 2014. The costs of being male: are there sex-specific effects of uniparental mitochondrial inheritance? *Phil Trans Roy Soc B* 369:20130440.
- Bergstrom CT, Pritchard J. 1998. Germline bottlenecks and the evolutionary maintenance of mitochondrial genomes. *Genetics* 149:2135-2146.
- Betancourt AJ, Kim Y, Orr HA. 2004. A pseudohitchhiking model of X vs. autosomal diversity. *Genetics* 168:2261-2269.
- Birky CW. 1995. Uniparental inheritance of mitochondrial and chloroplast genes: mechanisms and evolution. *Proc Natl Acad Sci USA* 92:11331-11338.
- Burton RS, Barreto FS. 2012. A disproportionate role for mtDNA in Dobzhansky-Muller incompatibilities? *Molecular Ecology* 21:4942-4957.
- Camus MF, Clancy DJ, Dowling DK. 2012. Mitochondria, maternal inheritance, and male aging. *Curr Biol.* 22:1717-1721.
- Camus MF, Wolf JBW, Morrow EH, Dowling DK. 2015. Single Nucleotides in the mtDNA Sequence Modify Mitochondrial Molecular Function and Are Associated with Sex-Specific Effects on Fertility and Aging. *Curr Biol.* 25:2717-2722.
- Charlesworth B, Charlesworth D. 2010. *Elements of evolutionary genetics*. Roberts and Company Publishers: Greenwood Village, Colorado.
- Charlesworth B, Coyne JA, Barton NH. 1987. The relative rates of evolution of sex chromosomes and autosomes. *Am Nat.* 130:113-146.

- Charlesworth D. 1981. A further study of the problems of the maintenance of females in gynodioecious species. *Heredity* 46:27-39
- Chase CD. 2007. Cytoplasmic male sterility: a window to the world of plant mitochondrial–nuclear interactions. *Trends Genet.* 23:81-90.
- Clancy DJ, Hime GR, Shirras AD. 2011. Cytoplasmic sterility in *Drosophila melanogaster* associated with a mitochondrial CYTB variant. *Heredity* 107:374-376.
- Connallon T, Clark AG. 2013. Antagonistic versus nonantagonistic models of balancing selection: characterizing the relative timescales and hitchhiking effects of partial selective sweeps. *Evolution* 67:908-917.
- Coyne JA, Orr HA. 2004. *Speciation*. Sinauer Associates: Sunderland MA.
- Crow JF. 1970. Genetic loads and the cost of natural selection. In Kojima K (ed.) *Mathematical topics in population genetics*, pp. 128-177. Springer-Verlag: Berlin
- Crow JF, Kimura M. 1970. *An introduction to population genetics theory*. Harper and Row: New York.
- Dean R, Zimmer F, Mank JE. 2015. Deficit of mitonuclear genes on the human X chromosome predates sex chromosome formation. *Genome Biol Evol.* 7:636-641.
- Dobler, R., Rogell, B., Budar, F. & Dowling, D. K. 2014. A meta-analysis of the strength and nature of cytoplasmic genetic effects. *Journal of Evolutionary Biology* 27: 2021-2034.
- Dowling DK, Meerupati T, Arnqvist G. 2010. Cytonuclear Interactions and the Economics of Mating in Seed Beetles. *American Naturalist* 176:131-140.
- Dowling DK. 2014. Evolutionary perspectives on the links between mitochondrial genotype and disease phenotype. *Biochim Biophys Acta* 1840:1393-403.
- Ellison CK, Burton RS. 2008. Interpopulation hybrid breakdown maps to the mitochondrial genome. *Evolution* 62:631-638.

