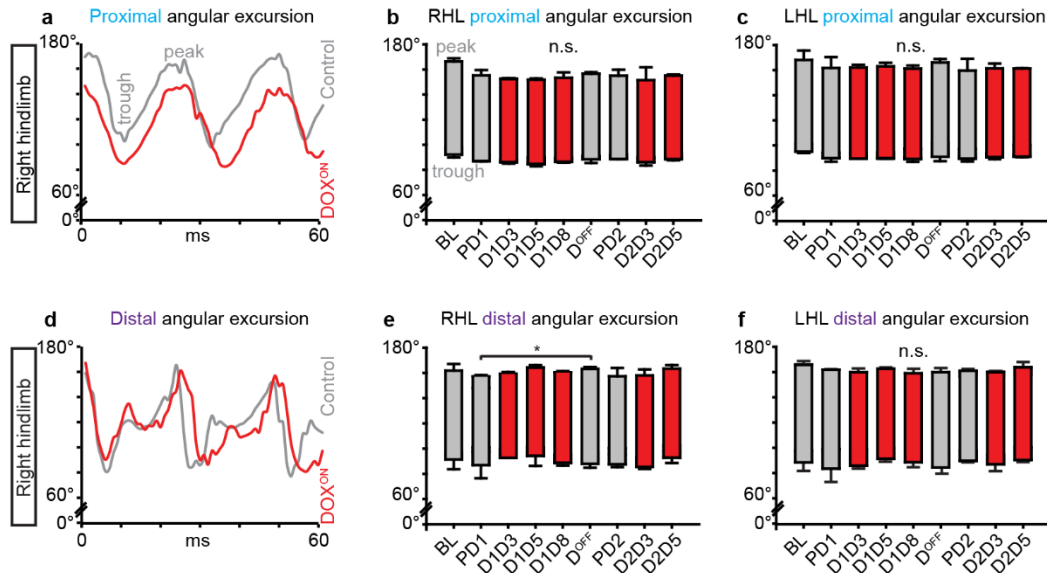


1 **Supplementary Figure 1.**



2

3 **Supplementary Figure 1. Silencing L2-L5 interneurons does not affect hindlimb range-of-**

4 **motion.** Representative traces of the proximal (a) and distal (d) hindlimb angle excursions (from peak to

5 trough) at Control (gray) and DOX^{ON} time points (red), respectively (right hindlimb shown). Silencing

