

1 **Hominoid arcade shape: pattern and magnitude of covariation**

2

3

4 Stefanie Stelzer¹, Philipp Gunz¹, Simon Neubauer¹, Fred Spoor^{1,2}

5

6 ¹*Department of Human Evolution, Max Planck Institute for Evolutionary Anthropology, Leipzig 04103,*
7 *Germany.*

8 ²*Department of Cell and Developmental Biology, University College London, London WC1E 6BT, UK.*

9

10

11

12

13

14

15

16

17

18

19

20

21 Keywords: maxilla, mandible, premaxilla, integration, jaw, canine

22

23 Correspondence: Stefanie Stelzer, Max Planck Institute for Evolutionary Anthropology, Department of
24 Human Evolution, Deutscher Platz 6, 04103 Leipzig, Germany. E-mail: stefanie_stelzer@eva.mpg.de

25 **Abstract**

26 The shape of the dental arcade and canine size distinguish extant humans from all apes.
27 Humans are characterized by a parabolic arcade with short postcanine tooth rows and small canines,
28 whereas apes have long, U-shaped arcades with large canines. The evolutionary and biomechanical
29 mechanisms underlying arcade shape differences between and within groups are not well understood. It
30 is unclear, for example, whether evolutionary changes in the covariation among modules comprising the
31 upper and lower jaws are the cause and/or consequence of different arcade shapes. Here we use 3D
32 geometric morphometric methods to explore to what extent the morphological differences in arcade
33 shape between living hominoids are related to differences in covariation of upper and lower jaws, and
34 the premaxilla and the maxilla. We show that all extant hominoids follow a very similar covariation
35 pattern between upper and lower dental arcades, as well as between the premaxilla and the maxilla. We
36 find comparably high magnitudes of covariation between the premaxilla and the maxilla in all groups.
37 Between the upper and lower jaws, levels of covariation are similar in apes (*Pan*, *Gorilla*, *Pongo*, and
38 *Hylobates*), but overall lower in extant humans. Our results demonstrate an independence of the
39 pattern of arcade shape covariation from dental spatial arrangements. Importantly, we show that a
40 shared hominoid pattern of covariation between premaxilla and maxilla together with the covariation of
41 upper and lower jaw is consistent with major evolutionary arcade shape changes in hominoids. We
42 suggest that with the reduction of canine and diastema size in hominins, the incisors move posteriorly
43 and the tooth row becomes more parabolic. Our study provides a framework for addressing questions
44 about fossil hominin dentognathic diversity, including inter- and intraspecific variation and associations
45 of upper and lower jaw morphology.

46

47 **Introduction**

48 Dentognathic morphology of extant humans and apes differs notably in canine size and arcade
49 shape. In apes, the arcades are U-shaped with narrow and near-parallel postcanine tooth rows.
50 Furthermore, dental arcade shape varies within and between ape species: posteriorly diverging or
51 converging postcanine tooth rows result in a range of arcade contours (Hellman, 1919). In some
52 individuals the arcade tapers in the premolar region to diverge again more posteriorly, giving it a saddle
53 shape (Hellman, 1919; Remane, 1921). The large lower canines are accommodated in a diastema
54 between the upper lateral incisor and the enlarged upper canine; in the mandible, the upper canine is
55 accommodated between the lower canine and the lower premolars (Angle, 1899; Hellman, 1942), where
56 canine size can vary strongly inter- and intraspecifically (Almquist, 1974; Leutenegger and Kelly, 1977;
57 Oxnard et al., 1985; Leutenegger and Shell, 1987; Plavcan and van Schaik, 1992; Plavcan, 1993; Kelley,
58 1995).

59 Modern human dental arcades are usually referred to as being parabolic (Angle, 1899; Broomell,
60 1902; Le Gros Clark, 1950; Genet-Varcin, 1969), elliptic (Black, 1902; Garn, 1968; Currier, 1969; Brader,
61 1972) or catenary (MacConaill and Scher, 1949; Scott, 1957; Engel, 1979), with upper and lower arcades
62 having slightly different shapes (Hellman, 1919; Engel, 1979). Incisors and canines are of almost equal
63 size (Schwartz, 1995) and their occlusal surfaces are on the same level as the postcanine dentition.

64 Intra- and interspecific differences in cranial and mandibular architecture among primates have
65 been related to differences in masticatory behavior, including varying mechanical stress (Hrdlicka, 1940;
66 Hylander, 1972; Carlson and Vangerven, 1977; Hylander, 1979; Weijs and Hillen, 1984; Bouvier, 1986;
67 Weijs and Hillen, 1986; Armelagos et al., 1989; Ravosa, 1990; Corruccini, 1991; Herring, 1993; Anapol
68 and Lee, 1994; Larsen, 1995; Cassidy et al., 1998; Sardi et al., 2004; Lieberman, 2008; von Cramon-
69 Taubadel, 2011; Prasad et al., 2013). These biomechanical arguments emphasize that relative position

70 and length of the dental arcade influence the stress distribution (Witzel and Preuschoft, 2002). In this
71 view, phenotypic variation of the upper and lower jaw is seen as a consequence rather than a
72 precondition for different force distributions. In apes relatively long tooth rows and prognathic dental
73 arcades seem to be biomechanically advantageous, whereas in humans the parabolic arcade is seen as a
74 side effect of reduction in bite- and chewing forces and the reorganization of the face (Preuschoft, 1989;
75 Witzel and Preuschoft, 1999) with little biomechanical significance (Preuschoft and Witzel, 2004). Recent
76 studies in capuchin monkeys (Makedonska et al., 2012) and modern humans (Noback and Harvati, 2015)
77 showed that, at least within groups, dental arcade shape (in contrast to size and position) is largely
78 independent from masticatory forces. These authors found correlations between diet and shape of the
79 temporalis muscle and the cranium, but none between subsistence and maxillary arch shape.

80 The evolutionary and biomechanical mechanisms underlying the differences in arcade shape
81 between extant humans and apes, as well as those underlying the within-group variability, are not well
82 understood. The evolutionary trajectory of our lineage is interesting in this regard, as arcade shape
83 varies considerably among fossil hominins (e.g., Weidenreich, 1936; Tobias, 1967; Johanson et al., 1978;
84 Johanson and White, 1979; Greenfield, 1992). Besides a general reduction in absolute canine size, early
85 hominins like *Australopithecus anamensis* and *A. afarensis* possess primitive features such as a small
86 diastema between the upper incisor and the canine, as well as parallel to slightly diverging tooth rows
87 (Schwartz, 1995; White et al., 2000; Ward et al., 2001; Kimbel and Deleuzene, 2009). Some early *Homo*
88 specimens retain long and almost parallel tooth rows, whereas others have shorter postcanine rows and
89 non-projecting frontal tooth rows (Tobias, 1991; Wood, 1991; Rightmire, 1993; Kimbel et al., 1997;
90 Clarke, 2012; Leakey et al., 2012; Spoor et al., 2015). Large-scale differences in jaw shapes have been
91 used to support arguments about species diversity in early *Homo* (Spoor et al., 2015). Moreover, it has
92 been argued that high variation in the anterior maxillary region, the premaxilla, indicates that it might
93 have been subject to selection in hominin evolution (Villmoare et al., 2014). Understanding the

94 mechanisms contributing to the variation and covariation of jaw shapes in extant hominoids is thus
95 informative for interpreting the hominin fossil record.

96 Morphological covariation is the statistical interrelationship of morphometric variables
97 (Mitteroecker and Bookstein, 2007). It is the consequence of developmental, evolutionary, genetic, and
98 functional processes (Cheverud, 1996a) that in combination lead to integration of structures. Integrated
99 traits must covary, however, traits that covary are not necessarily integrated (Villmoare et al., 2014).
100 Although not equivalent (compare Porto et al., 2009; Young et al., 2010; Grabowski et al., 2011),
101 integration and covariation are mutually dependent in that covarying structures influence the way an
102 organism can react to evolutionary forces. On the other hand, evolution can also act on the covariation
103 of structures. Using 3D geometric morphometric methods we explore the pattern and magnitude of the
104 covariation between and within upper and lower arcades in extant hominoids. We seek to understand if
105 and how the morphological differences in arcade shape between living hominoids are related to
106 differences in covariation of upper and lower jaws.

107 Previous studies of cranial integration and covariation have consistently demonstrated
108 conserved patterns among primates and even mammals (Cheverud, 1996b; Ackermann and Cheverud,
109 2000; Lieberman et al., 2000; Marroig and Cheverud, 2001; Ackermann, 2002; Gonzalez-Jose et al.,
110 2004; Marroig et al., 2004; Ackermann, 2005; Goswami, 2006; Gunz and Harvati, 2007; Mitteroecker and
111 Bookstein, 2008; Porto et al., 2009; Makedonska et al., 2012; Villmoare et al., 2014). In this study we
112 assess (1) whether the same applies to hominoid upper and lower dental arcades, or (2) whether the
113 variation in extant arcade shapes is associated with a variation of the underlying patterns of covariation.

114 The magnitude of covariation has been related to material properties of food, in that taxa
115 relying on a mechanically challenging diet have stronger integrated upper and lower jaws (Marroig and
116 Cheverud, 2001; Makedonska et al., 2012). *Pan*, *Pongo*, *Gorilla*, and *Hylobates* consume different

117 proportions of leaves, fruit, other plant parts, or insects, and their dietary preferences are dependent on
118 season, food availability, habitat and sex. In general, however, *Pan*, *Pongo*, and *Hylobates* are more
119 frugivorous, whereas *Gorilla* is more reliant on leaves (MacKinnon, 1974; Rijksen, 1978; Gittins and
120 Raemaekers, 1980; Watts, 1984; Galdikas, 1988; Tutin and Fernandez, 1993; Knott, 1998; McConkey et
121 al., 2003; Rogers et al., 2004; Boesch et al., 2006; Doran-Sheehy et al., 2009). Morphologically, this
122 difference is reflected in relative dental sizes. Frugivores have relatively larger incisors, while foliovores
123 have relatively larger molars (Hylander, 1975; Kay and Hylander, 1978). In comparison to apes, modern
124 humans have a more generalist diet, where extra-oral food processing via tool use or cooking is thought
125 to have reduced the masticatory effort since the emergence of our species or even earlier (e.g. Brace et
126 al., 1987; Wrangham et al., 1999; Richards et al., 2001; Teaford et al., 2002; Henry, 2010). While a
127 correlation with diet seems likely, some authors have suggested that the lower covariation magnitudes
128 in the human cranium (Marroig et al., 2009; Porto et al., 2009) and pelvis (Grabowski et al., 2011) may
129 indicate a general relaxation of integration in the hominin lineage. These authors argued that the
130 change in magnitude of integration might have paved the way for the development of the
131 morphological changes characterizing our lineage. In addition to the two aforementioned aims, we
132 therefore also assess (3) whether there are differences in the magnitude of covariation between the
133 groups, and (4) whether there are differences between females and males within groups.