- Engelstädter J, Charlat S. 2006. Outbreeding selects for spiteful cytoplasmic elements. *Proc Roy Soc B*. 273:923-929.
- Ewens WJ. 2004. *Mathematical population genetics. I. Theoretical introduction*. 2nd ed. Springer: New York.
- Eyre-Walker A. 2017. Mitochondrial replacement therapy: are mito-nuclear interactions likely to be a problem? *Genetics* 205:1365-1372.
- Frank SA. 1989. The evolutionary dynamics of cytoplasmic male sterility. *Am Nat*. 133:345-376.
- Frank SA, Hurst LD. 1996. Mitochondria and male disease. *Nature* 383:224.
- Froman DP, Kirby JD. 2005. Sperm mobility: phenotype in roosters (*Gallus domesticus*) determined by mitochondrial function. *Biology of Reproduction* 72:562-567.
- Gemmell NJ, Metcalf VJ, Allendorf FW. 2004. Mother's curse: the effect of mtDNA on individual fitness and population viability. *Trends Ecol Evol*. 19:238-244.
- Gillespie JH. 2004. Why $k = 4N_{us}$ is silly. In *The Evolution of Population Biology*, Singh RS, Uyenoyama MK (eds.). Cambridge University Press: Cambridge, UK, pp. 178-192.
- Gregorius HR, Ross MD. 1984. Selection with gene-cytoplasm interactions. I. Maintenance of cytoplasm polymorphisms. *Genetics* 107:165-178.
- Haldane JBS. 1931. A mathematical theory of natural selection. VIII. Stable metapopulations. *Proc Camb Philos Soc*. 27:137-142.
- Haldane JBS. 1957. The cost of selection. *J Genet*. 55:511-524.
- Hedrick P. 2012. Reversing mother's curse revisited. *Evolution* 66:612-6116.
- Immonen E, Collet M, Goenaga J, Arnqvist G. 2016. Direct and indirect genetic effects of sex-specific mitonuclear epistasis on reproductive ageing. *Heredity* 116:338-347.
- Innocenti P, Morrow EH, Dowling DK. 2011. Experimental Evidence Supports a Sex-Specific Selective Sieve in Mitochondrial Genome Evolution. *Science* 332:845-848.

- Jacobs MS, Wade MJ. 2003. A synthetic review of the theory of gynodioecy. *Am Nat.* 161:837-851.
- James AC, Ballard JWO. 2003. Mitochondrial genotype affects fitness in *Drosophila simulans*. *Genetics* 164:187-194.
- Kimura M. 1979. Model of effectively neutral mutations in which selective constraint is incorporated. *Proc Nat Acad Sci USA* 76:3440-3444.
- Kimura M. 1985. The role of compensatory neutral mutations in molecular evolution. *J Genet.* 64:7-19.
- Kimura M, Ohta T. 1971. *Theoretical aspects of population genetics*. Princeton University Press: Princeton, NJ.
- Kuijper B, Lane N, Pomiankowski A. 2015. Can paternal leakage maintain sexually antagonistic polymorphism in the cytoplasm. *J Evol Biol.* 28:468-480.
- Lande R. 1994. Risk of population extinction from fixation of new deleterious mutations. *Evolution* 48:1460-1469.
- Lande R. 1998. Risk of population extinction from fixation of deleterious and reverse mutations. *Genetica* 102:21-27.
- Martikainen MK, Grady JP, Ng YS, Alston CL, Gorman GS, Taylor RW, McFarland R, Turnbull DM. 2017. Decreased male reproductive success in association with mitochondrial dysfunction. *European J Hum Genet.* 25:1162-1164.
- Maynard Smith J. 1971. What use is sex? *J Theor Biol.* 30:319-335.
- Maynard Smith J. 1976. What determines the rate of evolution? *Am Nat.* 110:331-338.
- McCandlish DM, Stoltzfus A. 2014. Modeling evolution using the probability of fixation: history and implications. *Quarterly Rev Biol.* 89:225-252.