6 L2-L5 interneurons did not affect the peak-to-trough excursion of the right (b) or left (c)

7 proximal hindlimb angle. Similarly, the conditional silencing did not alter the excursion of the

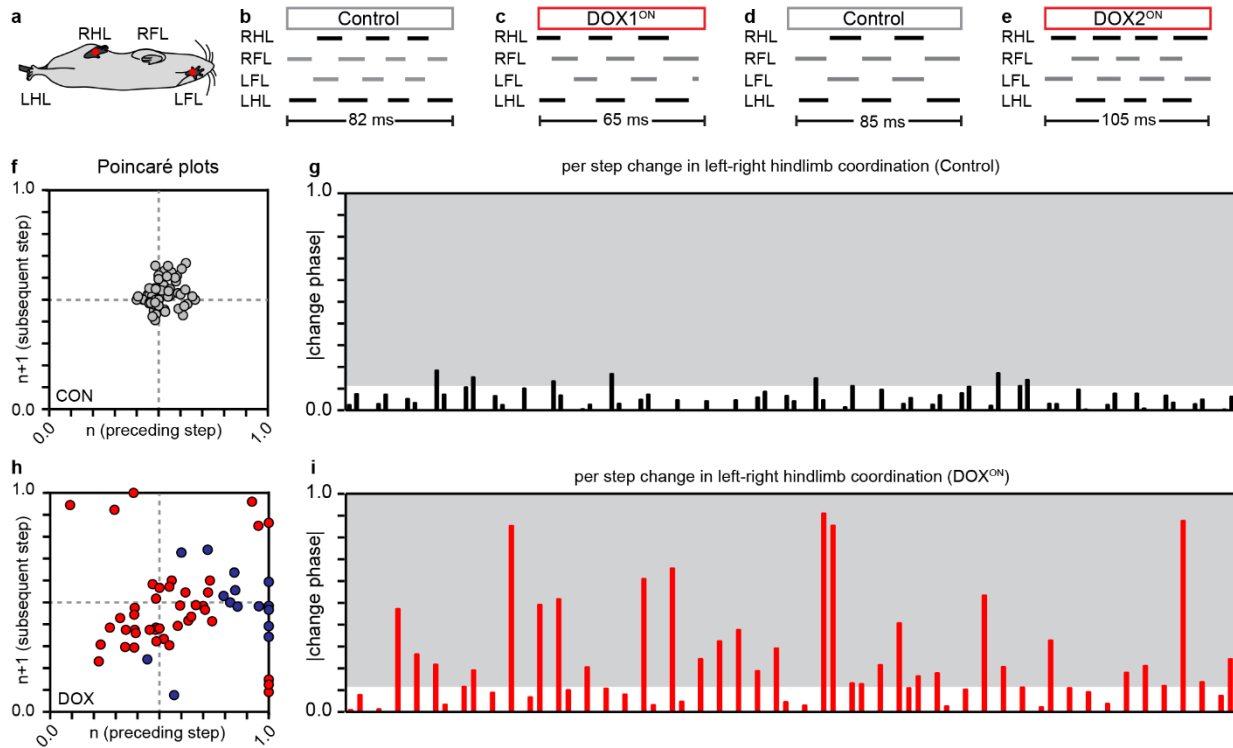
8 distal hindlimb angle for the right (e) or left hindlimb (f), respectively. There was a slight, but