134 It has been argued that within the maxilla, the premaxilla represents its own module that has
135 been a target of selection in the hominin lineage leading to higher variation in the anterior region in
136 hominins compared to other anthropoids (Villmoare et al., 2014). The reduction of canine and thereby
137 diastema size is associated with an earlier closure of the premaxillary sutures (McCollum and Ward,
138 1997; Braga, 1998). We explore whether those differences are associated with a change in the pattern
139 or the magnitudes of covariation. We therefore also assess (5) the pattern and magnitudes of
140 covariation between the premaxilla and the maxilla.

141

142 **Materials and methods**

143 *Samples*

144 Our samples are summarized in Table 1 and in the Supplementary Online Material (SOM) Table
145 S1. The modern human sample ($n=53$) comprises geographically diverse pre-industrial populations, and
146 was obtained from the Institute of Anatomy at Leipzig University (ULAC), Germany; the Natural History
147 Museum London, UK; the National Historical Museum, Buenos Aires, Argentina; the American Museum
148 of Natural History, New York; and the Smithsonian Institution, Washington, D.C., USA. The *Pan*
149 *trogodytes* sample ($n=44$) is comprised of specimens from Côte d'Ivoire, Liberia, Gabon, Cameroon and
150 Equatorial Guinea, housed at the Senckenberg Museum, Frankfurt, Germany; at the Max Planck Institute
151 for Evolutionary Anthropology in Leipzig (MPI-EVA), Germany; the Phyletisches Museum, Jena,
152 Germany; and the Smithsonian Institution; plus one zoo specimen housed in the Phyletisches Museum
153 Jena. The *Gorilla* sample ($n=53$) includes *Gorilla gorilla* specimens from Cameroon, Gabon, and the
154 Congo, and *Gorilla beringei* from Rwanda, housed at the Museum für Naturkunde Berlin, Germany; the
155 Phyletisches Museum, Jena and the Smithsonian Institution. The *Pongo* sample ($n=52$) comprises *P.*
156 *pygmaeus* and *P. abelii* specimens from Borneo and Sumatra, respectively, and one specimen with
157 unknown provenance. The *Pongo* specimens are housed at the Museum für Naturkunde Berlin and the
158 Smithsonian Institution. The *Hylobates* specimens ($n=63$) include the species *H. agilis* (Sumatra), *H.*
159 *albibarbis* (Borneo), *H. klossii* (Sumatra), *H. lar* (Thailand, Myanmar), and *H. muelleri* (Borneo). The
160 specimens are housed at the Museum für Naturkunde Berlin and the Smithsonian Institution. Sex
161 attributions were taken from museum records when available. Sex of the remaining specimens of *Pan*,
162 *Gorilla* and *Pongo* was determined by several observers based on the formation of cranial
163 superstructures and canine size.

164 We used computed tomography (CT) of all specimens, including both industrial CT (BIR ACTIS
165 225/300 at the Max Planck Institute for Evolutionary Anthropology, Leipzig, Germany; isotropic voxel
166 sizes 0.03 – 0.093 mm) and medical CT (Vivantes Klinikum Berlin, Germany; CIMED, La Plata, Argentina;
167 the Senckenberg Museum, and the Smithsonian Institution, Washington, U.S.A. (modern human CT
168 scans obtained as part of Copes (2012)); pixel sizes 0.13 –0.47 mm, slice intervals 0.33 - 0.50 mm). Our
169 sample comprises scans of adult individuals (defined as third molars in occlusion) that preserved both
170 cranium and mandible and did not display malocclusions or other severe pathologies. So as to achieve
171 comparably large sample sizes for all groups that meet these strict criteria, we pooled the respective
172 species of *Gorilla*, *Pongo*, and *Hylobates* in our analyses. Species specific morphologies within genera are
173 known (e.g. Groves, 1970, 1972; Rörer-Ertl, 1984; Uchida, 1998; Guy et al., 2003; Schmittbuhl et al.,
174 2007, but see Groves et al. 1992; Courtenay et al. 1998 for a different view on *Pongo*), however, the
175 differences in dental arcade shape seem to be negligible, when compared to the shape differences
176 between genera, as can be seen in SOM Figures S1-S3, in which the species of each individual are color
177 coded to illustrate that they are randomly scattered within the convex hulls of their respective genera.

178

179 *Data*

180 The occlusal surfaces of teeth are modified by wear, which makes placing homologous
181 landmarks problematic. Hence, we took landmarks on the alveolar margin of every tooth and on the
182 cervix of the postcanine dentition in order to capture arcade shape, spatial arrangement of the teeth,
183 and dental size proportions. We recorded 224 homologous 3D landmarks on the mandibular and the
184 maxillary dental arcades (112 landmarks each) (Fig. 1). All measurements were taken in Avizo 7.1
185 (Visualization Sciences Group).

186 To capture overall length and breadth patterns, landmarks were placed on the distal, buccal,
187 mesial and lingual surfaces of the incisor, canine and premolar alveolus. On the molars, besides a distal
188 and a mesial landmark, we set two landmarks buccally and two lingually at the position of each root. On
189 the third molar we took one landmark buccally and lingually at the position of the mesial root. The
190 cervical landmarks were positioned distally, buccally, mesially, and lingually.

191 - Table 1 -

192 - Figure 1 -

193 *Analysis*

194 Landmarks on the upper and the lower jaw were superimposed separately applying Generalized
195 Procrustes Analysis (GPA) to extract shape information independent from rotation, translation and
196 scaling (Rohlf and Slice, 1990; Bookstein, 1991). We performed a principal component analysis (PCA) on
197 the Procrustes shape coordinates for mandibles and maxillae separately to assess taxonomic shape
198 differences.

199 Landmarks on the premaxilla (incisors) and the maxilla (canines, premolars, and molars) were
200 superimposed together to retain information on the exact position of the premaxilla relative to the
201 maxilla. In the analysis of the covariation between premaxilla and maxilla, only the alveolar, not the
202 cervical, landmarks were used to capture the morphology of the postcanine dentition.

203 Pattern of covariation A two-block partial least squares (PLS) analysis was used to quantify the
204 covariation of the mandibular and the maxillary arcade shape, or the premaxilla and the maxilla,
205 respectively (Rohlf and Corti, 2000; Bookstein et al., 2003). We used the mean of the original data and
206 the reflected relabeled landmark configuration in order to symmetrize the data and therewith remove
207 asymmetric shape variation that also arises from asymmetric measurement error (Klingenberg and
208 McIntyre, 1998; Mardia et al., 2000; Mitteroecker and Gunz, 2009). The PLS analysis computes

209 correlated pairs of vectors, so-called singular warps (SW), for the maxilla and for the mandible, or the
210 premaxilla and the maxilla, respectively, that account for the maximum covariance between the two
211 sets of landmarks (Wold, 1966; Rohlf and Corti, 2000; Bookstein et al., 2003).

212 We performed two PLS analyses per data set: one includes all groups with the data mean
213 centered for each group. This explores the overall pattern of covariation while accounting for large-scale
214 differences between the groups. The second PLS analysis was performed for every group separately,
215 mean centered by sex. If all groups follow a similar pattern of covariation, shape variation associated
216 with PLS axes of the pooled sample and within groups should be similar.

217 Magnitude of covariation We used two metrics to quantify the magnitude of covariation; the
218 covariance ratio (CR, Adams, 2016) and the correlation coefficient between the singular warp scores
219 (Bookstein et al., 2003). The CR has been proposed recently by Adams (2016) as an alternative to the RV
220 coefficient that overcomes some of the fundamental methodological issues associated with the RV
221 coefficient (Smilde et al., 2009; Fruciano et al., 2013; Bookstein, 2016). The CR quantifies the overall
222 pattern of covariation between the blocks. Therefore one does not know which aspect of covariation the
223 CR is quantifying. In contrast, the correlations of the singular warp scores pertain to the pattern
224 visualized in the respective singular warp plots, and are easier to interpret. As our sample sizes are not
225 equal across all groups, we used a resampling approach for computing the values for the CR. To get a
226 representative distribution, the CR was calculated for 30 randomly selected individuals in 1000 iterations
227 per group.

228 A permutation test (Good, 2000) was used to determine the significance of the mean
229 differences between the permuted CR values (Bonferroni-corrected for multiple testing, considered
230 significant at $\alpha < 0.05$).

231 Sexual dimorphism A permutation test (Good, 2000) was used to determine the significance of
232 the mean shape differences between male and female mandibles and maxillae. To do so, we randomly
233 selected individuals and assigned them to male or female and computed a mean shape and shape
234 difference between the permuted sexes 10 000 times (considered significant at $\alpha < 0.05$).

235 Allometry To assess the amount of the total shape variance that can be explained by jaw size we
236 separately computed multivariate regressions of upper and lower arcade shapes on the natural
237 logarithms of their centroid sizes. We evaluated the statistical significance of these regressions using a
238 permutation test based on the explained variance (Mitteroecker et al., 2013).

239 Intra-observer error All data were measured by one of the authors (S.S). Intra-observer error
240 was assessed by an analysis of repeated measurements: one specimen (*Gorilla gorilla*, ZMB 14645,
241 medical CT scan, voxel size 0.227x0.227x0.335 mm) was measured fifteen times. The largest Procrustes
242 distance between repeated measurements of this individual was considerably smaller than the smallest
243 Procrustes distance between different specimens of the same species. Specimen affinity is therefore not
244 affected by intra-observer error.

245 All analyses and visualizations were performed in Mathematica 8.0 (Wolfram Research Inc., 2010).

246

247 **Results**

248 *1. PCA*

249 Table 2 shows the results of the principal components (PC) analysis. The first PC (PC1, 66.9% of
250 total shape variance in maxillae, 70.8% in mandibles) separates humans from the other groups. Both,
251 maxillae (Fig. 2a) and mandibles (Fig. 2c), have short parabolic arcades and small canines on the positive
252 end (humans) compared to a U-shaped arch, long parallel tooth rows with large canines on the negative
253 side (apes).

254 For the maxillae, PC2 (11.0% of the total shape variance) describes the relation between a
255 relatively large postcanine dentition with relatively small incisors in the positive direction (*Gorilla*); and
256 relatively large incisors with a relatively small postcanine dentition in the negative direction (*Pan*) (Fig.
257 2a). For the mandibles (10.9% of the total shape variance), PC2 combines a long postcanine tooth row
258 with small incisors on the positive end (*Gorilla*), whereas the negative extreme of PC2 combines a short
259 postcanine tooth row with a large anterior dentition (*Pan*) (Fig. 2c).