- Meiklejohn CD, Holmbeck MA, Siddiq MA, Abt DN, Rand DM, Montooth KL. 2013. An Incompatibility between a Mitochondrial tRNA and Its Nuclear-Encoded tRNA Synthetase Compromises Development and Fitness in *Drosophila*. *Plos Genetics* 9.
- Meisel RP, Connallon T. 2013. The faster-X effect: integrating theory and data. *Trends Genet.* 29:537-544.
- Milot E, Moreau C, Gagnon A, Cohen AA, Brais B, Labuda D. 2017. Mother's curse neutralizes natural selection against a human genetic disease over three centuries. *Nature Ecology and Evolution* 1:1400-1406.
- Mossman JA, Tross JG, Li N, Wu Z, Rand DM. 2016a. Mitochondrial-nuclear interactions mediate sex-specific transcriptional profiles in *Drosophila*. *Genetics* 204:613-630.
- Mossman JA, Biancani LM, Zhu CT, Rand DM. 2016b. Mitonuclear epistasis for development time and its modification by diet in *Drosophila*. *Genetics* 203:463-484.
- Mossman JA, Tross JG, Jourjine NA, Li N, Wu Z, Rand DM. 2017. Mitonuclear interactions mediate transcriptional responses to hypoxia in *Drosophila*. *Mol Biol Evol.* 34:447-466.
- Nakada K, Sato A, Yoshida K, Morita T, Tanaka H, Inoue SI, Yonekawa H, Hayashi JI. 2006. Mitochondria-related male infertility. *Proceedings of the National Academy of Sciences of the United States of America* 103:15148-15153.
- Orr HA, Unckless RL. 2014. The population genetics of evolutionary rescue. *PLoS Genet.* 10:e1004551.
- Otto SP. 2004. Two steps forward, one step back: the pleiotropic effects of favoured alleles. *Proc Roy Soc Lond, B* 271:705-714.
- Otto SP, Day T. 2007. *A biologist's guide to mathematical modeling in ecology and evolution*. Princeton University Press: Princeton, NJ.

- Patel MR, Miriyala GK, Littleton AJ, Yang H, Trinh K, Young JM, Kennedy SR, Yamashita YM, Pallanck LJ, Malik HS. 2016. A mitochondrial DNA hypomorph of cytochrome oxidase specifically impairs male fertility in *Drosophila melanogaster*. *eLife* 5:e16923.
- R Core Team. 2016. R: A language and environment for statistical computing. R Foundation for Statistical Computing, Vienna, Austria.
- Rand, D. M., Clark, A. G. & Kann, L. M. 2001. Sexually antagonistic cytonuclear fitness interactions in *Drosophila melanogaster*. *Genetics* **159**: 173-187.
- Rand, D. M., Haney, R. A. & Fry, A. J. 2004. Cytonuclear coevolution: the genomics of cooperation. *Trends Ecol Evol.* 19:645-653.
- Reinhardt K, Dowling DK, Morrow EH. 2013. Medicine. Mitochondrial replacement, evolution, and the clinic. *Science* 341:1345-1346.
- Roubertoux PL, Sluyter F, Carlier M, Marcet B, Maarouf-Veray F, Chérif C, Marican C, Arrechi P, Godin F, Jamon M, Verrier B, Cohen-Salmon C. 2003. Mitochondrial DNA modifies cognition in interaction with the nuclear genome and age in mice. *Nature Genetics* 35:65-69.
- Roze D, Rousset F, Michalakis Y. 2005. Germline bottlenecks, biparental inheritance and selection on mitochondrial variants: a two-level selection model. *Genetics* 170:1385-1399.
- Schnable PS, Wise RP. 1998. The molecular basis of cytoplasmic male sterility and fertility restoration. *Trends in Plant Science* 3:175-180
- Smith S, Turbill C, Suchentrunk F. 2010. Introducing mother's curse: low male fertility associated with an imported mtDNA haplotype in a captive colony of brown hares. *Molecular Ecology* 19:36-43.
- Smith SRT, Connallon T. 2017. The contribution of the mitochondrial genome to sex-specific fitness variance. *Evolution* 71:1417-1424.