9 significant increase in the angular excursion at DOX^{OFF} as compared to Pre-DOX1 (*p<0.05;

10 mixed model ANOVA and Bonferroni *post hoc* t-test). Quantitative data are the mean ± S.D.

11 (N=6).

12 **Supplementary Figure 2.**

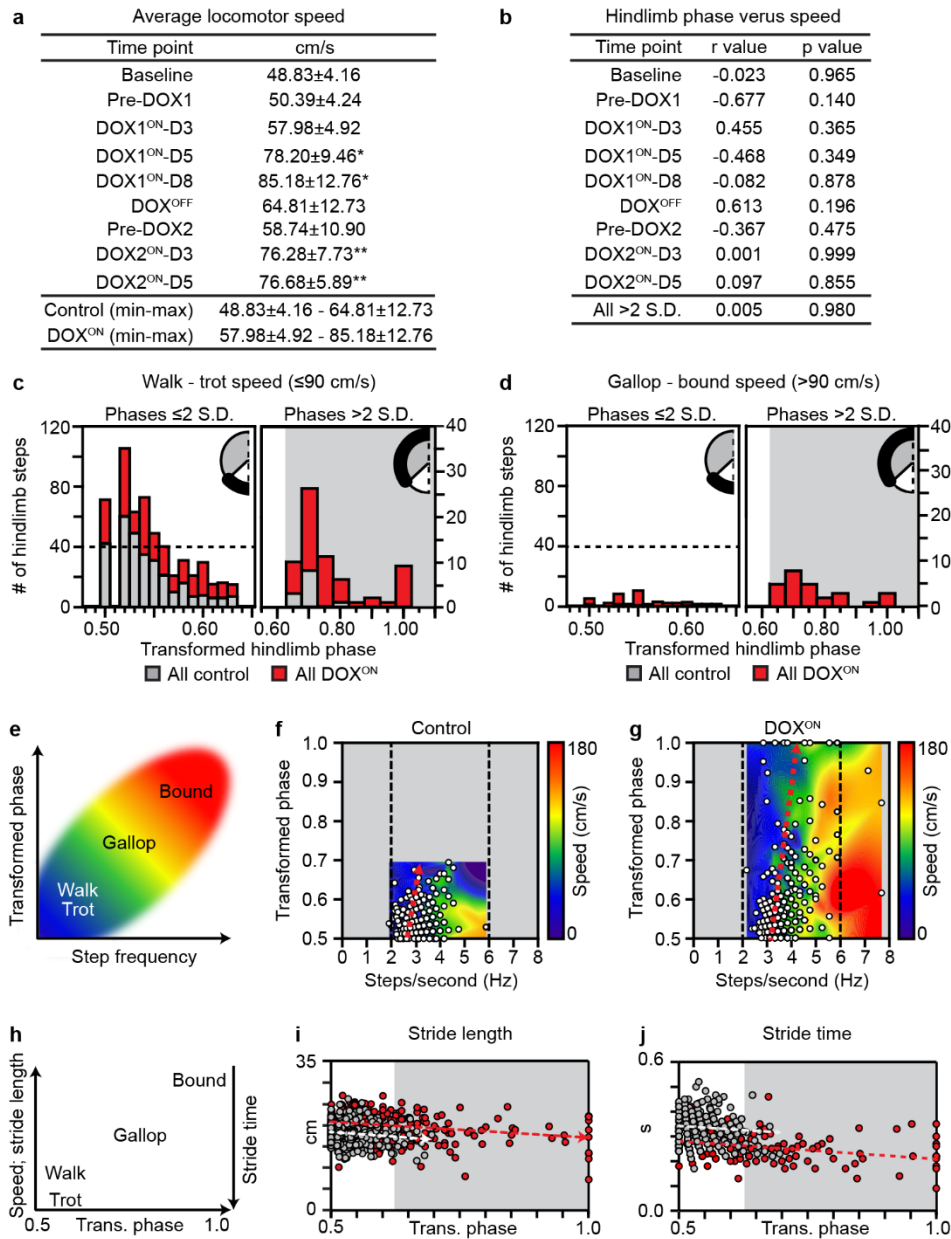


13

14 **Supplementary Figure 2. Silencing L2-L5 interneurons disrupts left-right hindlimb coordination on**

15 **a step-by-step basis.** (a) Ventral recordings of paw contacts were used to generate swing-stance graphs
 16 and gait analyses (LFL: left forelimb; RFL: right forelimb; LHL: left hindlimb; RHL: right hindlimb). (b-
 17 e) Swing-stance graphs from Control and DOX^{ON} time points, respectively, with the duration of the
 18 overall locomotor bout shown below. Each line denotes paw contact with the ground (stance phase) while
 19 the intervening space between each line represents the swing phase, when the paw is in the air. A subset
 20 of the data shown in Figure 5f was selected for expanded graphical representation of per-step changes in
 21 left-right hindlimb coordination (g,i, expanded for clarity; shaded region denotes changes beyond control
 22 variability) as well as Poincaré plot generation (f,h), which illustrate left-right hindlimb coordination
 23 across successive step cycles (x-axis, preceding step n ; y-axis, subsequent step $n+1$). $N=95$ and 96 per-
 24 step changes were sampled from Control and DOX^{ON} time points, respectively.

25 **Supplementary Figure 3.**



26 **Supplementary Figure 3. Silencing-induced changes in hindlimb coordination did not**

27 **correlate with speed or gait-related indices. (a)** Overall locomotor speed across time points (group

28 mean±S.D.). The speed was significantly enhanced during DOX1^{ON}-D5 and DOX1^{ON}-D8 as compared to