260 PC3 (5.8% of total shape variance in maxillae, 6.3% for mandibles) separates *Hylobates* from the
261 other hominoids. For the maxilla in the negative direction, relatively small postcanine teeth are
262 accompanied by relatively large canines and small incisors; the arcade is slightly V-shaped (*Hylobates*)
263 (Fig. 2b). For the mandibles, moderately sized canines are associated with relatively large postcanine
264 teeth and incisors in the positive direction of PC3 (other groups). The lower scores of PC3 describe
265 relatively small postcanine dentition associated with relatively small incisors and slightly enlarged
266 canines (*Hylobates*).

267 - Figure 2 –

268 - Table 2 –

269 2. Pattern and magnitudes of covariation between the upper and the lower arcades

270 Pattern of covariation (all groups, group-mean centered) Table 3 shows the percentages of the
271 explained covariance for the first five PLS components. On the positive side of PLS1 (61.4% of the total
272 covariance, $r = 0.73$), parabolic mandibles are associated with parabolic maxillae. On the negative side,
273 maxillary and mandibular arcades are U-shaped (Fig. 3a). Humans have positive PLS1 scores and are
274 separated from the apes that have decreasing scores along a diagonal line, starting at chimpanzees, to
275 gibbons, orang-utans and then gorillas.

276 At the negative extreme of PLS2 (21.6% of the total covariance, $r = 0.81$) large maxillary canines
277 are associated with large mandibular canines. The postcanine dentition is relatively small, short and
278 parallel. At the positive extreme, small mandibular canines and V-shaped arcades are associated with
279 small maxillary canines and arcades that are more rounded than the mandibles (Fig. 3b). Humans have
280 positive PLS2 scores and are separated from the apes along a diagonal line.

281 At the negative end of PLS3 (7.9% of the total covariance, $r = 0.73$), both arcades are broad and
282 converge distally (more pronounced in the maxilla). In the maxillary arcade, the front teeth lie on the
283 bicanine line. At the positive end of PLS3, both upper and lower arcades are V-shaped. The anterior
284 region is narrow and projecting anteriorly (more pronounced in the maxilla) (Fig. 3c). Separation among
285 groups along the PLS3 axis is less clear.

286 In all PLS dimensions the specimens of all five groups scatter around a diagonal line, indicating
287 that the patterns of covariation of all groups are similar along these PLS axes. In PLS2 *Hylobates*
288 specimens are shifted away from the hominid trajectory, but their trajectory has the same slope.

289 - Figure 3 -

290 - Table 3 -

291 Pattern of covariation (groups separate, sex-mean centered) Table 3 shows the percentages of
292 the explained covariance for the first five PLS components for each group. At the positive end of PLS1, in
293 all groups, rounded maxillae are associated with broad and rounded mandibles (Fig. 4 a-e). In the apes,
294 the maxillary arcade is anteriorly wider than the mandibular arcade. At the negative end of PLS1, long
295 and straight postcanine maxillary arcades are associated with straight mandibular postcanine arcades
296 that taper slightly in the premolar region. In *Hylobates*, PLS1 and PLS2 are interchanged, so that the
297 aforementioned shape changes are described by PLS2 (SOM Fig. S4). In addition, the maxillary arcade in
298 *Hylobates* also tapers in the premolar region. The anterior regions in both upper and lower arcades are

299 rounded and extend anteriorly. PLS scores of PLS2 and PLS3 and wireframes of the first three PLS
300 components are shown in SOM Figs. S4 and S5.

301 At the positive end of PLS2 in all groups, straight diverging posterior maxillary arcades are
302 associated with likewise diverging mandibular arcades. At the negative end of PLS2, straight converging
303 posterior maxillary tooth rows are associated with mandibular tooth rows that behave similarly. These
304 shape changes are described by PLS1 in *Hylobates*. In addition, on the positive side of this PLS
305 component, *Hylobates* is more straight than converging. In *Homo*, PLS2 and PLS3 are interchanged. The
306 incisal part of the upper and lower arcades in all groups are flat, with the exception of *Homo* and
307 *Hylobates*, where the mandibular arcades are more rounded anteriorly (SOM Figs. S4 and S5).

308 At the negative end of PLS3 in all groups, straight posterior tooth rows and flat anterior teeth in
309 the maxilla are associated with likewise shaped mandibular arcades. In *Pan*, *Gorilla* and *Pongo*, the
310 postcanine tooth rows are parallel, in *Homo* and *Hylobates* the arcades diverge in both upper and lower
311 jaws. At the positive end of PLS3, anteriorly extending incisors and large diastemata in the maxilla are
312 associated with long straight mandibular arcades, where the incisors extend anteriorly as well. In *Homo*
313 both arcades are more rounded in the postcanine row and less pointed in the anterior region (SOM Figs.
314 S4 and S5).

315 Overall, all groups show similar covariation patterns.

316

317 Morphological comparison between the variation within blocks and the covariation between
318 blocks The associated shape (co)variation between the maxillary arcades and the mandibular arcades
319 along PLS1 and PLS2 is comparable to the shape variation within the maxillae and within the mandibles
320 along PC1 (i.e., parabolic arcades with small canines versus long U-shaped arcades with large canines).

321 Also the shape changes along PLS3 are partly represented by the variation described by PC3 (broad
322 versus V-shaped arcades).

323

324 Magnitude of covariation (all groups combined) Results for the covariance ratio (CR) are given
325 in Table 4. The distribution of values for the permuted CRs from varied sample composition is shown
326 in Fig. 5. The apes overlap completely with CRs varying from 0.71 to 0.89. *Homo* shows values between
327 0.64 and 0.89. Median values are 0.81 (*Pongo*), 0.81 (*Pan*), 0.80 (*Gorilla*), 0.81 (*Hylobates*), and 0.76
328 (*Homo*). *Homo* is significantly different from every other group ($p < 0.02$, Bonferroni corrected). *Pan*,
329 *Gorilla*, *Pongo*, and *Hylobates* are not significantly different from each other ($p > 0.12$ at least) (Table 4).

330 The correlation coefficient of PLS1 is lowest in *Gorilla*, followed by *Homo*, *Pongo*, and *Pan*.
331 *Hylobates* show the highest value (Table 5). The correlation coefficient of PLS2 is lowest in *Hylobates*,
332 followed by *Homo*. *Pan*, *Pongo*, and *Gorilla* show higher values. In PLS3, the correlation coefficient is
333 lowest in *Hylobates* and *Homo*. *Gorilla* falls between the latter two and *Pongo* and *Pan*.

334 - Figure 5 -

335 - Table 4 -

336 - Table 5 -

337 Magnitude of covariation (groups separate) We used a subsample of 15, i.e., the smallest
338 number of subsamples (female *Homo*) minus one, and considered the bootstrapping distribution
339 resulting from 1000 iterations for the calculation of the CR (Table 6). Correlation coefficients are given in
340 Table 7.

341 Overall, female *Pan*, *Gorilla*, *Pongo* and *Homo* show higher mean values in the covariance ratio
342 than their male conspecifics (SOM Fig. S6) in the CR. In *Hylobates*, however, the males show higher
343 values.

344 In PLS1 female *Gorilla*, *Pongo*, and *Homo* show higher correlation coefficients than in their male
345 counterparts. *Hylobates* show the opposite pattern, so do male *Pan* even though the difference from
346 the females is small. In PLS2 male *Pan*, *Gorilla*, and *Homo* show higher values than the females. *Pongo*,
347 *Homo*, and *Hylobates* show the opposite pattern. In PLS3 males show higher values than females in all
348 groups.

349 - Table 6 -

350 - Table 7 -

351 3. Allometry

352 For the maxillary arcades, size explains 14.3% of the total variance in *Gorilla*, 12.6% in *Pongo*,
353 6.8% in *Pan*, 2.5% in *Homo*, and 2.4% in *Hylobates* ($p < 0.01$). For the mandibular arcades size explains
354 8.5% of the total variance in *Gorilla*, 9.7% in *Pongo*, 4.8% in *Pan*, 2.4% in *Homo*, and 3.6% in *Hylobates*
355 ($p < 0.01$) (Table 8).

356

357 4. Sexual dimorphism

358 Mean arcade shapes of female and males differ significantly (all p -values smaller than 0.05) in
359 *Gorilla*, *Pan* and *Pongo*. While the mandible shapes of female and male *Homo sapiens* are not
360 statistically different, the maxillae fall just below the significance level ($p = 0.047$). There is no significant
361 shape difference between female and male *Hylobates* (Fig. 6).

362 The main difference between male and female great apes is basal crown area of the canine. In
363 *Pongo* and *Gorilla*, this is associated with small differences in the anterior region. In females, the incisors
364 are shifted slightly posteriorly. In the postcanine dentition, the premolars are shifted anteriorly, whereas
365 the molar region remains unaffected. In humans, the subtle difference is also canine crown area.

366 - Figure 6 -

367 5. Pattern and magnitudes of covariation between the premaxilla and the maxilla

368 Pattern of covariation (all groups, group-mean centered) Table 9 shows the percentages of
369 explained covariance for the first five PLS dimensions. In PLS1 (84.1% of the total covariance, $r = 0.96$),
370 all groups scatter along a diagonal (Fig. 7a). At the negative end of PLS1 (gorillas followed by the other
371 non-human apes) large incisors are placed far anteriorly, creating a diastema between the maxilla and
372 the premaxilla. The posterior dentition is parallel, and canines are large. At the positive end (humans),
373 incisors and canines are small and incorporated in the dental arcade, and the posterior arcade is
374 parabolic (Fig. 7a).

375 In PLS2 (10.2% of the total covariance, $r = 0.59$), the groups still cluster along the diagonal, but
376 are shifted parallel from each other with large overlap among groups (Fig. 7b). At the negative end of
377 PLS2, large spaciouly arranged incisors are associated with rounded posterior arcades and there is no
378 diastema. At the positive end of PLS2, smaller incisors are positioned in closer proximity and are placed
379 anteriorly followed by a diastema and the posterior dentition is straight and parallel (Fig. 7b).

380 - Figure 7 -

381 - Table 9 -

382 Pattern of covariation (groups separate, sex-mean centered) Table 9 shows the percentages of
383 explained covariance for the first five PLS components for each group separately. SOM Figure S7 shows
384 the PLS scores for every group in the first three PLS dimensions.