- Stephan W. 1996. The rate of compensatory evolution. *Genetics* 144:419-426.
- Takahasi KR. 2009. Coalescent under the evolution of coadaptation. *Molecular Ecology* 18:5018-5029.
- Unckless RL, Herren JK. 2009. Population genetics of sexually antagonistic mitochondrial mutants under inbreeding. *J Theor Biol.* 260:132-136.
- Wade MJ. 2014. Paradox of mother's curse and the maternally provisioned offspring microbiome. *Cold Spring Harbor Perspectives in Biology* 6:a017541
- Wade MJ, Brandvain Y. 2009. Reversing mother's curse: selection on male mitochondrial fitness effects. *Evolution* 63:1084-1089.
- Wallace D, Singh G, Lott M, Hodge J, Schurr T, Lezza A, Elsas L, Nikoskelainen E. 1988. Mitochondrial DNA mutation associated with Leber's hereditary optic neuropathy. *Science* 242: 1427-1430.
- Wolff JN, Ladoukakis ED, Enríquez JA, Dowling DK. 2014. Mitonuclear interactions: evolutionary consequences over multiple biological scales. *Phil Trans Roy Soc B* 369:20130443.
- Wolff JN, Tompkins DM, Gemmell NJ, Dowling DK. 2016. Mitonuclear interactions, mtDNA-mediated thermal plasticity, and implications for the Trojan Female Technique for pest control. *Scientific Reports* 6:30016.
- Yee WKW, Rogell B, Lemos B, Dowling DK. 2015. Intergenomic interactions between mitochondrial and Y-linked genes shape male mating patterns and fertility in *Drosophila melanogaster*. *Evolution* 69:2876-2890.
- Yee WKW, Sutton KL, Dowling DK. 2013. In vivo male fertility is affected by naturally occurring mitochondrial haplotypes. *Current Biology* 23:R55-R56.
- Zhang H, Guillaume F, Engelstädter J. 2012. The dynamics of mitochondrial mutations causing male infertility in spatially structured populations. *Evolution* 66:3179-3188.

TABLES AND FIGURES

Table 1. Relative fitness as a function of the mito-nuclear genotype.

	Nuclear Genotype		
	<i>AA</i>	<i>Aa</i>	<i>aa</i>
Male relative fitness			
<i>M</i> cyotype	1	$1 - t_m/2$	$1 - t_m$
<i>m</i> cyotype	$1 - s_m$	$1 - s_m/2$	1
Female relative fitness			
<i>M</i> cyotype	1	1	1
<i>m</i> cyotype	$1 + s_f$	$1 + s_f$	$1 + s_f$

FIGURES & LEGENDS

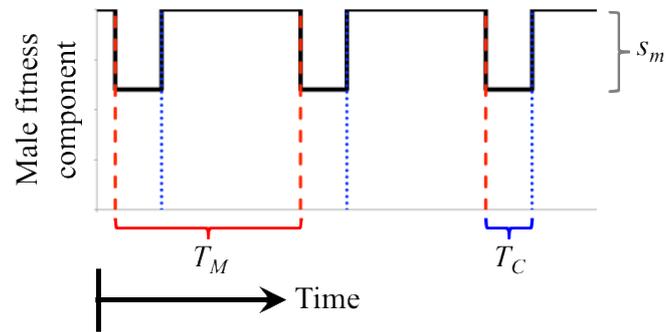


Figure 1. Fluctuations in male fitness components resulting from coevolution between a mitochondrial and nuclear locus. The figure illustrates the case of sequential substitutions of male-harming mtDNA alleles, each followed by the substitution of a compensatory allele that restores male fitness. The mean time between substitutions at a mitochondrial locus is T_M . The mean lag between a mitochondrial substitution and a nuclear compensatory substitution is T_C . The time that males spend within the low-fitness state depends on the tempo of mitochondrial substitutions relative to the lag time of compensatory substitutions.