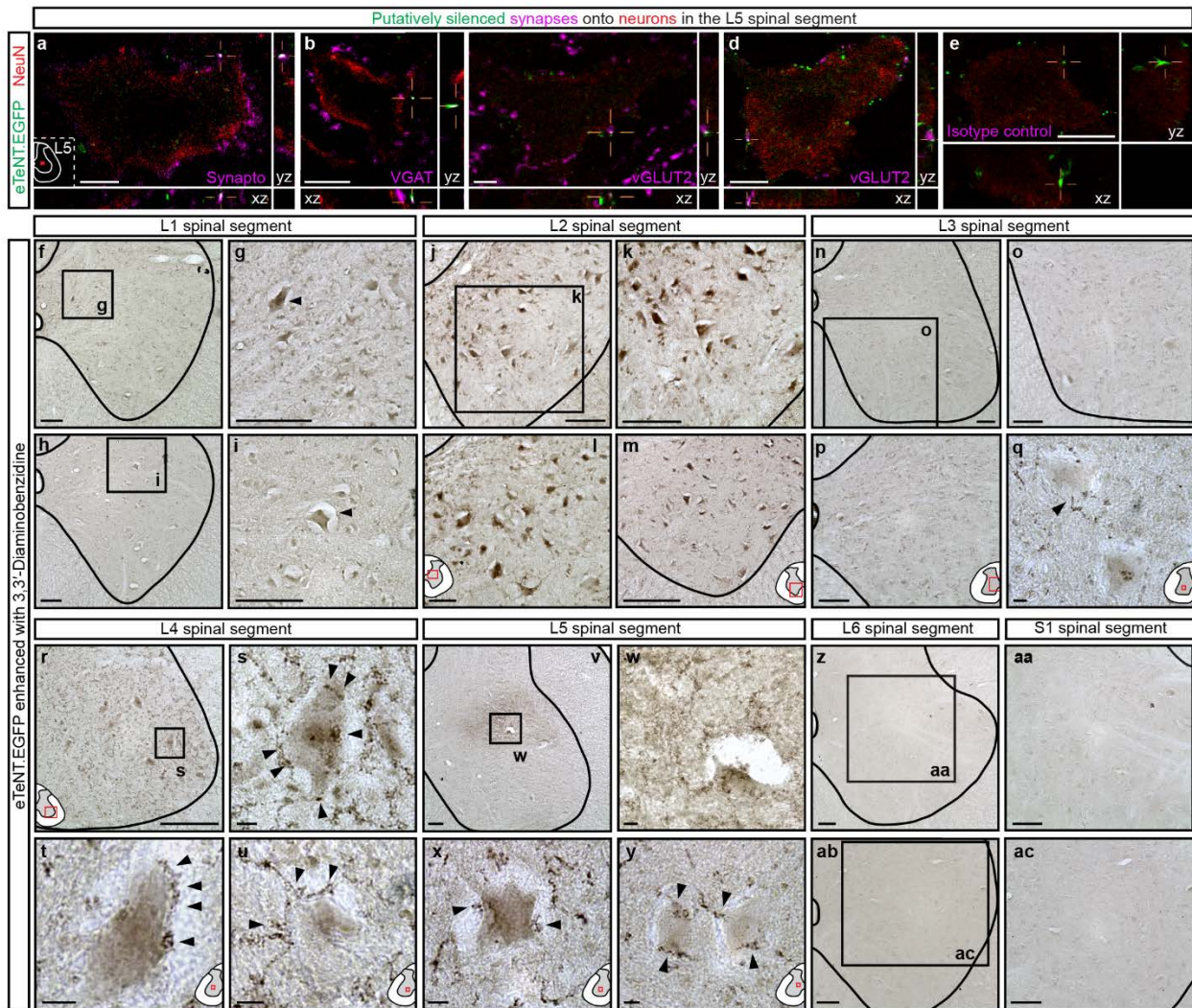
29 Pre-DOX1 (* $p \leq 0.5$, repeated measures ANOVA; Tukey's *post hoc* honest significant difference t-test) as

30 well as during DOX2^{ON}-D3 and DOX2^{ON}-D5 as compared to Pre-DOX2 (** $p \leq 0.01$). No significant

31 different differences were detected when comparing to Baseline and DOX^{OFF}. The overall speed range

32 (minimum-maximum) for the group at Control and DOX^{ON} time points, respectively, is listed below. **(b)**
33 Summary of Pearson correlations between averaged phase vs speed, per time point. **(c)** Left
34 panel shows the frequency distribution of hindlimb coordination values within the control
35 variability (right inset; 0.50-0.63) that occurred at ≤ 90 centimeters/second, a locomotor velocity
36 where the limbs typically alternate in a walk or trot gait. Right panel shows the frequency at
37 which phases > 0.63 , including synchrony at 1.0, occur at a speed where alternation usually
38 prevails (shaded region denotes phases beyond control variability). **(d)** Frequency distribution of
39 hindlimb phases at gallop-to-bound speeds (> 90 cm/sec). **(e,h)** Inter-relationship between various
40 gait indices. **(f,g)** Step frequency-phase relationship (white circles) mapped onto speed contour
41 plot (see methods for detail). Silencing-induced changes in hindlimb coordination did not
42 correlate with increased step frequency. **(i)** Similarly, changes to hindlimb coordination did not
43 correlate with increased stride length (Control, $r_S = -0.068$ in dashed white line; DOX^{ON}, $r_S = -$
44 0.125 in dashed red line; Spearman Rank correlation) or decreased stride time **(j)**, Control $r_S = -$
45 0.036 ; DOX^{ON} $r_S = -0.338$).