385 At the negative end of PLS1 for every group, anteriorly expanding incisors are associated with
386 straight, parallel posterior tooth rows. In *Pan*, *Gorilla*, and *Pongo*, diastemata are present. At the
387 positive end there are no diastemata between the premaxillae and the maxillae. The incisors are
388 incorporated in the dental arcade. The posterior row is more rounded (SOM Fig. S8, top row).

389 In the negative direction of PLS2, rather flat anterior regions are associated with wide, more
390 rounded posterior arcades. At the positive end, the incisal region extends anteriorly, creating a diastema
391 between the premaxilla and the maxilla. The posterior arcade is straight. *Gorilla* differs in showing
392 straight posterior arcades in both directions; *Homo* shows no diastema and the posterior arcades are
393 parabolic in both directions (SOM Fig. S8, middle row).

394 PLS3 describes the spacing of the incisors. In the negative direction, incisors are narrowly
395 positioned. In the positive direction, incisors are widely spaced (SOM Fig. S8, bottom row).

396 The associated shape changes along all three PLS dimensions in the separate PLS analysis
397 correspond to the shape changes observed in the combined analysis.

398

399 Magnitudes of covariation (all groups) Magnitudes of covariation (covariance ratio) are slightly
400 smaller between the premaxilla and the maxilla than between the upper arcade and the lower arcade in
401 the apes (Fig. 8, Table 10). In *Homo*, the values are comparable.

402 *Gorilla* is significantly different from *Homo* and *Hylobates* ($p < 0.015$, Bonferroni corrected), but
403 not from *Pongo*. The difference between *Gorilla* and *Pan* is close to the significance level ($p \leq 0.0559$).
404 *Pan*, *Pongo*, *Homo*, and *Hylobates* are not significantly different from each other ($p > 0.27$ at least) (Table
405 10).

406 The correlation coefficient in PLS1 is lowest in *Pan* and *Hylobates*, followed by *Pongo*, *Homo* and
407 *Gorilla* (Table 11). In PLS2, the correlation coefficient is lowest in *Gorilla*, followed by *Pongo*. *Homo* and

408 *Hylobates* show similar values. *Pan* shows the highest correlation coefficient. In PLS3, *Gorilla* shows the
409 highest value, followed by *Pongo*, *Pan*, and *Hylobates*. *Homo* shows the lowest correlation coefficient.

410 - Figure 8 –

411 - Table 10 –

412 - Table 11 -

413 Magnitudes of covariation (groups separate) We used a subsample of 15, i.e., the smallest
414 number of subsamples (female *Homo*) minus one, and considered the bootstrapping distribution
415 resulting from 1000 iterations for the calculation of the CR (SOM Table S2). Correlation coefficients are
416 given in SOM Table S3.

417 *Pan* and *Gorilla* show a marked sexual dimorphism with females showing lower values than
418 males in the covariance ratio (SOM Table S2). In *Pongo*, *Homo*, and *Hylobates*, male and female
419 distributions overlap. In all groups, magnitudes of covariation are in general slightly smaller between the
420 premaxilla and the maxilla than between the upper and the lower arcades. Only male *Gorilla* show
421 higher magnitudes (SOM Fig. S9).

422 In PLS1 male *Pan*, *Gorilla*, *Hylobates* show higher correlation coefficients than their female
423 counterparts. *Homo* shows the opposite pattern, in *Pongo* values are similar (SOM Table S3). In PLS2
424 male *Pan*, *Gorilla*, and *Pongo* show higher values than females. In *Homo* and *Hylobates* the opposite is
425 true. In PLS3 correlation coefficients are low in all groups except for *Pan*, where males show higher
426 values than females.

427

428 **Discussion**

429 In this study we sought to explore the pattern and magnitude of the covariation between the
430 upper and the lower jaw, as well as between the premaxilla and the maxilla in extant hominoids. In the
431 first three PLS components, that together explain more than 90% of the total covariance of the upper
432 and the lower arcade, the data points scatter along the diagonal (Fig. 3). Such an arrangement is
433 interpreted as the same pattern of covariation (e.g. Mitteroecker and Bookstein, 2008). While retaining
434 the same slope, *Hylobates* are shifted from the hominid trajectory in PLS2 (Fig. 3b), probably owing to
435 the presence of absolutely and relatively large canines in males and females in this group.

436 We also performed separate PLS analyses which showed that the related shape changes of the
437 mandibular and the maxillary arcades are similar across species (Fig. 4 and SOM Figs. S4 and S5). In
438 general, this result suggests a common hominoid pattern of covariation and that the evolutionary shape
439 changes that led to hominoid arcade variation are not associated with modification of the underlying
440 covariation of arcades. Importantly, our data demonstrate an independence of the pattern of
441 covariation, which is similar in the analyzed species, from dental spatial arrangements, which are
442 different in the analyzed species. This conclusion is in accordance with earlier studies that suggest
443 conserved covariation patterns in primates and other mammals in cranial parts other than the jaws
444 (Cheverud, 1996b; Ackermann and Cheverud, 2000; Lieberman et al., 2000; Marroig and Cheverud,
445 2001; Ackermann, 2002; Gonzalez-Jose et al., 2004; Marroig et al., 2004; Ackermann, 2005; Goswami,
446 2006; Gunz and Harvati, 2007; Mitteroecker and Bookstein, 2008; Porto et al., 2009; Makedonska et al.,
447 2012; Villmoare et al., 2014).

448 To evaluate the magnitude of the covariation we used the covariance ratio (CR, Adams, 2016)
449 and the correlations between upper and lower arcades, or the premaxilla and the maxilla, respectively.
450 For the CR we considered the distributions from permutations. *Homo* stands out in showing lower
451 overall CR values (Fig.5, Table 4). Covariation magnitudes of individual PLS axes are quantified by the
452 correlation coefficient. When the first PLS is considered, which explains almost two thirds of the total

453 covariance (i.e., arcade shapes from U-shaped to parabolic), *Gorilla* shows the weakest covariation,
454 followed by *Homo*, *Pongo*, and *Pan* which show similar covariation magnitudes. *Hylobates* has slightly
455 higher values (Table 5). Thus the magnitude of covariation does not reflect the shape gradient from U-
456 shaped to parabolic, and might be more taxon specific than arcade shape specific. In PLS2, which
457 explains almost 22% of the total covariance (i.e., canine size), *Homo* and *Hylobates* display lower
458 correlations than the other groups. Interestingly, while it seems as if absolute canine size itself is of
459 minor importance for the magnitude of covariation, this result might indicate that sexual dimorphism in
460 canine size contributes to the magnitude of covariation, in a way that less dimorphic taxa show lower
461 magnitudes of covariation. In PLS3, that explains about 8% of the total covariance (i.e., spatial
462 arrangement of anterior dentition and postcanine divergence), *Homo* and *Hylobates* also show lower
463 values, followed by *Gorilla* which falls in-between the latter and *Pan* and *Pongo* (Fig. 5d). These
464 differences in covariation magnitudes might be related to different strategies of incorporating large
465 canines into the dental arcade. While in *Pan*, *Pongo* and *Gorilla* intercanine distance is enlarged, which
466 results in a flat anterior region, in *Hylobates* the incisal region is extended anteriorly, resulting in a V-
467 shaped arcade. The lower magnitudes in *Homo* might be the consequence of the generally reduced
468 canines. In combination with the results for the overall measure for the magnitude of covariation these
469 findings imply that the lower magnitude in modern humans is not a reflection of the large-scale
470 differences between humans and apes.

471 While hominoids do show variation in dental morphology and dental size proportions, the
472 pattern of the interplay between upper and lower arcades remained generally unchanged. This suggests
473 that selection acted on maintaining a proper fit between the upper and the lower arcade. The lower
474 magnitudes of maxillo-mandibular covariation in modern humans which we report here may be related
475 to a systemic change in the hominin lineage. It was shown previously that magnitudes of covariation in
476 the cranium and the pelvis are relaxed in modern humans in general compared to other primates and

477 mammals (Marroig et al., 2009; Porto et al., 2009; Grabowski et al., 2011). Following this argument, a
478 relaxation initially allowed new body plans characteristic of hominins to emerge, potentially including
479 the development of the parabolic arcade shape. If lower magnitudes can account for the development
480 of hominin morphologies then they should be identifiable before new morphologies appear, i.e., at or
481 shortly after the split from the chimpanzee lineage. While the resolution of the current fossil record
482 might not be good enough to determine when this relaxation occurred in the hominin lineage, fossil
483 evidence from about 2.1 to 1.5 million years ago suggests that bipedal adaptations, brain enlargement
484 and short and rounded dental arcades are not necessarily associated with each other. While brain size of
485 *H. habilis*, *H. rudolfensis* and *H. erectus* largely overlap in range, they show distinct facial morphologies
486 including the primitive morphology of long and parallel post-canine tooth rows in *H. habilis*, the derived
487 condition in *H. erectus*, and the distinct morphology of *H. rudolfensis* with short and parallel post-canine
488 tooth rows plus a flat anterior region (Spoor et al., 2015). While these seemingly conflicting lines of
489 evidence clearly need more data, another, mutually non-exclusive, explanation for a lower covariation in
490 *Homo* compared to all apes could be diet. It has been found that those platyrrhine species consuming
491 softer foods show lower magnitudes of covariation than species reliant on harder foods (Marroig and
492 Cheverud, 2001; Makedonska et al., 2012). A similar relationship could contribute to the difference we
493 found between apes and modern humans. This would be in accordance with the notion that at least in
494 *H. sapiens* the masticatory effort was reduced due to extra-oral food processing via tool use and cooking
495 (e.g. Brace et al., 1987; Wrangham et al., 1999; Richards et al., 2001; Teaford et al., 2002; Henry, 2010).
496 This explanation, however, does not sufficiently explain the differences we observe between ape
497 genera, which show similar magnitudes of covariation but at the same time different dietary habits.

498 We also analyzed the pattern and magnitudes of covariation between the premaxilla and the
499 maxilla. Corroborating the results of Villmoare et al. (2014) the magnitudes of covariation between the
500 premaxilla and the maxilla are in general slightly lower than between the upper and the lower jaw.

501 In the first PLS components, all groups follow a similar pattern of covariation between the
502 premaxilla and the maxilla. When canines are large, incisors are placed anteriorly to create a diastema
503 for the lower canine. The posterior dentition is long and parallel. With the reduction of canine and
504 diastema size, incisors move posteriorly and the posterior arcades get more parabolic (PLS1). This
505 finding of a shared pattern of covariation between the premaxilla and the maxilla as well as *Homo* falling
506 within the apes' range of covariation magnitude is in accordance with Villmoare et al. (2014). Either
507 result shows that the variation in hominin premaxilla is not associated with a change in the pattern of
508 character covariation. Previous studies suggested that there is a relation between anterior tooth size
509 and premaxilla size (Bromage, 1989; Mooney and Siegel, 1991; Lockwood, 1997; Braga, 1998). Our
510 results show that canine and diastema size contributes substantially to maxillary arcade shape.