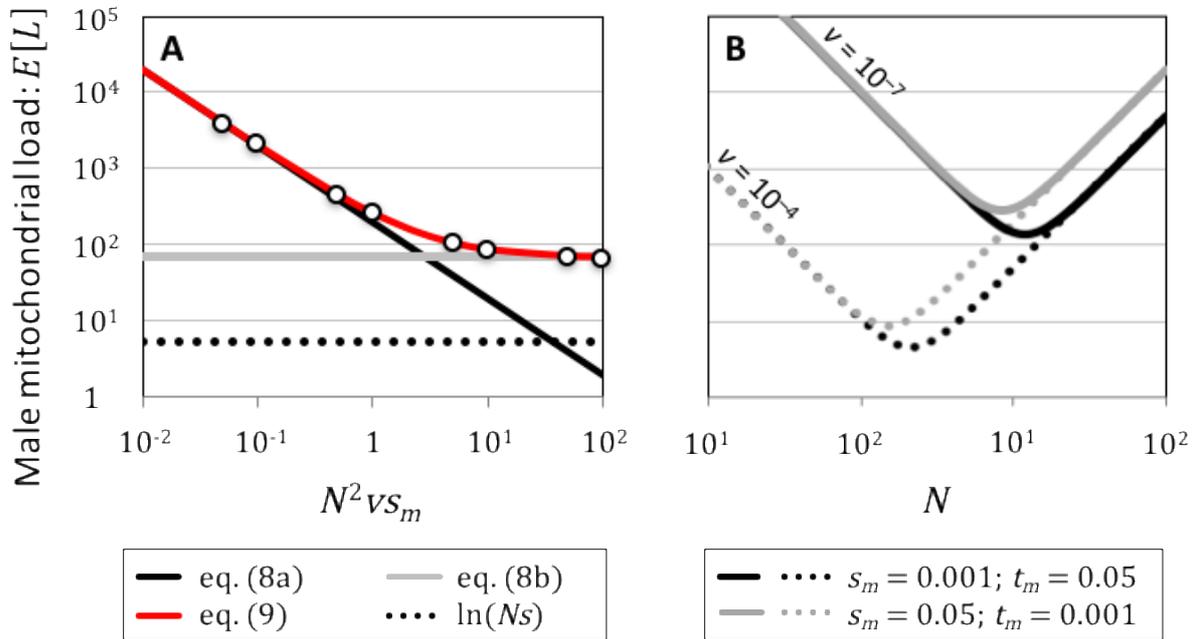


Figure 2. The male mitochondrial load under male-harming/female-neutral mtDNA substitutions. **(A)** Results are shown for $N = 10,000$ and $s_m = t_m = 0.02$, with a variable mutation rate at the compensatory locus (v). Open circles show the average load from 1,000 Wright-Fisher forward simulations of pairs of mito-nuclear substitutions. Mitochondrial load results are contrasted against the classical substitution load (Haldane’s (1957) “cost of selection”, represented by the broken line, $C = \ln(Ns)$), which shows the load arising from the substitution of an unconditionally beneficial mtDNA mutation with fitness benefit of $s = 0.02$, and “effective” initial frequency of $1/Ns$ (*i.e.*, following Maynard Smith 1971, 1976; see the Supplementary Material). **(B)** Male mitochondrial load as a function of population size (N). Theoretical curves, which are based on eq. (9), illustrate the impacts of the compensatory mutation rate (v) and selection parameters (s_m, t_m) on the magnitude of the load. The minimum for each curve corresponds to N_{crit} , the population size that minimizes the male load (see eq. (10)).

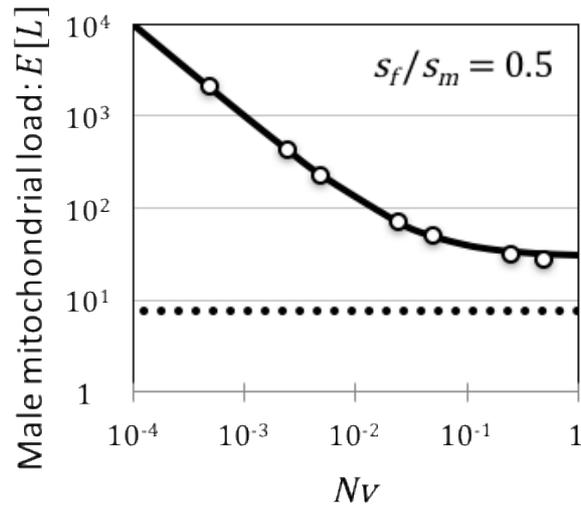


Figure 3. The male mitochondrial load under sexually antagonistic mtDNA substitutions. Results are shown for $N = 100,000$, $s_m = t_m = 0.02$, and variable compensatory mutation rate, ν . The solid line is the approximation from eq. (11). Open circles show the average load from 1,000 Wright-Fisher forward simulations of pairs of mito-nuclear substitutions. Mitochondrial load results are contrasted against the classical substitution load (Haldane’s (1957) “cost of selection”, represented by the broken line, $C = \ln(Ns)$), which shows the load arising from the substitution of an unconditionally beneficial mtDNA mutation with fitness benefit of $s = 0.02$, and “effective” initial frequency of $1/Ns$ (*i.e.*, following Maynard Smith 1971, 1976; see the Supplementary Material).