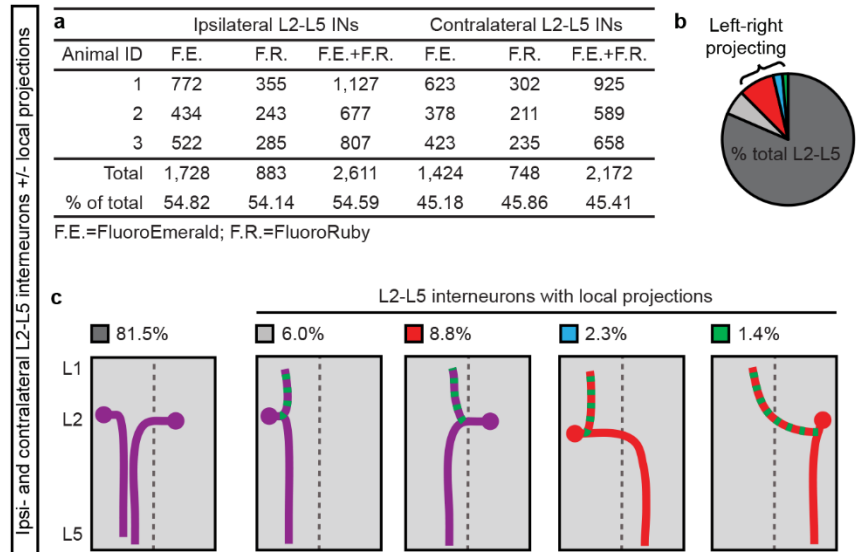
46 **Supplementary Figure 4.**



47
 48 **Supplementary Figure 4. Immunohistochemical interrogation of conditionally silenced L2-L5**
 49 **interneurons.** Cross-sections throughout the z-stack confirming co-localization of eTeNT.EGFP with
 50 synaptophysin (**a**, synapto), vesicular GABA transporter (**b**, VGAT), or vesicular glutamate transporter 2
 51 (**c-d**) in the xz-yz planes (white signal denoted in cross-hairs). (**e**) Isotype control. (**f-ac**) eTeNT.EGFP
 52 immunoreactivity assessed throughout the lumbosacral enlargement using 3,3'-Diaminobenzidine
 53 enhancement. (**f-i**) Few double-infected neurons were detected at L1 (black arrowheads). (**j-m**) Numerous
 54 double-infected neurons were detected throughout the intermediate gray matter at spinal L2 (black
 55 arrowheads). (**n-q**) Sparse, terminal-like structures were detected at L3 (**q**, arrowhead). No double-
 56 infected neurons were detected. (**r-u**) Terminal-like structures were detected throughout intermediate and

57 ventral gray matter at spinal L4 through L5 (arrowheads). (**v-y**) Cross-sections distal to lentiviral vector
58 injection. (**z-ac**) No eTeNT.EGFP immunoreactivity was detected at L6-S1. Images acquired at 10, 20,
59 and 40x magnifications. Scale bar=10 μm (**a-e, q, s-u, w-x, y**) or 100 μm (**f-p, r, v, z-aa, ab-ac**).