511 To assess differences related to sexual dimorphism, we evaluated magnitudes of covariation for
512 females and males in each group between the upper and the lower jaw (SOM Fig. S6) and between the
513 premaxilla and the maxilla (SOM Fig. S9). Magnitudes of the latter overlap almost completely in all
514 groups except for *Pan* and *Gorilla*, where males show higher values. Mean magnitudes between the
515 upper and the lower jaw are higher in females except for *Hylobates*, where males have higher values.
516 Due to small sample sizes in the within-group comparisons these results should be treated with caution.
517 Additionally, an uncertainty about modern human sex attribution must be considered, since their sexing
518 is based on traditional morphological methods, rather than actual records (dissection, autopsy or
519 graveyard). In general, it is probable that the mechanisms responsible for the results presented here
520 might be multiple and different for each taxon as there are different patterns and magnitudes of
521 dimorphism between species (Plavcan, 2002).

522 Our results suggest that variation in hominoid arcade shape is not the consequence of a change
523 of the underlying covariation patterns. Our results provide a model for the evolutionary arcade shape

524 change in the hominin lineage: with the loss of interlocking canines and the decreased need for space in
525 the antagonistic arcade in the hominin lineage, the tooth rows become more rounded and eventually
526 parabolic. Driven by the necessity of the upper and the lower jaw to form an effective masticatory
527 apparatus, the morphological covariation between the mandibular and the maxillary arcade has been
528 maintained.

529 The strong focus on dentognathic structures in palaeoanthropology is due to their good
530 preservation and their taxonomic significance. Nonetheless, our limited understanding of the degree
531 and pattern of intraspecific variation has resulted in different views regarding the number of species
532 that can be recognized in the fossil record (e.g. Wood, 1992; Rightmire, 1993; Leakey et al., 2012).
533 Another factor adding to differences in hypodigm composition is that there is a lack of consensus over
534 which maxillae and mandibles can be accommodated within a single species (Spoor et al., 2015). The
535 results of the current study will help address these issues, as they provide reference data which
536 document the intraspecific and interspecific variation of the upper and lower dental arcades in extant
537 taxa. Moreover, the degree and pattern of covariation between upper and lower jaws can help with
538 identifying the probability that certain maxillae and mandibles in the fossil record are conspecific. Using
539 a smaller landmark set, we have shown recently that this approach can indeed be used to recognize
540 distinct morphs within early *Homo*, identifying those pairs of maxillae and mandibles which, based on
541 extant hominid variation, can be excluded statistically from belonging to a single species (Spoor et al.,
542 2015).

543

544 **Conclusion**

545 We found that *Pan*, *Gorilla*, *Pongo*, *Hylobates*, and *Homo* share a very similar covariation pattern
546 between the upper and the lower arcade, as well as between the premaxilla and the maxilla, that is

547 independent from dental spatial arrangements. We suggest that in the hominin lineage, with the
548 reduction of canine and diastema size, the incisors move posteriorly and the whole arcade becomes
549 parabolic. The upper and the lower arcades keep track of each other through high magnitudes of
550 covariation. Modern humans show lower magnitudes of covariation between the upper and the lower
551 jaw compared to apes, which might be related to a mechanically less challenging diet in living humans.
552 Alternatively or additionally, lower magnitudes might follow from a general relaxation in covariation
553 attributable to a systemic change in the hominin lineage that enabled the development of new shapes in
554 the first place.

555

556 **Acknowledgements**

557 We are grateful to Alexander Stoessel for his constructive feedback on this manuscript and the
558 anonymous reviewers and the editors for the helpful comments. We thank the following curators and
559 museums for access to their specimens: Ingo Bechmann and Christine Feja, Anatomisches Institut der
560 Universität Leipzig; Ottmar Kullmer and Friedemann Schrenk, Senckenberg Museum, Frankfurt/Main;
561 Frieder Mayer and Nora Lange, Museum für Naturkunde, Berlin; the Ivorian authorities and Christophe
562 Boesch, MPI-EVA, Leipzig; the Natural History Museum, London; S. Ivan Perez, Facultad de Ciencias
563 Naturales y Museo, Universidad Nacional de la Plata, Buenos Aires; the Smithsonian's Division of
564 Mammals (Dr. Kristofer Helgen); Martin Fischer, Phyletisches Museum Jena; Lynn Copes for providing
565 the modern human scans and the Human Origins Program (Dr. Matt Tocheri), Washington, D.C., for the
566 scans of USNM specimens used in this research ([http://humanorigins.si.edu/evidence/3d-](http://humanorigins.si.edu/evidence/3d-collection/primate)
567 [collection/primate](http://humanorigins.si.edu/evidence/3d-collection/primate)). These scans were acquired through the generous support of the Smithsonian 2.0
568 Fund and the Smithsonian's Collections Care and Preservation Fund. We thank Jean-Jacques Hublin for
569 supporting this research, which was funded by the Max Planck Society.

570

571 **References**

- 572 Ackermann, R.R., 2002. Patterns of covariation in the hominoid craniofacial skeleton: implications for
573 paleoanthropological models. *J. Hum. Evol.* 43, 167-187.
- 574 Ackermann, R.R., 2005. Ontogenetic integration of the hominoid face. *J. Hum. Evol.* 48, 175-197.
- 575 Ackermann, R.R., Cheverud, J.M., 2000. Phenotypic covariance structure in tamarins (genus *Saguinus*): A
576 comparison of variation patterns using matrix correlation and common principal component
577 analysis. *Am. J. Phys. Anthropol.* 111, 489-501.
- 578 Adams, D.C., 2016. Evaluating modularity in morphometric data: challenges with the RV coefficient and
579 a new test measure. *Methods Ecol. Evol.* 7, 565-572.
- 580 Almquist, A.J., 1974. Sexual differences in the anterior dentition in African primates. *Am. J. Phys.*
581 *Anthropol.* 40, 359-367.
- 582 Anapol, F., Lee, S., 1994. Morphological adaptation to diet in platyrrhine primates. *Am. J. Phys.*
583 *Anthropol.* 94, 239-261.
- 584 Angle, E.H., 1899. Classification of malocclusion. *The Dental Cosmos* 41, 248-264.
- 585 Armelagos, G.J., Van Gerven, D.P., Goodman, A.H., Calcagno, J.M., 1989. Post-Pleistocene facial
586 reduction, biomechanics and selection against morphologically complex teeth: a rejoinder to
587 Macchiarelli and Bondioli. *Hum. Evol.* 4, 1-7.
- 588 Black, G.V., 1902. *Descriptive Anatomy of the Human Teeth*, 5th edition. SS White Manufacturing
589 Company, Philadelphia.
- 590 Boesch, C., Bi, G., Bertin, Z., Anderson, D.P., Stahl, D., 2006. Food choice in Tai chimpanzees: are cultural
591 differences present? In: Hohmann, G., Robbins, M., Boesch, C. (Eds.), *Feeding Ecology in Apes*
592 *and Other Primates*. Cambridge University Press, Cambridge.
- 593 Bookstein, F., 2016. The inappropriate symmetries of multivariate statistical analysis in geometric
594 morphometrics. *Evol. Biol.* 43, 277-313.
- 595 Bookstein, F.L., 1991. *Morphometric Tools for Landmark Data: Geometry and Biology*. Cambridge
596 University Press, Cambridge.

597 Bookstein, F.L., Gunz, P., Mitteroecker, P., Prossinger, H., Schaefer, K., Seidler, H., 2003. Cranial
598 integration in *Homo*: singular warps analysis of the midsagittal plane in ontogeny and evolution.
599 J. Hum. Evol. 44, 167-187.

600 Bouvier, M., 1986. A biomechanical analysis of mandibular scaling in Old World monkeys. Am. J. Phys.
601 Anthropol. 69, 473-482.

602 Brace, C.L., Rosenberg, K.R., Hunt, K.D., 1987. Gradual change in human tooth size in the Late
603 Pleistocene and Postpleistocene. Evolution 41, 705-720.

604 Brader, A.C., 1972. Dental arch form related with intraoral forces: PR= C. Am. J. Orthod. 61, 541-561.

605 Braga, J., 1998. Chimpanzee variation facilitates the interpretation of the incisive suture closure in South
606 African Plio-Pleistocene hominids. Am. J. Phys. Anthropol. 105, 121-135.

607 Bromage, T.G., 1989. Ontogeny of the early hominid face. J. Hum. Evol. 18, 751-773.

608 Broomell, I.N., 1902. Anatomy and Histology of the Mouth and Teeth. P. Blackiston's Son & Co,
609 Philadelphia.

610 Carlson, D.S., Vangerven, D.P., 1977. Masticatory function and post-Pleistocene evolution in Nubia. Am.
611 J. Phys. Anthropol. 46, 495-506.

612 Cassidy, K.M., Harris, E.F., Tolley, E.A., Keim, R.G., 1998. Genetic influence on dental arch form in
613 orthodontic patients. Angle Orthod. 68, 445-454.

614 Cheverud, J.M., 1996a. Developmental integration and the evolution of pleiotropy. Am. Zool. 36, 44-50.

615 Cheverud, J.M., 1996b. Quantitative genetic analysis of cranial morphology in the cotton-top (*Saguinus*
616 *oedipus*) and saddle-back (*S. fuscicollis*) tamarins. J. Evol. Biol. 9, 5-42.

617 Clarke, R.J., 2012. A *Homo habilis* maxilla and other newly-discovered hominid fossils from Olduvai
618 Gorge, Tanzania. J. Hum. Evol. 63, 418-428.

619 Copes, L., 2012. Comparative and experimental investigations of cranial robusticity in mid-Pleistocene
620 hominins. Doctoral dissertation. Arizona State University.

621 Corruccini, R.S., 1991. Anthropological aspects of orofacial and occlusal variations and anomalies. In:
622 Kelley, M.A., Larsen, C.S. (Eds.), Advances in Dental Anthropology. Wiley-Liss, New York, pp. 295-
623 323.

624 Courtenay, J., Groves, C.P., Andrews, P., 1998. Interior intra-island variation? An assessment of the
625 differences between Bornean and Sumatran orangutans. In: Schwartz, J.H. (Ed.), *Orang-utan*
626 *Biology*. Oxford University Press, New York, pp. 19-29.

627 Currier, J.H., 1969. A computerized geometric analysis of human dental arch form. *Am. J. Orthod.* 56,
628 164-179.

629 Doran-Sheehy, D., Mongo, P., Lodwick, J., Conklin-Brittain, N.L., 2009. Male and female western gorilla
630 diet: preferred foods, use of fallback resources, and implications for ape versus Old World
631 monkey foraging strategies. *Am. J. Phys. Anthropol.* 140, 727-738.

632 Engel, G.A., 1979. Preformed arch wires: reliability of fit. *Am. J. Orthod.* 76, 497-504.

633 Fruciano, C., Franchini, P., Meyer, A., 2013. Resampling-based approaches to study variation in
634 morphological modularity. *PLOS ONE* 8, e69376.

635 Galdikas, B.M.F., 1988. Orangutan Diet, Range, and Activity at Tanjung Puting, Central Borneo. *Int. J.*
636 *Primatol.* 9, 1-35.

637 Garn, D.H., 1968. Human dental arch form determination from cranial anatomy through conic section
638 geometry. M. S. Thesis. Temple University School of Dentistry, Philadelphia.

639 Genet-Varcin, E., 1969. *A la Recherche du Primate Ancêtre de l'Homme: Primates et Évolution*. Editions
640 Boubée, Paris.

641 Gittins, S.P., Raemaekers, J.J., 1980. Siamang, lar and agile gibbons. In: Chivers, D.J. (Ed.), *Malayan Forest*
642 *Primates: Ten Years Study in Tropical Rainforest*. Plenum, New York, pp. 63-195.

643 Gonzalez-Jose, R., Van der Molen, S., Gonzalez-Perez, E., Hernandez, M., 2004. Patterns of phenotypic
644 covariation and correlation in modern humans as viewed from morphological integration. *Am. J.*
645 *Phys. Anthropol.* 123, 69-77.

646 Good, P., 2000. *Permutation Tests: A Practical Guide To Resampling Methods For Testing Hypotheses*.
647 Springer, New York.

648 Goswami, A., 2006. Morphological integration in the carnivoran skull. *Evolution* 60, 169-183.

649 Grabowski, M.W., Polk, J.D., Roseman, C.C., 2011. Divergent patterns of integration and reduced
650 constraint in the human hip and the origins of bipedalism. *Evolution* 65, 1336-1356.

- 651 Greenfield, L.O., 1992. Origin of the human canine: a new solution to an old enigma. Yearb. Phys.
652 Anthropol. 35, 153-185.
- 653 Groves, C.P., 1970. Population Systematics of Gorilla. J. Zool. 161, 287-300.
- 654 Groves, C.P., 1972. Systematics and phylogeny of gibbons. In: Rumbaugh, D.M. (Ed.), Gibbon and
655 Siamang. Karger, Basel, pp. 1-89.
- 656 Groves, C.P., Westwood, C., Shea, B.T., 1992. Unfinished business, Mahalanobis and a clockwork orang.
657 J. Hum. Evol. 22, 327-340.
- 658 Gunz, P., Harvati, K., 2007. The Neanderthal "chignon": variation, integration, and homology. J. Hum.
659 Evol. 52, 262-274.
- 660 Guy, F., Brunet, M., Schmittbuhl, M., Viriot, L., 2003. New approaches in hominoid taxonomy:
661 Morphometrics. Am. J. Phys. Anthropol. 121, 198-218.
- 662 Hellman, M., 1919. Dimensions vs. form in teeth and their bearing on the morphology of the dental
663 arch. Int. J. Orthod. Oral Surg. 5, 615-651.
- 664 Hellman, M., 1942. Factors influencing occlusion. Angle Orthod. 12, 3-27.
- 665 Henry, A.G., 2010. Plant foods and the dietary ecology of Neandertals and modern humans. Doctoral
666 dissertation. George Washington University.
- 667 Herring, S.W., 1993. Functional morphology of mammalian mastication. Am. Zool. 33, 289-299.
- 668 Hrdlicka, A., 1940. Lower jaw. Am. J. Phys. Anthropol. 27, 281-308.
- 669 Hylander, W., 1972. Functional significance of Eskimo craniofacial morphology. Am. J. Phys. Anthropol.
670 37, 441.
- 671 Hylander, W.L., 1975. Incisor Size and Diet in Anthropoids with Special Reference to Cercopithecidae.
672 Science 189, 1095-1098.
- 673 Hylander, W.L., 1979. Functional significance of primate mandibular form. J. Morphol. 160, 223-239.
- 674 Johanson, D.C., White, T.D., 1979. A systematic assessment of early African hominids. Science 203, 321-
675 330.
- 676 Johanson, D.C., White, T.D., Coppens, Y., 1978. A new species of the genus *Australopithecus* (Primates:
677 Hominidae) from the Pliocene of eastern Africa. Kirtlandia 28, 1-14.

678 Kay, R.F., Hylander, W.L., 1978. The dental structure of mammalian folivores with special reference to
679 primates and Phalangerioidea (Marsupialia). In: Montgomery, G.C. (Ed.), *The Ecology of Arboreal*
680 *Folivores*. Smithsonian Institution Press, Washington, pp. 173-191.

681 Kelley, J., 1995. Sexual dimorphism in canine shape among extant great apes. *Am. J. Phys. Anthropol.* 96,
682 365-389.

683 Kimbel, W.H., Deleuzene, L.K., 2009. "Lucy" redux: A review of research on *Australopithecus afarensis*.
684 *Yearb. Phys. Anthropol.* 52, 2-48.

685 Kimbel, W.H., Johanson, D.C., Rak, Y., 1997. Systematic assessment of a maxilla of *Homo* from Hadar,
686 Ethiopia. *Am. J. Phys. Anthropol.* 103, 235-262.

687 Klingenberg, C.P., McIntyre, G.S., 1998. Geometric morphometrics of developmental instability:
688 Analyzing patterns of fluctuating asymmetry with Procrustes methods. *Evolution* 52, 1363-1375.

689 Knott, C.D., 1998. Changes in orangutan caloric intake, energy balance, and ketones in response to
690 fluctuating fruit availability. *Int. J. Primatol.* 19, 1061-1079.

691 Larsen, C.S., 1995. Biological changes in human populations with agriculture. *A. Rev. Anthropol.* 24, 185-
692 213.

693 Le Gros Clark, W.E., 1950. Hominid characters of the australopithecine dentition. *J. Roy. Anthropol. Inst.*
694 80, 37-54.

695 Leakey, M.G., Spoor, F., Dean, M.C., Feibel, C.S., Anton, S.C., Kiarie, C., Leakey, L.N., 2012. New fossils
696 from Koobi Fora in northern Kenya confirm taxonomic diversity in early *Homo*. *Nature* 488, 201-
697 204.

698 Leutenegger, W., Kelly, J.T., 1977. Relationship of sexual dimorphism in canine size and body size to
699 social, behavioral, and ecological correlates in anthropoid primates. *Primates* 18, 117-136.

700 Leutenegger, W., Shell, B., 1987. Variability and sexual dimorphism in canine size of *Australopithecus*
701 and extant hominoids. *J. Hum. Evol.* 16, 359-367.

702 Lieberman, D.E., 2008. Speculations about the selective basis for modern human craniofacial form. *Evol.*
703 *Anthropol.* 17, 55-68.

704 Lieberman, D.E., Ross, C.F., Ravosa, M.J., 2000. The primate cranial base: ontogeny, function, and
705 integration. *Am. J. Phys. Anthropol.* 113.S31, 117-169.

706 Lockwood, C.A., 1997. Variation in the face of *Australopithecus africanus* and other African hominoids.
707 Doctoral dissertation. University of the Witwatersrand.

708 MacConaill, M.A., Scher, E.A., 1949. The ideal form of the human dental arcade, with some prosthetic
709 application. Dent. Rec. 69, 285-302.

710 MacKinnon, J., 1974. Behavior and Ecology of Wild Orangutans (*Pongo pygmaeus*). Anim. Behav. 22, 3-
711 74.

712 Makedonska, J., Wright, B.W., Strait, D.S., 2012. The effect of dietary adaption on cranial morphological
713 integration in capuchins (Order Primates, Genus *Cebus*). PLOS ONE 7, e40398.

714 Mardia, K.V., Bookstein, F.L., Moreton, I.J., 2000. Statistical assessment of bilateral symmetry of shapes.
715 Biometrika 87, 285-300.

716 Marroig, G., Cheverud, J.M., 2001. A comparison of phenotypic variation and covariation patterns and
717 the role of phylogeny. Ecology, and ontogeny during cranial evolution of New World monkeys.
718 Evolution 55, 2576-2600.

719 Marroig, G., Shirai, L.T., Porto, A., de Oliveira, F.B., De Conto, V., 2009. The Evolution of Modularity in
720 the Mammalian Skull II: Evolutionary Consequences. Evol. Biol. 36, 136-148.

721 Marroig, G., Vivo, M., Cheverud, J.M., 2004. Cranial evolution in sakis (Pithecia, Platyrrhini) II:
722 evolutionary processes and morphological integration. Evol. Biol. 17, 144-155.

723 McCollum, M.A., Ward, S.C., 1997. Subnasalveolar anatomy and hominoid phylogeny: evidence from
724 comparative ontogeny. Am. J. Phys. Anthropol. 102, 377-405.

725 McConkey, K.R., Ario, A., Aldy, F., Chivers, D.J., 2003. Influence of forest seasonality on gibbon food
726 choice in the rain forests of Barito Ulu, Central Kalimantan. Int. J. Primatol. 24, 19-32.

727 Mitteroecker, P., Bookstein, F., 2007. The conceptual and statistical relationship between modularity
728 and morphological integration. Syst. Biol. 56, 818-836.

729 Mitteroecker, P., Bookstein, F., 2008. The evolutionary role of modularity and integration in the
730 hominoid cranium. Evolution 62, 943-958.

731 Mitteroecker, P., Gunz, P., 2009. Advances in Geometric Morphometrics. Evol. Biol. 36, 235-247.

732 Mitteroecker, P., Gunz, P., Windhager, S., Schaefer, K., 2013. A brief review of shape, form, and
733 allometry in geometric morphometrics, with applications to human facial morphology. *Hystrix*
734 24, 59-66.

735 Mooney, M.P., Siegel, M.I., 1991. Premaxillary-maxillary suture fusion and anterior nasal tubercle
736 morphology in the chimpanzee. *Am. J. Phys. Anthropol.* 85, 451-456.

737 Noback, M.L., Harvati, K., 2015. Covariation in the human masticatory apparatus. *Anat. Rec.* 298, 64-84.

738 Oxnard, C.E., Lieberman, S.S., Gelvin, B.R., 1985. Sexual dimorphisms in dental dimensions of higher
739 primates. *Am. J. Primatol.* 8, 127-152.