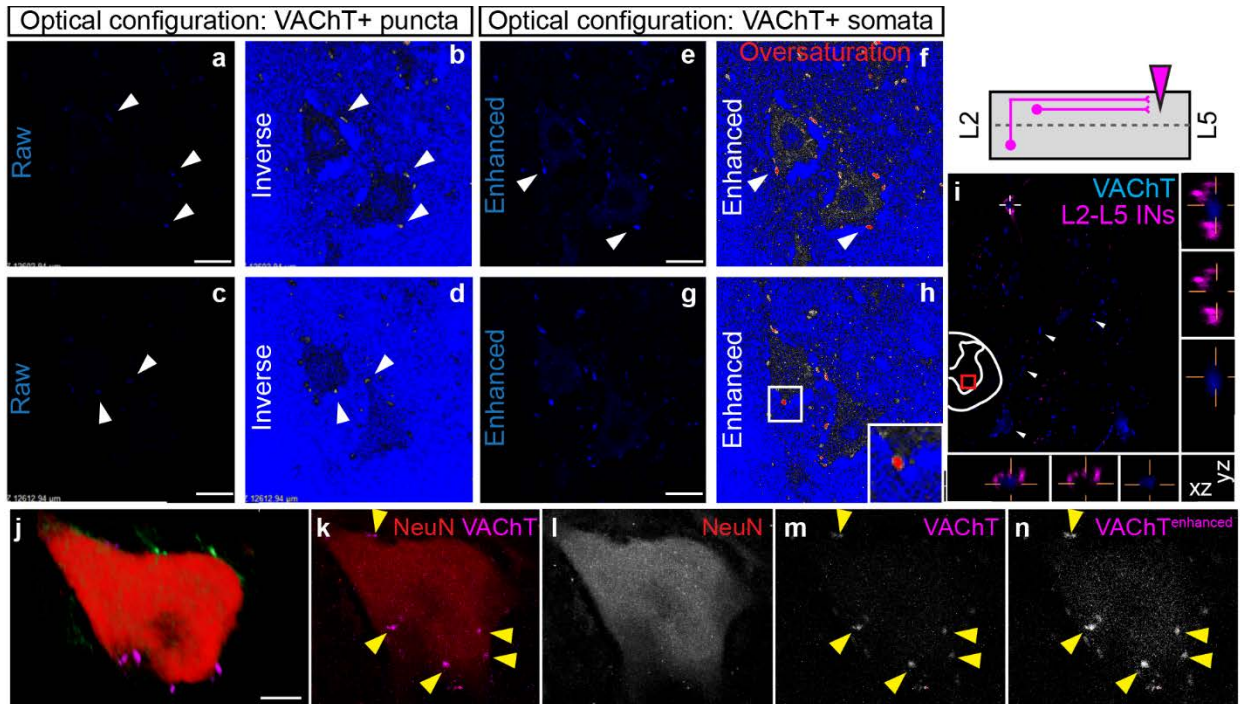
60 **Supplementary Figure 5.**



61 **Supplementary Figure 5. The majority of L2-L5 interneurons lack local projections in the**
 62 **rostral lumbar spinal cord. (a)** Data shown are absolute cell counts and percent total of L2
 63 interneurons with ipsilateral or contralateral projections to spinal L5 following bilateral
 64 injections of FluoroEmerald (F.E.) and FluoroRuby (F.R.). No significant difference was found
 65 between ipsi- and contralateral L2-L5 interneurons (total ipsilateral vs total contralateral: $p > 0.4$;
 66 independent t-test between means of equal variance). **(b,c)** Summary of the L2-L5 projection
 67 patterns observed following triple tracer (CTB) injections. Of the total L2-L5 interneurons
 68 labeled at L2 following bilateral L5 injections, approximately 80% did not have local projections
 69 within one (rostral) segment of their cell body (dark gray, $81.49 \pm 2.36\%$). Almost 20% of the L2-
 70 L5 interneurons had projections within one segment of their cell body ($18.51 \pm 2.36\%$). Of this
 71 proportion, approximately 12.5% had direct projections between the left and right sides of the
 72 spinal cord **(c, red, blue, and green)**. Data shown represent the proportions of projection patterns
 73 observed relative to percent total L2-L5 interneurons that were labeled.

74

75 **Supplementary Figure 6.**



76

77 **Supplementary Figure 6. Putatively silenced synapses onto caudal lumbar motor neurons.**

78 (a-d) Optimal optical configurations for VACHT immunoreactivity in pre-synaptic puncta vs
 79 motor neuron somata (not shown: CTB-labeled motor neurons from hindlimb intramuscular
 80 injections; see “ghost” neurons in b and d). Images shown in (a-b) and (c-d) reflect VACHT-
 81 positive immunoreactivity with optimal confocal configurations set for the pre-synaptic puncta.
 82 Left panels reflect raw VACHT signal (blue). Right panels reflect the inverted image to show
 83 saturation of the fluorophore based on image acquisition settings. Ideal optical configurations for
 84 fluorophore detection yield gray-white signal (white arrowheads). Images shown in (e-f) and (g-
 85 h) reflect VACHT-positive immunoreactivity with optical configurations set for the somata. Note
 86 that when VACHT signal is detected in the somata, the pre-synaptic puncta become oversaturated
 87 (red signal, white arrowheads). Images shown reflect one optical slice (0.4 μ m) through a 20-30
 88 slice z-stack captured at 100x (scale bar=20 μ m). A subset of L2-L5 interneurons showed

89 somatic co-localization with VAcHT (i). (j-n) *Post hoc* image modifications reveal VAcHT-
90 positive immunoreactivity in putative NeuN-positive motor neuron shown in Figure 7e.