740 Plavcan, J.M., 1993. Canine size and shape in male anthropoid primates. *Am. J. Phys. Anthropol.* 92, 201-
741 216.

742 Plavcan, J.M., 2002. Taxonomic variation in the patterns of craniofacial dimorphism in primates. *J. Hum.*
743 *Evol.* 42, 579-608.

744 Plavcan, J.M., van Schaik, C.P., 1992. Intrasexual competition and canine dimorphism in anthropoid
745 primates. *Am. J. Phys. Anthropol.* 87, 461-477.

746 Porto, A., de Oliveira, F.B., Shirai, L.T., De Conto, V., Marroig, G., 2009. The evolution of modularity in
747 the mammalian skull I: morphological integration patterns and magnitudes. *Evol. Biol.* 36, 118-
748 135.

749 Prasad, M., Hussain, M.Z., Shetty, S.K., Kumar, T.A., Khaur, M., George, S.A., Dalwai, S., 2013. Median
750 mandibular flexure at different mouth opening and its relation to different facial types: A
751 prospective clinical study. *J. Nat. Sci. Biol. Med.* 4, 426-430.

752 Preuschoft, H., 1989. Biomechanical approach to the evolution of the facial skeleton of hominoid
753 primates. *Fortschr. Zool.* 35, 421-431.

754 Preuschoft, H., Witzel, U., 2004. Functional structure of the skull in hominoidea. *Folia Primatol.* 75, 219-
755 252.

756 Ravosa, M.J., 1990. Functional assessment of subfamily variation in maxillomandibular morphology
757 among Old World monkeys. *Am. J. Phys. Anthropol.* 82, 199-212.

758 Remane, A., 1921. Beiträge zur Morphologie des Anthropoidengebisses. *Archiv für Naturgeschichte.*
759 *Abteilung A* 87, 1-179.

760 Richards, M.P., Pettitt, P.B., Stiner, M.C., Trinkaus, E., 2001. Stable isotope evidence for increasing
761 dietary breadth in the European mid-Upper Paleolithic. *Proc. Natl. Acad. Sci.* 98, 6528-6532.

762 Rightmire, G.P., 1993. Variation among early *Homo* crania from Olduvai Gorge and the Koobi Fora
763 region. *Am. J. Phys. Anthropol.* 90, 1-33.

764 Rijksen, H.D., 1978. A fieldstudy on Sumatran orang utans (*Pongo pygmaeus abelii*, Lesson 1827):
765 Ecology, behaviour and conservation. Doctoral dissertation. Mededelingen
766 Landbouwhogeschool Wageningen, Netherlands.

767 Rogers, M.E., Abernethy, K., Bermejo, M., Cipolletta, C., Doran, D., McFarland, K., Nishihara, T., Remis,
768 M., Tutin, C.E.G., 2004. Western gorilla diet: A synthesis from six sites. *Am. J. Primatol.* 64, 173-
769 192.

770 Rohlf, F.J., Corti, M., 2000. Use of two-block partial least-squares to study covariation in shape. *Syst.*
771 *Biol.* 49, 740-753.

772 Rohlf, F.J., Slice, D., 1990. Extensions of the Procrustes method for the optimal superimposition of
773 landmarks. *Syst. Zool.* 39, 40-59.

774 Rörer-Ertl, O., 1984. Orang-Utan-Studien. Hironymus Verlag, Neuried.

775 Sardi, M.L., Ramirez Rozzi, F., Pucciarelli, H.M., 2004. The Neolithic transition in Europe and North Africa.
776 The functional craneology contribution. *Anthropol. Anz.* 62, 129-145.

777 Schmittbuhl, M., Rieger, J., Le Minor, J.M., Schaaf, A., Guy, F., 2007. Variations of the mandibular shape
778 in extant hominoids: Generic, specific, and subspecific quantification using elliptical Fourier
779 analysis in lateral view. *Am. J. Phys. Anthropol.* 132, 119-131.

780 Schwartz, J.H., 1995. *Skeleton Keys: An Introduction To Human Skeletal Morphology, Development, and*
781 *Analysis.* Oxford University Press, New York.

782 Scott, J.H., 1957. The shape of the dental arches. *J. Dent. Res.* 36, 996-1003.

783 Smilde, A.K., Kiers, H.A.L., Bijlsma, S., Rubingh, C.M., van Erk, M.J., 2009. Matrix correlations for high-
784 dimensional data: the modified RV-coefficient. *Bioinformatics* 25, 401-405.

785 Spoor, F., Gunz, P., Neubauer, S., Stelzer, S., Scott, N., Kwekason, M., Dean, C., 2015. Reconstructed
786 *Homo habilis* holotype OH 7 suggests deep-rooted species diversity in early *Homo*. *Nature* 513,
787 83-86.

788 Teaford, M.F., Ungar, P.S., Grine, F.E., 2002. Paleontological evidence for the diets of African Plio-
789 Pleistocene hominins with special reference to early *Homo*. In: Ungar, P.S., Teaford, M.F. (Eds.),
790 Human Diet: Its Origin and Evolution. Bergin & Garvey, Westport, CT, pp. 143-166.

791 Tobias, P.V., 1967. Olduvai Gorge. Vol. 2. The Cranium and Maxillary Dentition of *Australopithecus*
792 (*Zinjanthropus*) *boisei*. Cambridge University Press, Cambridge.

793 Tobias, P.V., 1991. Olduvai Gorge IV: The Skulls, Endocasts and Teeth of *Homo habilis*. Cambridge
794 University Press, Cambridge.

795 Tutin, C.E.G., Fernandez, M., 1993. Composition of the Diet of Chimpanzees and Comparisons with That
796 of Sympatric Lowland Gorillas in the Lope Reserve, Gabon. *Am. J. Primatol.* 30, 195-211.

797 Uchida, A., 1998. Variation in tooth morphology of Gorilla gorilla. *J. Hum. Evol.* 34, 55-70.

798 Villmoare, B.A., Dunmore, C., Kilpatrick, S., Oertelt, N., Depew, M.J., Fish, J.L., 2014. Craniofacial
799 modularity, character analysis, and the evolution of the premaxilla in early African hominins. *J.*
800 *Hum. Evol.* 77, 143-154.

801 von Cramon-Taubadel, N., 2011. Global human mandibular variation reflects differences in agricultural
802 and hunter-gatherer subsistence strategies. *Proc. Natl. Acad. Sci.* 108, 19546-19551.

803 Ward, C.V., Leakey, M.G., Walker, A., 2001. Morphology of *Australopithecus anamensis* from Kanapoi
804 and Allia Bay, Kenya. *J. Hum. Evol.* 41, 255-368.

805 Watts, D.P., 1984. Composition and Variability of Mountain Gorilla Diets in the Central Virungas. *Am. J.*
806 *Primatol.* 7, 323-356.

807 Weidenreich, F., 1936. The mandibles of *Sinanthropus pekinensis*: a comparative study. *Palaeontologia*
808 *Sinica*, Series D 3, 162.

809 Weijs, W.A., Hillen, B., 1984. Relationships between masticatory muscle cross-section and skull shape. *J.*
810 *Dent. Res.* 63, 1154-1157.

811 Weijs, W.A., Hillen, B., 1986. Correlations between the cross-sectional area of the jaw muscles and
812 craniofacial size and shape. *Am. J. Phys. Anthropol.* 70, 423-431.

813 White, T.D., Suwa, G., Simpson, S., Asfaw, B., 2000. Jaws and teeth of *Australopithecus afarensis* from
814 Maka, Middle Awash, Ethiopia. *Am. J. Phys. Anthropol.* 111, 45-68.

- 815 Witzel, U., Preuschoft, H., 1999. The bony roof of the nose in humans and other primates. *Zool. Anz.*
816 238, 103-115.
- 817 Witzel, U., Preuschoft, H., 2002. Function-dependent shape characteristics of the human skull.
818 *Anthropol. Anz.* 60, 113-135.
- 819 Wold, H., 1966. Estimation of principal components and related models by iterative least squares. In:
820 Krishnaiah, P.R. (Ed.), *Multivariate Analysis*. Academic Press, New York, pp. 391-420.
- 821 Wolfram Research Inc., 2010. *Mathematica*. Wolfram Research Inc., Champaign, Illinois.
- 822 Wood, B., 1991. *Hominid Cranial Remains From Koobi Fora*. Clarendon Press, Oxford.
- 823 Wood, B., 1992. Origin and evolution of the genus *Homo*. *Nature* 355, 783-790.
- 824 Wrangham, R.W., Jones, J.H., Laden, G., Pilbeam, D., Conklin-Brittain, N., 1999. The raw and the stolen:
825 Cooking and the ecology of human origins. *Curr. Anthropol.* 40, 567-594.
- 826 Young, N.M., Wagner, G.P., Hallgrímsson, B., 2010. Development and the evolvability of human limbs.
827 *Proc. Natl. Acad. Sci.* 107, 3400-3405.
- 828

829 **Figure captions**

830 Figure 1. Data acquisition. Landmarks are placed on the alveolar margin (blue) and on the cervix
831 of the postcanine dentition (orange) of the maxilla (a) and the mandible (c); b) and d) show the
832 wireframe of the symmetrized data.

833 Figure 2. Arcade shape space of maxillae (a, b) and mandibles (c). *Homo* is shown in yellow, *Pan*
834 in blue, *Gorilla* in black, *Pongo* in red, and *Hylobates* in green. The wireframes show the shape variation
835 along PC1, PC2 and PC3. In PC3, *Hylobates* mandibles plot similarly to the maxillae and are therefore not
836 shown.

837 Figure 3. Two-block partial least squares (PLS) analysis between the maxillary and the
838 mandibular arcade. Group colors as in Figure 2. The wireframes show the shape changes along SW1 (a),
839 SW2 (b), and SW3 (c). All groups scatter along the diagonal. *Hylobates* shows a shift along SW2, parallel
840 with the hominid trajectory.

841 Figure 4. Two-block partial least squares (PLS) analysis between the maxillary and the
842 mandibular arcade for each group separately: *Pan* (a), *Gorilla* (b), *Pongo* (c), *Homo* (d), and *Hylobates*
843 (e). Males are shown in dark, females in light colors. The wireframes show the shape changes along
844 SW1.