91

92

93 **Supplementary Table 1.**

	Stance time		Swing time		Stride time		Stride length	
	r value	p value	r value	p value	r value	p value	r value	p value
Baseline	-0.023	0.965	0.574	0.234	0.508	0.304	0.648	0.164
Pre-DOX1	-0.677	0.140	0.935	0.054	0.732	0.098	-0.505	0.307
DOX1 ^{ON} -D3	0.455	0.365	-0.640	0.171	-0.699	0.123	-0.169	0.750
DOX1 ^{ON} -D5	-0.468	0.349	-0.860	0.252	-0.354	0.492	-0.865	0.234
DOX1 ^{ON} -D8	-0.082	0.878	-0.422	0.404	-0.129	0.808	-0.306	0.555
DOX ^{OFF}	0.613	0.196	-0.326	0.529	-0.668	0.147	0.389	0.446
Pre-DOX2	-0.367	0.475	0.273	0.601	0.630	0.180	0.010	0.985
DOX2 ^{ON} -D3	0.001	0.999	-0.528	0.282	-0.768	0.074	-0.720	0.106
DOX2 ^{ON} -D5	0.097	0.855	-0.738	0.094	-0.727	0.102	-0.694	0.126
All phases >2 S.D.	-0.031	0.862	-0.247	0.158	-0.091	0.609	-0.100	0.573

94 **Supplementary Table 1. Disruption in hindlimb phase did not correlate with speed-related**

95 **gait parameters.** Time point comparisons showed hindlimb phase did not significantly correlate

96 with speed-associated gait measures such as stance, swing, and stride time as well as stride

97 distance. All phase values >2 S.D. also did not significantly correlate with speed-related gait

98 measures (averaged data; Pearson correlation with the Bonferroni correction for multiple

99 comparisons to reduce the likelihood of Type I errors).

100 **Supplementary Table 2.**

	Stance time		Swing time		Stride time		Stride length	
	$r_{y(2\cdot1)}$ value	p value	$r_{y(2\cdot1)}$ value	p value	$r_{y(2\cdot1)}$ value	p value	$r_{y(2\cdot1)}$ value	p value
Baseline	0.663	0.223	0.766	0.131	0.862	0.060	0.842	0.073
Pre-DOX1	-0.414	0.323	0.645	0.051	0.306	0.487	0.246	0.582
DOX1 ^{ON} -D3	0.025	0.965	-0.544	0.274	-0.801	0.342	-0.753	0.071
DOX1 ^{ON} -D5	-0.700	0.110	-0.809	0.261	-0.858	0.054	-0.867	0.027
DOX1 ^{ON} -D8	-0.961	0.072	-0.463	0.430	-0.669	0.215	-0.809	0.095
DOX ^{OFF}	-0.327	0.488	0.014	0.978	-0.284	0.553	-0.267	0.579
Pre-DOX2	0.633	0.206	0.045	0.939	0.599	0.241	0.534	0.312
DOX2 ^{ON} -D3	-0.966	0.072	-0.557	0.329	-0.852	0.067	-0.883	0.423
DOX2 ^{ON} -D5	-0.915	0.243	-0.744	0.146	-0.840	0.072	-0.930	0.180
All phases >2 S.D.	-0.039	0.827	-0.258	0.148	-0.140	0.436	-0.162	0.368

101 **Supplementary Table 2. Hindlimb phase versus gait after controlling for speed.** Part
102 correlations were performed, where the relationship between phase and gait (e.g., stance time)
103 was measured after controlling for the effect of speed on that gait variable. Hindlimb phase
104 significantly correlated with stride distance at DOX1^{ON}-D5 only. This represents approximately
105 2.8% of the total phase versus gait comparisons analyzed. All hindlimb phase values >2 S.D. did
106 not significantly correlate with gait (averaged data; Part correlation with Bonferroni correction
107 for multiple comparisons).