845 Figure 5. Distribution of the covariance ratio (CR) (sample size 30, 1000 iterations). *Homo* shows
846 lower overall values than the apes

847 Figure 6. Female (red) and male (blue) mean shapes for the maxilla (a-e) and the mandible (f-j).
848 Mean shape differences statistically significant at $\alpha \leq 0.05$; *p*-values given below the wireframe. Female
849 and male arcades are significantly different between *Pan*, *Gorilla*, *Pongo*, and *Homo* (maxillae). *Homo*
850 mandibles and *Hylobates* are not significantly different. Smaller canines (females) are associated with a

851 more posteriorly placed anterior arcade. In the postcanine row, the premolars are placed more
852 anteriorly, the molars remain unaffected.

853 Figure 7. PLS analysis between the premaxilla (anterior, black wireframe) and the maxilla
854 (posterior, red wireframe). Group colors as in Figure 2.

855 Figure 8. Distributions of the CR (sample size 30, 1000 iterations) between the premaxilla and
856 the maxilla.

857 SOM Figure S1. Species distribution within the genus *Gorilla* in arcade shape space of maxillae
858 (a, b) and mandibles (c). *G. gorilla* is shown in black, *G. beringei* in gray. The wireframes show the shape
859 variation along PC1, PC2 and PC3.

860 SOM Figure S2. Species distribution within the genus *Pongo* in arcade shape space of maxillae (a,
861 b) and mandibles (c). *P. pygmaeus* is shown in light red, *P. abelii* in yellow, *Pongo* sp. in dark red. The
862 wireframes show the shape variation along PC1, PC2 and PC3.

863 SOM Figure S3. Species distribution within the genus *Hylobates* in arcade shape space of
864 maxillae (a, b) and mandibles (c). *H. muelleri* is shown in green, *H. lar* in pink, *H. klossii* in orange, *H.*
865 *albibarbis* in blue, *H. agilis* in yellow. The wireframes show the shape variation along PC1, PC2 and PC3.

866 SOM Figure S4. Shape changes along SW1, SW2, SW3 between the maxillary arcade (black) and
867 mandibular arcade (red) for each group separately. The left wireframe in each group is the negative
868 extreme, the right wireframe is the positive extreme for each SW.

869 SOM Figure S5. PLS scores of the maxillary and the mandibular arcade for each group
870 separately. Dark colors in each group are males, light colors females.

871 SOM Figure S6. Sexual dimorphism in the magnitude of covariation. For each taxon, distributions
872 (sample size 15, 1000 iterations) and single values (horizontal lines) are given. Light group colors are

873 females, dark group colors are males. Female hominids show higher magnitudes (distributions and
874 single values) than their male counterparts. *Hylobates* show the opposite pattern. In *Pan*, the single
875 values are almost the same.

876 SOM Figure S7. PLS scores of the premaxilla and the maxilla for each group separately. Dark
877 colors in each group are males, light colors females.

878 SOM Figure S8. Shape changes along SW1 (a), SW2 (b), and SW3 (c) between the premaxilla
879 (black) and the maxilla (red). The left column is the negative extreme, the right column is the positive
880 extreme.

881 SOM Figure S9. Sexual dimorphism in the covariance ratio (CR) between the premaxilla and the
882 maxilla. For each taxon distributions subsample size is 15 (1000 iterations); light group colors are
883 females, dark group colors are males.

884

Table 1. Sample composition

Taxon	♀	♂	Unknown sex
<i>Pan</i>	25	19	-
<i>Homo</i>	16	24	13
<i>Gorilla</i>	22	31	-
<i>Pongo</i>	25	27	-
<i>Hylobates</i>	29	30	4

Table 2. Results of the PC analyses.

	Eigenvalues	% variance	Cumulative %
<i>Maxillae</i>			
PC1	0.00789125	66.958	66.958
PC2	0.00129874	11.02	77.978
PC3	0.00067848	5.757	83.735
<i>Mandibles</i>			
PC1	0.01059464	70.795	70.795
PC2	0.00163547	10.928	81.723
PC3	0.00093735	6.264	87.987

Table 3. Percentage of the explained covariance between the upper and the lower arcade for the first five PLS components. Symmetrized data; combined PLS: group-mean centered; separate PLS: sex-mean centered.

% covariance combined PLS		% covariance separate PLS				
		<i>Pan</i>	<i>Gorilla</i>	<i>Pongo</i>	<i>Homo</i>	<i>Hylobates</i>
PLS1	61.39	68.44	51.79	57.80	65.53	79.40
PLS 2	21.55	11.74	19.31	15.00	13.27	8.78
PLS 3	7.92	6.26	13.06	10.85	6.55	4.44
PLS 4	3.46	3.50	3.83	6.38	3.41	1.36
PLS 5	1.38	2.60	2.65	2.39	2.09	1.27

Table 4. Results for the CR and the permutation tests for the upper and the lower jaw. Subsample size for the permutation is 30, 1000 resamples. Below, Bonferroni corrected p -values from permutation test of group mean differences (considered significant at $\alpha < 0.05$).

	<i>Pan</i>	<i>Gorilla</i>	<i>Pongo</i>	<i>Homo</i>	<i>Hylobates</i>
CR	0.777	0.771	0.786	0.726	0.789
Min	0.712	0.715	0.739	0.635	0.721
Max	0.894	0.867	0.886	0.892	0.891
Median	0.805	0.804	0.812	0.755	0.814
Mean	0.805	0.802	0.813	0.757	0.814
SD	0.031	0.025	0.024	0.044	0.030
<i>Pan</i>	15	14.580	0.165	0.015	0.150
<i>Gorilla</i>	14.536	15	0.270	0.015	0.165
<i>Pongo</i>	0.120	0.240	15	0.015	11.613
<i>Homo</i>	0.015	0.015	0.015	15	0.015
<i>Hylobates</i>	0.210	0.150	10.984	0.015	15

Table 5. Results for the correlation coefficient (r) between the upper and the lower arcade.

	PLS1	PLS2	PLS3
<i>Pan</i>	0.757	0.841	0.858
<i>Gorilla</i>	0.645	0.853	0.757
<i>Pongo</i>	0.728	0.851	0.828
<i>Homo</i>	0.695	0.664	0.668
<i>Hylobates</i>	0.831	0.569	0.625

Table 6

Table 6. Results for the CR and the permutation tests grouped by species and sex (subsample size 15, 1000 resamples) for the upper and the lower arcade. Below, Bonferroni corrected p -values from permutation test of sex mean differences (considered significant at $\alpha < 0.05$).

	<i>Pan</i> M	<i>Pan</i> F	<i>Gorilla</i> M	<i>Gorilla</i> F	<i>Pongo</i> M	<i>Pongo</i> F	<i>Homo</i> M	<i>Homo</i> F	<i>Hylo</i> M	<i>Hylo</i> F
CR	0.837	0.831	0.720	0.809	0.771	0.834	0.756	0.859	0.836	0.795
Min	0.786	0.797	0.682	0.749	0.737	0.778	0.715	0.843	0.771	0.719
Max	0.926	0.921	0.912	0.910	0.893	0.933	0.904	0.872	0.940	0.935
Median	0.847	0.872	0.804	0.840	0.809	0.858	0.793	0.864	0.862	0.844
Mean	0.853	0.869	0.803	0.839	0.810	0.857	0.796	0.862	0.861	0.842
SD	0.030	0.023	0.038	0.029	0.023	0.031	0.033	0.007	0.032	0.040
p	0.950		0.002		0.002		0.002		0.214	

Table 7. Correlation coefficients (r) grouped by species and sex for the upper and the lower arcade.

	PLS1	PLS2	PLS3
<i>Pan</i> M	0.924	0.810	0.879
<i>Pan</i> F	0.910	0.780	0.827
<i>Gorilla</i> M	0.660	0.906	0.876
<i>Gorilla</i> F	0.764	0.780	0.800
<i>Pongo</i> M	0.764	0.703	0.797
<i>Pongo</i> F	0.852	0.784	0.705
<i>Homo</i> M	0.745	0.858	0.860
<i>Homo</i> F	0.907	0.741	0.806
<i>Hylobates</i> M	0.865	0.779	0.802
<i>Hylobates</i> F	0.787	0.837	0.728

Table 8. Percentage of the total variance explained by size.

Taxon	Maxillae		Mandibles	
	%	<i>p</i>	%	<i>p</i>
<i>Pan</i>	6.80	<0.001	4.84	<0.001
<i>Gorilla</i>	14.25	<0.001	8.53	<0.001
<i>Pongo</i>	12.58	<0.001	9.68	<0.001
<i>Homo</i>	2.49	<0.001	2.38	<0.004
<i>Hylobates</i>	2.36	<0.001	3.55	<0.001
Total	5.38	<0.001	8.39	<0.001

Table 9. Percentage of the explained covariance between the premaxilla and the maxilla for the first five PLS components. Symmetrized data; combined PLS: group-mean centered; separate PLS: sex-mean centered.

% covariance combined PLS		% covariance separate PLS				
		<i>Pan</i>	<i>Gorilla</i>	<i>Pongo</i>	<i>Homo</i>	<i>Hylobates</i>
PLS1	84.14	44.28	85.99	67.19	79.75	74.83
PLS2	10.18	39.06	7.37	17.07	7.72	13.70
PLS3	2.20	6.44	2.21	6.47	5.41	3.95
PLS4	1.16	3.55	1.42	3.87	2.50	3.11
PLS5	0.74	1.96	1.06	1.85	1.42	1.27

Table 10. Results for the CR and the permutation tests between the premaxilla and the maxilla. Subsample size for the permutation is 30. 1000 resamples. Below, Bonferroni corrected p -values from permutation test of group mean differences (considered significant at $\alpha < 0.05$).

	<i>Pan</i>	<i>Gorilla</i>	<i>Pongo</i>	<i>Homo</i>	<i>Hylobates</i>
CR	0.678	0.731	0.702	0.694	0.646
Min	0.611	0.621	0.620	0.596	0.576
Max	0.803	0.882	0.844	0.834	0.809
Median	0.707	0.751	0.727	0.719	0.688
Mean	0.707	0.749	0.726	0.719	0.687
SD	0.031	0.044	0.037	0.038	0.042
<i>Pan</i>	15	0.030	8.571	5.784	0.300
<i>Gorilla</i>	0.060	15	0.135	0.015	0.015
<i>Pongo</i>	8.796	0.120	15	3.267	0.270
<i>Homo</i>	5.619	0.015	3.282	15	2.742
<i>Hylobates</i>	0.554	0.015	0.360	2.892	15

Table 11. Results of the correlation coefficient (r) between the premaxilla and the maxilla.

	PLS1	PLS2	PLS3
<i>Pan</i>	0.752	0.885	0.395
<i>Gorilla</i>	0.815	0.707	0.668
<i>Pongo</i>	0.798	0.752	0.492
<i>Homo</i>	0.814	0.794	0.060
<i>Hylobates</i>	0.758	0.796	0.207