108 **Supplementary Table 3.**

		p value	U² value
Baseline vs	Pre-DOX1	p>0.50	-0.13369
	DOX1 ^{ON} -D3	0.05<p<0.10	0.16895
	DOX1 ^{ON} -D5	0.002<p<0.005	0.33339
	DOX1 ^{ON} -D8	0.10<p<0.20	0.13157
	All-DOX1 ^{ON}	p<0.001	0.56176
	DOX ^{OFF}	p>0.5	0.07047
Pre-DOX1 vs	DOX1 ^{ON} -D3	0.20<p<0.50	0.08912
	DOX1 ^{ON} -D5	0.01<p<0.02	0.24507
	DOX1 ^{ON} -D8	0.02<p<0.05	0.21514
	All-DOX1 ^{ON}	p<0.001	0.59762
	DOX ^{OFF}	p>0.50	0.00498
	Pre-DOX2	p>0.50	0.01499
Baseline + Pre-DOX1 vs	All-DOX1 ^{ON}	p<0.001	0.59762
Pre-DOX2 vs	DOX ^{OFF}	p>0.50	0.07400
	DOX2 ^{ON} -D3	0.002<p<0.005	0.31224
	DOX2 ^{ON} -D5	p<0.001	0.39267
	All-DOX2 ^{ON}	p<0.001	1.29965

109
110 **Supplementary Table 3. Silencing L2-L5 interneurons functionally uncouples the left and**
111 **right hindlimbs during overground stepping.** Using the non-parametric two-sample U² test,
112 we tested the null hypothesis that the two samples (e.g. Baseline vs DOX1^{ON}-D5) came from two
113 populations with the same directions (in other words, degree of concentration or dispersion).
114 This is an indication of whether the limbs are coupled (phases concentrated in same direction) or
115 uncoupled (phases are dispersed). Silencing the L2-L5 interneurons significantly decreased the
116 concentration of the phase values (reduced clustering at 0.5) and caused an increased dispersion
117 throughout the coordination range. This suggests the hindlimbs became functionally uncoupled
118 during overground locomotion. (Critical value of Watson’s U²_(0.05,∞,∞) = 0.1869; Appendix D,
119 Table D.44)²⁰.

120 **Supplementary Table 4.**

			p value	U² value
Forelimb stepping	Baseline vs	Pre-DOX1	p>0.50	0.01735
		DOX1 ^{ON} -D3	p>0.50	0.03821
		DOX1 ^{ON} -D5	p>0.50	-0.03716
		DOX1 ^{ON} -D8	p>0.50	0.06999
		All-DOX1 ^{ON}	p>0.50	0.00771
		DOX ^{OFF}	p>0.50	0.02585
	Pre-DOX1 vs	DOX1 ^{ON} -D3	p>0.50	0.02560
		DOX1 ^{ON} -D5	p>0.50	0.03907
		DOX1 ^{ON} -D8	p>0.50	0.02254
		All-DOX1 ^{ON}	p>0.50	0.07072
DOX ^{OFF}		p>0.50	0.03725	
Hindlimb swimming	Baseline vs	Pre-DOX1	p>0.50	0.02585
		DOX1 ^{ON} -D3	p>0.50	-0.96664
		DOX1 ^{ON} -D5	p>0.50	-0.54443
		DOX1 ^{ON} -D8	p>0.50	-0.48521
		All-DOX1 ^{ON}	p>0.50	-1.62815
		DOX ^{OFF}	0.02<p<0.05	0.20163
	Pre-DOX1 vs	DOX1 ^{ON} -D3	p>0.50	-1.3616
		DOX1 ^{ON} -D5	p>0.50	-0.79496
		DOX1 ^{ON} -D8	p>0.50	-0.92183
		All-DOX1 ^{ON}	p>0.50	-2.69285
DOX ^{OFF}		p>0.50	0.07581	

121 **Supplementary Table 4. The conditional silencing of L2-L5 interneurons does not uncouple**

122 **the forelimbs during stepping nor the hindlimbs during swimming.** Following methods

123 described above, the null hypothesis was not rejected for time point comparisons of forelimb

124 stepping and hindlimb swimming. Silencing the L2-L5 interneurons did not change the

125 concentration of the phase values at 0.5. Note that in hindlimb swimming, Baseline was

126 significantly different from DOX^{OFF} wherein the phases were more clustered at 0.5. (Critical

127 value of Watson's $U^2_{(0.05, \infty, \infty)} = 0.1869$; Appendix D, Table D.44)²⁰.